

# 1 **A strategy to exploit surrogate sire technology in** 2 **livestock breeding programs**

3

4 Paolo Gottardo<sup>1</sup>, Gregor Gorjanc<sup>1</sup>, Mara Battagin<sup>1</sup>, R Chris Gaynor<sup>1</sup>, Janez Jenko<sup>1</sup>,  
5 Roger Ros-Freixedes<sup>1</sup>, C Bruce A Whitelaw<sup>1</sup>, Alan J Mileham<sup>2</sup>, William O Herring<sup>3</sup>,  
6 John M Hickey<sup>1§</sup>

7

8 <sup>1</sup>The Roslin Institute and Royal (Dick) School of Veterinary Studies, The University  
9 of Edinburgh, Easter Bush, Midlothian, Scotland, UK

10 <sup>2</sup>Genus PLC, 1525 River Rd., DeForest, Wisconsin 53532, USA

11 <sup>3</sup>The Pig Improvement Company, Genus PLC, 100 Bluegrass Commons Blvd., Ste  
12 2200, Hendersonville, Tennessee 37075, USA

13 <sup>§</sup>Corresponding author

14

15 Email addresses:

16 PG: [paolo.gottardo@roslin.ed.ac.uk](mailto:paolo.gottardo@roslin.ed.ac.uk)

17 GG: [gregor.gorjanc@roslin.ed.ac.uk](mailto:gregor.gorjanc@roslin.ed.ac.uk)

18 MB: [mara.battagin@roslin.ed.ac.uk](mailto:mara.battagin@roslin.ed.ac.uk)

19 RCG: [chris.gaynor@roslin.ed.ac.uk](mailto:chris.gaynor@roslin.ed.ac.uk)

20 JJ: [janez.jenko@roslin.ed.ac.uk](mailto:janez.jenko@roslin.ed.ac.uk)

21 RRF: [roger.ros@roslin.ed.ac.uk](mailto:roger.ros@roslin.ed.ac.uk)

22 CBAW: [bruce.whitelaw@roslin.ed.ac.uk](mailto:bruce.whitelaw@roslin.ed.ac.uk)

23 AJM: [Alan.Mileham@genusplc.com](mailto:Alan.Mileham@genusplc.com)

24 WOH: [William.Herring@genusplc.com](mailto:William.Herring@genusplc.com)

25 JMH: [john.hickey@roslin.ed.ac.uk](mailto:john.hickey@roslin.ed.ac.uk)

26

## 27 **Abstract**

28 In this work, we performed simulations to develop and test a strategy for exploiting  
29 surrogate sire technology in animal breeding programs. Surrogate sire technology  
30 allows the creation of males that lack their own germline cells, but have transplanted  
31 spermatogonial stem cells from donor males. With this technology, a single elite male  
32 donor could give rise to huge numbers of progeny, potentially as much as all the  
33 production animals in a particular time period.

34 One hundred replicates of various scenarios were performed. Scenarios followed a  
35 common overall structure but differed in the strategy used to identify elite donors and  
36 how these donors were used in the product development part.

37 The results of this study showed that using surrogate sire technology would  
38 significantly increase the genetic merit of commercial sires, by as much as 6.5 to 9.2  
39 years' worth of genetic gain compared to a conventional breeding program. The  
40 simulations suggested that a strategy involving three stages (an initial genomic test  
41 followed by two subsequent progeny tests) was the most effective of all the strategies  
42 tested.

43 The use of one or a handful of elite donors to generate the production animals would  
44 be very different to current practice. While the results demonstrate the great potential  
45 of surrogate sire technology there are considerable risks but also other opportunities.  
46 Practical implementation of surrogate sire technology would need to account for  
47 these.

## 48 **Introduction**

49 In this study, we performed simulations to develop a strategy for exploiting surrogate  
50 sire technology [1-2] in animal breeding programs (Fig. 1). Surrogate sire technology  
51 allows the creation of males that lack their own germline cells, but have transplanted  
52 spermatogonial stem cells from other donor males. The concept requires the  
53 production of recipient males with an ablated germ line. Rodent males can have their  
54 germline ablated using chemotoxic drugs or localised irradiation of the testes, but,  
55 importantly for use in livestock breeding, this ablation is incomplete and recipient  
56 sperm output is mixture of donor and recipient cells [3]. The mammalian NANOS2  
57 gene seems to be absolutely required for the maintenance of germ line cells in males  
58 only [4]. In mice, Nanos 2 knock out males the testes completely lack germ-line cells,  
59 but there is no effect in females [4]. NANOS 2 knock out pigs have been produced  
60 using CRISP/Cas9 gene editing [1] and boars homozygous for the knockout likely  
61 provide ideal recipients for the surrogate sire concept."

62 With this technology, a single elite male donor could give rise to huge  
63 numbers of progeny, potentially as much as all the production animals in a particular  
64 time period. This potential offers many advantages. Firstly, it would reduce the  
65 genetic lag between the elite nucleus animals and the production animals. Secondly, it  
66 could enable better matching of specific management plans to the genetics. Thirdly, as  
67 we outline in the discussion it could enable exploitation of combining ability. The  
68 latter could increase production on farm and increase investment and innovation in  
69 breeding by enabling a greater ability to protect intellectual property.

70 Typically, animal breeding programs are implicitly or explicitly organized in  
71 pyramid structures with layers (Fig. 2). The top layer is the nucleus, which is

72 improved using recurrent selection. Nowadays most selection decisions are made  
73 using genomic based testing rather than traditional phenotype based testing [4–8]. The  
74 middle layer is the multiplication, where the nucleus genetics is multiplied and  
75 sometimes crosses between purebred lines are produced. The base layer is the  
76 commercial sector, where the majority of animals are kept for production. The  
77 commercial producers often make a final cross between the terminal line sires and the  
78 maternal line dams.

79         The need to generate huge numbers of production animals and the limited  
80 number of progeny that a male can produce means that large numbers of nucleus  
81 animals must contribute genetics to the subsequent layers and that one to several  
82 generations are required for multiplication. These factors give rise to a genetic lag, a  
83 difference in genetic mean between the nucleus and commercial layers. This lag can  
84 also be represented with the number of years of genetic gain [9], e.g., ~4 years in a pig  
85 breeding program. Surrogate sire technology would allow a single elite nucleus male  
86 to give rise to very large numbers of commercial animals, by donating spermatogonial  
87 stem cells to its commercial surrogates [1]. This could shorten the lag between the  
88 nucleus, multiplication, and commercial layers.

89         Using surrogate sire technology in this way would require that animal  
90 breeding programs identify elite donor males and create surrogate sires. This process  
91 should take place in a sufficiently small amount of time so that the extra genetic gain  
92 would not be significantly reduced by the extra time required for the identification of  
93 donors and creation of surrogate sires.

94         A restructured animal breeding program with surrogate sire technology would  
95 be conceptually similar to a plant breeding program that produces clonally propagated

96 individual lines or inbred lines or hybrid lines (Fig. 2). These programs seek: (i) to  
97 identify the best individual (note: here we take individual to mean clonal, inbred or  
98 hybrid lines), or a handful of individuals, from a population of individuals; and (ii) to  
99 disseminate this individual very widely in the commercial layer [10]. To identify the  
100 best individual, plant breeders typically use multiple stage testing and selection. As  
101 the breeding program progresses through these stages the number of individuals being  
102 tested is reduced and the precision of these tests increases. The small number of  
103 individuals in the final stages are intensively tested in large replicated experiments  
104 that are repeated across several environments and years. This ensures that the  
105 commercially released individual is well characterized and carries a minimal risk of  
106 major undetected weakness. This is necessary because this individual will have a huge  
107 footprint in the commercial layer. Similar levels of evaluation would be needed with  
108 surrogate sire technology in animal breeding programs.

109         The objective of this study was to develop a strategy for exploiting surrogate  
110 sire technology in animal breeding programs. This strategy involved a subtle, but  
111 important, reorganisation to combine components of traditional animal and plant  
112 breeding programs. The reorganization is similar to the two-part breeding program  
113 that we recently proposed for the incorporation of genomic selection into plant  
114 breeding programs [11]. The reorganization involves an explicit partitioning of a  
115 breeding program into a population improvement component and a product  
116 development component. The population improvement component is similar to the  
117 currently used recurrent genomic selection in many animal breeding nucleus  
118 populations. The product development component is similar to traditional plant  
119 breeding programs and involves a number of stages of testing to identify the elite  
120 donors. The product development component could make use of testing for combining

121 ability, if that was appropriate for the particular species of interest.

122           With a focus on application in pig breeding, several alternative versions of the  
123 reorganized breeding program were compared to different variants of a conventional  
124 breeding program using simulation. The alternative versions varied: (i) the number of  
125 stages of testing; (ii) the number of donor candidates tested at subsequent stages; (iii)  
126 the accuracy of the genomic test at the first stage; and (iv) the accuracy of progeny  
127 test in later stages. The results showed that using surrogate sire technology would  
128 significantly increase the genetic merit of commercial sires, by as much as between  
129 6.5 and 9.20 years' worth of genetic gain compared to different variants of a  
130 conventional breeding program. The simulations suggested that an identification  
131 strategy involving three stages (a genomic test followed by two subsequent progeny  
132 tests) was the most effective of all the strategies tested. The use of one or a handful of  
133 elite donors to generate the production animals would be very different to current  
134 practice. While the results demonstrate the great potential of surrogate sire technology  
135 there are considerable risks and these are discussed.

136

## 137 **Methods**

138           Simulation was used to evaluate the impact of a strategy for exploiting  
139 surrogate sire technology in animal breeding programs. One hundred replicates of  
140 various scenarios were performed. Scenarios followed a common overall structure but  
141 differed in the strategy used to identify elite donors and how these donors were used  
142 (Fig. 3, 4).

143           Conceptually, the simulation scheme was divided into historical and future  
144 phases. The historical phase represented historical evolution and recent animal  
145 breeding efforts up to the present day, under the assumption that animal populations  
146 have evolved for tens of thousands of years, followed by 22 recent generations of  
147 modern animal breeding with selection on genomic breeding values in a nucleus  
148 population. The future phase represented 20 future generations of modern animal  
149 breeding, with selection on genomic breeding values in a nucleus population that  
150 subsequently supplied genetic improvement to multiplication and commercial layers.  
151 The historical animal breeding generations were denoted -21 to 0 and the future  
152 animal breeding generations were denoted 1 to 20. The multiplier and commercial  
153 layers were not explicitly simulated but were instead represented with the average  
154 genetic merit of nucleus males that would give rise to multiplication and commercial  
155 animals while accounting for the time lag. Specifically, we only focused on a breeding  
156 program that produced terminal males in a scheme that closely resembled a pig  
157 breeding program.

158 Simulations involved the following four steps:

159 (i)    Generating genome,

- 160 (ii) Generating a quantitative trait and breeding values,  
161 (iii) Generating an animal breeding program,  
162 (iv) Selection and dissemination to the commercial layer with the conventional or  
163 surrogate sires strategy.

164 Results are presented as the mean of one hundred replicates for each scenario  
165 and encompass the genetic merit of nucleus males that would give rise to commercial  
166 animals at a given time point.

## 167 **Genome**

168 Whole-genome sequences were generated using the Markovian Coalescent  
169 Simulator (MaCS) [12] and AlphaSim [13] for 400 base haplotypes for each of ten 10  
170 chromosomes. Chromosomes (each 100 cM long and comprising  $10^8$  base pairs) were  
171 simulated using a per site mutation rate of  $2.5 \times 10^{-8}$ , a per site recombination rate of  
172  $1.0 \times 10^{-8}$ , and an effective population size ( $N_e$ ) that varied over time in accordance  
173 with estimates that are representative of livestock populations [e.g., 14–17] as  
174 follows:  $N_e$  was set to 100 in the final generation of the coalescent simulation, to  $N_e =$   
175 1256, 1000 years ago, to  $N_e = 4350$ , 10,000 years ago, and to  $N_e = 43,500$ , 100,000 years  
176 ago, with linear changes in between these time-points. The resulting sequences had  
177 approximately 540,000 segregating sites.

## 178 **Quantitative trait**

179 A quantitative trait was simulated by randomly sampling 10,000 causal loci  
180 from the genome in the base population, with the restriction that 1,000 were sampled  
181 from each of the 10 chromosomes. For these loci, the allele substitution effect was  
182 randomly sampled from a normal distribution with a mean of 0 and standard deviation  
183 of 0.01 (1.0 divided by the square root of the number of loci).



## 184 **Breeding values**

185 True breeding values were computed as a sum of effects at causal loci. To  
186 simulate selection without the full computational burden and complexity of simulating  
187 training sets and estimation with best linear unbiased prediction, we simulated pseudo  
188 estimates of breeding values by adding a level of noise to true breeding values.  
189 Different levels of noise were added to achieve a targeted accuracy. For the genomic  
190 tests we simulated accuracies of 0.50, 0.70 and 0.90. For the progeny tests we  
191 simulated accuracies as a function of the number of progeny [24] used in the different  
192 scenarios (described below).

## 193 **Breeding program**

194 A pedigree of 42 generations for the nucleus population was simulated. Each  
195 generation included 1,000 (**SmallScenario**) or 5,000 (**BigScenario**) individuals with  
196 equal sex ratio. The different numbers of individuals were used to quantify impact of  
197 nucleus population size on the benefit of surrogate sire technology. All females (500  
198 for the SmallScenario or 2,500 for the BigScenario) and 50 males were selected as the  
199 parents of each generation. This selection was based on a genomic test. In the first  
200 generation of the recent historical animal breeding population (i.e., generation -22),  
201 the chromosomes of each individual were sampled from the 400 base haplotypes. In  
202 later generations (i.e., generations -21 to 20), the chromosomes of each individual  
203 were sampled from parental chromosomes with recombination (assuming no  
204 interference). A recombination rate of 1 Morgan per chromosome was used, resulting  
205 in a 10 Morgan genome.

## 206 **Scenarios**

207 Two different strategies were used to identify males from the nucleus who

208 would give rise to commercial animals, either through conventional multiplication or  
209 surrogate sires. The conventional multiplication strategy used the top 50, 200, or 500  
210 males in both the SmallScenario and the BigScenario. Males were chosen based on a  
211 genomic test. The surrogate sires strategy used multi-stage testing. Males were chosen  
212 based on an initial genomic test (S0), followed by one or two subsequent progeny  
213 tests (S1 and S2). As is the case with plant breeding programs, as the testing  
214 progressed through the stages we reduced the number of tested individuals and  
215 increased accuracy of tests. Based on the tests the best individual or set of individuals  
216 were identified and used as elite donors of spermatogonial stem cells to surrogate  
217 sires.

218 To quantify the impact of different amounts of testing resources and different  
219 allocation of these resources we simulated different accuracies of the genomic test at  
220 S0, different numbers of donor candidates tested with different number of progeny at  
221 S1 and S2. At S0 we simulated a genomic test with an accuracy of 0.50, 0.70, and  
222 0.90. To ensure that each breeding program had the same costs, we assumed that a  
223 total of 14,000 progeny were available for progeny testing stages.

224 With single progeny test (S1) we used the 14,000 progeny to test 14 donor  
225 candidates each with 1,000 progeny, 28 donor candidates each with 500 progeny,  
226 56 donor candidates each with 250 progeny, 112 donor candidates each with  
227 125 progeny, 224 donor candidates each with 63 progeny, or 448 donor candidates  
228 each with 31 progeny.

229 With two progeny tests (S1 and S2) we used either 2,000, 4,000, or 6,000  
230 progeny for the first test (S1) and the remaining 12,000, 10,000, or 8,000 for the  
231 second test (S2). At S1 either 100, 200, or 400 donor candidates were tested. Thus,

232 when 2,000 progeny were used at S1 the 100, 200, or 400 donor candidates were each  
233 tested with 20, 10, or 5 progeny respectively. When 4,000 progeny were used at S1  
234 the 100, 200, or 400 donor candidates were each tested with 40, 20, or 10 progeny  
235 respectively. When 6,000 progeny were used at S1 the 100, 200, or 400 donor  
236 candidates were each tested with 60, 30, or 15 progeny respectively. At S2 we tested  
237 either 10 or 20 donor candidates advanced from S1. When 12,000 progeny remained  
238 to be used at S2 the 10 or 20 donor candidates were each tested with 1,200 or 600  
239 progeny respectively. When 10,000 progeny remained to be used at S2 the 10 or 20  
240 donor candidates were each tested with 1,000 or 500 progeny respectively. When  
241 8,000 progeny remained to be used at S2 the 10 or 20 donor candidates were each  
242 tested with 800 or 400 progeny respectively. From each of these testing strategies we  
243 chose either 1 or 5 donors of spermatogonial stem cells for surrogate sires in the  
244 commercial layer.

245 All of these different factors (two sizes of a breeding program [Small, Big],  
246 three conventional strategy scenarios [50, 200, 500 males], six surrogate sires strategy  
247 scenarios with two-stage testing, 18 surrogate sires strategy scenarios with three-stage  
248 testing, and using one or five donors) gave 102 different scenarios for each level of  
249 genomic test accuracy. The map of all these scenarios and used resources is  
250 summarized in Fig. 4.

## 251 **Time assumptions**

252 The time taken to transfer germplasm from the nucleus to the commercial  
253 layer was assumed to be 3.5 years for the conventional strategy (but see the note  
254 below about “dilution”), 3.5 years for the surrogate sires strategy with two-stage  
255 testing, and 4.5 years for the surrogate sires strategy with three-stage testing. The

256 different steps that underlie these time frames are presented in Fig. 3. We based our  
257 parameters on pigs and assumed 6 months for a male to reach sexual maturity, 4  
258 months for a successful gestation, and 8 months to collect terminal line phenotypes on  
259 progeny. Based on these parameters we assumed 12 months to progeny test a sexually  
260 mature male. When the donors are identified we assumed that it takes a further 12  
261 months to produce surrogate sires from these. Finally, we assumed a 12 months for  
262 the commercial progeny to pass through gestation and complete their growth. We  
263 assumed that the conventional program involved two rounds of multiplication that  
264 each take 12 months to complete.

265         Although we assumed that the genetic improvement with the conventional  
266 strategy is delivered to the commercial population in 3.5 years, we assumed an  
267 additional component of genetic lag, because the genetic merit of the sires entering  
268 the multiplier layer is “diluted” by the lagged genetic merit of females in the  
269 multiplier layer (i.e., we assumed no selection of females in the multiplier). Such a  
270 dilution would not occur with the surrogate sires strategy, because the multiplication  
271 layer does not arise. To account for this extra genetic lag in the conventional strategy  
272 we “diluted” genetic merit of commercial sires as follows:

$$\bar{a}_t^d = 0.5\bar{a}_t + 0.5 \sum_{i=1}^6 \bar{a}_{t-i} w_i$$

273 where  $\bar{a}_t$  is the average genetic merit of used nucleus males in generation  $t$  and  $w_i$  is  
274 the relationship coefficient between the commercial sire and his maternal male  
275 ancestor in the generation  $i$ . We only accounted for 6 generations with  $w_i$  ranging  
276 from 0.5 in  $t - 1$  generation to 0.015625 in the  $t - 6$  generation. This “dilution”  
277 increased genetic lag of the conventional strategy by an equivalent of ~1.04 years’

278 worth of extra genetic gain.

## 279 **Comparison of different scenarios**

280 To ensure that sufficient numbers of generations had been traversed for  
281 “dilution”, we chose to present the results in terms of the genetic merit of terminal  
282 sires used in the commercial layer emerging from generation 11 and each subsequent  
283 generation. We report genetic merit in units of the standard deviation of true breeding  
284 values of the nucleus animals in the base generation ( $\sigma_b$ ), i.e., as  $(\bar{a}_t - \bar{a}_b)/\sigma_b$ , where  
285  $\bar{a}_t$  is the average true breeding value of the nucleus males that gave rise to  
286 commercial sires in year  $t$  and  $\bar{a}_b$  is the average true breeding value of nucleus  
287 animals in the base generation. Calculating the genetic merit of commercial sires in  
288 this way allowed the different strategies to be compared in terms of genetic merit of  
289 the commercial sires at the same year. Finally, we have converted the standardized  
290 genetic merit into years’ worth of genetic gain by calculating the number of years it  
291 takes the conventional breeding program when selecting the top 50 males to deliver  
292 the same level of genetic merit to the commercial layer.

## 293 **Results**

294 The surrogate sires strategy increased the genetic merit of terminal sires used in the  
295 commercial layer. The genetic merit of commercial surrogate sires from the surrogate  
296 sires strategy was as much as 6.5 to 9.2 years' worth of genetic gain higher than the  
297 genetic merit of commercial sires from the conventional multiplication strategy. In  
298 both the SmallScenario and BigScenario the three-stage testing strategy was the best  
299 strategy for identifying elite donors. The best performing three-stage testing strategy  
300 involved a genomic test at the first stage, 100 donor candidates tested each with 60  
301 progeny at the second stage, and 20 donor candidates tested each with 400 progeny at  
302 the third stage (see Table 1 for details). The benefit of surrogate sires strategy was  
303 greatest when the genomic test accuracy was lowest and when the conventional  
304 strategy required large proportions of the nucleus males to be used for multiplication.

305 In what follows the results are divided into three sub-sections for ease of  
306 presentation: (i) comparison of the conventional strategy and the best performing  
307 surrogate sires strategies; (ii) comparison of two-stage testing scenarios of the  
308 surrogate sires strategy; and (iii) comparison of three-stage testing scenarios of the  
309 surrogate sires strategy. To avoid clutter in the figures or tables we do not show  
310 standard errors across the 100 replicates of the simulated scenarios because the  
311 standard errors were small in all instances less than 0.009 YGG.

### 312 **Comparison of the conventional and the best performing surrogate sires** 313 **strategies**

314 Fig. 5 and S1 show the average genetic merit of commercial sires derived from  
315 the best performing surrogate sires strategy scenario and the conventional strategy  
316 against time, for three different genomic test accuracies (0.5, 0.7, and 0.9) and the

317 SmallScenario and the BigScenario. The conventional strategy used the top 50, 200,  
318 or 500 males in multiplication. At all points in time and for all three genomic  
319 accuracies commercial sires derived from the best performing surrogate sires strategy  
320 scenario had a higher genetic merit than those derived from the conventional strategy.  
321 This benefit was greater when more males were used for multiplication in the  
322 conventional strategy. The benefit of using surrogate sires strategy decreased as the  
323 genomic test accuracy increased. Across time the difference between the two  
324 strategies was almost constant. These trends were common both in the SmallScenario  
325 and the BigScenario, although with differences in magnitude.

326 Table 1 enumerates some of the main results than can be observed in Fig. 4  
327 and S1. Across all scenarios tested the best performing surrogate sires strategy  
328 scenario involved first a genomic test of all donor candidates followed by two  
329 subsequent progeny tests and the use of a single elite donor. The benefit of surrogate  
330 sires strategy above the conventional strategy was greater when more males were used  
331 for multiplication with the conventional strategy. When the genomic test accuracy  
332 was low (0.5) the best strategy was to first progeny test 100 candidates on 6,000  
333 progeny and then to test 20 candidates on 8,000 progeny. This testing and subsequent  
334 production of surrogate sires was assumed to take one additional year compared to the  
335 conventional strategy. After accounting for this extra time and for the dilution in the  
336 conventional multiplication process, we observed that in the SmallScenario the  
337 surrogate sires strategy delivered on average between 6.5 and 9.2 years' worth of  
338 extra genetic gain in commercial sires compared to the conventional strategy that uses  
339 respectively between 50 and 500 males in multiplication. For the BigScenario the  
340 equivalent values were between 2.7 and 4.1 years' worth of extra genetic gain.

341           When the genomic test accuracy was higher ( $> 0.5$ ) the optimal allocation of  
342 testing resources was slightly different. Instead of first progeny testing 100  
343 candidates, as was the case when the genomic test accuracy was low, progeny testing  
344 200 candidates was the best performing scenario. All other scenario parameters were  
345 the same as when the genomic test accuracy was low. The benefit of surrogate sires  
346 strategy decreased with the increasing genomic test accuracy and the magnitude of  
347 benefit differed significantly between the SmallScenario and the BigScenario  
348 (Table 1).

349           On average the surrogate sires strategy in SmallScenario delivered between  
350 6.5 and 9.2 years' worth of extra genetic gain in commercial sires when the genomic  
351 test accuracy was 0.5. When the genomic test accuracy was 0.7 these values reduced  
352 to between 4.5 and 7.2 years and when the genomic test accuracy was 0.9 they further  
353 reduced to between 2.4 and 5.0 years.

354           On average the surrogate sires strategy in BigScenario delivered between 2.7  
355 and 4.1 years' worth of extra genetic gain in commercial sires when the genomic test  
356 accuracy was 0.5. When the genomic test accuracy was 0.7 these values reduced to  
357 between 2.1 and 3.5 years and when the genomic test accuracy was 0.9 they further  
358 reduced to between 1.20 and 2.50 years.

359           The differences in the SmallScenario and the BigScenario were due to the  
360 different proportions of males used in multiplication to give rise to commercial sires.  
361 In the SmallScenario 10% to 100% of males were used while the in the BigScenario  
362 2% to 20% of males were used.



363 For simplicity of presentation and based on the consistency of trends described  
364 above, in the following sections we only present comparisons to the conventional  
365 strategy in which 50 males were used in multiplication.

### 366 **Comparison of two-stage testing scenarios of the surrogate sires strategy**

367 Tables 2 and 3 show the performance of different two-stage testing scenarios  
368 of the surrogate sires strategy. Performance is measured as the average years' worth  
369 of extra genetic gain in the commercial sires delivered by the surrogate sires strategy  
370 compared to the conventional strategy for both the SmallScenario (Table 2) and the  
371 BigScenario (Table 3). Consistent with the results reported in the previous sub-section  
372 the benefit of surrogate sires strategy was always lower when the genomic test  
373 accuracy was higher. In some scenarios, the benefit was minimal. In all cases, there  
374 was an intermediate optimum for the numbers of candidates tested. Using five elite  
375 donors was always worse than using one. This behaviour was observed in both the  
376 SmallScenario and the BigScenario although with some interesting differences. The  
377 BigScenario showed a general shrinkage of years' worth of genetic gain compared to  
378 the SmallScenario, resulting in a general increase in the number of scenarios that  
379 showed a small benefit of the surrogate sires strategy.

380 At all levels of genomic test accuracy the best scenario was to screen  
381 candidates based on genomic test, progeny test 112 candidates each with 125 progeny,  
382 and use the best candidate as a single elite donor. With the genomic test accuracy of  
383 0.5, 0.7, and 0.9 this scenario gave respectively 5.3, 3.6, or 2.2 years' worth of extra  
384 genetic gain in commercial sires in the SmallScenario (Table 2) and respectively 2.5,  
385 2.0 or 1.1 years' worth of extra genetic gain in commercial sires in the BigScenario  
386 (Table 3).

387           Just as for the case of selecting one elite donor of spermatogonial cells for  
388 surrogate sires, when selecting five elite donors, progeny testing 112 candidates each  
389 with 125 progeny gave the highest benefit. With the genomic test accuracy of 0.5, 0.7,  
390 and 0.9 this scenario gave respectively 3.6, 2.6 and 1.2 years' worth of extra genetic  
391 gain in the SmallScenario (Table 2) and respectively 2.0, 1.1 and 1.0 in the  
392 BigScenario (Table 2).

### 393 **Comparison of three-stage testing scenarios of the surrogate sires strategy**

394           Tables 4 and 5 shows the performance of different three-stage testing  
395 scenarios of the surrogate sires strategy in the SmallScenario when either one or five  
396 elite donors used. By varying several parameters, we tested 216 (108 for the  
397 SmallScenario and 108 for the BigScenario) different scenarios of three-stage testing  
398 with fixed total progeny testing resources. These resources were the same as for the  
399 two-stage testing scenarios described in the previous sub-section. The parameters with  
400 the three-stage testing scenarios were the genomic test accuracy for the first stage, the  
401 split of resources between the two subsequent progeny tests, the number of tested  
402 donor candidates, the number of progeny per tested donor candidate at each progeny  
403 test stage, and the number of elite donors used for production of surrogate sires.

404           The three-stage testing gave a greater benefit than the two-stage testing. As for  
405 the two-stage testing, using one elite donor for surrogate sires gave a greater benefit  
406 than using five elite donors and the benefit of surrogate sires strategy was greater  
407 when the genomic test accuracy was lower. A total of 14,000 progeny were split  
408 across the two stages of progeny testing. Increasing the resources in the first progeny  
409 test increased benefit of surrogate sires strategy. For example, with the SmallScenario  
410 when the genomic test accuracy was 0.5, 6,000 progeny were used in the first progeny

411 test, 8,000 were used in the second progeny test, and when one elite donor was used in  
412 the end, the benefit was 6.5 years' worth of extra genetic gain in commercial sires  
413 above the conventional strategy that uses 50 nucleus males in multiplication. This was  
414 a greater benefit than the 5.8 years' worth of extra genetic gain for the scenario that  
415 split the 14,000 progeny into 4,000 for the first progeny test and 10,000 for the second  
416 progeny test, which was in turn better than the 5.4 years' worth of extra genetic gain  
417 for the scenario that split the 14,000 progeny into 2,000 for the first progeny test and  
418 12,000 for the second progeny test. This trend of greater benefit when more progeny  
419 were dedicated to the first progeny test was observed for almost all tested scenarios.

420 For the SmallScenario the difference between testing 100 or 200 donor  
421 candidates at the first progeny test was not consistent. That said, when the genomic  
422 test accuracy was 0.5, allocating 100 candidates to the first progeny test was usually  
423 better than allocating 200, and allocating 200 candidates was usually better than  
424 allocating 400. At higher genomic test accuracies, there were little differences  
425 between allocating 100 or 200 candidates to the first progeny test, but both of these  
426 sets of scenarios were usually better than allocating 400 candidates to the first  
427 progeny test.

428 In the SmallScenario allocating 20 elite donor candidates to the second  
429 progeny test was almost always better than allocating 10 candidates. A total of 54  
430 scenarios were evaluated for SmallScenario. In only 6 of these scenarios allocating 10  
431 candidates was better than allocating 20.

432 Overall for the SmallScenario, when the genomic test accuracy was 0.5, the  
433 best three-stage testing scenario used 6,000 progenies in the first progeny test of 100  
434 candidates each with 60 progeny, 8,000 progenies in the second progeny test of 20

435 candidates each with 400 progeny, and used a single elite donor for surrogate sires.  
436 This scenario gave a benefit of 6.50 years' worth of extra genetic gain in commercial  
437 sires compared to the conventional strategy. The same distribution of testing resources  
438 was also the joint best when five, instead of one, elite donors were used for surrogate  
439 sires.

440 The same trends as for the SmallScenario were observed also for the  
441 BigScenario, but with smaller benefit of the surrogate sire strategy (See table S1 and  
442 S2).

443

## 444 **Discussion**

445           The results of this paper suggest that a surrogate sires strategy could be very  
446 beneficial for the dissemination of genetic gain in animal breeding. In summary, our  
447 results indicate that benefits of the as much as 6.5 to 9.2 years' worth of genetic gain  
448 in commercial sires could be realized with surrogate sires compared to the  
449 conventional multiplication. It was best to identify elite donors for surrogate sires via  
450 a three-stage testing strategy involving a first screen with a genomic test followed by  
451 two subsequent progeny tests. The benefits of a surrogate sires strategy were greater  
452 when genomic test accuracy was low and when the conventional strategy used a large  
453 proportion of males in multiplication. To discuss these results we divide the  
454 discussion into four sections: (i) possible explanations for the observed trends; (ii)  
455 justification and impact of assumptions; (iii) the potential impact of surrogate sires on  
456 the redesign of animal breeding programs; and (iv) risks and opportunities of using  
457 surrogate sires.

### 458 **Possible explanations for the observed trends**

459           That surrogate sire technology generates such a benefit in terms of years'  
460 worth of genetic gain can be explained in the context of the breeders' equation. While  
461 the surrogate sires strategy does not rely on the selection of the best individuals and  
462 using them as parents of the next generation, it does rely on the identification of the  
463 best individuals from a cohort and using them as donors of spermatogonial cells for  
464 surrogate sires, which is another form of the selection problem. In any cohort, the best  
465 few individuals will be some number of standard deviations above the cohort average.  
466 For example, when surrogate sires technology delivered 6.5 years' worth of additional  
467 genetic gain in commercial sires the best nucleus male was on average 2.7 standard

468 deviations above the cohort mean. In contrast, the best 50 nucleus males were 2.0  
469 standard deviations above the cohort mean. Given that the breeding program  
470 proceeded at a rate of genetic progress of 0.4 standard deviations per year, choosing  
471 the best male as a donor for surrogate sires rather than the best 50 males produced  
472 surrogate sires that were better for more than 8 years' worth of genetic gain. However,  
473 accounting for the imperfect accuracy of identifying donors with the surrogate  
474 strategy or the best 50 males for multiplication with the conventional strategy and the  
475 time to generate commercial sires with either strategy the final result was 6.5 years'  
476 worth of genetic gain.

477         With constant progeny test accuracies the benefit of the surrogate sires  
478 strategy depended on the proportion of male candidates that the conventional strategy  
479 used to give rise to commercial sires. If the breeding program needed to use a large  
480 proportion of its nucleus male candidates (e.g., the best 200 or 500) the benefit of  
481 surrogate sires strategy was greater than if it needed to use a few. Again, this result is  
482 entirely consistent with the breeders' equation. Specifically, it can be explained in the  
483 context of selection intensity being a nonlinear function of the percentage of selected  
484 individuals, i.e., selection intensity increases almost linearly down to 20 or 10%  
485 selected, but increases sharply (nonlinearly) thereafter. While both conventional and  
486 surrogate sires strategies exploit the tail of distribution with high selection intensities,  
487 the surrogate sires strategy also exploits the steeper part. This explains why the  
488 benefit of surrogate sires was higher in the SmallScenario than in the BigScenario. In  
489 the SmallScenario we had 500 candidates and selected 100 with the conventional  
490 strategy (percentage selected 20% and selection intensity 1.4) or 1 with the surrogate  
491 sires strategy (percentage selected 0.2% and selection intensity 3.2). In the  
492 BigScenario we had 2,500 candidates and selected 100 with the conventional strategy

493 (percentage selected 4% and selection intensity 2.2) or 1 with the surrogate sires  
494 strategy (percentage selected 0.04% and selection intensity 3.6). The same logic also  
495 explains why selecting five as opposed to one donor for surrogate sires gave a lower  
496 benefit.

497         The observed differences in the performance of different surrogate sire  
498 strategies can also be explained in the context of the breeders' equation. When the  
499 genomic test accuracy used in the first stage of testing was lower the benefit of  
500 surrogate sires strategy was higher. Under the conventional strategy, the average  
501 genetic merit of the nucleus males that gave rise to commercial sires was lower when  
502 the genomic test accuracy was lower than when it was higher. With surrogate sires  
503 strategy this reduction in genetic merit due to the low genomic test accuracy is  
504 compensated by the subsequent progeny tests. This is in line with the analysis of  
505 Dickerson and Hazel [18], who compared the use of progeny test as a supplement to  
506 earlier culling on own or sibling performance. Their conclusion was that progeny  
507 testing is warranted when heritability is low in which case accuracy of estimated  
508 breeding values from own or sibling phenotypes (or genomic prediction in our study)  
509 is low. Genomic selection can be thought of as a light touch first screen, the purpose  
510 of which is to identify the top group of animals, which are then tested on many  
511 progeny. The purpose of subsequent progeny tests is then a search for the best  
512 individual within this group.

513         This same logic also explains why the three-stage testing was better than the  
514 two-stage testing. Both schemes started with a genomic test that was followed by one  
515 progeny test with the two-stage testing or two subsequent progeny tests with the three-  
516 stage testing. With the three-stage testing the first progeny test serves to use a portion

517 of resources to evaluate many candidates relatively accurately in order to discard most  
518 candidates. Then the second progeny test uses the remaining resources to even more  
519 accurately identify the final candidate. In terms of the breeders' equation the first  
520 progeny test maximizes selection intensity, while the second maximizes accuracy.  
521 The three-stage testing appears to address both of these parameters more optimally  
522 than the two-stage testing.

523         There is a substantial body of literature on multi-stage selection [19–23] which  
524 the observed trends in this study are consistent with. It is well known that increasing  
525 the number of progeny per candidate increases accuracy [24,25] and that the number  
526 of candidates to be tested is important and the trade-off between the two must be  
527 found. In our simulations, we found the optimum at progeny testing 112 candidates,  
528 given a fixed amount of resources, in our case 14,000 progeny. This optimum was  
529 consistent across the different levels of genomic test accuracy. However, the level of  
530 genomic test accuracy heavily influenced the amount of extra genetic gain, because  
531 higher accuracy directly translates to higher genetic gain. These trends are consistent  
532 with the long-established multi-stage testing in plant breeding [9]. Most plant  
533 breeding programs use multi-stage testing to identify elite single genotype (e.g.,  
534 inbred line) that is then deliver to the commercial layer. Typically, these programs  
535 initially screen many individuals imprecisely at the first stage. At each subsequent  
536 stage they reduce the number of tested individuals, but the testing precision is  
537 increased.

### 538 **Justification and impact of assumptions**

539         There is a huge range of possible strategies for the identification of donors for  
540 surrogate sires and we only evaluated a small subset in this study. We choose the



541 tested range of scenarios because we believe they could demonstrate the properties of  
542 surrogate sires strategy. They show that in some circumstances surrogate sires can  
543 deliver a large benefit and in others small benefit. We chose the three levels of  
544 genomic test accuracy as these levels reflect what might be possible in breeding  
545 programs of various sizes. To ensure that all strategies used an equal set of resources  
546 we set the total number of progeny involved in progeny testing to 14,000. We chose  
547 this number as it was divisible in many ways and thereby enabled several strategies to  
548 be compared and because this number was similar the 10,000 progeny that would be  
549 used by an animal breeding program that each year tested 100 candidates each with  
550 100 progeny, a scale of progeny testing that was not uncommon in some animal  
551 breeding programs before the advent of genomic selection.

552         With the two-stage testing the total testing resources were distributed across  
553 many or few candidates. As expected, testing an intermediate to high number of  
554 candidates (i.e., 112 to 224) on a relatively small number of progeny (i.e., 125 to 63)  
555 gave higher benefits than testing a few candidates (e.g., 14) on many progeny or a  
556 very high number of candidates (448) on few progeny (31). These trends fit the  
557 expectations from the breeders' equation and occur due to the interplay between  
558 selection intensity and accuracy. However, when the chosen elite donors of  
559 spermatogonial cells for surrogate sires are to be used to produce huge numbers of  
560 progeny in the commercial layer, the risk of a donor carrying some major defect that  
561 was not identified by the testing process must also be minimized. For this reason, it is  
562 unlikely that a strategy in which donors are tested with a single stage of progeny  
563 testing using a ~200 or less progeny would ever be used by a commercial breeding  
564 program.

565           It was this logic that motivated us in our design of the three-stage testing  
566 scheme. Our intuition was that the first progeny test would evaluate many candidates  
567 with relatively low accuracy, while the second progeny test would evaluate a handful  
568 of individuals with high accuracy, i.e., 10 or 20 candidates each with respectively 800  
569 or 400 progeny. Using many progeny ensures high accuracy, but also a high degree of  
570 certainty that the final donor(s) would not carry any major defects.

571           A major assumption of this study was the amount of time it took to identify  
572 elite donors and then to make surrogate sires. It is likely that the different time  
573 assumptions could be shortened or lengthened for both the conventional  
574 multiplication strategy and the surrogate sires strategy in several ways and depending  
575 on the assumed species. The benefit of surrogate sires strategy would change  
576 accordingly.

577           Finally, we choose to model a pig breeding program in this study because this  
578 is the species that we are most familiar with. The benefits may be greater or smaller  
579 for other species. The benefits depend on the ratio of existing reproductive rates of  
580 males versus that enabled by surrogate sire technology, the time and cost associated  
581 with performing progeny tests, the levels of accuracy that can be obtained by genomic  
582 prediction and the relative cost and technical possibilities of surrogate sire technology  
583 itself in a particular species. Incidentally, in this study we did not account for the cost  
584 aspects of surrogate sire technology itself. Undoubtedly developing the technology  
585 itself will be hugely expensive and these costs of development may impact its  
586 eventual commercial cost. That said, in time many biotechnologies which are initially  
587 expensive become much cheaper (e.g., nowadays genotyping and animal cloning are  
588 both relatively inexpensive compared to their former costs) and we anticipate that

589 surrogate sire technology will follow a similar pattern. However, given we have  
590 ignored the cost component of surrogate sire technology its benefit may be  
591 overestimated based on our results compared to a study which would account for such  
592 costs.

### 593 **The potential impact of surrogate sires on the redesign of animal breeding** 594 **programs**

595 Animal breeding programs maximize the genetic merit of commercial animals  
596 within the available financial, physical, technical, and physiological constraints.  
597 Implicitly a breeding program has two objectives: (i) improving the mean of the  
598 population; and (ii) delivering a product to the commercial producers. In dairy cattle  
599 for example, before the advent of genomic selection, breeders used progeny testing  
600 schemes that intensively evaluated relatively small numbers of candidate males and  
601 used the best of these as parents to improve the population, but also as a commercial  
602 and breeding product to be used by the commercial layer. In doing so, dairy cattle  
603 breeders maximised selection accuracy, but were constrained in their ability to  
604 increase selection intensity and decrease generation interval. However, commercial  
605 producers used well tested sires and therefore an individual producer could rely on  
606 using relatively few sires, who together could serve entire geographic regions. The  
607 advent of genomic selection changed this paradigm. Under genomic selection progeny  
608 testing of a small number of candidates has been replaced with a genomic testing of a  
609 large number of candidates. Those with best predictions are used as parents to  
610 improve the “open” nucleus population, but are also sold to commercial layer as a  
611 team of sires product (i.e., a group of sires sold together rather than a single sire sold  
612 on its own). In doing so, dairy cattle breeders increased selection intensity and

613 reduced generation interval, but are constrained in their ability to achieve very high  
614 accuracy. Given that each candidate male has not had their merit assessed based on  
615 phenotypes of their progeny, there is a risk that certain sires are not that good or may  
616 carry mutations that are highly detrimental (e.g., a *de-novo* mutation that prevents  
617 progeny from lactating) [26–28]. To overcome this risk, breeders recommend that  
618 commercial producers use semen of a larger number (i.e., a team) of sires and limit  
619 their use of any one sire.

620 A surrogate sires strategy would need to exploit aspects of both genomic and  
621 progeny testing. Genomic testing can be used to drive the population improvement  
622 and, as demonstrated in the present paper, to identify a set of candidates that could  
623 enter a progeny testing scheme as part of the product development. The role of the  
624 progeny testing is to ensure that the chosen elite donors that give rise to surrogate  
625 sires released to the commercial sector are good animals, that they are not  
626 significantly worse than it is predicted by a genomic test and that they do not carry  
627 detrimental mutations. As demonstrated by the results of the present study two  
628 subsequent progeny tests used resources more efficiently than a single progeny test.  
629 Such multi-stage testing has a long history of use in plant breeding which also has a  
630 long history delivering products to commercial producers in a way that is highly  
631 analogous to what surrogate sires would enable for animal producers.

632 The majority of commercial producers for all of the major crops (maize,  
633 wheat, rice) use inbred lines or their hybrids. These inbred or hybrid lines can be  
634 grown on huge areas. Plant and animal breeding designs have diverged somewhat  
635 over the years owing to differences in biology, economics, and technical possibilities.  
636 Surrogate sire technology, combined with genomic selection, could result in a

637 coalescence of designs across plant and animal breeding. One such design that could  
638 apply to both is the two-part scheme recently proposed by Gaynor et al. (2017) [11].  
639 In this scheme, rapid recurrent selection based on genomic testing is used to increase  
640 the mean of the population, while multi-stage testing (genomic and phenotypic) is  
641 used to periodically extract, test, and develop a product from the population. The  
642 population improvement component resembles the nucleus of animal breeding  
643 programs, while the product development component resembles the multi-stage  
644 testing to derive inbred or hybrid lines of plant breeding programs. The latter could  
645 also be seen as an improved multiplication layer of animal breeding programs that  
646 exploit breed complementarity to deliver a commercial product.

647         In the present work, we focused on the use of surrogate sires to produce  
648 commercial animals (e.g., a terminal sire in a pig population). To do this, donors for  
649 surrogate sires were chosen based on their general combining ability. The strategy  
650 could also be extended to exploit specific combining ability to produce a relatively  
651 homogenous set of females from a maternal line that are crossed with single terminal  
652 male (via surrogate sires). Use of specific combining ability is widespread in hybrid  
653 crops where it exploits complementarity of pairs of individuals and heterosis  
654 generated by specific pairs of individuals. The surrogate sires strategy proposed in the  
655 present paper could be extended to exploit specific combining ability by adding  
656 additional stages that progeny tests specific crosses as is conducted in hybrid plant  
657 breeding programs. Because in livestock the parents are outbred (compared to crops  
658 where they are often inbred), a tiered strategy may be needed in the maternal line(s)  
659 that homogenizes dam haplotypes. For example, using a single surrogate sire,  
660 grandsire, and great-grandsire on the maternal population would give a pool of  
661 females that carried one of two haplotypes for 87.5% ( $0.5 + 0.25 + 0.125$ ) of their

662 genome. The terminal surrogate sire would be chosen based on a specific combining  
663 ability to these haplotypes.

#### 664 **Risks and opportunities of using surrogate sires**

665 Surrogate sires present risks and opportunities to commercial production. The  
666 most obvious risk relates to the genetic homogeneity of commercial animals if a  
667 single surrogate sire, or a set of very closely related surrogate sires were used. If a  
668 disease emerged that this homogenous group of animals was susceptible to, it could  
669 have a major detrimental impact on the commercial production. Having such large  
670 groups of homogeneous animals would also increase the selection pressure on disease  
671 pathogens to evolve pathogenicity to the group. Plant breeders and commercial crop  
672 growers have extensive experience in managing the potential to have genetic  
673 homogeneity across large segments of the production area. They have developed  
674 strategies to minimize the risk of disease outbreaks and other failures such as crop  
675 rotation, using multiple varieties on a farm, creating varietal blends consisting of  
676 multiple genotypes, and taking holistic strategies to pathogen management [29]. Aside  
677 from rotation, which is practically impossible in the animal sector, these strategies  
678 might have important roles in ensuring the effectiveness of surrogate sires in  
679 livestock.

680 A further risk of the homogenisation of the commercial population relates to  
681 genetic diversity. The genetic diversity contained in current populations is potentially  
682 a useful reserve of genetic diversity that could be used in breeding programs in case  
683 the nucleus genetic diversity was to become inappropriate at some point in the future  
684 (e.g., due to a disease catastrophe or because it became exhausted). Homogenisation  
685 of the commercial population would remove this safety net requiring greater care to

686 be taken in the preservation of genetic diversity. Genebanks using frozen semen, eggs,  
687 or embryos are well established ways to preserve genetic diversity. There are also new  
688 ways which include the use of cultured primordial germ cells [30].

689 Undetected but highly deleterious mutations also pose a risk for the use of  
690 surrogate sires. While it is unlikely that this would arise after sufficient testing, it is  
691 not impossible. One such route could be through the occurrence of one or more such  
692 mutations arising as somatic mutations after the animal had been tested, leading to a  
693 mosaicism, which might affect sets of surrogate sires from the donor.

694 The most obvious opportunity emanating from surrogate sires also relates to  
695 the genetic homogeneity of commercial animals and can also draw on practices that  
696 are well established in crop production. In crops, management plans are supplied to a  
697 farmer alongside the seed (e.g., <https://catalog.extension.oregonstate.edu/em9004>).  
698 These plans are specifically tailored to the variety genotype based on extensive sets of  
699 field trials. They include recommendations for target market, expected performance,  
700 optimum sowing date, seeding rate, soil type and water, fertilizer, pesticide and  
701 fungicide requirements. These management plans complement the genetics of the  
702 variety and increase the benefit obtained from the genetic potential in a generic  
703 environment. Similar management plans could be developed for surrogate sires and  
704 the benefits would be similarly expected to exceed the benefit that was observed in  
705 the present study for the genetics alone (e.g., 6.5 year's worth of genetic gain). The  
706 phenotype data collected to development of the management plans would also serve  
707 to further test and validate a particular donor.

708 Another obvious opportunity emanating from surrogate sires that also relates  
709 to the genetic homogeneity is the potential for increasing the product homogeneity. In

710 animal production, product uniformity is an important topic. In meat animals, for  
711 instance, uniformity has economic benefits because excessive variability in carcass  
712 weight or conformation is penalized by slaughterhouses [31,32]. A genetically  
713 homogeneous commercial population, achieved through the use of surrogate sires,  
714 could aid product uniformity. However, if this was to be achieved, most of the  
715 increase in uniformity would need to emanate from matching very specific  
716 management plans to the homogenous genetics because homogeneous genetics in  
717 itself has limited ability to increase phenotypic homogeneity. Van Vleck [33] showed  
718 that in the context of cloned animals, if heritability is 25%, then the phenotypic  
719 standard deviation among clones would be 87% of that of uncloned animals and only  
720 if heritability is 100%, will clone mates have complete uniformity.

721         Compared to the conventional multiplication the surrogate sires strategy  
722 enables shorter lag between nucleus and commercial layer and requires a smaller  
723 number of parents contributing to the commercial layer. This offers several  
724 advantages including the ability to rapidly change the entire genetics in the  
725 commercial layer. This could be used to rapidly respond to sudden changes in  
726 requirements such as pressure from a new disease or the emergence of a new market  
727 for the product that has specific requirements (e.g., meat marbling).

728         The surrogate sire strategy would be costly to implement in practice because it  
729 would require capacity in advanced molecular biology and infrastructure for progeny  
730 testing. However, it presents other opportunities through which costs can be saved.  
731 For example, multiplier populations to produce terminal sires would not need to be  
732 maintained. This would free up resources for other investment in breeding programs,  
733 such as more progeny testing of donor candidates.



734           The surrogate sires strategy presents breeding programs with an enhanced  
735 opportunity to protect its intellectual property via limited release of males (thereby  
736 limiting the access of competitors to the broader source germplasm) and by  
737 exploitation of specific combining ability. This protection would give the breeding  
738 companies incentive to invest more and help to avoid the commonly observed market  
739 failure in some breeding industries. When intellectual property is properly protected,  
740 breeding companies are anecdotally reported to share the benefits two-thirds to the  
741 farmers and one third to the breeding company. Such sharing more than offsets the  
742 purchase cost to a producer, while it also gives profit to the breeder. Perhaps the most  
743 spectacular example of the benefits of such ways to reward investment in intellectual  
744 property are seen in maize which has seen a 6-fold increase in productivity since  
745 hybrid breeding was introduced in the 1930's [34]. By releasing hybrids breeding  
746 organisations can protect the intellectual property that is their source germplasm. This  
747 in turn enables them to invest heavily in breeding activities (e.g., technology, field  
748 testing networks) that in turn drive accelerated genetic gains.

749           At least two barriers exist that may prevent the deployment of this technology  
750 in in real livestock breeding program. Firstly, genome editing currently appears to be  
751 the technology that is most likely to enable genome editing to be implemented in  
752 practice [2]. Globally, the future of governmental regulation of genome editing  
753 technology is currently uncertain which places uncertainty on the possibility for  
754 practical implementation of surrogate sire technology in real livestock breeding  
755 program. Secondly, effective deployment of surrogate sire technology will require  
756 partitioning of animal breeding programs into population improvement and product  
757 development parts. Product development will require deployment of extensive  
758 progeny testing schemes. Over the past decade the advent of genomic selection has

759 removed progeny testing schemes from many breeding programs. Reinstating such  
760 schemes would be costly and further work will be needed to demonstrate the exact  
761 return on investment.

762         Finally, the results of this study raise an important question for existing  
763 breeding programs that use artificial insemination for dissemination. As noted above,  
764 genomic selection has led to the removal of progeny testing schemes from many  
765 livestock breeding programs. Our results raise some doubts about the merit of this.  
766 They show that when a breeding program releases a small number of individuals that  
767 are deployed widely there is a benefit to progeny testing these individuals. The degree  
768 of benefit depends on the accuracy of genomic selection, the number of individuals  
769 released and their subsequent usage, and the accuracy and the number of stages in a  
770 progeny testing scheme and the relative time taken to perform a progeny test.  
771 Determining whether the removal of progeny testing schemes from genomic selection  
772 driven livestock breeding programs was the right thing to do in retrospect is beyond  
773 the scope of the present study but is an interesting question for future research.

774

## 775 **Conclusions**

776           The results of this study showed that using the surrogate sires strategy could  
777 significantly increase the genetic merit of commercial sires, by as much as 6.5 to 9.2  
778 years' worth of genetic gain, compared to the conventional multiplication strategy.  
779 The simulations suggest that identifying elite donors for surrogate sires should be  
780 based on three stages, the first of which uses a genomic test followed by two  
781 subsequent progeny tests. The use of one or a handful of elite donors to generate  
782 surrogate sires that in turn give rise to all production animals would be very different  
783 to current practice. While the results demonstrate the great potential of surrogate sires  
784 strategy there are considerable risks as well as opportunities. Practical implementation  
785 of surrogate sires strategy would need to account for these.

786

787 **Competing interests**

788 The authors declare that they have no competing interests.

789 **Authors' contributions**

790 JMH conceived the study. JMH, MB, and PG designed the study. PG performed the  
791 analysis. JMH and PG wrote the first draft of the manuscript. GG, MB, RCG, JJ,  
792 RRF, CBAW, AJM, and WOH helped to interpret the results and refine the  
793 manuscript. All authors read and approved the final manuscript.

794 **Acknowledgements**

795 The authors acknowledge the financial support from the BBSRC ISPG to The Roslin  
796 Institute BBS/E/D/30002275, from Grant Nos. BB/N015339/1, BB/L020467/1,  
797 BB/M009254/1, from Genus PLC and from Innovate UK. This work has made use of  
798 the resources provided by the Edinburgh Compute and Data Facility (ECDF)  
799 (<http://www.ecdf.ed.ac.uk>).

800

## 801 **References**

- 802 1. Park K-E, et al. (2017) Generation of germline ablated male pigs by CRISPR/Cas9  
803 editing of the NANOS2 gene. *7*:40176.
- 804 2. Oatley Jon M. (2017) Recent advances for spermatogonial stem cell transplantation  
805 in livestock. *Reproduction, fertility and Development 30*:44-49.  
806
- 807 3. Zhang Z., Shao S., and Meistrich M. L. (2006) Irradiated mouse testes efficiently  
808 support spermatogenesis derived from donor germ cells of mice and rats. *J.*  
809 *Androl.***27**: 365–375.  
810
- 811 4. Tsuda M., Sasaoka Y., Kiso M., Abe K., Haraguchi S., Kobayashi S., and Saga Y.  
812 (2003) Conserved role of nanos proteins in germ cell development. *Science* **301**:  
813 1239–1241.  
814
- 815 4. De Roos APW. (2011) Recent trends in genomic selection in dairy cattle. 62nd  
816 Annu. Meet. EAAP. Stavanger, Norway: Wageningen Academic Publishers: p.  
817 Contribution 01–7, p. 4.
- 818 5. Pryce JE, Daetwyler HD (2012) Designing dairy cattle breeding schemes under  
819 genomic selection: a review of international research. *Anim Prod Sci* 52(3):107–114.
- 820 6. Meuwissen T, Hayes B, Goddard M (2013) Accelerating Improvement of  
821 Livestock with Genomic Selection. *Annu Rev Anim Biosci* 1(1):221–237.
- 822 7. Van Eenennaam AL, Weigel KA, Young AE, Cleveland MA, Dekkers JCM (2014)  
823 Applied Animal Genomics: Results from the Field. *Annu Rev Anim Biosci* 2(1):105–  
824 139.
- 825 8. Knol EF, Nielsen B, Knap PW (2016) Genomic selection in commercial pig  
826 breeding. *Anim Front* :6-15.
- 827 9. Visscher P, Pong-Wong R, Whittemore C, Haley C (2000) Impact of biotechnology  
828 on (cross)breeding programmes in pigs. *Livestock Production Science* 65(1):57–70.
- 829 10. Bernardo R (2014) Essentials of Plant Breeding. Stemma Press (2014), pp. 252,  
830 £145.00 (pb). ISBN 13:9780972072427.
- 831 11. Gaynor RC, Gorjanc G, Bentley AR, Howell P, Jackson R, et al. (2017) A two-  
832 part strategy for using genomic selection to develop inbred lines. *Crop Science*  
833 *57*:2372-2386.
- 834 12. Chen GK, Marjoram P, Wall JD (2009) Fast and flexible simulation of DNA  
835 sequence data. *Genome Research* 19(1):136–142.
- 836 13. Faux A-M, et al. (2016) AlphaSim: Software for Breeding Program  
837 Simulation. *The Plant Genome* 9(3). doi:10.3835/plantgenome2016.02.0013.

- 838 14. Rodríguez-Ramilo ST, Fernández J, Toro MA, Hernández D, Villanueva B (2015)  
839 Genome-Wide Estimates of Coancestry, Inbreeding and Effective Population Size in  
840 the Spanish Holstein Population. *PLOS ONE*10(4):e0124157.
- 841 15. Hill WG (2009) Understanding and using quantitative genetic variation. *Philos*  
842 *Trans R Soc Lond B Biol Sci* 365(1537):73.
- 843 16. Brotherstone S, Goddard M (2005) Artificial selection and maintenance of genetic  
844 variance in the global dairy cow population. *Philos Trans R Soc Lond B Biol*  
845 *Sci* 360(1459):1479.
- 846 17. Hayes BJ, Visscher PM, McPartlan HC, Goddard ME (2003) Novel Multilocus  
847 Measure of Linkage Disequilibrium to Estimate Past Effective Population Size.  
848 *Genome Res* 13(4):635–43.
- 849 18. G.E. Dickerson, Hazel LN. Effectiveness of selection on progeny performance as  
850 a supplement to earlier culling in livestock. *Journal of Agricultural Research* 69:459.
- 851 19. Cunningham EP (1975) Multi-stage index selection. *Theoretical and Applied*  
852 *Genetics* 46:55–61.
- 853 20. Saxton AM (1983) A comparison of exact and sequential methods in multi-stage  
854 index selection. *Theoretical and Applied Genetics* 66(1):23–28.
- 855 21. Ducrocq V, Colleau J J (1989) Optimum truncation points for independent culling  
856 level selection on a multivariate normal distribution, with an application to dairy  
857 cattle selection. *Genet Sel Evol* 21(2):185–198.
- 858 22. Xie C, Xu S, Mosjidis JA (1997) Multistage selection indices for maximum  
859 genetic gain and economic efficiency in red clover. *Euphytica* 98(1):75–82.
- 860 23. Mi X, Utz HF, Technow F, Melchinger AE (2014) Optimizing Resource  
861 Allocation for Multistage Selection in Plant Breeding with R Package  
862 Selectiongain. *Crop Science* 54(4):1413–1418.
- 863 24. Robertson A (1957) Optimum Group Size in Progeny Testing and Family  
864 Selection. *Biometrics* 13(4):442–450.
- 865 25. Meuwissen THE, Goddard ME (1997) Optimization of progeny tests with prior  
866 information on young bulls. *Livestock Production Science* 52(1):57–68.
- 867 26. Milan D, Jeon J-T, Looft C, Amarger V, Robic A, Thelander M, et al. (2000) A  
868 Mutation in PRKAG3 Associated with Excess Glycogen Content in Pig Skeletal  
869 Muscle. *Science* 288(5469):1248-51.
- 870 27. VanRaden PM, Olson KM, Null DJ, Hutchison JL (2011) Harmful recessive  
871 effects on fertility detected by absence of homozygous haplotypes. *Journal of Dairy*  
872 *Science* 94(12):6153–61.
- 873 28. Kadri NK, Sahana G, Charlier C, Iso-Touru T, Guldbbrandtsen B, Karim L, et al.  
874 (2014) A 660-Kb Deletion with Antagonistic Effects on Fertility and Milk Production  
875 Segregates at High Frequency in Nordic Red Cattle: Additional Evidence for the

- 876 Common Occurrence of Balancing Selection in Livestock. *PLOS Genetics*  
877 10:e1004049.
- 878 29. McDonald BA, Linde C (2002) The population genetics of plant pathogens and  
879 breeding strategies for durable resistance. *Euphytica* 124:163–80.
- 880 30. Nandi S, Whyte J, Taylor L, Sherman A, Nair V, Kaiser P, et al. (2016)  
881 Cryopreservation of specialized chicken lines using cultured primordial germ cells.  
882 *Poultry Science* 95:1905–11.
- 883 31. Mulder HA, Bijma P, Hill WG (2007) Prediction of Breeding Values and  
884 Selection Responses With Genetic Heterogeneity of Environmental Variance.  
885 *Genetics* 175:1895.
- 886 32. Mulder HA, Bijma P, Hill WG (2008) Selection for uniformity in livestock by  
887 exploiting genetic heterogeneity of residual variance. *Genetics Selection*  
888 *Evolution* 40(1):37.
- 889 33. van Vleck LD (1999) Implications of cloning for breed improvement strategies:  
890 are traditional methods of animal improvement obsolete? *J Anim Sci* 77 Suppl 2:111–  
891 121.
- 892 34. Duvick DN (2005) Genetic Progress in yield of United States maize (*Zea mays*  
893 L.). *Maydica* 50:193–202.
- 894

895 **Figures**

896 **Fig. 1** Schematic depicting the possible application of spermatogonial stem cell  
897 transplantation methodology in pig production (depiction inspired by Oatley et al.,  
898 2018 [2])

899 **Fig. 2** Example animal (left) and plant (right) breeding schemes

900 **Fig. 3** Timeline of the different strategies to identify and disseminate genetic  
901 improvement

902 **Fig. 4** Map of the scenarios used in the study

903 **Fig.5** Average genetic merit of commercial sires derived from the best performing  
904 surrogate sire strategy scenario and the conventional strategy (top 50, 200 and 500  
905 males) for SmallScenario (a and b) and BigScenario (c and d) plotted against time

906

907 **Tables**

908 **Table 1** Average Years' worth of Genetic Gain (YGG) of the best performing  
909 surrogate sire strategy scenario above the conventional strategy that uses either 50,  
910 200, or 500 males

911 **Table 2** Average Years' worth of Genetic Gain (YGG) with the two-stage testing  
912 scenarios of the surrogate sire strategy above the conventional strategy that uses 50  
913 males (SmallScenario)

914



915 **Table 3** Average Years' worth of Genetic Gain (YGG) with the two-stage testing  
916 scenarios of the surrogate sire strategy above the conventional strategy that uses 50  
917 males (BigScenario)

918 **Table 4** Average Years' worth of Genetic Gain (YGG) with the three-stage testing  
919 scenarios of the surrogate sire strategy with one elite donor above the conventional  
920 strategy that uses 50 males (SmallScenario)

921 **Table 5** Average Years' worth of Genetic Gain (YGG) with the three-stage testing  
922 scenarios of the surrogate sire strategy with five elite donors above the conventional  
923 strategy that uses 50 males (SmallScenario)

924

925

926 **Supplementary material**

927 **Fig. S1** Average genetic merit of commercial sires derived from the best performing  
928 surrogate sire strategy scenario and the conventional strategy (top 50, 200 and 500  
929 males) for SmallScenario (a) and BigScenario(b) plotted against time

930 *Table S1* Average Years' worth of Genetic Gain (YGG) with the three-stage testing  
931 scenarios of the surrogate sire strategy with one elite donor above the conventional  
932 strategy that uses 50 males (BigScenario)

933 **Table S2** Average Years' worth of Genetic Gain (YGG) with the three-stage testing  
934 scenarios of the surrogate sire strategy with five elite donors above the conventional  
935 strategy that uses 50 males (BigScenario)

936

937

938

939

940

941

942

943

944

945

946

947

948

949

950

Fig. 1 Schematic depicting the possible application of spermatogonial stem cell transplantation methodology in pig production (depiction inspired by Oatley et al., 2018[2])

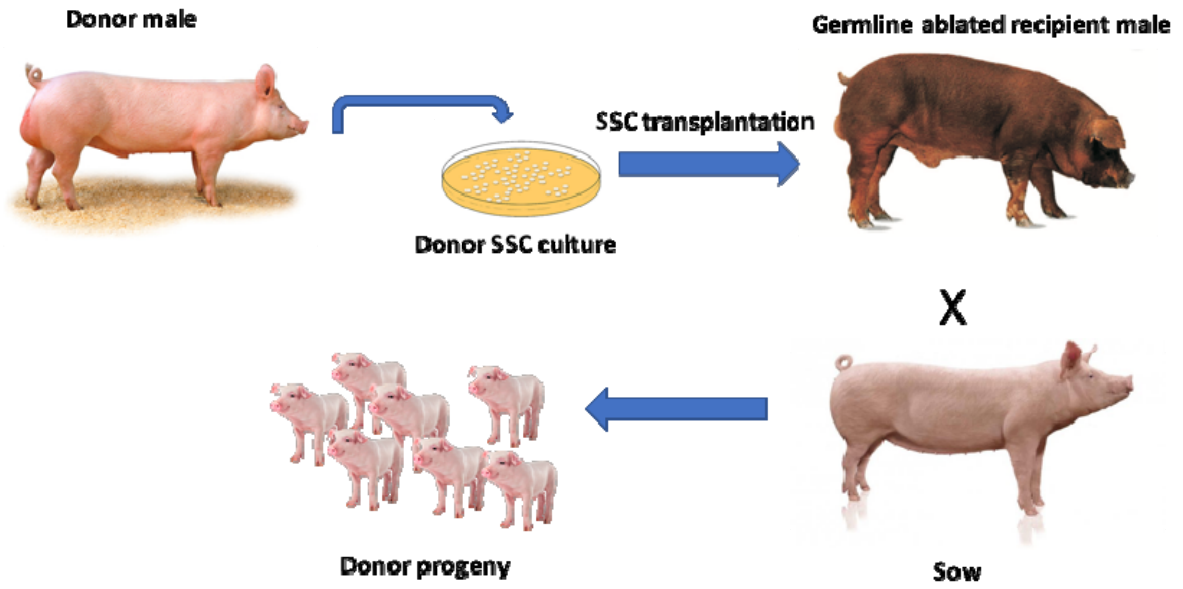


Fig. 2 Typical animal (left) and plant (right) breeding schemes

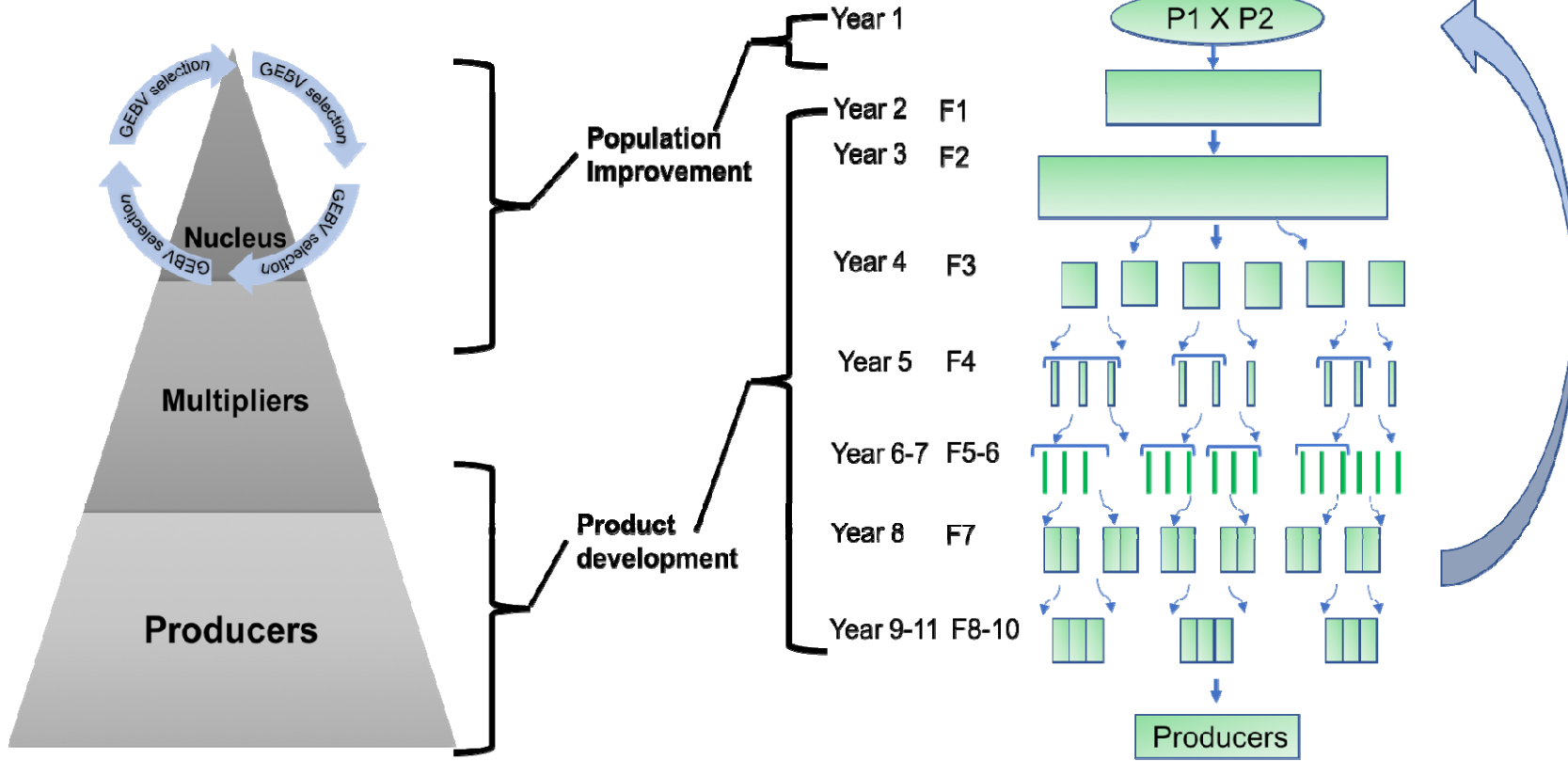
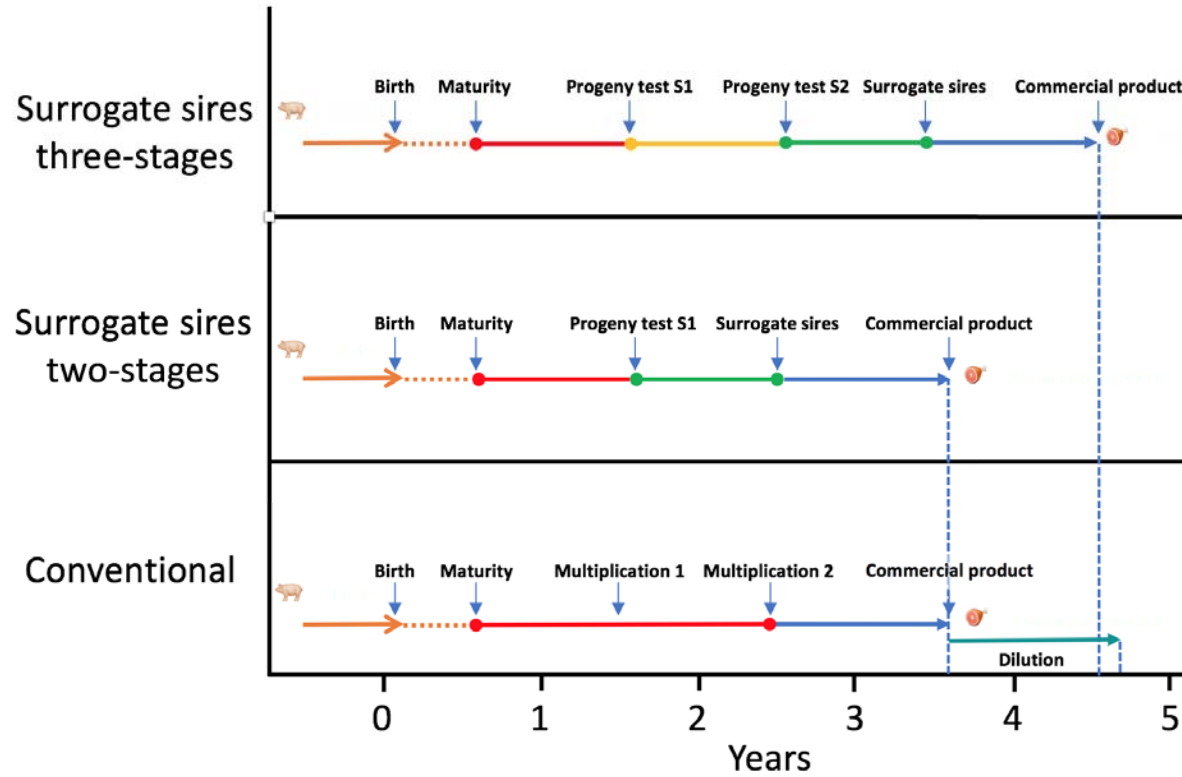


Fig. 3 Timeline of the different strategies to identify and disseminate genetic improvement



*Fig. 4 Map of the scenarios used in the study*

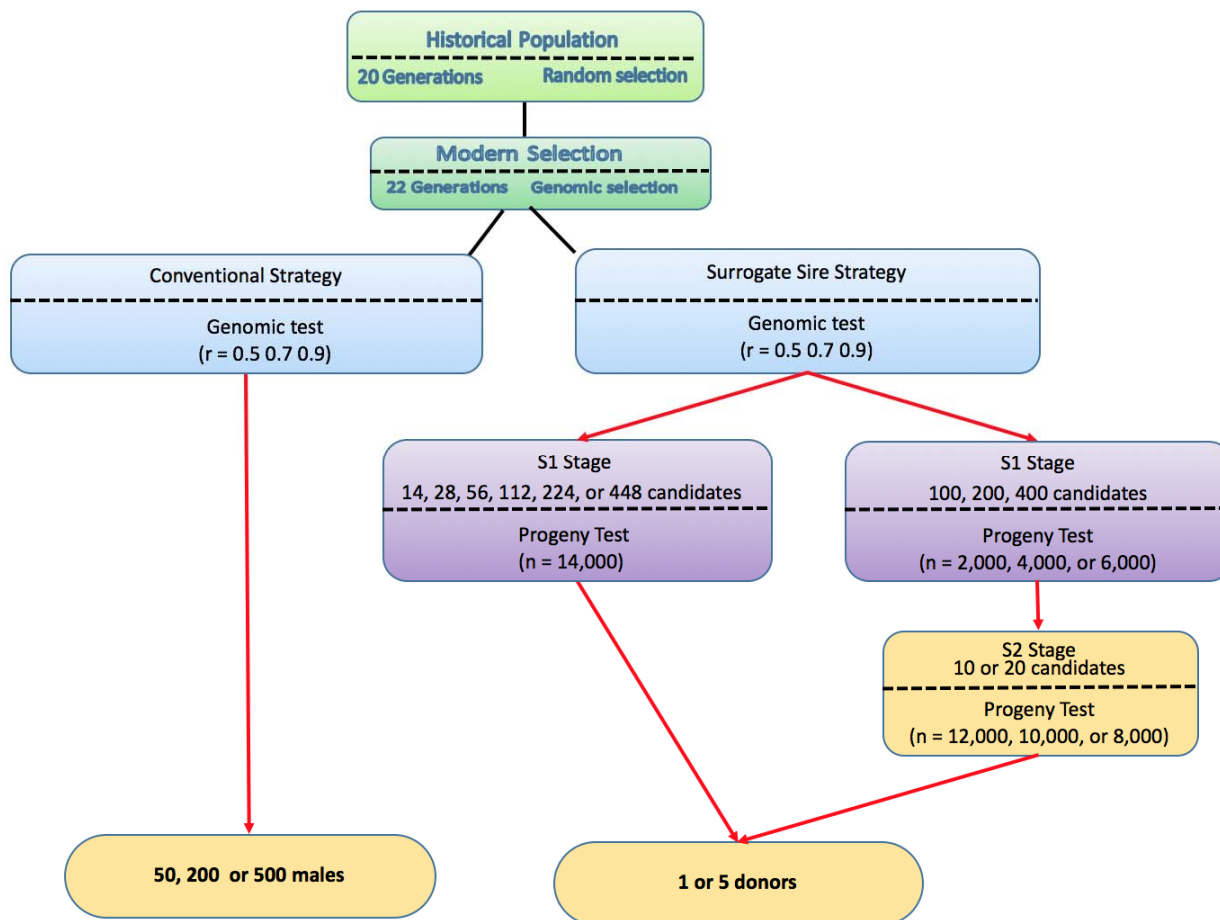
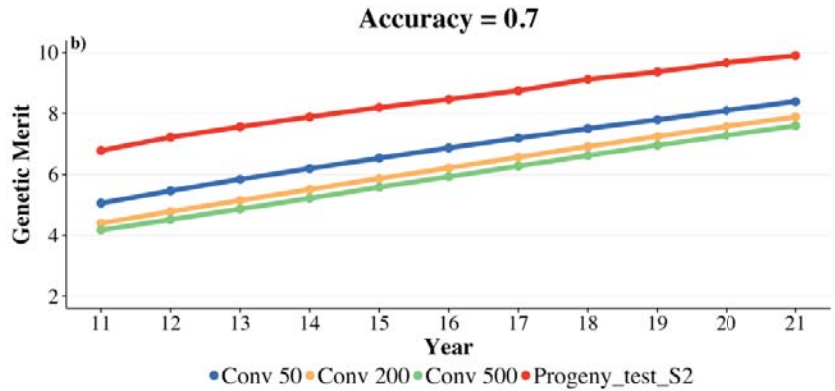
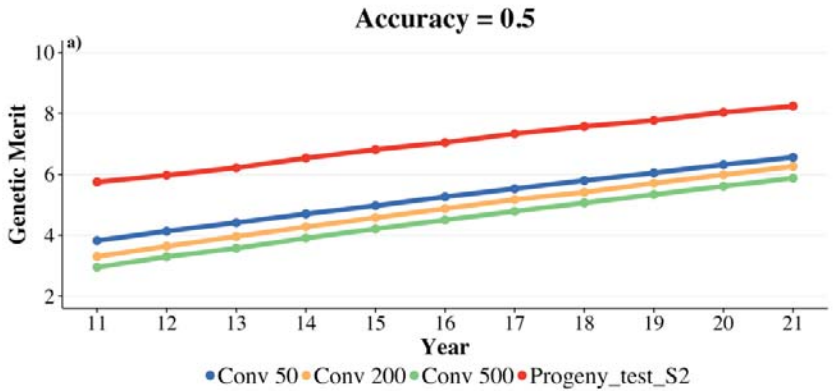




Fig. 5 Average genetic merit of commercial sires derived from the best performing surrogate sire strategy scenario and the conventional strategy (top 50, 200 and 500 males) for SmallScenario (a and b) and BigScenario (c and d) plotted against time



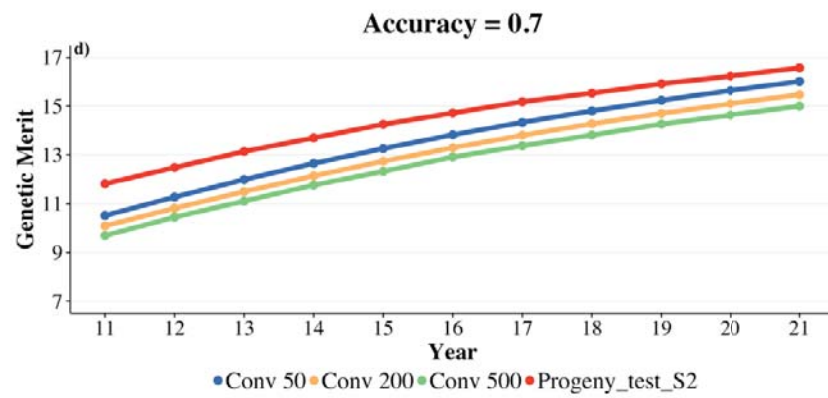
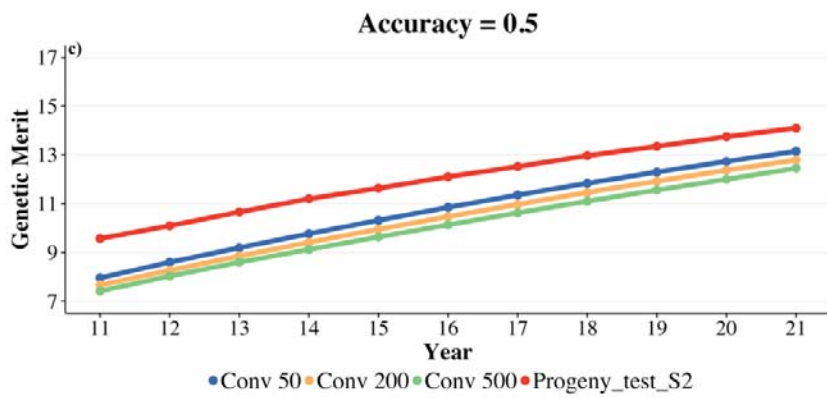
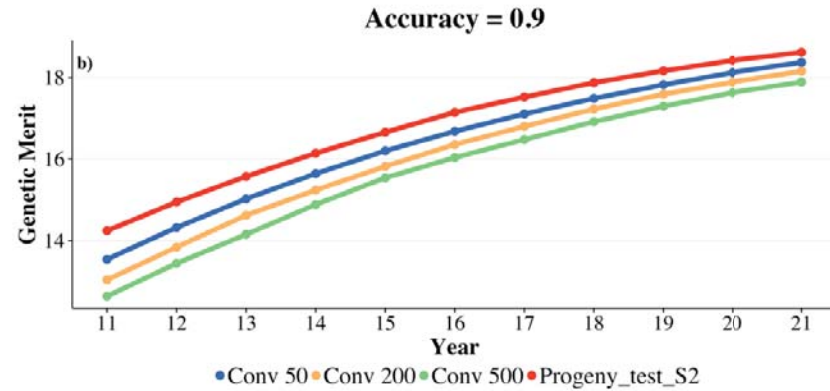
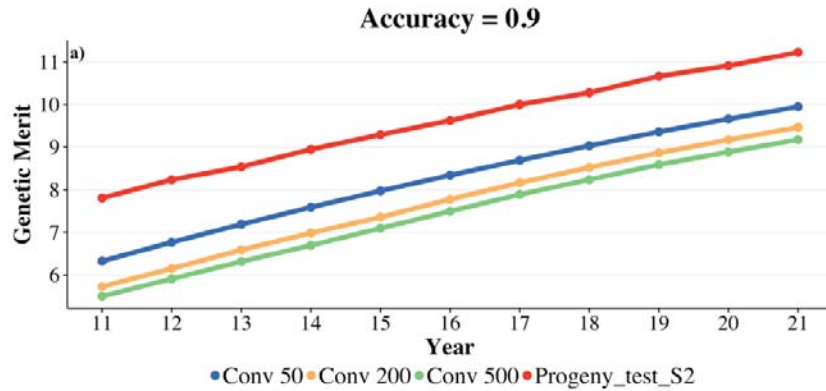


Fig. S1 Average genetic merit of commercial sires derived from the best performing surrogate sire strategy scenario and the conventional strategy (top 50, 200 and 500 males) for SmallScenario (a) and BigScenario(b) plotted against time



*Table 1 Average Years' worth of Genetic Gain (YGG) of the best performing surrogate sire strategy scenario above the conventional strategy that uses either 50, 200, or 500 males*

Genomic test accuracy	Males progeny tested S1	Males progeny tested S2	Progeny test resources <sup>1</sup>	Donors used	YGG <sub>50</sub>	YGG <sub>200</sub>	YGG <sub>500</sub>
Small Scenario							
0.5	100	20	6000S1 / 8000S2	1	6.5	7.5	9.2
0.7	200	20	6000S1 / 8000S2	1	4.5	6.5	7.2
0.9	200	20	6000S1 / 8000S2	1	2.4	4.5	5.0
Big Scenario							
0.5	100	20	6000S1 / 8000S2	1	2.7	3.5	4.1
0.7	200	20	6000S1 / 8000S2	1	2.1	2.5	3.5
0.9	200	20	6000S1 / 8000S2	1	1.2	1.7	2.5

<sup>1</sup>Total number of progeny allocated in the first progeny test (S1) and in the second progeny test (S2)

*Table 2 Average Years' worth of Genetic Gain (YGG) with the two-stage testing scenarios of the surrogate sire strategy above the conventional strategy that uses 50 males (SmallScenario)*

Males Tested	Progeny/Male	Donors used	YGG <sub>0.5</sub> <sup>1</sup>	YGG <sub>0.7</sub> <sup>1</sup>	YGG <sub>0.9</sub> <sup>1</sup>
14	1000	1	4.1	3.0	1.8
28	500	1	4.7	3.0	1.2
56	250	1	5.1	3.5	2.2
112	125	1	5.3	3.6	2.2
224	63	1	4.8	2.8	1.3
448	31	1	3.8	2.1	1.1
14	1000	5	2.9	1.9	0.2
28	500	5	3.1	2.1	0.5
56	250	5	3.6	2.4	1.1
112	125	5	3.6	2.6	1.2
224	63	5	3.4	1.9	0.3
448	31	5	2.8	1.6	0.2

<sup>1</sup>Genomic test accuracy at the initial stage (S0) 0.5, 0.7, and 0.9

*Table 3 Average Years' worth of Genetic Gain (YGG) with the two-stage testing scenarios of the surrogate sire strategy above the conventional strategy that uses 50 males (BigScenario)*

Males Tested	Progeny/Male	Donors used	YGG <sub>0.5</sub> <sup>1</sup>	YGG <sub>0.7</sub> <sup>1</sup>	YGG <sub>0.9</sub> <sup>1</sup>
14	1000	1	2.3	1.7	0.7
28	500	1	2.4	1.9	0.8
56	250	1	2.5	2.0	1.0
112	125	1	2.5	2.0	1.1
224	63	1	2.0	1.8	0.8
448	31	1	1.9	1.5	0.4
14	1000	5	1.7	1.2	0.5
28	500	5	1.7	1.2	0.7
56	250	5	1.9	1.1	1.0
112	125	5	2.0	1.1	1.0
224	63	5	1.8	1.0	0.5
448	31	5	1.0	0.8	0.3

<sup>1</sup>Genomic test accuracy at the initial stage (S0) 0.5, 0.7, and 0.9

*Table 4 Average Years' worth of Genetic Gain (YGG) with the three-stage testing scenarios of the surrogate sire strategy with one elite donor above the conventional strategy that uses 50 males (SmallScenario)*

Progeny test resources <sup>1</sup>	Males progeny tested S1	Progeny/Male S1	Males progeny tested S2	Progeny/Male S2	YGG <sub>0.5</sub> <sup>2</sup>	YGG <sub>0.7</sub> <sup>2</sup>	YGG <sub>0.9</sub> <sup>2</sup>
2000S1/12000S2	100	20	10	1200	5.3	3.5	2.2
			20	600	5.4	3.6	2.4
	200	10	10	1200	4.9	3.2	2.2
			20	600	5.1	3.3	2.1
	400	5	10	1200	4.5	3.7	2.0
			20	600	4.7	2.7	1.4
4000S1/10000S2	100	40	10	1000	5.5	3.6	2.2
			20	500	5.8	4.0	2.3
	200	20	10	1000	5.3	3.5	2.4
			20	500	5.4	3.8	2.3
	400	10	10	1000	4.3	3.3	1.6
			20	500	4.5	3.5	1.4
6000S1/8000S2	100	60	10	800	5.9	4.1	2.0
			20	400	6.5	4.2	2.2
	200	30	10	800	5.3	4.2	2.1
			20	400	5.7	4.5	2.4
	400	15	10	800	5.0	3.4	1.6
			20	400	5.8	3.5	1.2

<sup>1</sup>Number of total progeny allocated in the first progeny test (S1) and in the second progeny test(S2)

<sup>2</sup>Genomic test accuracy at the initial stage (S0) 0.5, 0.7, and 0.9

*Table 5 Average Years' worth of Genetic Gain (YGG) with the three-stage testing scenarios of the surrogate sire strategy with five elite donors above the conventional strategy that uses 50 males (SmallScenario)*

Progeny test resources <sup>1</sup>	Males progeny tested S1	Progeny/Male S1	Males progeny tested S2	Progeny/Male S2	YGG <sub>0.5</sub> <sup>2</sup>	YGG <sub>0.7</sub> <sup>2</sup>	YGG <sub>0.9</sub> <sup>2</sup>
2000S1/12000S2	100	20	10	1200	4.1	2.1	1.1
			20	600	4.4	2.2	1.2
	200	10	10	1200	3.0	2.1	1.2
			20	600	3.7	2.5	1.3
	400	5	10	1200	2.2	1.5	1.0
			20	600	2.2	1.4	1.0
4000S1/10000S2	100	40	10	1000	4.4	2.4	1.3
			20	500	4.5	2.5	1.2
	200	20	10	1000	4.1	2.2	1.1
			20	500	4.1	2.7	1.2
	400	10	10	1000	4.2	1.7	1.0
			20	500	4.2	2.0	1.8
6000S1/8000S2	100	60	10	800	4.5	3.1	1.6
			20	400	5.0	3.2	1.8
	200	30	10	800	4.6	2.1	1.3
			20	400	5.0	2.2	1.4
	400	15	10	800	4.1	1.7	1.2
			20	400	4.6	2.2	1.2

<sup>1</sup>Number of total progeny allocated in the first progeny test(S1) and in the second progeny test(S2)

<sup>2</sup>Genomic test accuracy at the initial stage (S0) 0.5, 0.7, and 0.9



*Table S1 Average Years' worth of Genetic Gain (YGG) with the three-stage testing scenarios of the surrogate sire strategy with one elite donor above the conventional strategy that uses 50 males (BigScenario)*

Progeny test resources <sup>1</sup>	Males progeny tested S1	Progeny/Male S1	Males progeny tested S2	Progeny/Male S2	YGG <sub>0.5</sub> <sup>2</sup>	YGG <sub>0.7</sub> <sup>2</sup>	YGG <sub>0.9</sub> <sup>2</sup>
2000S1/12000S2	100	20	10	1200	2.2	1.5	0.9
			20	600	2.3	1.7	0.9
	200	10	10	1200	2.1	1.6	1.0
			20	600	2.2	1.1	0.9
	400	5	10	1200	2.2	1.3	0.9
			20	600	2.3	1.5	0.8
4000S1/10000S2	100	40	10	1000	2.2	1.6	0.8
			20	500	2.1	1.6	0.8
	200	20	10	1000	2.1	2.1	0.9
			20	500	2.2	2.1	1.0
	400	10	10	1000	2.3	1.7	0.9
			20	500	2.3	2.0	0.9
6000S1/8000S2	100	60	10	800	2.5	2.0	1.0
			20	400	2.7	2.1	1.1
	200	30	10	800	2.4	1.9	1.1
			20	400	2.6	2.1	1.2
	400	15	10	800	2.3	2.0	0.9
			20	400	2.4	2.0	0.9

<sup>1</sup>Number of total progeny allocated in the first progeny test(S1) and in the second progeny test(S2)

<sup>2</sup>Genomic test accuracy at the initial stage (S0) 0.5, 0.7, and 0.9

*Table S2 Average Years' worth of Genetic Gain (YGG) with the three-stage testing scenarios of the surrogate sire strategy with five elite donors above the conventional strategy that uses 50 males (BigScenario)*

Progeny test resources <sup>1</sup>	Males progeny tested S1	Progeny/Male S1	Males progeny tested S2	Progeny/Male S2	YGG <sub>0.5</sub> <sup>2</sup>	YGG <sub>0.7</sub> <sup>2</sup>	YGG <sub>0.9</sub> <sup>2</sup>
2000S1/12000S2	100	20	10	1200	1.7	1.2	0.6
			20	600	1.6	1.2	0.8
	200	10	10	1200	1.6	1.2	0.6
			20	600	1.5	1.0	0.3
	400	5	10	1200	1.6	1.1	0.7
			20	600	1.6	1.2	0.8
4000S1/10000S2	100	40	10	1000	1.7	1.3	0.7
			20	500	1.7	1.2	0.6
	200	20	10	1000	1.6	1.3	0.4
			20	500	1.6	1.4	0.6
	400	10	10	1000	1.3	1.3	0.3
			20	500	1.4	1.4	0.4
6000S1/8000S2	100	60	10	800	1.8	1.1	0.4
			20	400	1.8	1.4	0.7
	200	30	10	800	1.6	1.2	0.5
			20	400	1.6	1.2	0.5
	400	15	10	800	1.7	1.2	0.7
			20	400	1.6	1.2	0.9

<sup>1</sup>Number of total progeny allocated in the first progeny test(S1) and in the second progeny test(S2)

<sup>2</sup>Genomic test accuracy at the initial stage (S0) 0.5, 0.7, and 0.9

