Age-related decline in behavioral discrimination of amplitude modulation frequencies compared to envelope-following responses

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1 Abstract

The ability to discriminate modulation frequencies is important for speech intelligibility because speech has amplitude and frequency modulations. Neurophysiological responses assessed by envelope following responses (EFRs) significantly decline at faster amplitude modulation frequencies (AMF) in older subjects. A typical assumption is that a decline in EFRs will necessarily result in corresponding perceptual deficits. To test this assumption, we investigated young and aged Fischer-344 rats' behavioral AMF discrimination abilities and compared to their EFRs. A modified version of prepulse inhibition (PPI) of acoustic startle reflex (ASR) was used to obtain behavioral performance. A PPI trial contains pulses of sinusoidal AM (SAM) at 128 Hz presented sequentially, a SAM prepulse with different AMF and a startle-eliciting-stimulus. To account for hearing threshold shift or agerelated synaptopathy, stimulus levels were presented at 10-dB lower or match to the aged peripheral neural activation (using auditory brainstem response wave I amplitude). When AMF differences and modulation depths were large, young and aged animals' behavioral performances were comparable. Aged animals' AMF discrimination abilities declined as the AMF difference or the modulation depth reduced, even compared to the young with peripheral matching. Young animals showed smaller relative decreases in EFRs with reduced modulation depths. The correlation of EFRs and AM perception was identified to be more consistent in young animals. The overall

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> results revealed larger age-related deficits in behavioral perception compared to EFRs, suggesting additional factors that affect perception despite smaller degradation in neural responses. Hence, behavioral and physiological measurements are critical in unveiling a more complete picture on the auditory function.

2

3 1. Introduction

Presbycusis is common and unavoidable in the elderly due to its properties 4 chronic deterioration and is asymptomatic early in life [66, 20]. It has of 5 been reported as the third most prevalent chronic disorder in the elderly $(\leq 65 \text{ years old})$ after hypertension and arthritis in the United States [40]. 7 Age-related changes in auditory structures and functions exist in both the 8 peripheral and central auditory systems [58, 59, 66, 6, 18, 72]. Age-related 9 degradation of the auditory periphery comprises loss or dysfunction of the 10 inner and outer hair cells [24, 59], alterations in the stria vascularis leading 11 to endocochlear potential reduction [8], and/or diminished auditory nerve 12 fibers (ANFs) and synapses [60]. Meanwhile, changes in excitatory/inhibitory 13 balance are reported and described as one of the main causes of age-related 14 auditory deficits in the central auditory system [6, 7, 53, 46]. Auditory central 15 degradation could result in degraded processing of complex sounds especially 16 in challenging situations, for example speech recognition in a cocktail party 17 [22].18

Human speech consists of complex and rapid modulations in amplitude 19 and frequency over time that are crucial for precise speech recognition [54, 20 61, 75]. Previously, our research team and others have revealed significant 21 age-related differences in temporal processing, assessed physiologically by en-22 velope following responses (EFRs) at the levels of the auditory midbrain and 23 brainstem, at faster AM frequencies (AMFs) [47, 52]. Psychoacoustic stud-24 ies using temporal modulation transfer functions (tMTFs) have also shown 25 that older adults have poor periodicity coding due to higher thresholds in 26 modulation depth and frequency modulation (FM) detection [25, 26]. We 27 have collected neurophysiological evidence from young and aged rats show-28 ing age-related differences in temporal processing of AM and FM [48, 47]. It 29 is assumed that larger EFR responses elicited by AM sounds are associated 30 with better perceptual performance [48, 2, 43]. However, there is a lack of be-31 havioral evidence that clarifies and confirms the relationship of physiological 32 and behavioral responses. 33

To assess and determine changes in neural processing related to auditory 34 impairments or brain disorders, the acoustic startle response (ASR) with its 35 modulation by a non-startling prepulse is broadly applied in behavioral sen-36 sory studies [37, 14, 62]. The ASR is a type of reflexive behavior manifested 37 as a transient contraction of facial and skeletal muscles in respond to a sud-38 den, brief and intensely loud sound [64, 39]. In rats, the ASR can be elicited 39 by an acoustic stimulus that is approximately more than 80 dB above the 40 hearing threshold [50]. Therefore, measurement of ASR can be used as an 41

indicator for the behavioral responsiveness or perception to acoustic stimuli. 42 Startle reflex behavior is convenient for age-related auditory studies because 43 it is an unconditioned reflex reaction and no animal training is required. It 44 has also been demonstrated that the ASR can be measured at any age past 45 juvenile in rats [67, 69]. The primary ASR circuit comprises the cochlear root 46 neurons, neurons in the caudal pontine reticular nucleus (PnC) and spinal 47 motor neurons [36, 10, 21]. This simple neural circuit has extremely short 48 latency because it involves only a few synapses located in the lower brainstem 49 [36, 10].50

The amplitude and probability of a startle movement following a SES can 51 be modulated by non-startling prepulses. A prepulse is a stimulus presented 52 prior to the SES. The amplitude of the ASR is attenuated significantly when 53 the prepulse is detected and processed by the subject [13]. Therefore, inhi-54 bition of the startle reaction using a prepulse is termed prepulse inhibition 55 (PPI). The magnitude of PPI is proportional to the subject's detectability 56 of the prepulse [33]. Prepulses have been used in the forms of acoustic [29], 57 visual [4] and tactile [51]. Animal studies have shown that auditory PPI is 58 associated with the function of the cochlear nucleus, the inferior and superior 59 colliculi (I/SC) and the pedunculopontine tegmental nucleus [36]. When a 60 prepulse is presented, the signal travels from the level of the cochlea to the 61 IC and then travels collaterally to the SC. Subsequently, the SC excites the 62 pedunculopontine tegmental nucleus, which inhibits the PnC, resulting in re-63 duced startle response [13, 36]. Hence, an interval of 20-500 ms between the 64

prepulse and the SES should provide sufficient time for the signal to inhibit
the ASR via PnC inhibition [13, 36, 37].

PPI can be induced by prepulses with various temporal characteristics. 67 Prepulse duration up to 100 ms are generally used in most PPI experiments 68 [32, 31, 17, 65]. Recently, other applications of the PPI paradigm were de-69 veloped using complex modulatory stimuli with relatively long duration, for 70 example 50-1000 ms gap prepulses in background noise [62]. Detection of an 71 amplitude modulated prepulse, which was presented during 1 s before the 72 SES, from a background of unmodulated noise has been demonstrated in 73 gerbils of two-month age [41]. Speech sounds of 100-300 ms have also been 74 used as prepulses in rats [15, 16]. Floody and Kilgard (2007) showed that 75 Sprague-Dawley rats of approximately four-month age were able to distin-76 guish syllable [pae] from [bae] with the application of the PPI paradigm. 77

In this study, we investigated AMF discrimination abilities of young and 78 aged F344 rats using the PPI paradigm. A modified test paradigm, adapted 79 from Floody and Kilgard's (2007) speech discrimination tasks, was applied 80 by replacing speech sounds with AM sounds. AM sounds modulated with 81 AMFs different from the AMF of background sounds were used as prepulses. 82 The behavioral results were then compared to EFRs of tMTFs recorded from 83 each of the tested animal. Sound levels that accounted for average sensation 84 level as well as sound levels that accounted for age-related cochlear synaptic 85 degeneration were used. As a whole, the results of this study should aid 86 in unveiling the relationship of neural AM processing and behavioral AM 87

⁸⁸ perception in aging.

⁸⁹ 2. Methods

90 2.1. Animals

Twelve young (3-11 months; mean b.w.: male = 264 g and female = 18391 g) and 14 aged (20-24 months; mean b.w.: male = 408 g and female = 242 g) 92 Fischer-344 (F344) rats obtained from Taconic (NIA colony) were used. All 93 animals were housed in the animal care facility during the period of this study 94 in a relatively quiet and standard condition. They were also maintained on 95 12-hour light and 12-hour dark cycle (light on at 6:00 and off at 18:00) with 96 water and food ad libitum. Behavioral experiments were performed during 97 the light phase of the light-dark cycle, mainly in between 13:00 and 18:00. 98 All protocols were approved by the Purdue Animal Care and Use Committee 99 (PACUC-1111000167). 100

101 2.2. Behavioral tests (ASR and PPI)

102 2.2.1. Setup and experimental procedure

All behavioral tests were performed in a sound attenuating cubicle (Med Associates) within a larger anechoic chamber (Industrial Acoustics). During the testing procedure, animals were placed on a grid rod animal holder on a motion-sensitive platform. Animals' startle responses were detected and transduced via an amplifier connecting to a TDT RZ6 system and the computer. The vertical movement of the platform, which resulted from a startle reaction, was converted into a voltage signal by a transducer.

Startle responses were measured from the beginning of each trial to 1.5 110 s after the offset of the SES. Acoustic stimuli, including background sounds 111 and prepulses, were generated by a TDT RZ6 system and presented via a 112 Fostex (FT28D Dome Tweeter) speaker. The SES was also generated by 113 the same TDT system and presented through a high frequency neodymium 114 compression driver (BMS speaker). Both speakers were placed behind the 115 animal holder. Stimulus presentation and response acquisition were manipu-116 lated by custom-written scripts using RPvdEx and MATLAB (MathWorks). 117 Calibration of the apparatus was carried out for frequencies 1-20 kHz using a 118 1/2" Bruel & Kjaer microphone connecting to Nexus preamplifier and an os-119 cilloscope (Tektronix). The microphone was placed inside the animal holder 120 at the middle of the cage, as recommended by the manual of Med Associates, 121 during the process of sound calibration. 122

For every animal that has not performed any behavioral PPI test before, 123 each of them was habituated to stay in the animal holder for 5-10 min for 3 124 successive days [68]. After 3 days of habituation, animals were then proceed 125 to perform an 8 kHz pure tone detection task or AMF discrimination task. 126 Each animal completed only one task (about 60 min) on one test day. A 127 complete task encompassed a total of 3 phases, which were named as phase 128 0, 1 and 2. In summary, phase 0 is an acclimation period for animals to 129 adapt to the animal holder, phase 1 is for habituation and association, and 130 phase 2 is the period in which the detection or discrimination task used for 131 analysis was carried out. 132

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133 2.2.2. 8 kHz pure tone detection task

Animals' abilities in detecting 8 kHz pure tones in a quiet background 134 were tested using prepulses of 8 kHz pure tones at sound levels of 25-75 135 dB SPL in 10-dB difference. In phase 0, animals underwent acclimation for 136 5 min. In phase 1, 30 trials of SES alone were performed for animals to 137 habituate to around 60 % of their initial startle responses [68]. Wideband 138 noise of 20 ms duration with zero rise fall times was used as the SES. The 139 intensity of the SES was set at 105 dB SPL for young animals and 115 dB 140 SPL for aged animals. The interval between the onset of each trial was 141 randomized between 15 and 30 sec so that animals could not estimate the 142 appearance of a SES. Phase 2 contains trials with a SES alone (served as 143 positive controls), trials with a prepulse placed before a SES and trials with 144 a prepulse alone (served as negative controls). The prepulses were 8 kHz 145 pure tones with a duration of 50 ms (5 ms rise fall times). The intensity of 146 a prepulse in each trial was pseudorandomized between 25 and 75 dB SPL 147 (10-dB gap). As each type of prepulse intensity repeated 9 times within one 148 complete task, a total of 72 trials were consisted in phase 2. Similar to phase 149 1. the intertrial interval in phase 2 was also randomized between 15 to 30 s. 150 Behavioral 8 kHz detection threshold was estimated for each animal by 151 comparing the ASR or RMS ratio measurements of no prepulse to the ASR 152 or RMS ratio measurements of 8 kHz prepulses at various sound levels. Sig-153 nificant decreases in the ASR or RMS ratio measurements of prepulses from 154 those of no prepulse were quantified using a one-sided t-test [41]. The mini-155

¹⁵⁶ mum sound levels that elicited a significant decrease in both of the measure¹⁵⁷ ment were averaged. This mean threshold was then taken as the behavioral
¹⁵⁸ 8 kHz detection threshold for the particular animal.

159 2.2.3. AMF discrimination task

AMF discrimination task was performed in a background of SAM tones. 160 An 8 kHz carrier (200 ms) with 128 Hz AMF at 100, 50 or 25 % AM depth 161 was presented as a background tone throughout the task. This SAM tone 162 was repeated multiple times (about 12-27 times) before a prepulse and a SES 163 were presented (Fig. 1). In phase 0, the background SAM tone was presented 164 at 1 /s for 5 min to allow animals to acclimate to the animal holder and the 165 background sounds. Phase 1, consisted of 20 trials, was used to habituate 166 animals in associating the prepulse, which has an AMF different from the 167 background, with a SES. In these 20 trials, the AMF of the prepulse was 168 set at the highest or lowest AMF (depending on the range of the AMF that 169 was tested in Phase 2) and presented alternatively. Fifty milliseconds after 170 the prepulse (200 ms) offset, the SES was released. The intertrial interval 171 was randomized between 15 and 30 s. The background AM tone was played 172 during the 15-30 s interval but became silent for 2.6 s after the generation of 173 a SES. The background AM tone was then resumed at the start of the next 174 trial. Phase 2 contained a total 81 trials (each trial type repeated 9 times) 175 and was used to measured PPI for AMF discrimination. The AMF of the 176 prepulse was varied from trial to trial to test animals' abilities in discrimi-177

nating it from the background AMF. The startle magnitude was expected to 178 be smaller if animals could discriminate the prepulse's AMF from the back-179 ground. In contrast, if animals could not discriminate the prepulse's AMF 180 from the background, the loud noise should trigger a relatively larger startle 181 response. All the trials in phase 2 could be categorized into four conditions: 182 (1) background only (negative control); (2) background and prepulse (nega-183 tive control); (3) background and SES (positive control); and (4) background, 184 prepulse and SES. Conditions (1) and (2) were negative controls because no 185 startle response should be induced in these two conditions. Condition (3) 186 served as a positive control since it contained a SES with no prepulse and a 187 large startle response should be triggered. In condition (4), reduced startle 188 response was expected if animals were able to discriminate a change in AMF 189 from the background. The AMFs that were tested in both young and aged 190 animals includes 16, 32, 64, 256, 512, 1024 Hz (\pm 3- to \pm 1-octave away from 191 128 Hz). A narrower AMF range was also tested in young animals and the 192 AMFs are 45, 64, 90, 181, 256 and 362 Hz (± 1.5 - to ± 0.5 -octave away from 193 128 Hz). The background SAM tones was randomly presented between 12 to 194 27 times (at 1/s for 12-27 s) from trial to trial in order to remove any other 195 possible cues that could be used by animals to predict the SES. The only 196 cue that should be used by animals to predict the SES would be based on 197 their abilities to distinguish a change in AMF from the background's AM. 198 Each animal repeated the same PPI behavioral test for 2 times to confirm 199 consistency. Overall, the experimental procedure, stimulus presentation and 200

²⁰¹ parameters for AMF discrimination task were designed by referring to the ²⁰² published literature [68, 56, 15].

In term of stimulus intensity, the background and the prepulse levels 203 were set at 85 dB SPL for aged animals and 75 dB SPL for young animals. 204 This 10-dB difference in the sound level used in young and aged animals 205 accounted for the average difference in sensation level at 8 kHz for young 206 and aged animals [49]. In addition, for the first set of AMFs at 100 or 50 %207 AM depth, we also tested young animals using sound levels that matched to 208 the aged's median ABR tone 8 kHz wave I amplitude at 85 dB SPL in order 209 to attain equivalent peripheral neural activation. This accounted for cochlear 210 synaptopathy and/or neuropathy as well as age-related differences in hearing 211 thresholds [60]. In this case, the average sound intensity was approximately 212 57.2 + - 5.1 dB SPL in the young based on the measurement of tone 8 kHz 213 ABR wave I amplitudes, which would be about 30 dB sensation level. 214

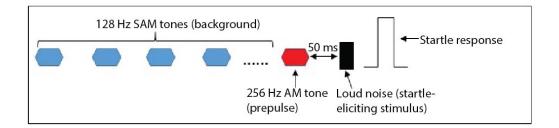


Figure 1: Presentation of background sounds, prepulse and startle-eliciting stimulus in a typical trial of the PPI behavioral task for AMF discrimination. The schematic shows an example of a PPI trial with multiple 128 Hz SAM tones presented in the background and a 256 Hz SAM tone used as a prepulse placing right before a startle-eliciting stimulus.

215 2.2.4. Startle response measurements and PPI calculation

Animal startle responses were recorded by the platform and then filtered 216 off-line with high-pass at 2 Hz and low-pass at 50 Hz. After filtering, a typical 217 startle response has a specific waveform as shown in Figure 2. Two different 218 methods were used to measure ASR responses [23]: (1) ASR magnitude: 219 the maximal peak-to-peak amplitude of transient voltage occurring within 220 300 ms after the offset of the SES; (2) ASR root mean square (RMS) ratio: 221 the RMS of the startle response (t_{ASR} , corresponding to a -100 to +200 ms 222 window relative to the first peak that occurred within 300 ms after the offset 223 of the SES) over the RMS of the baseline (t_{NF} , ref. Fig.2). The measured 224 mean ASR amplitude or mean RMS ratio for each trial type was estimated 225 as the average of all the ASR amplitudes or the RMS ratios after the highest 226 and lowest values were excluded [67]. This is to remove any possible outliers 227 as well as reduce variability of the responses. The percent of PPI (i.e. the 228 percent of startle magnitude reduced by the prepulse as compared to the 229 positive control) for each trial type was calculated using the below formula: 230 PPI % = [1- (ASR magnitude or RMS ratio to prepulse - baseline)/ (ASR 231 magnitude or RMS ratio of startle only -baseline)] x 100 %. 232

Magnitude or RMS ratio of baseline was measured from negative controls (trials with no SES) while startle only was measured from positive controls (trials of background and loud noise with no prepulse). A PPI % value that is close to or at 0 indicates that the prepulse does not have an inhibitory effect on animals' startle responses, which also indicates that animals could

not discriminate the prepulse from the background. However, a PPI % that
is near to 100 % indicates an almost complete inhibition of startle responses
by the prepulse.

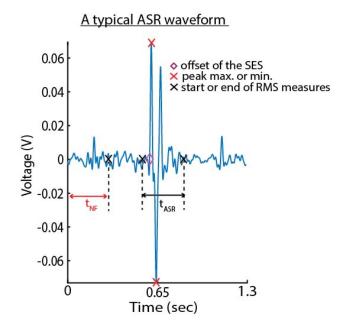


Figure 2: A typical acoustic startle response (ASR) waveform with distinct peaks and troughs that are above or below the noise floor (NF). The schematic shows an example of an ASR waveform obtained from a PPI trial. The offset of the startle-eliciting stimulus (SES), the start and end for root-mean-square (RMS) measures are labeled on the plot. For RMS ratio calculation, the time window of an ASR response is denoted by t_{ASR} while t_{NF} indicates the time window used for the noise floor. Both t_{ASR} and t_{NF} are 300 ms in duration.

241 2.3. Auditory evoked potentials

The experimental protocols used for ABR and EFR recordings were similar to previously described details in Parthasarathy and Bartlett (2012). All recordings were performed in a 9'x9' double-walled anechoic chamber (Industrial Acoustic Corporation). The animals were anesthetized using isofluorane

at 4 % and later maintained under 1.5-2 % isofluorane for placing the elec-246 trodes. Subdermal needle electrodes (Ambu) were placed on the animals' 247 scalps in a two-channel configuration. For channel 1, a positive electrode 248 was placed along the midline of the forehead in the the Cz to Fz position. 249 For channel 2, another positive electrode was placed horizontally along the 250 interaural line, which is above the location of the inferior IC. The negative 251 electrode was placed under the ipsilateral ear, along the mastoid, while the 252 ground electrode was placed in the nape of the neck. Electrode impedance 253 was confirmed to be less than 1 k Ω by testing with a low-impedance amplifier 254 (RA4LI, Tucker Davis Technologies or TDT). Before taking off isofluorane, 255 the animals were injected (intramuscular) with dexmedetomidine (Dexdomi-256 tor, 0.2 mg/kg), an α -adrenergic agonist acting as a sedative and an anal-257 gesic. Recording was then started after a 15-min waiting time for the effects 258 of isofluorane to wear off. The animals were maintained in an unanesthetized 259 and immobile condition during the whole session of recording. 260

Tone 8 kHz ABRs were recorded using brief 8 kHz pure tones of 2 ms 261 duration (0.5 ms \cos^2 rise/fall time), alternating polarity and presenting at 262 26.6/sec.The acquisition window was set to 30 ms and each ABR was 263 acquired as an average of 1500 repetitions (750 each polarity). Stimulus 264 intensity of the pure tone was decreased from 95 dB SPL to 15 dB SPL in 5-265 dB steps. This enabled us to obtain the animal's hearing threshold at 8 kHz 266 as well as the magnitude of wave I at each sound level, which was used as an 267 indicator for the amount of activated ANFs. The median of tone 8 kHz ABR 268

wave I amplitudes at 85 dB SPL from aged animals was used for stimulus 269 intensity matching of peripheral activation in young animals. Sinusoidally 270 amplitude modulated (SAM) tones with a 8 kHz carrier were used as acoustic 271 stimuli for EFRs. At 100 %, 50 % or 25 % modulation depth, the AMF of 272 the SAM tones was systematically increased from 16 to 2048 Hz in 0.5-octave 273 steps to generate the tMTF. The stimulus intensity was set at 75 dB SPL for 274 young animals and 85 dB SPL for aged animals. In young animals, sound 275 levels that matched to the aged's median ABR tone 8 kHz wave I amplitude 276 at 85 dB SPL were also recorded. 277

All stimuli were presented free-field to the right ear of the animal at a 278 distance of 115 cm from a speaker (Bower and Wilkins DM601). Stimuli 279 were generated using SigGenRP (TDT) at a 100-kHz sampling rate. Stimuli 280 presentation and response acquisition were conducted using BioSig software 281 (TDT). Waveforms were converted to sounds and delivered through a multi-282 channel processor (RX6, TDT) via the speaker. Digitized response waveform 283 was recorded with a multichannel recording and stimulation system (Rz5, 284 TDT). Responses were analyzed with BioSig and a custom-written program 285 in MATLAB. 286

All collected EFRs were low-pass filtered at 3000 Hz. EFRs were also high-pass filtered at 12 Hz for AMFs of 12-24 Hz, 30 Hz for AMFs of 32-64 Hz and 80 Hz for AMFs faster than 90 Hz. Filtered data were then exported as text files and analyzed using custom-written MATLAB scripts. Fast Fourier transform (FFT) were performed on time-domain waveforms from 10 to 190

ms relative to stimulus onset to exclude transient auditory brainstem re-292 sponses at the beginning. The maximum magnitude of the evoked response 293 at one of the three frequency bins (3 Hz/ bin) around AMF was measured 294 as the peak FFT amplitude. The noise floor was calculated as the average 295 magnitude of five frequency bins above and below the central three bins. A 296 peak response was taken to be significantly above noise level if the FFT am-297 plitude was at least 6 dB above the noise floor for the slower AMFs and at 298 least 10 dB above the noise floor for AMFs faster than 64 Hz to account for 299 the sharply decreasing noise floor. 300

301 2.4. Statistical analysis

Repeated measures ANOVAs (rmANOVAs) were performed to compare 302 ASR responses or FFT amplitudes of young and aged groups as well as 303 across different stimulus conditions using custom written scripts in SAS (Proc 304 MIXED, SAS Institute, Cary, NC, USA). Main effects and interactions ef-305 fects of each factor were analyzed based on comparisons of least squares (LS) 306 means. Data distributions were checked for normality using normal prob-307 ability plots of the residuals (proc UNIVARIATE). The differences in LS 308 means with a confidence level of 95 % was used when reporting significant 309 differences. LS means +/- standard error of mean (SEM) are shown in the 310 figures. 311

312 3. Results

313 3.1. 8 kHz tone detection in a quiet background

Prepulses of 8 kHz pure tones at sound intensities of 25-75 dB SPL, in 314 10-dB difference, were used to test animals' hearing sensitivities at 8 kHz. 315 The growth of PPI as a function of sound level, i.e. PPI values increased as 316 8 kHz prepulse intensity increased, was observed in young and aged animals 317 as shown in Fig. 3. For almost all of the sound levels, young animals had 318 larger PPI values than old animals although age-related differences were not 319 statistically significant. For each age group, PPI values at higher sound levels 320 were significantly larger than PPI values at lower sound levels, e.g. 75 > 35321 db SPL. Table 1 shows sound levels with PPI that are significantly different 322 from each other in young and aged animals for each of the measurement. 323 In addition, SEM of aged animals tended to be larger at lower sound levels 324 (25-45 dB SPL). This indicates that young animals were more behaviorally 325 consistent at perceiving 8 kHz tones at lower sound levels because of having 326 better hearing sensitivity. In young animals, the mean PPI values at each 327 sound level were significantly larger than 0 when tested using a t-test. How-328 ever, the mean PPI values were significantly larger than 0 in aged animals at 329 higher sound levels. Statistical analysis using rmANOVA revealed a signif-330 icant main effect of sound level for the measurement of ASR magnitude (F 331 = 17.52, p < 0.05) and ASR RMS ratio (F = 13.05, p < 0.05). However, no 332 significant age or age*sound level effect was observed for both measurements. 333

Behavioral 8 kHz detection threshold estimation using the measurements 334 of ASR and RMS ratio was performed for each animal. Young animals gen-335 erally have lower 8 kHz detection thresholds than aged animals. The mean 336 8 kHz detection threshold of the young was 39.5 + - 0.2 dB SPL while the 337 mean 8 kHz detection threshold of the aged was 61.9 + -0.17 dB SPL. How-338 ever, these thresholds were higher than the 8 kHz hearing thresholds obtained 339 from ABRs elicited by brief 8 kHz tones. The measured mean tone 8 kHz 340 ABR threshold for the young was 25.5 + - 0.04 dB SPL and for the aged 341 was 37.2 + - 0.09 dB SPL. Statistical comparisons of hearing thresholds for 342 age vs. young or behavior vs. ABR were performed using rmANOVAs. The 343 results show main effect of Age (F = 12.44, p < 0.05) and Measure type (F 344 = 22.61, p < 0.05) but no significant interaction effect. 345

Sound level (dB SPL)	25	35	45	55	65
ASR magnitue					
Young	55,65,75	65, 75	65, 75	75	
Aged	45, 55, 65, 75	55,65,75	65, 75	75	
RMS ratio					
Young	65, 75	65, 75	65, 75	75	
Aged	$\begin{array}{c} 65,\ 75 \\ 55,\ 65,\ 75 \end{array}$	55, 65, 75	75		75

Table 1: For 8 kHz prepulse detection, PPI values of lower sound levels were mostly significantly different from PPI values of higher sound levels. This table shows sound levels with PPI that are significantly different from each other within each age group according to the results of rmANOVAs for Figure 3.

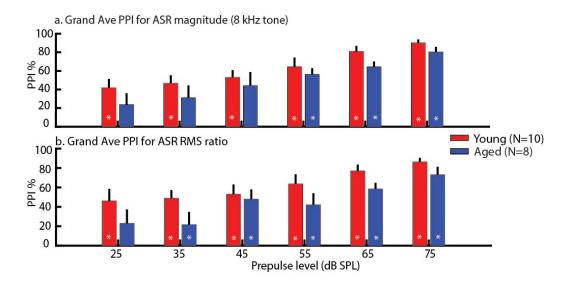


Figure 3: Prepulse inhibition (PPI) using prepulses of 25-75 dB SPL 8 kHz pure tones in a quiet background showed similar growth in PPI as sound intensity increased in young and aged animals. PPI values of higher sound intensities were larger than those of lower sound intensities. The black asterisks indicate p < 0.05 for PPI comparison between age groups and at the same sound level. The white asterisks in bars indicate p < 0.05 for mean PPI not equal to zero using a t-test. All statistically significant differences were obtained using least squares means comparison from rmANOVA and PPI comparison between sound levels within an age group is summarized in Table 1.

346 3.2. Behavioral discrimination of AMFs

347 3.3. In young animals

The first set of frequencies tested in young animals for AMF discrimination includes the range of 16-1024 Hz with 1-octave difference. Each AMF is 1, 2 or 3 octaves higher or lower than 128 Hz AM. The same AMF discrimination task was performed by fixing AM depths of all SAM tones at either 100, 50 or 25 %. The PPI results obtained with these three AM depths using either ASR magnitude or RMS ratio were shown in Figure 4. When comparing PPI values among different AM depths but at one single AMF,

higher inhibition was observed for larger AM depths compared to smaller 355 AM depths, e.g. 100 % > 50 % > 25 %. Statistical significance for PPI 356 values being higher at larger AM depths compared to smaller AM depths 357 was observed at most AMFs. In addition, when comparing PPI values across 358 different AMFs but within the same AM depth, a trend of higher PPI was 359 observed at AMFs that were further away from 128 Hz for 50 and 25 % AM 360 depths. At 25 % AM depth, grand average PPIs of almost all the tested 361 AMFs generally had larger SEMs. This indicates that behavioral variability 362 among young animals in AMF discrimination increased when AM depth re-363 duced. According to the results of t-tests, the mean PPI values at each AMF 364 at 100 and 50 % depth were all significantly different from 0 indicating signif-365 icant inhibitory effects. In contrast, the mean PPI values at 25 % depth were 366 not significantly different from 0 at most AMFs except 1024 Hz. In addition, 367 a significant main effect of AM depth was obtained from rmANOVA for the 368 measurements of ASR magnitude (F = 10.51, p < 0.05) and RMS ratio (F 369 = 14.54, p < 0.05). 370

The second set of frequencies tested on the young includes the range of 45-362 Hz separated in 0.5-octave difference. Each AMF is 0.5-, 1- or 1.5-octave away from 128 Hz AM. In Figure 5, PPI values at 100 % depth were relatively higher than 50 % depth. When comparing PPI across different AMFs at 50 % AM depth, a trend of increased PPI was observed when AMFs were further away from 128 Hz. Moreover, for 50 % AM depth, grand average PPI of most AMFs had larger SEM indicating variability among young animals in

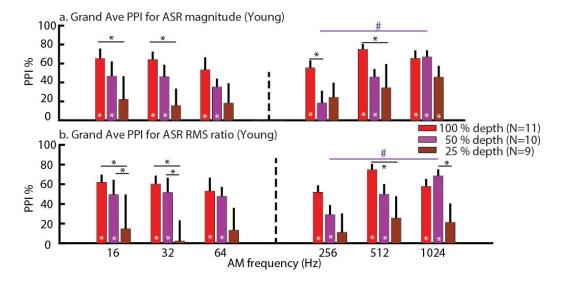


Figure 4: In young animals, PPI values were higher for larger AM depths compared to lower AM depths (e.g. 100 % > 50 % > 25 %) at various AMFs (16-1024 Hz in 1-octave difference). For 50 % AM depths, PPI tended to increase as AMFs were further away from 128 Hz. The black asterisks indicate p < 0.05 for PPI comparison between different AM depths within the same AMF while the pound signs indicate p < 0.05 for PPI comparison between different AMFs but within the same AM depth. All statistically significant differences were obtained using least squares means comparison from rmANOVA. The white asterisks in bars indicate p < 0.05 for PPI not equal to zero using a t-test.

AMF discrimination increased as AM depth reduced. The mean PPI values were significantly larger than 0 for almost all AMFs at 100 % depth but not for 50 % depth. According to rmANOVA, there is a significant main effect of AM depth for both ASR magnitude measurement (F = 17.69, p < 0.05) and RMS ratio measurement (F = 11.74, p < 0.05).

383 3.4. Young vs. aged animals

AMF discrimination was tested in young and aged animals using stimulus intensity of either 75 (young) or 85 db SPL (aged). The tests were performed at either 100 or 50 % AM depth. Young animals were also tested at sound

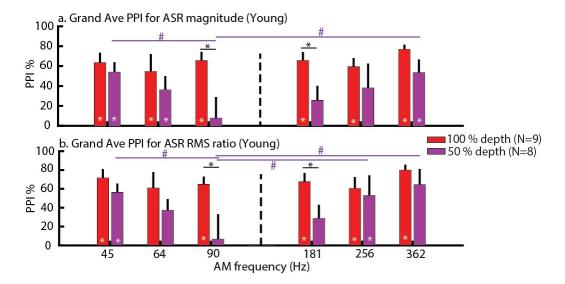


Figure 5: A trend of higher PPI was observed for 100 % AM depth compared to 50 %. The PPI results were obtained from a more difficult task in which the AMF range was set at 0.5-1.5 octave away from 128 Hz. Indications for the asterisk and the pound signs are similar to Figure 4.

levels (an average of about 55.3 db SPL) that matched to the aged median 387 tone 8 kHz ABR wave I amplitude to achieve equivalent peripheral neural 388 activation. This accounted for cochlear synaptopathy and/or neuropathy 389 [60] as well as age-related differences in hearing thresholds because ABR 390 wave I amplitude reflects the amount of activated and synchronized auditory 391 neurons [55, 9]. Figure 6 shows the results of PPI obtained at 100 % AM 392 depth. There was a trend of aged PPI values at 85 dB SPL being lower 393 than PPI of the young at 75 dB SPL and at matched peripheral activation. 394 Young PPI values at 75 dB SPL and at matched peripheral activation were 395 similar except at 1024 Hz AMF. Statistical analysis using rmANOVA revealed 396 significant main effect of AMF for PPI measured with ASR magnitude (F =397

³⁹⁸ 4.1, p < 0.05) and RMS ratio (F = 3.42, p < 0.05).

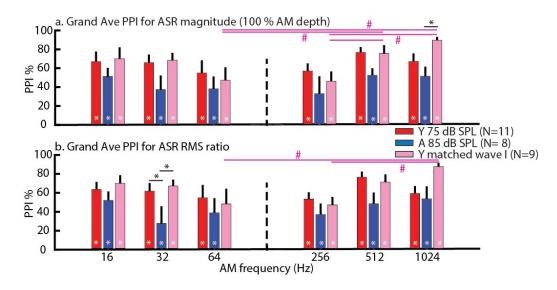


Figure 6: **PPI was detectable in aged animals for almost all AMF differences for one octave spacing and 100% AM depth.** There was a trend of aged PPI values being lower than the young at 75 dB SPL and at matched peripheral activation. The pound signs indicate p < 0.05 for PPI comparison between different AMFs within the same age group. All statistically significant differences were obtained using least squares means comparison from rmANOVA. In the legend, Y indicates young animals while A indicates aged animals. The white asterisks in bars indicate p < 0.05 in t-test for mean PPI not equal to zero. In the legend, Y indicates young animals while A indicates aged animals.

Figure 7 shows the results of PPI obtained at 50 % AM depth. In the 399 young 75 dB SPL, PPI values were generally smaller than for 100 % depth 400 (cf. Fig 6), but still showed PPI significantly higher than zero. By contrast, 401 the PPI responses for the aged 85 dB SPL and the young with peripheral 402 matching were not significantly above zero at some AMFs (e.g. 16, 256 and 403 512 Hz). When AM depth reduced to 50 %, AMF discrimination abilities 404 for the aged at 85 dB SPL and the young at matched peripheral activation 405 reduced, especially at 256 Hz AMF. According to rmANOVAs, there was a 406

significant main effect of AMF obtained from rmANOVAs for PPI measured using the ASR magnitude method (F = 6.71, p < 0.05) and the ASR RMS ratio method (F = 7.55, p < 0.05). The rmANOVA results for the ASR RMS ratio also showed a significant main effect of Age (F = 9.28, p < 0.05).

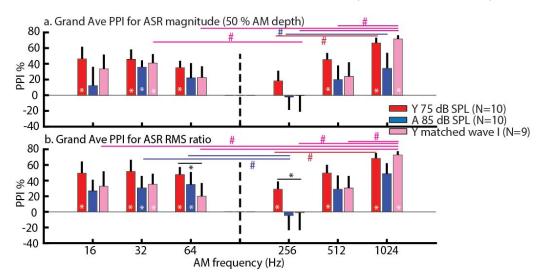


Figure 7: For AMF discrimination at 50 % AM depth, a trend of higher PPI values in young animals (75 dB SPL) across AMFs was observed. Aged animals had PPI close to baseline or in negative values especially when responses were measured using RMS ratio. PPI values of young animals at 75 dB SPL or matched wave I were mostly not significantly different from the aged. The black asterisks indicate p < 0.05 for PPI comparison between age groups but at the same AMF. All statistically significant differences were obtained using least squares means comparison from rmANOVA. The white asterisks in bars indicate p < 0.05 in t-test for mean PPI not equal to zero. In the legend, Y indicates young animals while A indicates aged animals.

411 3.5. Electrophyiological responses for AMF perception

Electrophyiological responses elicited by AMFs ranging from 16-2048 Hz were recorded in both young and aged animals via EFRs using 8 kHz tone carriers (Fig. 8a). Sound levels were set at 75 dB SPL for the young and 85 dB SPL for the aged, which has been shown to evoke peak EFR responses in

most animals [47]. Fig. 8a shows EFRs of tMTFs with 100, 50 or 25 % AM 416 depth in young and aged animals. At 100 % AM depth, the young EFRs 417 were generally higher than the aged even though the stimulus level used in 418 the aged was 10 dB SPL louder. For aged animals, their EFRs at 100 % AM 419 depth were similar to the young EFRs at 50 % AM depth. Moreover, the aged 420 EFRs at 50 % AM depth were also similar to the young EFRs at 25 % AM 421 depth. However, when EFRs of tMTFs were recorded at equivalent peripheral 422 activation, the aged EFRs at 100 % AM depth were significantly higher than 423 the young EFRs at 100 % AM depth (Fig. 8b). Although differences were 424 smaller, the aged EFRs at 50 % AM depth were still significantly larger than 425 the young EFRs at 50 % AM depth. According to statistical analysis using 426 rmANOVA for EFRs recorded at equivalent peripheral activation, the main 427 effects of age and AMF as well as their interaction effect were statistically 428 significant (p < 0.05). At 100 % AM depth, the F-values of age and AMF 429 main effects were 19.97 and 52.92, respectively. The interaction effect of 430 age*AMF had an F-value of 5.68. For 50 % AM depth, the F-values of 431 age and AMF main effects were 6.68 and 179.12, respectively while the F-432 value for the interaction effect of age*AMF was 2.13. We did not perform 433 statistical analysis for EFRs in Fig. 8a because the emphasis was to observe 434 the trends and how EFRs of tMTFs with different AM depths were distinct 435 or overlapped. 436

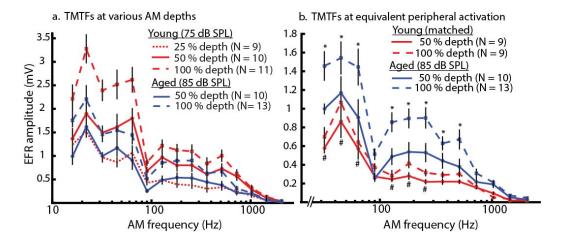


Figure 8: Young animals' EFR amplitudes were generally larger at 75 dB SPL compared to aged animals at 85 dB SPL but their EFR amplitudes were lower than the aged at equivalent peripheral activation. (a) EFRs of temporal modulation transfer functions (tMTFs) with 100, 50 or 25 % AM depth recorded in young and aged animals, respectively. (b) EFRs of tMTFs with 100 or 50 % AM depth recorded in both age groups at matched peripheral activation. The asterisks indicate p < 0.05 for comparison of EFR amplitudes between young and aged animals for tMTFs with 100 % AM depth while the pound signs indicate p < 0.05 for comparison of EFR amplitudes between young and aged animals for tMTFs with 100 % AM depth while the pound signs indicate p < 0.05 for comparison of EFR amplitudes between young and aged animals for tMTFs with 100 % AM depth while the pound signs indicate p < 0.05 for comparison of EFR amplitudes between young and aged animals for tMTFs with 50 % AM depth. All statistically significant differences were obtained using least squares means comparison from rmANOVA.

437 3.6. Relationship of EFRs and behavioral PPI

To identify the relationship between neurophysiological responses and be-438 havioral AMF discrimination at each of the tested AMFs, changes in each 439 of these measures due to a change in temporal salience of AM depth were 440 compared simultaneously. The changes in behavioral PPI or the changes in 441 EFR amplitudes as temporal salience of AM depth dropped from 100 to 50 %442 were measured at each of the tested AMF and in each age group. As shown 443 in Figure 9, changes in PPI values were plotted on the left ordinate while 444 changes in EFR amplitudes were plotted on the right ordinate. The changes 445

in PPI values (ΔPPI) were measured as PPI % at 100 % AM depth minus
PPI % at 50 % AM depth from the same animals. The changes in EFRs
(EFR ratio) were measured as EFR amplitudes at 50 % depth divided EFR
amplitudes at 100 % depth from the same animal as well.

For young animals (75 dB SPL), consistent smaller changes in EFRs and 450 PPIs due to a decrease in stimulus AM depth were observed. This indicates 451 that their abilities in AMF discrimination and EFR responses to the tested 452 AMFs were not much affected by a reduction in AM depth. For aged animal 453 (85 dB SPL), the trend of EFR ratio over AMF behaved similarly to young 454 animals (75 dB SPL) but their Δ PPIs were larger compared to young animals 455 (75 dB SPL). There was a larger change in behavioral AMF discrimination 456 performance due to a reduction in AM depth although changes in EFRs 457 were relatively smaller. The trend observed in young animals seemed to 458 hold even when they were tested at matched peripheral activation. The 459 changes in behavioral PPI were slightly larger compared to those at 75 dB 460 SPL. Overall, a smaller change in EFR correlated with a smaller change in 461 behavioral PPI value in young animals at both 75 dB SPL and at equivalent 462 peripheral activation. However, this correlation was no longer consistent in 463 aged animals. 464

465 4. Discussion

466 4.1. Behavioral PPI audiometry versus ABRs

The paradigm of behavioral ASR and PPI has been used to assess audi-467 tory behavior in rodents [56, 65, 63, 42, 45, 19, 62, 41]. Using standard PPI 468 techniques in the absence of a background sound, both younger and older 469 animals exhibited PPI whose amplitude increased with increasing salience of 470 the prepulse (Fig. 3). For a 25 dB prepulse, PPI was significantly larger 471 than 0 in younger animals, comparable to their ABR thresholds and consis-472 tent with previous studies [42]. As expected based on the ABR thresholds, 473 PPI magnitudes tended to be smaller in older animals for lower prepulse 474 levels, but still grew with increasing level and achieved similar peak PPI. 475 Therefore, animals of all ages tested exhibited the PPI behavior and to a 476 similar degree. 477

478 4.2. Aging effects on PPI of ASR

Age-dependent reduction on startle responses elicited by acoustic stim-479 uli in rodents, including F344 rats, have been reported in previous studies 480 [56, 69, 45, 30, 38]. It has been suggested that age-related changes in ASR 481 cannot be directly attributed to hearing loss because different ASR ampli-482 tudes were obtained from young adult rats of different strains with similar 483 hearing sensitivities [56]. In our study, we observed comparable PPI val-484 ues, especially at supra-threshold prepulse levels, for 8 kHz detection task 485 in young and aged animals (Fig. 3). This is different that the reduction of 486

PPI efficiency associated with aging reported in F344 rats by Rybaklo et al. (2012). At 100 % AM depth (Fig. 6), the aged and young had similar PPI values for AMF differences of 2-3 octaves. For 1 octave AMF difference, PPI tended to be reduced in the aged 85 dB SPL and the young with peripheral matching (Fig. 6). When AM depth salience decreased (Fig. 6), the observed age-related reductions of PPI further suggest a deficit in temporal processing leading to impaired perception.

494 4.3. AM frequency discrimination

Amplitude modulation is used by humans and animals to aid in auditory 495 object formation [5, 3]. Many studies have used tMTFs as a measure of 496 temporal acuity of the auditory system in psychoacoustic [71, 26, 1, 35] as 497 well as in electrophysiological studies [12, 47, 52]. AM depth sensitivity 498 as a function of AMF has been demonstrated as similar for rats [35] and 499 other mammals, including humans [71] and chinchillas [27]. A progressive 500 decrease in AM depth sensitivity (behavioral threshold became worse) of a 501 noise carrier modulated between 5-2000 Hz were observed in rats [35] and 502 rats having better AM depth sensitivity at AMFs of 10-60 Hz was also found 503 to be similar to humans [71]. The behavioral tMTFs of humans [44], rats 504 [35], barn owls [11] and chinchilla [57] showed a low-pass characteristic for 505 AM detection resembling the electrophyiological tMTFs in F344 rats shown 506 in this study (Fig. 8) and in our previous study [47]. For low modulation 507 depths (25%), there was little evidence of discrimination in young animals 508

for most AMFs. Despite this, PPI was evident for 1024 Hz AM (Fig. 4a), 509 suggesting that AM discrimination even at low modulation depths (25%) is 510 possible at AMF well above those that thalamic and cortical neurons can 511 phase-lock to [34], suggesting that spectral cues and rate coding may be 512 used. As task difficulty increased by reducing AM depth (Fig. 7), aged 513 animals performed worse. Young animals tested at equivalent peripheral 514 activation (55.3 dB SPL) performed better than the aged 85 dB SPL (Fig. 515 7) implying that peripheral activation by itself does not fully account for 516 behavioral performance. 517

518 4.4. Correlation of behavioral auditory responses and the underlying neural 519 responses

When the temporal salience of AM depth was decreased from 100 to 50 520 % depth, the degree of the EFR phase-locking to the SAM stimuli decreased 521 (Fig. 8 and 9). If EFR amplitudes have a strong link to behavioral perfor-522 mance, we expect that this should result in a decline in temporal perception 523 (Fig. 9). When we compared changes in EFRs versus changes in behavioral 524 PPI values due to a change in AM depth, Figure 9 reveals that both neuro-525 physiological and behavioral changes in young animals were correlated at 75 526 dB SPL as well as at softer sound levels (equivalent peripheral activation). 527 A relative smaller change in behavioral PPI was associated with a relative 528 smaller change in neural responses to SAM stimuli at the tested AMFs in the 529 young 75 dB SPL. However, this correlation was no longer seemed to hold in 530

the aged 85 dB SPL. A relatively smaller reduction in EFRs was observed to 531 result in a larger decline in behavioral PPI in aged animals. This observa-532 tion is analogous to the findings of Xu and Gong (2014). When behavioral 533 frequency difference limens (FDLs) and two-tone evoked frequency-following 534 responses (FFRs) were measured in normal hearing young adults, they ob-535 served that frequency difference of two-tone, which was able to evoked FFRs, 536 was smaller than behavioral FDL threshold [74]. Therefore, these and our re-537 sults show that the neurophysiological measurements of EFRs or FFRs may 538 be more sensitive than behavioral measurements because a smaller change 539 in stimulus parameters can be detected physiologically but the response is 540 not expressed behaviorally. Other behavioral tasks may be more sensitive, 541 or it may be that phase-locking physiological measures are too sensitive [28]. 542 These data also suggest that age-related degradation that exists beyond the 543 auditory brainstem and midbrain could have a larger contribution to the de-544 cline in behavioral perception [73]. In addition, since we performed tone 8 545 kHz ABR wave I amplitude matching to achieve equivalent peripheral ac-546 tivation, which accounts for age-related increase of hearing threshold and 547 age-related neuropathy/synaptopathy [60, 70], age-related decline in behav-548 ioral AMF discrimination should be due to more of a central effect and less 549 to a peripheral effect. 550

In conclusion, we examined the relationship of behavioral AM perception and neurophysiological responses to similar stimuli by measuring PPI of ASRs and EFRs. The young behavioral performance in discriminating dif-

ferent AMFs dropped gradually as salience of AM depth reduced from 100 to 554 25 % depth. Comparable behavioral performances at AMFs 1-2 octaves away 555 from 128 Hz were observed in young and aged animals when AMF spacing 556 was larger and at 100 % AM depth. At 50 % AM depth, age-related decline 557 of EFRs was smaller but aged animals' AMF discrimination performance was 558 highly compromised. When physiological and behavioral results were com-559 pared, the correlation of AM processing and AM perception were identified 560 to be more consistent in the young, including even when peripheral activa-561 tion was matched. Overall, the results reveal a larger age-related deficit in 562 behavioral perception compared to auditory evoked potentials using similar 563 SAM stimuli. This suggests that behavioral and physiological measurements 564 should be combined to capture a more complete view on the auditory function 565 and aid in identifying the localization of age-related auditory deficits. 566

567 5. References

- ⁵⁶⁸ [1] Bacon, S. P. and Viemeister, N. F. (1985). Temporal modulation transfer
 ⁵⁶⁹ functions in normal-hearing and hearing-impaired listeners. *Audiology*,
 ⁵⁷⁰ 24(2):117–34.
- ⁵⁷¹ [2] Boettcher, F. A., Poth, E. A., Mills, J. H., and Dubno, J. R. (2001).
 ⁵⁷² The amplitude-modulation following response in young and aged human
 ⁵⁷³ subjects. *Hearing Research*, 153(1-2):32–42.
- 574 [3] Bohlen, P., Dylla, M., Timms, C., and Ramachandran, R. (2014). De-

tection of Modulated Tones in Modulated Noise by Non-human Primates.
Journal of the Association for Research in Otolaryngology, 15(5):801–21.

- ⁵⁷⁷ [4] Buckland, G., Buckland, J., Jamieson, C., and Ison, J. R. (1969). Inhibi-⁵⁷⁸ tion of startle response to acoustic stimulation produced by visual prestim-
- ulation. Journal of Comparative and Physiological Psychology, 67(4):493–
 6.
- ⁵⁸¹ [5] Bürck, M. and van Hemmen, J. L. (2009). Neuronal identification of signal
 ⁵⁸² periodicity by balanced inhibition. *Biological Cybernetics*, 100(4):261–70.
- [6] Caspary, D. M., Ling, L., Turner, J. G., and Hughes, L. F. (2008). Inhibitory neurotransmission, plasticity and aging in the mammalian central
 auditory system. J Exp Biol, 211(Pt 11):1781–91.
- ⁵⁸⁶ [7] Caspary, D. M., Schatteman, T. A., and Hughes, L. F. (2005). Age⁵⁸⁷ related changes in the inhibitory response properties of dorsal cochlear nu⁵⁸⁸ cleus output neurons: Role of inhibitory inputs. *Journal of Neuroscience*,
 ⁵⁸⁹ 25(47):10952–9.
- [8] Chen, G. D., Li, M., Tanaka, C., Bielefeld, E. C., Hu, B. H., Kermany,
 M. H., Salvi, R., and Henderson, D. (2009). Aging outer hair cells (OHCs)
 in the Fischer 344 rat cochlea: function and morphology. *Hearing Research*,
 248(1-2):39–47.
- ⁵⁹⁴ [9] Chen, T. and Chen, S. (1991). Generator study of brainstem auditory

- evoked potentials by a radiofrequency lesion method in rats. *Experimental brain research*, 85(3):537–42.
- ⁵⁹⁷ [10] Davis, M., Gendelman, D. S., Tischler, M. D., and Gendelman, P. M.
- (1982). A primary acoustic startle circuit: lesion and stimulation studies.
 The Journal of Neuroscience, 2(6):791–805.
- [11] Dent, M., Klump, G., and Schwenzfeier, C. (2002). Temporal modulation transfer functions in the barn owl (Tyto alba). Journal of *Comparative Physiology A: Sensory, Neural, and Behavioral Physiology*,
 187(12):937–43.
- [12] Fay, R. (1980). Psychophysics and neurophyiology of temporal factors
 in hearing by the goldfish amplitude-modulation detection. Journal of *Neurophysiology*, 44(2):312–32.
- [13] Fendt, M., Li, L., and Yeomans, J. S. (2001). Brain stem circuits mediating prepulse inhibition of the startle reflex. *Psychopharmacology*, 156(23):216–24.
- [14] Fitch, R. H., Threlkeld, S. W., McClure, M. M., and Peiffer, A. M.
 (2008). Use of a modified prepulse inhibition paradigm to assess complex
 auditory discrimination in rodents. *Brain Research Bulletin*, 76(1):1–7.
- [15] Floody, O. R. and Kilgard, M. P. (2007). Differential reductions in
 acoustic startle document the discrimination of speech sounds in rats. *The Journal of the Acoustical Society of America*, 122(4):1884–7.

- ⁶¹⁶ [16] Floody, O. R., Ouda, L., Porter, B. A., and Kilgard, M. P. (2010). Effects
- of damage to auditory cortex on the discrimination of speech sounds by rats. *Physiology & Behavior*, 101(2):260–8.
- [17] Friedman, J. T., Peiffer, A. M., Clark, M. G., Benasich, A. A., and Fitch,
- R. H. (2004). Age and experience-related improvements in gap detection
 in the rat. Developmental Brain Research, 152(2):83–91.
- ⁶²² [18] Frisina, R. D. (2010). Aging changes in the central auditory system. In
- Palmer, A. R. and Rees, A., editors, *The Oxford Handbook of Auditory* Science: The Auditory Brain, pages 415–36. Oxford University Press.
- [19] Gaese, B. H., Nowotny, M., and Pilz, P. K. (2009). Acoustic startle
 and prepulse inhibition in the Mongolian gerbil. *Physiology & Behavior*,
 98(4):460–6.
- [20] Gates, G. A. and Mills, J. H. (2005). Presbycusis. The Lancet,
 366(9491):1111–20.
- [21] Gomez-Nieto, R., Horta-Junior, J. d. A. C., Castellano, O., MillianMorell, L., Rubio, M. E., and Lopez, D. E. (2014). Origin and function
 of short-latency inputs to the neural substrates underlying the acoustic
 startle reflex. *Frontiers in Neuroscience*, 8.
- [22] Gordon-Salant, S. (2005). Hearing loss and aging: New research findings and clinical implications. The Journal of Rehabilitation Research and
 Development, 42(4s):9.

- 637 [23] Grimsley, C. A., Longenecker, R. J., Rosen, M. J., Young, J. W., Grims-
- ley, J. M., and Galazyuk, A. V. (2015). An improved approach to separat-
- ing startle data from noise. *Journal of Neuroscience Methods*, 253:206–17.
- [24] Harding, G. W., Bohne, B. A., and Vos, J. D. (2005). The effect of
 an age-related hearing loss gene (Ahl) on noise-induced hearing loss and
 cochlear damage from low-frequency noise. *Hearing Research*, 204(1-2):90–
 100.
- [25] He, N. J., Mills, J. H., Ahlstrom, J. B., and Dubno, J. R. (2008).
 Age-related differences in the temporal modulation transfer function with
 pure-tone carriers. *The Journal of the Acoustical Society of America*,
 124(6):3841–9.
- [26] He, N.-j., Mills, J. H., and Dubno, J. R. (2007). Frequency modulation
 detection: Effects of age, psychophysical method, and modulation waveform. *The Journal of the Acoustical Society of America*, 122(1):467–77.
- [27] Henderson, D., Salvi, R., Pavek, G., and Hamernik, R. (1984). Amplitude modulation thresholds in chinchillas with high-frequency hearing
 loss. J Acoust Soc Am, 75(4):1177–83.
- ⁶⁵⁴ [28] Henry, K. S., Abrams, K. S., Forst, J., Mender, M. J., Neilans, E. G.,
 ⁶⁵⁵ Idrobo, F., and Carney, L. H. (2016). Midbrain Synchrony to Envelope
- ⁶⁵⁶ Structure Supports Behavioral Sensitivity to Single-Formant Vowel-Like

Sounds in Noise. Journal of the Association for Research in Otolaryngology.

- [29] Hoormann, J., Falkenstein, M., Hohnsbein, J., and Blanke, L. (1992).
 The human frequency-following response (FFR) normal variability and
 relation to the click-evoked brain-stem response. *Hearing Research*,
 59(2):179–88.
- [30] Ison, J., Bowen, G., Pak, J., and Gutierrez, E. (1997). Changes in
 the strength of prepulse inhibition with variation in the startle baseline
 associated with individual differences and with old age in rats and mice. *Psychobiology*, 25(3):266-74.
- [31] Ison, J. R., Allen, P. D., Rivoli, P. J., and Moore, J. T. (2005). The
 behavioral response of mice to gaps in noise depends on its spectral components and its bandwidth. *The Journal of the Acoustical Society of America*,
 117(6):3944.
- [32] Ison, J. R. and Bowen, G. (2000). Scopolamine reduces sensitivity to
 auditory gaps in the rat, suggesting a cholinergic contribution to temporal
 acuity. *Hearing Research*, 145(1-2):169–76.
- [33] Ison, J. R. and Hoffman, H. S. (1983). Reflex modification in the domain of startle: II. The anomalous history of a robust and ubiquitous
 phenomenon. *Psychological bulletin*, 94(1):3–17.

- ⁶⁷⁷ [34] Joris, P. X., Schreiner, C. E., and Rees, A. (2004). Neural processing of
 ⁶⁷⁸ amplitude-modulated sounds. *Physiological Reviews*, 84:541–77.
- [35] Kelly, J. B., Cooke, J. E., Gilbride, P. C., Mitchell, C., and Zhang,
- H. (2006). Behavioral limits of auditory temporal resolution in the rat:
 amplitude modulation and duration discrimination. *Journal of comparative*
- ⁶⁸² psychology (Washington, D.C. : 1983), 120(2):98–105.
- [36] Koch, M. (1999). The neurobiology of startle. Progress in Neurobiology,
 59(2):107–28.
- [37] Koch, M. and Schnitzler, H.-U. (1997). The acoustic startle response
 in rats—circuits mediating evocation, inhibition and potentiation. *Be- havioural Brain Research*, 89(1):35–49.
- [38] Krauter, E., Wallace, J., and Campbell, B. (1981). Sensory-motor function in the aging rat. *Behavioral and neural biology*, 31(4):367–92.
- [39] Landis, C. and Hunt, W. A. (1939). The startle patter. New York, NY:
 Farrar & Rinehart.
- [40] Li-Korotky, H. S. (2012). Age-related hearing loss: quality of care for
 quality of life. *Gerontologist*, 52(2):265–71.
- [41] Lingner, A., Kugler, K., Grothe, B., and Wiegrebe, L. (2013).
 Amplitude-modulation detection by gerbils in reverberant sound fields. *Hearing Research*, 302:107–12.

697	[42] Longenecker, R., Alghamdi, F., Rosen, M., and Galazyuk, A. (2016).
698	Prepulse inhibition of the acoustic startle reflex vs. auditory brainstem
699	response for hearing assessment. Hearing Research, 339:80–93.
700	[43] Mamo, S. K., Grose, J. H., and Buss, E. (2016). Speech-evoked ABR:
701	Effects of age and simulated neural temporal jitter. Hearing Research,
702	333:201–9.
703	[44] O'Connor, K. N., Johnson, J. S., Niwa, M., Noriega, N. C., Marshall,

- E. A., and Sutter, M. L. (2011). Amplitude modulation detection as a function of modulation frequency and stimulus duration: comparisons between macaques and humans. *Hearing research*, 277(1-2):37–43.
- ⁷⁰⁷ [45] Ouagazzal, A.-M., Reiss, D., and Romand, R. (2006). Effects of age⁷⁰⁸ related hearing loss on startle reflex and prepulse inhibition in mice on
 ⁷⁰⁹ pure and mixed C57BL and 129 genetic background. *Behavioural Brain*⁷¹⁰ *Research*, 172(2):307–15.
- [46] Ouda, L., Profant, O., and Syka, J. (2015). Age-related changes in the
 central auditory system. *Cell and Tissue Research*, 361(1):337–58.
- [47] Parthasarathy, A. and Bartlett, E. (2012). Two-channel recording of
 auditory-evoked potentials to detect age-related deficits in temporal processing. *Hearing Research*, 289(1-2):52–62.
- ⁷¹⁶ [48] Parthasarathy, A. and Bartlett, E. L. (2011). Age-related auditory
 ⁷¹⁷ deficits in temporal processing in F-344 rats. *Neuroscience*, 192:619–30.

- ⁷¹⁸ [49] Parthasarathy, A., Datta, J., Torres, J. A., Hopkins, C., and Bartlett,
- E. L. (2014). Age-related changes in the relationship between auditory
- ⁷²⁰ brainstem responses and envelope-following responses. Journal of the As-
- sociation for Research in Otolaryngology, 15(4):649-61.
- ⁷²² [50] Pilz, P. K., Schnitzler, H. U., and Menne, D. (1987). Acoustic startle
 ⁷²³ threshold of the albino rat (Rattus norvegicus). Journal of comparative
 ⁷²⁴ psychology (Washington, D.C. : 1983), 101(1):67–72.
- [51] Pinckney, L. A. (1976). Inhibition of the startle reflex in the rat by prior
 tactile stimulation. Animal Learning & Behavior, 4(4):467–72.
- [52] Purcell, D. W., John, S. M., Schneider, B. A., and Picton, T. W. (2004).
 Human temporal auditory acuity as assessed by envelope following responses. *The Journal of the Acoustical Society of America*, 116(6):3581–93.
- [53] Rabang, C. F., Parthasarathy, A., Venkataraman, Y., Fisher, Z. L.,
 Gardner, S. M., and Bartlett, E. L. (2012). A Computational Model of
 Inferior Colliculus Responses to Amplitude Modulated Sounds in Young
 and Aged Rats. *Frontiers in Neural Circuits*, 6:77.
- [54] Rosen, S. (1992). Temporal information in speech: acoustic, auditory
 and linguistic aspects. *Philos Trans R Soc Lond B Biol Sci*, 336(1278):367–
 73.

- [55] Rowe, M. J. (1981). The brainstem auditory evoked response in neurological disease: a review. *Ear and Hearing*, 2(1):41–51.
- 740 [56] Rybalko, N., Bureš, Z., Burianová, J., Popelář, J., Poon, P. W., and
- ⁷⁴¹ Syka, J. (2012). Age-related changes in the acoustic startle reflex in Fischer
- $_{742}$ 344 and Long Evans rats. Exp Gerontol, 47(12):966–73.
- ⁷⁴³ [57] Salvi, R. J. (1982). Detection of sinusoidally amplitude modulated
 ⁷⁴⁴ noise by the chinchilla. *The Journal of the Acoustical Society of Amer-*⁷⁴⁵ *ica*, 71(2):424–9.
- [58] Schuknecht, H. F. (1964). Further observations on the pathology of
 presbycusis. Arch Otolaryngol, 80:369–82.
- [59] Schuknecht, H. F. and Gacek, M. R. (1993). Cochlear pathology in
 presbycusis. Ann Otol Rhinol Laryngol, 102:1–16.
- [60] Sergeyenko, Y., Lall, K., Liberman, M. C., and Kujawa, S. G. (2013).
 Age-related cochlear synaptopathy: an early-onset contributor to auditory
 functional decline. *Journal of Neuroscience*, 33(34):13686–94.
- [61] Shannon, R. V., Zeng, F. G., Kamath, V., Wygonski, J., and Ekelid,
 M. (1995). Speech recognition with primarily temporal cues. *Science*,
 270(5234):303–4.
- [62] Steube, N., Nowotny, M., Pilz, P. K. D., and Gaese, B. H. (2016). Dependence of the Startle Response on Temporal and Spectral Characteris-

- tics of Acoustic Modulatory Influences in Rats and Gerbils. Frontiers in
 Behavioral Neuroscience, 10:133.
- ⁷⁶⁰ [63] Suta, D., Rybalko, N., Shen, D.-W., Popelář, J., Poon, P. W. F., and
- Syka, J. (2015). Frequency discrimination in rats exposed to noise as
 juveniles. *Physiology & Behavior*, 144:60–5.
- [64] Swerdlow, N. R., Braff, D. L., and Geyer, M. A. (1999). Cross-species
 Studies of Sensorimotor Gating of the Startle Reflex. Annals of the New
 York Academy of Sciences, 877:202–16.
- [65] Swetter, B. J., Fitch, R. H., and Markus, E. J. (2010). Age-related
 decline in auditory plasticity: Experience dependent changes in gap detection as measured by prepulse inhibition in young and aged rats. *Behavioral Neuroscience*, 124(3):370–380.
- [66] Syka, J. (2002). Plastic Changes in the Central Auditory System After
 Hearing Loss, Restoration of Function, and During Learning. *Physiol Rev*,
 82(3):601–36.
- ⁷⁷³ [67] Syka, J. (2010). The Fischer 344 rat as a model of presbycusis. *Hear*⁷⁷⁴ *Res*, 264:70–8.
- [68] Valsamis, B. and Schmid, S. (2011). Habituation and Prepulse Inhibition of Acoustic Startle in Rodents. *Journal of Visualized Experiments*,
 (55):e3446.

- ⁷⁷⁸ [69] Varty, G., Hauger, R., and Geyer, M. (1998). Aging Effects on the
 ⁷⁷⁹ Startle Response and Startle Plasticity in Fischer F344 Rats. *Neurobiology*⁷⁸⁰ of Aging, 19(3):243-51.
- [70] Viana, L. M., O'Malley, J. T., Burgess, B. J., Jones, D. D., Oliveira,
 C. A. C. P., Santos, F., Merchant, S. N., Liberman, L. D., and Liberman, M. C. (2015). Cochlear neuropathy in human presbycusis: Confocal
 analysis of hidden hearing loss in post-mortem tissue. *Hearing Research*,
 327:78–88.
- [71] Viemeister, N. F. (1979). Temporal modulation transfer functions based
 upon modulation thresholds. *The Journal of the Acoustical Society of America*, 66(5):1364–80.
- [72] Walton, J. P. (2010). Timing is everything: Temporal processing deficits
 in the aged auditory brainstem. *Hearing Research*, 264:63–9.
- [73] Wojtczak, M., Nelson, P. C., Viemeister, N. F., and Carney, L. H. (2011).
 Forward Masking in the Amplitude-Modulation Domain for Tone Carriers:
 Psychophysical Results and Physiological Correlates. *Jaro-Journal of the*Association for Research in Otolaryngology, 12:361–373.
- [74] Xu, Q. and Gong, Q. (2014). Frequency difference beyond behavioral
 limen reflected by frequency following response of human auditory Brainstem. *BioMedical Engineering OnLine*, 13(1):114–27.

- ⁷⁹⁸ [75] Zeng, F. G., Nie, K., Stickney, G. S., Kong, Y. Y., Vongphoe, M., Bhar-
- ⁷⁹⁹ gave, A., Wei, C., and Cao, K. (2005). Speech recognition with amplitude
- and frequency modulations. Proc Natl Acad Sci U S A, 102(7):2293-8.

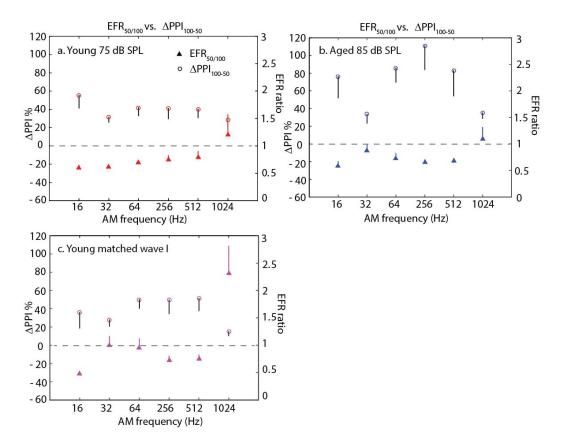


Figure 9: Greater changes of behavioral PPI values compared to changes of EFRs in aged animals when salience of AM depth reduced. Left ordinate indicates the measure of Δ PPI, which is the difference of PPI % at 100 % AM depth versus 50 % AM depth. Right ordinate indicates the measure of EFR ratio, which is the ratio of EFR amplitude at 50 % AM depth versus 100 % AM depth. The change in PPI value or EFR amplitude due to a change in AM depth was measured from the same animal in (a) young animals (75 dB SPL), (b) aged animals (85 dB SPL), and (c) young animals at equivalent peripheral activation. The paired changes were then averaged and the means of paired differences +/- SEM were plotted.