# Candidate genes-based investigation of susceptibility to Human African Trypanosomiasis in Côte d'Ivoire.

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#### 27 Abstract

28 Human African Trypanosomiasis (HAT) or sleeping sickness is a Neglected Tropical Disease. Long regarded as an invariably fatal disease, there is increasing evidence that infection by T. 29 b. gambiense can result in a wide range of clinical outcomes, including latent infections, 30 which are long lasting infections with no parasites detectable by microscopy. The 31 determinants of this clinical diversity are not well understood but could be due in part to 32 parasite or host genetic diversity in multiple genes, or their interactions. A candidate gene 33 association study was conducted in Côte d'Ivoire using a case-control design which included 34 a total of 233 subjects (100 active HAT cases, 100 controls and 33 latent infections). All three 35 possible pairwise comparisons between the three phenotypes were tested using 96 SNPs in16 36 candidate genes (IL1, IL4, IL4R, IL6, IL8, IL10, IL12, IL12R, TNFA, INFG, MIF, APOL1, 37 HPR, CFH, HLA-A and HLA-G). Data from 77 SNPs passed quality control. There were 38 39 suggestive associations at three loci in *IL6* and *TNFA* in the comparison between active cases and controls, one SNP in each of APOL1, MIF and IL6 in the comparison between latent 40 41 infections and active cases and seven SNP in IL4, HLA-G and TNFA between latent infections and controls. No associations remained significant after Bonferroni correction, but the 42 Benjamini Hochberg false discovery rate test indicated that there were strong probabilities 43 that at least some of the associations were genuine. 44

The excess of associations with latent infections despite the small number of samples available suggests that these subjects form a distinct genetic cluster different from active HAT cases and controls, although no clustering by phenotype was observed by principle component analysis. This underlines the complexity of the interactions existing between host genetic polymorphisms and parasite diversity.

50 Keywords: Single nucleotide polymorphism; Candidate genes; Association study; Human
51 African trypanosomiasis; Sleeping sickness, Cote d'Ivoire

#### 52 Author summary

53 Since it was first identified, human African trypanosomiasis (HAT) or sleeping sickness has been described as invariably fatal. Recent data however suggest that infection by T. b. 54 gambiense can result in a wide range of clinical outcomes in its human host including long 55 lasting infections, that can be detected by the presence of antibodies, but in which parasites 56 cannot be seen by microscopy; these cases are known as latent infections. While the factors 57 determining this varied response have not been clearly characterized, the effectors of the 58 immune responses have been partially implicated as key players. We collected samples from 59 people with active HAT, latent infections and controls in endemic foci in the Côte d'Ivoire. 60 We tested the role of single nucleotide polymorphisms (SNPs) in 16 genes on 61 susceptibility/resistance to HAT by means of a candidate gene association study. There was 62 some evidence that variants of the genes for IL4, IL6, APOL1, HLAG, MIF and TNFA 63 64 modified the risk of developing HAT. These proteins regulate the inflammatory response to many infections or are directly involved in killing the parasites. In this study, the results were 65 statistically weak and would be inconclusive on their own, however other studies have also 66 found associations in these genes, increasing the chance that the variants that we have 67 identified play a genuine role in the response to trypanosome infection in Côte D'Ivoire. 68

#### 69 Introduction

Sleeping sickness, or human African Trypanosomiasis (HAT), is caused by *Trypanosoma brucei gambiense* and *T.b. rhodesiense* and is transmitted by tsetse flies (*Glossina spp*). *T. b. gambiense* is associated with a more chronic disease that can take decades to become patent, and *T.b. rhodesiense* causes an acute disease within months of infection. The chronic form of the disease caused by the *T. b. gambiense* is classically characterized by an early hemolymphatic stage (stage 1) associated with non-specific symptoms such as intermittent fevers and headaches, followed by a meningo-encephalitic stage (stage 2) in which the

parasite invades the central nervous system and causes neurological disorders and death if left 77 78 untreated. This chronic form is found in Western and Central Africa while the acute form caused by T. b. rhodesiense is endemic to Eastern Africa [1,2]. However, recent observations 79 are increasingly indicating that infection by T. b. gambiense can result in a wide range of 80 clinical outcomes in its human host [3,4]. Self-cure processes have been described and 81 reviewed in Checchi et al. [3]. Furthermore, some authors argued that HAT is not invariably 82 fatal [5], supporting observations made by Garcia et al. [4] who followed individuals who 83 remained seropositive for HAT but without detectable parasites for two years. This clinical 84 diversity is not well understood but could be due to parasite genetic diversity [6,7], human 85 immune gene variability [8] or their interaction [9]. It has been suggested that genetic 86 polymorphisms of the parasite could be associated with asymptomatic and very chronic 87 infections [10]. Nevertheless, genes involved in the host immune response have been 88 89 implicated in the control of infection or susceptibility to HAT [11,12] and also T. congolense infections in experimental models [13,14]. Among the genes implicated in the pathogenesis of 90 91 the disease, are IL1, IL4, IL6, IL8, IL10, IL12, IFNG, TNFA, CFH and MIF [8,11,15-17].

Some studies have found associations between certain polymorphisms in genes encoding 92 cytokines. For example, IL4 plays a role in susceptibility to T. brucei infection [18], while 93 94 polymorphisms in the IL6 and IL10 genes have been associated with a decreased risk of developing HAT [11,15]. On the other hand, polymorphisms in TNFA genes have been 95 associated with an increased risk of developing the disease [11,15]. Furthermore, MacLean et 96 97 al. [19] reported in their study that plasma levels of IFNG significantly decrease during the late stage of the T.b. rhodesiense disease [19]. Human blood contains trypanolytic factors 98 (TLF1 and TLF2) that are lytic to almost all African trypanosomes except T. b. rhodesiense 99 and T. b. gambiense [20]. Human TLF1 contains two primate-specific proteins, 100 101 apolipoprotein L1 (APOL1) and haptoglobin-related protein (HPR). APOL1 expression is

induced by *T. b. gambiense* infection but expression is not associated with susceptibility to
 sleeping sickness [21]. Macrophage migration inhibitory factor (MIF) contributes to
 inflammation-associated pathology in the chronic phase of *T. brucei* infection [16].

105 Although genes directly involved in the immune response, like genes encoding cytokines, are 106 very important candidates, genes implicated in the regulation of immunity also have a critical 107 role. Thus, Courtin et *al.* [12] have shown a genetic association between *HLA-G* 108 polymorphisms and susceptibility to HAT.

There is now cumulative evidence that polymorphisms in genes involved in the control of the
immune response and genes implicated in the regulation of immunity could play a role in
HAT infection outcome [11,12].

112 We report a candidate gene association study of the role of single nucleotide polymorphisms

113 (SNPs) in IL1, IL4, IL4R, IL6, IL8, IL10, IL12, IL12R, TNFA, INFG, MIF, HPR, CFH,

114 *APOL1, HLA-A*, and *HLA-G* genes on susceptibility/resistance to HAT.

115

## 116 **Results**

# 117 Characteristics of the population

A total of 100 cases (or former cases), 100 controls and 33 latent infections were enrolled into
the study (Table 1). The mean age (range) of the study population was 38.8 (6–84) years. The
sex ratio (male: female) was 0.88 (109/124).

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## 124 Table1: Population characteristics

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					127
	Cases	Controls	Latent infections	Whole popu	ul <b>atie</b> n
	(n=100)	(n=100)	(n= 33)	(n=233)	129
Age mean	41,8	36,8	36	38,8	130
	50	10	1.4	100	131
Males	53	42	14	109	132
Females	47	58	19	124	133
	65	0	10	0.2	134
IL+	65	0	18	83	135
TL-	35	100	15	150	136
					137

**Table 1:** Population characteristics. Cases: CATT+ve, microscopy +ve; Controls: CATT-ve and microscopy –ve. Trypanolysis negative (TL-ve) cases were from subjects who were retrospectively sampled for the purpose of this study and who had not been subject to the trypanolysis test when originally identified as HAT cases by CATT and microscopy. Latent infection: subjects who were CATT positive (CATTpl  $\geq 1/4$ ) but parasites could not be detected by microscopy at repeated tests for at least two years.

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# 145 Association study

Three association analyses were run on the three possible pairwise combinations of the controls, latent infections and active HAT cases. After filtering out SNP loci that were not in Hardy Weinberg equilibrium (P <0.001), or had > 10% missing genotypes or had minor allele frequencies of zero, 77 SNP loci remained in the latent infection vs active case comparison and 74 SNP loci remained in the other two comparisons. One individual was filtered out of the case sample and one out of the latent infection sample because of low genotyping rates (<90%).

F<sub>ST</sub> between ethnic groups showed that allele frequency differences between the ethnic groups were small, indicating that ethnicity was unlikely to confound results (median Fst - 155 0.00011763, Fst maximum 0.015). Scatter plots of the first two principal components between 156 the different phenotypes (cases, latent infections and controls) and different ethnic groups 157 show that this population is homogeneous and that the samples did not cluster by phenotype 158 or by ethnic groups (Fig 1) and consequently the data were not stratified by ethnicity.

Fig 1. Multidimensional scaling (ms) plots of the genotype data by ethnicity (upper plot) and
phenotype (lower plot). The plots show no evidence of clustering either by ethnicity or
phenotype.

There were suggestive associations at three loci in *IL6* and *TNFA* in the comparison between 162 active cases and controls (Table 2), three loci in APOL1, MIF and IL6 in the comparison 163 between latent infections and active cases (Table 3) and five loci in *IL4* (Fig 2), and one each 164 in HLA-G and TNFA between latent infections and controls (Table 4). After Bonferroni 165 166 correction, none of these associations remained significant, however Bonferroni correction is very conservative, particularly since there was some linkage between adjacent marker SNP in 167 some genes. The Benjamini-Hochberg false discovery rate (FDR) test, which shows the 168 probability that an observation is a false positive, indicated that at least some of the 169 suggestively positive samples may be true positives. Under the FDR the rs62449495 SNP in 170 IL6 had an 80% chance of being a true positive and the TNFA-308 rs1800629 SNP had a 71% 171 chance of being a true positive (Table 2). The strongest association was with APOL1 172 rs73885319, which is also known as the G1 allele, which had a 90% chance of being a true 173 positive in the comparison between latent infections and controls (Table 3). Complete results 174 175 for all tests that passed quality control are shown in supplementary data tables S1, S2 and S3.

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Fig 2. Positions of SNP genotyped within IL4. SNP with suggestive associations (uncorrected
 p <0.05) are indicated with a star.</li>

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186 Co	185 5 5	184 Tal	CHI 22 22 22 7	182 Tal	7 181	CHF 7	180 Tal
ons= Consequence from Ensen ntrols minor allele frequency o rrected p value, FDR_BH = Fals pothesis is that allele frequenc	132,009,154 132,010,726 132,016,554 29,799,116 132,009,710 31,543,031 132,014,109	<b>ble 4</b> : Associatic 8 BP	۲ BP 36,661,906 24,235,455 22,764,338	ble 3: Associatic	22,767,137	۶ BP 22,764,338 31,543,031	ble 2: Associatic
	rs2243250 rs734244 rs2243282 rs1611139 rs2070874 rs1800629 rs2243270	on tests betwe SNP	SNP rs73885319 rs36086171 rs62449495	on tests betwo	rs2069830	SNP 3 rs62449495 . rs1800629	on tests betwe
nbl Var of cont	IL4 IL4 HLAG IL4 TNFA IL4	en late Gene	Gene APOL1 MIF IL6	en late	IL6	Gene IL6 TNFA	en cas
iant predi rols; OR = overv Rate	5' Intronic 1' 5' UTR 5' Intronic	nt infectio Cons	Cons Missens e 5 ' 5'	nt infection	Missens e	Cons 5'	es (n=99)
lictor, Min A = minor allele, = odds ratio, P = p-value, [L9 te Benjamini-Hochberg (It is 1 controls are the same), OR	$\circ$ $\land$	ons (n= Min A	Min G A G A	ons (n=	-	A Min A	and co
	0.42 0.25 0.26 0.34 0.08 0.36	32) and Latent	Cases 0.20 0.29 0.04	32) and	0.11	Cases 0.04 0.09	ntrols (n
	0.27 0.41 0.14 0.14 0.50 0.19 0.23	controls (n: Controls	Latent 0.41 0.13 0.13	active HAT	0.06	Controls 0.13 0.19	=100)
Maj A = 95-U95] s probat	<b>໑໑</b> ݦ໑ݦݦ⊣	=100) Maj A	Maj A J	cases (	C	G G A	
. = major allele, Latent minor allele frequ 5] = confidence interval of odds ratio, Bc ability of falsely rejecting the null hypoth ds ratio, [L95-U95] = confidence interval	0.0159 0.0215 0.0347 0.0354 0.0354 0.0372 0.0396 0.0415	P	P 0.0012 0.0062 0.0217	n=99)	0.0369	P 0.0025 0.0071	
	1.00 1.00 1.00 1.00 1.00 1.00	Bonf_Corr	Bonf_Corr 0.10 0.52 1.00		1.00	Bonf_Corr 0.20 0.58	
	0.48 0.48 0.48 0.48 0.48 0.48	FDR_BH	FDR_BH 0.10 0.26 0.61		0.81	FDR_BH 0.20 0.29	
	2.02 0.49 0.53 2.13 0.53 0.37 1.88	OR	OR 0.37 2.90 0.29		2.17	OR 0.30 0.44	
iency o onf_Cor iesis. Tl	1.13 0.26 0.29 1.04 0.30 0.14 1.02	261	L95 0.20 1.30 0.10		1.02	L95 0.13 0.24	
f Laten r= Bon ıe null	3.64 0.92 0.96 4.36 0.96 0.99 3.45	U95	U95 0.68 6.46 0.80		4.61	U95 0.68 0.80	
t cases; ferroni	0.80 0.01 1.00 0.39 0.69 0.51 1.00	HWE_p	HWE_p 1.00 0.05 1.00		1.00	HWE_p 1.00 0.51	

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#### 191 **DISCUSSION**

In this study, we investigated the role of single nucleotide polymorphisms on susceptibility/resistance to HAT. We genotyped 96 SNPs within sixteen genes that were investigated to test genetic association with HAT in Côte d'Ivoire. For consistency, all samples were commercially genotyped using two platforms: Genome Transcriptome de Bordeaux, France and LGC Genomics UK.

Our results suggest that three SNPs were associated with the development of HAT by controls, another three were associated with the development of active HAT by people with latent infections, and seven SNP were associated with the risk of developing latent infections five of them in *IL4* although none of these results remained significant after Bonferroni correction. Only two of the SNP with suggestive associations were coding and missense (II6 rs2069830 and APOL1 rs73885319) since, with some exceptions, SNP were primarily selected as tags that might be linked to functional SNP rather than for any putative function.

The most striking feature of the results is that most associations were found in the 204 comparisons with latent infections, despite limited power, with only 33 of these samples 205 206 being available compared with 100 each for the cases and controls. This illustrates the increased power to be gained from using well defined phenotypes. Given the evidence that 207 some people in West Africa can self-cure their T. b. gambiense infections [5] it is likely that 208 209 some of the control subjects are resistant to infection or have recovered from infection, whilst others are naïve and susceptible. These two groups may have very different genetic 210 backgrounds which could confound the association studies. The data also suggest that the 211 latent infections may represent a genetically distinctive group. Although the principle 212 component analysis did not identify any cluster associated with latent infections (Fig 1) 213 further studies of their genetic and immunological profiles are required to test this hypothesis. 214

215 Associations with *IL6* 

The minor (A) allele of IL6 rs62449495 appeared to protect against progression from latent 216 infections to active HAT (Table 3) and against the development of active HAT by controls 217 but these associations did not remain significant after Bonferroni correction (Table 2). The 218 major allele of rs2069830 was also protective against the development of active HAT by 219 controls before but not after Bonferroni correction (Table 2). The rs2069830 SNP causes a 220 proline to serine change, but this is predicted to be benign by both Sift and Polyphen 221 according to the Ensembl Variant Effect predictor. IL6 plays a key role in the acute 222 223 inflammatory response and in regulation of the production of acute phase proteins such as Creactive protein [22]. It contributes to the inflammatory response, regulates haematopoiesis, 224 which may contribute to the anaemia associated with HAT, and modifies the permeability of 225 the blood brain barrier, which may contribute to the development of the stage 2 invasion of 226 the CNS [22,23]. The results of our association study are consistent with the involvement of 227 228 IL6 variants in the susceptibility to the disease as Courtin et al [15] have reported before. 229 However, it should be noted that the polymorphisms of IL6 in our study (rs62449495 and 230 rs2069830) are different of those found by Courtin *et al.* [15] with  $IL6_{4339}$  = rs2069849. They showed that in DRC, IL64339 SNP was significantly associated with a decreased risk of 231 developing the disease with a P-value = 0.0006 before the Bonferroni correction (0.04 after 232 Bonferroni Correction). 233

# 234 Association with TNFA

Our results also indicate that subjects carrying the A allele of *TNFA* (rs1800629) A/G had a lower risk of developing active HAT (Table 2) or a latent infection (Table 4), suggesting the possibility of a protective effect. This SNP is also known as the *TNF*-308 SNP and the minor A allele is associated with higher plasma levels of *TNFA* [24]. In a previous study in Côte d'Ivoire, Courtin et *al*. [11] showed that the distribution of the *TNFA*-308G/A polymorphism did not differ significantly between cases and controls in the total population, but found that,

under a recessive model the AA genotype was associated with risk of HAT in the 39 cases 241 and 57 controls who had been living in the endemic area of Sinfra for less than 10 years 242 (before Bonferroni correction). In contrast, in our study, all participants had lived in the 243 244 endemic area all their lives and the association was additive rather than recessive, furthermore the MAF in Courtin's studies was 24%, whilst in ours it was 14%, suggesting significant 245 differences in population structure between the two studies. Varying results in TNFA 246 associations studies are not uncommon, multiple candidate gene association studies of TNFA-247 308 (rs1800629) and malaria have found inconsistent results in different populations [25,26]. 248 Another explanation could be that the TNFA-308 has no effect but is in linkage disequilibrium 249 (LD) with another unidentified polymorphism. Varying LD across populations might lead to 250 different findings [27]. Despite the conflicting results from association study, animal models 251 indicate that *TNFA* is likely to be a key mediator in the control of *T. brucei* infections [28] 252 253 and a direct dose dependent lytic effect of TNFA on purified T.b. gambiense parasites has been reported suggesting an involvement in parasite growth control [29,30]. Given the 254 255 experimental evidence for a role for TNFA, it is possible that inconsistent results of association studies are a consequence of different TNFA alleles only making a small 256 difference to infection outcome and that therefore larger studies would be needed to detect 257 258 associations and also to heterogeneity in regulation of TNFA between populations.

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## 260 Association with IL4

Five of the seven SNP that were suggestively associated with the comparison between latent infections and controls were in *IL4* (Table 4, Fig 2). Three of these SNP were adjacent to each other at the 5' end of the gene (Fig 2), although linkage between them was modest ( $r^2 < 0.5$ ). While the individual associations did not remain significant after Bonferroni correction the observation that five of the sixteen SNP tested in *IL4* had suggestive associations increases the probability of a genuine association with *IL4* and the risk of developing a latent infection.

A study using three *IL4* SNP in DRC did not find any associations, but that was in a comparison between cases and controls [15]. In B10.Q mice deletion in *IL4* lead to increased *T. brucei* parasitaemia levels but longer survival time [18]. The relationship between *IL4* polymorphisms and acute HAT infection requires further study.

## 271 Association with APOL1

The suggestive association with APOL1 G1 allele rs73885319 and protection against 272 273 progression from latent infection to active HAT is consistent with the association found in Guinea [31,32], although in those studies this SNP was also associated with increased risk of 274 controls developing a latent infection as well. Moreover, APOL1 expression is induced by T. 275 b. gambiense infection but not associated with differential susceptibility to sleeping sickness 276 [21]. APOL1 rs73885319 is also known as the G1 allele of APOL1 and is associated with 277 278 kidney disease in African Americans and the relatively high frequency of this deleterious 279 allele was assumed to be due to selection by HAT [33]. The data presented here is consistent 280 with that hypothesis, but no support was found for the role of the G2 allele of APOL1 (rs71785313) in HAT although it is implicated in kidney disease. 281

## 282 Association with MIF

Although, our data show that subjects carrying the G allele of *MIF* (rs36086171) G/A had a risk of developing active HAT (table 3), we did not find a significant difference after correction (BONF = 0.52). Macrophage inhibitory factor (MIF) is a ubiquitously expressed protein that has proinflammatory, hormonal and enzymatic activities [34]. It is implicated in many inflammatory diseases [35]. It functions by recruiting myeloid cells to the site of inflammation [36], by inducing their differentiation towards M1 cells secreting TNF [37] and by suppressing p53-dependent apoptosis of inflammatory cells. While there are no human studies directly linking *MIF* to HAT, murine studies show that *MIF* plays a role as a mediator
of the inflammation which is a key feature in trypanosomiasis-associated pathology [16,38].

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## 293 Association with *HLA-G*

HLA-G molecule plays an important role on immune response regulation and has been 294 associated with the risk of human immunodeficiency virus (HIV) infection [39], human 295 papilloma virus [40] and herpes simplex virus type 1 [41]. The results of our association study 296 297 are consistent with the involvement of HLA-G genetic variants in the susceptibility to the disease [12]. The rs1611139 T allele of the HLA-G gene showed a suggestive association with 298 increased risk of controls a latent infection. This result could be due to differences in selection 299 pressures possibly driven by variability in the immuno-pathology of the diseases. It is known 300 that cytokines such as IL-10 can induce HLA-G expression by affecting mRNA transcripts 301 302 and protein synthesis by human monocytes and trophoblasts, thus having a significant impact on parasitic infections [42]. 303

## 304 Conclusion

The results discussed in this paper should be used with caution as no loci remained significantly associated with HAT after the Bonferroni correction for multiple testing, although some had high probabilities of being associated using a false discovery rate test. Some of the SNP loci identified here were also significant in other studies. Multiple independent observations of marginally significant effects suggest that these effects may be genuine but that the effect size is not large enough to be detected by the numbers available to be tested.

Our data support the findings from Guinea about the role of the *APOL1* G1 allele and also suggest that the polymorphisms of *IL4*, *IL6*, *HLA-G* and *TNFA* present interesting candidates for the investigation of the genetic susceptibility / resistance to HAT. The large number of suggestive associations with latent infections despite the small number of samples indicate the people with latent infections form a genetically distinctive group that merit further investigation.

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#### 319 Material and Methods

## 320 Population study, definition of phenotypes and study design

The study took place in western-central Côte d'Ivoire in the main HAT foci of Bonon, 321 Bouafle, Zoukougbeu, Oume, and Sinfra. Samples were collected in three stages, (i) 322 previously archived samples; existing collections of previously archived samples were 323 centralized to CIRDES (Centre International de Recherche-Développement sur l'Elevage en 324 zone Subhumide) Burkina-Faso. (ii) retrospectively collected samples; study sites were 325 revisited to consent and resample previously diagnosed and treated patients and (iii) 326 327 prospectively collected samples including new HAT patients. Three phenotypes were considered: (i) cases, defined as individuals in whom the presence of trypanosomes was 328 329 confirmed by microscopy; (ii) controls, defined as individuals living in the endemic area who are card agglutination test for trypanosomiasis (CATT) negative and trypanolysis test (TL) 330 negative, and without evidence of previous HAT infection and (iii) latent infections, defined 331 as subjects who were CATT positive (CATTpl  $\geq 1/4$ ) but in whom parasites could not be 332 detected by microscopy at repeated tests for at least two years. 333

A total of 233 individuals with 100 former HAT cases, 100 controls and 33 latent infections were included in the study. Samples were frozen directly in the field at -20°C and kept at that temperature until use. For each individual, an aliquot of plasma was used to perform the immune trypanolysis test that detects Litat 1.3 and Litat 1.5 variable surface antigens specific for *T.b. gambiense* [43]. All control individuals (n=100) included in this study were negative on the trypanolysis test (TL-ve) (Table 1). This study was one of five studies of populations of HAT endemic areas in Cameroon, Côte d'Ivoire, Guinea, Malawi and Uganda by the TrypanoGEN consortium [44]. The studies were designed to have 80% power to detect odds ratios (OR) >2 for loci with disease allele frequencies of 0.15 - 0.65 and 100 cases and 100 controls with the 96 SNPs genotyped. Power calculations were undertaken using the genetics analysis package gap in R [45].

# 345 Selection of SNPs and genotyping

Genomic DNA was obtained from peripheral blood samples. Extraction was performed using the Qiagen DNA extraction kit (QIAamp DNA Blood Midi Kit) according to the manufacturer's instructions and quantified by Nanodrop assay. DNA was stored at -20 °C until analysis.

Ninety-six SNPs were genotyped in IL1, IL4, IL4R, IL6, IL8, IL10, IL12, IL12R, TNFA, 350 INFG, MIF, HPR, CFH, APOL1, HLA-A, and HLA-G. The SNPs were selected by two 351 strategies: 1) SNP in IL4, IL6, IL8, HLAG and IFNG were designed as markers for linkage 352 353 disequilibrium (LD) scans of each gene [46], 2) SNPs in other genes were selected based on reports in the literature that they were associated with trypanosomiasis or other infectious 354 diseases. The LD scans were designed using 1000 Genomes Project data [47], merged with 355 low fold coverage (8-10x) whole genome shotgun data generated from 230 residents living in 356 regions (DRC, Guinea Conakry, Côte D'Ivoire and Uganda) where trypanosomiasis is 357 endemic (TrypanoGEN consortium, European Nucleotide Archive 358 study EGAS00001002482). Loci with minor allele frequency < 5% in the reference data were 359 excluded and an  $r^2$  of 0.5 was used to select SNP in linkage disequilibrium. 360

DNA was genotyped by two commercial service providers: INRA- Site de Pierroton, Plateforme Genome Transcriptome de Bordeaux, France and ii- LGC genomics Hoddesden UK. At INRA multiplex design (two sets of 40 SNPs) was performed using Assay Design Suite v2.0 (Agena Biosciences). SNP genotyping was achieved with the iPLEX Gold genotyping kit (Agena Biosciences) for the Mass Array iPLEX genotyping assay, following
the manufacturer's instructions. Products were detected on a Mass Array mass
spectrophotometer and data were acquired in real time with Mass Array RT software (Agena
Biosciences). SNP clustering and validation was carried out with Typer 4.0 software (Agena
Biosciences). At LGC Genomics SNP were genotyped using the PCR based KASP assay[48].

#### 370 Statistical analysis

All statistical analyses were performed using the Plink 1.9 and R v 3.2.1 software. Individuals 371 were excluded who had > 10% missing SNP data. SNP loci were excluded that > 10%372 missing genotypes or if the control samples were not in Hardy-Weinberg equilibrium 373 (p<0.001). Case-control association analysis using SNP alleles/genotypes was undertaken 374 using Fisher's exact test. The difference between the ethnic groups was estimated using the 375 376 F<sub>ST</sub>, with values potentially varying from zero (no population differentiation) to one (complete differentiation) [47]. Ethnicity was not used as a risk factor for HAT because we 377 observed no significant differences in  $F_{ST}$  between ethnic groups ( $F_{ST}$  maximum 0.015) or 378 379 clustering by linguistic group by multidimensional scaling in Plink. Some studies have used ethnicity and age as risk factors for HAT but found no significant association [11,15]. 77 SNP 380 remained in at least one comparison after filtering. P values were adjusted for multiple testing 381 using the Bonferroni corrections and Benjamini Hocheberg false discovery rate test as 382 implemented in Plink. 383

#### 384 Ethics Statement

The population of the study was informed. All adult subjects provided written informed consent. For children, a parent or guardian of any child participant provided written informed consent on their behalf. The protocol of the study was approved by the traditional authorities (chief and village committee) and by National ethics committee hosted by the Public Health Ministry of Côte d'Ivoire with the number: N°38/MSLS/CNERm-dkn 5th May 2014. This study is part of the TrypanoGen project which aims to a better understanding of genetic determinism of human susceptibility to HAT and the TrypanoGen-CI samples were archived in the TrypanoGen Biobank hosted by CIRDES in Bobo-Dioulasso, Burkina Faso [44].

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Acknowledgements: We thank subjects who generously donated their specimens and the
field workers from the Ivorian HAT foci for their dedication in collecting and processing
these specimens.

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