Introgression across evolutionary scales suggests reticulation contributes to Amazonian tree diversity

Running title: Reticulate evolution in Amazonian trees

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Abstract

Hybridization has the potential to generate or homogenize biodiversity and is a particularly common

phenomenon in plants, with an estimated 25% of species undergoing inter-specific gene flow.

However, hybridization in Amazonia's megadiverse tree flora was assumed to be extremely rare

despite extensive sympatry between closely related species, and its role in diversification remains

enigmatic because it has not yet been examined empirically. Using members of a dominant

Amazonian tree family (*Brownea*, Fabaceae) as a model to address this knowledge gap, our study

recovered extensive evidence of hybridization among multiple lineages across phylogenetic scales.

More specifically, our results uncovered several historical introgression events between *Brownea*

lineages and indicated that gene tree incongruence in *Brownea* is best explained by introgression,

rather than solely by incomplete lineage sorting. Furthermore, investigation of recent hybridization

using ~19,000 ddRAD loci recovered a high degree of shared variation between two *Brownea* species

which co-occur in the Ecuadorian Amazon. Our analyses also showed that these sympatric lineages

exhibit homogeneous rates of introgression among loci relative to the genome-wide average, implying

a lack of selection against hybrid genotypes and a persistence of hybridization over time. Our results

demonstrate that gene flow between multiple Amazonian tree species has occurred across temporal

scales, and contrasts with the prevailing view of hybridization's rarity in Amazonia. Overall, our

results provide novel evidence that reticulate evolution influenced diversification in part of the

Amazonian tree flora, which is the most diverse on Earth.

Keywords: Hybridization; Speciation; Angiosperms; Rainforest; Population genomics; Phylogenomics;

Introduction

Reproductive isolation is often seen as a prerequisite for speciation and as a defining feature of

species (Barraclough 2019; Mayr 1942). Despite this, hybridization between species is known to

occur and has several different outcomes, from the erosion of evolutionary divergence (Kearns *et al.,*

2018; Vonlanthen *et al.,* 2012) to the formation of entirely new 'hybrid species' (Mallet 2007).

Neotropical rainforests harbour the highest levels of plant diversity on Earth (Antonelli & Sanmartín

2011), and to date there has been little convincing evidence that hybridization consistently occurs

between tree species therein. Indeed, the prevailing view has been that hybridization between tropical

tree species is an exceptionally rare event (Ashton 1969; Ehrendorfer 1970; Gentry 1982). Although a

few observations of reproductive biology support this, such as the intersterility between lineages of

Inga, a species-rich genus in the legume family (Koptur 1984), evidence for hybridization has been

poorly tested empirically in tropical floras. The only potential example of hybridization among

neotropical trees which has been documented using DNA sequence data involves two species of

Carapa (Scotti-Saintagne *et al.,* 2013) found in Amazonia, the largest expanse of rainforest in the

world, containing at least 6,800 tree species (Cardoso *et al.,* 2017).

One factor that suggests hybridization might occur more frequently in neotropical rainforest tree

species, if their reproductive isolation is not absolute, is the remarkable level of sympatry found for

closely related species. In several neotropical lineages, including *Inga*, *Guatteria* and *Swartzia*, many

recently diverged species co-occur (Dexter *et al.,* 2017), often to a remarkable degree. One such

example of this is the co-occurrence of 19 *Inga* species in a single hectare of the Ecuadorian Amazon

(Valencia, Balslev, & Miño 1994). As such, for many rainforest taxa the opportunity for hybridization

is constantly present.

Hybridization can have a range of evolutionary consequences. In many cases, hybridization simply

results in the formation of sterile, maladapted offspring with poor reproductive fitness, allowing

genetic isolation between species to be maintained (i.e., reinforcement (Hopkins 2013)). In other

cases, there may be a permanent movement of genetic material from one lineage to another

(Rieseberg & Wendel 1993), which is known as 'introgression'. This transfer of genetic material

through hybridization may confer a selective advantage to resultant offspring (Taylor & Larson 2019),

in which case it is referred to as 'adaptive introgression'. Adaptive introgression is most often

observed between closely-related taxa during the invasion of new habitats (Suarez-Gonzalez, Lexer,

& Cronk 2018; e.g. Whitney, Randell, & Rieseberg 2010). Furthermore, hybridization can lead to

rapid evolutionary radiations. This occurs through the re-assembly of standing genetic variation which

has accumulated between diverging lineages, and which has already been subject to selection. This

'combinatorial' process is much more rapid than the gradual accumulation of variation through

mutation, and the passage of these variants through hybridization often fuels rapid diversification

events (Marques, Meier, & Seehausen 2019). This has been demonstrated in a wide range of taxa,

including sunflowers, African cichlid fish and Darwin's finches (Lamichhaney *et al.,* 2018; Meier *et*

al., 2017; Rieseberg *et al.,* 2003).

Introgression can occur at different rates in different regions of the genome (Payseur 2010). Regions

under divergent selection may remain distinct due to reduced fitness of hybrid genotypes at such loci,

resulting in a low rate of introgression. Conversely, regions under little or no selection tend to

introgress more freely, becoming homogenised between species. Moreover, if there is selection for

hybrid genotypes (as in adaptive introgression), the rate of introgression for a region may be further

increased relative to the rest of the genome (Gompert, Parchman, & Buerkle 2012). Such a process

has been demonstrated in temperate-zone tree species, where divergence between hybridizing lineages

is maintained through environmentally-driven selection (Hamilton & Aitken 2013; Sullivan *et al.,*

2016). This explains why species that hybridize can remain as biologically distinct (and

taxonomically identifiable) groups despite undergoing genetic exchange with other species (Abbott *et*

al., 2013; Seehausen *et al.,* 2014).

Brownea, a member of the legume family (Fabaceae), is a characteristic tree genus of lowland

neotropical rainforests and contains around 27 species distributed across northern South America.

Previous work indicates that there is a broad degree of phylogenetic incongruence evident in this

genus (Schley *et al.,* 2018) which might indicate hybridization, although this could also result from

incomplete lineage sorting. However, there are numerous *Brownea* hybrids in cultivation (e.g. *B. x*

crawfordii (Crawford & Nelson 1979) and *B. hybrida* (Backer 1911)), as well as several instances of

putative hybridization among *Brownea* lineages in the wild. The most notable of these instances is

that proposed between the range-restricted *Brownea jaramilloi* (Pérez *et al.,* 2013) and the wide-

ranging *B. grandiceps*, which co-occur in the Ecuadorian Amazon (Fig. 1). There are multiple

morphological distinctions between these two species, including differences in inflorescence colour

and structure, growth habit, tree height and leaf morphology (Pérez *et al.,* 2013). Although they co-

occur, these species favour different habitats: *B. jaramilloi* grows on ridge tops and hillsides, whereas *B. grandiceps* shows a slight preference for swamps and valleys but is more evenly distributed (pers. obs. & Klitgaard 1991; Pérez *et al.,* 2013). Despite this, hybridization appears to occur, as evidenced by the existence of a putative hybrid between these two species known as *B.* "rosada" (Fig. 1). *Brownea "*rosada*"* displays an intermediate morphology between its two parental species, producing pink flowers. The hypothesis of a *B. jaramilloi* x *B. grandiceps* hybrid has not yet been tested, however, using molecular data.

As a member of the legume family, which dominates Amazonian forests (ter Steege *et al.,* 2013), and with its apparent propensity for hybridization, *Brownea* is an excellent system with which to study the phylogenetic patterns and genomic architecture of introgression in neotropical rainforest trees. Systematically documenting hybridization at a range of time scales and taxonomic levels within this group could reveal how admixture has contributed to the assembly of one of the world's richest floras. Accordingly, this investigation aims to answer the following questions:

1. Is there evidence of introgression at a deep phylogenetic level in these lineages of neotropical rainforest tree?
2. Is there evidence of more recent gene flow, and if so, does this occur evenly across most of the genome?

Materials and Methods

Phylogenetic analysis

Sequences from 226 nuclear genes were used to elucidate the evolutionary relationships between *Brownea* species using a phylogenomic approach. To do so, 23 of 27 lineages were sampled within *Brownea*, including the three subspecies of *B. coccinea* and *B.* "rosada", a putative hybrid lineage of *B. grandiceps* and *B. jaramilloi*. In total, 59 accessions were sampled within *Brownea*, as well as an additional 13 outgroup taxa from the genera *Macrolobium*, *Heterostemon, Paloue* and *Browneopsis*,

which form part of the 'Brownea clade' (Fabaceae, subfamily Detarioideae (LPWG, 2017)). The list

of accessions and their associated information can be found in Supporting Information Table S1.

DNA sequence data were generated using leaf material collected from herbarium specimens and

silica-gel dried accessions. Genomic DNA was extracted using the CTAB method (Doyle & Doyle

1987), and sequencing libraries were prepared using the NEBNext® Ultra™ II DNA Kit (New

England Biolabs, Massachusetts, USA) with a modified protocol to account for fragmented DNA, and

including a ~600bp size-selection step. Hybrid bait capture was performed using the MyBaits protocol

(Arbor Biosciences, Michigan, USA) to target 289 phylogenetically-informative nuclear genes, using

a bait kit designed for the legume subfamily Detarioideae within which *Brownea* is nested (Ojeda *et*

al., 2019). The final library pools were sequenced either on the Illumina MiSeq platform (Illumina,

San-Diego, USA) at RBG Kew, or on the Illumina HiSeq platform by Macrogen Inc. (Seoul, South

Korea).

DNA sequencing reads were quality-checked with the program *FASTQC* v0.11.3 (Andrews 2010),

and were subsequently trimmed using *Trimmomatic* v.0.3.6 (Bolger, Lohse, & Usadel 2014) in order

to remove adapter sequences and to quality-filter reads. *Trimmomatic* settings permitted <4

mismatches, a palindrome clip threshold of 30 and a simple clip threshold of 6. Bases with a quality

score <28 and reads shorter than 36 bases long were removed from the dataset. Following quality-

filtering, loci were assembled using *SPAdes* v3.11.1 (Bankevich *et al.,* 2012) by the HybPiper pipeline

v1.2 (Johnson *et al.,* 2016), and potentially paralogous loci were removed using the Python (Python

Software Foundation 2010) script '*paralog_investigator.py*', distributed with the HybPiper pipeline.

All sequences were aligned by gene region using *MAFFT* v7.215 (Katoh & Standley 2013). In order

to infer gene trees for phylogenetic network analysis, the 226 recovered gene regions were further

refined to include only 20 taxa, representing a single accession per lineage in *Brownea*, using

Macrolobium colombianum as the outgroup taxon. Where applicable, samples were chosen by

comparing the sequence recovery of conspecific accessions and choosing the individual with the best

gene recovery. This resulted in 220 single-accession-per-lineage gene alignments. Further details of

phylogenomic sampling, labwork, sequencing and data processing are detailed in Supporting

Information Methods S1.

Gene trees were generated for each of the 226 gene alignments using *RAxML* v.8.0.26 (Stamatakis

2014) with 1,000 rapid bootstrap replicates and the GTRCAT model of nucleotide substitution, which

is a parameter-rich model and hence is most suitable for large datasets. *Macrolobium colombianum*

was used to root both the full and single-accession dataset analyses since it was the *Macrolobium*

accession that was present in the most gene alignments. The gene trees from both datasets were used

to generate species trees under the multi-species coalescent model in the heuristic version of *ASTRAL*

v.5.6.1 (Zhang *et al.,* 2018) using the default parameters. Monophyly was not enforced for individuals

belonging to the same species (the '-*a*' option in *ASTRAL*). Finally, discordance between gene trees

was calculated and visualised for the full dataset using the Java program *PhyParts* v0.0.1 (2015)

(https://bitbucket.org/blackrim/phyparts). The pattern of incongruence between gene trees for each

node was then mapped onto the *ASTRAL* species tree using the Python script *PhyPartsPieCharts* v1.0

(https://github.com/mossmatters/MJPythonNotebooks).

Inferring ancient introgression

Phylogenetic networks were inferred for 220 gene trees from the single-accession-per-lineage dataset

to understand whether introgression occurred over *Brownea*'s evolutionary history. Networks were

inferred with the program *SNaQ!,* implemented in the *Julia* v0.6.4 (Bezanson *et al.,* 2017) package

PhyloNetworks v0.11.0 (Solís-Lemus, Bastide, & Ané 2017). This program facilitates testing of

whether the observed incongruence between gene trees is better explained by a model describing only

ILS or by a model describing introgression. Networks were estimated by calculating quartet

concordance factors (CF) for each node from the single-accession-per-lineage gene trees, since *SNaQ!*

requires that each tip of the phylogenetic trees represent a single lineage. The network with the

number of hybridization events (*h*) best describing the data was chosen using negative log

pseudolikelihood comparison. Finally, the observed gene tree CFs were compared with those of the

best-fit phylogenetic network (i.e., the model including hybridization) and those expected under a

coalescent model (i.e., a 'tree-like' model which accounts for incomplete lineage sorting but not

hybridization). The fit of the observed gene tree topologies to either the 'network-like' or the 'tree-

like' model was assessed using the Tree Incongruence Checking in R (TICR) test (Stenz *et al.,* 2015).

This was done with the function '*test.one.species.tree'* in the R v3.4.4 (R Development Core Team

2013) package *PHYLOLM* (Ho & Ané 2014). Proportions of genes contributed between lineages by

hybridization events were taken from the 'Gamma value' output of *PhyloNetworks*. Further details of

phylogenetic network analysis are contained in Supporting Information Methods S1.

Population-level introgression

Having examined the degree of historical reticulation among *Brownea* lineages, population genomic

data were generated with ddRAD sequencing and used to investigate recent introgression at a finer

taxonomic scale. More specifically, the degree of shared genetic variation was visualised for

individuals of the ecologically divergent species *B. grandiceps* and *B. jaramilloi*, which co-occur in

the Ecuadorian Amazon and putatively hybridize in the wild. Following this, in order to make

inferences about the potential evolutionary significance of recent introgression, the rates at which

different loci introgress relative to the rest of the genome were estimated for these taxa using genomic

clines. In order to do this, 171 specimens in total were genotyped using ddRADseq (Peterson *et al.,*

2012). Sampling consisted of 128 individuals of *Brownea grandiceps*, 40 individuals of *B. jaramilloi*

and three individuals of *B.* "rosada" (the putative hybrid of *B. grandiceps* and *B. jaramilloi*),

representing the relative abundance of each species in the forest plot from which they were sampled.

Leaf material was collected from *Brownea* trees in the Yasuní National Park 50ha forest plot in Napo,

Ecuador, and dried in a herbarium press. The sample list is shown in Supporting Information Table

S2.

Genomic DNA was extracted from dried leaf material with the CTAB method and purified using a

QIAGEN plant minikit (QIAGEN, Hilden, Germany) column cleaning stage. Sequencing libraries

were prepared by digesting the DNA template using the restriction enzymes *EcoRI* and *mspI* (New

England BioLabs, Massachusetts, USA), in accordance with the ddRADseq protocol. Samples were

then ligated to universal Illumina P2 adapters and barcoded using 48 unique Illumina P1 adapters.

Samples were pooled and size-selected to between 375-550 bp using a Pippinprep electrophoresis

machine (Sage Science, Massachusetts, USA). Following amplification and multiplexing, sequencing

was performed using a single-lane, paired-end 150bp run on the HiSeq 3/4000 platform.

DNA sequencing reads from the ddRADseq genotyping were processed *de novo* (i.e., without the use

of a reference genome) using the Stacks pipeline v2.1 (Catchen *et al.,* 2011). Reads were quality

filtered using Stacks by removing reads which were of poor quality (i.e., had a phred score <10),

following which 'stacks' of reads were created, and SNPs were identified among all *de novo* 'loci'

and across individuals. This was done using a minimum coverage depth (the '*-m'* flag in Stacks) of

three and a within-individual mismatch distance (*-M*) of seven nucleotides. Individuals with

sequencing coverage under 7.5x were removed. Loci found in fewer than 40% of individuals and sites

with a minor allele frequency threshold of 5% were filtered out using the *'populations'* module of

Stacks to account for genotyping error. Model parameters for the various programs within the Stacks

pipeline were chosen using the recommendations in Paris *et al*. (2017), as well as through running the

pipeline with multiple different parameter settings. This resulted in a dataset containing 22,046 loci

with 120,085 SNPs for 171 individuals. A dataset containing only one SNP per locus was also

extracted using the Stacks *populations* module for use in analyses which assumed no linkage

disequilibrium. This subsetting resulted in a dataset containing 19,130 loci with 19,130 SNPs for 171

individuals. Details of population genomic sampling, library preparation, sequencing and data

filtering are shown in Supporting Information Methods S2.

In order to understand the patterns of introgression at the population level, the full ddRAD dataset was

used to visualise the degree of shared variation between *B. grandiceps* and *B. jaramilloi*. This was

performed using principal component analysis implemented in R v3.4.4, followed by plotting with

ggplot2 (Wickham 2016), and by producing a neighbour net plot in *SPLITSTREE* v4.14.6 (Huson &

Bryant 2005). Furthermore, the single-SNP-per-locus dataset containing 19,130 RAD loci was used

to infer population structure with the program *fastSTRUCTURE* v1.0 (Raj, Stephens, & Pritchard

2014). The number of populations (*K*) was chosen using the value which provided the largest

improvement in marginal likelihood. In addition, a *fastSTRUCTURE* run incorporating 40 random

individuals from each species was performed to account for any bias which may have been incurred

by differences in sample size. Finally, *NEWHYBRIDS* v1.1 (2002)

(https://github.com/eriqande/newhybrids) was used to categorise individuals into different hybrid

classes, using three runs on a subset of 500 randomly selected ddRAD loci due to the computational

limits of the program. Runs were performed with 50,000 MCMC sweeps following 50,000 burn-in

sweeps under the default parameters of the program.

The relative 'rate' and 'direction' of introgression for each locus between the two *Brownea* species

was estimated using Bayesian estimation of genomic clines (*bgc*) v1.03 (Gompert & Buerkle 2011).

For each locus, *bgc* compares the probability of ancestry at the locus relative to an individual's

genome wide ancestry, thereby allowing it to estimate two parameters for each locus. These

parameters are α, which roughly equates to the 'direction' of introgression, and β, which may be

summarised as the 'rate' of introgression for a locus (Gompert & Buerkle 2011; Gompert *et al.,*

2012). In order to estimate these parameters from the single-SNP dataset consisting of 19,130 RAD

loci, 50,000 MCMC generations with a 50% burn-in were used in *bgc*. Admixture proportions (i.e.,

mean *Q* values) generated by *fastSTRUCTURE* were used to assign each individual to three

populations (*Brownea grandiceps*, *B. jaramilloi* and admixed). Convergence was checked for the

MCMC output from *bgc* in Tracer v1.6 (Rambaut *et al.,* 2015) and with the R package *coda*

(Plummer *et al.,* 2006) using Geweke's diagnostic (Geweke 1991). Loci with significant 'excess

ancestry' were identified by ascertaining whether the 99% posterior probability estimates of the α and

β parameters included zero (i.e., by identifying positive or negative non-zero estimates of the

parameters). In addition, loci which were extreme 'introgression outliers' were identified for both

parameters by identifying loci whose median estimates were not included in the 99% posterior

probability credible intervals (Gompert & Buerkle 2011). Further details of visualising shared

variation, hybrid category assignment and *bgc* analysis are shown in Supporting Information Methods

S2.

<u>Results</u>

<u>Phylogenetic analysis</u>

Using DNA sequence data from 226 nuclear gene regions, we produced a 'species' tree using

ASTRAL (Zhang *et al.,* 2018) (Fig. 2) based on gene trees inferred with the program *RAxML*

(Stamatakis 2014). This resulted in well-supported relationships between major subclades (>0.9 local

posterior probability (PP)), with lower support for inter-specific relationships. *ASTRAL* inferred a high

degree of discordance among gene tree topologies at many nodes (quartet score = 0.52), with multiple

alternative bipartitions reconstructed at most nodes. This is evident from the presence of many more

conflicting gene trees (numbers below branches) than congruent gene trees (numbers above branches)

in Supporting Information Fig. S1.

<u>Ancient introgression in Brownea species</u>

Phylogenetic networks estimated to ascertain whether diversification in *Brownea* was tree-like or

reticulate pinpointed two hybridization events within the genus (Fig. 3). This is indicated by the -log

pseudolikelihood values in Supporting Information Fig. S2, since the number of hybridization events

(*h*) which best describe the data give the largest improvement in -log pseudolikelihood. In Fig. S2, -

log pseudolikelihoods increased steadily between $h = 0$ and $h = 2$, after which the increasing number

of hybridization events only made minimal improvements to -log pseudolikelihood.

The inferred phylogenetic network (Fig. 3) showed a broadly similar topology to the species tree in

Fig. 2, with the addition of two hybridization events. The first hybridization event occurred between

the lineage leading to the Venezuelan accessions of *B. grandiceps/B. leucantha* and *B. birschelli*,

suggesting that the lineage leading to *B. birschelli* has in the past contributed around 34% of the

genetic material present in the common ancestor of the Venezuelan *B. grandiceps* and *B. leucantha*.

The second inferred occurrence of hybrid ancestry occurs between the ancestors of the subclade

containing the Colombian accessions of *B. coccinea/B. santanderensis* and the lineage leading to *B.*

enricii, which contributed around 37% of the genetic material belonging to the ancestor of the

aforementioned subclade.

Tree Incongruence Checking in R (TICR) analysis indicated that a network-like model (i.e., one

including introgression) best described the patterns of incongruence in the single-accession-per-

species gene trees inferred during this study. This method suggested an excess of outlier quartets ($P=$

1.240×10^{-19}, $X^2 = 91.149$), which differed significantly from the CF values expected under a

coalescent model describing only incomplete lineage sorting. As such, a tree-like model was able to

be rejected as an explanation for the observed relationships between taxa in *Brownea*, suggesting that

hybridization occurred over the course of its evolutionary history and was responsible for the

observed incongruence.

Introgression dynamics across loci

Our population genomic analyses revealed a broad degree of shared variation between *B. grandiceps*

and *B. jaramilloi*, along with a homogeneous distribution of introgression rates among loci. In total,

ddRAD sequencing resulted in 640,898,950 reads between 350-550bp in length for 171 individuals.

Among these reads, 28,503,796 (4.45%) were discarded due to poor recovery. There was a mean of

3,747,947 reads per individual, with an average coverage depth across all samples of 27.5x. Relatively

high levels of shared genetic variation were observed among taxa, along with low levels of genetic

differentiation and only marginal differences in the amount of genetic variation, as shown by the

population genetic statistics calculated by the Stacks pipeline for the two *Brownea* species (including

B. "rosada", which is grouped with *B. grandiceps*) (Supporting Information Table S3). The F_{st}

calculated between *B. grandiceps/B.* "rosada" and *B. jaramilloi* was 0.111, representing a low degree

of fixation, and so a high amount of shared variation.

Patterns of shared genetic variation visualised with principal component analysis (PCA) and

SPLITSTREE showed a distinct signature of admixture between *B. grandiceps* and *B. jaramilloi*. The

PCA of SNP variation inferred using the R package *ggplot2* (Fig. 4a) indicated that the first principal

component (PC1) explained the largest proportion of the genetic variation among all the principal

components (20.4%). Individuals of *B. grandiceps* are tightly clustered along PC1, where the

individuals of *B. jaramilloi* show much more variability, with many individuals forming an

intergradation between the two main species clusters. Additionally, the *B.* "rosada" accessions appear

to have clustered more closely to *B. grandiceps* than to *B. jaramilloi* along PC1. PC2, which

explained 10.7% of the variation in the SNP data, shows a similar degree of variability in both *B.*

grandiceps and *B. jaramilloi*, with two accessions of *B.* "rosada" shown to cluster in between both

species. This pattern is reflected by the implicit network built using *SPLITSTREE* (Supporting

Information Fig. S3). *SPLITSTREE* recovered a clustering of individuals into two main groups,

largely representing *B. grandiceps* and *B. jaramilloi,* with four putative hybrid individuals displaying

an intermediate relationship between the two species clusters.

The *fastSTRUCTURE* analysis (Raj *et al.,* 2014) performed to further examine the degree of shared

genetic variation indicated that there was a large amount of shared ancestry between the genotyped

individuals (Fig. 4b), with evidence of extensive backcrossing due to the widely differing proportions

of ancestry in different individuals. Marginal likelihood comparison indicated that the best value of *K*

(i.e., the number of populations) was two ($\Delta_{marginal\ likelihood} = 0.085$, Supporting Information Fig. S4A),

which was the value that resulted in the largest increase in marginal likelihood. Since the 'best' value

of *K* is an estimate, f*astSTRUCTURE* plots generated with other *K* values are shown in Supporting

Information Fig. S4B. Figure 4b shows that most individuals of *B. jaramilloi* have varying fractions

of *B. grandiceps* ancestry in their genome. In addition, the individuals identified as *B.* "rosada" appear

to have inherited most of their ancestry from *B. grandiceps*, with only a minimal contribution from *B.*

jaramilloi. The same pattern was recovered from a *fastSTRUCTURE* run incorporating 40 random

individuals from each species, performed to account for any bias which may have been incurred by

differences in sample size (Supporting Information Fig. S4C).

The *NEWHYBRIDS* analysis revealed that most hybrid individuals were the result of continued

hybridization, with pure *B. grandiceps* making up 73.9% of the genotyped population, and pure *B.*

jaramilloi making up 21.9% for the 500 subsampled loci used in this analysis. There were no F1 (first

generation) hybrids identified by the subset of loci analysed, and all hybrid individuals were either F2

hybrids (0.592%) or had a broad distribution of probabilities across hybrid classes (3.55%),

suggesting backcrossing. In the latter, the probability of any one hybrid class did not exceed 90%,

which was the threshold used to categorize individuals as belonging to a certain class.

The Bayesian estimation of genomic clines (*bgc*) analysis, which was used to determine whether any

loci showed outlying rates of admixture relative to the genome-wide average, recovered a signal of

asymmetric introgression, but did not detect any differential rates of introgression between loci. Of

the 19,130 loci under study, *bgc* recovered 251 loci (1.3%) with mostly *B. jaramilloi* alleles among all

individuals across species (i.e. with positive α estimates) and 1089 loci (5.69%) with mostly *B.*

grandiceps alleles among all individuals across species (i.e., with negative α estimates). However, no

loci displayed extreme rates of introgression relative to the average rate across the genome (i.e., there

were no statistically outlying β parameter estimates). The MCMC runs from which these results were

drawn showed adequate mixing (Supporting Information Fig. S5A-C).

<u>Discussion</u>

Our study represents the first clear case of reticulated evolution among Amazonian trees documented

at multiple phylogenetic levels. This contrasts with previous work, which suggested that hybridization

is an extremely rare phenomenon in Amazonian trees. We demonstrate that within *Brownea*, a

characteristic Amazonian tree genus, reticulated evolution has occurred over the course of its

evolutionary history, with evidence of introgression deep in the history of the genus and more

recently, between the closely related *B. grandiceps* and *B. jaramilloi*.

Introgression has occurred at deep phylogenetic levels in **Brownea**

Phylogenetic network analysis suggested that introgression has taken place between *Brownea* lineages in the past, with two separate hybridization events inferred (Fig. 3). The concordance factors obtained from gene trees were not adequately explained by a tree-like model (i.e., one accounting only for incomplete lineage sorting), suggesting that a network-like model (i.e., one including introgression) best describes the diversification patterns within *Brownea* (P= 1.240 x 10^{-19}, X^2 = 91.149). Hybridization mainly occurs between individuals which are geographically overlapping, or have been so at some point in the past (Burbrink & Gehara 2018). Our findings are thus corroborated by the fact that both historical introgression events were between taxa that co-occur (e.g., *B. grandiceps, B. leucantha* and *B. birschelli* on the Northern coast of Venezuela, and *B. coccinea, B. santanderensis* and *B. enricii* within the Colombian cordilleras). This is also in agreement with previous work (Schley *et al.,* 2018), which indicated that the 'stem' lineages within *Brownea* shared a broad distribution across northern South America, which may have further facilitated earlier hybridization. Our study also recovered a low degree of support for bifurcating inter-specific relationships (Fig. 2) and a high degree of incongruence among gene trees (Supporting Information Fig. S1). These results can be partially explained by introgression, rather than being caused purely by the stochasticity of lineage sorting in large populations, which is suggested to be a common phenomenon in many rainforest trees with large population sizes (Pennington & Lavin 2016). It seems likely that amongst closely-related species of *Brownea* divergence has not progressed to the point of complete reproductive isolation, as has been shown in temperate *Quercus* species (e.g. Lepais & Gerber 2011). Moreover, the low quartet scores and minimal gene-tree concordance found at the species level in this study mirror those observed in *Lachemilla*, a montane neotropical plant genus, in which gene tree discordance was shown to be explained by both historical and recent hybridization (Morales-Briones, Liston, & Tank 2018).

Since the inferred events of ancient introgression both involved the common ancestors of two sets of present-day species, they are likely to have occurred several million years in the past, before the

divergence of the descendent species. The inferred introgression events are likely to have occurred

since the Miocene period (~23Ma), as date estimates from previous work suggest that *Brownea*

originated around this time (Schley *et al.,* 2018). Accordingly, it is unlikely that the inferred

hybridization events are due to 'accidental' hybridization between co-occurring species, merely

producing maladapted F1 hybrids which will eventually be outcompeted, fortifying the boundaries

between species (i.e. 'reinforcement' (Hopkins 2013)). Rather, our results suggest a persistence of

introgression over evolutionary time within *Brownea*.

Recent hybridization also occurs between* Brownea *species, resulting in persistent hybrids and

introgression across the genome

This investigation also uncovered a substantial signal of recent introgression between *B. grandiceps*

and *B. jaramilloi* in the Yasuní National Park 50ha plot located in the Ecuadorian Amazon. The low

F_{st} estimate for the *B. grandiceps/B.* "rosada" and *B. jaramilloi* populations (0.11), in addition to the

principal component analysis (Fig. 4a), the *fastSTRUCTURE* analysis (Fig. 4b) and the *SPLITSTREE*

plot (Supporting Information Fig. S3) indicated a high degree of shared variation between these

lineages. All these approaches show that individuals cluster into two main groups, largely

representing *B. grandiceps* and *B. jaramilloi*, as well as a set of individuals forming an intergradation

between the two main clusters. The individuals which form a part of this intergradation were mostly

identified as *B. jaramilloi*, although hybrid individuals are also observed in the *B. grandiceps* cluster.

The higher number of variant sites and the higher nucleotide diversity (π) recovered for *B. jaramilloi*

(Supporting Information Table S3) could also reflect the hybrid ancestry of many individuals

identified as this species. A similar introgression-driven increase in nucleotide diversity has been

shown in closely-related species of *Mimulus* which undergo asymmetric introgression (Sweigart &

Willis 2003). In addition, *B. jaramilloi* and *B. grandiceps* showed evidence of multi-generational

backcrossing in both the *fastSTRUCTURE* and *NEWHYBRIDS* analyses, further suggesting that

hybridization does not always result in reinforcement and that hybrids persist over time.

The *bgc* analysis indicated that most introgression was asymmetric, with more loci containing mostly

B. grandiceps alleles (5.96% of all loci) than loci containing mostly *B. jaramilloi* alleles (1.3% of all

loci). Importantly, this *bgc* analysis also indicated that gene flow occurs largely at the same rate

across loci, since there were no outlying estimates of the β parameter. The excess of *B. grandiceps*

ancestry mirrors the asymmetry of introgression suggested by Fig. 4b and is likely driven by the

uneven population sizes of the two species. *Brownea jaramilloi* has only ever been observed in a

small part of the western Amazon (Pérez *et al.,* 2013) and is much less populous than *B. grandiceps*,

which occurs across northern South America. More specifically, the disproportionate donation of

alleles from *B. grandiceps* may be due to 'pollen swamping', whereby pollen transfer from another,

more populous species is more likely than pollen transfer from less numerous conspecifics (Buggs &

Pannell 2006). Pollen swamping can serve as a mechanism of invasion (e.g., in *Quercus* (Petit *et al.,*

2004)), and the associated asymmetric introgression may lead to the extinction of the rarer species,

especially when hybrids are inviable or sterile (Balao *et al.,* 2015). However, due to the ongoing

backcrossing and introgression evident between *B. grandiceps* and *B. jaramilloi* it appears that

hybrids are not always sterile, or at least that 'foreign' alleles are not always subject to strong

purifying selection. The best evidence of this is the absence of loci with extreme values of the β

parameter recovered by our *bgc* analysis. Extreme deviations in β are mainly expected in the presence

of gene flow when selection against hybrids is strong, resulting in underdominance and a paucity of

heterozygous sites (Gompert *et al.,* 2012). As such, our inferred lack of outlying β values suggests

persistence of hybrid genotypes.

Further to this, the observed asymmetry in introgression could be caused by selection favouring

hybrid genotypes. The transfer of genomic regions which confer a selective advantage from a donor to

a recipient species (i.e. adaptive introgression) can result in a disproportionate amount of one species'

genome being present in hybrid individuals (Yang *et al.,* 2018). This occurs when viable hybrids only

tend to backcross with one of the parental species, thereby introducing the other species' genetic

material into the first in an asymmetrical fashion. Selection has been suggested as a driver for

asymmetric introgression in multiple plant genera, including *Helianthus* (Scascitelli *et al.,* 2010) and

Iris (Arnold *et al.,* 2010). Accordingly, it is possible that selectively advantageous alleles are passing

between species (e.g., between *B. jaramilloi* and *B. grandiceps*) through backcrossing events over

many generations as observed by the *fastSTRUCTURE* and *NEWHYBRIDS* analyses in this study

(Mallet 2005). This may facilitate adaptation to new niches due to the widening of the pool of

variation upon which selection can act (e.g. Whitney *et al.,* 2010). However, it is difficult to ascertain

whether adaptive introgression has occurred without measuring the impact of introgression on

variation in the phenotype and its fitness effects (Suarez-Gonzalez *et al.,* 2018).

The contribution of hybridization to neotropical rainforest tree diversity

Studies such as ours that document persistent hybridization across evolutionary time between tree

species within Amazonia, and within tropical rainforests in general, are rare. Indeed, it was previously

suggested that inter-specific hybrids between rainforest tree species are poor competitors, and that

fertile hybrid populations are nearly non-existent (Ashton 1969). While there is some evidence of

introgression in tropical trees (Scotti-Saintagne *et al.,* 2013), most available studies substantiating this

are based on trees from other tropical regions or habitats (e.g., *Shorea* in Asia (Kamiya *et al.,* 2011;

Kenzo *et al.,* 2019), or *Rhizophora* in Indo-Pacific mangroves (Lo 2010)). Many of these other

instances appear to occur only in degraded habitats, or involve infertile first-generation hybrids with

minimal backcrossing, which contrasts with the findings of our study. Within *Brownea* we uncovered

introgression across taxonomic, spatial and temporal scales, with evidence of backcrossing and the

persistence of hybrid genotypes.

It is possible that a lack of selection against hybrids allows the passage of variation between *Brownea*

species, which may persist over evolutionary time and could explain why we found evidence of

introgression at both macroevolutionary and microevolutionary scales. This is congruent with a

'syngameon' model, where large, disparate populations of individuals belonging to multiple

interfertile species occasionally hybridize and exchange genetic variants. This can aid in the

maintenance of genetic diversity in species which exist in very low population densities, as is typical

of tropical rainforest trees, thereby preventing Allee effects. This has been shown in temperate tree species (Cronk & Suarez-Gonzalez 2018), although it typically occurs at the edge of species ranges and its prevalence in hyperdiverse systems such as tropical rainforests is a topic of recent debate (e.g. Cannon & Lerdau 2019). Furthermore, while the passage of adaptive variation between syngameons of closely related species can seed rapid, speciose radiations in other taxa (Marques *et al.,* 2019; Meier *et al.,* 2017), whether this occurs between tree species in neotropical rainforest remains a key knowledge gap.

While it is difficult to determine whether hybridization between tree species occurs across many syngameons of closely related Amazonian lineages, or whether it is a tendency unique to certain genera such as *Brownea*, it may be prudent to review how relationships among tropical tree lineages are inferred. Accordingly, the use of phylogenetic networks in resolving the relationships between such groups may provide additional insight into whether reticulate evolution has contributed to diversification within Amazonian rainforest, which is among the most species-rich environments on Earth (Burbrink & Gehara 2018; Solís-Lemus *et al.,* 2017).

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- **Mobilization permit**: 037-2016-MOV-FLO-MAE-DPAO;

- **Export permit**: 208-2019-EXP-CM-FAU-DNB/MA; and

- **Genetic research permit (Contrato Marco)**: MAE-DNB-CM-2018-0082

References

Abbott, R. J., Albach, D., Ansell, S., Arntzen, J. W., Baird, S. J., Bierne, N., ... Buggs, R. (2013)
Hybridization and speciation. *Journal of Evolutionary Biology,* **26**, 229-246.

Anderson, E. C., Thompson, E. A. (2002) A model-based method for identifying species hybrids
using multilocus genetic data. *Genetics,* **160**, 1217-1229.

Andrews, S. (2010) FastQC: a quality control tool for high throughput sequence data. Available
online at http://www.bioinformatics.babraham.ac.uk/projects/fastqc.

Antonelli, A., Sanmartín, I. (2011) Why are there so many plant species in the Neotropics? *Taxon,* **60**,
403-414.

Arnold, M. L., Tang, S., Knapp, S. J., & Martin, N. H. (2010) Asymmetric introgressive hybridization
among Louisiana Iris species. *Genes,* **1**, 9-22.

Ashton, P. S. (1969) Speciation among tropical forest trees: some deductions in the light of recent
evidence. *Biological Journal of the Linnean Society,* **1**, 155-196.

Backer, C. A. (1911) *Brownea hybrida* Hort. ex Backer. In: *Schoolflora voor Java*pp. 418 Visser &
Co., Weltevreden, Java, Dutch East Indies.

Balao, F., Casimiro-Soriguer, R., García-Castaño, J. L., Terrab, A., & Talavera, S. (2015) Big thistle
eats the little thistle: does unidirectional introgressive hybridization endanger the conservation of
Onopordum hinojense? *New Phytologist,* **206**, 448-458.

Bankevich, A., Nurk, S., Antipov, D., Gurevich, A. A., Dvorkin, M., Kulikov, A. S., ... Prjibelski, A.
D. (2012) SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing.
Journal of Computational Biology, **19**, 455-477.

Barraclough T. G. (2019) *The Evolutionary Biology of Species* Oxford University Press, Oxford.

Bezanson, J., Edelman, A., Karpinski, S., & Shah, V. B. (2017) Julia: A fresh approach to numerical
computing. *SIAM Review, 59*, 65-98.

Bolger, A. M., Lohse, M., & Usadel, B. (2014) Trimmomatic: a flexible trimmer for Illumina
sequence data. *Bioinformatics, 30*, 2114-2120.

Buggs, R. J., Pannell, J. R. (2006) Rapid displacement of a monoecious plant lineage is due to pollen
swamping by a dioecious relative. *Current Biology, 16*, 996-1000.

Burbrink, F. T., Gehara, M. (2018) The biogeography of deep time phylogenetic reticulation.
Systematic Biology, 67, 743-744.

Cannon, C. H., Lerdau, M. T. (2019) Demography and destiny: The syngameon in hyperdiverse
systems. *Proceedings of the National Academy of Sciences of the United States of America, 116*,
8105.

Cardoso, D., Sarkinen, T., Alexander, S., Amorim, A. M., Bittrich, V., Celis, M., ... Forzza, R. C.
(2017) Amazon plant diversity revealed by a taxonomically verified species list. *Proceedings of the
National Academy of Sciences of the United States of America, 114*, 10695-10700.

Catchen, J. M., Amores, A., Hohenlohe, P., Cresko, W., & Postlethwait, J. H. (2011) Stacks: building
and genotyping Loci de novo from short-read sequences. *G3 (Bethesda, Md.), 1*, 171-182.

Crawford, T., Nelson, E. (1979) Irish Horticulturists. I: WH Crawford. *Garden History, 7*, 23-26.

Cronk, Q. C., Suarez-Gonzalez, A. (2018) The role of interspecific hybridization in adaptive potential
at range margins. *Molecular ecology, 27*, 4653-4656.

Dexter, K. G., Lavin, M., Torke, B. M., Twyford, A. D., Kursar, T. A., Coley, P. D., ... Pennington, R.
T. (2017) Dispersal assembly of rain forest tree communities across the Amazon basin. *Proceedings
of the National Academy of Sciences of the United States of America, 114*, 2645-2650.

Doyle, J. J., Doyle, J. L. (1987) Genomic plant DNA preparation from fresh tissue- CTAB method.
Phytochemical Bulletin, 19, 11-15.

Ehrendorfer, F. (1970) Evolutionary patterns and strategies in seed plants. *Taxon, 19*, 185-195.

Gentry, A. H. (1982) Neotropical floristic diversity: phytogeographical connections between Central
and South America, Pleistocene climatic fluctuations, or an accident of the Andean orogeny? *Annals
of the Missouri Botanical Garden, 69*, 557-593.

Geweke J. (1991) *Evaluating the accuracy of sampling-based approaches to the calculation of
posterior moments* Federal Reserve Bank of Minneapolis, Research Department, Minneapolis, MN,
USA.

Gompert, Z., Buerkle, C. A. (2011) Bayesian estimation of genomic clines. *Molecular Ecology, 20*,
2111-2127.

Gompert, Z., Parchman, T. L., & Buerkle, C. A. (2012) Genomics of isolation in hybrids.
Philosophical Transactions of the Royal Society of London Series B: Biological sciences, 367, 439-
450.

Hamilton, J. A., Aitken, S. N. (2013) Genetic and morphological structure of a spruce hybrid (*Picea*
sitchensis × *P. glauca*) zone along a climatic gradient. *American Journal of Botany,* **100**, 1651-1662.

Ho, L. S. T., Ané, C. (2014) A linear-time algorithm for Gaussian and non-Gaussian trait evolution
models. *Systematic Biology,* **63**, 397-408.

Hopkins, R. (2013) Reinforcement in plants. *New Phytologist,* **197**, 1095-1103.

Huson, D. H., Bryant, D. (2005) Application of phylogenetic networks in evolutionary studies.
Molecular Biology and Evolution, **23**, 254-267.

Johnson, M. G., Gardner, E. M., Liu, Y., Medina, R., Goffinet, B., Shaw, A. J., ... Wickett, N. J.
(2016) HybPiper: Extracting coding sequence and introns for phylogenetics from high-throughput
sequencing reads using target enrichment. *Applications in Plant Sciences,* **4**, 1600016.

Kamiya, K., Gan, Y. Y., Lum, S. K., Khoo, M. S., Chua, S. C., & Faizu, N. N. (2011) Morphological
and molecular evidence of natural hybridization in *Shorea* (Dipterocarpaceae). *Tree Genetics &*
Genomes, **7**, 297-306.

Katoh, K., Standley, D. M. (2013) MAFFT multiple sequence alignment software version 7:
improvements in performance and usability. *Molecular Biology and Evolution,* **30**, 772-780.

Kearns, A. M., Restani, M., Szabo, I., Schrøder-Nielsen, A., Kim, J. A., Richardson, H. M., ...
Omland, K. E. (2018) Genomic evidence of speciation reversal in ravens. *Nature Communications,* **9**,
906.

Kenzo, T., Kamiya, K., Ngo, K. M., Faizu, N., Lum, S. K. Y., Igarashi, S., ... Ichie, T. (2019)
Overlapping flowering periods among *Shorea* species and high growth performance of hybrid
seedlings promote hybridization and introgression in a tropical rainforest of Singapore. *Forest*
Ecology and Management, **435**, 38-44.

Klitgaard, B. B. (1991) Ecuadorian *Brownea* and *Browneopsis* (Leguminosae-Caesalpinioideae):
taxonomy, palynology, and morphology. *Nordic Journal of Botany,* **11**, 433-449.

Koptur, S. (1984) Outcrossing and pollinator limitation of fruit set: breeding systems of neotropical
Inga trees (Fabaceae: Mimosoideae). *Evolution,* **38**, 1130-1143.

Lamichhaney, S., Han, F., Webster, M. T., Andersson, L., Grant, B. R., & Grant, P. R. (2018) Rapid
hybrid speciation in Darwin's finches. *Science,* **359**, 224-228.

Legume Phylogeny Working Group (2017) A new subfamily classification of the Leguminosae based
on a taxonomically comprehensive phylogeny: The Legume Phylogeny Working Group (LPWG).
Taxon, **66**, 44-77.

Lepais, O., Gerber, S. (2011) Reproductive patterns shape introgression dynamics and species
succession within the European white oak species complex. *Evolution,* **65**, 156-170.

Lo, E. (2010) Testing hybridization hypotheses and evaluating the evolutionary potential of hybrids in
mangrove plant species. *Journal of Evolutionary Biology,* **23**, 2249-2261.

Mallet, J. (2007) Hybrid speciation. *Nature,* **446**, 279-283.

Mallet, J. (2005) Hybridization as an invasion of the genome. *Trends in Ecology & Evolution,* **20**,
229-237.

Marques, D. A., Meier, J. I., & Seehausen, O. (2019) A combinatorial view on speciation and
adaptive radiation. *Trends in Ecology & Evolution,* **34**, 531-544.

Mayr E. (1942) *Systematics and the Origin of Species* Columbia University Press, New York, New
York, USA.

Meier, J. I., Marques, D. A., Mwaiko, S., Wagner, C. E., Excoffier, L., & Seehausen, O. (2017)
Ancient hybridization fuels rapid cichlid fish adaptive radiations. *Nature Communications,* **8**, 14363.

Morales-Briones, D. F., Liston, A., & Tank, D. C. (2018) Phylogenomic analyses reveal a deep
history of hybridization and polyploidy in the Neotropical genus Lachemilla (Rosaceae). *New*
Phytologist, **218**, 1668-1684.

Ojeda, D., Koenen, E., Cervantes Arango, S., de la Estrella, M., Banguera-Hinestroza, E., Janssens,
S., ... Hardy, O. (2019) Phylogenomics within the Anthonotha clade (Detarioideae, Leguminosae)
reveals a high diversity in floral trait shifts and a general trend towards organ number reduction.
bioRxiv, , 511949.

Paris, J. R., Stevens, J. R., & Catchen, J. M. (2017) Lost in parameter space: a road map for stacks.
Methods in Ecology and Evolution, **8**, 1360-1373.

Payseur, B. A. (2010) Using differential introgression in hybrid zones to identify genomic regions
involved in speciation. *Molecular Ecology Resources,* **10**, 806-820.

Pennington, R. T., Lavin, M. (2016) The contrasting nature of woody plant species in different
neotropical forest biomes reflects differences in ecological stability. *New Phytologist,* **210**, 25-37.

Pérez, Á J., Klitgård, B. B., Saslis-Lagoudakis, C., & Valencia, R. (2013) *Brownea jaramilloi*
(Leguminosae: Caesalpinioideae), a new, over-looked species endemic to the Ecuadorian Amazon .
Kew Bulletin, **68**, 157-162.

Peterson, B. K., Weber, J. N., Kay, E. H., Fisher, H. S., & Hoekstra, H. E. (2012) Double digest
RADseq: an inexpensive method for de novo SNP discovery and genotyping in model and non-model
species. *PloS one,* **7**, e37135.

Petit, R. J., Bodénès, C., Ducousso, A., Roussel, G., & Kremer, A. (2004) Hybridization as a
mechanism of invasion in oaks. *New Phytologist,* **161**, 151-164.

Plummer, M., Best, N., Cowles, K., & Vines, K. (2006) CODA: convergence diagnosis and output
analysis for MCMC. *R news,* **6**, 7-11.

Python Software Foundation (2010) Python Language Reference, version 2.7. *http://www.python.org,*
.

R Development Core Team (2013) R: A language and environment for statistical computing.
Available at: http://www.R-project.org/.

Raj, A., Stephens, M., & Pritchard, J. K. (2014) fastSTRUCTURE: variational inference of population
structure in large SNP data sets. *Genetics,* **197**, 573-589.

Rambaut, A., Suchard, M. A., Xie, D., & Drummond, A. J. (2015) Tracer.
http://tree.bio.ed.ac.uk/software/tracer/, **1.6**.

Rieseberg, L. H., Wendel, J. F. (1993) Introgression and its consequences in plants. *Hybrid Zones and*
the Evolutionary Process, **70**, 70-109.

Rieseberg, L. H., Raymond, O., Rosenthal, D. M., Lai, Z., Livingstone, K., Nakazato, T., ... Lexer, C.
(2003) Major ecological transitions in wild sunflowers facilitated by hybridization. *Science (New*
York, N.Y.), **301**, 1211-1216.

Scascitelli, M., Whitney, K., Randell, R., King, M., Buerkle, C., & Rieseberg, L. (2010) Genome scan
of hybridizing sunflowers from Texas (*Helianthus annuus* and *H. debilis*) reveals asymmetric patterns
of introgression and small islands of genomic differentiation. *Molecular Ecology,* **19**, 521-541.

Schley, R. J., de la Estrella, M., Pérez-Escobar, O. A., Bruneau, A., Barraclough, T., Forest, F., &
Klitgård, B. (2018) Is Amazonia a 'museum'for Neotropical trees? The evolution of the Brownea
clade (Detarioideae, Leguminosae). *Molecular Phylogenetics and Evolution,* **126**, 279-292.

Scotti-Saintagne, C., Dick, C. W., Caron, H., Vendramin, G. G., Guichoux, E., Buonamici, A., ...
Lemes, M. R. (2013) Phylogeography of a species complex of lowland Neotropical rain forest trees
(Carapa, Meliaceae). *Journal of Biogeography,* **40**, 676-692.

Seehausen, O., Butlin, R. K., Keller, I., Wagner, C. E., Boughman, J. W., Hohenlohe, P. A., ...
Brännström, Å (2014) Genomics and the origin of species. *Nature Reviews Genetics,* **15**, 176-192.

Smith, S. A., Moore, M. J., Brown, J. W., & Yang, Y. (2015) Analysis of phylogenomic datasets
reveals conflict, concordance, and gene duplications with examples from animals and plants. *BMC*
Evolutionary Biology, **15**, Published online: DOI 10.1186/s12862-015-0423-0.

Solís-Lemus, C., Bastide, P., & Ané, C. (2017) PhyloNetworks: a package for phylogenetic networks.
Molecular Biology and Evolution, **34**, 3292-3298.

Stamatakis, A. (2014) RAxML version 8: a tool for phylogenetic analysis and post-analysis of large
phylogenies. *Bioinformatics (Oxford, England),* **30**, 1312-1313.

Stenz, N. W., Larget, B., Baum, D. A., & Ané, C. (2015) Exploring tree-like and non-tree-like
patterns using genome sequences: an example using the inbreeding plant species *Arabidopsis thaliana*
(L.) Heynh. *Systematic Biology,* **64**, 809-823.

Suarez-Gonzalez, A., Lexer, C., & Cronk, Q. C. B. (2018) Adaptive introgression: a plant perspective.
Biology Letters, **14**, 10.1098/rsbl.2017.0688.

Sullivan, A. R., Owusu, S. A., Weber, J. A., Hipp, A. L., & Gailing, O. (2016) Hybridization and
divergence in multi-species oak (*Quercus*) communities. *Botanical Journal of the Linnean Society,*
181, 99-114.

Sweigart, A. L., Willis, J. H. (2003) Patterns of nucleotide diversity in two species of *Mimulus* are
affected by mating system and asymmetric introgression. *Evolution,* **57**, 2490-2506.

Taylor, S. A., Larson, E. L. (2019) Insights from genomes into the evolutionary importance and
prevalence of hybridization in nature. *Nature Ecology & Evolution,* **3**, 170-177.

ter Steege, H., Pitman, N. C., Sabatier, D., Baraloto, C., Salomao, R. P., Guevara, J. E., ... Silman, M.
R. (2013) Hyperdominance in the Amazonian tree flora. *Science,* **342**, 1243092.

Valencia, R., Balslev, H., & Miño, G. P. Y. (1994) High tree alpha-diversity in Amazonian Ecuador.
Biodiversity & Conservation, **3**, 21-28.

Vonlanthen, P., Bittner, D., Hudson, A. G., Young, K. A., Müller, R., Lundsgaard-Hansen, B., ...
Seehausen, O. (2012) Eutrophication causes speciation reversal in whitefish adaptive radiations.
Nature, **482**, 357-362.

Whitney, K. D., Randell, R. A., & Rieseberg, L. H. (2010) Adaptive introgression of abiotic tolerance
traits in the sunflower *Helianthus annuus. New Phytologist,* **187**, 230-239.

Wickham H. (2016) *ggplot2: elegant graphics for data analysis* Springer.

Yang, W., While, G. M., Laakkonen, H., Sacchi, R., Zuffi, M. A., Scali, S., ... Uller, T. (2018)
Genomic evidence for asymmetric introgression by sexual selection in the common wall lizard.
Molecular Ecology, **27**, 4213-4224.

Zhang, C., Rabiee, M., Sayyari, E., & Mirarab, S. (2018) ASTRAL-III: polynomial time species tree
reconstruction from partially resolved gene trees. *BMC Bioinformatics,* **19**, 153.

Data Accessibility

The data that support the findings of this study are openly available from online repositories. All raw

reads generated with the targeted bait capture and ddRADseq methods are available on the NCBI

Sequence Read Archive with the accession numbers SAMN13439069- SAMN13439140 and

SAMN13441804- SAMN13441974, respectively, under the Bioproject number PRJNA592723. All

full phylogenomic sequence alignments, single-accession-per-species alignments and tree files, bgc

input files, Stacks output files and the Detarioideae bait kit sequence file are found on Dryad

(https://doi.org/10.5061/dryad.k3j9kd53w). Data are under embargo until publication, and any further

data required are available from the corresponding author upon reasonable request.

Author Contributions

R.J.S. conceived this study and performed all analyses, supervised by T.B., F.F. and B.K. Population

genomic data were generated by R.J.S., and phylogenomic data were generated by R.J.S., I.K. and I.L.

Baits for hybrid capture were provided by M.d.l.E. R.J.S. wrote the first draft of the manuscript with

contributions from R.T.P., O.A.P.E., A.J.H., M.d.l.E., I.L., T.B., F.F. and B.K.

<u>Figures</u>

Figure 1: Two co-occurring *Brownea* lineages (*Brownea grandiceps* (photograph © Rowan Schley) and
Brownea jaramilloi (photograph © Xavier Cornejo)) as well as their putative hybrid *Brownea* "rosada"
(photograph © J. L. Clark).

Figure 2: *ASTRAL* species tree inferred from *RAxML* gene trees using the multi-species coalescent model.
Numbers at nodes of the tree signify local posterior probability (LPP) values, a measure of support for each
quadripartition. Taxa within the red box belong to the 'Grandiceps' subclade, taxa within the green box belong
to the 'Coccinea' subclade, taxa within the pink box represent the 'Stenantha' subclade and those within the
blue box belong to the 'Chocóan' subclade. Images show inflorescences of species within *Brownea*- species
shown are, from top to bottom: *B. grandiceps* (photograph © Rowan Schley), *B. jaramilloi* (photograph ©
Xavier Cornejo), *B. macrophylla* (photograph © Bente Klitgaard), *B. coccinea* subsp. *capitella* (photograph ©
Xavier Cornejo), *B. longipedicellata* (photograph © Domingos Cardoso) and *B. multijuga* (photograph © Bente
Klitgaard).

Figure 3: Phylogenetic network with two hybridization events ($h = 2$), estimated using *SNaQ!* in the Julia
package *PhyloNetworks*. Light blue horizontal branches indicate inferred hybridization events, and numbers
next to the branches show the estimated proportion of genes contributed by each lineage in the hybridization
event. Red branches signify the ancestral lineage and what proportion of the modern lineage's genes were
contributed from it. Taxa were chosen in order to represent one accession per lineage, as per the assumptions of
PhyloNetworks, and the individual with the highest gene recovery was used for each lineage. Coloured boxes
containing taxa are described in the legend of Fig. 2.

Figure 4:
a) Principal component analysis of genotype data for all SNPs inferred at all loci. Individuals are coded by colour and point shape: red circles denote individuals identified as *B. grandiceps*, yellow circles denote individuals identified as *B. jaramilloi,* and blue triangles denote putative hybrid individuals (*B.* "rosada"). The amount of the genetic variation explained by each principle component is shown next to axes, which are labelled PC1 and PC2.

b) *fastSTRUCTURE* plot, indicating ancestry proportions for two populations (*K* = 2). The 'populations' correspond to two different species: *B. grandiceps* (shown in red) and *B. jaramilloi* (shown in yellow). Each individual accession is represented by a column, and the proportion of ancestry from either species (mean *Q*) is proportional to the length of different coloured bars in each column. Individuals identified as *B.* "rosada" are

748   marked with a blue box and a blue triangle beneath their column. All taxa to the left and right of these boxes
749   were identified as *B. grandiceps* and *B. jaramilloi*, respectively.