

1 **Title:** Asymmetric migration decreases stability but increases resilience in a
2 heterogeneous metacommunity

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16 **Abstract:** Many natural communities are spatially distributed, forming a network of
17 subcommunities linked by migration. Migration patterns are often asymmetric and
18 heterogeneous, with important consequences on the ecology and evolution of the species. Here
19 we investigated experimentally how asymmetric migration and heterogeneous structure affect a
20 simple metacommunity of budding yeast, formed by one strain that produces a public good and a
21 non-producer strain that benefits from it. We find that asymmetric migration increases the
22 fraction of producers in all subpopulations of the metacommunity. Furthermore, asymmetric
23 migration decreases the metacommunity's tolerance to challenging environments, but increases
24 its resilience to transient perturbations. This apparent paradox occurs because tolerance to a
25 constant challenge depends on the weakest subpopulations of the network, while resilience to a
26 transient perturbation depends on the strongest ones.

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28 **One Sentence Summary:** Asymmetric migration decreases the stability of experimental yeast
29 metacommunities but increases their resilience to transient shocks.

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32 **Main Text:**

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34 Natural populations are spatially distributed, often in a way in which neighboring populations are
35 linked to each other by migration. These complex populations are called metapopulations (1–6)
36 or, when they contain several coexisting species, metacommunities (7, 8). Natural
37 metacommunities often have heterogeneous connectivity (some subpopulations have more
38 neighbors than others) and asymmetric migration patterns (the net flow of individuals between
39 any two subpopulations can be non-zero). Extreme examples are dendritic networks, in which
40 several nodes of each level connect to a single node in the next one (9). For example, river basins
41 give rise to dendritic metacommunities, with all tributaries being connected to their parent river
42 (and with highly asymmetric migration due to water currents) (9–15). Besides dendritic
43 networks, many natural metacommunities have heterogeneous connectivity and asymmetric
44 migration patterns (16, 17).

45 A major challenge is understanding how the structure of metacommunities influences the
46 ecology and evolution of the involved species. Spatial structure usually increases the biodiversity
47 of metacommunities (8, 18–23), and may enhance particular interactions such as cooperation
48 (24–26). It may also have profound effects on the ability of a species to survive environmental
49 deterioration and transient perturbations. This question has been studied extensively, especially
50 for metapopulations but also for metacommunities (20, 27–33). Given the complexity of these
51 systems, results are mixed: Depending on the conditions, spatial structure may increase (20, 27–
52 30) or decrease (29–32) the system’s ability to survive in a challenging environment. Results
53 from stability theory in dynamical systems may help identify the main factors determining each
54 outcome (34). For example, well-mixed populations and communities have been shown to cross
55 a tipping point as the environment deteriorates, leading to a sudden collapse of the population
56 rather than a smooth decline towards extinction (35–37). This detailed understanding allows
57 predicting how different factors affect the stability of the populations, and has also helped to
58 demonstrate that generic indicators such as critical slowing down can predict the collapse of the
59 system (34, 35, 37–39). Extending this approach to metacommunities requires taking into
60 account the effect of migration on the density of the subpopulations. This effect is often
61 neglected, as the most studied effect of migration is to propagate species to locations where they
62 are not present (5, 6, 40–42). Yet in many cases migration may be strong enough to have a

63 significant effect in the density of the subpopulations—the so-called mass effects (7, 43)—,
64 which in turn may determine their survival and composition (via density-dependent selection).

65 We addressed these questions experimentally taking advantage of the high-throughput and short
66 generation times of microbial microcosms, which allow us to study metacommunities over
67 hundreds of generations. We chose a simple community formed by two strains of budding yeast
68 (*Saccharomyces cerevisiae*) growing on sucrose. These cells cannot metabolize sucrose directly;
69 one of the strains (the producer) produces an enzyme that breaks down sucrose into glucose and
70 fructose, which can be metabolized by the cells. This reaction does not take place inside the cell,
71 but in the periplasmic space between the cell membrane and the cell wall, so most of the
72 products (~99%) diffuse away, acting as a public good that can be used by any nearby cell. Cells
73 that do not produce the enzyme can benefit from the public good without paying the cost of
74 producing it (**Fig 1A**) (44, 45). This simple community therefore contains three important
75 interactions: cooperation among individuals of the producer strain, facilitation between the
76 producer strain and the non-producer one, and competition for resources. Also, the production of
77 a public good gives rise to an Allee effect (i.e. the growth rate of the population is higher at
78 intermediate densities than at low densities, due to the accumulation of public good). Because of
79 this Allee effect, the population size does not diminish smoothly when conditions deteriorate, but
80 undergoes a catastrophic collapse when the density of producers falls below a critical threshold
81 (35, 36). This sudden collapse makes our experimental system ideal to study the stability of the
82 metacommunity.

83 Additionally, this system presents both frequency- and density-dependent selection. When the
84 population contains enough producers to make the public good plentiful, non-producers have the
85 advantage of not paying the production cost, so they will increase in frequency. However, if too
86 few producers are present the public good will be so scarce that the small amount imported by
87 producers before it diffuses away will allow them to grow faster than the non-producers. These
88 two effects create negative frequency-dependent selection, in which each strain is at a
89 disadvantage when too frequent, and the population tends toward an intermediate fraction of both
90 strains (**Fig. 1A, left inset**) (44, 46). This equilibrium fraction of producers in turn depends on
91 the overall density of the population, giving rise to density-dependent selection: the same
92 fraction of producers in a denser population entails more producer cells, hence more public good

93 and greater advantage for the non-producers. Therefore, the equilibrium fraction of producers is
94 lower in denser populations (**Fig. 1A, right inset**)(36).

95 This density-dependent selection may have important consequences in metacommunities,
96 because asymmetric migration may lead to unequal densities in different nodes, and hence
97 unequal fractions of producers. To study this effect, we compared isolated well-mixed
98 communities to metacommunities with heterogeneous connectivity, in which a central node is
99 connected to 9 side nodes (star network in **Fig. 1B**). As a control, we also tested
100 metacommunities with homogeneous connectivity and symmetric migration (fully connected
101 networks in **Fig. 1B**). In every time step, a fraction m of the cells in each node migrate to
102 neighboring nodes, distributing evenly among them (**Fig. 1B**) (47). This fraction m of migrants is
103 independent of the number of neighboring nodes, as is for example the case for organisms with a
104 specialized dispersal stage. This migration scheme leads to asymmetric migration in
105 heterogeneous networks, because the proportion of migrants traversing a link in each direction
106 depends on the degree of the two connected nodes, with net migration flowing from the less
107 connected to the most connected one (**Fig. 1B, center**). In the star network, net migration flows
108 from the side nodes towards the center, which should lead to lower density on the sides and
109 higher in the center, and therefore an increased fraction of producers in the sides and decreased
110 in the center.

111 To determine how heterogeneous networks affect the frequency of producers in the
112 metacommunity, we performed experiments comparing isolated nodes, 10-node star networks,
113 and 10-node fully-connected networks. All populations underwent a daily dilution-migration-
114 growth procedure: At the beginning of each day, all cultures were diluted in fresh medium by a
115 factor 650. A fraction m of the remaining cells in each node then migrated to neighboring nodes,
116 distributing uniformly among them (**Fig. 1B**). The cells then grew for 23 hours, until the next
117 dilution-migration step. We chose a migration rate $m=0.6$, which corresponds to around 6% per
118 generation (cells undergo around 10 generations in every growth cycle).

119 We found that network heterogeneity increases the overall fraction of producers. Regardless of
120 the initial fraction, isolated populations and fully connected networks converge to having around
121 8% producers, while star networks show a two-fold increase over this value (**Fig. 1C**).

122 Heterogeneous metacommunity structure therefore favors the public-goods producers in this
123 system.

124 Furthermore, producer fraction increased in all nodes of the heterogeneous networks, including
125 the central one (**Fig. 2A**). As predicted, migration in the star network resulted in lower density
126 for the side nodes and higher density for the central node, as compared to isolated populations
127 (**Fig. 2B**). Yet both side and central nodes showed an increased fraction of producers (**Fig. 2A**).
128 This increase in producer fraction is expected for the side nodes, as they experience a higher
129 effective dilution rate due to asymmetric migration, thus leading to a decrease in cell density that
130 favors producers due to the density-dependent selection (**Fig. 1A**). In contrast, the increased
131 density in the central node should produce a decrease in the fraction of producers, as is indeed
132 the case at the beginning of the experiment (**Fig. 2A**, days 1-5). However, the central node
133 receives a large number of migrants from the side nodes. Therefore, once the fraction of
134 producers in the side nodes is high enough, immigration into the central node increases its
135 fraction of producers in spite of its high cell density. The heterogeneous network structure in our
136 star network therefore increases the producer fraction in all nodes throughout the network.

137 To further understand these effects, we built a simple phenomenological model that incorporates
138 negative frequency-dependent selection and density dependent selection. In this model, both
139 strains grow logistically up to a common carrying capacity K . Their growth rates increase with
140 the amount of available public good, which we assume to be proportional to the density of
141 producers (N_p). We assumed Michaelis-Menten dynamics for this increase, with k_M being the
142 density of producers needed to produce enough public good to bring growth rate to half its
143 maximum value. Because of the small fraction of sugars imported directly by producers, they
144 benefit from an extra quantity ε of public good (44). Finally, producers pay a small cost c for
145 producing the public good (**Fig. 2C**). We used this model to simulate daily growth followed by
146 650x dilution and migration. This simple model successfully reproduces the increase in producer
147 fraction that we observed experimentally (**Fig. 2D,E**).

148 This phenomenological model also predicts the impact of heterogeneous structure on the
149 metacommunity's stability in the face of deteriorating environments. In isolated populations,
150 increasing the daily dilution factor eventually leads to a catastrophic collapse of the population

151 (35, 36). Our model predicts that migration in the star network will favor this collapse, which
152 will occur with milder dilution rates when migration rate is higher (**Fig. 3A**). This anticipated
153 collapse happens because the lower density of the side nodes makes them incapable of sustaining
154 the combined burden of dilution and net outward migration. Once the side nodes have collapsed,
155 the central node receives no inward flux yet still has an outward flux of migrants, thus causing
156 the central node to go extinct soon thereafter. Our model therefore predicts that heterogeneous
157 metacommunities will go extinct in milder environmental conditions than isolated populations,
158 despite the higher producer fraction present in the heterogeneous network.

159 To experimentally test this prediction of premature collapse, we compared the survival ability of
160 isolated populations with that of the 10-node star networks over daily dilution factors from 400 to
161 2000. As predicted by the model, star networks collapse at lower dilution rates (**Fig 3B**). For
162 example, at a dilution rate of 1300 all four isolated populations survived, whereas none of the
163 three star networks survived (**Fig 3B**, inset). Therefore, isolated populations are better able to
164 survive challenging environments than populations connected in a heterogeneous network.

165 Previous reports have shown that a system close to a catastrophic collapse is less capable of
166 recovering from harmful shocks (35, 36, 38, 46). In line with this, one would expect
167 heterogeneous networks to be less likely to recover from perturbations than isolated populations,
168 since for a given dilution rate the heterogeneous networks are closer to the tipping point. We
169 investigated this prediction with the model, finding that it is only fulfilled in the immediate
170 vicinity of the heterogeneous network's tipping point. For most conditions, asymmetric
171 migration increases the resilience of the metacommunity to a transient shock (in particular a
172 transient decrease in population density) (**Fig. 4A**). To test whether this result depends on the
173 nature of the perturbation, we investigated the metacommunity's resilience to both dilution
174 shocks and growth rate shocks (i.e. decreasing the growth rate during one cycle). In both cases,
175 we find higher resilience for star networks than for isolated nodes (**Fig. 4B**).

176 We tested experimentally this counterintuitive prediction of higher resilience in the
177 metacommunity, by subjecting yeast populations to a growth-inhibiting high-salt environment
178 (32 g/L) during one day. As predicted by the model, all three heterogeneous networks recovered
179 after the shock, while four of the five isolated populations went extinct (**Fig. 4C**). We therefore

180 find that, despite being less able to survive sustained exposure to challenging environments,
181 heterogeneous networks are more resilient to transient environmental perturbations.

182 The surprising resilience of our star network is due to the increase in both density and producer
183 fraction at the central node, which combine to increase the total number of producers present in
184 the population. The salt shock leads to a smaller population in every node, which could take the
185 density of producers below the threshold required for population survival. The increased number
186 of producers allows the central node to survive perturbations that would drive isolated
187 populations extinct. The side nodes of the network are not so resilient (because of their lower
188 density), but they can be reseeded from the central node once the shock is over (**Fig 4D**; note that
189 the density of the central node still decreases during the first cycle after the shock, as a
190 consequence of the outbound migration which is reseeding the side nodes, which is not yet
191 compensated by any significant influx from them).

192 These results highlight that stability and resilience may be determined by different factors in a
193 complex system. In our heterogeneous metacommunities, stability depends on the weakest
194 elements of the system—the side nodes—while resilience depends on the strongest one.

195 Our findings are a direct consequence of density-dependent selection, which is often linked to
196 frequency-dependent selection but not equivalent to it. In the absence of density dependence, our
197 system would have the same equilibrium producer fraction across the whole network, regardless
198 of its topology and migration scheme. Many systems subject to frequency-dependent selection
199 also exhibit density-dependent selection, yet this effect is often neglected in theoretical models
200 (for example the replicator equation (48), which only considers fractions). Our results therefore
201 highlight the important consequences of feedback between ecological and evolutionary dynamics
202 in spatially distributed populations.

203 Our migration scheme links asymmetric migration with heterogeneous connectivity, because the
204 proportion m of outgoing migrants does not depend on the number of neighbors of their home
205 node. This would be true for example for organisms with an specialized dispersal stage.
206 However, migration patterns may differ across species. An opposite assumption would be that
207 flux between two nodes is symmetric regardless of the connectivity. Many natural systems will
208 probably be between these two extremes, and as long as heterogeneous connectivity produces

209 some degree of asymmetric migration our qualitative results will be relevant. Future work that
210 investigates alternative migration schemes will help disentangle the relative contributions of
211 asymmetric migration and heterogeneous connectivity.

212 The seemingly paradoxical result of lower distance to the tipping point but higher resilience to
213 transient perturbations may be a common feature of asymmetry in networked systems: collapse
214 in steady state is dictated by the weaker elements, while resilience to transient perturbations is
215 dictated by the stronger ones. This is especially important given the pervasiveness of
216 heterogeneous networks (such as scale-free networks) in nature, and may have parallels in other
217 complex systems such as power grids or human populations (49).

218

219 **Materials and Methods**

220 Strains

221 Both strain derive from haploid cells BY4741 (mating type a, EUROSCARF). The producer
222 strain, JG300B (44), has a wild-type SUC2 gene, and can therefore produce invertase for the
223 breakdown of sucrose. It has a mutated HIS3 gene (*his3ΔI*), therefore being a histidine
224 auxotroph. It also expresses YFP constitutively. The non-producer strain, JG210C (44), has a
225 deletion of the SUC2 gene, so it does not produce invertase. It has a wild-type HIS3 gene and
226 expresses dTomato constitutively.

227

228 Culture conditions

229 Before every experiment, we picked a single colony of each cell type from a YPD agar
230 plate¹, and cultured it in 5 mL of YNB+Nitrogen² + CSM-his³ supplemented with 2% glucose⁴
231 and 8 μg/mL histidine⁵ in a 50-mL Falcon tube at 30C and 50% humidity with shaking at 250
232 rpm for 24 hours. We then mixed the two strains at different fractions and diluted them x100 in
233 YNB+Nitrogen + CSM-his supplemented with 2% sucrose⁶, 0.001% glucose and 8 μg/mL
234 histidine. We then incubated them for 24 hours at 30C and 50% humidity with shaking at 250
235 rpm (5 mL of culture in 50 mL Falcon tubes). The first day of the experiment we determined the
236 fraction of each strain in each co-culture with flow cytometry, and mixed different co-cultures in
237 order to achieve the desired starting fraction of producers for the experiment. This procedure
238 ensured that the cells started in a physiological state characteristic of the co-culture of the two
239 strains.

240 Experiments were performed in flat-bottom 96-well plates, with 200 μL of medium per well
241 (YNB + CSM-his + 2% sucrose + 0.001% glucose + 8 μg/mL histidine). Plates were covered
242 with Parafilm (Bemis Flexible Packaging, Neenah, WI, USA) to limit evaporation, and incubated

¹ YPD agar plates: Teknova, Hollister, CA, USA; cat. no. Y1030

² YNB+Nitrogen: Sunrise Science, CA, USA; cat no. 1501-250

³ CSM: Sunrise Science, CA, USA; cat no. 1001-100

⁴ Glucose: Sigma-Aldrich/Millipore Sigma, St Louis, MO, USA; cat. no. G8270-1KG

⁵ Histidine: Sigma-Aldrich/Millipore Sigma, St Louis, MO, USA; cat. no. 53319-25G

⁶ Sucrose: Macron Fine Chemicals; cat. no. 8360-06

243 for 23 hours at 30C and 50% humidity with 800 rpm shaking. After every incubation period,
244 cells were diluted in fresh medium by the corresponding dilution factor, and migration was
245 performed following the scheme described in the main text. Dilution and migration were
246 performed at room temperature (~23C) and using two intermediate plates, to prevent pipeting of
247 small volumes and ensure accurate dilutions.

248

249 Measurements

250 Total density of every subpopulation was measured at the end of each growth cycle by
251 measuring the optical density at 600 nm in a plate reader (Varioskan Flash, Thermo Fisher
252 Scientific). Fraction of producers was also determined at the end of each growth cycle by flow
253 cytometry (Macs Quant VYB, Miltenyi Biotec, Bergisch Gladbach, Germany).

254

255 Model

256 We simulated the system using a model that reproduces the discrete cycles of the
257 experiment: The populations grow during each cycle governed by the equations shown in Fig.
258 2C (we used Matlab's function ode45 to solve the differential equations numerically). Then, all
259 populations are divided by the dilution rate, and migration is performed as detailed in Fig. 1B.
260 Then the next growth cycle is simulated.

261 To determine the model's parameters, we experimentally determined the parameters $r=0.5$
262 h^{-1} and $K=90000$ cells/ μ L. From previous works we know that $c<0.1$, $\varepsilon \approx k_M$ and that the lag
263 phase of yeast should be between 1 and 4 hours (44). Within these constraints, we manually
264 fitted the exact values of c , ε and the growth cycle duration to reproduce the trends shown in Fig.
265 2, as well as the collapse dilution rates shown in Fig. 3. We found a good agreement for $c=0.07$,
266 $\varepsilon = 14$ cells/ μ L, $k_M = 26$ cells/ μ L and a growth cycle of 22 hours (i.e. a lag phase of 1 hour).
267 A more detailed fit was unnecessary, given that this simple phenomenological model does not
268 capture the quantitative details of the system.

269 Note that ε is in units of the density of producer cells that should exist in the culture to
270 bring the concentration of public good to a level that matches the amount that each producer cell
271 keeps for itself.

272 To perform the resilience tests, we first let the metacommunity reach a stable state. If this
273 stable state presented oscillations (see **Fig. S2**), we chose a cycle in which population density
274 was minimum (given that we expect the metacommunity to be least resilient at this point). Then
275 we perturbed the metacommunity during a single cycle, either by imposing an additional dilution
276 factor ΔD (so for one cycle the dilution factor was $D'=D*\Delta D$), or by reducing the growth rate by
277 Δr (so for one cycle the growth rate was $r'=r-\Delta r$). After the perturbation cycle all parameters
278 went back to normal, and the simulation continued until the metapopulation either recovered or
279 went extinct.

280

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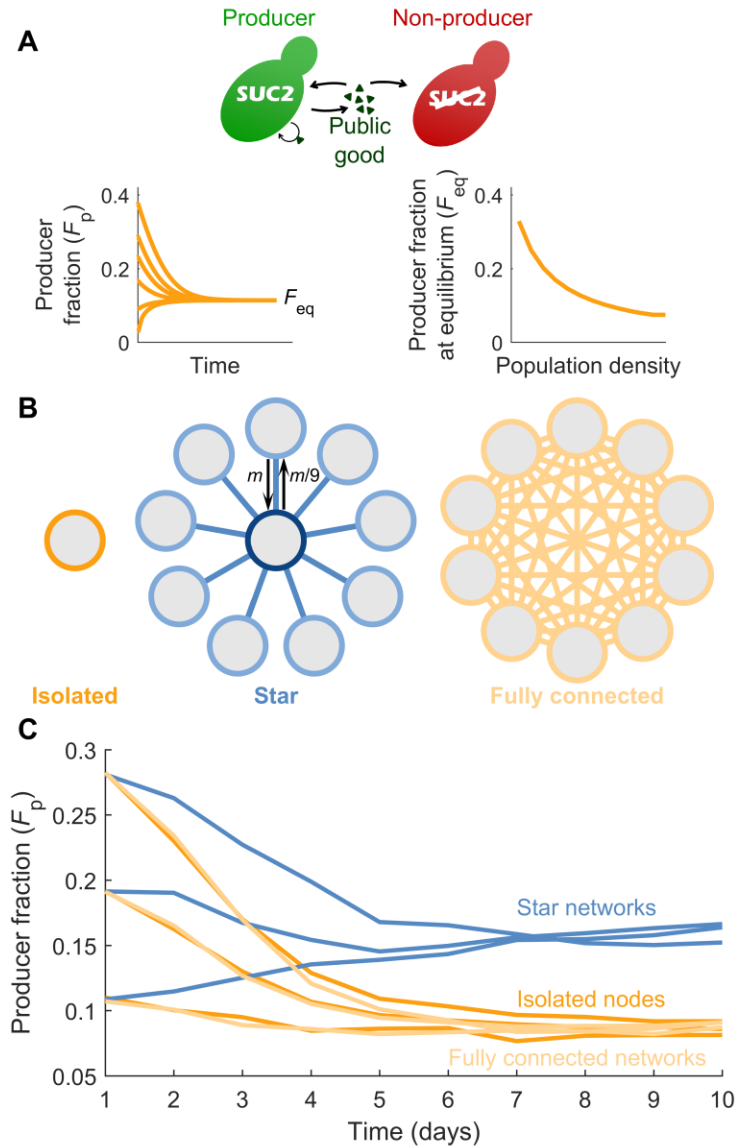
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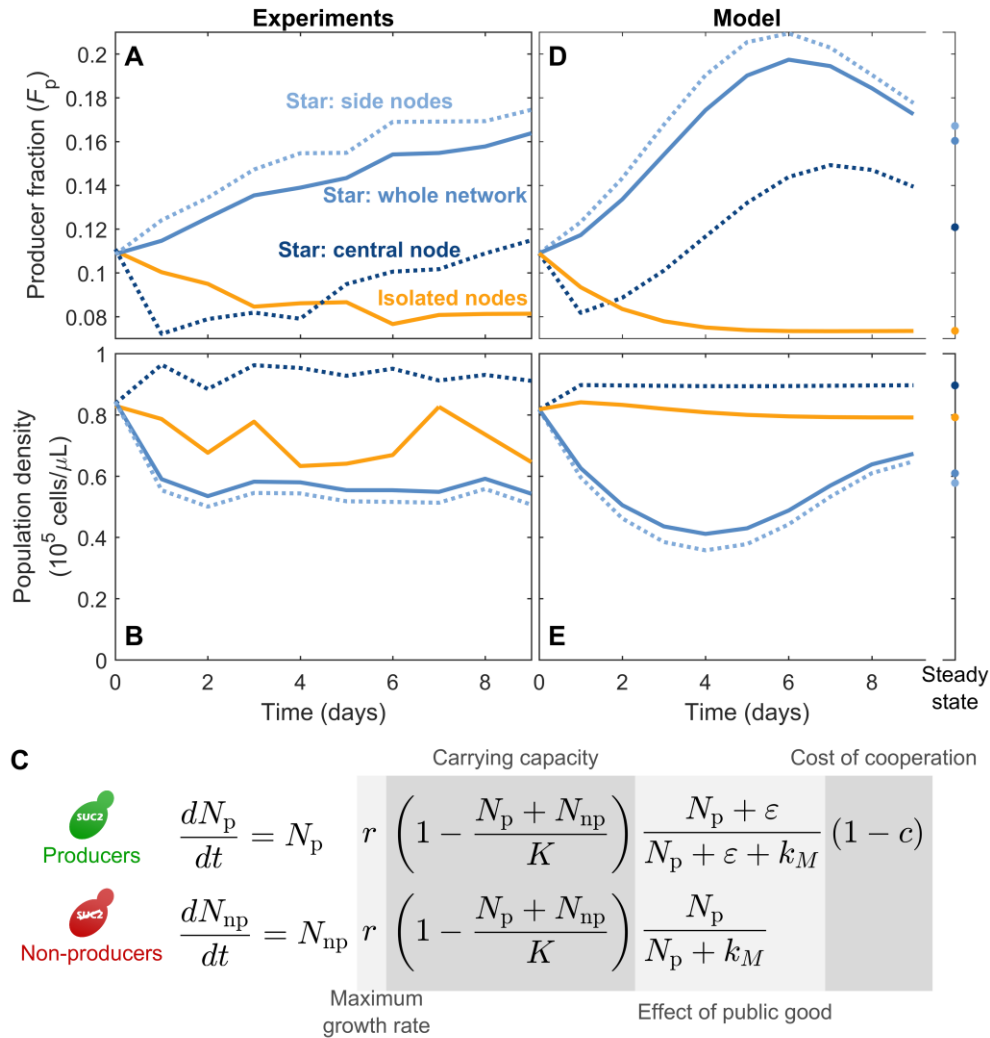
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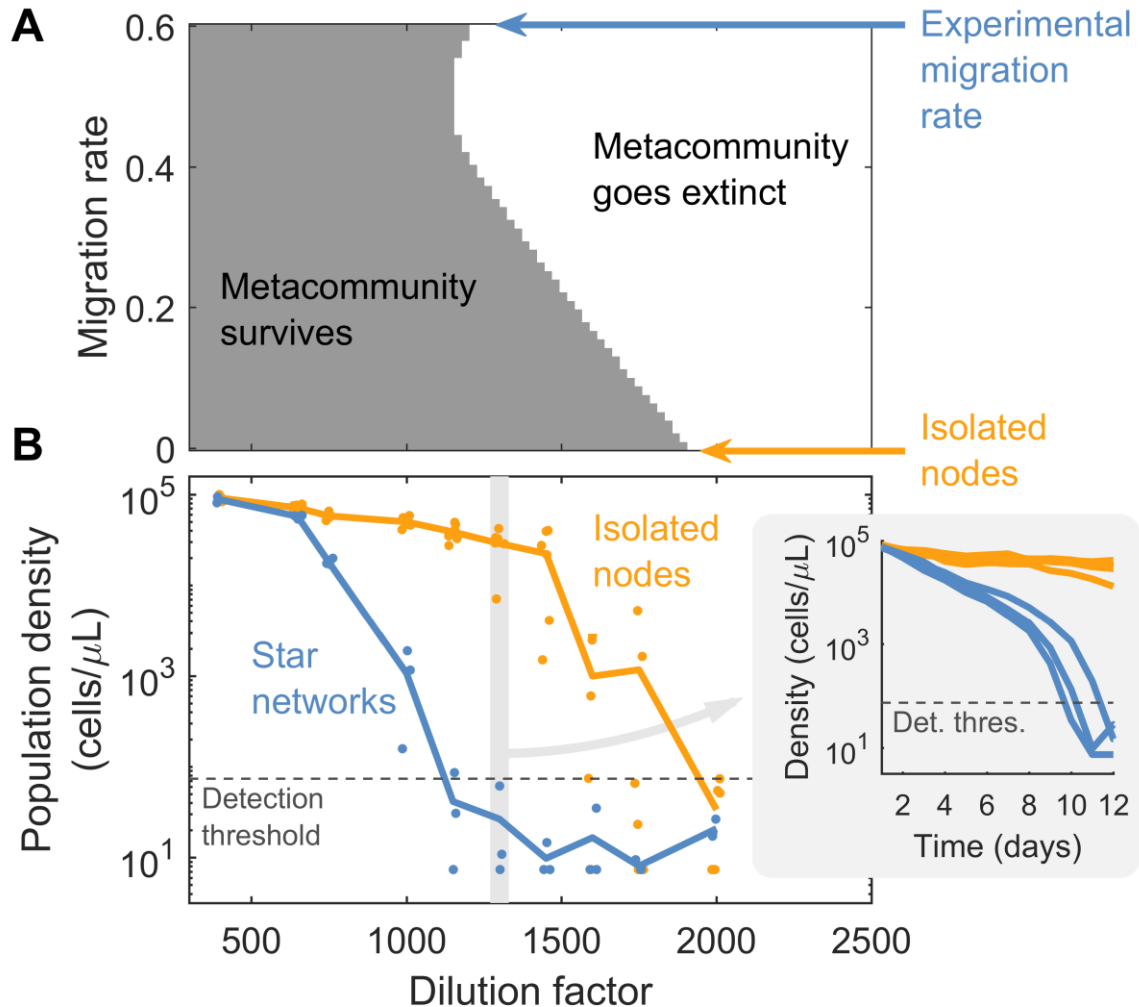
403 **Figure 1. Heterogeneous metacommunity structure increases the fraction of producers.** (A)
 404 Schematic of the public-goods game. Producers (green) produce a public good (triangles),
 405 keeping a small fraction (~1%) for themselves and releasing the rest, which also benefits non-
 406 non-producers (red). **Left inset:** Illustration of the time evolution of the fraction of producers (F_C) in
 407 a well-mixed population. **Right inset:** Illustration of the dependency of the equilibrium fraction
 408 of producers ($F_{C,eq}$) on the density of a well-mixed population. Both illustrations were computed
 409 using the model in Fig. 2. (B) Migration scheme for isolated nodes (no migration), star networks
 410 (all side nodes send a fraction m of migrants towards the central node, which in turn sends a
 411 fraction $m/9$ towards each side node), and fully connected networks (every node sends a fraction
 412 $m/9$ of migrants towards each other node). (C) Experimental results showing the fraction of
 413 producers (computed as total number of producers over total number of cells for the whole
 414 network) as a function of time for the three network topologies, starting from three initial
 415 conditions. Each line corresponds to one 10-node network (or 10 isolated nodes).
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418 **Figure 2. The increase in producer fraction in all nodes of a star network is captured by a**
 419 **simple phenomenological model. (A)** Experimental results showing the time evolution of the
 420 fraction of producers in the star network on average (solid blue line), each type of node within
 421 the star network (dashed dark blue line for central nodes and dashed light blue line for side
 422 nodes) and in an isolated population (yellow line). **(B)** Same as (a), but for the total density of
 423 cells. **(C)** Equations the phenomenological model. N_p , N_{np} are the densities of producers and non-
 424 producers, respectively (in cells/ μ L); r is the maximum growth rate; K is the carrying capacity
 425 (common for both strains); ε corresponds to the amount of enzyme imported by producers
 426 before the rest diffuses away (expressed in units of equivalent producer cells/ μ L, see Methods);
 427 k_M is the amount of producers needed to produce enough enzyme to bring the growth of non-
 428 producers to half its maximum value; c is the cost of producing the public good. Model
 429 parameters: $r=0.5 \text{ h}^{-1}$, $K=90000 \text{ cells}/\mu \text{ L}$, $c=0.07$, $\varepsilon = 14 \text{ cells}/\mu \text{ L}$, $k_M = 26 \text{ cells}/\mu \text{ L}$, growth
 430 cycle 22 hours, dilution factor 650 and migration rate $m=0.6$ per cycle (~ 0.06 per generation).
 431 **(D)** Same as (a), but as predicted by the model (points show the steady state after 100 cycles).
 432 **(E)** Same as (b), but as predicted by the model.

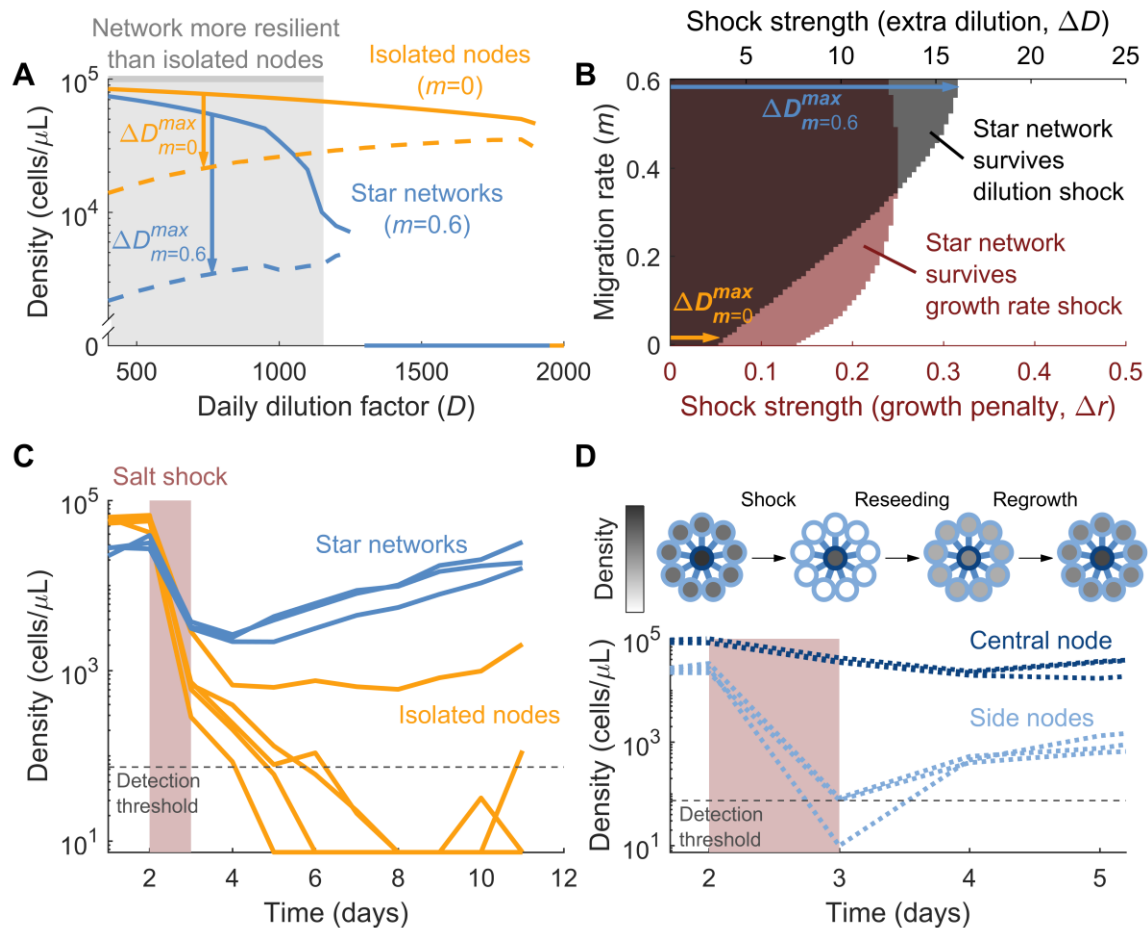
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435 **Figure 3. Heterogeneous metapopulations are less able to survive challenging environments**
436 **than isolated populations.** (A) The grey area indicates the conditions (dilution factor and
437 migration rate) in which the model predicts that a 10-node star network will survive after 1000
438 growth/dilution/migration cycles (note that $m = 0$ is also equivalent to isolated nodes). The rest
439 of model parameters are the same as in Fig. 2. (B) Experimental equilibrium population densities
440 for 10-nodes star networks (blue) and isolated nodes (yellow), as a function of dilution factor.
441 Each dot is either an individual network or a set of 10 isolated nodes; lines are the average.
442 **Inset:** Experimental time series of population densities for dilution factor 1300, for star networks
443 (blue) and isolated nodes (yellow). See **Supplementary Figure S1** for the complete time series.
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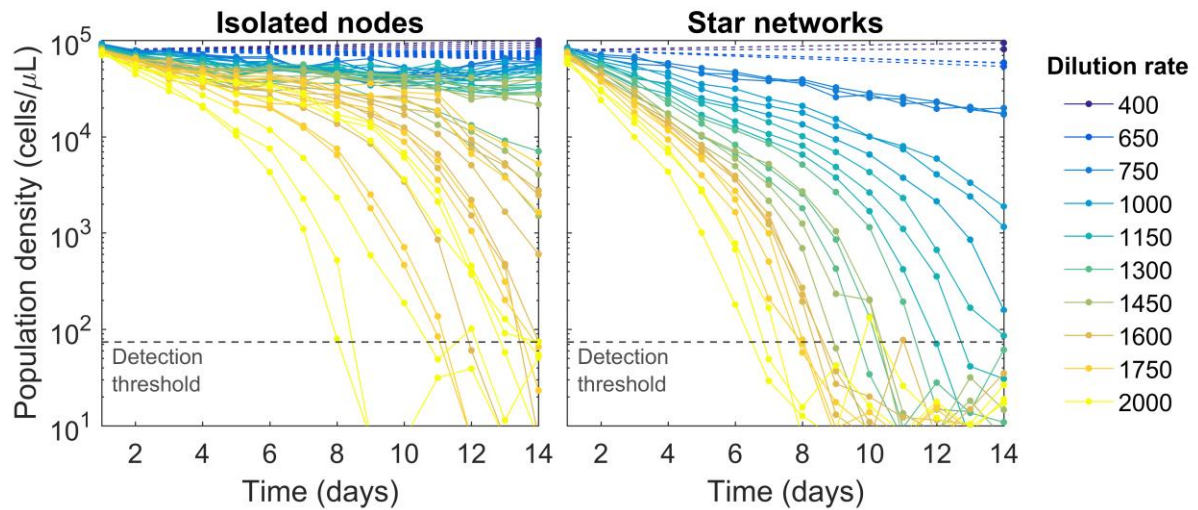
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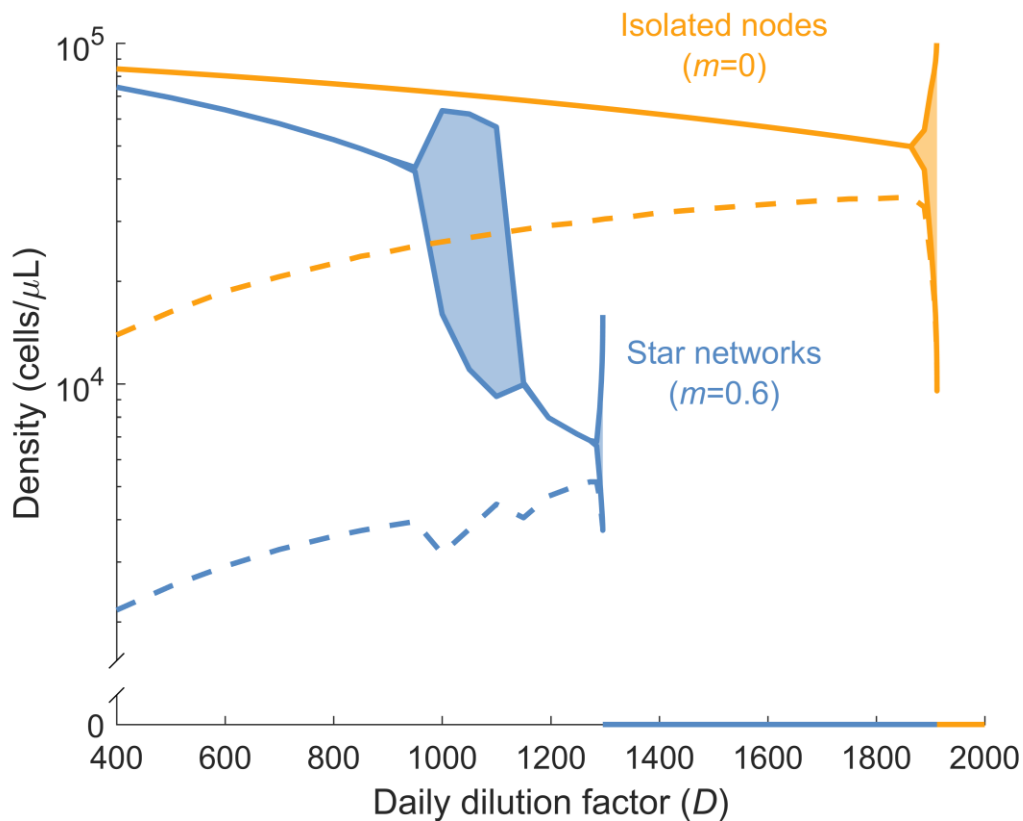
447 **Figure 4. Star networks are more resilient to perturbations than isolated populations. (A)**
 448 Bifurcation diagram for the metacommunity. Solid lines show the average population density in
 449 equilibrium as a function of dilution rate (yellow: isolated nodes; blue: 10-node star network).
 450 Dashed lines show the minimum density from which the metapopulation can recover. Arrows
 451 show the maximum dilution shock that each metapopulation can withstand (ΔD^{max}), when daily
 452 dilution rate is $D=750$. Grey area marks the region where the star network can withstand a
 453 greater shock than the isolated nodes. Model parameters are as in Fig. 2. See **Supplementary**
 454 **Figure S2** for a more detailed version of this figure. **(B)** Model prediction for survival of a 10-
 455 node star network with daily dilution factor $D=750$ after a perturbation, as a function of
 456 perturbation strength and migration rate. Rest of parameters are as in Fig. 2. **Black:** Shock
 457 corresponds to one cycle with increased dilution factor, $D'=750*\Delta D$. **Red:** Shock corresponds to
 458 one cycle with reduced growth $r'=0.5-\Delta r$. **(C)** Experimental results showing the time evolution
 459 of the average density in star networks (blue) and isolated populations (yellow). Each line
 460 corresponds to one 10-node network or to 10 isolated nodes. All populations are subject to a 750
 461 dilution factor and are initially in equilibrium. They are then perturbed by a salt shock (32g/L
 462 concentration of NaCl, shaded in red). **(D) Top:** Cartoon of star network recovering from a
 463 perturbation. Grayscale indicates population density in every node. **Bottom:** Same as (B), but
 464 only for the star network and separating the center and side nodes.

465 **Supplementary Figures:**
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468 **Figure S1. Time series show that star networks collapse at lower dilution rates than**
469 **isolated nodes. Left:** Population density over time for isolated nodes at different dilution rates.
470 Dashed lines indicate cases in which only initial and final densities were measured. Data at 14
471 days correspond to the points shown in main text **Figure 3B. Right:** Same as (Left), but for star
472 network (population density is averaged across each network).
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Figure S2. Bifurcation diagram for the metacommunity, accounting for oscillatory states.

Solid lines show the average population density in equilibrium as a function of dilution rate (yellow: isolated nodes; blue: 10-node star network). For some dilution factors the system reaches a limit cycle, presenting stable oscillations. These cases are represented by a shaded patch, the top line corresponding to the peak population density and the bottom line corresponding to the valley population density. Dashed lines show the minimum density from which the metapopulation can recover (assuming same producer frequencies and same relative densities between center and side nodes as the population in equilibrium; when oscillations are present, we took the population at the lowest point of the cycle as a reference). Model parameters are as in Fig. 2.