Floral herbivory does not reduce pollination-mediated fitness in shelter rewarding Royal Irises

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Running title: Effect of florivory on pollination

ABSTRACT:

Florivory, the damage to flowers by herbivores can affect fitness both directly and indirectly. Flowers consumed by florivores may fail to produce fruit or produce lower seed set because of direct damage to reproductive organs. In addition, eaten flowers are less attractive to pollinators because of reduced or modified advertisement, which reduces pollination services. While observational data are abundant, experimental evidence is scarce and results are contrasting. We tested experimentally the effect of florivory on both pollinator visitation and reproductive success in three species of the Royal Irises, which have large flowers that are attractive to pollinators, and potentially also for florivores. We hypothesized that florivory will reduce pollen deposition due to reduced attractiveness to pollinators, while fruit set and seed set will depend on the extent of florivory. We performed artificial florivory in two experiments over two years. In the first experiment, each of the three floral units of a single *Iris* flower was subject to either low or high artificial florivory, or left un-touched as control. We counted the number of pollen grains deposited on each of the three stigmas as a measure of pollinator visitation. In the second experiment, three flowers of the same plant received low, high, or no artificial florivory and were further recorded for fruit and seed production. In 2016, high artificial florivory revealed lower number of pollen grains on stigmas of *Iris atropurpurea*, but in 2017 there was no difference. Similarly, number of pollen grains in high artificial was lower than low florivory in 2017 in *I. petrana*. No significant effect of florivory was found on pollen grain deposition, fruit set or seed set. The results remained consistent across species and across years. The results undermine the assumption that flower herbivory is necessarily antagonistic interaction and suggests that florivores may not be strong selection agents on floral reproductive biology in the *Oncocyclus* irises.

43   **KEYWORDS**: herbivory, *Iris* section *Oncocyclus*, fitness, natural selection, pollen limitation

INTRODUCTION

Flowers of animal-pollinated plants are the major means of plants to advertise and attract

pollinators. Floral traits serve as signals to the pollinators, usually fit to the most-efficient

pollinator (Fenster et al., 2004). Flowers advertise through visual as well as olfactory signals, and

even by acoustic signature, in order to stand out of the canopy or other flowering species

(Schiestl and Johnson, 2013). Consequently, flower traits that increase attraction are selected by

pollinators (Harder and Johnson, 2009). For example, floral size, contributing to the visibility of

the flower, is positively selected by pollinators (Sletvold et al., 2010, 2016; Campbell et al.,

1991; Conner and Rush, 1996; Harder and Johnson, 2009; Lavi and Sapir, 2015). However,

flowers are costly organs that require investment in resources for production and maintenance.

Other selection agents also act on floral traits, either in concert with, or in contrast to the

selection exerted by pollinators (Strauss and Whittall, 2006). For example, floral size is under

contrasting selection regimes, positive by pollinators and negative by drought and water loss

(Galen, 2000; Carroll et al., 2001). Color polymorphism is also thought to be maintained by the

combined effect of mutualists (pollinators) and antagonists (Carlson and Holsinger, 2010), or by

opposing selection directions by herbivores and pathogens (Frey, 2004). Although many studies

have examined floral adaptations to pollinators, the role of non-pollinator selection agents in

shaping floral evolution and plant reproductive success is still understudied.

   Florivory, the damage of flowers by herbivores, is widespread across plant taxa and

ecosystems (Gonzáles et al., 2016; Burgess, 1991). Florivores comprise of various taxonomic

orders of animals that consume the entire flower (or the buds) or floral parts (bracts, sepals,

petals, nectaries, stamens, pistils or pollen). Florivory may affect plant fitness directly or

indirectly, by reducing fruiting or seed-set, and can consequently affect population dynamics

(Louda and Potvin, 1995). Florivory reduces fitness directly when reproductive parts are

consumed, whereas indirect effect can be through effects on pollinator behavior and hence

reduced pollination services. An increasing number of studies suggest that florivory can decrease

pollinator visitation rate and pollination success (reviewed in Gonzáles et al., 2016, but see Zhu

et al., 2017).

Methods for studying the effect of florivory and its consequences on pollination primarily

focus on the effect of florivore presence or the extent of florivory on pollinator visits, using

natural encounters and manipulative studies. For example, Kirk et al. (1995) placed black spots

on flowers to simulate presence of florivore beetle and found that bees were likely to avoid

flowers with mimicry of florivore presence. Although artificial florivory performed in the field

can provide an estimate to the direct effect of florivory on pollination, few studies implement this

method. Moreover, behavior of pollinators and/or measure of maternal fitness (measured as the

number of visits or seed set, respectively) are often used to estimate the effect of florivory on

pollination (e.g., Cardel and Koptur, 2010).Pollen deposition on stigma can be a direct evidence

for pollination success, but has rarely been incorporated in studies that investigate the effect of

florivory on pollination. Therefore, controlled manipulative experiments that measure pollen

deposition and fitness are needed to estimate directly the interaction of florivory and pollination

success.

In a survey of the literature, we found only a small number of studies that used controlled

experimental florivory that mimics natural florivory by artificially manipulating or removing

parts of the petals and tested for effect on pollination. Of these, seven studies found negative

effect on pollination success, one study found mixed effects that depended on flower morph

(Carper et al., 2016), and one study found no effect (Tsuji et al., 2016). Thus, it remains unclear

whether florivory affects plant fitness indirectly via negative effects on pollinators. As an

example, McCall (2008) found that both natural and artificial petal damage indeed reduced

fitness, and while it deterred pollinator activity, the effect was a result of petal physical damage

rather than reduced pollinators' activity, as pollen addition treatment did not recover fruit-set.

Hence, for fully understanding the multiple effects of florivory on fitness and the interplay

between direct and indirect effects it requires further experiments in diverse plant species.

We studied the effect of florivory on pollination success in three species of the Royal Irises (*Iris*

section *Oncocyclus*) in their natural habitats in Israel (Fig.1 A–C). The Royal Irises comprise

about 30 species across the Middle East, and most are narrow endemics (Rix, 1997; Mathew,

1989). This group of irises is well studied as a system for pollination by shelter reward to male

Eucera bees (Fig.1 D; Sapir et al., 2005; Sapir et al., 2006; Watts et al., 2013; Monty et al., 2006;

Vereecken et al., 2013; Lavi and Sapir, 2015). Flowers of the Royal Irises grow singly on a stem,

but a plant (genet) comprises one to hundreds of stems (ramets) in a well-defined patch. Flowers

of the Royal Irises comprise of three identical units, each bearing one upright and one pendant

petal (standard and fall, respectively), and one petaloid style, curved above the fall to create a

tunnel where the reproductive organs reside (Fig. 1). The three stigmas, located at the top of the

entrance of each of the pollination tunnels. Pollen is deposited on stigmas by bees that move

among flowers as they seek shelter (Sapir et al., 2005). The three style lobes are merged at the

base to form one united style. A previous study showed that pollinating one style is sufficient to

produce seeds in all three carpels in the ovary (Watts et al., 2013), thus, although each

pollination tunnel functions as a single ecological unit of pollination, pollinator visit in one

tunnel affects the fitness of the whole flower. All the species are self-incompatible and require

pollination to set fruits (Sapir et al., 2005).

Figure 1 – (A) *Iris atropurpurea* in Netanya; (B) *Iris petrana* in Yeruham; (C) *Iris lortetii* in
Malkiya; (D) Male *Eucera* bee, the specific pollinator of the Royal Irises, sheltering within a
pollination tunnel of *Iris petrana*; (E-G) Natural florivory in flowers of *Iris atropurpurea* (E), *I.*
petrana (F) and *I. lortetii* (G).
119

During a survey in the year 2015 and also based on previous observations we found that the

Royal Irises are eaten by various florivores, from snails and true bugs to grasshoppers, birds and

goats, and the intensity of damage ranges from a few superficial scratches or poke marks, to

>90% of damage to floral tissue (M. Ghara and Y. Sapir, unpubl. Res.; Fig. 1 E–G). From our

observations, it appears that all flower parts are potentially eaten, including tepals, the petaloid

style, anthers and ovaries. While damage to reproductive organs may obviously reduce fitness

directly, we asked whether florivory of the advertising floral parts affects fitness indirectly

through reduced attraction to pollinators. In order to estimate the effect of florivory on pollinator

visitation and pollination success, we manipulated the flowers to simulate two levels of florivory,

i.e., high (more than 50% damage) and low (10-30% damage), and compared them to control

flowers without damage. We asked the following questions: (1) Does florivory affect pollinator

visitation? (2) Does florivory affect fruit set and seed set? We used pollen deposition on the

stigma as a surrogate for estimating pollinator visitation and used fruit- and seed-set to estimate

overall effect of florivory on (maternal) fitness. Accounting for two measures of pollination

success simultaneously is likely to reveal an indication of the possible effect of florivory on

pollination-mediated fitness in the Royal Irises.

MATERIALS AND METHODS

Study species and sites — Three species of the Royal Irises (Iridaceae), *Iris atropurpurea* Baker,

I. petrana Dinsm., and *I. lortetii* Barbey ex Boiss, were used in this study. Experiments were

conducted in the natural environment at the largest population for each of the species. *Iris*

atropurpurea was studied in Netanya Iris Reserve (32.28°N, 34.84°E, alt. 35 m), located on

stable coastal sand dunes in Mediterranean climate and consisting of mostly low shrub

vegetation. Population size is estimated >1,000 plants (Yardeni et al., 2016). Flowering season,

and hence experiment time, is earlier compared to other species of the Royal Irises, starting as

early as mid-January, and peaks in February. Experiments in *I. atropurpurea* in Netanya were

conducted between 12 February 2016 and 02 March 2016, and between 19 February 2017 and 07

March 2017. *Iris petrana* was studied in Yeruham Iris Reserve (31.02°N, 34.97°E, alt. 560 m), a

large population (estimated >10,000 plants) growing on sandy loess hills over Neogene

sandstone in arid climate. Vegetation is sparse desert shrubs, mostly *Retama raetam* (Forssk.)

Webb and *Anabasis articulata* (Forssk.) Moq. Flowering season is in March, and experiments in

I. petrana were conducted between 05 March and 14 March in 2016 and between 19 March and

02 April in 2017. The shift in dates in 2017 resulted due to a delay in flowering period by about

two weeks. *Iris lortetii* was studied in two sub-populations near Malkiya in the upper Galilee

(central coordinates: 33.09°N, 35.52°E, alt. 620 m). Populations of *I. lortetii* are sparse and

relatively small, thus, two sub-populations at a distance of 3 km of each other were pooled to

achieve a sufficient sample size. Plants are growing on Eocene limestone in mesic Mediterranean

climate and vegetation is open woodland dominated by *Quercus calliprinos* Webb and *Pistacia*

atlantica Desf. trees, accompanied by dense herbaceous vegetation. *Iris lortetii* is the late

blooming species among the Israeli species of the Royal Irises; experiments were conducted

between 29 March and 07 April in 2016. The experiments were not conducted on *Iris lortetii* in

2017 because of high rhizome herbivory in 2016 and therefore a potential decrease in sample

size.

Plants for the experiments were randomly selected in a dense part of the population in

Netanya, or along transects in Yeruham. In Malkiya the plants are sparse and plants in all genets

located were used. The three experiments described below were conducted simultaneously in

time with only a single experiment conducted in each genet to avoid the joint effect of several

treatments.

Florivory manipulations and effect on pollination —

To study the effect of florivory on pollen deposition, a single bud in a genet was bagged with

a cloth bag and upon anthesis the bag was removed and each flower unit was given one of three

treatments. In each treatment, the fall treated was paired with the standard in the opposite side of

the flower. This way, a putative pollinator approaching the flower, assuming it is approaching in

perpendicular angle to the pollination tunnel, is seeing the projection of the fall in the proximate

part of the flower and the standard in the distant part. It is important to note that a confounding

effect of such design is that the flower's symmetry is changed. However, this design is

advantageous because it enables to control for differences among flowers in, e.g., size of the

pollination tunnel or colour. We employed two florivory treatments in each flower: High damage

–the lower petal and its opposite upper petal were manually damaged up to 50% or more of their

area, pierced by an awl of 6-8 mm in diameter. Low damage – 10-30% of the petals' area were

pierced using awl. The third pair of petals was left un-touched as control (Fig. 2).

Figure 2 – Artificial florivory manipulations exemplified in *Iris atropurpurea*. (A) Within flower
manipulation – each floral unit treated as either high florivory (>50% petal cut), low florivory
(10-30% petal removed by hole puncher), or control (no treatment). These treatments were used
for testing the effect on pollination. (B–D) Flowers used for testing the effect of florivory on
fitness. (B) Flower treated as high florivory. (C) Flower treated as low florivory. (D) Control
flower.

188
To control for the possible effect of the contact between the metal awl and the flower the awl

was rubbed on the petal surface in the control treatments. In addition, because damaging the

petals required holding a layer of tissue paper against the awl, we also gently rubbed tissue paper

under the surface of the petal in the control treatment. Note that florivory was done on the petals

and not on the pollination tunnel, to explicitly address the hypothesis that change in visual

display disrupt pollination services in the presence of florivory. Flowers that had natural

florivory were not used. To prevent naturally occurring florivores from damaging the flowers,

the stem of the flower was coated with a layer of double sided sticky paper tape, as well as a

layer of Petroleum jelly (Vaseline). Occasionally we found insects trapped on the Vaseline layer,

and in some rare cases, we found florivores that passed this barrier mainly because the sticky

tape lost its stickiness owing to sand in the air that got stuck to the tape. Flowers found to be

damaged naturally (mostly by flying insects, snails, or mammals) were discarded in order to

account for the effect of controlled artificial florivory only.

After manipulating the flowers they left open for two consecutive evenings to enable

pollinators to visit naturally. On the morning of the third day, the entire stigmas of the three

pollination units were collected in vials containing 1 ml of 70% ethanol. Collected stigmas were

brought to the laboratory for pollen counting and kept at room temperature. The vial containing

the stigma with pollen was washed until there was no remaining pollen in alcohol. This was to

ensure that pollen was not lost due to the protocol followed.Pollen grains were stained using a

drop of basic fucshin (Calberla's stain). Each stigma was then dissected in a drop 70% aqueous

glycerol (Dafni et al., 2005), mounted on microscope glass slide, and the pollen present on the

stigma was counted under dissecting microscope (WILD Heerbrugg Switzerland M5-72558).

Florivory manipulations and effect on maternal fitness — To study the effect of florivory on

seed set, three buds of the same genet, roughly of the same developmental stage (i.e., before

emerging from bracts) were selected and bagged to avoid bud florivory. Upon anthesis, each of

the three flowers was randomly assigned to one of the florivory treatments described above, i.e.,

high, low, or control. To control for the effect of visual attraction mediated by flower size (Lavi

and Sapir, 2015) we measured flower length as a surrogate for display size of each flower before

manipulation. Flower length was measured from the bottom of the lower petal to the top end of

the upper one. The flowers were left open to enable naturally occurring pollination. At the end of

the season, approximately three weeks after the end of the flowering in each site, the fruits were

collected and brought to the laboratory. In 2016, fruits of *I. atropurpurea, I. petrana* and *I.*

lortetii were collected in 21 March, 02 April and 27 April, respectively. In 2017, fruits were

collected in 12 March and 22 March for *I. atropurpurea* and on 13 April and 27 April for *I.*

petrana. Fruits were kept in paper bags at room temperature until seed maturation. Fitness was

recorded as presence or absence of a fruit (binomial data), and as the number of viable seeds

(count data).

Data analyses —Data were analyzed in R (R Development Core Team, 2014) using R-studio

interface. Because each stigma is not independent of the other two stigmas of the same flower,

we normalized the number of pollen grains on stigma by subtracting the number from the mean

number of pollen grains on the three stigmas of the same flower. These normalized values of

number of pollen grains deposited on stigmas were analyzed using generalized linear models

(GLMs) with year and treatment effects nested within species (note that in *I. lortetii* experiment

was done only in 2016). For analyses using fruit or seeds as explained variables, we used GLM

and incorporated flower size as a covariate. For seeds as a response variable, we used only the

subset of flowers that set a fruit. In order to account for non-normal distribution, we used GLM

with binomial distribution errors for fruits and GLM with quasi-Poisson distribution errors for

seeds.

RESULTS

Pollen deposition on stigma — Using absolute number of pollen grains deposited on the stigma,

we found significant differences between species and years (GLM with quasi-Poisson

distribution; $F_{2,685}=4.39$, $P=0.013$ for species and $F_{2,685}=25.09$, $P<0.001$ for year effect, nested

within species). In *Iris atropurpurea*, 62 and 44 flowers were manipulated in 2016 and 2017,

respectively. Of these, 49 (26.3%) of the stigmas did not receive pollen at all in 2016 and 8

(5.3%) in 2017. Flower units with artificial florivory, either high or low, received lower number

of pollen grains, compared to control, un-manipulated units in 2016 (Contrast analysis:

P=0.024), but the difference in number of pollen grains between high and low damage was not

significant (P=0.115; Fig. 3 A). In 2017 higher number of pollen grains was deposited on

stigmas overall, but with no differences among treatments (P=0.65; Fig. 3 B).

In *I. petrana*, 66 and 49 flowers were manipulated in 2016 and 2017, respectively. All stigmas

received pollen grains in 2016, but in 2017, 51 (34%) did not receive any pollen grain. Number

of pollen grains deposited on the stigmas was an order of magnitude larger than in *I.*

atropurpurea in 2016 (Fig. 3 A), but in 2017 number of pollen grains was slightly less than in *I.*

atropurpurea (Fig. 3 B).No significant treatment effect was found in 2016 ($F_{2,195}$=0.60, P=0.242;

Fig. 3 A). In 2017, however, we found significant higher number of pollen grains deposited on

stigmas in flower units that manipulated in the low florivory treatment, compared to high

florivory and control treatments (Contract analysis: P<0.001; Fig. 3 B).

In *Iris lortetii*, 11 flowers were manipulated in 2016. As in *I. petrana* in 2016, all stigmas

received pollen and stigmas in units treated by high artificial florivory received pollen grains in a

similar level as the control, untreated units, both higher than medium artificial florivory

treatment (Fig. 3 A). However, this difference was not significant as well ($F_{2,30}$=0.914, P=0.412).

Figure 3 – Mean number of pollen grains counted on stigma (± standard errors) as a function of florivory treatment in three species. (A) Pollen grains in 2016 experiment; (B) Pollen grains in 2017 experiment. Bars are fractions ± standard errors. Same letters above bars denote non-significant difference (P>0.05) between means. n.s. – not significant. Significance test was performed using GLM and contrasts analyses on relative number of pollen grains, normalized for each stigma by the mean number of pollen grains in all three stigmas of the same flower.

Fruit and seed sets — In *Iris atropurpurea*, 80 flowers, out of 96 flowers treated, were included

in the final analyses in 2016, because 16 of the flowers were either not found or damaged. Of

these, only 13 flowers set fruits, indicating extreme pollinator limitation and lack of pollinator

visitations. Treatment did not affect either fruit-set or number of seeds ($F_{2,75}$=0.69, P=0.501, and

$F_{2,8}$=0.26, P=0.774, respectively; Fig. 4 A and C). In both analyses flower size did not affect

significantly (P=0.266 and P=0.554 for fruits and seeds, respectively). In 2017, 124 flowers were

included in the experiment, of which four were damaged. Of the remaining 120 flowers, 23

flowers set fruits. As in 2016, no effect of the treatment was found, neither on fruit-set, nor on

number of seeds ($F_{2,116}$=0.37, P=0.695, and $F_{2,19}$=0.95, P=0.404, respectively; Fig. 4 B and D).

Similar to 2016, flower size as covariate did not affect fruit-set or seed-set (P=0.188 and

P=0.857, respectively).

Figure 4 – Fitness as a function of artificial florivory manipulations. (A) Fruit-set (fraction of
flowers that set fruits) in three species in 2016 and (B) in two species in 2017. (C) Seed-set
(mean number of seeds in a fruit) in 2016 and (D) in 2017. Bars are means ± standard errors.

In *I. petrana* population in Yeruham, 132 flowers were treated in 2016, but 29 flowers of all

treatments were eaten by goats that entered the reserve illegally and ate wilting flowers and

young fruits in the pre-dispersal stage. Of the remaining 103 treated flowers, 31 flowers set

fruits. Flower length (before treatment) significantly affected fruit set ($F_{1,98}$=5.09, P=0.026), but

with no significant interaction with treatment ($F_{2,96}$=0.26, P=0.770). Although control flowers

produced almost twice fraction of fruits compared to florivory treatment (34% vs. 18%; contrasts

analysis: P=0.022; Figure 4 A), this difference was not significant when controlled for flower

size ($F_{2,98}$=1.89, P=0.156). Number of seeds was not affected by treatment ($F_{2,98}$=1.89, P=0.156,

n=31; Fig. 4 C). Flower size did not affect seed-set ($F_{1,26}$=1.23, P=0.278). In 2017, only six

flowers were eaten or not found, out of 188 flowers treated. As in 2016, no effect of the

treatment was found, neither on fruit-set, nor on number of seeds ($F_{2,133}$=1.64, P=0.199, and

$F_{2,29}$=0.72, P=0.495, respectively; Fig. 4 B, D). As opposed to 2016, flower size as covariate did

not affect fruit-set but did affect seed-set (P=0.472 and P<0.001, respectively). No interaction

was found between flower size and florivory treatment in their effect on seed-set (P=0.463).

In the two sites of *Iris lortetii*, 61 out of 62 flowers treated (12 in Avivim and 50 in Malkiya)

were found at the end of the season and included in the analyses. No significant difference was

found among treatments ($F_{2,57}$=0.21, P=0.811; Fig. 4 A). Flower size affected fruit-set

($F_{1,57}$=4.42, P=0.40). Number of seeds was not significantly affected by treatment ($F_{2,12}$=0.37,

P=0.698; Fig. 4 C), and neither by flower size ($F_{1,12}$=0.92, P=0.356).

DISCUSSION

Florivory, the damage herbivores cause to floral organs, can affect fitness either directly by

consuming pollen or ovules or by physiological costs, or indirectly, by reducing plant attraction

signal for the pollinators (Burgess, 1991; McCall and Irwin, 2006). Here we tested for both direct

and indirect effects of florivory on fitness by executing artificial florivory and measuring both

fitness and pollination success. Our results do not fully support the hypothesis that florivory

affects pollination success in the Royal Irises either directly or indirectly. Instead, we show that

artificial damage to reproductive tissues in three species of the royal irises had inconsistent effect

on pollen deposition, but no effect on fitness (Fig. 3, 4).

Florivory has small effect on pollinators' behavior, but no overall effect on fitness. Limited

cost in reducing pollinator's services, no effect on fruit/seed-set. Adapted against florivory?

While numerous studies are concerned with the effect of florivory on plant fitness, controlled,

artificial florivory was rarely performed. Most studies examined flowers that were naturally

attacked by florivores (e.g., Meindl et al., 2013; Ruane et al., 2014; Eliyahu et al., 2015) or used

experimental florivore removal or prevention (e.g., Krupnick et al., 1999; Theis and Adler, 2012;

Althoff et al., 2013). Only a few studies implemented methods similar to ours, using cutting

flowers to simulate florivory. These studies revealed mixed results. For example, Sõber et al.

(2010) experimentally showed a correlation between extent of florivory and pollinator visitations

at both population and plant level. On the other hand, Tsuji et al. (2016) found no evidence for

pollinators discrimination against experimentally damaged flowers. Interestingly, mixed results

can be found within the same system: Carper et al. (2016) found differences between

heterostylous morphs in pollinator responses to artificial damage, and found no effect on fitness.

Our study adds to the puzzle by providing yet another piece of evidence that florivory itself does

not deter pollinators, nor reduces fitness.

Negative effect of florivory on pollination may act in two avenues. One possible effect of

florivory on pollinator behavior is the deterring of pollinators from eaten flowers. This is

achieved by either avoidance of flowers where florivore is visually detected (Kirk et al., 1995) or

by a change in volatile compounds (Kessler et al., 2013). Another possible effect of florivory on

pollination is mediated by the effect of the overall advertisement size of the flower and reducing

visual signaling for the pollinators (Sánchez-Lafuente, 2007). This may reduce number of visits

and lead to pollinator limitation or pollen limitation, which in turn reduces fruit-set and seed-set,

respectively (Sapir et al., 2015). This study was conducted in natural populations of which at

least one iris species, *Iris atropurpurea* experience strong pollinator limitation. It is likely that

the effect of pollen limitation in this population obscures the effect of florivory (McCall, 2010).

Thus, we propose that selection mediated by pollinator/pollen limitation is stronger than

selection pressure exerted by florivory. A previous study that tested for pollen limitation in two

Iris species showed that pollinator limitation provides conditions for pollinator-mediated

selection (Lavi and Sapir, 2015). Although for *Iris petrana* we found no evidence for pollen

limitation, still no effect of florivory was found, which contradicts this hypothesis. Our mixed

results suggest that while pollinators may be a selection agent on flower traits (as in Lavi and

Sapir, 2015), florivores do not act as selection agents because pollen limitation balances their

effect (cf. Jogesh et al., 2017), but this connection cannot be generalized beyond our specific

system.

Pollinators are thought to be the major selection agent on floral traits through their positive

effect on fitness; this, however, was challenged by observations on contrasting effects of abiotic

conditions or antagonistic biotic interactions (Herrera, 1996, reviewed in Strauss and Whittall,

2006). Strauss and Whittall (2006) proposed two scenarios of such mutualistic-antagonistic

effect, in which overall selection acts as either directional or stabilizing on floral traits. Based on

our results, we propose a third scenario, where florivory affects fitness at the same level as the

mean effect of pollinators and regardless floral trait. In this case, pollinator-mediated selection

will govern trait evolution, but the presence of florivory reduces or diminishes effect size (Figure

5). We speculate that florivory may not affect the direction of selection but the intensity of it.

Thus, we suggest that testing for the net selection mediated by pollinators should control

experimentally for the effect of florivores. In a follow-up experiment, we intend to test for net

pollinator-mediated selection on floral color in the Royal Irises. Floral pigment concentration is

expected to deter florivores and attract pollinators (de Jager and Ellis, 2014; McCall et al., 2013),

and given the results of the current study we assume that florivory may not necessarily reduce

fitness; instead, it is expected that a weak directional pollinator-mediated selection on floral color

will be detected after controlling for florivory.

Figure 5 – Hypothetical floral trait evolution as a function of selection by both mutualists (pollinators) and antagonists (florivores). (A) When pollinators and florivores exert selection in the same direction, the joint selection favors the same trait optimum. (B) When pollinators and florivores exert opposing selection on a trait, an intermediate trait optimum is favored. A and B are adapted from figure 7.1 in Strauss and Whittall (2006). (C) When florivores have no preference, or their effect is similar to mean fitness derived from pollinators' effect, trait will have optimum fitness selected by pollinators like in A.

While few previous studies have detected only slight (or no) effect of florivory on pollination, it may be further hypothesized that the Royal Irises are unique and thus do not represent a general rule relevant to other species. The Royal Irises present a unique pollination syndrome in which pollination is performed by night sheltering male bees (Sapir et al., 2005; Watts et al., 2013). Previous studies suggest that floral display itself is not necessarily the major attractant for pollinators of the Royal Irises (Lavi and Sapir, 2015). While our florivory-like manipulations were performed on petals, it is likely that these manipulations did not affect pollinator choice

because the shelter itself (the pollination tunnel) was not damaged. Because we have observed

florivory of stigma and anthers (M. Ghara pers. obs.), we suspect that florivory may affect male

and female fitness through consumption of reproductive organs, but we have yet to assess the

cost of florivory on reproductive organs. Nonetheless, while natural florivory is widespread in all

species and most populations of the Royal Irises in Israel, the estimated proportion of flowers of

which pollination tunnels were eaten is rather small (M. Ghara, manuscript in preparation). Thus,

our manipulation on petals accurately mimics natural florivory and our conclusions on the role of

florivory in selection on flowers of the Royal Irises are valid.

Finally, our study provides an application to conservation. Of the three species studied, two

(*Iris atropurpurea* and *I. lortetii*) are rare and endangered species (Sapir, 2016a, b).

Understanding the relative contribution of biotic interactions to population dynamics may shed

light on the factors affecting species survival in a way that may contribute for evidence-based

management. The study presented here suggests that reduced mutualistic interactions, namely,

pollination services, rather than antagonistic florivory, threatens the maintenance of positive

population growth in the Royal Irises.

CONCLUSIONS

Our experiments manipulations closely represent natural florivory in the Royal Irises. Florivores,

largely considered as antagonists, do not seem to effect pollination in all the species we

investigated. This is in contrast to some studies that found a negative effect of florivory on

pollination where damage to flowers petals show reduction in fruit set. Perhaps, this is because

of the unique pollination system in the Royal Irises where reward is shelter and the choice of

shelter is carried out during the dusk and probably florivory cannot be truly assessed during

those hours. Assessment of more study systems is thus required to arrive at a general conclusion

of whether florivory affects pollination.

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LITERATURE CITED

Althoff, D. M., W. Xiao, S. Sumoski, and K. A. Segraves. 2013. Florivore impacts on plant

reproductive success and pollinator mortality in an obligate pollination mutualism. *Oecologia,*

173: 1345-1354.

Bartkowska, M. P., and M. O. Johnston. 2015. Pollen limitation and its influence on natural

selection through seed set. *Journal of Evolutionary Biology,* 28: 2097-2105.

Botto-Mahan, C., P. A. Ramírez, C. G. Ossa, R. Medel, M. Ojeda-Camacho, and A. V.

González. 2011. Floral herbivory affects female reproductive success and pollinator visitation in

the perennial herb *Alstroemeria ligtu* (Alstroemeriaceae). *International Journal of Plant*

Sciences, 172: 1130-1136.

Burgess, K. 1991. *Florivory: The ecology of flower feeding insects and their host plants,*

PhD Thesis, Harvard University.

Campbell, D. R., N. M. Waser, M. V. Price, E. A. Lynch, and R. J. Mitchell. 1991.

Components of phenotypic selection: pollen export and flower corolla width in *Ipomopsis*

aggregata. Evolution, 45: 1458-1467.

Cardel, Y. J., and S. Koptur. 2010. Effects of florivory on the pollination of flowers: an

experimental field study with a perennial plant. *International Journal of Plant Sciences,* 171:

283-292.

Carlson, J. E., and K. E. Holsinger. 2010. Natural selection on inflorescence color

polymorphisms in wild Protea populations: The role of pollinators, seed predators, and intertrait

correlations. *American Journal of Botany,* 97: 934-944.

Carper, A. L., L. S. Adler, and R. E. Irwin. 2016. Effects of florivory on plant-pollinator

interactions: Implications for male and female components of plant reproduction. *American*

Journal of Botany, 103: 1061-1070.

Carroll, A. B., S. G. Pallardy, and C. Galen. 2001. Drought stress, plant water status, and

floral trait expression in fireweed, *Epilobium angustifolium* (Onagraceae). *American Journal of*

Botany, 88: 438-446.

Conner, J. K., and S. Rush. 1996. Effects of flower size and number on pollinator visitation

to wild radish, *Raphanus raphanistrum. Oecologia,* 105: 509-516.

Dafni, A., P. G. Kevan, and B. C. Husband. 2005**.** Practical Pollination Biology. Ontario:

Enviroquest.

de Jager, M. L., and A. G. Ellis. 2014. Floral polymorphism and the fitness implications of

attracting pollinating and florivorous insects. *Annals of Botany,* 113: 213-222.

Eliyahu, D., A. C. McCall, M. Lauck, and A. Trakhtenbrot. 2015. Florivory and nectar-

robbing perforations in flowers of pointleaf manzanita *Arctostaphylos pungens* (Ericaceae) and

their effects on plant reproductive success. *Arthropod-Plant Interactions,* 9: 613-622.

Fenster, C. B, W. S. Armbuster, P. Wilson, M. R. Dudash, and J. D. Thomson. 2004.

Pollination syndromes and floral specialization. *Annual Review of Ecology, Evolution and*

Systematics, 35: 375-403.

Frey, F. M. 2004. Opposing natural selection from herbivores and pathogens may maintain

floral-color variation in *Claytonia virginica* (Portulacacea). *Evolution,* 58: 2426-2437.

Galen, C. 2000. High and dry: drought stress, sex-allocation trade-offs, and selection on

flower size in the alpine wildflower *Polemonium viscosum* (Polemoniaceae). *The American*

Naturalist, 156: 72-83.

Gonzáles, W. L., L. H. Suárez, and E. Gianoli. 2016. Genetic variation in the reduction of

attractive floral traits of an annual tarweed in response to drought and apical damage. *Journal of*

Plant Ecology, 9: 629-635.

Harder, L. D., and S. D. Johnson. 2009. Darwin's beautiful contrivances: evolutionary and

functional evidence for floral adaptation. *New Phytologist,* 183: 530-545.

Herrera, C. M. 1996**.** Floral traits and plant adaptation to insect pollinators: a devil's

advocate approach. In: Lloyd DG, Barrett SCH, eds. *Floral biology: studies on floral evolution*

in animal-pollinated plants. New York: Chapman & Hall.

Jogesh, T., R. P. Overson, R. A. Raguso, and K. A. Skogen. 2017. Herbivory as an

important selective force in the evolution of floral traits and pollinator shifts. *AoB Plants*.

Kessler, D., C. Diezel, D. G. Clark, T. A. Colquhoun, and I. T. Baldwin. 2013. Petunia

flowers solve the defence/apparency dilemma of pollinator attraction by deploying complex

floral blends. *Ecology Letters,* 16: 299-306.

Kirk, W. D. J, M. Ali, and K. N. Breadmore. 1995. The effects of pollen beetles on the

foraging behaviour of honey bees. *Journal of Apicultural Research,* 34: 15-22.

Krupnick, G. A., A. E. Weis, and D. R. Campbell. 1999. The consequences of floral

herbivory for pollinator service to *Isomeris arborea. Ecology,* 80: 125-134.

Lavi, R., and Y. Sapir. 2015. Are pollinators the agents of selection for the extreme large

size and dark color in Oncocyclus irises? *New Phytologist,* 205: 369-377.

Louda, S. M., M. A. Potvin. 1995. Effect of inflorescence-feeding insects on the

demography and lifetime of a native plant. *Ecology,* 76: 229-245.

Mathew, B. 1989. *The Iris.* London: Batsford.

McCall, A. C. 2008. Florivory affects pollinator visitation and female fitness in *Nemophila*

menziesii. Oecologia, 155: 729-737.

McCall, A. C. 2010. Does dose-dependent petal damage affect pollen limitation in an annual

plant? *Botany,* 88: 601-606.

McCall, A. C, and R. E. Irwin. 2006. Florivory: the intersection of pollination and herbivory.

Ecology Letters, 9: 1351-1365.

McCall, A. C., S. J. Murphy, C Venner, and M. Brown. 2013. Florivores prefer white versus

pink petal color morphs in wild radish, *Raphanus sativus. Oecologia,* 172: 189-195.

Meindl, G. A., D. J. Bain, and T-L. Ashman 2013. Edaphic factors and plant–insect

interactions: direct and indirect effects of serpentine soil on florivores and pollinators.

Oecologia, 173: 1355-1366.

Monty, A., L. Saad, and G. Mahy . 2006. Bimodal pollination system in rare endemic

Oncocyclus irises (Iridaceae) of Lebanon. *Canadian Journal of Botany,* 84: 1327–1338.

Muola, A., D. Weber D, L. E. Malm, P. A. Egan, R. Glinwood, A. L. Parachnowitsch, and J.

A. Stenberg. 2017. Direct and Pollinator-Mediated Effects of Herbivory on Strawberry and the

Potential for Improved Resistance. *Frontiers in Plant Science,* 8: 823.

Pohl, N., G. Carvallo, C. Botto-Mahan, and R. Medel. 2006. Nonadditive effects of flower

damage and hummingbird pollination on the fecundity of *Mimulus luteus. Oecologia,* 149: 648-

655.

R Development Core Team. 2014**.** R: A Language and Environment for Statistical

Computing. Vienna: R Foundation for Statistical Computing.

Rix, M. 1997. Section *Oncocyclus* (Siemssen) Baker. In: The species group of the British

Iris Society, ed. *A Guide to Species Irises*: Cambridge University Press.

Ruane L. G., A. T. Rotzin, and P. H. Congleton. 2014. Floral display size, conspecific

density and florivory affect fruit set in natural populations of *Phlox hirsuta*, an endangered

species. *Annals of Botany,* 113: 887-893.

Sánchez-Lafuente, A. M. 2007. Corolla herbivory, pollination success and fruit predation in

complex flowers: An experimental study with *Linaria lilacina* (Scrophulariaceae). *Annals of*

Botany, 99: 355-364.

Sapir, Y. 2016*a*. *Iris atropurpurea. The IUCN Red List of Threatened Species 2016*: e.

T13161450A18611400.

Sapir, Y. 2016*b*. *Iris lortetii. The IUCN Red List of Threatened Species 2016*:

e.T13161743A18612235.

Sapir, Y., A. Dorchin, and Y. Mandelik. 2015. Indicators of pollinator decline and pollen

limitation. In: Armon RH, Hänninen O, eds. *Environmental Indicators*: Springer Netherlands.

Sapir. Y., A. Shmida, and G. Ne'eman. 2005. Pollination of the *Oncocyclus* irises (*Iris*:

Iridaceae) by night-sheltering male bees. *Plant Biology,* 7: 417-424.

Sapir, Y., A. Shmida, and G. Ne'eman. 2006. Morning floral heat as a reward to the

pollinators of the *Oncocyclus* irises. *Oecologia,* 147: 53-59.

Schiestl, F. P., and S. D. Johnson. 2013. Pollinator-mediated evolution of floral signals.

Trends in Ecology and Evolution, 28: 307-315.

Sletvold, N., J. M. Grindeland, and Ågren J. 2010. Pollinator-mediated selection on floral

display, spur length and flowering phenology in the deceptive orchid *Dactylorhiza lapponica*.

New Phytologist, 188: 385–392.

Sletvold N., M. Tye M, and J. Ågren. 2017. Resource- and pollinator-mediated selection on

floral traits. *Functional Ecology,* 31: 135-141.

Sõber, V., M. Moora, and T. Teder. 2010. Florivores decrease pollinator visitation in a self-

incompatible plant. *Basic and Applied Ecology,* 11: 669-675.

Strauss, S. Y., and J. B. Whittall. 2006**.** Non-pollinator agents of selection on floral traits. In:

Harder LD, Barrett SCH, eds. *Ecology and Evolution of Flowers*. Oxford: Oxford University

Press.

Theis, N., and L. S. Adler. 2012. Advertising to the enemy: enhanced floral fragrance

increases beetle attraction and reduces plant reproduction. *Ecology,* **93**: 430-435.

Tsuji, K., M. K. Dhami, D. J. R. Cross, C. P. Rice, N. H.Romano, and T. Fukami. 2016.

Florivory and pollinator visitation: a cautionary tale. *AoB Plants,* 8.

Vereecken, N. J., A. Dorchin, A. Dafni, S. Hötling, S. Schulz, and S. Watts. 2013. A

pollinators' eye view of a shelter mimicry system. *Annals of Botany,* 111: 1155-1165.

Watts, S., Y. Sapir, B. Segal, and A. Dafni. 2013. The endangered *Iris atropurpurea*

(Iridaceae) in Israel: honey-bees, night-sheltering male bees and female solitary bees as

pollinators. *Annals of Botany,* 111: 395-407.

Yardeni, G., N. Tessler N, E. Imbert, and Y. Sapir. 2016. Reproductive isolation between

populations of *Iris atropurpurea* is associated with ecological differentiation. *Annals of Botany,*

118: 971-982.

Zhu, Y-R., M. Yang, J. C. Vamosi, W. S. Armbruster, T. Wan, and Y-B. Gong. 2017.

Feeding the enemy: loss of nectar and nectaries to herbivores reduces tepal damage and increases

pollinator attraction in *Iris bulleyana. Biology Letters,* **13**: 20170271.