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A Complex Evolutionary History for the Disease Susceptibility CDHR3 Locus

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1 Abstract

Selective pressures imposed by pathogens have varied among human populations throughout 2 3 their evolution, leading to marked inter-population differences at some genes mediating 4 susceptibility to infectious and immune-related diseases. A common polymorphism resulting in a C₅₂₉ versus T₅₂₉ change in the Cadherin-Related Family Member 3 (CDHR3) receptor is 5 associated with rhinovirus-C (RV-C) susceptibility and severe childhood asthma. Given the 6 7 morbidity and mortality associated with RV-C dependent respiratory infections and asthma, we hypothesized that the protective variant has been under selection in the human population. 8 9 Supporting this idea, a recent cross-species outbreak of RV-C among chimpanzees in Uganda, 10 which carry the ancestral 'risk' allele at this position, resulted in a mortality rate of 8.9%. Using publicly available genomic data, we sought to determine the evolutionary history and role of 11 selection acting on this infectious disease susceptibility locus. The protective variant is the 12 13 derived allele and is found at high frequency worldwide, with the lowest relative frequency in 14 African populations and highest in East Asian populations. There is minimal population structure among haplotypes, and we detect genomic signatures consistent with a rapid increase in 15 frequency of the protective allele across all human populations. However, given strong evidence 16 that the protective allele arose in anatomically modern humans prior to their migrations out of 17 18 Africa and that the allele has not fixed in any population, the patterns observed here are not consistent with a classical selective sweep. We hypothesize that patterns may indicate frequency-19 dependent selection worldwide. Irrespective of the mode of selection, our analyses show the 20 21 derived allele has been subject to selection in recent human evolution.

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23 Keywords

24 Selection, rhinovirus-C, asthma, CDHR3, population genetics, evolution

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26 Introduction

27 There is accumulating evidence to suggest that immunity-related genes are preferential targets of

natural selection (reviewed in $^{1-5}$), supporting the notion that infectious diseases have been

important selective forces on human populations⁶. However, for most candidate loci, the

30 mechanisms and phenotypic effects underlying the observed signatures of selection remain

31 elusive.

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32 Human rhinoviruses (RV) are found worldwide and are the predominant cause of the common cold. While many RV infections cause only minor illness, type C strains of the virus are 33 34 associated with higher virulence. Unlike RV-A and RV-B, RV-C utilizes the Cadherin Related Family Member 3 (CHDR3) receptor to gain entry into host cells. A missense variant in CDHR3 35 (rs6967330, C529T) results in a 10-fold difference in virus binding and replication in vitro⁷, and 36 experimental evidence points to varied cell surface expression between the two forms of the 37 receptor as a likely mechanism⁸. Adding clinical support that CDHR3 functions as an RV-C 38 receptor, the ancestral allele at rs6967330 has been found to be associated with an increased risk 39 of respiratory tract illness by RV-C in two birth cohorts⁹. The allele is also associated with a 40 severe form of childhood asthma, in accordance with the known role of RV, particularly RV-C, 41 in triggering asthma exacerbations¹⁰. Taken together, these studies suggest that host genetics 42 mediate differing susceptibility to RV-C infection and asthma by affecting interactions between 43 the virus and its receptor (e.g. whether or not the protein is expressed on the cell surface and is 44 45 thus visible to the virus).

A recent outbreak of lethal respiratory illness among wild chimpanzees was attributed to 46 47 human RV-C crossing species boundaries. All members of the infected chimpanzee population were invariant for the ancestral variant at the homologous position corresponding to the human 48 rs6967330 variant. The outbreak resulted in a staggering 8.9% mortality rate ¹¹, and suggests that 49 RV-C infection can result in a high mortality rate in a susceptible population. As respiratory 50 51 infections, particularly prior to the availability of modern medical interventions, represent significant threats to human health¹²⁻¹⁵, CDHR3, and in particular rs6967330, represent a 52 promising candidate target of natural selection. In the present study, we investigated the 53 evolutionary history of this locus for which there already exists strong experimental and clinical 54 55 data linking genotype with phenotypes that appear to modulate disease susceptibility. 56

57 **Results & Discussion**

58 Long-term evolution of the *CDHR3* locus

59 Examination of the locus in an alignment of 100 vertebrate genomes^{16,17} revealed that the

60 *CDHR3* locus is highly conserved, with homologs present in 85 species (Figure 1). Tyrosine is

61 the ancestrally encoded amino acid at the homologous position 529 in the human protein

62 sequence. There appears to have been a mutation leading to an A529T amino acid change

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introduced in a common ancestor of opossums, Tasmanian devils, and wallabies that results in
the encoding of a phenylalanine at the homologous position. Numerous vertebrates (elephant,
horse, rabbit, pika, naked mole-rat, rat, mouse, golden hamster, Chinese hamster, prairie vole,
and lesser Egyptian jerboa) spread throughout the species tree also encode a different amino acid
(histidine) at this position. Whether these mutations are fixed in each of these species requires
further sequencing, and the effect of these nonsynonymous substitutions on protein expression
and function remain to be explored.

Providing further evidence that the locus has been evolutionary constrained, rs6967330 has a positive Genomic Evolutionary Rate Profiling (GERP) score of 4.39, signifying a deficit of substitutions compared to neutral expectations. Values of the statistic calculated for bases within *CDHR3* range from -12.2 to 6.08 (mean = -0.15, sd = 2.59) and from -12.3 to 6.17 chromosomewide (mean =-0.09, sd = 1.91)^{17,18}. The apparent conservation of this locus suggests it is important for the function of CDHR3 and thus makes it an interesting candidate of natural selection.

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78 **Population diversity patterns of the** *CDHR3* **locus**

79 Excluding *Homo sapiens*, sequencing data from the remaining extant species comprising all hominids (great apes) are invariant at the position corresponding to $rs6967330^{19}$. Genotyping of 80 an additional 41 chimpanzees whose community experienced a severe respiratory outbreak of 81 RV-C in 2013 revealed that all individuals were homozygous for the ancestral allele¹¹. In 82 examining hominin genomes, we find that Neanderthals and Denisovans carry the ancestral 83 84 allele, while ancient specimens of anatomically modern humans carry both the ancestral and derived alleles. In low coverage sequencing data of *H. sapiens* estimated to have lived between 85 86 5000-8000 years ago (using sequencing reads from 45 aDNA samples for which we felt confident making diploid genotype calls) we estimate the allele frequencies at rs6967330 to be 87 34.4% A (ancestral) and 65.5% G (derived) (Table S1)²⁰. Higher coverage aDNA extracted from 88 a 7,000 year old skeleton found in Germany and an 8,000 year old skeleton from the Loschbour 89 rock shelter in Luxembourg reveals heterozygotes at the locus²¹. Finally, a 45,000 year old early 90 *H. sapiens* from western Siberia is a homozygote for the derived allele²². Collectively, these data 91 92 suggest that the derived allele likely arose in the evolutionary branch leading to anatomically 93 modern humans, although population data for Neanderthals and Denisovans remains scarce.

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94 The locus represents a shared polymorphism in contemporary worldwide populations. Based on whole genome sequence data for 2504 individuals from 26 different populations²³, we 95 96 find that the derived ('protective') G allele is, on average, very common in East Asian populations (EAS, 93.0%), followed by admixed American populations (AMR, 85.7%), South 97 98 Asian populations (SAS, 80.0%), and European populations (EUR, 79.2%). It is least common in African populations (AFR, 73.5%), albeit also at high frequency. At the individual population 99 100 level, allele frequencies of the derived G allele range from 68.8% ("Mende in Sierra Leone", MSL) to 95.3% ("Peruvians from Lima, Peru", PEL) (Figure 2). 101 102 We next investigated linkage disequilibrium (LD) patterns around CDHR3 with Haploview²⁴. A tight LD block was identified on chromosome 7 from 105,657,078-105,659,873, 103 104 with only moderate LD decay extending up to 105,680,022. Considering only biallelic SNPs 105 with a minor allele frequency (MAF) > 0.01, haplotypes within these blocks were identified with the Pegas package in $\mathbb{R}^{25,26}$. Relationships among haplotypes occurring at >1% were inferred 106 107 using network analysis (Figure 3). Eight haplotypes at frequency $\geq 1\%$ were identified from the region of high LD, with the majority of individuals in all populations carrying the same 108 haplotype with the derived G allele (n = 3848); two less frequent haplotypes carrying the derived 109 110 allele are found in various geographic regions. The ancestral allele is found in the remaining five haplotypes, which vary in their distributions across regions. Twenty-eight haplotypes were found 111 using the same criteria of a MAF > 0.01 and haplotype frequency of > 1% in the larger genomic 112 113 block containing moderate LD. Phylogenetic reconstruction of the resulting haplotypes in this region resulted in a clear separation of those carrying the ancestral and derived alleles. 114 Unexpectedly, Chimp, Neanderthal and Denisovan haplotypes are closer to H. sapiens A-115 haplotypes suggesting that natural selection shaped the haplotype pattern (Figure S1). 116 117

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World-wide Selection at the locus

Given the morbidity and mortality associated with viral respiratory infections^{13,15} and severe childhood asthma^{12,14} particularly prior to the availability of modern medical interventions, we hypothesized that the frequency of the protective G allele (resulting in a Cys₅₂₉) at rs6967330 might have increased more rapidly than under neutrality. We performed genome-wide scans for selection in the 26 populations of the 1000 Genomes Project²³. To do so, we used the integrated haplotype score (iHS)²⁷ and the number of segregating sites by length

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 $(nS_1)^{28}$, which are designed to capture rapid increases in frequency of selected variants and have 125 been applied in the context of detecting recent positive selection^{27,29,30}. We calculated these 126 127 neutrality statistics in each population independently and normalized by frequency-bin for all 128 loci with a DAF \ge 0.2 (Figure S2). Fourteen populations presented extreme values of iHS and $nS_{\rm L} \ge 95^{\rm th}$ percentile of the distribution in the population) at rs6967330, with 2 and 5 129 populations falling in the 99th percentile for the two statistics, respectively. CDHR3 lies in a 130 131 region of high recombination (1.8cM/Mb) and rs6967330 resides in a male specific recombination hotspot^{17,31}. This may explain the slightly different patterns observed in $nS_{\rm L}$ and 132 133 iHS statistics, as an excess of extreme values of iHS have been observed at regions of low recombination²⁸. 134

To investigate whether selection has acted on all populations simultaneously, we 135 implemented a multi-population (MP) combined statistic approach for iHS and $nS_{\rm L}$. For each 136 SNP in each population, we combined the percent ranks of iHS and $nS_{\rm L}$ using Fisher's method, 137 and defined the MP-iHS and MP- $nS_{\rm I}$ as the resulting γ^2 value. We obtained a MP-iHS score of 138 156 and a MP-nS_I score of 157 for rs6967330. Both scores are in the 99th percentile, regardless 139 140 of whether we include only segregating sites in 2 or more populations or limit it to those SNPs 141 segregating in all 26 populations examined. This suggests that the derived allele at rs6967330 is advantageous and that non-neutral processes have acted on this locus across global populations. 142 143

144 A complex evolutionary scenario

145 We originally hypothesized that selection pressures imposed by RV-C infection may have 146 resulted in a classic selective sweep at the locus; however, our results here are not consistent with such a model. Preliminary dating estimates of RV-C point to a recent origin in the last few 147 thousand years⁸. We infer that the protective (derived) allele was already present in ancestral 148 149 populations at intermediate frequencies during this time frame (65.5% in aDNA dated 5,000-8,000 years ago). Haplotype-based statistics such as iHS and nS_L have low power to detect 150 selection occurring on standing variation with intermediate frequency³². This suggests that 151 152 patterns of variation observed at rs6967330 are unlikely to have resulted from selection imposed by RV-C itself; our findings suggest instead that an alternative selective agent(s) may have 153 been/be acting on the CDHR3 locus prior to emergence of RV-C. These findings well parallel 154 155 those observed at the chemokine receptor gene-5 (CCR-5), where an unknown historical

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selective pressure maintained a deletion in this gene that attenuates infectivity and disease
progression of HIV (reviewed in ³³), because that mutation clearly predates the emergence of
AIDS.

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160 Frequency-dependent Selection

Several lines of evidence point to the derived allele at rs6967330 arising before the out-of-Africa 161 162 dispersal, including a lack of haplotype structure (Figure 3) and the presence of the allele at homozygous state in aDNA from anatomically modern humans dating back to ~45,000 year 163 old^{20-22} . While we find signatures of selection at the locus, the protective variant has not fixed in 164 any of the contemporary populations examined (Figure 2), contrary to expectations for a strongly 165 positively selected variant that arose prior to human migrations out of Africa³⁴. We therefore 166 wondered if balancing selection could explain observed patterns of variation at the locus, as 167 168 balancing selection can maintain functional diversity over long periods of time through frequency-dependent selection, heterozygote advantage, pleiotropy, and fluctuating selection³⁵. 169 Until recently, evidence for balancing selection in the human genome was limited to a few 170 171 classical cases such as the heterozygous advantage conferred by the *HbS* sickle cell mutation against malaria^{36,37}, genes of the major histocompatibility complex/human leukocyte antigen 172 complex³⁸, and the ABO blood group³⁹. Balancing selection, however, has recently been 173 174 recognized as more prevalent than previously thought, particularly in shaping human immune system phenotypes^{35,40,41}. To test whether the rs6967330 locus has been under long-term 175 balancing selection (i.e. selection occurring over at least hundreds of thousands of generations 176 ^{38,42,43}), we examined the distribution of β , a recently developed summary statistic designed to 177 detect clusters of alleles at similar frequencies, in 1000 Genomes Project data⁴⁴. β was similar 178 179 across all populations around the rs6967330 locus, and values were not indicative of long-term balancing selection (Figure S3). 180

Balancing selection operating over shorter timescales clearly also plays a role in shaping human diversity (e.g. *HbS* sickle cell mutation^{36,37}). Such a short-term balancing selection scenario could explain the signals we detected with haplotype-based statistics, as genomic signatures of short-term balancing selection are predicted to be indistinguishable from incomplete sweeps of positive selection ^{27,41}. In both of these scenarios, the selected allele rapidly increases in frequency, with the allele eventually fixing under positive selection or

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oscillating around a steady state in the case of balancing selection. As a reminder, haplotype-187 188 based selection methods suggest that rs6967330 rose in frequency faster than under neutrality, 189 both at the global and the individual population level. Furthermore, population differentiation as measured with F_{ST} at this locus is low relative to other SNPs with the same frequency (Figure 190 191 S4), a finding that is also consistent with short-term balancing selection across multiple populations^{35,38,41}. Based on our observation that the derived variant at rs6967330 was present at 192 193 high frequencies in ancient human specimens and in the homozygous state in the case of a 194 45,000-year-old fossil, we posit that this allele may have started to increase in frequency in 195 ancestral populations before reaching its current equilibrium frequency independently in worldwide populations. The derived allele ranges in frequency from 68.8-95.3% in modern 196 197 human populations, which is compatible with an equilibrium frequency being high. It is possible that as the allele frequency of the derived mutation reaches high frequencies in a population, 198 199 circulation of the viruses exploiting CDHR3 (past and present) are affected, with attendant 200 impacts on the selective pressures they impose on human populations. Such a frequency-201 dependent selection scenario seems more plausible than that of a heterozygote advantage given 202 that in Danish children with severe asthma, having even one copy of the risk variant was associated with increased risk of exacerbation and hospitalization¹⁰. If there were another virus 203 that exploited this receptor in past human populations, we might assume that heterozygotes 204 205 would similarly not be conferred protection.

206 In conclusion, our analyses combined *a priori* knowledge of a genetic variant underlying variable susceptibility to RV-C infections^{7,9,11} with population genetic analyses of whole genome 207 sequence data to investigate the evolutionary history of the locus in CDHR3. The conservation of 208 the protein, combined with its complex evolutionary history, exemplifies the biological 209 210 importance of CDHR3, which may or may not be ultimately relevant to its function as the cellular surface receptor for RV-C. We detected a worldwide signature of selection at this locus, 211 212 but also found that patterns of variation do not conform to the classic selective sweep model. 213 Instead, we posit the possibility of frequency-dependent balancing selection operating at this 214 locus which warrants more investigation into genotypic and biochemical effects of the variant (e.g. whether there is any phenotypic benefit to being heterozygous, trade-offs of different 215 genotypes, etc.). Irrespective of the mode of selection, our analyses show that the derived allele 216 217 has been advantageous and selected for in recent human evolution.

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219	Methods
220	Datasets. 1000 Genomes Project. Individual level phased sequencing data from the 1000
221	Genomes Project Phase 3 dataset were downloaded from
222	ftp://ftp.1000genomes.ebi.ac.uk/vol1/ftp/release/20130502/ ²³ .
223	
224	Ancient H. sapiens. BAM files for each of the 230 individuals included in the study by
225	Mathieson et. Al. ²⁰ were downloaded and converted to MPILEUP format using samtools ⁴⁵ . Low
226	coverage sequencing data at asthma susceptibility loci were manually inspected for each
227	individuals for which at least one read had mapped to the site. Manual diploid genotyping calls
228	were made for aDNA samples for which we felt confident making diploid genotype calls (Table
229	S1).
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231	Neandertal and Denisovan. Geontypes for the Vindija, Altai, and Denisovan genomes ⁴⁶
232	generated using snpAD, an ancient DNA damage-aware genotyper, were downloaded from
233	http://cdna.eva.mpg.de/neandertal/Vindija/VCF/.
234	
235	Great Apes. Genotypes of primates sequences were obtained from
236	https://eichlerlab.gs.washington.edu/greatape/data/ and converted to the corresponding human
237	regions with the LiftOver software ¹⁷ .
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239	Haplotype Networks. Indels and multi-allelic sites were filtered out with bcftools ⁴⁵ . Variants
240	having a Hardy-Weinberg equilibrium exact test p-value below 1×10^{-5} as calculated using the –
241	hwe midp function in PLINK1.9 ⁴⁷ in any of the 26 populations were removed from all
242	populations. The core haplotype surrounding rs6967330 was identified using biallelic markers
243	within 100kb of rs6967330 in Haploview ²⁴ from all 26 populations in the 1000 Genomes Phase 3
244	release. A large haplotype block was defined on Chromosome 7 from 105,657,078 to
245	105,680,022, and a smaller haplotype of Chromosome 7 from 105,657,078 to 105,659,873.
246	Haplotypes within the defined haplotype blocks were extracted from biallelic markers with a
247	minor allele frequency (MAF) > 0.01 with the Pegas package ^{25,26} . Haplotypes occurring at >1%
248	(at least 26 individuals) in the total 1000 Genomes Project dataset were constructed into

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networks. Genotypes from two high quality Neanderthal genomes and a Denisovan genome weresimilarly extracted and used in network analyses.

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252 Haplotype-Based Selection Scans. We computed iHS and nS_L using the method implemented in 253 Selink, a software to detect selection using whole-genome datasets (https://github.com/h-e-254 g/selink). As iHS and nS_I are sensitive to the inferred ancestral/derived state of an allele, we computed these statistics only when the derived state was determined unambiguously³². Results 255 were normalization by derived allele frequency (DAF) bins (from 0 to 1, increments of 256 $(0.025)^{27,32}$. We also minimized the false-positive discovery by excluding SNPs with a DAF 257 below 0.2, as the power to detect positive selection has been shown to be limited at such low 258 frequencies^{27,32}. For each statistic, we considered the percent rank at rs6967330 relative to the 259 genome-wide distribution in each population. 260

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In order to test selection shared among populations, we combined iHS and nS_L into a single 262 multi-population (MP) combined selection score, MP-iHS and MP- nS_L . The rationale behind 263 these composite approaches^{48–50} is that neutrality statistics, though expected to be correlated 264 among populations under neutrality, are more strongly correlated for positively selected variants 265 than for neutral variants. Indeed, under global positive selection, iHS and nS_L tend to become 266 negative in all populations while false positives will only be negative in a few populations. 267 268 Consequently, candidates genuinely selected in several populations should harbor extreme values for MP-iHS and MP- nS_L. For each SNP and each population, we determined the empirical p-269 value of iHS and nS_{I} , i.e. the rank of each statistic in the genome-wide distribution divided by 270 271 the number of SNPs. We then used the combine_pvalues function implementation of Fisher's method⁵¹ of the SciPy python package to compute MP-iHS and MP- nS_L for every SNP (passing 272 273 the criteria above to compute iHS and nS_L) found in at least two populations. 274 β Test for Long-Term Balancing Selection. β scores for each population in the 1000 Genomes 275

- 276 Project were obtained from <u>https://github.com/ksiewert/BetaScan</u>.
- 277
- Population Differentiation. Global F_{ST} was calculated with Selink (https://github.com/h-eg/selink).

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- 284

285 Author Contributions

- MBO, CSP, and ACP conceived the project. MBO, GL, and JCT performed the analyses. MBO
- drafted the paper and made the figures. All authors provided critical feedback, reviewed, and
- edited the manuscript.
- 289

290 **Competing interests**

291 The authors declare no competing interests.

References

- Barreiro, L. B. & Quintana-Murci, L. From evolutionary genetics to human immunology: how selection shapes host defence genes. *Nat Rev Genet* **11**, 17–30 (2010).
- Siddle, K. J. & Quintana-Murci, L. The Red Queen's long race: human adaptation to pathogen pressure. *Current Opinion in Genetics & Development* 29, 31–38 (2014).
- 3. Karlsson, E. K., Kwiatkowski, D. P. & Sabeti, P. C. Natural selection and infectious disease in human populations. *Nat Rev Genet* **15**, 379–393 (2014).
- Fumagalli, M. & Sironi, M. Human genome variability, natural selection and infectious diseases.
 Current Opinion in Immunology **30**, 9–16 (2014).
- 5. Quach, H. & Quintana-Murci, L. Living in an adaptive world: Genomic dissection of the genus Homo and its immune response. *Journal of Experimental Medicine* **214**, 877–894 (2017).
- Fumagalli, M. *et al.* Signatures of Environmental Genetic Adaptation Pinpoint Pathogens as the Main Selective Pressure through Human Evolution. *PLOS Genetics* 7, e1002355 (2011).
- 7. Bochkov, Y. A. *et al.* Cadherin-related family member 3, a childhood asthma susceptibility gene product, mediates rhinovirus C binding and replication. *PNAS* **112**, 5485–5490 (2015).
- Palmenberg, A. C. Rhinovirus C, Asthma, and Cell Surface Expression of Virus Receptor, CDHR3. J.
 Virol. JVI.00072-17 (2017). doi:10.1128/JVI.00072-17
- Bønnelykke, K. *et al.* Cadherin-related Family Member 3 Genetics and Rhinovirus C Respiratory Illnesses. *Am J Respir Crit Care Med* 197, 589–594 (2017).
- 10. Bønnelykke, K. *et al.* A genome-wide association study identifies CDHR3 as a susceptibility locus for early childhood asthma with severe exacerbations. *Nat Genet* **46**, 51–55 (2014).
- 11. Scully, E. J. *et al.* Lethal Respiratory Disease Associated with Human Rhinovirus C in Wild Chimpanzees, Uganda, 2013. *Emerg Infect Dis* **24**, 267–274 (2018).
- 12. Gern, J. E. The ABCs of Rhinoviruses, Wheezing, and Asthma. J. Virol. 84, 7418–7426 (2010).

- 13. Bryce, J., Boschi-Pinto, C., Shibuya, K. & Black, R. E. WHO estimates of the causes of death in children. *The Lancet* **365**, 1147–1152 (2005).
- 14. Busse, W. W., Lemanske, R. F. & Gern, J. E. The Role of Viral Respiratory Infections in Asthma and Asthma Exacerbations. *Lancet* **376**, 826–834 (2010).
- 15. Ferkol, T. & Schraufnagel, D. The Global Burden of Respiratory Disease. *Annals ATS* **11**, 404–406 (2014).
- 16. Karolchik, D. *et al.* The UCSC Table Browser data retrieval tool. *Nucleic Acids Res.* 32, D493-496 (2004).
- Rosenbloom, K. R. *et al.* The UCSC Genome Browser database: 2015 update. *Nucleic Acids Res* 43, D670–D681 (2015).
- Cooper, G. M. *et al.* Distribution and intensity of constraint in mammalian genomic sequence.
 Genome Res. 15, 901–913 (2005).
- Prado-Martinez, J. *et al.* Great ape genetic diversity and population history. *Nature* 499, 471–475 (2013).
- 20. Mathieson, I. *et al.* Genome-wide patterns of selection in 230 ancient Eurasians. *Nature* advance online publication, (2015).
- Lazaridis, I. *et al.* Ancient human genomes suggest three ancestral populations for present-day Europeans. *Nature* 513, 409–413 (2014).
- Fu, Q. *et al.* The genome sequence of a 45,000-year-old modern human from western Siberia.
 Nature 514, 445–449 (2014).
- 23. The 1000 Genomes Project Consortium. A global reference for human genetic variation. *Nature* 526, 68–74 (2015).
- Barrett, J. C., Fry, B., Maller, J. & Daly, M. J. Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* 21, 263–265 (2005).

- Paradis, E. pegas: an R package for population genetics with an integrated–modular approach.
 Bioinformatics 26, 419–420 (2010).
- 26. R Development Core Team. *R: A Language and Environment for Statistical Computing*. (R Foundation for Statistical Computing).
- 27. Voight, B. F., Kudaravalli, S., Wen, X. & Pritchard, J. K. A Map of Recent Positive Selection in the Human Genome. *PLoS Biol* **4**, e72 (2006).
- Ferrer-Admetlla, A., Liang, M., Korneliussen, T. & Nielsen, R. On detecting incomplete soft or hard selective sweeps using haplotype structure. *Mol Biol Evol* msu077 (2014). doi:10.1093/molbev/msu077
- 29. Sabeti, P. C. *et al.* Detecting recent positive selection in the human genome from haplotype structure. *Nature* **419**, 832–837 (2002).
- Ferrari, S. L., Ahn-Luong, L., Garnero, P., Humphries, S. E. & Greenspan, S. L. Two Promoter Polymorphisms Regulating Interleukin-6 Gene Expression Are Associated with Circulating Levels of C-Reactive Protein and Markers of Bone Resorption in Postmenopausal Women. *J Clin Endocrinol Metab* 88, 255–259 (2003).
- 31. Kong, A. *et al.* Fine-scale recombination rate differences between sexes, populations and individuals. *Nature* **467**, 1099–1103 (2010).
- 32. Fagny, M. *et al.* Exploring the Occurrence of Classic Selective Sweeps in Humans Using Whole-Genome Sequencing Data Sets. *Mol Biol Evol* **31**, 1850–1868 (2014).
- 33. Arenzana-Seisdedos, F. & Parmentier, M. Genetics of resistance to HIV infection: Role of coreceptors and co-receptor ligands. *Seminars in Immunology* **18**, 387–403 (2006).
- 34. Stephan, W. Signatures of positive selection: from selective sweeps at individual loci to subtle allele frequency changes in polygenic adaptation. *Molecular Ecology* **25**, 79–88 (2016).

- 35. Key, F. M., Teixeira, J. C., de Filippo, C. & Andrés, A. M. Advantageous diversity maintained by balancing selection in humans. *Current Opinion in Genetics & Development* **29**, 45–51 (2014).
- Allison, A. C. Protection Afforded by Sickle-cell Trait Against Subtertian Malarial Infection. *Br Med J* 1, 290–294 (1954).
- Shriner, D. & Rotimi, C. N. Whole-Genome-Sequence-Based Haplotypes Reveal Single Origin of the Sickle Allele during the Holocene Wet Phase. *The American Journal of Human Genetics* **102**, 547–556 (2018).
- Leffler, E. M. *et al.* Multiple Instances of Ancient Balancing Selection Shared Between Humans and Chimpanzees. *Science* 339, 1578–1582 (2013).
- Ségurel, L. *et al.* The ABO blood group is a trans-species polymorphism in primates. *PNAS* **109**, 18493–18498 (2012).
- 40. Ferrer-Admetlla, A. *et al.* Balancing Selection Is the Main Force Shaping the Evolution of Innate Immunity Genes. *The Journal of Immunology* **181**, 1315–1322 (2008).
- 41. Andrés, A. M. *et al.* Targets of Balancing Selection in the Human Genome. *Mol Biol Evol* **26**, 2755–2764 (2009).
- 42. Wiuf, C., Zhao, K., Innan, H. & Nordborg, M. The Probability and Chromosomal Extent of *trans*specific Polymorphism. *Genetics* **168**, 2363–2372 (2004).
- 43. Teixeira, J. C. *et al.* Long-Term Balancing Selection in LAD1 Maintains a Missense Trans-Species Polymorphism in Humans, Chimpanzees, and Bonobos. *Mol Biol Evol* **32**, 1186–1196 (2015).
- 44. Siewert, K. M. & Voight, B. F. Detecting Long-Term Balancing Selection Using Allele Frequency Correlation. *Mol Biol Evol* **34**, 2996–3005 (2017).
- 45. Li, H. *et al.* The Sequence Alignment/Map format and SAMtools. *Bioinformatics* **25**, 2078–2079 (2009).

- 46. Prüfer, K. *et al.* A high-coverage Neandertal genome from Vindija Cave in Croatia. *Science* eaao1887
 (2017). doi:10.1126/science.aao1887
- 47. Purcell, S. *et al.* PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* **81**, 559–75 (2007).
- Deschamps, M. *et al.* Genomic Signatures of Selective Pressures and Introgression from Archaic Hominins at Human Innate Immunity Genes. *Am J Hum Genet* **98**, 5–21 (2016).
- Grossman, S. R. *et al.* A Composite of Multiple Signals Distinguishes Causal Variants in Regions of Positive Selection. *Science* 327, 883–886 (2010).
- Grossman, S. R. *et al.* Identifying Recent Adaptations in Large-Scale Genomic Data. *Cell* 152, 703–713 (2013).
- 51. Fisher, R. A. Statistical Methods for Research Workers. in *Breakthroughs in Statistics* 66–70 (Springer, New York, NY, 1992). doi:10.1007/978-1-4612-4380-9_6
- 52. Marcus, J. H. & Novembre, J. Visualizing the geography of genetic variants. *Bioinformatics* **33**, 594–595 (2017).

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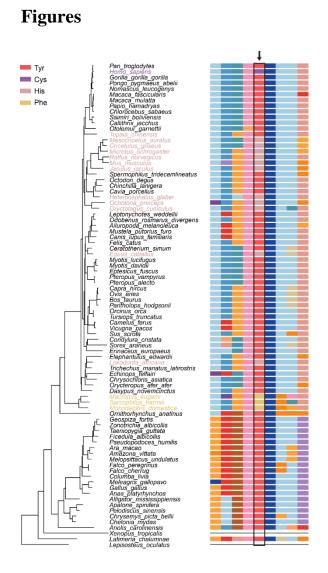


Figure 1. CDHR3 protein. (Left) Species tree for the multi sequence alignment of 85 species in the UCSC multiz alignment. (Right) Multi sequence protein alignment surrounding position 529 of the human CDHR3 protein. Residue 529 is outlined in black and designated with an arrow. Humans carry the Tyr and Cys alleles. Data obtained from the UCSC Genome Browser^{16,17}.

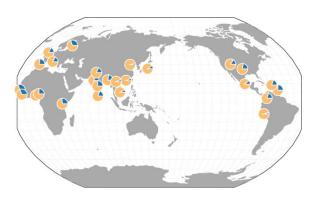


Figure 2. Global distribution of allele frequencies at rs6967330. Pie charts represent the relative allele frequencies of the ancestral A allele (blue) and the derived G allele (yellow) in each of the 26 populations of the 1000 Genomes Project²³. This image was generated through the *Geography of Genetic Variants* browser⁵².

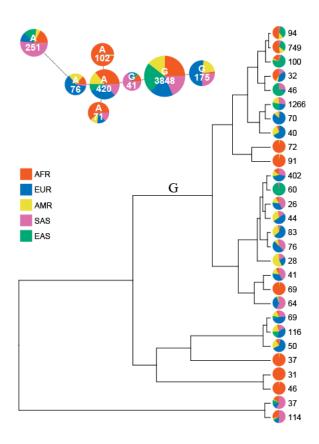


Figure 3. Haplotype structure of anatomically modern humans from the 1000 Genomes Project. (Top) Haplotype network from chromosome 7 between 105,657,078 and 105,659,873. Haplotype network was constructed from phased genome sequences of 2504 individuals with variation at 16 sites in the 2,795bp region. Only haplotypes occurring at > 1% are shown. Colors reflect super population designation of individuals. (Bottom) Unrooted tree from chromosome 7 between 105,657,078 and 105,680,022. Haplotypes of the same 2504 individuals were derived for a larger haplotype block. Again, only haplotypes occurring at > 1% are shown and colors reflect super population designation of individuals. The tree is based on 83 SNPs (68 informative) from the 22,945bp region.