Title

Mapping nonapoptotic caspase activity with a transgenic reporter in mice

Authors

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Abstract

The protease caspase-3 is a key mediator of apoptotic programmed cell death. But weak or transient caspase activity can contribute to neuronal differentiation, axonal pathfinding, and synaptic long-term depression. Despite the importance of sublethal, or nonapoptotic, caspase activity in neurodevelopment and neural plasticity, there has been no simple method for mapping and quantifying nonapoptotic caspase activity in rodent brains. We therefore

generated a transgenic mouse expressing a highly sensitive and specific fluorescent reporter of caspase activity, with peak signal localized to the nucleus. Surprisingly, nonapoptotic caspase activity was prevalent in healthy adult brains and influenced neuronal functional connectivity. We quantified the relationship between caspase activity and cell firing and morphology. We also notably observed a sex-specific persistent elevation in amygdalar caspase activity in females after restraint stress. This simple *in vivo* caspase activity reporter will facilitate systems-level studies of apoptotic and nonapoptotic phenomena in behavioral and pathological models.

Significance Statement

Caspase-3 is an enzyme that can cause cell death when highly active but can also perform important cellular functions, such as maturation and structural changes, when only weakly or transiently active. Despite the importance of this nonlethal type of caspase activity, there is no straightforward method to measure it in live rodents. We therefore developed mice that have a fluorescent reporter that is sensitive enough to detect nonlethal caspase activity. Surprisingly, we found that weak caspase activity is widespread in healthy brains and influences the synchrony of neuronal firing across different brain regions. We also observed increased caspase activity in female mice after severe stress. This simple, live-animal caspase activity reporter can subserve multiple applications in behavior and pathology research.

Keywords

caspases; *in vivo* reporter; nonapoptotic; mapping; amygdala; stress; sex differences

Introduction

When caspase-3 is highly active, a proteolytic cascade ensues that typically ends in cell death by apoptosis (Yuan et al., 1993; Unsain & Barker, 2015). Weak or transient caspase-3 activity, however, is nonlethal and may be a key component of neurodevelopment by directing neuronal differentiation (Fernando et al., 2005), axonal pathfinding and branching (Westphal et al., 2010; Campbell & Okamoto, 2013), and synaptic metaplasticity (Chen et al., 2012). In the past several years mounting evidence indicates that, in the adult brain, nonapoptotic caspase activity (NACA) is an important contributor to neuronal plasticity, by driving NMDA receptor-mediated long-term depression (Li et al., 2010; Jiao & Li, 2011). Caspase activity in mature neurons may also underlie dendritic pruning (Ertürk et al., 2014). Uncovering the brain regions that display NACA at baseline and how NACA changes in response to perturbation will enable studies of its role in brain function.

Preclinical studies in cultured neurons (Du et al., 2004) and neonatal (Zhu et al., 2006; Renolleau et al., 2007), young adult, and aged rodents (Liu et al., 2009, 2011) have revealed sex-dependent differences in cell death pathways. In the context of hypoxia-ischemia, for example, cell death in females depends predominantly on caspase activity, while in males cell death occurs primarily by a caspase-independent mechanism (Zhu et al., 2006; Liu et al., 2009). This may be due to the more pronounced caspase-3 activation seen in females following

hypoxia-ischemia and other cytotoxic insults (Liu et al., 2009; Du et al., 2004). Notably, these sex-dependent differences do not rely on estrogen levels or developmental stage (Liu et al., 2009). Although these studies were concerned with apoptotic caspase activity, they motivated us to investigate how sex influences baseline and stress-responsive NACA.

There is currently no tractable method for mapping and quantifying weak and transient caspase activity at cellular resolution throughout the brain. Such a method would enable systems-level studies of the relationship between NACA, neural circuits, and various behavioral and pathological models. Available transgenic fluorescent reporters of weak caspase activity that do not require reagents are limited to Drosophila (Bardet et al., 2008; Tang et al., 2015). In mice, the available caspase reporters rely on substrates for bioluminescence (Fu et al., 2013; Galbán et al., 2013) or beta-galactosidase activity (Khanna et al., 2010) which cannot be used for high-resolution studies of fixed tissues. And a FRET-based reporter in mice does not detect NACA (Yamaguchi et al., 2011). It was our goal to generate and validate a reagent-free caspase reporter in mouse brain that would produce a strong nuclear signal with no background activity.

In this paper, we describe a novel transgenic reporter of caspase activity in mice. After demonstrating the simplicity of separating apoptotic from nonapoptotic reporter signal in several models of programmed cell death, we find, surprisingly, that nonapoptotic signal is widespread in healthy adult brains. Mapping the brain reveals distinct anatomical regions that display prominent NACA. Subsequently, we focus on neurons in the anterior basolateral

amygdala, due to this region's involvement in stress-related behavioral responses, and we find sex-dependent baseline and stress-induced differences in NACA. Within this population of neurons, we then determine the relationship between caspase activity and cell firing, morphology, and functional connectivity. Taken together, our results suggest that NACA may underlie pathologic responses to severe behavioral stress.

Results

We designed a bimolecular fluorescence complementation reporter of caspase activity that comprises four elements intended to maximize the signal-to-noise ratio: CAG promoters, split inteins, PEST sequences, and localization signals (Figure 1A). Bidirectional CAG promoters provide strong, ubiquitous expression (Niwa et al., 1991; Okabe et al., 1997) of each half of the mCerulean reporter (Rizzo et al., 2004). Split inteins further boost the signal by enabling covalent reconstitution of mCerulean from the two nonfluorescent halves (Wang et al., 2012). To eliminate background signal, these halves are kept separate and continually degraded. This baseline state holds until active caspases cleave off the localization signals (Güttler et al., 2010) and PEST sequences (Li et al., 1998) by cutting canonical caspase cleavage sites (McStay et al., 2008) present between the mCerulean, localization, and PEST sequences. The liberated, stabilized mCerulean halves can then join via inteins and fluoresce, typically near the nucleus. This reporter construct was introduced into the C57BL/6 mouse genome by pronuclear microinjection, yielding at least 24 copies in homozygotes according to qPCR. This line was bred

onto a mixed background to increase the size and frequency of litters. After more than six years of breeding, the copy number has remained stable in both mouse lines.

Apoptosis produces a strong signal *in vivo* and *in vitro*. Reporter signal is evident in shedding skin and between the toes of P0 pups (Figure 1B). *In vitro*, application of 100-nM staurosporine to primary hippocampal cultures caused a robust increase in signal that preceded cell death (Figure 1C). We induced apoptosis *in vivo* with middle cerebral artery occlusion (Beraldo et al., 2013) and found heightened cortical signal ipsilateral to the lesion (Figure 1D). We also caused *in vivo* apoptosis by injecting 6-hydroxydopamine into the medial forebrain bundle (He et al., 2000) and observed bright reporter signal and TUNEL-positive cells in the substantia nigra pars compacta (Figure 1E). Likewise, bright signal was observed in the cerebellum of one-month-old *Arpc3* knockout mice, which demonstrate ataxia and loss of Purkinje cells at this age (Figure 1F; Kim et al., 2013).

Due to the extreme brightness of reporter signal during apoptosis, we could measure it in the red portion of the spectrum (570-640 nm) even though mCerulean emits primarily in the cyan range (470-550 nm) (Figure 2A, Table S1; Rizzo et al., 2004). Thus, it is simple to separate apoptotic (570-640 nm) from nonapoptotic (470-550 nm only) signal (Figure 2B and Methods). We confirmed that the prevalent baseline signal (eg, Figure 2A, top row) was nonapoptotic since TUNEL stain was negative for the brightest cells (Figure 2C), despite being positive for shrunken epithelial cells in choroid plexus (Figure 2D). We also verified that this nonapoptotic signal represents caspase activity by blocking its increase in acute hippocampal slices with a

caspase inhibitor (Figure 2E; Caserta et al., 2003; Yang et al., 2004). Further confirmation that the reporter depends on caspase activity was seen in the substantial reduction in baseline signal in caspase-3 knockouts (Figure 2F), with residual signal likely due to caspase-7.

We further characterized the nonapoptotic signal, which we henceforward will term NACA, in whole-brain scans using different imaging modalities (see Table S1). NACA was widespread and especially prominent in the amygdala, hippocampus, cortex, and Purkinje cells (Figure 3A-C). At higher resolution, we determined NACA was present in principal cells in the amygdala (Figure 3D), based on colocalization with the caspase target and neuronal marker myocyte enhancer factor 2 (Chen et al., 2012; Neely et al., 2009); and, in the hippocampus, NACA was mostly limited to neurons in the pyramidal layer (Figure 3E). Therefore, bringing the above findings together, we have generated a transgenic reporter of caspase activity that can differentiate apoptotic from nonapoptotic activity for neuronal mapping studies throughout the brain.

We focused our investigations on the aBLA because of its prominent NACA and its importance in behavioral stress (Roozendaal et al., 2009; Kim et al., 2016, 2017). There was a nonlinear relationship between NACA intensity and the intensity of the neuronal activity reporter cFos (Morgan et al., 1987; Dragunow & Robertson, 1987): as NACA intensity increases, there is a substantial reduction in cFos followed, intriguingly, by a smaller increase (Figure 4A, B). Also interesting, the relationship between NACA intensity and dendrite length appears to be linear and direct (Figure 4C-E), although this is based on limited evidence from sparse labeling of neurons. Since NACA correlated with key aspects of neuronal function and morphology, we

tested whether NACA was related to stress responses in the amygdala. We first noted that there was a baseline difference in NACA levels between males and females (Figure 4F) and therefore studied both sexes concurrently in a recently reported stress paradigm (Chu et al., 2016). Nine days after a 24-hour bout of restraint stress, we unexpectedly found a persistent increase in aBLA NACA only in females (Figure 4G). Furthermore, the mean NACA intensities in females (Figure 4G, right) corresponded to an increase in neuronal activity (Figure 4B) and a presumptive increase in dendritic length (Figure 4E).

Since aBLA NACA levels correlate with neuronal activity, morphology, and responses to stress, we tested the causal relationship between NACA and neuronal function by reducing baseline NACA in unstressed mice with a caspase inhibitor. We first determined that the inhibitor Q-VD-OPh reaches IC50 levels (Caserta et al., 2003) in brain when administered intraperitoneally at 5 mg/kg (Figure 5A) and shows no off-target activity at potential psychoactive targets at these concentrations (Figure 5B). Comparing multielectrode recordings of local field potentials (Dzirasa et al., 2013) in the basal amygdala, dorsal hippocampus, and prelimbic cortex 24 hours before treatment and 24 hours after treatment (Figure 5C-E), we detected a reduction in 15-30-Hz functional connectivity between the amygdala and prelimbic cortex (Figure 5F, G) and a specific reduction in 15-30-Hz amplitude in the amygdala (Figure 5H, I). Intriguingly, this reduction in amplitude after a caspase inhibitor appears to be consistent with an increase in neuronal firing and NACA after stress in females (Figure 4B, G). Hence, aBLA NACA may play an important role in neuronal function, morphology, and stress responsiveness, especially in females.

Discussion

We designed a transgenic fluorescent reporter sensitive and specific enough to detect NACA as well as caspase-mediated apoptosis in mouse brain. Due to the logarithmic increase in caspase activity during apoptosis (Li et al., 2010), it was possible to separate long-wavelength apoptotic signal from short-wavelength nonapoptotic signal. Focusing on nonapoptotic signal, NACA was relatively elevated in specific regions – including BLA, dorsal hippocampus, and medial frontal cortex – and specific cell types – most notably, excitatory neurons and Purkinje cells. We further characterized NACA in the aBLA by relating it to cell firing, morphology, and response to stress. There was a nonlinear relationship between NACA and levels of cFos, a widely-used proxy for neuronal activity (Morgan et al., 1987; Dragunow & Robertson, 1987): cFos levels decreased robustly with weak caspase activity but approached baseline levels with higher caspase activity. The association between NACA and cell morphology was apparently linear, with total dendrite length increasing with caspase activity. And we uncovered a sex-specific effect of stress on NACA: 9 days after prolonged restraint stress, NACA was elevated only in female mice. Finally, we confirmed there was a causal relationship between baseline NACA and neural activity by measuring functional connectivity in unstressed mice treated with a caspase inhibitor. Caspase inhibition caused a reduction in functional connectivity between the basal amygdala and prelimbic cortex, along with reduced 15-30-Hz power in the amygdala. Together these results demonstrate the utility of our caspase reporter mouse and the potential importance of NACA in studies of neural function and stress.

Our primary aim was to design a transgenic reporter that would enable mapping of all caspase activity across the brain using multiphoton and other forms of microscopy. Consequently, our reporter contains several design elements subserving this goal that in other contexts might constitute limitations or shortcomings. (1) There is no internal control, such as a contrasting, constitutively expressed fluorescent protein (Yamaguchi et al., 2011), because multiphoton absorption spectra typically overlap (Drobizhev et al., 2011), and any subsequent emission overlap from a bright control would impair the potentially weak signal of the reporter. (2) The reporter does not indicate the subcellular location of caspase activity, since, to facilitate mapping and cellwise quantification, we wanted peak signal to center on the nucleus regardless of the source of caspase activity. (3) Reporter signal is not specifically targeted to the nucleus because fluorescence in the soma can demonstrate shrinkage during apoptosis. (4) The cyan fluorophore overlaps the autofluorescence profile of neural tissue (Billinton & Knight, 2001). This design element enables crossing to other reporter strains, such as calcium sensor mice, which utilize green or red fluorescent proteins. In addition, the tail-end red emission of our mCerulean reporter during apoptosis can be detected by multiphoton microscopy, unlike the near-infrared tail-end emission of a red fluorophore, thereby facilitating separation of nonapoptotic from apoptotic signal. (5) The reconstituted reporter has a long half-life – probably >1 day (Corish & Tyler-Smith, 1999) – since a destabilized fluorescent protein would reduce our chances of detecting transient, weak caspase activity. (6) Our system is not controlled by Cre or other recombinases because this would limit genomic copy number to one, or perhaps two, which would constrain reporter signal output and make detection of weak

caspase activity less likely. (7) Likewise, the insertion site of our reporter construct is unknown, since we generated the mouse by pronuclear microinjection. But this random insertion enabled us to reach a homozygous copy number of at least 24, according to quantitative PCR, which has been stable for over six years of breeding. (8) Lastly, with such a sensitive reporter of caspase activity, tissue fixation must be performed with great care to maintain consistency of NACA signal and to avoid artifacts like “dark neurons” (Beevor, 1883; Cammermeyer, 1960; Jortner, 2006). In order to obtain reproducible results, we strongly recommend following the fixation procedure described in the Methods.

When we applied our careful sample preparation and microscopy techniques to the study of stress, we found a sex-specific persistent increase in NACA in the aBLA in females. Although surprising, if we suppose that caspases play a meaningful role in stress responsiveness, this result is not entirely unexpected. As mentioned above, the caspase pathway is engaged in apoptotic cell death in female brain, but a parallel cascade predominates in male brain (Zhu et al., 2006; Liu et al., 2009). Also, long-lived physiologic changes after stress tend to be confined to the amygdala. Perhaps the most widely reported alteration in the amygdala after stress is persistent dendritic hypertrophy (Roozendaal et al., 2009). This is consistent with our findings, since we saw a direct correlation between caspase activity and dendrite length in the aBLA. Furthermore, neurons in the aBLA fire more after stress and drive negative-valence defensive behaviors (Kim et al., 2016, 2017). This also is consistent with our results, since the increase in female aBLA NACA after stress corresponds to a subtle increase in cFos. Hence, caspase activity

may help explain alterations in neuronal morphology and activity, and, at least in females, the peculiar structural and functional responses to stress in the amygdala.

But are these structural and functional changes an adaptive coping mechanism or a pathologic reaction to severe stress? Since aBLA NACA, and presumably dendrite length and cFos, remained elevated 9 days after a single, prolonged bout of restraint stress; and because aBLA neurons drive anti-appetitive, pro-defensive behaviors (Kim et al., 2016, 2017); our results suggest there is a long-lived, negative-valence reaction to stress in females that is mediated by caspase activity. If there is no corresponding reaction in males by some other molecular pathway under these conditions, then caspase activity may help explain the increased prevalence among women of stress-related disorders, such as depression, anxiety, and posttraumatic stress disorder (PTSD) (Hasin et al., 2005; Seedat et al., 2009; Pedersen et al., 2014). For example, the lifetime risk of PTSD is fourfold higher in women, after controlling for the type of traumatic stress (Kessler et al., 1995). If caspases contribute to stress-related disorders, would caspase inhibitors have therapeutic value? We provide preliminary evidence that a caspase inhibitor can alter baseline NACA and thereby reduce 15-30-Hz amygdala-prelimbic functional connectivity and amygdala power. Of note, these neurophysiological changes are similar to what was observed when a mouse model of depression was treated chronically with fluoxetine (Dzirasa et al., 2013). Therefore, we propose that future studies address the causality of caspase activity in pathologic reactions to stress and the potential for caspase inhibitors in the treatment of stress-related disorders.

In conclusion, we have developed a transgenic reporter of caspase activity in mice, as well as tissue fixation and microscopy methods that enable the mapping and quantification of weak and transient nonapoptotic caspase activity. We utilized our system to demonstrate correlations between caspase activity and neuronal morphology and activity, and to uncover a sex-dependent effect of stress on the anterior basolateral amygdala. Our reporter and techniques can be used to test hypotheses relating to apoptotic and nonapoptotic caspase activity throughout the brain and the rest of the organism.

STAR Methods

Detailed methods are provided in the online version of this paper and include the following:

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Supporting Information

Supporting Information includes one table, one software resource, and one data resource and can be found with this article online at

Author Contributions

PJN, TFP, NMU, SK, GT, VFP, MAMP, IS, KD, GAJ, and MGC designed research. PJN, TFP, NMU, SK, GT, PF, VGO, MSG, FB, and IS performed research. PJN, TFP, WLR, IS, and JCS contributed new reagents. PJN, EC, JCS, and KD contributed analytic tools. PJN, SK, PF, VGO, IS, and KD analyzed data. PJN wrote the paper.

Acknowledgments

We are grateful to the following for their assistance and advice: Rainbo Hultman, Benjamin Sachs, Lauren Rochelle, Daniel Urban, Scott Robertson, Jue Fan, Nicole Calakos, Fan Wang, David Piston, Martin Fischer, Francisco Robles Guerrero, Ramona Rodriguiz, William Wetsel, Cheryl Bock, Meilang Flowers, Thomas L. Brown, Lauren Burianek, and Scott Soderling. Ki determinations were provided by the NIMH's Psychoactive Drug Screening Program, Contract #HHSN-271-2013-00017-C. Pharmacokinetic analysis was carried out by the Duke Cancer Institute PK/PD Core Laboratory. This work was supported by NIH grants: 1R21NS081513 (PJN), 8P41EB015897 (GAJ), 2R01MH079201 and 5R37MH073853 (MGC); and CIHR grants: MOP 93651, 12600, 89919 (MAMP). Support from the Pall Family Foundation and Lennon Family Foundation to MGC is greatly appreciated. PJN, NMU, GAJ, and MGC have filed a provisional method-of-use patent application relating to this research.

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Figures

Figure 1

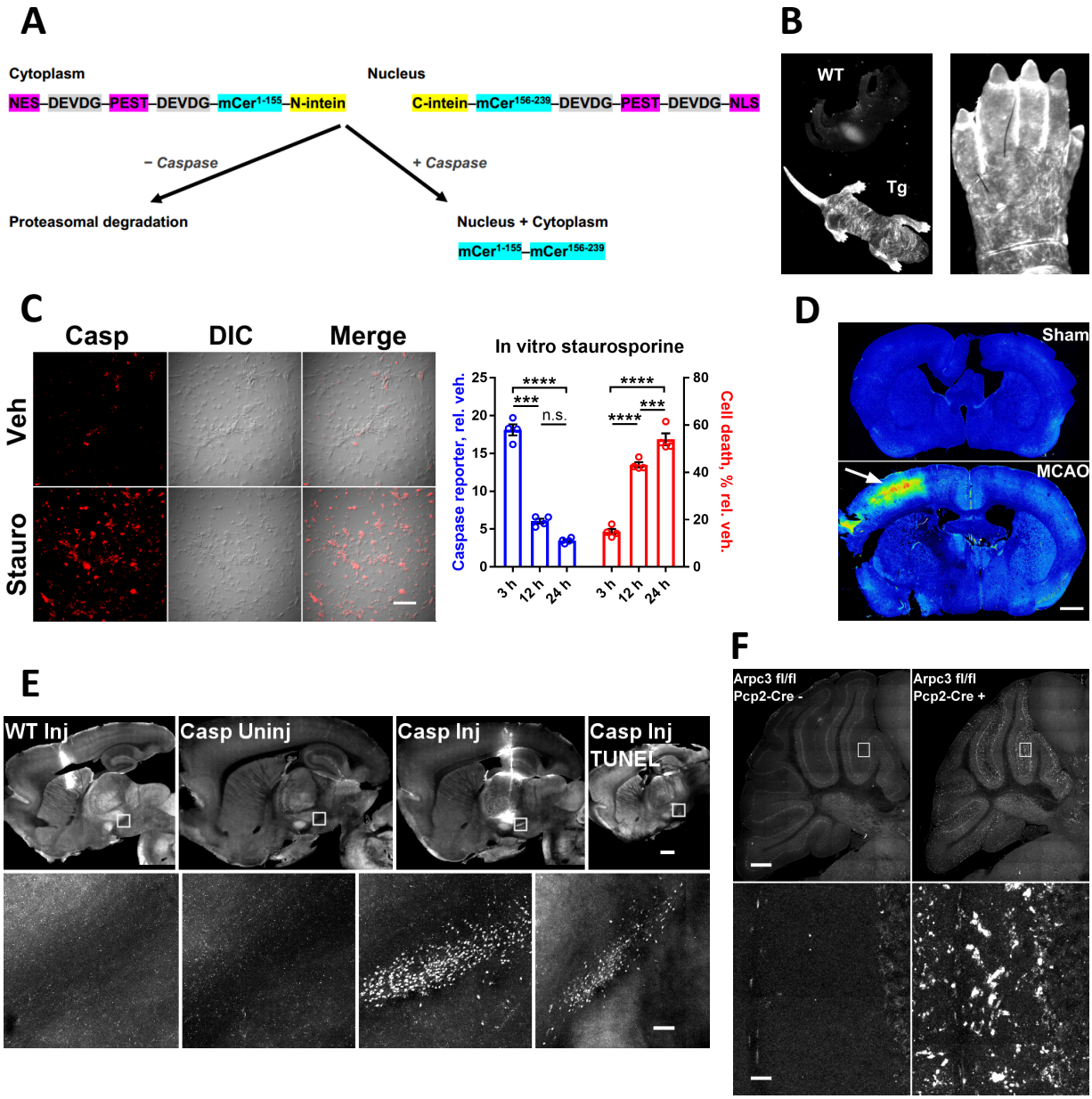


Figure 1. A transgenic reporter of apoptotic caspase activity.

(A) Schematic of caspase- and intein-mediated bimolecular fluorescence complementation. From left: NES, nuclear export sequence; DEVDG, caspase cleavage site; PEST, protein degradation sequence; mCer, mCerulean fluorophore; N-, C-intein, amino- and carboxy-terminal bacterial inteins; NLS, nuclear localization sequence.

(B) Caspase reporter signal is present in shedding skin of a transgenic P0 mouse (left, Tg) and in apoptotic webbing between its toes (right). Weak signal in wild-type (WT) P0 mouse is autofluorescence in the stomach.

(C) Representative data (left) and time course (right) of caspase reporter intensity and cell death in primary hippocampal neurons treated with 100-nM staurosporine. Two-way repeated-measures ANOVA with Tukey's multiple comparisons test (Reporter x Time $F(2, 12)=216.7$); mean $\pm$ SEM; n=4. n.s., not significant; ***, $p<0.001$; ****, $p<0.0001$. Scale bar, 100 µm.

(D) Caspase reporter distribution after middle cerebral artery occlusion (MCAO, arrow) versus sham. Red-blue is high-low signal intensity. Scale bar, 1 mm.

(E) Apoptotic reporter signal in the substantia nigra pars compacta ipsilateral to a 6-hydroxydopamine injection (Casp Inj), but not contralateral to the injection (Casp Uninj) or in injected wild-type (WT Inj), and confirmed by TUNEL stain (Casp Inj TUNEL). Scale bar, 1 mm (above), 100 µm (below).

(F) Apoptotic reporter signal in the cerebellar molecular layer of *Arpc3* conditional knockout mice, with and without Cre recombination. Scale bar, 400 µm (above), 25 µm (below).

See also Table S1.

Figure 2

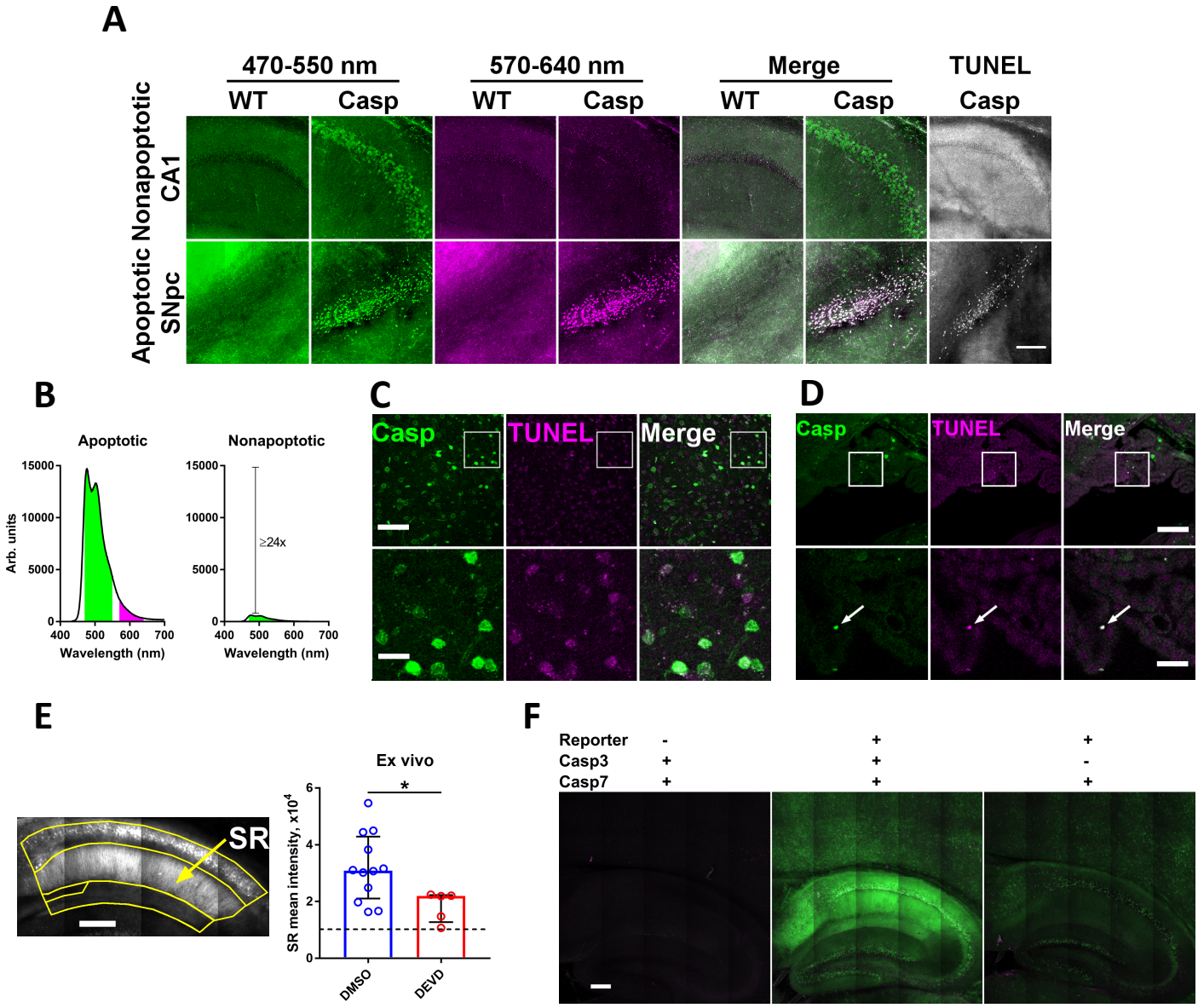


Figure 2. Separation of apoptotic and nonapoptotic reporter signal.

(A) Separation of nonapoptotic (above) from apoptotic (below) reporter signal by comparing short- (green) and long-wavelength (magenta) emission. Data are the same experiment as Figure 1E. Scale bar, 200 µm.

(B) Quantitative comparison of apoptotic and nonapoptotic reporter signal in short- (green) and long-wavelength (magenta) channels, derived from data used for Figure 4G (see Methods).

(C) Lack of colocalization between nonapoptotic reporter signal (green) and TUNEL stain (magenta) in the basolateral amygdala. Scale bar, 100 µm (above), 25 µm (below).

(D) Colocalization of reporter signal (green) with TUNEL stain (magenta) in apoptotic epithelial cells of the choroid plexus. Scale bar, 100 µm (above), 25 µm (below).

(E) Attenuation of reporter signal by a caspase inhibitor (3.7-µM Z-DEVD-FMK) in acute hippocampal slices. SR, stratum radiatum. Dashed line denotes threshold of detection. Mann-Whitney test (U=9); median ± IQR; n=12 DMSO, 5 DEVD. *, $p<0.05$. Scale bar, 200 µm.

(F) Comparison of representative reporter signal in acute hippocampal slices across 3 genotypes. Scale bar, 200 µm.

See also Table S1.

Figure 3

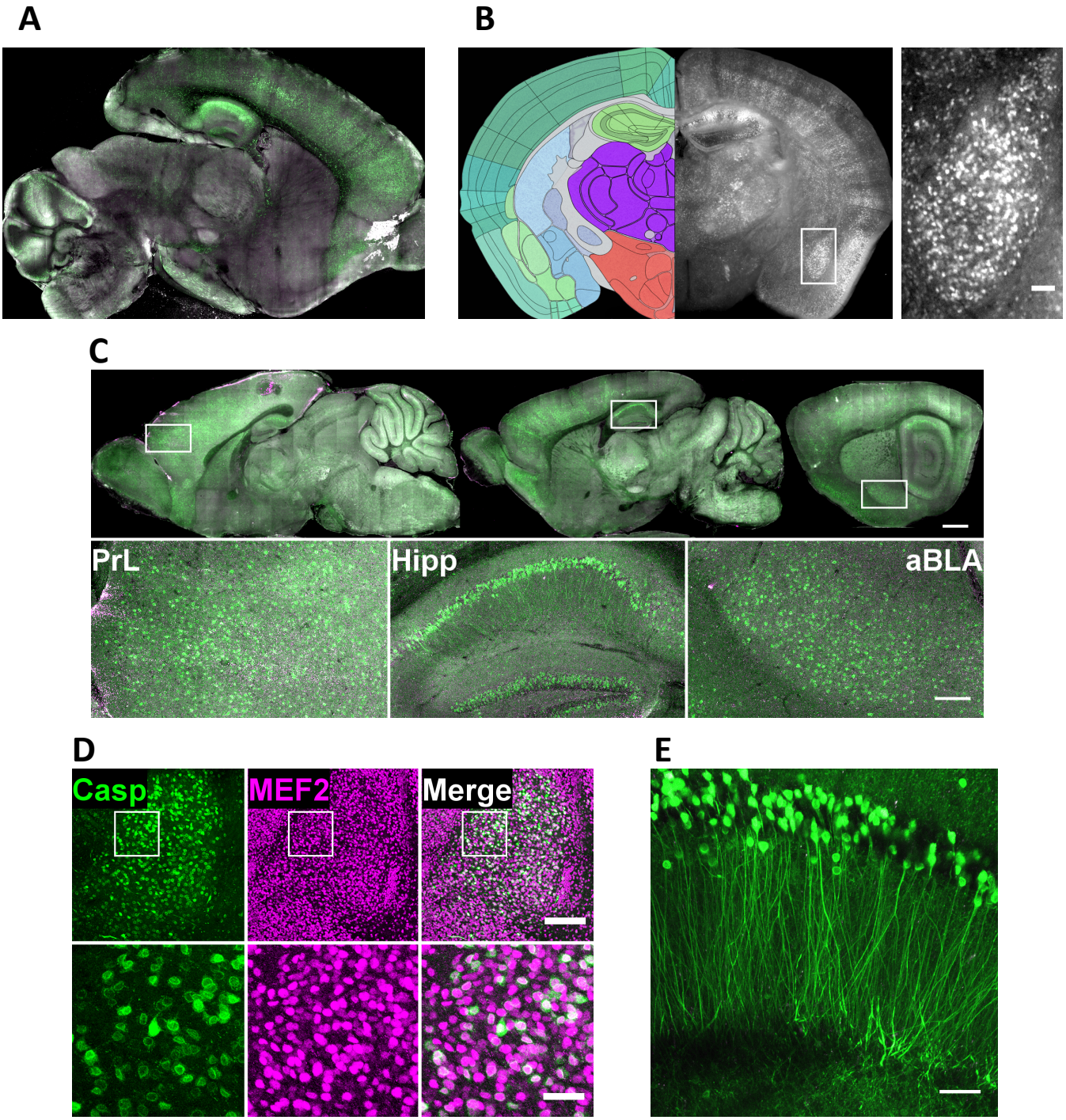


Figure 3. Regional and cellular location of nonapoptotic reporter signal.

(A) Nonapoptotic reporter signal (green) in a clarified sagittal section of P22 brain.

(B) Nonapoptotic reporter signal in a coronal section of P44 brain (right). Left atlas tracing image credit: Allen Institute (Lein et al., 2007). Inset: anterior basolateral amygdala. Scale bar, 100 µm.

(C) Nonapoptotic signal (green) is elevated in prelimbic cortex (PrL), hippocampus (Hipp), and anterior basolateral amygdala (aBLA). Scale bar, 1 mm (above), 200 µm (below).

(D) Colocalization of reporter signal (green) with the neuronal marker myocyte enhancer factor 2 (magenta) in basolateral amygdala. Scale bar, 200 µm (above), 50 µm (below).

(E) Hippocampal nonapoptotic reporter signal is predominantly in neurons of the stratum pyramidale. Scale bar, 50 µm.

See also Table S1.

Figure 4

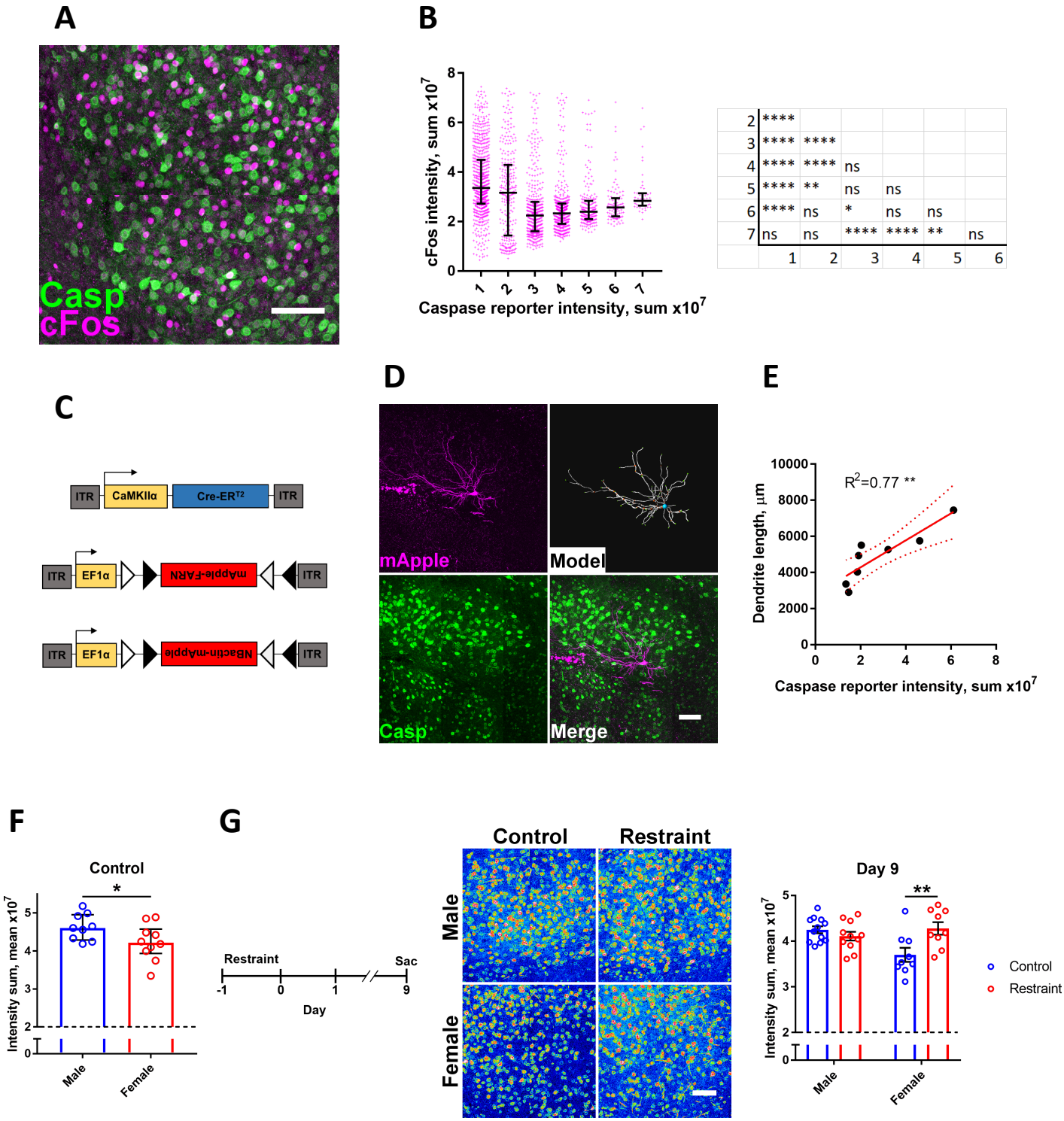


Figure 4. Functional and structural correlates of NACA.

(A, B) Representative data (A) of a nonlinear relationship (B) between NACA (green) and the neural activity marker cFos (magenta). (B, right) Stats table of all caspase reporter intensity bin comparisons. Kruskal-Wallis with Dunn's multiple comparisons test (K-W=404); median ± IQR; n=2251 cells from 5 amygdalae. Scale bar, 100 µm.

(C) Schemata of viral constructs for the sparse labeling of excitatory neurons. FARN, farnesylation sequence; NBactin, Actin-Chromobody®; triangles, loxP and lox2272.

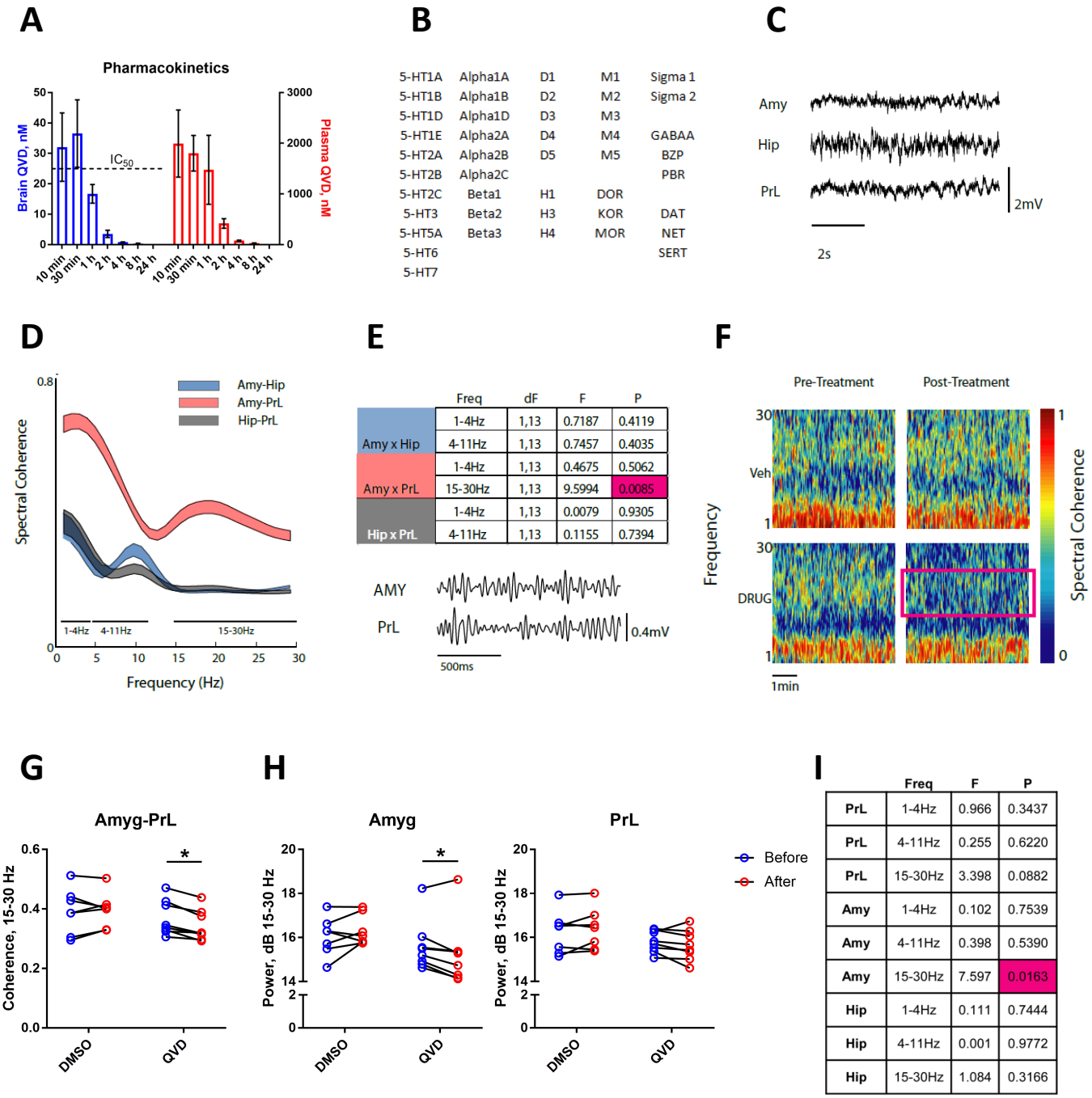
(D, E) Direct correlation between NACA (green) and dendrite length (magenta, modeled in gray). Linear regression (solid line) ± 95% CI (dotted lines) (F(1, 6)=20.25); n=8 cells from 6 amygdalae. Scale bar, 100 µm.

(F) Reduced baseline NACA in female mice. Dashed line represents threshold of detection. Mann-Whitney test (U=20); median ± IQR; n=9 M, 10 F.

(G) Left: Time course of the experiment. Middle: Representative caspase reporter intensity data (red-blue, high-low intensity). Right: Sex-specific increase in NACA 9 days after restraint stress. Dashed line denotes threshold of detection. Two-way ANOVA with Sidak's multiple comparisons test (Sex x Stress F(1, 37)=9.931); mean ± SEM; n=12 M Cont., 11 M Rest., 9 F Cont., 9 F Rest from 2 independent experiments. Scale bar, 100 µm.

ns, not significant; *, $p<0.05$; **, $p<0.01$; ****, $p<0.0001$. See also Table S1.

Figure 5



	Freq	dF	F	P
Amy x Hip	1-4Hz	1,13	0.7187	0.4119
	4-11Hz	1,13	0.7457	0.4035
Amy x PrL	1-4Hz	1,13	0.4675	0.5062
	15-30Hz	1,13	9.5994	0.0085
Hip x PrL	1-4Hz	1,13	0.0079	0.9305
	4-11Hz	1,13	0.1155	0.7394

	Freq	F	P
PrL	1-4Hz	0.966	0.3437
PrL	4-11Hz	0.255	0.6220
PrL	15-30Hz	3.398	0.0882
Amy	1-4Hz	0.102	0.7539
Amy	4-11Hz	0.398	0.5390
Amy	15-30Hz	7.597	0.0163
Hip	1-4Hz	0.111	0.7444
Hip	4-11Hz	0.001	0.9772
Hip	15-30Hz	1.084	0.3166

Figure 5. NACA contributes to functional connectivity in a stress-associated circuit.

(A) Pharmacokinetics of 5 mg/kg Q-VD-OPh in mouse brain and plasma after intraperitoneal injection. Mean ± SEM; 3-5 mice per time point.

(B) Receptors and transporters screened for interaction with Q-VD-OPh.

(C) Representative traces of local field potentials recorded from amygdala (Amy), hippocampus (Hip), and prelimbic cortex (PrL) in a freely behaving mouse.

(D) Spectral coherence between three area pairs (Amy-Hip, Amy-PrL, and Hip-PrL). Data are presented as 95% confidence intervals; 15 mice.

(E) Table of spectral coherence differences across functional connectivity bands depicted in (D), and representative β (15-30 Hz) frequency traces. Two-way repeated-measures ANOVA with Box-Cox transform; magenta highlights significant value.

(F) Representative spectral coherence data between basal amygdala and prelimbic cortex. Magenta box shows decreased coherence after caspase inhibitor treatment.

(G) Reduced functional connectivity between basal amygdala and prelimbic cortex after caspase inhibitor treatment (QVD). Two-way repeated-measures ANOVA with Sidak's multiple comparisons test (Drug x Time $F(1, 13)=6.594$).

(H) Decreased local field potential power after QVD treatment in basal amygdala (left), but not in prelimbic cortex (right). Two-way repeated-measures ANOVA with uncorrected Fisher's LSD (Drug x Time: left $F(1, 13)=6.811$, right $F(1, 13)=3.343$).

(I) Table of spectral power differences. Two-way repeated-measures ANOVA with Box-Cox transform; magenta highlights significant value.

In (D, E, G-I), n=7 DMSO, 8 QVD; *, $p<0.05$.

Supporting Information

Table S1

Figure	Specimen	Pixel size (µm)	z interval (µm)	Slices	Projection	Channels	Excitation (nm)	Emission (nm)	System
1B left	In vivo	9.29	0	1	N/A	Single	424-448	460-500	Mercury arc lamp, 1x/0.25NA
1B right	In vivo	9.29	0	1	N/A	Single	424-448	460-500	Mercury arc lamp, 1x/0.25NA
1C	In vitro	0.58	0	1	N/A	Single, merged	405 (red), 488 (gray)	430-470 (red), transmitted light (gray)	Diode laser (red), Ar laser (gray), 10x/0.4NA
1D	Fixed	2.49	0	1	N/A	Single	458	480-495	Ar laser, 10x/0.4NA
1E left 3	Clarified	3.29	20	51	Maximum	Single	858	570-640	Ti:sapphire laser, 20x/1.0NA
1E TUNEL	Declarified	3.29	20	11	Maximum	Single	1000	570-640	Ti:sapphire laser, 20x/1.0NA
1F	Fixed	0.82	10	21	Maximum	Single	858	570-640	Ti:sapphire laser, 20x/1.0NA
2A left 6	Clarified	3.29	20	51	Maximum	Single, merged	858	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA
2A TUNEL	Declarified	3.29	20	11	Maximum	Merged	1000	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA
2C	Fixed	0.82	5	41	Maximum	Single, merged	858	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA
2D	Fixed	0.82	5	41	Maximum	Single, merged	858	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA
2E	Ex vivo	0.82	10	15	Sum	Subtracted	858	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA
2F	Ex vivo	0.82	10	15	Sum	Merged	858	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA
3A	Clarified	3.29	20	51	Maximum	Merged	858	460-500, 520-550	Ti:sapphire laser, 20x/1.0NA
3B	Fixed	2.27	0	1	N/A	Single	450-490	500-550	Mercury arc lamp, 2.3x/0.57NA
3C	Fixed	3.29	20	14	Maximum	Merged	858	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA
3D	Fixed	0.82	5	51	Maximum	Single, merged	858	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA
3E	Ex vivo	0.2	2	21	Maximum	Merged	858	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA
4A	Fixed	0.55	5	51	Maximum	Merged	858	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA
4D	Fixed	0.55	5	51	Maximum	Subtracted, merged	858 (green), 1000 (magenta)	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA
4G	Fixed	0.55	5	51	Maximum	Subtracted	858	470-550, 570-640	Ti:sapphire laser, 20x/1.0NA

Table S1. Imaging parameters. Related to Figures 1-4.

STAR Methods

CONTACT FOR REAGENT AND RESOURCE SHARING

Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, PJ Nicholls (peter.nicholls@duke.edu).

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Mice

All procedures involving animals were carried out in accordance with National Institutes of Health standards as approved by the Duke University Institutional Animal Care and Use Committee. The caspase activity reporter transgene (Data S2) was synthesized and inserted into pCAGEN by GenScript. Pronuclear microinjection of the transgene into C57BL/6NHsd one-cell embryos was performed by Duke's Transgenic Mouse Facility. Reporter mice used in experiments were male and female, primarily 8-10 weeks old but ranging from P0 to 4 months old, homozygous on a C57BL/6 (Figures 1B-E, 2A, E, F, 3A, C, E) or mixed background (Figures 2C, D, 3B, D, 4A-G), and drug- and test-naïve. The mice in Figure 1F were P46 male caspase reporter hemizygous, floxed *Arpc3* homozygous, and *PCP2-Cre* wild-type or hemizygous. Caspase-3 knockout mice for Figure 2F were C57BL/6 congenic and bred to homozygosity for the caspase activity reporter. Finally, mice for the neurophysiology (Figure 5C-I) and

pharmacokinetics (Figure 5A) experiments were 2-3-month-old male C57BL/6J from Jackson Labs. Mice were housed in a barrier facility on a 12/12 light cycle with species-specific heat and humidity and fed a standard laboratory diet with food and water *ad libitum*. Mice were group housed 2-5 per micro-isolator cage on corn cob bedding, and cages were changed every two weeks. For the *ex vivo* (Figure 2E, F), neurophysiology, and pharmacokinetics experiments, cagemates were allocated randomly to experimental groups. For the aBLA NACA quantification in Figure 4F, male and female cages were alternated in the order of perfusion; the first half of samples perfused were randomized separately from the second half of samples perfused; the randomization was double-blind; and the first half of samples were imaged in their new order followed by the second half in their new order. For the aBLA NACA quantification in Figure 4G, alternate mice within a cage were restrained; the order of perfusion alternated between control and restraint within cages and male and female between cages; and the randomization and order of imaging were as in Figure 4F.

Primary Cultures (Figure 1C)

Primary cultures of hippocampal neurons (Figure 1C) were obtained from caspase reporter pregnant females on day E17 as previously described (Beraldo et al., 2010, 2013). Briefly, mouse hippocampal neurons of both sexes were harvested at day 17 of embryonic development (E17) and maintained in dishes coated with poly-L-lysine (Sigma-Aldrich, Canada) for 7 days in Neurobasal medium (Invitrogen) with 2% B-27 supplement (Invitrogen). Half of the medium was replaced every 2-3 days. All procedures were conducted in accordance with

approved animal use protocols at the University of Western Ontario (2016/103) following Canadian Council of Animal Care and National Institutes of Health guidelines.

METHOD DETAILS

Real-time PCR

Quantitative PCR for genotyping was performed with SYBR Select Master Mix (Life Technologies), 400-nM primers (synthesized by Integrated DNA Technologies), and a LightCycler (Roche). Later genotyping was performed on a StepOnePlus system (Applied Biosystems) with PowerUp SYBR Green Master Mix (Thermo Fisher Scientific) and 500-nM primers (IDT).

Microscopy

We used a Zeiss AxioZoom for Figures 1B, 3B; an Olympus confocal FV1000 for Figures 1C, D; and a Zeiss LSM 510 with an automated stage, Coherent Chameleon laser, and halogen lamp for all other microscopy. In two-color images, green represents shorter wavelengths and magenta longer wavelengths, as detailed in Table S1. Images were processed in ImageJ. For subtracted images, the magenta channel was linearly subtracted from the green channel, to remove autofluorescence and apoptotic reporter signal. Only linear window-level adjustments were made to images; and images within a figure panel were adjusted as a single image, except for

the TUNEL stain in Figures 1E, 2A. In Figures 1E, 2A, 3A, C, a linear unbiased MATLAB script (Data S1) was used to correct for a stable shading abnormality in our multiphoton system. Clarification was performed as previously described (Hama et al., 2011).

Hippocampal Cultures (Figure 1C)

On DIV 7, hippocampal neurons were treated with either vehicle or staurosporine (100 nM). Cells were imaged at 3, 12 and 24 hours after treatment, and caspase-3 activity was measured using fluorescence from the mCerulean reporter. Fluorescence of neuronal cultures from 4 independent embryos of either sex was quantified using an FV1000 Olympus confocal microscope (excitation 405 nm and emission 470-485 nm). Integrated fluorescence was divided by total cell number and normalized to control cells that were not subjected to staurosporine, using ImageJ software (NIH, Bethesda, MD).

Middle Cerebral Artery Occlusion (Figure 1D)

Three- to 4-month-old male caspase reporter homozygotes were subjected to 2-hour middle cerebral artery occlusion (MCAO) or were sham-operated. After 24-hour recovery, mice were perfused with 4% PFA in PBS. Perfused brains were incubated overnight at 4 C, after which 50-µm-thick coronal sections were prepared using a vibratome. Slices were immediately mounted with PBS on glass slides and imaged using an Olympus confocal FV1000 equipped with a 10x/0.4NA objective.

6-Hydroxydopamine Microinjections (Figures 1E, 2A)

In mice under constant isoflurane anesthesia, 1 µl of 6-hydroxydopamine (3 mg/ml) dissolved in saline containing 0.02% ascorbic acid was injected at coordinates AP=−1.2, ML=−1.2, and DV=−4.75 with a Hamilton syringe fitted with a pulled glass capillary needle at 0.4 µl/min using a Nanojet syringe pump. Fourteen hours after injection, mice were transcardially perfused with 4% PFA, and brains were fixed overnight in 4% PFA at 4 C.

Tissue Stains (Figures 2, 3)

1:500 Anti-MEF2D (AB2263, Millipore) and 1:500 Alexa-594 (Life Technologies), both overnight at 4 C, were used for immunohistochemistry in Figure 3D. The ApopTag TUNEL stain kit (Millipore) was used for Figures 1E, 2A, C, D.

Apoptotic vs. Nonapoptotic Signal Intensity (Figure 2B)

Based on the Cerulean emission spectrum available at

http://tsienlab.ucsd.edu/Documents.htm

and our observation that the 16-bit intensity mean of apoptotic reporter signal is 65535 in the 570-640-nm channel (0.06133 of total spectrum), and our calculation of 34555.6 for the mean

16-bit intensity mean in the 470-550-nm channel (0.7745 of total spectrum) for nonapoptotic signal in the 23 samples of one replicate of the experiment in Figure 4G, we found

(65535/0.06133)/(34555.6/0.7745) = 23.9

And since this value is limited by the maximal 16-bit intensity mean of 65535, the true quotient of apoptotic to nonapoptotic signal intensity would be greater than or equal to 24.

Acute Hippocampal Specimens (Figure 2E, F)

For *ex vivo* specimens, 3-4-week-old mice were anesthetized at time 0 and transcardially perfused with chilled oxygenated artificial CSF (in mM: NaCl 119, KCl 2.5, $NaHCO_3$ 26, NaH_2PO_4 1.25, glucose 11, $CaCl_2$ 0.5, $MgSO_4$ 1.3); then a vibratome-cut hemisphere was maintained in oxygenated artificial CSF with $CaCl_2$ 2 mM during multiphoton scanning. Z-DEVD-FMK was purchased from Millipore.

Measuring aBLA NACA (Figure 4)

For the stress intervention (Figure 4G), ~8-week-old male and female caspase reporter homozygotes housed 2-5 per cage were transferred to a nearby experimental procedure room within their barrier facility, where they acclimated overnight. At approximately 07:00 the next day, half of the mice were randomly selected and inserted into 50-ml tubes that had 0.25-in holes for breathing. For cages with an odd number of mice, selection was designed so that the total number of mice in each of the 4 experimental groups would be nearly equivalent.

Immediately after a mouse was placed in a tube, it was returned to its cage where it stayed with its restrained and unrestrained cagemates. All mice were deprived of food and water for the next 24 hours. Mice were observed, and tubes were rolled periodically to assess responsiveness. No mice became unresponsive during the test. Twenty-four hours later, restrained mice were released from their tubes, washed briefly in a room-temperature water bath, and returned to their cages with their unrestrained cagemates. For the next 9 days, all mice had access to food and water *ad libitum*. The released mice were again observed for distress or unresponsiveness, which were not detected during this test. Within a few hours of being released, restrained mice were indistinguishable from unrestrained mice upon gross inspection. After 9 days, all mice were transferred to a separate facility for transcardial perfusion.

Mice were perfused in order, such that restrained mice alternated with unrestrained mice and male cages alternated with female cages. Immediately after being weighed, each mouse was administered a mixture of ketamine 100 mg/kg, xylazine 10 mg/kg, and heparin 5000 U/kg in saline and given 10-15 minutes to attain a surgical plane of anesthesia. Mice were observed and tested by toe pinch for inadequate anesthesia, which was not noted in this experiment. No mice were given an additional dose of anesthetic. After 10-15 minutes of anesthesia, each mouse was pinned to a board in a chemical fume hood and rapidly dissected to reveal the liver and heart. Then the apex of the left ventricle was pierced with a 25-gauge needle attached to a 25-ml syringe containing 25 ml of 1:1 10% neutral buffered formalin : PBS plus 100 µl of 1000 U/ml heparin. Immediately, the solution was forced into the circulation at a moderate pace,

and the right atrium was snipped as it expanded. All 25 ml of formalin/PBS/heparin was forced through the circulation at a steady pace within 1-2 minutes. Each mouse was assessed for paleness of liver and stiffness of body; 1 mouse out of the 42 used in the stress experiment was determined to have a poor perfusion based on these criteria and was later excluded from data analysis before the randomization was unblinded.

Immediately after perfusion, the skin and mandible were carefully dissected from the skull, such that the skull was not compressed or dropped during the procedure. The skull was carefully placed in a 50-ml tube containing 25 ml of the perfusate solution, and each tube was placed on its side in a box on a rotator at ~60 rpm in a 4 C cold room for 48 hours. During this time, numbers on the tubes were replaced by letters in a random order by a person blind to the numbering system. The letter assignments of the numbers were known only by this individual, and the information was shared (i.e., the double-blind was broken) only after the completion of data analysis. After the tubes had been in the cold room for 2 days, brains were carefully extracted from skulls and placed in PBS. Brains were then cut into 200-µm coronal sections in PBS on a vibratome under dim room light. Sections corresponding to the aBLA were transferred to individual wells in a 24-well plate containing 1 ml of PBS/1% Triton X-100 per well. Plates were stored overnight in the dark in a 4 C cold room. The next day, sections were examined in plates by brief halogen transillumination on a dissecting microscope, and the ideal section for aBLA imaging was determined for each mouse in a blinded manner. Plates were then returned to the dark in a 4 C cold room for overnight storage.

The next day, 3 sections were mounted per gelatin-coated slide, with 150-µm square coverslips Scotch Magic-taped to both ends supporting a 170-µm coverslip again secured by a single layer of Scotch Magic tape. Sections were kept wet with PBS/1% Triton X-100. Each slide was Krazy-glued to the bottom of a 10-cm tissue culture dish which was then filled with 20 ml of PBS/1% Triton X-100 and placed in the dark at room temperature. Samples were transported to a multiphoton microscope, and an additional 20 ml of PBS/1% Triton X-100 was added to each plate. Then a single 2x2 tiled image centered on the predetermined ideal aBLA was obtained for each specimen according to the parameters listed in Table S1. Multiphoton scans were analyzed with Imaris (Bitplane) to quantify the reporter signal in each nucleus using the Spots function with the following parameters: Estimated Diameter = 15 µm, Background Subtraction = true, "Quality" above 4000. These parameters were determined by trial and error during preliminary experiments. The Intensity Sum was found for each nucleus and the mean Intensity Sum was calculated for each aBLA. This entire experiment was replicated, with 19 mice in the first round and 23 mice in the second round. As mentioned previously, when the data were plotted, 1 sample in the second round had a mean Intensity Sum more than 2.5 SD greater than the mean for all 42 samples; this 1 poorly perfused sample was excluded from the analysis. For each round of the replication, the double-blind was broken only after the data analysis was complete.

For the male-female comparison in Figure 4F, the protocol was the same as above except there was no stress intervention. For the cFos study in Figure 4A, B, the mice were unstressed and there was only 1 experimental group, so there was no blinding. Sections were stained overnight

at 4 C with 1:500 c-Fos ((9F6) Rabbit mAb #2250, Cell Signaling Technology) primary antibody, and again overnight at 4 C with 1:500 Alexa Fluor 594 (Thermo Fisher Scientific) secondary antibody. The Imaris Spots function parameters were the same as for Figure 4G and were applied to both channels. The Colocalize Spots function was used to remove repeated nuclei that met criteria for both channels, so that each nucleus was counted only once during the statistical analysis.

For the viral injections in Figure 4C-E, since mice were unstressed and constituted 1 experimental group, there was no blinding prior to data analysis. Mice were anesthetized with continuous 1.5% isoflurane and injected stereotactically in the BLA at AP -1.4, ML +/- 3.1, DV -3.9 with 0.5 µl of AAV10 containing 1:1 10,000-fold diluted CaMKII-Cre-ER : undiluted EF1a-CreDIO-FARN-mApple or 1:10 undiluted CaMKII-Cre-ER : undiluted EF1a-CreDIO-Actin-Chromobody®-mApple at a rate of 0.25 µl/min. The needle was held in place for an additional 3 minutes. Two weeks later, mice were injected IP with 20 mg/kg tamoxifen. And 1-2 weeks after tamoxifen administration, mice were perfused as described above. Dendrite length was calculated with Imaris' Filaments function, while blind to caspase reporter intensity, which was subsequently measured using Imaris' Spots function. Only bipolar, pyramidal-type neurons – having the morphology of principal cells – located in the aBLA were included in the statistical analysis.

Pharmacokinetics (Figure 5A)

LC-MS/MS Analysis

Sample preparation

Aliquots of 10 µL of plasma or 100 µL brain tissue homogenate (1 part tissue (g) + 2 parts water (mL)) were mixed with 10 µL of 100-nM Q-DEVD-OPh (internal standard) and 500 µL of deionized water. After liquid-liquid extraction (LLE) of analytes by 1 mL of dichloromethane, the lower layer was removed and 800 µL evaporated under a gentle stream of nitrogen. The residue was reconstituted in 100 µL of 70% mobile phase A/30% acetonitrile (see below); 5 µL (plasma) or 50 µL (brain homogenate) was injected into LC-MS/MS system.

LC conditions

Shimadzu 20A series liquid chromatography system; column: Agilent, Eclipse Plus C8, 3 x 50 mm, 1.8 µm particle size, at 45 C; mobile phase A: 98:2 H_2O:acetonitrile (0.1% formic acid); mobile phase B: acetonitrile; elution gradient: 0-1 min 0-95% B, 1-1.5 min 95% B, 1.5-1.7 min 95-0% B; run time: 4 min.

ESI-MS/MS conditions

AB/SCIEX API 4000 QTrap tandem-mass spectrometer; positive mode electrospray (e.g., declustering potential, temperature, gas flow) and mass spectrometer (parent/daughter ions,

quadrupole potential) parameters were optimized by direct infusion of pure analytes into the ESI-LC/MS system. MRM transitions (m/z units) monitored for quantification: Q-VD-OPh 514.0/226.6 and Q-DEVD-OPh (internal standard) 800.3/256.5. Calibration samples in 0.38 – 1000 nM (plasma) and 0.5 – 61.3 nM (brain tissue) ranges were prepared by adding known amounts of pure Q-VD-OPh into plasma or brain tissue homogenate and were analyzed along with study samples. Linear response was observed in the concentration ranges measured (r=0.9997 and 0.9984) and the low limits of quantification (LLOQ) were 0.38 nM and 0.5 nM (+/- 80% accuracy) in plasma and brain tissue, respectively.

Binding (Figure 5B)

NIMH's Psychoactive Drug Screening Program (Besnard et al., 2012) determined the binding affinities of the receptors and transporters listed in Figure 5B.

Neurophysiology (Figure 5C-I)

Chronic Multi-Circuit Electrode Implants

All mice used in this study were 2-4-month-old male C57BL/6J from Jackson Labs. Mice were group housed (3-5 animals per cage) in a climate-controlled facility for rodents maintained on a 12-hour light-dark cycle and fed with food and water ad libitum. Electrodes were arranged in bundles and implanted as previously described (Dzirasa et al., 2013). Briefly, mice were

anesthetized using isoflurane and placed in a stereotaxic frame. The isoflurane/oxygen flow rate was maintained at >1.5% saturation level and adjusted as needed to ensure full anesthesia as determined by tail-pinch response during the entire duration of surgery. Ground screws were placed in the skull above the cerebellum and the anterior cranium. HML coated tungsten wires were arranged in bundles and micro-wires were implanted into mice. All electrodes were referenced to the ground wire connected to the ground screws in the cranium. Electrode bundles were implanted as follows (all coordinates are measured from bregma):

Brain area	AP coordinates (mm)	ML coordinates (mm)	DV coordinates (mm)
PrL	+1.8	±0.25	–1.75
Amyg	–1.46	±2.85	–3.9
DHipp	–2.4	±1.63	–1.25

Neurophysiological Recordings

Acute drug treatment and brain network function

After 2-3 weeks of recovery from surgery, all mice were subjected to neurophysiological recordings. For 7 days, all mice were habituated to the handling required for connecting cables and performing saline injections. During the habituation period all mice were taken to the behavior room in their home cage and manually connected to the recording cables; after 15 minutes of recovery from handling, mice were given 20 minutes in the cage followed by a saline

injection and another 60 minutes in the cage. After 7 days of habituation all mice were connected to the recording cables for the multi-channel Cerebus acquisition system (Blackrock Microsystems Inc., UT). Following 15 minutes of recovery from handling while connecting the recording cables, brain activities as local field potential (LFPs) (band-pass filtered at 0.3-250 Hz) were recorded and stored at 1000 Hz. LFP activity was recorded for 20 minutes while mice were in the cage before the saline/drug/DMSO injections. Behavioral data with automated tracking was also acquired (Neuromotive, Blackrock Microsystems Inc., UT). Data were analyzed using custom-written MATLAB scripts.

Brain network function before, during and after acute drug treatment

Following 7 days of habituation, all mice were subjected to neurophysiological recordings for 3 days as follows: first day, all mice were injected with saline; second day, 8 and 7 mice were injected with drug or DMSO, respectively; third day, all mice were injected with saline. All mice were recorded in their cage for 20 minutes prior to injections. Data were collected and stored for further analysis.

Statistical Analysis

For all mice, 20 minutes of home cage brain activity was compared with 20 minutes of brain activity following saline or drug/DMSO treatment. Four LFP channels for each structure were pseudorandomly chosen for analysis. LFP cross-structural coherence was calculated from the

LFP pairs used for LFP oscillatory power analysis with the MATLAB (MathWorks, Natick, MA) mscohere function at a 1-second sliding window with a 1-second step.

LFPs and coherence activity were averaged for mice in the 6 frequencies that demonstrated bands of increased connectivity for 20 minutes: delta 1-4 Hz (Amy-Hip, Amy-PrL, and Hip-PrL), theta 4-11 Hz (Amy-Hip and Hip-PrL), and beta 15-30 Hz (PrL-Hip). The LFP power and coherence activity data were averaged across the recording session. Data were compared before and after treatment in the two groups using a RM-ANOVA with Box-Cox transformation, followed by a Wilcoxon sign-rank test with false discovery rate correction.

QUANTIFICATION AND STATISTICAL ANALYSIS

Data analysis was performed with Microsoft Excel, Imaris (Bitplane), and MATLAB; and statistical analysis was done with GraphPad Prism 7. All statistical details can be found in the figure legends and figures. $p<0.05$ was the threshold of significance. Due to a poor perfusion, one data point in the experiment shown in Figure 4G was more than 2.5 SD greater than the mean for all 42 samples and was excluded from the analysis prior to unblinding.

DATA AND SOFTWARE AVAILABILITY

MATLAB script for unbiased shading correction of images containing at least ~100 tiles: Data S1

FASTA sequence of the caspase activity reporter pApo: Data S2

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
Rabbit polyclonal anti-MEF2D	Millipore	Cat#AB2263
Rabbit monoclonal anti-c-Fos (clone 9F6)	Cell Signaling Tech.	Cat#2250
Bacterial and Virus Strains		
pAAV10-CaMKII-Cre-ER	This paper	N/A
pAAV10-EF1a-CreDIO-FARN-mApple	This paper	N/A
pAAV10-EF1a-CreDIO-Actin-Chromobody®-mApple	This paper	N/A
Biological Samples		
Chemicals, Peptides, and Recombinant Proteins		
Z-DEVD-FMK	Millipore	Cat#264156
Q-VD-OPh	Cayman Chemical	Cat#15260
Critical Commercial Assays		
ApopTag® Red In Situ Apoptosis Detection Kit	Millipore	Cat#S7165
PowerUp™ SYBR™ Green Master Mix	Thermo Fisher Sci.	Cat#A25742
Deposited Data		
Experimental Models: Cell Lines		
Mouse: Caspase reporter hippocampal neurons	This paper	N/A

Experimental Models: Organisms/Strains		
Mouse: Caspase activity reporter+/+: Mixed background	This paper	N/A
Mouse: Arpc3fl/fl x PCP2-Cre+/-	Lab of Scott Soderling	N/A
Mouse: Casp3^{tm1Flv}-/-: C57BL/6 congenic	Jackson Laboratory	N/A
Oligonucleotides		
Genotyping primer: CMV enhancer forward: CCCACTTGGCAGTACATCAA	This paper	N/A
Genotyping primer: CMV enhancer reverse: GCCAAGTAGGAAAGTCCCATAA	This paper	N/A
Control primer: Actb intron forward: GCCGTATTAGGTCCATCTTGAG	This paper	N/A
Control primer: Actb intron reverse: GGTTTGGACAAAGACCCAGA	This paper	N/A
Recombinant DNA		
Caspase activity reporter: pApo	This paper	Data S2
Software and Algorithms		
Shading correction algorithm: tiling_correction3	This paper	Data S1
Other		

Data S1

```
function tiling_correction3(filename,tile_dim)

% --- set threshold for highlights
threshold=0;

% --- Get lsm file name
[path name ext]=fileparts(filename);

% --- Generate nii file name
```

```
niiout=[path '/' name '.nii'];

% --- if not already a nii then convert
if ~exist(niiout,'file')
    display('Creating NIfTI')
    % --- Write ImageJ macro
    fid=fopen(fullfile(path,'lsm_to_nii.ijm'),'w+');
    display(fullfile(path,'lsm_to_nii.ijm'))
    fprintf(fid,'%s \n',['run("NIfTI-1", "save=',niiout,'");']);
    fprintf(fid,'%s \n','run("Quit");');
    fclose(fid);

    % --- run imageJ macro
    system(horzcat('java -Xmx10000m -jar
/Applications/ImageJ/ImageJ64.app/Contents/Resources/Java/ij.jar -ijpath
/Applications/ImageJ ',filename,' -macro ',fullfile(path,'lsm_to_nii.ijm')));
end

% --- Load nii, separate channels and convert to double precision
orig=load_nii(niiout);
voxdim=orig.hdr.dime.pixdim(2:4);
orig=orig.img;
[xres yres slices channels]=size(orig);

% --- make mtile stack for each channel unless it has already been done
if ~exist(horzcat(path,'/',name,'_mtile.nii'),'file')
    for b=1:channels
        mtile=zeros(tile_dim,tile_dim,slices,channels);
        for x=0:xres/tile_dim-1
            for y=0:yres/tile_dim-1
                display(horzcat('Working on tile
(',num2str(x),',',num2str(y),')'));

mtile=mtile+orig((tile_dim*x)+1:tile_dim*(x+1),(tile_dim*y)+1:tile_dim*(y+1),
:,:);
            end
        end
        mtile=mtile./(x*y);
    end
    % --- save mtile for inspection
    mtilenii=make_nii(mtile,[1 1 1],[0 0 0],64);
    output=horzcat(path,'/',name,'_mtile.nii');
    save_nii(mtilenii,output)
end

% --- if mtile is not in memory, load it
if ~exist('mtile','var')
    mtile=load_nii(horzcat(path,'/',name,'_mtile.nii'));
    mtile=reshape(mtile.img,tile_dim,tile_dim,slices,channels);
end

% --- apply correction
for b=1:channels
    for x=0:xres/tile_dim-1
        for y=0:yres/tile_dim-1
            for zz=1:slices
```

```
                mtile(:,:,zz,b)=mtile(:,:,zz,b)./max(max(mtile(:,:,zz,b)));
            end
            display(horzcat('Working on tile
(',num2str(x),',',num2str(y),')'));

orig((tile_dim*x)+1:tile_dim*(x+1),(tile_dim*y)+1:tile_dim*(y+1),:,b)=orig((t
ile_dim*x)+1:tile_dim*(x+1),(tile_dim*y)+1:tile_dim*(y+1),:,b)./mtile(:,:,:,b
);
        end
    end
end

% --- separate channels, do subtraction and downsample
c1=squeeze(orig(:,:,:,1));
c2=squeeze(orig(:,:,:,2));
combined=c2-(c1.*2);
combined_ds=zeros(round(size(combined)./2));
for i=1:slices
    combined_ds(:,:,i)=imresize(combined(:,:,i),.5,'bicubic');
end

% --- Create color 2 channel image
c1=max(c1,[],3);
c2=max(c2,[],3);
color=reshape(cat(4,c1,c2),size(c1,1),size(c1,2),1,1,channels);
nii=make_nii(color,voxdim,[0 0 0],16);
save_nii(nii,horzcat(path,'/',name,'_corrected_color','.nii'));

% --- threshold, z sum and save subtraction
thresh=sum((combined_ds.*double(combined_ds>threshold)),3);
thresh(thresh==0)=NaN;
nii=make_nii(thresh,voxdim.*2,[0 0 0],16);
save_nii(nii,horzcat(path,'/',name,'_corrected_subtraction','.nii'));

% --- cleanup
if exist(fullfile(path,'lsm_to_nii.ijm'),'file')
    system(horzcat('rm ',fullfile(path,'lsm_to_nii.ijm')));
end
```

Data S2

```
>pApo
GTCGACGTCGAGGGATCTTCATAAGAGAAGAGGGACAGCTATGACTGGGAGTAGTCAGGAGAGGAGGAAAAATCTGG
CTAGTAAAACATGTAAGGAAAATTTTAGGGATGTTAAAGAAAAAAATAACACAAAACAAAATATAAAAAAAATCTAA
CCTCAAGTCAAGGCTTTTCTATGGAATAAGGAATGGACAGCAGGGGGCTGTTTCATATACTGATGACCTCTTTATAG
CCACCTTTGTTCATGGCAGCCAGCATATGGCATATGTTGCCAAACTCTAAACCAAATACTCATTCTGATGTTTTAAA
TGATTTGCCCTCCCATATGTCCTTCCGAGTGAGAGACACAAAAAATTCCAACACACTATTGCAATGAAAATAAATTT
CCTTTATTAGCCAGAAGTCAGATGCTCAAGGGGCTTCATGATGTCCCCATAATTTTTGGCAGAGGGAAAAAGATCTC
AGTGGTATTTGTGAGCCAGGGCATTGGCCACACCAGCCACCACCTTCTGATAGGCAGCCTGCACCTGAGGAGTGGCG
CGCCTCACACTTTCCTTTTCTTCTTTGGTCCCCCTCCGTCTACCTCATCGCCCCCGCCGACGTTGATCCTGGCGCTG
GCGCAAGCAGCAGGGTGTCTATCCATGCCGCTCTCCTGGGCGCAGCTCATGGGCAGGGTGCCGGCGGCCTGCTCCTC
CACCTCGGGAGGGAAGCCGTGAGAATTTCCCCCTCCGTCTACCTCATCGCCCCCGCCCTTGTACAGCTCGTCCATGC
CGAGAGTGATCCCGGCGGCGGTCACGAACTCCAGCAGGACCATGTGATCGCGCTTCTCGTTGGGGTCTTTGCTCAGC
TTGGACTGGGTGCTCAGGTAGTGGTTGTCGGGCAGCAGCACGGGGCCGTCGCCGATGGGGGTGTTCTGCTGGTAGTG
```

GTCGGCGAGCTGCACGCTGCCGTCCTCGATGTTGTGGCGGATCTTGAAGTTGGCCTTGATGCCGTTCTTCTGCTTGT
CGGATTTGTTAAAACAATTGGCGGCGATCGCCCCATTGGCTAGCAGAAAATTATGGTCTTGGGGAAGACCAATATCA
AATATTCTTTGCACTCCGAGGGAACGACGACCGATAACTTTAACCATGGTGGCGGCGAGCTCTTTGCCAAAATGATG
AGACAGCACAATAACCAGCACGTTGCCCAGGAGCTGTAGGAAAAAGAAGAAGGCATGAACATGGTTAGCAGAGGCTC
TAGAGCCGCCGGTCACACGCCAGAAGCCGAACCCCGCCCTGCCCCGTCCCCCCCGAAGGCAGCCGTCCCCCTGCGGC
AGCCCCGAGGCTGGAGATGGAGAAGGGGACGGCGGCGCGGCGACGCACGAAGGCCCTCCCCGCCCATTTCCTTCCTG
CCGGCGCCGCACCGCTTCGCCCGCGCCCGCTAGAGGGGGTGCGGCGGCGCCTCCCAGATTTCGGCTCCGCCAGATTT
GGGACAAAGGAAGTCCCTGCGCCCTCTCGCACGATTACCATAAAAGGCAATGGCTGCGGCTCGCCGCGCCTCGACAG
CCGCCGGCGCTCCGGGGCCGCCGCGCCCCTCCCCCGAGCCCTCCCCGGCCCGAGGCGGCCCCGCCCCGCCCGGCACC
CCCACCTGCCGCCACCCCCCGCCCGGCACGGCGAGCCCCGCGCCACGCCCCGCACGGAGCCCCGCACCCGAAGCCGG
GCCGTGCTCAGCAACTCGGGGAGGGGGGTGCAGGGGGGGGTTACAGCCCGACCGCCGCGCCCACACCCCCTGCTCAC
CCCCCCACGCACACACCCCGCACGCAGCCTTTGTTCCCCTCGCAGCCCCCCCGCACCGCGGGGCACCGCCCCCGGCC
GCGCTCCCCTCGCGCACACGCGGAGCGCACAAAGCCCCGCGCCGCGCCCGCAGCGCTCACAGCCGCCGGGCAGCGCG
GGCCGCACGCGGCGCTCCCCACGCACACACACACGCACGCACCCCCCGAGCCGCTCCCCCCCGCACAAAGGGCCCTC
CCGGAGCCCTTTAAGGCTTTCACGCAGCCACAGAAAAGAAACGAGCCGTCATTAAACCAAGCGCTAATTACAGCCCG
GAGGAGAAGGGCCGTCCCGCCCGCTCACCTGTGGGAGTAACGCGGTCAGTCAGAGCCGGGGCGGGCGGCGCGAGGCG
GCGCGGAGCGGGGCACGGGGCGAAGGCAACGCAGCGACTCCCGCCCGCCGCGCGCTTCGCTTTTTATAGGGCCGCCG
CCGCCGCCGCCTCGCCATAAAAGGAAACTTTCGGAGCGCGCCGCTCTGATTGGCTGCCGCCGCACCTCTCCGCCTCG
CCCCGCCCCGCCCCTCGCCCCGCCCCGCCCCGCCTGGCGCGCGCCCCCCCCCCCCCCGCCCCCATCGCTGCACAAAA
TAATTAAAAAATAAATAAATACAAAATTGGGGGTGGGGAGGGGGGGGAGATGGGGAGAGTGAAGCAGAACGTGGGGC
TCACCTCGACACTAGTTATTAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCATATATGGAGTTCCGCGTTA
CATAACTTACGGTAAATGGCCCGCCTGGCTGACCGCCCAACGACCCCCGCCCATTGACGTCAATAATGACGTATGTT
CCCATAGTAACGCCAATAGGGACTTTCCATTGACGTCAATGGGTGGACTATTTACGGTAAACTGCCCACTTGGCAGT
ACATCAAGTGTATCATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCGCCTGGCATTATGCCC
AGTACATGACCTTATGGGACTTTCCTACTTGGCAGTACATCTACGTATTAGTCATCGCTATTACCATGGGTCGAGGT
GAGCCCCACGTTCTGCTTCACTCTCCCCATCTCCCCCCCCTCCCCACCCCCAATTTTGTATTTATTTATTTTTTAAT
TATTTTGTGCAGCGATGGGGGCGGGGGGGGGGGGGCGCGCGCCAGGCGGGGCGGGGCGGGGCGAGGGGCGGGGCGG
GGCGAGGCGGAGAGGTGCGGCGGCAGCCAATCAGAGCGGCGCGCTCCGAAAGTTTCCTTTTATGGCGAGGCGGCGGC
GGCGGCGGCCCTATAAAAAGCGAAGCGCGCGGCGGGCGGGAGTCGCTGCGTTGCCTTCGCCCCGTGCCCCGCTCCGC
GCCGCCTCGCGCCGCCCGCCCCGGCTCTGACTGACCGCGTTACTCCCACAGGTGAGCGGGCGGGACGGCCCTTCTCC
TCCGGGCTGTAATTAGCGCTTGGTTTAATGACGGCTCGTTTCTTTTCTGTGGCTGCGTGAAAGCCTTAAAGGGCTCC
GGGAGGGCCCTTTGTGCGGGGGGGAGCGGCTCGGGGGGTGCGTGCGTGTGTGTGTGCGTGGGGAGCGCCGCGTGCGG
CCCGCGCTGCCCGGCGGCTGTGAGCGCTGCGGGCGCGGCGCGGGGCTTTGTGCGCTCCGCGTGTGCGCGAGGGGAGC
GCGGCCGGGGGCGGTGCCCCGCGGTGCGGGGGGGCTGCGAGGGGAACAAAGGCTGCGTGCGGGGTGTGTGCGTGGGG
GGGTGAGCAGGGGGTGTGGGCGCGGCGGTCGGGCTGTAACCCCCCCCTGCACCCCCCTCCCCGAGTTGCTGAGCACG
GCCCGGCTTCGGGTGCGGGGCTCCGTGCGGGGCGTGGCGCGGGGCTCGCCGTGCCGGGCGGGGGGTGGCGGCAGGTG
GGGGTGCCGGGCGGGGCGGGGCCGCCTCGGGCCGGGGAGGGCTCGGGGGAGGGGCGCGGCGGCCCCGGAGCGCCGGC
GGCTGTCGAGGCGCGGCGAGCCGCAGCCATTGCCTTTTATGGTAATCGTGCGAGAGGGCGCAGGGACTTCCTTTGTC
CCAAATCTGGCGGAGCCGAAATCTGGGAGGCGCCGCCGCACCCCCTCTAGCGGGCGCGGGCGAAGCGGTGCGGCGCC
GGCAGGAAGGAAATGGGCGGGGAGGGCCTTCGTGCGTCGCCGCGCCGCCGTCCCCTTCTCCATCTCCAGCCTCGGGG
CTGCCGCAGGGGGACGGCTGCCTTCGGGGGGGACGGGGCAGGGCGGGGTTCGGCTTCTGGCGTGTGACCGGCGGCTC
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CTCATCATTTTGGCAAAGAATTCgccgccaccatgGACATCGACGAGCTGGCCCTGAAGTTCGCCGGCCTGGACCTG
GGCGGGGGCGATGAGGTAGACGGAGGGGGAAATTCTCACGGCTTCCCTCCCGAGGTGGAGGAGCAGGCCGCCGGCAC
CCTGCCCATGAGCTGCGCCCAGGAGAGCGGCATGGATAGACACCCTGCTGCTTGCGCCAGCGCCAGGATCAACGTCG
GCGGGGGCGATGAGGTAGACGGAGGGGGAGTGAGCAAGGGCGAGGAGCTGTTCACCGGGGTGGTGCCCATCCTGGTC
GAGCTGGACGGCGACGTAAACGGCCACAAGTTCAGCGTGTCCGGCGAGGGCGAGGGCGATGCCACCTACGGCAAGCT
GACCCTGAAGTTCATCTGCACCACCGGCAAGCTGCCCGTGCCCTGGCCCACCCTCGTGACCACCCTGACCTGGGGCG
TGCAGTGCTTCGCCCGCTACCCCGACCACATGAAGCAGCACGACTTCTTCAAGTCCGCCATGCCCGAAGGCTACGTC
CAGGAGCGCACCATCTTCTTCAAGGACGACGGCAACTACAAGACCCGCGCCGAGGTGAAGTTCGAGGGCGACACCCT
GGTGAACCGCATCGAGCTGAAGGGCATCGACTTCAAGGAGGACGGCAACATCCTGGGGCACAAGCTGGAGTACAACG
CCATCAGCGACAACGTCTATATCACCGCCaaatttgcggaatatTGCCTCAGTTTTGGCACCGAAATTTTAACCGTT
GAGTACGGCCCATTGCCCATTGGCAAAATTGTGAGTGAAGAAATTAATTGTTCTGTGTACAGTGTTGATCCAGAAGG
GAGAGTTTACACCCAGGCGATCGCCCAATGGCATGACCGGGGAGAGCAGGAAGTATTGGAATATGAATTGGAAGATG
GTTCAGTAATCCGAGCTACCTCTGACCACCGCTTTTTAACCACCGATTATCAACTGTTGGCGATCGAAGAAATTTTT
GCTAGGCAACTGGACTTGTTGACTTTAGAAAATATTAAGCAAACTGAAGAAGCTCTTGACAACCATCGTCTTCCCTT
TCCATTACTTGACGCTGGGACAATTAAATAAGCGGCCGCACTCCTCAGGTGCAGGCTGCCTATCAGAAGGTGGTGGC

TGGTGTGGCCAATGCCCTGGCTCACAAATACCACTGAGATCTTTTTCCCTCTGCCAAAAATTATGGGGACATCATGA
AGCCCCTTGAGCATCTGACTTCTGGCTAATAAAGGAAATTTATTTTCATTGCAATAGTGTGTTGGAATTTTTTGTGT
CTCTCACTCGGAAGGACATATGGGAGGGCAAATCATTTAAAACATCAGAATGAGTATTTGGTTTAGAGTTTGGCAAC
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AACATCCCTAAAATTTTCCTTACATGTTTTACTAGCCAGATTTTTCCTCCTCTCCTGACTACTCCCAGTCATAGCTG
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AAATTGTTATCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGGGTGCCTAATGAG
TGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTCCAGTCGGGAAACCTGTCGTGCCAGCGGATCCGC
ATCTCAATTAGTCAGCAACCATAGTCCCGCCCCTAACTCCGCCCATCCCGCCCCTAACTCCGCCCAGTTCCGCCCAT
TCTCCGCCCCATGGCTGACTAATTTTTTTTATTTATGCAGAGGCCGAGGCCGCCTCGGCCTCTGAGCTATTCCAGAA
GTAGTGAGGAGGCTTTTTTGGAGGCCTAGGCTTTTGCAAAAAGCTAACTTGTTTATTGCAGCTTATAATGGTTACAA
ATAAAGCAATAGCATCACAAATTTCACAAATAAAGCATTTTTTTCACTGCATTCTAGTTGTGGTTTGTCCAAACTCA
TCAATGTATCTTATCATGTCTGGATCCGCTGCATTAATGAATCGGCCAACGCGCGGGGAGAGGCGGTTTGCGTATTG
GGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCTCGGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCA
AAGGCGGTAATACGGTTATCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAAGGC
CAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATAGGCTCCGCCCCCCTGACGAGCATCACAAAAATCGAC
GCTCAAGTCAGAGGTGGCGAAACCCGACAGGACTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGC
TCTCCTGTTCCGACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGGCGCTTTCTCAATG
CTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAGCTGGGCTGTGTGCACGAACCCCCCGTTCAGC
CCGACCGCTGCGCCTTATCCGGTAACTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCA
GCCACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAGTGGTGGCCTAACTACGG
CTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCTTCGGAAAAAGAGTTGGTAGCTCTT
GATCCGGCAAACAAACCACCGCTGGTAGCGGTGGTTTTTTTGTTTGCAAGCAGCAGATTACGCGCAGAAAAAAAGGA
TCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGAAAACTCACGTTAAGGGATTTTGGT
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GATACCGCGAGACCCACGCTCACCGGCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAA
GTGGTCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCTAGAGTAAGTAGTTCGCCAGTT
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CTCCGGTTCCCAACGATCAAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTCCTC
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