Long-range inputs and H-current regulate different modes of operation in a multiscale model of mouse M1 microcircuits

Salvador Dura-Bernal[1]
Samuel A Neymotin[1,2]
Benjamin A Suter[3]
Gordon M G Shepherd[4]
William W Lytton[1,5]

[1] Dept. Physiology & Pharmacology, SUNY Downstate, Brooklyn, NY
[2] Brown University
[3] IST Austria
[4] Department of Physiology, Northwestern University
[5] Dept. Neurology, Kings County Hospital Center, Brooklyn, NY

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Abstract

We developed a biologically detailed multiscale model of mouse primary motor cortex (M1) microcircuits, incorporating data from several recent experimental studies. The model simulates at scale a cylindrical volume with a diameter of 300 m and cortical depth 1350 m of M1. It includes over 10,000 cells distributed across cortical layers based on measured cell densities, with close to 30 million synaptic connections. Neuron models were optimized to reproduce electrophysiological properties of major classes of M1 neurons. Layer 5 corticospinal and corticostriatal neuron morphologies with 700+ compartments reproduced cell 3D reconstructions, and their ionic channel distributions were optimized within experimental constraints to reproduce in vitro recordings. The network was driven by the main long-range inputs to M1: posterior nucleus (PO) and ventrolateral (VL) thalamus PO, primary and secondary somatosensory cortices (S1, S2), contralateral M1, secondary motor cortex (M2), and orbital cortex (OC). The network local and long-range connections depended on pre- and post-synaptic cell class and cortical depth. Data was based on optogenetic circuit mapping studies which determined that connection strengths vary within layer as a function of the neuron's cortical depth. The synaptic input distribution across cell dendritic trees – likely to subserve important neural coding functions – was also mapped using optogenetic methods and incorporated into the model. We employed the model to study the effect on M1 of increased activity from each of the long-range inputs, and of different levels of H-current in pyramidal tract-projecting (PT) corticospinal neurons. Microcircuit dynamics and information flow were quantified using firing rates, oscillations, and information transfer measures (Granger causality and normalized transfer entropy). Simulation results support the hypothesis that M1 operates along a continuum of modes characterized by the degree of activation of intratelencephalic (IT), corticothalamic (CT) and PT neurons. The particular subset of M1 neurons targeted by different long-range inputs and the level of H-current in PT neurons regulated the different modes. Downregulation of H-current facilitated synaptic integration of inputs and increased PT output activity. Therefore, VL, cM1 and M2 inputs with downregulated H-current promoted an PT-predominant mode; whereas PO, S1 and S2 inputs with upregulated H-current favored an IT/CT-predominant mode, but a mixed mode where upper layer IT cells in turn activated PT cells if

H-current was downregulated. This evidenced two different pathways that can generate corticospinal output: motor-related inputs bypassing upper M1 layers and directly projecting to PT cells, and sensory-related inputs projecting to superficial IT neurons in turn exciting PT cells. These findings provide a hypothetical mechanism to translate action planning into action execution. Overall, our model serves as an extensible framework to combine experimental data at multiple scales and perform in silico experiments that help us understand M1 microcircuit dynamics, information flow and biophysical mechanisms, and potentially develop treatments for motor disorders.

1 Introduction

Primary motor cortex (M1) microcircuitry exhibits many differences with the tranditional canonical microcircuit proposed by Douglas and Martin [10]. These include differences in cytoarchitecture – *e.g.,* narrower layer 4 and wider layer 5B [57] –, in terms of cell classes – *e.g.,* corticospinal neurons – or connectivity – *e.g.,* strong asymmetric connection from upper layer IT cells to PT cells [1]. Corticocortical and corticostriatal cells in superficial layers seem to play a role in recurrently processing information at a local level and within thalamocortical regions, whereas deep corticospinal cells act output information (motor commands) from M1 to spinal cord circuits [55, 1]. These distinctions have profound consequences in terms of understanding neural dynamics and information processing [47], and should therefore be considered when developing detailed biophysical models of M1.

We developed a detailed model of M1 microcircuits that combines data at multiple scales from multiple studies: including cell classes morphology, physiology, densities and proportions; and long-range, local, and dendritic-level connectivity. Most data was obtained from studies on the same species and region – mouse M1 – and combined into a single theoretical framework. To integrate all these experimental findings we developed several methodological novelties including specifying connections as a function of normalized cortical depth (NCD) instead of the standard layers, and incorporating projection-specific dendritic distributions of synaptic inputs extracted from subcellular Channelrhodopsin-2- Assisted Circuit Mapping (sCRACM) studies [17, 51]. Additionally, we developed a novel tool, NetPyNE, to help develop, simulate and analyze data-driven biological network models in NEURON.

Our model, which contains over 10 thousand neurons, 30 miliion synapses and models a cylindric cortical volume with a depth of 1350 μm and a diameter of 300 μm constitutes the most biophysically detailed model of mouse M1 microcircuits available. It enables studying the non-intuitive dynamics and interactions occurring across multiple scales, with a level of resolution and precision which is not yet available experimentally. Cortical circuits can have extremely disparate responses to changes in their cellular and subcellular properties; detailed simulation provides a means to systematically explore the circuit-level consequences of manipulations of the cellular components and wiring of the system.

In this study we explore the effect on M1 of increased activity from the different long-range inputs projectig into M1; and of regulation of H-current in PT corticospinal cells. We evaluate the hypothesis that different that M1 operates along a continuum of modes characterized by the level of activation of intratelencephalic (IT), corticothalamic (CT) and PT neurons, and that long-range inputs and H-current modulates these which modes predominate.

2 Methods

2.1 Morphology and physiology of neuron classes

Seven excitatory pyramidal cell and two interneuron cell models were employed in the network. Their morphology and physiological responses are summarized in Fig. 1.

In previous work we developed layer 5B PT corticospinal cell and L5 IT corticostriatal cell models

that reproduce in vitro electrophysiological responses to somatic current injections, including sub- and super-threshold voltage trajectories and f-I curves [39, 50]. To achieve this we optimized the parameters of the Hodgkin-Huxley neuron model ionic channels – Na, Kdr, Ka, Kd, HCN, CaL, CaN, KCa – within a range of values constrained by literature. The corticospinal and corticostriatal cell model morphologies had 706 and 325 compartments, respectively, digitally reconstructed from 3D microscopy images. We further improved the PT model by 1) increasing the concentration of Ca2+ channels ("hot zones") between the nexus and apical tuft, following parameters published in [15]; 2) lowering dendritic Na+ channel density in order to increase the threshold required to elicit dendritic spikes, which then required adapting the axon sodium conductance and axial resistance to maintain a similar f-I curve; 3) replacing the HCN channel model and distribution with a more recent implementation by Migliore [37]. The new HCN channel reproduced a wider range of experimental observations than our previous implementation [24], including the change from excitatory to inhibitory effect in response to synaptic inputs of increasing strength [13]. This was achieved by including a shunting current proportional to I_{h}. We tuned the HCN parameters (lk and $v_{rev_l k}$) – originally developed for a CA3 pyramidal neuron [37] – and passive parameters to reproduce these experimental findings using the corticospinal cell, while keeping its somatic f-I curve consistent with recordings [50].

The network model includes five other excitatory cell classes: layer 2/3, layer 4, layer 5B and layer 6 IT neurons and layer 6 CT neurons. Since our focus was on the role of L5 neurons, other cell classes were implemented using simpler models as a trade-off to enable running a larger number of exploratory network simulations. Previously we had optimized 6-compartment neuron models to reproduce somatic current clamp recordings from two IT cells in layers 5A and 5B. The layer 5A cell had a lower f-I slope (X) and higher rheobase (X) than that in layer 5B (X and X). Based on our own and published data, we found two broad IT categories based on projection and intrinsic properties: corticocortical IT cells found in upper layers 2/3 and 4 which exhibited a lower f-I slope (avg X) and higher rheobase (avg X) than IT corticostriatal cells in deeper layers 5A, 5B and 6 (avg X and X) [57, 50, 41]. CT neurons' f-I rheobase and slope (X and X) was closer to that of corticocortical neurons [41]. We therefore employed the layer 5A IT model for layers 2/3 and 4 IT neurons and layer 6 CT neurons, and the layer 5B IT model for layers 5A, 5B and 6 IT neurons. We further adapted cell models by modifying their apical dendrite length to match the average cortical depth of the layer, thus introducing small variations in the firing responses of neurons across layers.

We implemented models for two major classes of GABAergic interneurons [14]: parvalbumin-expressing fast-spiking (PV) and somatostatin-expressing low-threshold spiking neurons (SOM). We employed existing simplified 3-compartment (soma, axon, dendrite) models [25] and increased their dendritic length to better match the average f-I slope and rheobase experimental values of cortical basket (PV) and Martinotti (SOM) cells (Neuroelectro online database [52]).

2.2 Microcircuit composition: neuron locations, densities and ratios

We modeled in full scale a cylindric volume of mouse M1 cortical microcircuits with a 300 μm diameter and 1350 μm height (cortical depth) with a total of 10,171 neurons Fig. 1A. The cylinder diameter was chosen to approximately match the horizontal dendritic span of a corticospinal neuron located at the center, consistent with the approach used in the HBP model of rat S1 microcircuits [34]. Mouse cortical depth and boundaries for layers 2/3, 4, 5A, 5B and 6 were based on our published experimental data [55, 1, 57]. Although traditionally M1 has been considered an agranular area lacking layer 4, we recently identified M1 pyramidal neurons with the expected prototypical physiological, morphological and wiring properties of layer 4 neurons [57], and therefore incorporated this layer in the model.

Cell classes present in each layer were determined based on mouse M1 studies [14, 50, 1, 57, 41, 25, 38]. IT cell populations were present in all layers, whereas the PT cell population was confined to layer 5B, and the CT cell population only occupied layer 6. SOM and PV interneuron populations were distributed in each layer, with layer 4 and 5B sharing the same populations. Neuronal densities (neurons per mm^3) for each layer Fig. 1A were taken from a histological and imaging study of

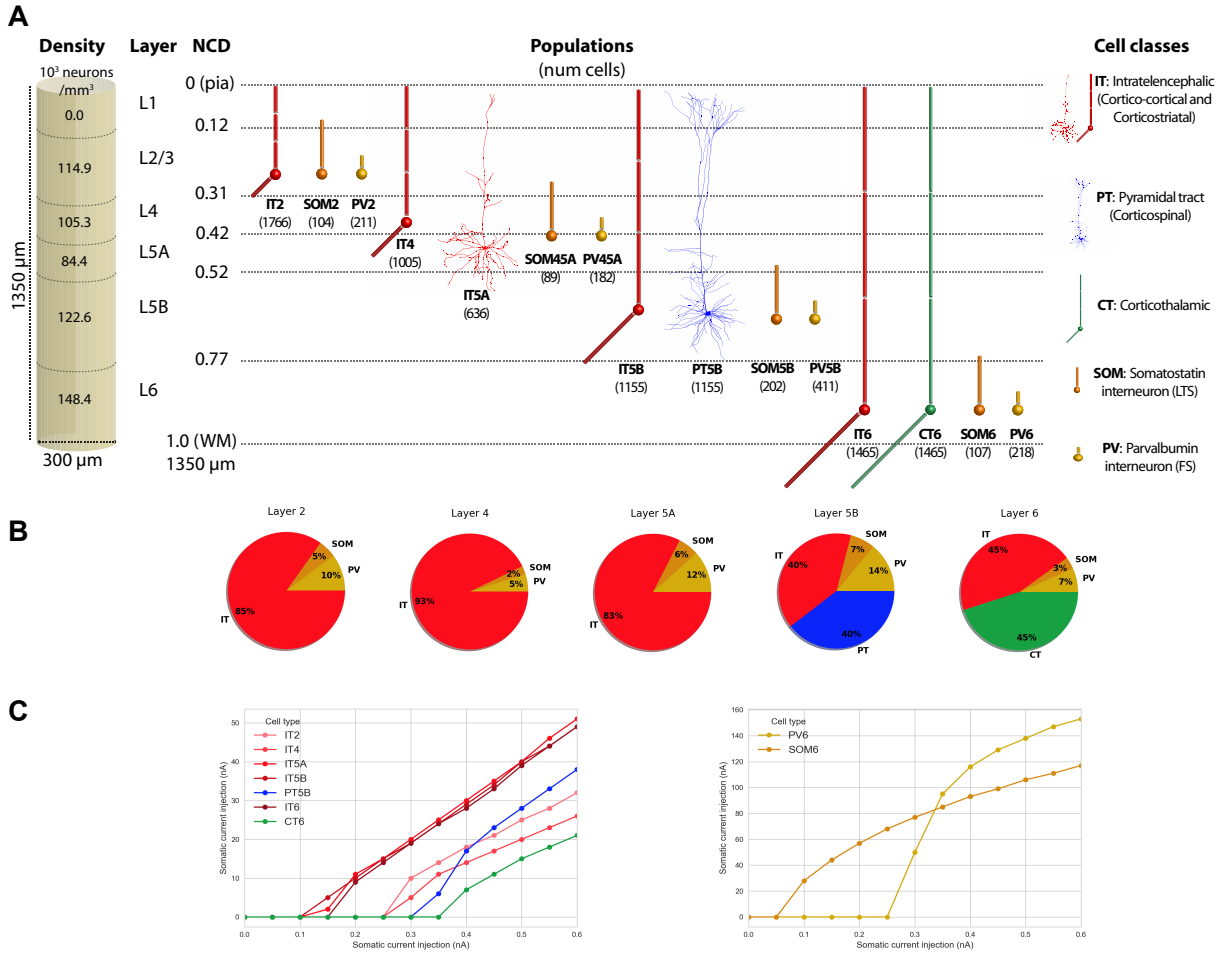

Figure 1: **Microcircuit dimensions and neuron densities, composition, classes, morphologies and f-I response.** A) Volume, cell density per layer, populations, number of cells, and morphologies; B) Proportion of cell classes per layer; C) f-I curve for each excitatory and inhibitory cell class.

mouse agranaular cortex [53]. The proportion of excitatory to inhibitory neurons per layer was obtained from mouse S1 data [28]. The proportion of IT to PT and IT to CT cells in layers 5B and 6, respectively, were both estimated as 1:1 [14, 50, 58]. The ratio of PV to SOM neurons per layer was estimated as 2:1 based on mouse M1 and S1 studies [22, 54]. The number of cells for each population was calculated based on the modeled cylinder dimensions, layer boundaries and neuronal proportions and densities per layer.

2.3 Local connectivity

We calculated local connectivity between M1 neurons Fig. 2 by combining data from multiple studies. Data on excitatory inputs to excitatory neurons (IT, PT and CT) was primarily derived from mapping studies using whole-cell recording, glutamate uncagingbased laser-scanning photostimulation (LSPS) and channelrhodopsin-2-assisted circuit mapping (CRACM) analysis [55, 1, 57, 58]. Connectivity data was postsynaptic cell class-specific and employed normalized cortical depth (NCD) instead of layers as the primary reference system. Unlike layer definitions which are interpreted differently, NCD provides a well-defined, consistent and continuous reference system, depending only on two readily-identifiable landmarks: pia (NCD=0) and white matter (NCD=1). Incorporating NCD-based connectivity into our

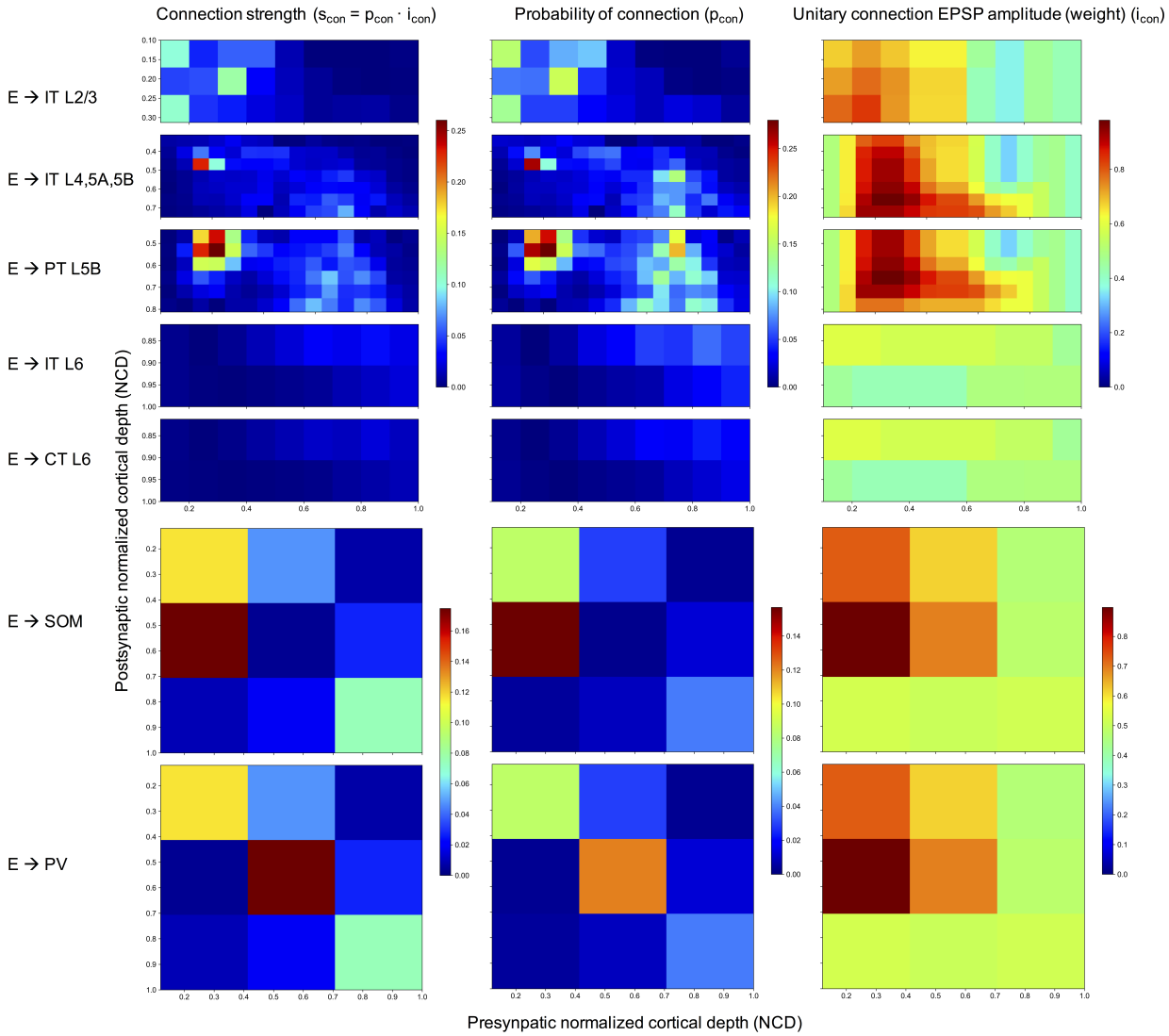

Figure 2: **Microcircuit local connectivity.** A) Volume, cell density per layer, populations, number of cells, and morphologies; B) Proportion of cell classes per layer; C) f-I curve for each excitatory and inhibitory cell class.

model allowed us to capture wiring patterns down to a 100 μm spatial resolution, well beyond traditional layer-based cortical models. M1 connectivity varied systematically within layers. For example, the strength of inputs from layer 2/3 to L5B corticospinal cells depends significantly on cell soma depth, with upper neurons receiving much stronger input [1].

Connection strength thus depended on presynaptic NCD and postsynatic NCD and cell class. For postsynaptic IT neurons with NCD ranging from 0.0 to 0.3125 (layer 2/3) and 0.8125 to 1.0 (layer 6) we determined connection strengths based on data from [55] with cortical depth resolution of 140 μm-resolution. For postsynaptic IT and PT neurons with NCD between 0.3125 and 0.8125 (layers 4, 5A and 5B) we employed connectivity strength data from [1] with cortical depth resolution of 100 μm. For postsynaptic CT neurons in layer 6 we used the same connection strengths as for layer 6 IT cells [55], but reduced to 62% of original values, following published data on the circuitry of M1 CT neurons [58]. Diagonal elements in the connectivity matrices were increased by 20% to compensate for the

underestimates resulting from LSPS CRACM analysis. Our data [58] also suggested that connection strength from layer 4 to layer 2/3 IT cells was similar to that measured in S1, so for these projections we employed values from Lefort's S1 connectivity strength matrix [28]. Experimentally, these connections were found to be four times stronger than in the opposite direction – from layer 2/3 to layer 4 – so we decreased the latter in the model to match this ratio.

Following previous publications [23, 28], we defined connection strength (s_{con}) between two populations, as the product of their probability of connection (p_{con}) and the unitary connection somatic EPSP amplitude in mV (i_con), i.e. $s_{con} = p_{con} \times i_{con}$. We employed this equivalence to disentangle the connection s_{con} values provided by the above LSPS studies into p_{con} and i_{con} values that we could use to implement the model. First, we rescaled the LSPS raw current values in pA [1, 55, 57, 58] to match s_{con} data from a paired recording study of mouse M1 L5 excitatory circuits [23]. Next, we calculated the M1 NCD-based i_{con} matrix by interpolating a layerwise unitary connection EPSP amplitude matrix of mouse S1 [28], and thresholding values between 0.3 and 1.0 mV. Finally, we calculated the probability of connection matrix as $p_{con} = s_{con}/i_{con}$.

To implement i_{con} values in the model we calculated the required NEURON connection weight of an excitatory synaptic input to generate a somatic EPSP of 0.5 mV at each neuron segment. This allowed us to calculate a scaling factor for each segment that converted i_{con} values into NEURON weights, such that the somatic EPSP response to a unitary connection input was independent of synaptic location. This is consistent with experimental evidence showing synpatic conductances increased with distance from soma, to normalize somatic EPSP amplitude of inputs within 300 μm of soma [32]. Following this study, scaling factor values above 4.0 – such as those calculated for PT cell apical tufts – were thresholded to avoid overexcitability in the network context where each cell receives hundreds of inputs that interact nonlinearly [48, 3]. For morphologically detailed cells (layer 5A IT and layer 5B PT), the number of synaptic contacts per unitary connection (or simply, synapses per connection) was set to five, an estimated average consistent with the limited mouse M1 data [18] and rat S1 studies [5, 34]. Individual synaptic weights were calculated by dividing the unitary connection weight (i_{con}) by the number of synapses per connection. Although the method does not account for nonlinear summation effects [48], it provides a reasonable approximation and enables employing a more realistic number and spatial distribution of synapses, which may be key for dendritic computations [30]. For the remaining cell models, all with six compartments or less, a single synapse per connection was used.

For excitatory inputs to inhibtory cell types (PV and SOM) we started with the same values as for IT cell types but adapted these based on the specific connectivity patterns reported for mouse M1 interneurons [2, 58] Fig. 2. Following the layer-based description in these studies, we employed three major subdivisions: layer 2/3 (NCD 0.12 to 0.31), layers 4, 5A and 5B (NCD 0.31 to 0.77) and layer 6 (NCD 0.77 to 1.0). We increased the probability of layer 2/3 excitatory connections to layers 4, 5A and 5B SOM cells by 50% and decreased that to PV cells by 50% [2]. We implemented the opposite pattern for excitatory connections arising from layer 4,5A,5B IT cells such that PV interneurons received stronger intralaminar inputs than SOM cells [2]. The model also accounts for layer 6 CT neurons generating relatively more inhibition than IT neurons [58]. Inhibitory connections from interneurons (PV and SOM) to other cell types were limited to neurons in the same layer [22], with layers 4, 5A and 5B combined into a single layer [38]. Probability of connection decayed exponentially with the distance between the pre- and post-synaptic cell bodies with length constant of 100 μm [12, 11].

Excitatory synapses consisted of colocalized AMPA (rise, decay τ: 0.05, 5.3 ms) and NMDA (rise, decay τ: 15, 150 ms) receptors, both with reversal potential of 0 mV. The ratio of NMDA to AMPA receptors was 1.0 [**?**], meaning their weights were each set to 50% of the connection weight. NMDA conductance was scaled by $1/(1 + 0.28Mg \exp(0.062V))$; Mg = 1mM [21]. Inhibitory synapses from SOM to excitatory neurons consisted of a slow GABAA receptor (rise, decay τ: 2, 100 ms) and GABAB receptor, in a 90% to 10% proportion; synapses from SOM to inhibitory neurons only included the slow GABAA receptor; and synapses from PV to other neurons consisted of a fast GABAA receptor (rise, decay τ: 0.07, 18.2). The reversal potential was -80 mV for GABAA and -95 mV for GABAB. The GABAB synapse was modeled using second messenger connectivity to a G protein-coupled inwardly-rectifying potassium channel

(GIRK) [9]. The remaining synapses were modeled with a double-exponential mechanism.

Connection delays were estimated as 2 ms plus a variable delay depending on the distance between the pre- and postsynaptic cell bodies assuming a propagation speed of 0.5 m/s.

2.4 Long-range input connectivity

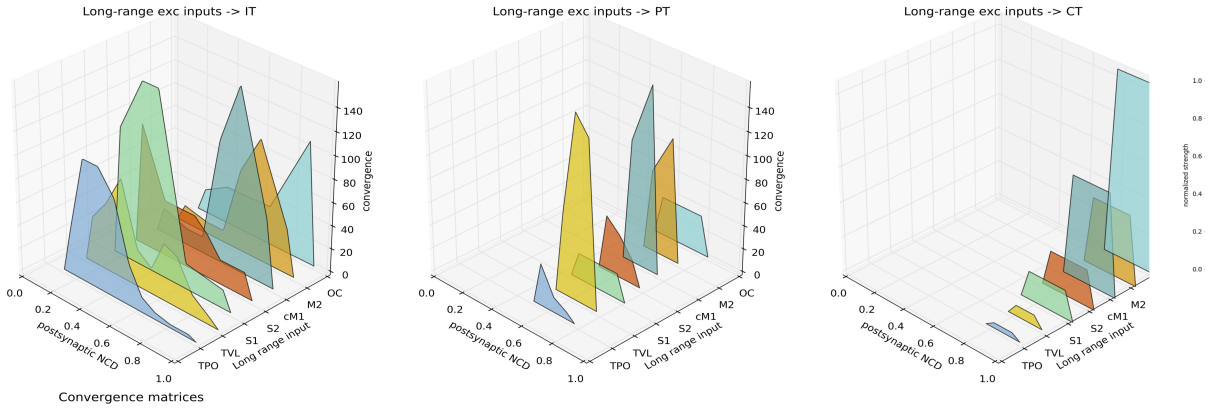

Figure 3: **Long-range inputs to M1 microcircuit.**

We added long-range input connections from seven regions that are known to project to M1: thalamus posterior nucleus (TPO), ventro-lateral thalamus (TVL), primary somatosensory cortex (S1), secondary somatosensory cortex (S2), contralateral primary motor cortex (cM1), secondary motor cortex (M2) and orbital cortex (OC). Each region consisted of a population of 1000 [7, 5] spike-generators (NEURON VecStims) that generated independent random Poisson spike trains with uniform distributed rates between 0 and 2 Hz [56, 16]; or 0 to 20 Hz [19, 20] when simulating increased input from a region. Previous studies – using the LSPS CRACM method – provided a measure of normalized input strength from these regions as a function of postsynaptic cell type and layer or NCD. Broadly, TPO [57, 58, 38], S1 [33] and S2 [51] projected strongly to IT cells in layers 2/3, 4 and 5A; TVL projected strongly to PT cells and moderately to IT cells [57, 58, 38]; cM1 and M2 projected strongly to IT and PT cells in layers 5B and 6 [17]; and OC projected strongly to layer 6 CT and IT cells [17]. We implemented these relations by estimating the maximum number of synaptic inputs from each region and multiplying that value by the normalized input strength for each postsynaptic cell type and NCD range. This resulted in a convergence value – average number of synaptic inputs to each postsynaptic cell – for each projection Fig. 3. We fixed all connection weights (unitary connection somatic EPSP amplitude) to 0.5 mV, consistent with rat and mouse S1 data [18, 7].

To estimate the maximum number of synaptic inputs per region, we made a number of assumptions based on the limited data available Fig. 3. First, we estimated the average number of synaptic contacts per cell as 8234 by rescaling rat S1 data [36] based on our own observations for PT cells [50] and contrasting with related studies [45, 8]; we assumed the same value for all cell types so we could use convergence to approximate long-range input strength. We assumed 80 % of synaptic inputs were excitatory vs. 20 % inhibitory [8, 34]; out of the excitatory inputs, 80 % were long-range vs. 20 % local [34, 49]; and out of the inhibitory inputs, 30 % were long-range vs. 70 % local [49]. Finally, we estimated the percentage of long-range synaptic inputs arriving from each region based on mouse brain mesoscale connectivity data [40] and other studies [35, 5, 36, 59, 4]: TVL, S1, cM1 each contributed 15 %, whereas TPO, S2, M2 and OC each accounted for 10 %.

2.5 Dendritic distribution of synaptic inputs

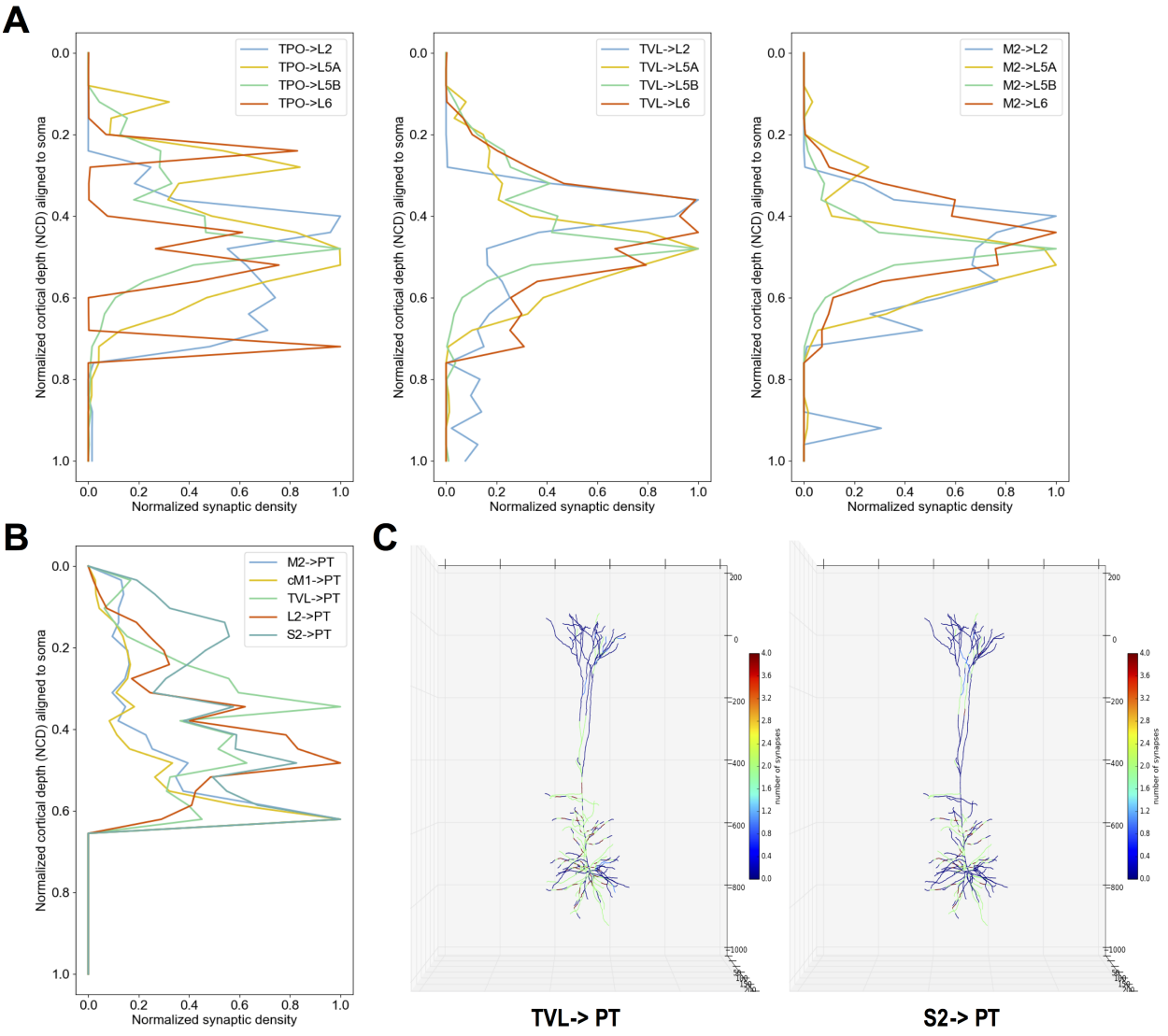

Figure 4: **Dendritic distribution of synaptic inputs.**

Experimental evidence demonstrates the location of synapses along dendritic trees follows very specific patterns of organization that depend on the brain region, cell type and cortical depth [42, 51]; these are likely to result in important functional effects [26, 27, 48]. We employed subcellular Channelrhodopsin-2-Assisted Circuit Mapping (sCRACM) data to estimate the synaptic density along the dendritic arbor – 1D radial axis – for inputs from TPO, TVL, M2 and OC to layers 2/3, 5A, 5B and 6 IT and CT cell [17] (Fig. 4A), and from layer 2/3 IT, TVL, S1, S2, cM1 and M2 to PT neurons [51] (Fig. 4B). To approximate radial synaptic density we divided the sCRACM map amplitudes by the dendritic length at each grid location, and averaged across rows. Once all network connections had been generated, synaptic locations were automatically calculated for each cell based on its morphology and the pre- and postsynaptic cell type-specific radial synaptic density function (Fig. 4C). Synaptic inputs from PV to excitatory cells were located perisomatically (50 μm around soma); SOM inputs targeted apical dendrites of excitatory neurons [38, 22]; and all inputs to PV and SOM cells targeted apical dendrites. For projections where no data synaptic distribution data was available – IT/CT, S1, S2 and cM1 to IT/CT cells – we assumed a uniform dendritic length distribution.

2.6 Model implementation, simulation and analysis

The model was developed using parallel NEURON [31] and NetPyNE [44] (www.neurosimlab.org/netpyne), a Python package to facilitate the development of biological neuronal networks in the NEURON simulator. NetPyNE emphasizes the incorporation of multiscale anatomical and physiological data at varying levels of detail. It converts a set of simple, standardized high-level specifications in a declarative format into a NEURON model. NetPyNE also facilitates organizing and running parallel simulations by taking care of distributing the workload and gathering data across computing nodes. It also provides a powerful set of analysis methods so the user can plot spike raster plots, LFP power spectra, information transfer measures, connectivity matrices, or intrinsic time-varying variables (eg. voltage) of any subset of cells. To facilitate data sharing, the package saves and loads the specifications, network, and simulation results using common file formats (Pickle, Matlab, JSON or HDF5), and can convert to and from NeuroML, a standard data format for exchanging models in computational neuroscience. Simulations were run on XSEDE supercomputers Comet and Stampede using the Neuroscience Gateway (NSG) and our own resource allocation.

3 Results

3.1 Cell type-, layer and sublayer-specific spontaneous firing rates and physiological oscillations

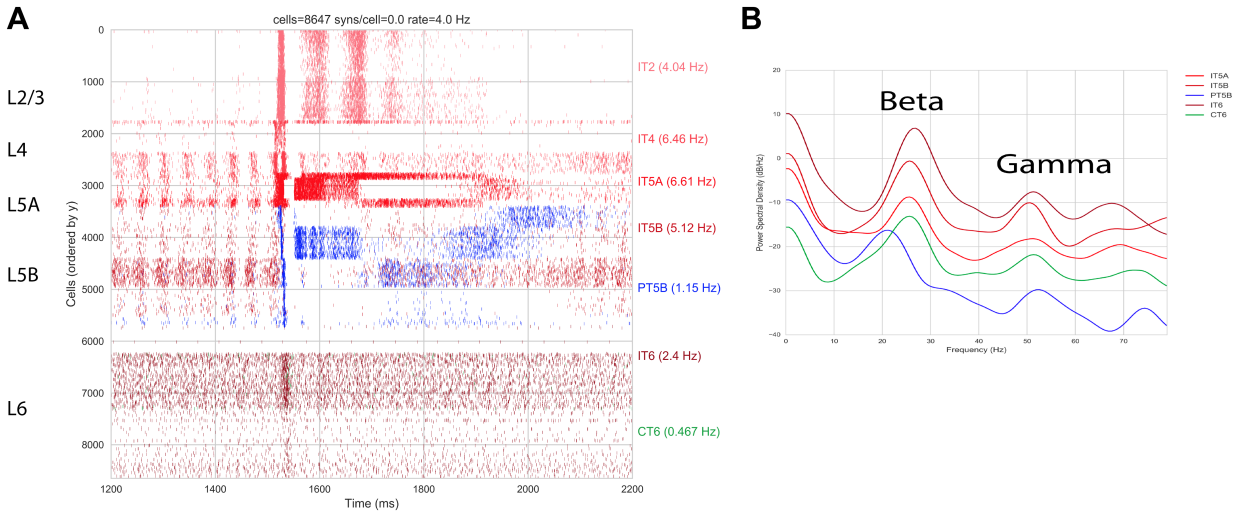

Figure 5: **Cell type-, layer and sublayer-specific spontaneous firing rates and physiological oscillations.** A) Raster plot with cells arranged by cortical depth. Note higher activity in upper vs lower layer 5B PT cells. B) Firing rate power density spectrum.

Simulations of M1 (Fig. 5A) exhibit spontaneous firing rates between 0 and 10 Hz for all populations, consistent with the experimental litearature [19, 29]. The sublaminar specificity of connections is evidenced by the gradual decrease of PT activity as cortical depth increases within layer 5B, reflecting experimental observations [1]. The circuit also showed emergence of physiological oscillations in the beta and gamma range (Fig. 5B) consistent with in vivo data [6, 43]. These oscillations emerged in the absence of rhythmic external inputs.

3.2 Sensory-related pathway and H-current modulation

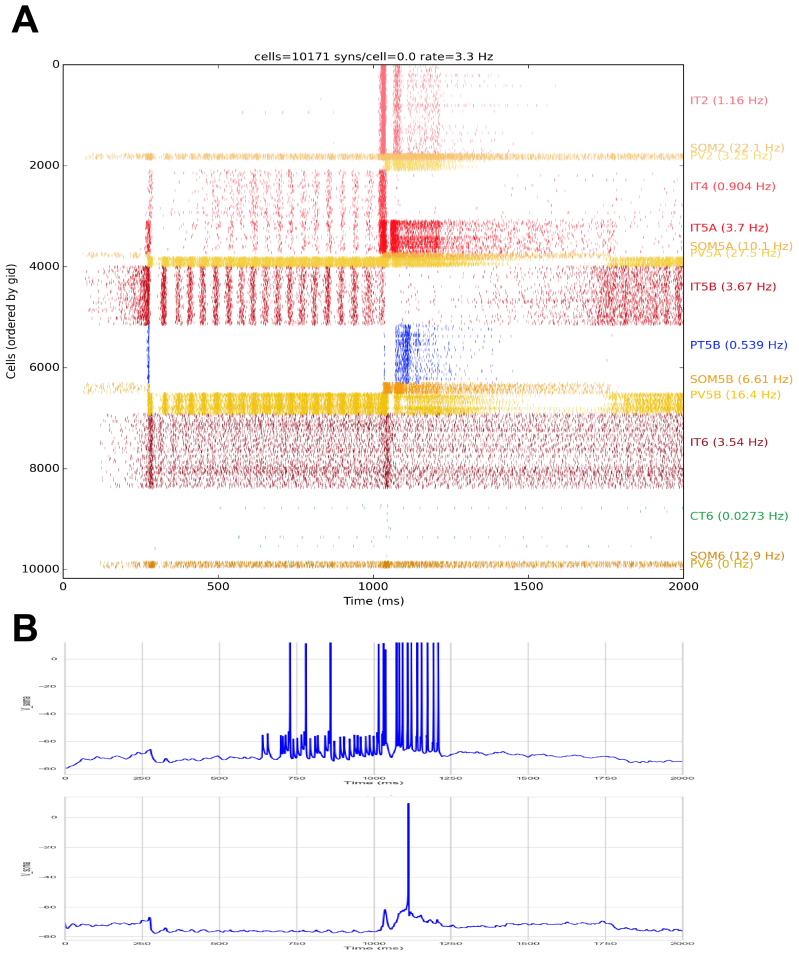

Figure 6: **Sensory input with upregulated H-current showing IT-predominant mode (low PT).** A) Raster plot. B) IT5A vs P5B example voltage traces.

Differnet combinations of long-range inputs and H-current levels lead to different modes of operation in M1. Increased rates (20 Hz) in PO, S1 and S2 inputs together with an upregulated H-current favored an IT/CT-predominant mode (with low PT activity) Fig. 6. The same long-range input levels with H-current downregulated lead to a mixed mode where upper layer IT cells in turn activated PT cells Fig. 7. Downregulation of H-current facilitated synaptic integration of inputs and increased PT output activity. This constitutes one of the pathways that can generate corticospinal output: sensory-related inputs projecting to superficial IT neurons in turn exciting PT cells.

3.3 Motor-related pathways and H-current modulation

Increased activation of VL, cM1 and M2 inputs with downregulated H-current promoted a PT-predominant mode Fig. 8. This evidences a second pathway that can result in corticospinal output: motor-related inputs bypassing upper M1 layers and directly projecting to PT cells. As with sensory-related inputs, upregulation of H-curerent in corticospinal cells also decreased their activity.

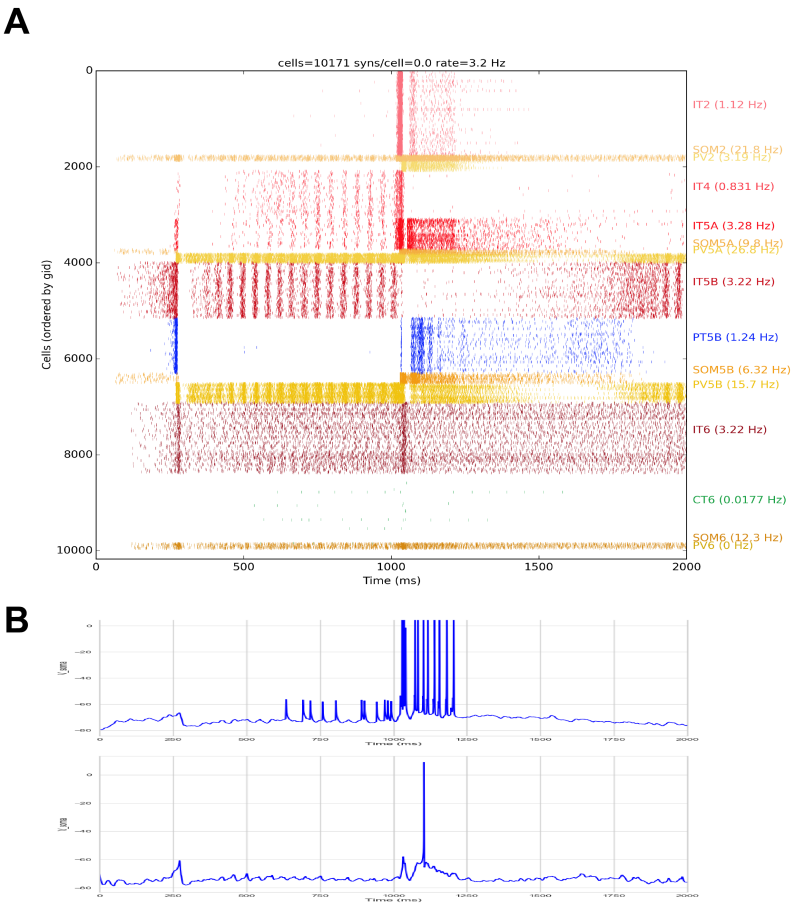

Figure 7: **Sensory input with downregulated H-current showing mixed mode (both IT and PT active** A) Raster plot. B) IT5A vs P5B example voltage traces. C) Information transfer (Granger and/or nTE) between long range inputs, upper layer ITs and L5B PTs. 4) Comparison of stimulus response delay and average firing rate of upper IT vs PT (N = 25 = 5 wiring x 5 input seeds)

4 Discussion

In this study we developed the most detailed computational model of mouse M1 microcircuits and corticospinal neurons up to date, based on an accumulated set of experimental studies. Overall, the model incorporates quantitative experimental data from 15 publications, thus integrating previously isolated knowledge into a unified framework. Unlike the real brain, this in silico system serves as a testbed that can be probed extensively and precisely, and can provide accurate measurements of neural dynamics at multiple scales. We employed the model to evaluate how long-range inputs from surrounding regions and molecular/pharmacological-level effects (e.g. regulation of HCN channel) modulate M1 dynamics.

We simulated the inputs from the main cortical and thalamic regions that project to M1, based on recent experimental data from optogenetic mapping studies. Consistent with experimental data [17, 51], this study evidenced two different pathways that can generate corticospinal output: motor-related inputs from VL, cM1 or M2 regions bypassing upper M1 layers and directly projecting to corticospinal cells, and sensory-related inputs from PO, S1 and S2 regions projecting to superficial IT neurons in turn exciting layer 5B corticospinal cells. Activation of OC region lead to increased layer 6 IT and CT activity and physiological oscillations in the beta and gamma range. Downregulation of corticospinal HCN lead to increased firing rate, as observed experimentally. This has been hypothesized as a potential mechanism to

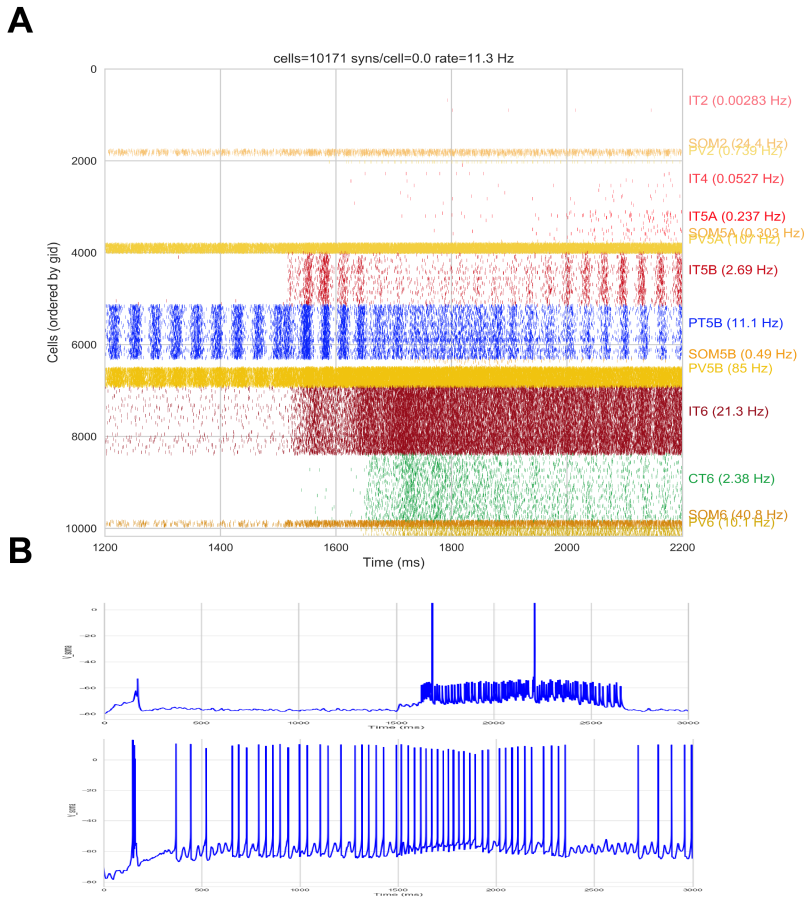

Figure 8: **Motor input with downregulated H-current showing PT-predominant mode.** A) Raster plot. B) IT5A vs P5B example voltage traces.

translate action planning into action execution [46].

Our study provides insights to help decipher the neural code underlying the brain circuits responsible for producing movement, and help understand motor disorders, including spinal cord injury. The computational model provides a framework to integrate experimental data and can be progressively extended as new data becomes available. This provides a useful tool for researchers in the field, who can use the framework to evaluate hypothesis and guide the design of new experiments.

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