

1 Article

2 ***Symbiodinium* genomes reveal adaptive evolution of**

3 **functions related to symbiosis**

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## 28 **Abstract**

29 Symbiosis between dinoflagellates of the genus *Symbiodinium* and reef-building corals forms  
30 the trophic foundation of the world's coral reef ecosystems. Here we present the first draft  
31 genome of *Symbiodinium goreau* (Clade C, type C1: 1.03 Gbp), one of the most ubiquitous  
32 endosymbionts associated with corals, and an improved draft genome of *Symbiodinium*  
33 *kawagutii* (Clade F, strain CS-156: 1.05 Gbp), previously sequenced as strain CCMP2468, to  
34 further elucidate genomic signatures of this symbiosis. Comparative analysis of four available  
35 *Symbiodinium* genomes against other dinoflagellate genomes led to the identification of 2460  
36 nuclear gene families that show evidence of positive selection, including genes involved in  
37 photosynthesis, transmembrane ion transport, synthesis and modification of amino acids and  
38 glycoproteins, and stress response. Further, we identified extensive sets of genes for meiosis  
39 and response to light stress. These draft genomes provide a foundational resource for  
40 advancing our understanding *Symbiodinium* biology and the coral-algal symbiosis.

41

## 42 Introduction

43 Coral reefs provide habitats for one-quarter to one-third of all marine species<sup>1</sup>. Although  
44 typically surrounded by nutrient-poor waters, coral reefs show high rates of primary  
45 productivity, with the fixed carbon supporting not only the biomass of reef organisms but  
46 also commercial and recreational fisheries and aquaculture. Reef-building corals rely on the  
47 symbiosis between the coral animal *per se* and photosynthetic dinoflagellates of the genus  
48 *Symbiodinium*. This symbiosis is based on mutual nutrient exploitation, with corals providing  
49 shelter and inorganic nutrients to their algal partners, while *Symbiodinium* supply their coral  
50 hosts with photosynthates that can meet up to 95% of the corals' energy requirements<sup>2,3</sup>.

51 The relationship between *Symbiodinium* and their host determines not only the rate of  
52 coral-reef growth (calcium carbonate deposition), but also how the system responds to  
53 environmental stress<sup>3</sup>. Many studies have shown that coral-*Symbiodinium* mutualism is  
54 susceptible to environmental factors including temperature, light and salinity. Exposure to  
55 ultraviolet radiation, thermal stress or a combination of both can initiate photoinhibition,  
56 decoupling of carbon flow between symbiont and host, oxidative damage and breakdown of  
57 the symbiosis, a phenomenon known as coral bleaching<sup>4</sup>. Unless the symbiosis is soon re-  
58 established the coral host is at risk of starvation, disease and eventual death<sup>5</sup>. In recent  
59 decades, coral bleaching has led to large-scale mortality on coral reefs around the world, with  
60 the most recent global coral bleaching event (2014-2016) now confirmed as the longest and  
61 most severe on record<sup>6,7</sup>.

62 Despite the critical importance of this coral-dinoflagellate symbiosis, little is known  
63 about the underlying molecular mechanisms (apart from photosynthesis and carbon  
64 exchange), largely due to the lack of comprehensive understanding of what molecules,  
65 pathways and functions *Symbiodinium* can contribute. Genomes of *Symbiodinium* (and of

66 dinoflagellates more broadly) are known for their idiosyncratic features including non-  
67 canonical splice sites, extensive methylation<sup>8</sup> and large sizes, up to 250 Gbp<sup>9</sup>. Their plastid  
68 genomes occur as plasmid-like minicircles<sup>10-12</sup>; their mitochondrial genomes harbor only  
69 three protein-coding genes and lack stop codons, and both mitochondrial<sup>13-15</sup> and nuclear<sup>16</sup>  
70 transcripts are extensively edited.

71 *Symbiodinium* are classified into nine clades<sup>17-19</sup>, with members of Clades A, B, C and  
72 D responsible for the vast majority of associations with scleractinian corals<sup>20</sup>. Draft genomes  
73 have been published for representatives of Clades A, B and F<sup>17-19</sup>, with sequence comparisons  
74 demonstrating them to be highly divergent<sup>18</sup>. Genome sequences are still lacking for Clade C,  
75 the most ubiquitous and diverse clade associated with tropical reef corals<sup>21</sup>, at least some sub-  
76 clades (“types”) of which are ecologically partitioned<sup>22</sup>.

77 Here we report draft genomes of two *Symbiodinium* from the Pacific Ocean: *S. goreau*  
78 (type C1; isolated from the acroporid coral *Acropora tenuis*) from the Great Barrier Reef, and  
79 *S. kawagutii* CS-156 (=CCMP2468, Clade F) from Hawaii. *Symbiodinium* type C1 is one of  
80 two “living ancestors” (along with type C3) of Clade C<sup>21</sup>, and one of the most dominant types  
81 associated with reef corals in both Indo-Pacific and Caribbean waters<sup>20</sup>. *S. goreau* has been  
82 reported from >150 coral species on Australia’s Great Barrier Reef, representing >80% of the  
83 studied coral genera in this region<sup>23</sup> across environments from reef flats to lower mesophotic  
84 depths<sup>23-25</sup>. In contrast, *S. kawagutii* CS-156 (=CCMP2468) was isolated during attempts to  
85 culture the symbiont from *Montipora verrucosa* (Todd LaJeunesse, *personal*  
86 *communication*). This isolate has yet to be verified to occur in mutualistic symbiosis with any  
87 coral, and appears incapable of establishing experimental symbiosis with cnidarian hosts<sup>26</sup>.  
88 Instead *S. kawagutii* may be exclusively a symbiont of foraminifera, or occur free-living at  
89 low environmental densities but proliferate opportunistically in culture. As some genome data

90 have been published for *S. kawagutii* CCMP2468<sup>18</sup>, we used these in combination with new  
91 data from the present study to generate a refined genome assembly. The genomes of *S.*  
92 *goreaui* and *S. kawagutii* offer a platform for comparative genomic analyses between two of  
93 the most-recently diverged *Symbiodinium* lineages Clades C and F, and published genome  
94 sequences in the more-basal Clades A and B.

95 Adopting a comparative approach using both genome and transcriptome data, we  
96 systematically investigated genes and functions that are specific to *Symbiodinium* vis-à-vis  
97 other dinoflagellates, and their association with the establishment and maintenance of  
98 symbiosis. We also computationally identify genes and functions for which there is evidence  
99 of adaptive selection in *Symbiodinium*. This is the most-comprehensive comparative analysis  
100 so far of *Symbiodinium* genomes, and the first to include a prominent endosymbiont of corals  
101 of Indo-Pacific and Caribbean reefs.

## 102 **Results**

### 103 ***Genomes of S. goreaui and S. kawagutii***

104 We sequenced and generated two draft *Symbiodinium* genome assemblies *de novo*, for *S.*  
105 *goreaui* (Clade C, 1.03 Gbp) and for *S. kawagutii* (Clade F, 1.05 Gbp). Details of data  
106 generation and assembly statistics are shown in Supplementary Tables S1 and S2  
107 respectively. Our *S. goreaui* assembly consists of 41,289 scaffolds (N50 length 98,034 bp).  
108 For *S. kawagutii*, we first verified that our data (from isolate CS-156) and the published data  
109 (from the synonym isolate CCMP2468) are indeed from the same culture of origin (see  
110 Supplementary Methods and Supplementary Figure S1). Compared to the published assembly  
111 (Lin et al.<sup>18</sup>), independent mapping of their ten fosmid sequences<sup>18</sup> onto our preliminary CS-  
112 156 assembly yielded up to 43-fold and 37-fold fewer gaps and mismatches respectively  
113 (Supplementary Figure S2). We later combined both datasets in a single *de novo* assembly,

114 yielding 16,959 scaffolds (N50 length 268,823 bp). Genome-size estimates based on *k*-mer  
115 coverage are 1.19 Gbp for *S. goreau* and 1.07 Gbp for *S. kawagutii* (Supplementary Table  
116 S3), comparable to those for other sequenced *Symbiodinium* genomes. We also recovered  
117 sequences putatively derived from their plastid genomes (Supplementary Tables S4, S5 and  
118 S6) including their distinct core conserved regions (Supplementary Table S7), and from their  
119 mitochondrial genomes; see Supplementary Note for details.

120 The repeat content of the assembled genomes ranged from 16.0% (*S. kawagutii*) to  
121 27.9% (*S. microadriaticum*); a large peak in transposable element (TE) abundance observed  
122 at high divergence (Kimura distance<sup>27</sup> 15-25) in all genomes (Supplementary Figure S3)  
123 suggests that most extant transposable elements are remnants of an ancient burst of TE  
124 activity. TE activity has been broadly linked to genome size in plants<sup>28-30</sup>, so reduced TE  
125 activity may be connected with the relative compactness of *Symbiodinium* genomes in  
126 comparison to those of other dinoflagellates. However, as these genomes are still in draft, the  
127 impact of assembly completeness on the patterns of repeat divergence cannot be dismissed.

128 Using a stringent threshold to remove genome scaffolds of potential bacterial or viral  
129 origin (Methods), we predict 35,913 and 26,609 high-quality gene models respectively for *S.*  
130 *goreau* and *S. kawagutii* (Supplementary Table S8). Usage profiles of codons and amino  
131 acids are shown in Supplementary Figures S4 and S5 respectively, and non-canonical splice  
132 sites in Supplementary Table S9 and Supplementary Figure S6. Although we report fewer  
133 genes, the majority (67.0% and 64.4% respectively for *S. goreau* and *S. kawagutii*) have  
134 transcriptome support, and we generally recovered more of the 458 conserved core eukaryote  
135 genes (Supplementary Figure S7), 371 of which are common to all four *Symbiodinium* based  
136 on the predicted gene models (Figure 1a; Supplementary Table S10); similar results are  
137 observed for the corresponding genome sequences (Supplementary Figure S7). About 94% of

138 the predicted genes have introns, similar to *S. microadriaticum* (98.2%) and *S. minutum*  
139 (95.3%); the earlier *S. kawagutii* genome assembly<sup>18</sup> may have underestimated the proportion  
140 of intron-containing genes (Supplementary Table S8) due to a less-stringent approach to gene  
141 prediction. All coding sequences have higher G+C content (56.7% in *S. goreau* and 55.0% in  
142 *S. kawagutii*) than does the genome overall, comparable to coding sequences from other  
143 *Symbiodinium* (57.7% in *S. microadriaticum* and 52.7% in *S. minutum*).

#### 144 ***Sequence divergence and synteny***

145 Despite the seemingly high number of protein-coding genes, an earlier analysis of syntenic  
146 blocks<sup>17</sup> found only several hundred blocks conserved in a pairwise manner among three  
147 published *Symbiodinium* genomes. Here we included our two new genome sequences in this  
148 analysis, and focused further on syntenic collinear blocks, requiring each block to contain the  
149 same genes in the same order and orientation with no gene losses (Methods). The *S. goreau*  
150 and *S. kawagutii* genomes share the most collinear blocks, 889 blocks implicating 8621  
151 genes; 62 of these blocks are of size >15, with the largest containing 76 genes  
152 (Supplementary Table S11). Thus substantial proportions of genes in these two genomes  
153 occur in clusters: for cluster size  $\geq 5$  genes, 32.4% and 24.0% of *S. kawagutii* and *S. goreau*  
154 genes respectively. These are likely to be underestimates, as the assemblies remain  
155 fragmentary. At the other end of the spectrum, the genomes of *S. microadriaticum* and *S.*  
156 *goreau* share only 86 collinear blocks of size  $\geq 5$ , with maximum size 12 and implicating  
157 588 genes in total (Supplementary Table S11). These results suggest that (a) although Clades  
158 C and F are divergent, they nonetheless are the most-closely related among the four analysed  
159 *Symbiodinium* genomes (in line with their phylogenetic relationship); and (b) C and F are  
160 more divergent from Clade A than from Clade B (in line with their phylogenetic  
161 relationship). Thus whole-genome sequences support and extend earlier conclusions, based

162 on common marker sequences, about phylogenetic relationships among *Symbiodinium*  
163 clades<sup>31-33</sup> and the remarkable divergence among *Symbiodinium* lineages<sup>17,18</sup>.

#### 164 ***Gene and protein functions***

165 All annotated genes from *S. goreau* and *S. kawagutii* genomes are listed in Supplementary  
166 Tables S12 and S13 respectively. Of the 35,913 proteins predicted in *S. goreau*, 31,646  
167 (88.1%) show similarity (BLASTP,  $E \leq 10^{-5}$ ) to sequences in UniProt; among these, 29,198  
168 (81.3% of 35,913) and 19,718 (54.9%) are annotated with Gene Ontology (GO) terms or  
169 Pfam domains (Supplementary Tables S12 and S14). For *S. kawagutii*, 21,947 of 26,609  
170 proteins (82.5%) find a match in UniProt (Supplementary Tables S13 and S14). *Protein*  
171 *kinase* (Pfam PF00069), *reverse transcriptase* (PF07727), *ion transport protein* (PF00520)  
172 and *ankyrin repeats* (PF12796) are among the most-abundant domains in *Symbiodinium*,  
173 appearing among the ten most-abundant for each of the four genomes (Supplementary Table  
174 S15). Ankyrin repeat motifs are important in the recognition of surface proteins, and more  
175 generally in protein-protein interactions<sup>34</sup>. Thus proteins potentially involved in host-  
176 symbiont interaction (with phosphorylation, ion transport and protein recognition/interaction  
177 domains) are well represented in the predicted *Symbiodinium* proteomes.

178 We compared functions of proteins predicted from these four *Symbiodinium* genomes  
179 to a set of 27 more-narrowly scoped eukaryotes: 17 alveolates (ten other dinoflagellates, four  
180 ciliates, two apicomplexans and *Perkinsus marinus*), stramenopiles (two diatoms) and  
181 Archaeplastida (four rhodophytes, three chlorophytes and *Arabidopsis*). This 31-taxon set  
182 (1,136,347 proteins; Supplementary Tables S16 and S17) represents lineages in which one or  
183 more endosymbioses are implicated in plastid origin<sup>35-37</sup>; these proteins were clustered (based  
184 on sequence similarity) into 56,530 groups of size two or greater (Supplementary Table S17;  
185 see Methods). Using this 31-taxon dataset as background, we assessed the over- or under-



186 representation of protein domains within our various groups of *Symbiodinium* proteins. We  
187 found 270 domains (Supplementary Table S18) to be significantly overrepresented (adjusted  
188  $p \leq 0.05$ ) in *Symbiodinium*. Interestingly, many domains e.g. *C-5 cytosine-specific DNA*  
189 *methylase* (PF00145), *planctomycete cytochrome c* (PF07635) and RNA polymerase *sigma-*  
190 *70 region 2* (PF04542) are also overrepresented in the four *Symbiodinium* genomes in a  
191 similar comparison against 880,909 proteins in a 15-taxon set that includes ten other  
192 dinoflagellates and the immediate outgroup *Perkinsus marinus* (Supplementary Table S19).  
193 Thus compared to related eukaryotes and to other dinoflagellates, *Symbiodinium* is enriched  
194 in functions involved in methylation of cytosine, (photosynthetic) energy production and  
195 RNA polymerisation. Hydroxymethylation of uracil is common (12-70%) in dinoflagellate  
196 genomes<sup>38</sup>; while cytosine methylation has been described in *Symbiodinium*<sup>39,40</sup>, our findings  
197 suggest that cytosine methylation is more prominent in *Symbiodinium* than in these other  
198 dinoflagellates.

199       Activation of some retrotransposons is part of the stress-response mechanism in  
200 diatoms<sup>41</sup>, plants<sup>42,43</sup> and other eukaryotes<sup>44,45</sup>. The *reverse transcriptase* domain (PF07727)  
201 is enriched in *Symbiodinium* in both the 31-taxon and 15-taxon sets, suggesting that  
202 retrotransposition could be a prominent mechanism of stress response in *Symbiodinium* and  
203 dinoflagellates. Although we set a stringent threshold for removing putative bacterial or viral  
204 sequences (see Methods), 40 (~0.1%) of the final 41,289 genome scaffolds of *S. goreau* have  
205 significant hits (BLASTN  $E \leq 10^{-20}$ ) to the virus genomes<sup>46</sup> isolated from the same *S. goreau*  
206 (type C1) strain, with 16 identical regions (76-609 bp) distributed in nine scaffolds of lengths  
207 ranging from 1092 to 7,338,656 bp. Whether this indicates introgression of viral sequences  
208 remains to be determined.

## 209 ***Positive selection of Symbiodinium genes***

210 Using a branch-site model based on the ratio of substitution rates in non-synonymous (dN) to  
211 synonymous (dS) sites<sup>47</sup> (Methods and Supplementary Figure S8), we identified  
212 *Symbiodinium* genes under positive selection in comparison to ten other dinoflagellates, with  
213 *Perkinsus marinus* as the outgroup (15 taxa: Supplementary Tables S16 and S17). The  
214 reference species tree (Figure 2a) was computed following Price and Bhattacharya<sup>48</sup>. We then  
215 based our analysis of adaptive evolution on all orthologous sets for which the protein tree is  
216 topologically congruent with our reference tree (Methods).

217 Of our 44,282 homologous sets, 2460 containing 7987 *Symbiodinium* proteins show  
218 evidence of positive selection in one or more *Symbiodinium* lineages; 1069 of these sets are  
219 annotated with GO terms (Supplementary Table S20). Figure 2b shows the terms (level 3) in  
220 the three GO hierarchies that are shared by  $\geq 5\%$  of these 1069 sets. In the Biological Process  
221 hierarchy, metabolic processes are highly represented (*primary metabolic process* [292] and  
222 *macromolecule compound metabolic process* [243] are among the four most-frequent terms),  
223 followed by *oxidation reduction* [96] and transport (*establishment of localization* [90],  
224 *transport* [90], and *transmembrane transport* [80]). Highly represented terms in the  
225 Molecular Function hierarchy point to binding of diverse molecules and ions e.g. *protein*  
226 *binding* [173] and metabolism (*hydrolase* [390], *transferase* [344]). In Cellular Component,  
227 *cell part* [113], *membrane* [86] and *membrane part* [59] are the most frequent. Thus in  
228 *Symbiodinium* as represented by these four assemblies, broad aspects of metabolism, and  
229 transport including across membranes, show evidence of positive selection, in line with their  
230 recognised importance in cnidarian-dinoflagellate symbioses<sup>17</sup>.

231 We further assessed enrichment of GO terms against all annotated terms specifically in  
232 the four *Symbiodinium* genomes (Supplementary Table S21) compared with the other

233 dinoflagellates in this study. Here we consider themes among Biological Process terms. The  
234 first theme is that functions associated with photosynthetic light reactions are enriched among  
235 the positively selected *Symbiodinium* genes; *photosynthesis*, *light reaction* and *Photosystem*  
236 *II assembly* are significantly over-represented (adjusted  $p \leq 0.05$ ), as are Cellular Component  
237 terms related to plastid functions e.g. *thylakoid*, *photosynthetic membrane*, *intracellular*  
238 *membrane-bounded organelle* (Supplementary Table S21). Coral bleaching has been  
239 associated with the loss of light-harvesting proteins and the subsequent inactivation of  
240 photosystem II (PSII) in *Symbiodinium* under combined light and temperature stress<sup>49-51</sup>.  
241 These authors reported that coral bleaching associated with algal photobleaching can be  
242 ameliorated, at least in part, by thermal acclimation of *Symbiodinium* to improve the thermal  
243 tolerance of PSII.

244 The second emerging theme involves the transport of ions and metabolites across  
245 membranes. *Intracellular transport*, *cytosolic transport*, *transition metal ion transport* and  
246 *copper ion transport* as well as terms related to transmembrane transport of amino acids,  
247 organic acids and carboxylic acids are significantly enriched (Supplementary Table S21);  
248 these functions underpin multiple physiological processes, including but not limited to pH  
249 regulation, calcification and photosynthetic carbon fixation<sup>52</sup>. Investigated *Symbiodinium* are  
250 enriched in membrane transporters compared with other dinoflagellates<sup>17</sup>. These biological  
251 processes are especially relevant to the maintenance and regulation of coral-dinoflagellate  
252 symbiosis<sup>52,53</sup>, possibly including its sensitivity and/or response to environmental stress.

253 The third theme is the enrichment of functions related to the biosynthesis and  
254 modification of amino acids and glycoproteins (Supplementary Table S21) e.g. *protein*  
255 *phosphorylation*, *peptide biosynthesis process*, *protein ADP-ribosylation*, *protein*  
256 *glycosylation*, *D-amino acid metabolic process* and *glycoprotein biosynthetic process*. Corals

257 lack the capacity to synthesise a number of amino acids (e.g. cysteine in *Acropora*  
258 *digitifera*<sup>54</sup>), thus selection acting on the synthesis of amino acids may indicate the critical  
259 role of *Symbiodinium* in supplying amino acids both for self-preservation and for the coral  
260 host. Glycoprotein molecules are often surface-localised and in microbes form the basis of  
261 microbe-associated molecular patterns (MAMPs) which, in conjunction with a host-  
262 associated pattern recognition receptor, mediate recognition by a host<sup>55</sup>. Both in culture and  
263 *in hospite*, *Symbiodinium* exude glycoconjugates<sup>56-59</sup>. Where investigated, cell-surface glycan  
264 profiles are stable over time within a *Symbiodinium* culture but can differ between clades  
265 within a host<sup>60</sup>. *N*-acetyl and mannosyl residues are prominent constituents of *Symbiodinium*  
266 cell-surface glycans, making them candidates for MAMPs that could participate in the  
267 establishment of symbiosis. Lin et al.<sup>18</sup> reported a *S. kawagutii* glycan biosynthesis pathway  
268 distinct from that of *S. minutum*, again pointing to a possible role of glycans in specificity of  
269 host recognition<sup>60,61</sup>. Neubauer et al.<sup>62</sup> demonstrated that the thrombospondin type 1 repeat  
270 (TSR) from the sea anemone *Aiptasia pallida* contains binding sites for glycosaminoglycan,  
271 and that blocking TSR led to decreased colonisation by *S. minutum*. Our results offer the first  
272 evidence of positive selection of functions underlying the biosynthesis and modification of  
273 amino acids and glycoproteins, suggesting that these functions are critical in the  
274 establishment of cnidarian-dinoflagellate symbioses.

275 Our fourth emerging theme is stress response. Enriched terms annotated for the  
276 positively selected genes include *cell redox homeostasis*, *translation initiation* and 22 terms  
277 describing the negative regulation of gene expression, transcription, RNA biosynthesis and  
278 cellular biosynthetic and metabolic processes (Supplementary Table S21). Environmental  
279 stressors can disrupt the cellular redox homeostasis and break down the coral-dinoflagellate  
280 symbiosis. Negative regulation of transcription may represent a stress response that  
281 safeguards the genome from accumulating DNA damage<sup>63</sup>; a similar stress response has been

282 observed in coral<sup>64</sup>. Other enriched functions that may be related to stress response include  
283 *mitotic nuclear division, translation*, and various processes of nucleotide biosynthesis and  
284 modification e.g. *RNA methylation, rRNA methylation, DNA replication, RNA processing*,  
285 and *deoxyribonucleotide biosynthetic process*. Our results thus provide evidence that stress  
286 response is under positive selection in *Symbiodinium*, presumably (given their lifestyle) in  
287 relation to the establishment and/or maintenance of symbiosis.

### 288 ***Do Symbiodinium have sex?***

289 *Symbiodinium* have been hypothesised to reproduce sexually and to have a diploid life stage<sup>65</sup>  
290 but definitive evidence for sex, e.g. karyogamy and meiosis, has yet to be observed<sup>31,66-68</sup>.  
291 The ability to reproduce sexually offers increased efficiency of selection and adaptation<sup>69</sup>. So  
292 far, the strongest evidence supporting meiotic potential in *Symbiodinium* comes from patterns  
293 of population-genetic variation revealed in allozymes, randomly amplified polymorphic DNA  
294 and other molecular markers<sup>22,31,70-73</sup>. Indeed, for some markers a higher degree of genetic  
295 variation has been observed in certain *Symbiodinium* clades than in dinoflagellates known to  
296 reproduce sexually<sup>70</sup>. More recently, differential gene expression analysis<sup>74</sup> using a  
297 heterologous culture from which our sequenced *S. goreau* was derived revealed an  
298 enrichment of gene functions related to meiosis under thermal stress, suggesting a switch  
299 from mitosis to meiosis under stress conditions.

300 Schurko and Logsdon<sup>75</sup> described a “meiosis detection toolkit”, a set of 51 genes  
301 specific or related to meiosis<sup>76,77</sup> that collectively point to a capacity for meiosis even in  
302 divergent or specialised eukaryotic genomes<sup>78</sup>. Incomplete genome coverage or assembly,  
303 sequence divergence, paralogy, patterns of overlapping function and evolutionary  
304 specialisation mean that not all 51 need to be present or detectable for a lineage to be  
305 assessed as probably sexual, or only recently asexual<sup>75,77</sup>. Thirty-one of these genes were

306 earlier identified in *Symbiodinium* Clades A and B<sup>76</sup>. Here, BLASTP search ( $E \leq 10^{-5}$ ) of  
307 predicted proteins in these four *Symbiodinium* genomes recovered matches corresponding to  
308 48 of the of 51 toolkit genes in *S. microadriaticum*, 47 in *S. minutum* and *S. goreau*, and 46  
309 in *S. kawagutii* (Figure 3a and Supplementary Table S22). Eight of the 11 meiosis-specific  
310 proteins were detected in all four *Symbiodinium*. REC114, SAD1 and XRS2 found weaker  
311 matches ( $E \geq 10^{-14}$ ) in one to two genomes, although confirmatory UniProt domains were  
312 usually present (Supplementary Table S22). RAD17 is the *Schizosaccharomyces pombe*  
313 homolog of *S. cerevisiae* RAD24<sup>79</sup>, for which we find highly significant matches ( $E \leq 10^{-127}$ )  
314 in all four *Symbiodinium*. Moreover, 15 of the 51 genes show evidence of positive selection  
315 in *Symbiodinium* against other dinoflagellates (Supplementary Table S22). Our data imply  
316 that these four *Symbiodinium* are, or until recently have been, capable of meiosis.

### 317 ***Response to light stress***

318 Mycosporine-like amino acids (MAAs) are UV-protective compounds that, in corals and  
319 other marine organisms, also act as antioxidants scavenging reactive oxygen species (ROS).  
320 Up to five MAAs have been reported in *Symbiodinium* (Clades A, B and C) isolated from  
321 cnidarian hosts<sup>80,81</sup>. The MAA biosynthetic pathway involves dehydroquinase synthase  
322 (DHQS), *O*-methyltransferase (*O*-MT), an ATP-grasp and non-ribosomal peptide synthetase  
323 (NRPS)<sup>82,83</sup>. In cyanobacteria, a short-chain dehydrogenase may play a role in converting  
324 shinorine to palythine-serine<sup>84</sup>. Genes encoding these four MAA-biosynthetic enzymes were  
325 reported absent from the *S. kawagutii* genome<sup>18</sup>. Here, using known proteins in bacteria,  
326 fungi and cnidarians as queries, we recovered all five enzymes including the short-chain  
327 dehydrogenase from the *S. microadriaticum*, *S. goreau* and *S. kawagutii* genomes  
328 (Supplementary Table S23); ATP-grasp was not found in *S. minutum*. These enzymes were  
329 earlier reported absent from *S. kawagutii*, and it was proposed that their absence can be  
330 compensated via coral-*Symbiodinium* co-evolution<sup>18</sup>; this hypothesis remains to be

331 investigated, but we note that this *S. kawagutii* isolate has not been observed in association  
332 with an animal host<sup>26</sup>.

333 Scytonemin is a UV-blocker first reported in terrestrial cyanobacteria<sup>83,85</sup>, and in  
334 contrast to MAAs was thought to be synthesised exclusively by cyanobacteria<sup>86</sup>. The genome  
335 of *Nostoc punctiforme* contains an 18-gene operon that specifies proteins of scytonemin  
336 biosynthesis and regulation, including the synthesis of aromatic amino-acid precursors<sup>87,88</sup>. Its  
337 expression is up-regulated by UV radiation<sup>89</sup>. Homologs of six of these 18 genes have been  
338 described in the coral *Acropora ditigifera*, and were considered putative instances of lateral  
339 genetic transfer<sup>90</sup>. We find 12 of these 18 genes in the genomes of *S. goreau* and in *S.*  
340 *kawagutii*, 11 in *S. microadriaticum* and ten in *S. minutum* (Figure 3b, Supplementary Table  
341 S24).

342 Genes responsible for biosynthesis of tryptophan (*trpA*, *trpB*, *trpC*, *trpD* and *trpE*) and  
343 the two key enzymes of chorismate (and aromatic amino acid) biosynthesis, *aroG* and *aroB*  
344 (dehydroquinate synthase, also important for MAA biosynthesis), are found in all  
345 *Symbiodinium* genomes, albeit so far in different scaffolds; these genes are also present in  
346 *Arabidopsis thaliana* although not in corals or *Hydra* which, like most other animals, are  
347 unable to synthesise tryptophan. The recovery of more of these 18 genes in *Symbiodinium*  
348 than in corals or other animals (Figure 3b) could reflect the impact of endosymbiotic  
349 association of ancestral cyanobacteria during the course of plastid evolution in all  
350 photosynthetic eukaryotes including dinoflagellates<sup>35-37</sup>. The presence of multiple gene copies  
351 (Supplementary Table S24) also implicates genetic duplication. Our findings suggest that  
352 *Symbiodinium* has the capacity to produce scytonemin, either by itself or jointly within the  
353 holobiont.

## 354 **Discussion**

355 *Symbiodinium* can form associations with a wide range of cnidarian hosts (as well as some  
356 other marine invertebrates and protists) across broad geographic and time scales<sup>91</sup>. The  
357 symbiosis between corals and *Symbiodinium* relies on compatible host-symbiont recognition  
358 and sustainable nutrient exchange, both of which are vulnerable to external environmental  
359 factors including temperature and light. A sustainable coral-*Symbiodinium* association  
360 requires an adaptive capacity in the face of environmental extremes.

361 In this study we generated the first draft genome of *S. goreau* (Clade C), a much-  
362 improved draft genome of *S. kawagutii* (Clade F) and high-quality gene models for both.  
363 Comparative analysis revealed remarkable divergence among the genomes of *Symbiodinium*  
364 from four clades, consistent with previous single-gene phylogenetic relationships. We  
365 identified 2460 *Symbiodinium* gene families that are under positive selection, many of which  
366 encode functions directly relevant to the establishment and/or maintenance of symbiosis with  
367 the coral host. We also identified complete, or near-complete, sets of genes indicative of the  
368 presence of meiosis and several mechanisms of stress tolerance, functions that have until now  
369 been considered absent from *S. kawagutii*. Our results demonstrate the remarkable genomic  
370 capacity of *Symbiodinium* to synthesize key metabolites that are essential to the  
371 establishment of symbiosis with coral hosts, and to respond to environmental stress.

372 *S. goreau* (type C1) belongs to one of the most globally dominant clades (Clade C) on  
373 coral reefs, and analysis of its draft genome has provided important new insights into coral-  
374 algal symbiosis. This genomic resource is already demonstrating utility in the identification  
375 of symbiont fractions in *de novo* sequencing of coral tissues<sup>92,93</sup>, and will provide a  
376 foundation for targeted studies into the molecular biology, physiology and of this crucial  
377 symbiosis and its adaptation to a changing environment.



## 378 **Methods**

### 379 ***Biological materials and sequencing data***

380 *Symbiodinium goreau* (Clade C, type C1; AIMS-aten-C1-MI-cfu-B2, now AIMS culture  
381 collection SCF055-01) is a single-cell monoclonal culture first isolated from the coral  
382 *Acropora tenuis* at Magnetic Island (Queensland, Australia) at 3 m depth<sup>94</sup>. *Symbiodinium*  
383 *kawagutii* CCMP2468 (Clade F) was maintained as a monoclonal culture. Genomic DNA  
384 was extracted from these isolates using the Qiagen DNeasy Plant Mini Kit. We generated a  
385 total of 116.0 Gb of sequence data (2 paired-end libraries of 230- and 500-bp inserts, plus 3  
386 mate-pair libraries of 3-, 6- and 9-Kbp inserts) for *S. goreau*, and a total of 92.2 Gb of  
387 sequence data (1 paired-end library of 230-bp inserts, plus 3 mate-pair libraries of 4-, 6- and  
388 9-Kbp inserts) for *S. kawagutii*, in each case using the Illumina HiSeq 2500 Rapid Chemistry  
389 platform. See Supplementary Methods and Supplementary Table S1 for details.

### 390 ***Genome assembly and annotation***

391 We adopted a combined approach to *de novo* genome assembly (Supplementary Methods) in  
392 which multiple assembly programs (CLC Genomics Workbench (Qiagen), SPAdes<sup>95</sup> and  
393 ALLPATHS-LG<sup>96</sup>) were first used independently. The quality of each assembly was assessed  
394 based on full-length recovery of phylogenetic markers and known coding sequences. Once  
395 the best assembly (the master assembly) was identified, other assemblies were used to refine  
396 it via merging scaffolds and filling gaps. We adopted a comprehensive *ab initio* approach for  
397 gene prediction using all available dinoflagellate proteins, as well as all *Symbiodinium* genes  
398 and transcriptomes, as guiding evidence. Our approach combines evidence-based methods i.e.  
399 PASA (with TransDecoder)<sup>97</sup>, AUGUSTUS<sup>98</sup> and MAKER<sup>99</sup>, and unsupervised machine-  
400 learning (GeneMark-ES<sup>100</sup> and SNAP<sup>101</sup>; alternative splice sites were specified in these  
401 methods by modifying the relevant scripts. We then used EvidenceModeler<sup>102</sup> to combine

402 multiple sets of predicted genes, allocating a heavier weight for evidence-based predictions  
403 than for those produced by unsupervised approaches; for details see Supplementary Methods.  
404 Final genome assemblies, predicted gene models and proteins are available at  
405 <https://cloudstor.aarnet.edu.au/plus/index.php/s/6yziMf2ygWjGu0L>.

406 We adopted multiple approaches to assess genome completeness. Established methods  
407 including CEGMA<sup>103</sup> and BUSCO<sup>104</sup> are based on conserved genes in a limited number of  
408 eukaryote model organisms that are distantly related to dinoflagellates. The use of these  
409 methods resulted in relatively low recovery of conserved eukaryote genes in *Symbiodinium*  
410 (e.g. 33-42% of BUSCO genes; Supplementary Figure S7) when run at default setting. We  
411 further assessed completeness using BLAST based on predicted proteins from the gene  
412 models and the assembled genome scaffolds. For each genome, we followed Baumgarten et  
413 al.<sup>105</sup> and searched (BLASTP,  $E \leq 10^{-5}$ ) against the predicted proteins using the 458 CEGMA  
414 proteins<sup>103</sup>. We also searched against the CEGMA proteins using the genome scaffolds  
415 (BLASTX  $E \leq 10^{-5}$ ), against genome scaffolds using the 458 CEGMA proteins (TBLASTN,  
416  $E \leq 10^{-5}$ ), and against genome scaffolds using the 458 CEGMA transcripts (TBLASTX,  $E \leq$   
417  $10^{-5}$ ) (Supplementary Table S10, Supplementary Figure S7).

#### 418 ***Identification and removal of bacterial and viral sequences***

419 Bacterial and viral sequences were identified and removed following Aranda et al.<sup>17</sup>. Briefly,  
420 we used our genome scaffolds (BLASTN,  $E \leq 10^{-20}$ ) to query the complete and draft bacterial  
421 genomes in NCBI, and the viral genomes in NCBI and PhAnToMe (<http://phantome.org>). We  
422 applied more-stringent criteria than did Aranda et al.<sup>17</sup> to identify putative contaminating  
423 sequences, removing from the final assembly any scaffold sequence that contains regions that  
424 match a bacterial or viral genome (BLASTN bit score > 1000,  $E \leq 10^{-20}$ ) where such regions

425 constitute >10% of the overall scaffold length (Supplementary Methods, Supplementary  
426 Figure S9).

#### 427 ***Analysis of genome synteny and collinearity***

428 Using all predicted genes and their associated genomic positions, we assessed the number of  
429 syntenic collinear blocks (i.e. regions with the same genes coded in the same order, free from  
430 rearrangement or loss) shared pairwise among genomes of *S. microadriaticum* (Clade A)<sup>17</sup>, *S.*  
431 *minutum* (B)<sup>19</sup>, *S. goreauii* (C) and *S. kawagutii* (F). We used BLASTP ( $E \leq 10^{-5}$ ) to search  
432 for similar proteins within each genome, and among all of them. Next we used MCScanX<sup>106</sup>  
433 with parameter -s 5 to sort the BLASTP matches (alignments) based on genomic positions; to  
434 minimise the number of collinear gene pairs arising from tandem repeats, we report only  
435 collinear blocks that consist of five or more genes.

#### 436 ***Functional annotation of gene models***

437 We adopted a similar approach to Aranda et al.<sup>17</sup> to annotate gene models. Protein sequences  
438 predicted using the standard genetic code were used as query (BLASTP,  $E \leq 10^{-5}$ ) first  
439 against Swiss-Prot, and those with no Swiss-Prot hits subsequently against TrEMBL (both  
440 databases from UniProt release 2016\_10). Gene Ontology (<http://geneontology.org/>) terms  
441 associated with Swiss-Prot and TrEMBL hits were obtained using the UniProt-GOA mapping  
442 (release 2016\_10).

#### 443 ***Identification of homologous protein groups and gene families***

444 Our workflow for delineation of sets of putatively homologous proteins, multiple sequence  
445 alignment, generation of protein-family and reference trees, and analysis of selection is  
446 shown in Supplementary Figure S8. Using predicted proteins from 31 phylogenetically diverse  
447 organisms including *Symbiodinium* for which genome and/or transcriptome data are available  
448 (Supplementary Table S25), we identified sets of putatively homologous proteins using

449 OrthoFinder<sup>107</sup> and consider the corresponding gene sets (families) to be homologs.  
450 Following Harlow et al.<sup>108</sup> and Beiko et al.<sup>109</sup>, we consider sequences within single-copy sets  
451 (i.e. those in which each genome is represented no more than once) to be orthologs. We refer  
452 to sets that contain proteins only from *Symbiodinium*, plus the *Symbiodinium* singletons, as  
453 *Symbiodinium*-specific. For enrichment analysis of annotated features (GO terms or Pfam  
454 domains), we compared the features within the *Symbiodinium*-specific set against those in  
455 each background set (i.e. the 31-taxon set and, separately, the 15-taxon set) using a  
456 hypergeometric test; features with an adjusted<sup>110</sup>  $p$ -value  $< 0.05$  were considered significant.

#### 457 ***Analysis of positive selection in Symbiodinium genes***

458 For the 15-taxon set we sorted the 311,651 protein sets into 1,654 single-copy (ortholog) and  
459 16,836 multi-copy sets. Multiple sequence alignments were carried out using MAFFT v7.245  
460 at -linsi mode<sup>111</sup>; questionably aligned columns and rows were removed from these  
461 alignments using trimAl<sup>112</sup> with the -automated1 option.

462 Branch-site models (BSMs; see below) require a reference topology. We follow Price  
463 and Bhattacharya<sup>48</sup> to generate the reference species tree. The trimmed single-copy protein  
464 alignments were concatenated prior to maximum-likelihood (ML) inference of the species  
465 phylogeny using IQTREE<sup>113</sup>; each alignment represents a partition for which the best  
466 evolutionary model was determined independently. Support for each node was assessed using  
467 2000 rapid bootstraps. The species tree so generated (Figure 2a) is similar to that of Price and  
468 Bhattacharya<sup>48</sup>, with very strong support ( $\geq 96\%$  bootstrap replicates) for all internal nodes;  
469 the *Symbiodinium* and Suessiales (*Symbiodinium* + *Polarella glacialis*) clades received 100%  
470 bootstrap support.

471 Of all trimmed protein alignments, those with  $\geq 60$  aligned positions and  $\geq 4$  sequences  
472 were used in subsequent analysis. For multi-copy protein sets, we imposed further filtering

473 criteria. We first inferred individual ML trees for the multi-copy sets using IQ-TREE, and  
474 each resulting protein tree was compared with the reference species tree. Those congruent  
475 with the reference species tree at genus level, and in which all *Symbiodinium* are resolved as  
476 an exclusive monophyletic clade, were judged paralog-free and used in subsequent BSM  
477 analysis (Supplementary Figure S8). Among the 16,836 multi-copy sets of the 15-taxon  
478 analysis, 1788 (10.6%) resolve all *Symbiodinium* sequences into an exclusive monophyletic  
479 clade and are topologically congruent at genus level with the reference species tree (i.e.  
480 contain co-orthologs but not paralogs) and were retained, while the remaining 15,048 failed  
481 one or both of these filtering criteria (i.e. contain presumed paralogs) and were not analysed  
482 further (Supplementary Figure S8). The percentages of missing data and parsimoniously  
483 informative sites in all filtered protein alignments for the 15-taxon set are detailed in  
484 Supplementary Table S26. For each filtered alignment, we used the corresponding coding-  
485 sequence alignment (codon alignment) generated using PAL2NAL<sup>114</sup> in the BSM analysis.  
486 Some predicted protein sequences in MMETSP<sup>115</sup> do not match their corresponding CDS,  
487 sometimes due to problematic translation and other times due to a frameshift. For these, we  
488 used MACSE<sup>116</sup> to derive the codon alignments.

489 We applied the branch-site model (BSM) implemented in the *codeml* program in  
490 PAML 4.9<sup>117</sup> to detect positive selection signal unique to the *Symbiodinium* lineage. BSMs  
491 allow the dN/dS ratio  $\omega$  to vary among both sites and branches, making it possible to infer  
492 selection at both. We computed two models: a null model with fixed  $\omega = 1$ , and an alternative  
493 model that estimates  $\omega$  in our defined foreground branches (here, the node that leads to all  
494 *Symbiodinium* lineages). We then compared the likelihoods of these two models to determine  
495 the better fit. To reduce false positives we applied *q*-value estimation for false discovery rate  
496 control, as implemented in R package *qvalue* to adjust *p*-values. Instances with an adjusted *p*

497  $\leq 0.05$  were considered significant. See Supplementary Note and Supplementary Figure S10  
498 for analysis of gene gain and gene loss in *Symbiodinium*.

## 499 **References**

- 500 1. Plaisance, L., Caley, M.J., Brainard, R.E. & Knowlton, N. The diversity of coral reefs:  
501 what are we missing? *PLoS ONE* **6**, e25026 (2011).
- 502 2. Falkowski, P.G., Dubinsky, Z., Muscatine, L. & Porter, J.W. Light and the  
503 bioenergetics of a symbiotic coral. *Bioscience* **34**, 705-709 (1984).
- 504 3. Muscatine, L. & Porter, J.W. Reef corals: mutualistic symbioses adapted to nutrient-  
505 poor environments. *Bioscience* **27**, 454-460 (1977).
- 506 4. Glynn, P.W. Coral reef bleaching: facts, hypotheses and implications. *Global Change*  
507 *Biol.* **2**, 495-509 (1996).
- 508 5. Hoegh-Guldberg, O. Climate change, coral bleaching and the future of the world's coral  
509 reefs. *Mar. Freshw. Res.* **50**, 839-866 (1999).
- 510 6. Albright, R. et al. Ocean acidification: linking science to management solutions using  
511 the Great Barrier Reef as a case study. *J. Environ. Manage.* **182**, 641-650 (2016).
- 512 7. Hughes, T.P. et al. Global warming and recurrent mass bleaching of corals. *Nature* **543**,  
513 373-377 (2017).
- 514 8. Lin, S. Genomic understanding of dinoflagellates. *Res. Microbiol.* **162**, 551-569 (2011).
- 515 9. LaJeunesse, T.C., Lambert, G., Andersen, R.A., Coffroth, M.A. & Galbraith, D.W.  
516 *Symbiodinium* (Pyrrophyta) genome sizes (DNA content) are smallest among  
517 dinoflagellates. *J. Phycol.* **41**, 880-886 (2005).
- 518 10. Barbrook, A.C., Voolstra, C.R. & Howe, C.J. The chloroplast genome of a  
519 *Symbiodinium* sp. Clade C3 isolate. *Protist* **165**, 1-13 (2014).
- 520 11. Howe, C.J., Nisbet, R.E.R. & Barbrook, A.C. The remarkable chloroplast genome of  
521 dinoflagellates. *J. Exp. Bot.* **59**, 1035-1045 (2008).
- 522 12. Zhang, Z., Green, B.R. & Cavalier-Smith, T. Single gene circles in dinoflagellate  
523 chloroplast genomes. *Nature* **400**, 155-159 (1999).
- 524 13. Nash, E.A., Nisbet, R.E.R., Barbrook, A.C. & Howe, C.J. Dinoflagellates: a  
525 mitochondrial genome all at sea. *Trends Genet.* **24**, 328-335 (2008).
- 526 14. Waller, R.F. & Jackson, C.J. Dinoflagellate mitochondrial genomes: stretching the rules  
527 of molecular biology. *BioEssays* **31**, 237-245 (2009).
- 528 15. Jackson, C.J. et al. Broad genomic and transcriptional analysis reveals a highly derived  
529 genome in dinoflagellate mitochondria. *BMC Biol.* **5**, 41 (2007).

- 530 16. Liew, Y.J., Li, Y., Baumgarten, S., Voolstra, C.R. & Aranda, M. Condition-specific  
531 RNA editing in the coral symbiont *Symbiodinium microadriaticum*. *PLoS Genet.* **13**,  
532 e1006619 (2017).
- 533 17. Aranda, M. et al. Genomes of coral dinoflagellate symbionts highlight evolutionary  
534 adaptations conducive to a symbiotic lifestyle. *Sci. Rep.* **6**, 39734 (2016).
- 535 18. Lin, S. et al. The *Symbiodinium kawagutii* genome illuminates dinoflagellate gene  
536 expression and coral symbiosis. *Science* **350**, 691-694 (2015).
- 537 19. Shoguchi, E. et al. Draft assembly of the *Symbiodinium minutum* nuclear genome  
538 reveals dinoflagellate gene structure. *Curr. Biol.* **23**, 1399-1408 (2013).
- 539 20. LaJeunesse, T.C. et al. Low symbiont diversity in southern Great Barrier Reef corals,  
540 relative to those of the Caribbean. *Limnol. Oceanogr.* **48**, 2046-2054 (2003).
- 541 21. LaJeunesse, T.C. "Species" radiations of symbiotic dinoflagellates in the Atlantic and  
542 Indo-Pacific since the Miocene-Pliocene transition. *Mol. Biol. Evol.* **22**, 570-581  
543 (2005).
- 544 22. Thornhill, D.J., Lewis, A.M., Wham, D.C. & LaJeunesse, T.C. Host-specialist lineages  
545 dominate the adaptive radiation of reef coral endosymbionts. *Evolution* **68**, 352-367  
546 (2014).
- 547 23. Tonk, L., Bongaerts, P., Sampayo, E.M. & Hoegh-Guldberg, O. *SymbioGBR*: a web-  
548 based database of *Symbiodinium* associated with cnidarian hosts on the Great Barrier  
549 Reef. *BMC Ecol.* **13**, 7 (2013).
- 550 24. Bongaerts, P. et al. Prevalent endosymbiont zonation shapes the depth distributions of  
551 scleractinian coral species. *R. Soc. Open Sci.* **2**, 140297 (2015).
- 552 25. Chan, Y.L. et al. Generalist dinoflagellate endosymbionts and host genotype diversity  
553 detected from mesophotic (67-100 m depths) coral *Leptoseris*. *BMC Ecol.* **9**, 21 (2009).
- 554 26. Yuyama, I., Higuchi, T. & Mezaki, T. *Symbiodinium kawagutii* (clade F) coats the  
555 surface of *Aropora solitaryensis*, resulting in the formation of a sheet-like crust. *Proc.*  
556 *13th Intl Coral Reef Symp.*, 49-56 (2016).
- 557 27. Kimura, M. A simple method for estimating evolutionary rates of base substitutions  
558 through comparative studies of nucleotide sequences. *J. Mol. Evol.* **16**, 111-120 (1980).
- 559 28. Vitte, C., Panaud, O. & Quesneville, H. LTR retrotransposons in rice (*Oryza sativa*, L.):  
560 recent burst amplifications followed by rapid DNA loss. *BMC Genomics* **8**, 218 (2007).
- 561 29. Tenailon, M.I., Hufford, M.B., Gaut, B.S. & Ross-Ibarra, J. Genome size and  
562 transposable element content as determined by high-throughput sequencing in maize  
563 and *Zea luxurians*. *Genome Biol. Evol.* **3**, 219-229 (2011).
- 564 30. Lee, S.I. & Kim, N.S. Transposable elements and genome size variations in plants.  
565 *Genomics Inform* **12**, 87-97 (2014).

- 566 31. LaJeunesse, T.C. Investigating the biodiversity, ecology, and phylogeny of  
567 endosymbiotic dinoflagellates in the genus *Symbiodinium* using the ITS region: in  
568 search of a “species” level marker. *J. Phycol.* **37**, 866-880 (2001).
- 569 32. Pochon, X. & Gates, R.D. A new *Symbiodinium* clade (Dinophyceae) from soritid  
570 foraminifera in Hawai'i. *Mol. Phylogenet. Evol.* **56**, 492-497 (2010).
- 571 33. van Oppen, M.J.H., Mieog, J.C., Sánchez, C.A. & Fabricius, K.E. Diversity of algal  
572 endosymbionts (zooxanthellae) in octocorals: the roles of geography and host  
573 relationships. *Mol. Ecol.* **14**, 2403-2417 (2005).
- 574 34. Mosavi, L.K., Cammett, T.J., Desrosiers, D.C. & Peng, Z.Y. The ankyrin repeat as  
575 molecular architecture for protein recognition. *Protein Sci.* **13**, 1435-1448 (2004).
- 576 35. Chan, C.X., Gross, J., Yoon, H.S. & Bhattacharya, D. Plastid origin and evolution: new  
577 models provide insights into old problems. *Plant Physiol.* **155**, 1552-1560 (2011).
- 578 36. Howe, C.J., Barbrook, A.C., Nisbet, R.E.R., Lockhart, P.J. & Larkum, A.W.D. The  
579 origin of plastids. *Phil. Trans. R. Soc. B* **363**, 2675-2685 (2008).
- 580 37. Yoon, H.S. et al. Tertiary endosymbiosis driven genome evolution in dinoflagellate  
581 algae. *Mol. Biol. Evol.* **22**, 1299-1308 (2005).
- 582 38. Rae, P.M. Hydroxymethyluracil in eukaryote DNA: a natural feature of the Pyrrophyta  
583 (dinoflagellates). *Science* **194**, 1062-1064 (1976).
- 584 39. Blank, R.J., Huss, V.R. & Kersten, W. Base composition of DNA from symbiotic  
585 dinoflagellates: a tool for phylogenetic classification. *Arch. Microbiol.* **149**, 515-520  
586 (1988).
- 587 40. ten Lohuis, M.R. & Miller, D.J. Hypermethylation at CpG-motifs in the dinoflagellates  
588 *Amphidinium carterae* (Dinophyceae) and *Symbiodinium microadriaticum*  
589 (Dinophyceae): evidence from restriction analyses, 5-azacytidine and ethionine  
590 treatment. *J. Phycol.* **34**, 152-159 (1998).
- 591 41. Maumus, F. et al. Potential impact of stress activated retrotransposons on genome  
592 evolution in a marine diatom. *BMC Genomics* **10**, 624 (2009).
- 593 42. Ito, H. et al. A stress-activated transposon in *Arabidopsis* induces transgenerational  
594 abscisic acid insensitivity. *Sci. Rep.* **6**, 23181 (2016).
- 595 43. Ramallo, E., Kalendar, R., Schulman, A.H. & Martínez-Izquierdo, J.A. *Remel*, a *Copia*  
596 retrotransposon in melon, is transcriptionally induced by UV light. *Plant Mol. Biol.* **66**,  
597 137-150 (2008).
- 598 44. Faure, E., Best-Belpomme, M. & Champion, S. X-irradiation activates the *Drosophila*  
599 1731 retrotransposon LTR and stimulates secretion of an extracellular factor that  
600 induces the 1731-LTR transcription in nonirradiated cells. *J Biochem* **120**, 313-319  
601 (1996).



- 602 45. Hunter, R.G., Gagnidze, K., McEwen, B.S. & Pfaff, D.W. Stress and the dynamic  
603 genome: steroids, epigenetics, and the transposome. *Proc. Natl. Acad. Sci. U. S. A.* **112**,  
604 6828-6833 (2015).
- 605 46. Weynberg, K.D. et al. Prevalent and persistent viral infection in cultures of the coral  
606 algal endosymbiont *Symbiodinium*. *Coral Reefs* **36**, 773-784 (2017).
- 607 47. Kimura, M. Preponderance of synonymous changes as evidence for the neutral theory  
608 of molecular evolution. *Nature* **267**, 275-276 (1977).
- 609 48. Price, D.C. & Bhattacharya, D. Robust Dinoflagellata phylogeny inferred from public  
610 transcriptome databases. *J. Phycol.* **53**, 725-729 (2017).
- 611 49. Takahashi, S., Yoshioka-Nishimura, M., Nanba, D. & Badger, M.R. Thermal  
612 acclimation of the symbiotic alga *Symbiodinium* spp. alleviates photobleaching under  
613 heat stress. *Plant Physiol.* **161**, 477-485 (2013).
- 614 50. Lesser, M.P. Elevated temperatures and ultraviolet radiation cause oxidative stress and  
615 inhibit photosynthesis in symbiotic dinoflagellates. *Limnol. Oceanogr.* **41**, 271-283  
616 (1996).
- 617 51. Lesser, M.P. Oxidative stress causes coral bleaching during exposure to elevated  
618 temperatures. *Coral Reefs* **16**, 187-192 (1997).
- 619 52. Barott, K.L., Venn, A.A., Perez, S.O., Tambutte, S. & Tresguerres, M. Coral host cells  
620 acidify symbiotic algal microenvironment to promote photosynthesis. *Proc. Natl. Acad.*  
621 *Sci. U. S. A.* **112**, 607-612 (2015).
- 622 53. Zoccola, D. et al. Bicarbonate transporters in corals point towards a key step in the  
623 evolution of cnidarian calcification. *Sci. Rep.* **5**, 9983 (2015).
- 624 54. Shinzato, C. et al. Using the *Acropora digitifera* genome to understand coral responses  
625 to environmental change. *Nature* **476**, 320-323 (2011).
- 626 55. Davy, S.K., Allemand, D. & Weis, V.M. Cell biology of cnidarian-dinoflagellate  
627 symbiosis. *Microbiol. Mol. Biol. Rev.* **76**, 229-261 (2012).
- 628 56. Lin, K.L., Wang, J.T. & Fang, L.S. Participation of glycoproteins of zooxanthellal cell  
629 walls in the establishment of a symbiotic relationship with the sea anemone, *Aiptasia*  
630 *pulchella*. *Zool. Stud.* **39**, 172-178 (2000).
- 631 57. Markell, D.A., Trench, R.K. & Iglesias-Prieto, R. Macromolecules associated with the  
632 cell walls of symbiotic dinoflagellates. *Symbiosis* **12**, 19-31 (1992).
- 633 58. Markell, D.A. & Trench, R.K. Macromolecules exuded by symbiotic dinoflagellates in  
634 culture: amino acid and sugar composition. *J. Phycol.* **29**, 64-68 (1993).
- 635 59. Markell, D.A. & Wood-Charlson, E.M. Immunocytochemical evidence that symbiotic  
636 algae secrete potential recognition signal molecules *in hospite*. *Mar. Biol.* **157**, 1105-  
637 1111 (2010).

- 638 60. Logan, D.D.K., LaFlamme, A.C., Weis, V.M. & Davy, S.K. Flow-cytometric  
639 characterization of the cell-surface glycans of symbiotic dinoflagellates (*Symbiodinium*  
640 spp.). *J. Phycol.* **46**, 525-533 (2010).
- 641 61. Wood-Charlson, E.M., Hollingsworth, L.L., Krupp, D.A. & Weis, V.M. Lectin/glycan  
642 interactions play a role in recognition in a coral/dinoflagellate symbiosis. *Cell.*  
643 *Microbiol.* **8**, 1985-1993 (2006).
- 644 62. Neubauer, E.F. et al. A diverse host thrombospondin-type-1 repeat protein repertoire  
645 promotes symbiont colonization during establishment of cnidarian-dinoflagellate  
646 symbiosis. *eLife* **6**, e24494 (2017).
- 647 63. Ljungman, M. The transcription stress response. *Cell Cycle* **6**, 2252-2257 (2007).
- 648 64. Mohamed, A.R. et al. The transcriptomic response of the coral *Acropora digitifera* to a  
649 competent *Symbiodinium* strain: the symbiosome as an arrested early phagosome. *Mol.*  
650 *Ecol.* **25**, 3127-3141 (2016).
- 651 65. Fitt, W.K. & Trench, R.K. Endocytosis of the symbiotic dinoflagellate *Symbiodinium*  
652 *microadriaticum* Freudenthal by endodermal cells of the scyphistomae of *Cassiopeia*  
653 *xamachana* and resistance of the algae to host digestion. *J. Cell Sci.* **64**, 195-212  
654 (1983).
- 655 66. Freudenthal, H.D. *Symbiodinium* gen. nov. and *Symbiodinium microadriaticum* sp.  
656 nov., a zooxanthella: taxonomy, life cycle, and morphology. *J. Protozool.* **9**, 45-52  
657 (1962).
- 658 67. Taylor, F.J.R. Implications and extensions of the serial endosymbiosis theory of the  
659 origin of eukaryotes. *Taxon* **23**, 229-258 (1974).
- 660 68. Trench, R.K. Diversity of symbiotic dinoflagellates and the evolution of microalgal-  
661 invertebrate symbioses. *Proc. 8th Intl Coral Reef Symp.* **2**, 1275-1286 (1997).
- 662 69. de Visser, J.A. & Elena, S.F. The evolution of sex: empirical insights into the roles of  
663 epistasis and drift. *Nat. Rev. Genet.* **8**, 139-149 (2007).
- 664 70. Baillie, B.K., Belda-Baillie, C.A. & Maruyama, T. Conspecificity and Indo-Pacific  
665 distribution of *Symbiodinium* genotypes (Dinophyceae) from giant clams. *J. Phycol.* **36**,  
666 1153-1161 (2000).
- 667 71. Pettay, D.T., Wham, D.C., Pinzón, J.H. & LaJeunesse, T.C. Genotypic diversity and  
668 spatial-temporal distribution of *Symbiodinium* clones in an abundant reef coral. *Mol.*  
669 *Ecol.* **20**, 5197-5212 (2011).
- 670 72. Reichman, J.R., Wilcox, T.P. & Vize, P.D. PCP gene family in *Symbiodinium* from  
671 *Hippopus hippopus*: low levels of concerted evolution, isoform diversity, and spectral  
672 tuning of chromophores. *Mol. Biol. Evol.* **20**, 2143-2154 (2003).
- 673 73. Santos, S.R. & Coffroth, M.A. Molecular genetic evidence that dinoflagellates  
674 belonging to the genus *Symbiodinium* Freudenthal are haploid. *Biol. Bull.* **204**, 10-20  
675 (2003).

- 676 74. Levin, R.A. et al. Sex, scavengers, and chaperones: transcriptome secrets of divergent  
677 *Symbiodinium* thermal tolerances. *Mol. Biol. Evol.* **33**, 2201-2215 (2016).
- 678 75. Schurko, A.M. & Logsdon, J.M., Jr. Using a meiosis detection toolkit to investigate  
679 ancient asexual "scandals" and the evolution of sex. *BioEssays* **30**, 579-589 (2008).
- 680 76. Chi, J., Parrow, M.W. & Dunthorn, M. Cryptic sex in *Symbiodinium* (Alveolata,  
681 Dinoflagellata) is supported by an inventory of meiotic genes. *J. Eukaryot. Microbiol.*  
682 **61**, 322-327 (2014).
- 683 77. Malik, S.B., Pightling, A.W., Stefaniak, L.M., Schurko, A.M. & Logsdon, J.M., Jr. An  
684 expanded inventory of conserved meiotic genes provides evidence for sex in  
685 *Trichomonas vaginalis*. *PLoS ONE* **3**, e2879 (2008).
- 686 78. Speijer, D., Lukeš, J. & Eliáš, M. Sex is a ubiquitous, ancient, and inherent attribute of  
687 eukaryotic life. *Proc. Natl. Acad. Sci. U. S. A.* **112**, 8827-8834 (2015).
- 688 79. Griffiths, D.J., Barbet, N.C., McCready, S., Lehmann, A.R. & Carr, A.M. Fission yeast  
689 *rad17*: a homologue of budding yeast *RAD24* that shares regions of sequence similarity  
690 with DNA polymerase accessory proteins. *EMBO J.* **14**, 5812-5823 (1995).
- 691 80. Banaszak, A.T., LaJeunesse, T.C. & Trench, R.K. The synthesis of mycosporine-like  
692 amino acids (MAAs) by cultured, symbiotic dinoflagellates. *J. Exp. Mar. Biol. Ecol.*  
693 **249**, 219-233 (2000).
- 694 81. Rosic, N.N. & Dove, S. Mycosporine-like amino acids from coral dinoflagellates. *Appl.*  
695 *Environ. Microbiol.* **77**, 8478-8486 (2011).
- 696 82. Balskus, E.P. & Walsh, C.T. The genetic and molecular basis for sunscreen  
697 biosynthesis in cyanobacteria. *Science* **329**, 1653-1656 (2010).
- 698 83. Gao, Q. & Garcia-Pichel, F. Microbial ultraviolet sunscreens. *Nat. Rev. Microbiol.* **9**,  
699 791-802 (2011).
- 700 84. D'Agostino, P.M. et al. Comparative profiling and discovery of novel glycosylated  
701 mycosporine-like amino acids in two strains of the cyanobacterium *Scytonema* cf.  
702 *crispum*. *Appl. Environ. Microbiol.* **82**, 5951-5959 (2016).
- 703 85. Rastogi, R.P., Sonani, R.R. & Madamwar, D. Cyanobacterial sunscreen scytonemin:  
704 role in photoprotection and biomedical research. *Appl. Biochem. Biotechnol.* **176**, 1551-  
705 1563 (2015).
- 706 86. Garcia-Pichel, F. & Castenholz, R.W. Characterization and biological implications of  
707 scytonemin, a cyanobacterial sheath pigment. *J. Phycol.* **27**, 395-409 (1991).
- 708 87. Balskus, E.P. & Walsh, C.T. Investigating the initial steps in the biosynthesis of  
709 cyanobacterial sunscreen scytonemin. *J. Am. Chem. Soc.* **130**, 15260-15261 (2008).
- 710 88. Soule, T., Stout, V., Swingley, W.D., Meeks, J.C. & Garcia-Pichel, F. Molecular  
711 genetics and genomic analysis of scytonemin biosynthesis in *Nostoc punctiforme*  
712 ATCC 29133. *J. Bacteriol.* **189**, 4465-4472 (2007).

- 713 89. Soule, T., Garcia-Pichel, F. & Stout, V. Gene expression patterns associated with the  
714 biosynthesis of the sunscreen scytonemin in *Nostoc punctiforme* ATCC 29133 in  
715 response to UVA radiation. *J. Bacteriol.* **191**, 4639-4646 (2009).
- 716 90. Shinzato, C., Mungpakdee, S., Satoh, N. & Shoguchi, E. A genomic approach to coral-  
717 dinoflagellate symbiosis: studies of *Acropora digitifera* and *Symbiodinium minutum*.  
718 *Front. Microbiol.* **5**, 336 (2014).
- 719 91. Frankowiak, K. et al. Photosymbiosis and the expansion of shallow-water corals. *Sci.*  
720 *Adv.* **2**, e1601122 (2016).
- 721 92. Bongaerts, P. et al. Deep reefs are not universal refuges: reseeded potential varies  
722 among coral species. *Sci. Adv.* **3**, e1602373 (2017).
- 723 93. Thomas, L., Kennington, W.J., Evans, R.D., Kendrick, G.A. & Stat, M. Restricted gene  
724 flow and local adaptation highlight the vulnerability of high-latitude reefs to rapid  
725 environmental change. *Global Change Biol.* **23**, 2197-2205 (2017).
- 726 94. Howells, E.J. et al. Coral thermal tolerance shaped by local adaptation of  
727 photosymbionts. *Nat. Clim. Change* **2**, 116-120 (2012).
- 728 95. Bankevich, A. et al. SPAdes: a new genome assembly algorithm and its applications to  
729 single-cell sequencing. *J. Comput. Biol.* **19**, 455-477 (2012).
- 730 96. Gnerre, S. et al. High-quality draft assemblies of mammalian genomes from massively  
731 parallel sequence data. *Proc. Natl. Acad. Sci. U. S. A.* **108**, 1513-1518 (2011).
- 732 97. Haas, B.J. et al. Improving the *Arabidopsis* genome annotation using maximal  
733 transcript alignment assemblies. *Nucleic Acids Res.* **31**, 5654-5666 (2003).
- 734 98. Stanke, M. et al. AUGUSTUS: *ab initio* prediction of alternative transcripts. *Nucleic*  
735 *Acids Res.* **34**, W435-439 (2006).
- 736 99. Holt, C. & Yandell, M. MAKER2: an annotation pipeline and genome-database  
737 management tool for second-generation genome projects. *BMC Bioinformatics* **12**, 491  
738 (2011).
- 739 100. Lomsadze, A., Ter-Hovhannisyan, V., Chernoff, Y.O. & Borodovsky, M. Gene  
740 identification in novel eukaryotic genomes by self-training algorithm. *Nucleic Acids*  
741 *Res.* **33**, 6494-6506 (2005).
- 742 101. Korf, I. Gene finding in novel genomes. *BMC Bioinformatics* **5**, 59 (2004).
- 743 102. Haas, B.J. et al. Automated eukaryotic gene structure annotation using  
744 EVIDENCEModeler and the Program to Assemble Spliced Alignments. *Genome Biol.* **9**,  
745 R7 (2008).
- 746 103. Parra, G., Bradnam, K. & Korf, I. CEGMA: a pipeline to accurately annotate core  
747 genes in eukaryotic genomes. *Bioinformatics* **23**, 1061-1067 (2007).

- 748 104. Simão, F.A., Waterhouse, R.M., Ioannidis, P., Kriventseva, E.V. & Zdobnov, E.M.  
749 BUSCO: assessing genome assembly and annotation completeness with single-copy  
750 orthologs. *Bioinformatics* **31**, 3210-3212 (2015).
- 751 105. Baumgarten, S. et al. The genome of *Aiptasia*, a sea anemone model for coral  
752 symbiosis. *Proc. Natl. Acad. Sci. U. S. A.* **112**, 11893-11898 (2015).
- 753 106. Wang, Y. et al. MCScanX: a toolkit for detection and evolutionary analysis of gene  
754 synteny and collinearity. *Nucleic Acids Res.* **40**, e49 (2012).
- 755 107. Emms, D.M. & Kelly, S. OrthoFinder: solving fundamental biases in whole genome  
756 comparisons dramatically improves orthogroup inference accuracy. *Genome Biol.* **16**,  
757 157 (2015).
- 758 108. Harlow, T.J., Gogarten, J.P. & Ragan, M.A. A hybrid clustering approach to  
759 recognition of protein families in 114 microbial genomes. *BMC Bioinformatics* **5**, 45  
760 (2004).
- 761 109. Beiko, R.G., Harlow, T.J. & Ragan, M.A. Highways of gene sharing in prokaryotes.  
762 *Proc. Natl. Acad. Sci. U. S. A.* **102**, 14332-14337 (2005).
- 763 110. Benjamini, Y. & Yekutieli, D. The control of the false discovery rate in multiple testing  
764 under dependency. *The Annals of Statistics* **29**, 1165-1188 (2001).
- 765 111. Katoh, K., Kuma, K.-I., Toh, H. & Miyata, T. MAFFT version 5: improvement in  
766 accuracy of multiple sequence alignment. *Nucleic Acids Res.* **33**, 511-518 (2005).
- 767 112. Capella-Gutiérrez, S., Silla-Martínez, J.M. & Gabaldón, T. trimAl: a tool for automated  
768 alignment trimming in large-scale phylogenetic analyses. *Bioinformatics* **25**, 1972-1973  
769 (2009).
- 770 113. Nguyen, L.T., Schmidt, H.A., von Haeseler, A. & Minh, B.Q. IQ-TREE: a fast and  
771 effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Mol.*  
772 *Biol. Evol.* **32**, 268-274 (2015).
- 773 114. Suyama, M., Torrents, D. & Bork, P. PAL2NAL: robust conversion of protein sequence  
774 alignments into the corresponding codon alignments. *Nucleic Acids Res.* **34**, W609-612  
775 (2006).
- 776 115. Keeling, P.J. et al. The Marine Microbial Eukaryote Transcriptome Sequencing Project  
777 (MMETSP): illuminating the functional diversity of eukaryotic life in the oceans  
778 through transcriptome sequencing. *PLoS Biol.* **12**, e1001889 (2014).
- 779 116. Ranwez, V., Harispe, S., Delsuc, F. & Douzery, E.J. MACSE: Multiple Alignment of  
780 Coding SEquences accounting for frameshifts and stop codons. *PLoS ONE* **6**, e22594  
781 (2011).
- 782 117. Yang, Z. PAML 4: phylogenetic analysis by maximum likelihood. *Mol. Biol. Evol.* **24**,  
783 1586-1591 (2007).

784

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## 793 **Author contributions**

794 HL, MAR and CXC conceived the study and designed the experiments. HL, TGS, RGP and  
795 CXC conducted all computational analyses. VHB and BL established the algal cultures and  
796 extracted the DNA. HL, TGS, RGP, IC, MAR and CXC analysed and interpreted the results.  
797 HL and CXC prepared all figures, tables, and the first draft of this manuscript. SF and CRV  
798 provided analytical tools and scripts. HL, TGS, MAR and CXC wrote the manuscript. PB, IC,  
799 DGB, DJM, MJHvO and CRV assisted in experimental design and writing of the manuscript.  
800 All authors reviewed, commented on and approved the final manuscript.

## 801 **Data availability**

802 All sequence data generated from this study will be available at the NCBI Short Read  
803 Archive (accessions XXXX and XXXX). Assembled genomes, predicted gene models and  
804 proteins are available at <https://cloudstor.aarnet.edu.au/plus/index.php/s/6yziMf2ygWjGu0L>.

## 805 **Code availability**

806 The customised scripts for AUGUSTUS and PASA used in this study were previously  
807 published in Aranda et al.<sup>17</sup>; they are available at <http://smic.reefgenomics.org/download/>.

## 808 **Competing financial interests**

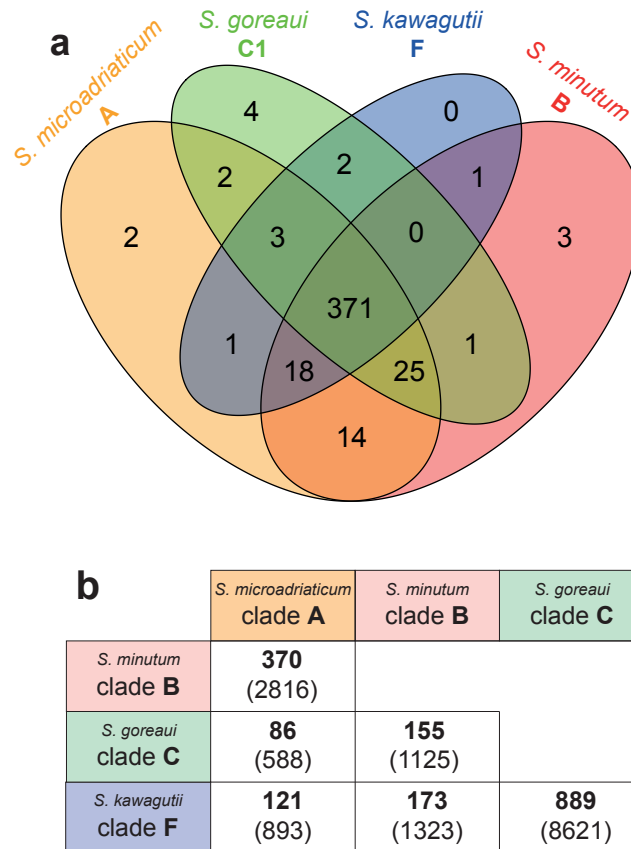
809 The authors declare no competing financial interests.

## 810 **Figure legends**

811 **Figure 1. Comparison of *Symbiodinium* genomes.** (a) Number of recovered core eukaryote  
812 genes in each genome based on CEGMA. (b) Number of identified syntenic collinear blocks  
813 (and the corresponding number of implicated genes) between each genome pair.

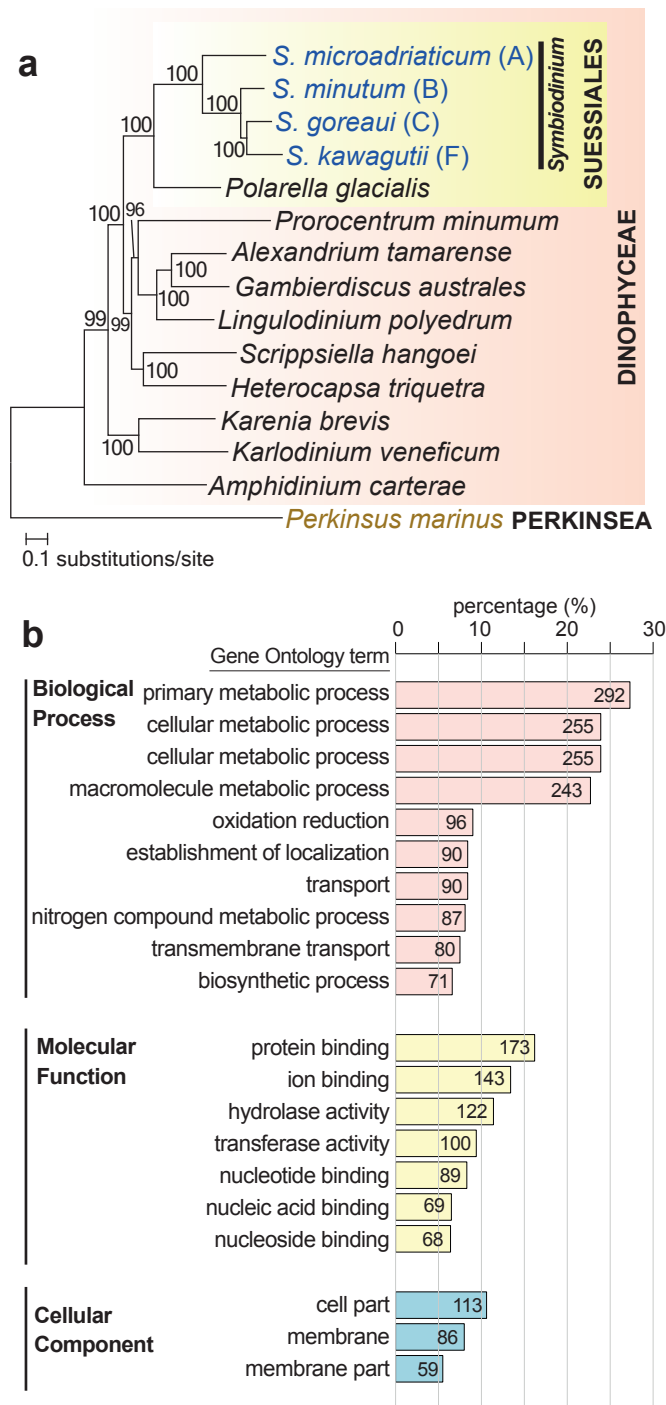
814 **Figure 2. Testing for positive selection acting on *Symbiodinium* genomes.** (a) The  
815 reference 15-species supertree of *Symbiodinium*, dinoflagellates and *Perkinsus marinus* (as  
816 outgroup) based on single-copy orthologous genes. Support based on 2000 rapid bootstraps is  
817 shown on each internal node, and the branch length is the number of substitutions per site. (b)  
818 Percentage of positively selected genes with annotated GO (level 3) terms in *Symbiodinium*,  
819 shown for principal hierarchies Biological Function, Molecular Function and Cellular  
820 Component.

821 **Figure 3. Recovery of genes in *Symbiodinium*.** (a) Meiosis-related genes recovered in the  
822 genomes of *S. microadriaticum* (Clade A), *S. minutum* (Clade B), *S. goreaui* (Clade C) and *S.*  
823 *kawagutii* (Clade F). (b) Scytonemin biosynthesis genes in *Symbiodinium* genomes relative to  
824 the coral *Acropora digitifera*, sea anemone *Nematostella vectensis*, hydra (*Hydra*  
825 *magnipapillata*) and the green plant *Arabidopsis thaliana*. The order of the 18-gene cluster in  
826 the cyanobacteria *Nostoc punctiforme* is used as a reference, with presence (+) and absence (-  
827 ) of a gene in each species are shown. Figure modified from Shinzato et al.<sup>90</sup>.

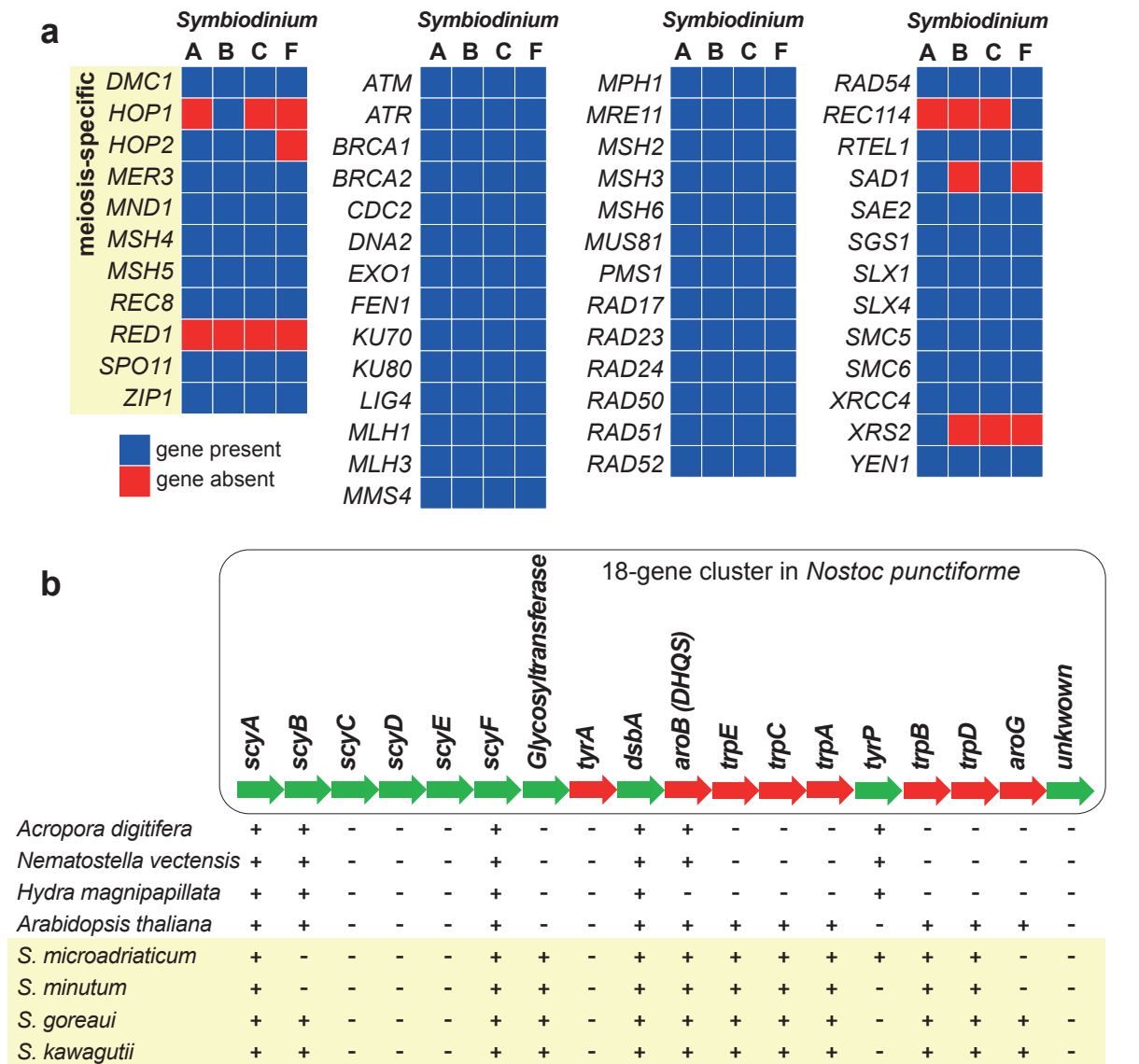


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