Microtubule Stutter: a Transient Dynamic Instability Phase that is Strongly Associated with Catastrophe

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ABSTRACT

Microtubules (MT) are cytoskeletal fibers that undergo dynamic instability, a remarkable process involving phases of growth and shortening separated by approximately random transitions (catastrophe and rescue). Dissecting dynamic instability mechanism(s) requires first characterizing and quantifying these dynamics, a subjective process that often ignores complexity in MT behavior. Using a novel machine-learning based tool (STADIA), we show that microtubule behavior consists not only of growth and shortening, but also a transient intermediate phase we term 'stutter.' Quantifying stutter and other behaviors with STADIA demonstrates that stutters precede most catastrophes in simulations and experiments. Moreover, we show that the anti-catastrophe factor CLASP2γ works by promoting stuttering MTs to return to growth. STADIA enables more comprehensive and objective analysis of MT dynamics compared to previous methods. The identification of stutters as a distinct and quantifiable phase of dynamic instability provides new opportunities for analyzing mechanisms of microtubule dynamics and their regulation by binding proteins.

INTRODUCTION

Microtubules (MTs) are protein-based biological polymers that have a central role in fundamental eukaryotic processes including cellular organization, chromosome separation during cell division, and intracellular transport (Goodson and Jonasson 2018). Crucial to the function of MTs in these processes is a well-known behavior termed dynamic instability (DI), where the polymers switch stochastically between periods of growth and shortening as can be seen in traditional MT length-history plots (**Figure 1 A,B**) (Mitchison and Kirschner 1984; Desai and Mitchison 1997). Accurate quantification of MT DI behavior is essential for understanding its significance and mechanism and also for investigating the activities of DI-regulating proteins and pharmaceutical agents (e.g., chemotherapy drugs, fungicides).

Problems with current methods of quantifying dynamic instability
Traditionally, MTs have been treated as two-state polymers; that is, MTs have been considered to be either growing or shortening, with abrupt, instantaneous transitions called catastrophes and rescues between these two phases (**Figure 1 A,B,D**). In this framework, MT behavior is characterized by four quantities called DI parameters (Walker et al. 1988):

- V_{growth} - velocity of growth, commonly measured as the mean of the growth rates over the set of growth phases
- V_{short} - velocity of shortening, commonly measured as the mean of the shortening rates over the set of shortening phases
- F_{cat} - frequency of catastrophe, commonly measured as the number of catastrophes (transitions from growth to shortening) per time in growth
- F_{res} - frequency of rescue, commonly measured as the number of rescues (transitions from shortening to growth) per time in shortening

In this approach to quantifying MT dynamics, the velocity of an individual growth or shortening phase is typically determined as the slope of a line drawn between the points of catastrophe and rescue (see e.g. (Zanic 2016)).

While determination of DI parameters as described above is now a standard way to quantify MT behavior (Portran et al. 2017; Kapoor et al. 2019), there are aspects of MT behavior that are not captured using this approach. First, it has long been recognized that both growth and shortening rates are variable. This variability occurs both with and without MT binding proteins (MTBPs), and it is observed both within and between individual growth phases and similarly for shortening phases (see e.g., (Gildersleeve et al. 1992; Pedigo and Williams 2002; Schek et al. 2007; Lawrence et al. 2018)). In addition, Odde et al.'s analysis of the variability in growth and shortening rates suggested that the two-state (growth and shortening) model well approximated experimentally observed MT behavior at time scales greater than ~ 1 minute, but underestimated the observed variability at time scales less than ~ 1 minute (David J. Odde, Buettner, and Cassimeris 1996). These observations raise the concern that current DI analysis methods that categorize an entire period between nucleation (or rescue) and catastrophe as a single growth or shortening phase could cause finer, but functionally significant, aspects of MT behavior to be missed.

Second, pauses, attenuation phases, and other intermediate states have been observed in experiments and proposed in models, but the way these behaviors have been identified and defined has varied. Pauses are commonly observed *in vivo* (e.g., (Sammak and Borisy 1988; Schulze and Kirschner 1988; Shelden and Wadsworth 1993; Dhamodharan and Wadsworth 1995; Waterman-Storer and Salmon 1997; Gierke, Kumar, and Wittmann 2010; Applegate et al. 2011)). Pauses have also been observed *in vitro* in the presence of MTBPs (e.g., (Moriwaki and Goshima 2016)), cell extracts (e.g. (Keller et al. 2008)), drugs (e.g., (Toso et al. 1993)), and occasionally for purified tubulin (e.g., (Walker et al. 1988)). Identification of pause phases in length-history data frequently relies on arbitrary fixed thresholds. A length-change threshold of 0.5 microns has been common, but the exact way in which this threshold was applied to data has varied among research groups (Sammak and Borisy 1988; Dhamodharan and Wadsworth 1995; Dhamodharan et al. 1995; Rusan et al. 2001; Kamath, Oroudjev, and Jordan 2010; Fees, Estrem, and Moore 2017). Others have used combinations of thresholds on the speed of length change (e.g., in pixels per frame, or microns per minute), length change itself, and/or number of data points involved (Panda et al. 1996; Yenjerla, Lopus, and Wilson 2010; Matov et al. 2010; Gierke, Kumar, and Wittmann 2010; Kiris, Ventimiglia, and Feinstein 2010; Mahrooghy et al. 2015; Moriwaki and Goshima 2016). Recognition of states other than growth and shortening has also led various authors to consider theoretical three- or four-state models in which the additional states are pauses or an intermediate state (D. J. Odde, Cassimeris, and Buettner 1995; David J. Odde, Buettner, and Cassimeris 1996; Tran, Walker, and Salmon 1997; Jánosi, Chréien, and Flyvbjerg 2002; Maly 2002; Keller et al. 2008; Smal et al. 2010; Blackwell et al. 2017). Thus, it is clear that many researchers are interested in methods for quantifying states beyond growth and shortening in data and the inclusion of such states in the development of theory, but it is less clear how these states should be defined.

Third, recent improvements in imaging technology have enabled acquisition of MT growth data with both high temporal and high spatial resolution which allows for the possibility of analyzing length-history data at finer scales. These data have verified the intrinsic variability of MT behavior. They have also demonstrated that both growth and shortening phases can include significant time periods (e.g., a few seconds in duration or longer) during which MTs do not change appreciably in length (**Figure 1 C,E;** see also (Maurer et al. 2014; Duellberg, Cade, and Surrey 2016; Duellberg et al. 2016; Rickman et al. 2017)). These slow-down periods likely overlap with pauses discussed above. Significantly, these slow-down periods can also occur in association with catastrophe, making it difficult to determine with reasonable precision where transitions between phases begin and end. To illustrate this problem, consider the zoomed-out length-history plots that are typically used for DI analysis (**Figure 1 B,D**). Examination of these plots can make the task of determining when transitions occur look trivial. However, the zoomed-in views made possible by high-resolution data acquisition (**Figure 1 C,E**) demonstrate the difficulty of identifying the point of transition and/or categorizing this behavior.

Thus, many researchers have recognized that MT dynamic instability behavior is more complex than a simple two-state system of growth and shortening with abrupt transitions. The four traditional DI parameters (V_{growth}, V_{short}, F_{cat}, and F_{res}) would be sufficient to quantify such a two-state system, but are not sufficient to quantify all aspects of observed MT dynamic instability. How to analyze such dynamics

has not been obvious. One approach has been to exclude slow-down periods from quantification of DI parameters, since including them in either growth or shortening phases would reduce the magnitude of measured values of V_{growth} and V_{short} (e.g., (Rickman et al. 2017)). However, there are two problems with this approach. First, it leaves researchers to make subjective judgments or use 'in-house' software to identify both the periods to exclude and the points where phase transitions occur (e.g., (Yenjerla, Lopus, and Wilson 2010; Zanic 2016; Jonasson et al. 2019)). This approach impacts precision and reproducibility. Second, and perhaps more significantly, entirely excluding these behaviors from analysis could potentially result in the loss of information critical for understanding the mechanisms of the phase transitions or their regulation by MT binding proteins. Thus, capturing and quantifying these alternative behaviors is a key step towards explaining the recognized variations in growth and shortening rates, improving the precision of these metrics, and elucidating mechanisms of dynamic instability.

Summary of results

Using established statistical methods, we developed the <u>S</u>tatistical <u>T</u>ool for <u>A</u>utomated <u>D</u>ynamic <u>I</u>nstability <u>A</u>nalysis (STADIA), an automated tool for comprehensively characterizing and quantifying MT behavior. Applying STADIA to *in silico* and *in vitro* MT length-history data demonstrated the prevalence of an intermediate phase that we propose calling 'stutter'. Stutter is a transient DI phase where MTs exhibit rapid low-amplitude fluctuations but with an overall rate of change in MT length that is markedly less in magnitude compared to classically recognized growth and shortening phases. Stutters, as recognized and quantified by STADIA, overlap with previously observed behaviors such as pre-catastrophe slow-downs (e.g., (Maurer et al. 2014)) and short pauses (e.g., (Guo et al. 2018)), but differ from pauses during which no dynamics are occurring (e.g., (Gierke, Kumar, and Wittmann 2010)). More specifically, stutters differ from non-dynamic pauses in that during stutters subunit-scale dynamics continue. Significantly, we observed that most catastrophes *in silico* and *in vitro* are preceded by stutters, and that the MT stabilizing protein CLASP2γ reduces catastrophe by increasing the fraction of stutters that return to growth rather than enter shortening phases. These results indicate that classical methods of analyzing MT length-history data may miss mechanistically significant aspects of MT behavior and that our automated DI analysis tool, STADIA, is able to recognize and quantify these behaviors. The relationship of our results to previous work is further discussed at the end of the Results & Discussion section. We conclude that identification of stutters as a phase distinct from growth and shortening warrants their future inclusion in DI analyses, and serves as a necessary step forward in gaining a better understanding of MTs, their dynamics, and their regulation by MT binding proteins.

<u>RESULTS & DISCUSSION</u>

We first present a brief overview of STADIA and its analysis procedure (readers are encouraged to refer to the Methods for more detailed information; see also (Patel et al. 2020)). We then use STADIA to analyze MT dynamics as they are observed in simulations (*in silico*) and in experiments (*in vitro*), including detection and quantification of a transient, intermediate phase that we term 'stutter'. We use this observation as a foundation on which to study the relationship between stutter and the phase transitions, showing that stutter is strongly associated with catastrophe. We further test the functional significance of this observation and demonstrate the utility of STADIA in studying MT-binding proteins

by using STADIA to analyze the dynamics of MTs growing in the presence of the anti-catastrophe factor CLASP2γ, thus examining for the first time its effect on stutter.

STADIA: A Novel Tool for Analyzing Dynamic Instability Behavior of MTs

To meet the goal of more precisely identifying, categorizing, and quantifying the range of MT behaviors, we created the <u>S</u>tatistical <u>T</u>ool for <u>A</u>utomated <u>D</u>ynamic <u>I</u>nstability <u>A</u>nalysis (STADIA). Specific aims for the development of STADIA were that it be: **1)** Automated to create a consistent and reproducible method with minimal user input; **2)** Impartial such that it does not presuppose MT dynamics are restricted to two states (i.e., being limited to growth and shortening); **3)** Adaptive to handle varying time durations of phases and the stochastic nature of phase changes; **4)** Compatible with classical DI analysis, enabling comparison to and continuity with previous work; **5)** Capable of analyzing MT time-series data sourced from both computational simulations and laboratory experiments.

The resulting software, STADIA, is a data-driven tool that uses time-series analysis to characterize and quantify MT behavior. STADIA can be run in two modes: Diagnostic Mode (outlined in the Methods, useful for performing preliminary analyses and tuning analysis parameters), and Automated Mode (outlined below; used for performing full DI analysis). The process, presently implemented in MATLAB, has three major stages (**SuppMat Figure S1**):

I. <u>Segmentation</u>: STADIA creates a continuous piecewise linear approximation of MT length-history data, where segment endpoints mark moments of significant change, i.e., transitions between periods of sustained behavior. These transitions include extreme events such as traditionally recognized catastrophes and rescues as well as more subtle changes in the rate of MT length change (**Figure 2**). The level of accuracy of the approximation is dictated by user-defined parameters.

II. <u>Classification</u>: STADIA then classifies the individual segments from the linear approximation in Stage I using an unsupervised clustering method, *k*-means, and bundles segment clusters with similar characteristics into DI phases (**Figure 3, SuppMat Figures S2,S3,S4,S5**). The number of DI phases identified is informed by running STADIA in the Diagnostic Mode.

III. <u>Phase and Transition Analysis</u>: STADIA then applies the segment classifications from Stages I and II to length-history plots and considers chronological ordering to perform quantitative phase and transition analysis (this includes measuring the frequencies of catastrophe, rescue, and other possible transitions) (**Figure 3 G,H, Table 1, SuppMat Figure S9**).

More information about the process by which STADIA analyzes and quantifies dynamic instability and its limitations is provided in the Methods and **SuppMat Figure S1**.

In the analysis below, we used STADIA to analyze length history data sourced from both simulation and experiment (*in silico* and *in vitro*). The simulation dataset was obtained using our detailed kinetic Monte Carlo model of MT dynamics (Margolin et al. 2012). Please see the Data Acquisition subsections of the Methods section for additional information about the simulations and the physical experiments.

Constraining STADIA to perform two-state analysis, for comparison with classical DI measurements
In initial testing, we used STADIA to analyze the *in silico* and *in vitro* datasets under settings where we forced the classification stage of STADIA to assume that MT dynamics consist of only two states: growth and shortening phases ('Strictly two-state' analysis in **Table 1**). That is, all positive slope segments were classified as growth and all negative slope segments were classified as shortening. As might be expected, the DI parameters measured by STADIA under these constrained conditions are similar, but not identical, to those measured through a more traditional DI analysis method ('Peak-Valley Method' in **Table 1**, described in the Methods section).

Both of the methods used here begin by identifying major peaks and valleys (which correspond to catastrophes and rescues) in the length-history data such that the length change between a peak and valley exceeds a user defined threshold (**Figure 2 A,B**). While the segmentation process in the more traditional peak-valley method stops at this point, STADIA seeks to make further improvements by iteratively identifying additional vertices (segment endpoints, **Figure 2 D**) until the difference between the piecewise linear approximation and the raw length-history data is less than the user-defined maximum error tolerance. Effectively, as shown in **Figure 2**, the piecewise linear approximation produced by the segmentation stage of STADIA resembles the raw data more closely than does the approximation from classical methods.

Consequently, there are two fundamental differences between the segmentations resulting from classical methods (e.g. as implemented in our peak-valley method) and Stage I of STADIA. First, an individual segment of growth or shortening as identified by classical methods (**Figure 2 A,B**) may be composed of multiple segments of varying slopes in the STADIA analysis (**Figure 2 D**). Second, the refined approximation resulting from STADIA identifies segments of shallower slope that are not separated out from longer growth and shortening segments in classical methods.

Thus, STADIA can produce measurements of the traditional DI parameters (i.e., V_{growth}, V_{short}, F_{cat}, and F_{res}), but does so by using a finer linear approximation of the length-history data resulting in differences in the measured values of the DI parameters (**Table 1**). However, behaviors such as those captured in Figure 1C and 1E indicate that the traditional DI parameters alone are inadequate to capture the full range of MT dynamics as revealed by data acquired at high temporal resolution, indicating the possible need for more complex considerations when analyzing DI behaviors. These observations provided a solid foundation on which to proceed with using STADIA to analyze DI without preconceptions about how many phases (sustained and distinguishable behaviors) exist in MT length-history data.

Evidence for multiple types of behavior within the groups of positive and negative slope segments
After segments have been identified, the next task is to classify them. To perform the classification process in STADIA, we rely on an unsupervised machine learning algorithm, *k*-means clustering (Lloyd 1982; Macqueen 1967). This data driven approach does not presuppose that the clusters correspond to any particular DI phase; after the segments are assigned to clusters, the DI phase to which each cluster belongs will be determined based on cluster metrics, i.e., the average characteristics of segments in the cluster. The *k*-means algorithm requires that the number of clusters, *k*, be provided in advance (see the

Methods for more information regarding *k*-means clustering and its use in this analysis). Though various approaches exist for determining the *k*-value with which to perform the clustering (reviewed by (Steinley 2006; Pham, Dimov, and Nguyen 2005)), STADIA uses a measurement called the gap statistic (Tibshirani, Walther, and Hastie 2001). Analysis of the initial gap statistic data (generated by running STADIA in Diagnostic Mode; see Methods for more information) provided preliminary evidence that MT dynamics can be objectively divided into more behaviors than simply growth and shortening.

More specifically, we performed the gap statistic analysis for the two datasets (*in silico* and *in vitro*) already analyzed in Table 1, by separately processing positively- and negatively-sloped segments (i.e., growing and shortening segments). If MT growth and shortening each correspond to one behavior (with variation), one would expect to obtain a value of *k*=1 for each of the positive slope group and the negative slope group. However, with the *in silico* dataset, the gap statistic suggested *k*=3 to be the optimal number of clusters for each of these groups (**Figure 3 A,D & SuppMat Figures S4,S5**). These observations indicate that the *in silico* dataset contained multiple clusters of growth and shortening behaviors. In total, when including the near zero-slope segments (which were separated out before analysis of the positive and negative segments), 7 distinct clusters were identified in simulation data (**SuppMat Figure S7**).

Consistent with the *in silico* results, the analysis of the *in vitro* experimental data suggested *k*=3 for positive slopes (**Figure 3 B** and **SuppMat Figure S4**). However, differences were found between the *in silico* and *in vitro* data in analysis of the negative slopes: the optimal number of clusters was identified to be two (*k*=2) in both experimental datasets (**Figure 3 E** and **SuppMat Figure S5**), in contrast to the three clusters identified in the simulation data. This observation can be explained by the fact that for technical reasons, the *in vitro* dataset contains the beginning of shortening phases, but not the full loss of MTs to near-zero length, which was available in the simulation data. Consistent with this explanation, inspection of the clustering results for the negative slope segments in **Figure 3** shows that segments belonging to Negative Slope Cluster 3 (the cluster with the longest time durations) in the *in silico* data in **Figure 3 D** were not captured for the *in vitro* data in **Figure 3 E**. Therefore, we can only conclude that there are *at least* two clusters with negative slopes for the *in vitro* data. For illustration purposes, a 'ghost' region is added to **Figure 3 E** where we expect the missing third negative slope cluster to reside. Thus, including the flat slope segments, we find evidence for at least 6 distinct clusters in the experimental DI data: three clusters of growth, two clusters of shortening, plus a cluster that corresponds to the small number of flat slope segments that do not belong to either the positive or the negative slope segment groups (**SuppMat Figure S7**). Note that an additional cluster of shortening segments might be identified if full depolymerization events were captured in experiments.

In summary, application of *k*-means clustering with gap statistic analysis to DI data from either simulations or experiments leads to a similar conclusion: the data argue against the idea that MT DI can be characterized as a two-state process consisting only of growth and shortening with instantaneous transitions. More specifically, the results provide evidence for considering multiple types of growth behavior (3 clusters) and multiple types of shortening behavior (3 clusters, or 2 clusters for the truncated experimental data). Note that STADIA's identification of the cluster boundaries is driven by

the dataset itself, in contrast to many existing DI analysis methods that use arbitrary pre-defined thresholds. In the next section, we examine the differences between these clusters of length-history segments to determine how the segments in these clusters differ from each other and how these clusters might correspond to recognizably different phases of DI behavior.

Growth and shortening phases consistent with classical DI analysis are among the multiple types of behavior identified by STADIA

After using STADIA in Diagnostic Mode to perform gap statistic analysis and thus gain information about the optimal number of clusters to use in the *k*-means clustering process, we used STADIA in the Automated Mode with the suggested number of clusters (3 growth, and 2 or 3 shortening) to perform a full analysis of MT behavior. In the Automated Mode, STADIA first determines the centroid of each cluster of length-history segments. It then categorizes each segment identified from the segmentation stage as belonging to one cluster or another (see **SuppMat Figure S1** for an outline of the full analysis process; see the Methods section for more details). To study the relationships between these clusters of length-history segments and recognizable phases of DI, we examined the average characteristics of the segments in each group.

This analysis showed that, for both the *in silico* and *in vitro* data, some of the clusters correspond to the well-recognized growth and shortening phases of DI. More specifically, two of the positive segment clusters (positive slope clusters 1 and 2 from **Figure 3 A and B**) have slopes (rates of length change) similar to growth rates reported in classical DI analysis (compare STADIA results in **Figure 3 C** and **Table 1** to classical analysis results in **Table 1**). Similarly, negative slope cluster 2 (*in silico* and *in vitro*, **Figure 3 D and E**) and negative slope cluster 3 (*in silico*, **Figure 3 D**) have slopes similar to shortening rates reported in classical DI analysis (compare **Figure 3 F** and **Table 1**). Based on this information, in the classification stage of STADIA, length-history segments were assigned to the growth phase if they belonged to one of the clusters with a steep positive slope (Positive Slope Cluster 1 or 2 in **Figure 3 C,I**), and to the shortening phase if they belonged to a cluster with a steep negative slope (Negative Slope Cluster 2 or 3 in **Figure 3 F,I**).

The identification of two clusters within the bundled growth phase (and for the *in silico* data, within the bundled shortening phase) was unexpected. It is notable that in each case (both positive and negative slopes), the clusters differ primarily by duration (brief or sustained, **Figure 3 I** and **SuppMat Figure S8**). This observation may be evidence of different behaviors of tapered or split tips (e.g., as observed by (Coombes et al. 2013; Doodhi et al. 2016; Aher et al. 2018)) relative to the rest of the MT; such structures might be able to extend or retract faster than the bulk MT lattice in the absence of lateral bonds. Future work will investigate whether the differences between brief and sustained growth (or shortening) relate to tip structure.

In the next section, we consider the length-history segments that have much shallower slopes, which set them apart from the other growth and shortening behaviors discussed above.

STADIA detects and quantifies 'stutters': a dynamic phase distinct from growth and shortening

Examination of **Figure 3 A-F** shows that, in addition to clusters with slopes that correspond to rates of length change seen in classical growth or shortening behaviors, STADIA also identifies clusters with much shallower slopes (Positive Slope Cluster 3 and Negative Slope Cluster 1 in **Figure 3 A-F; Table 1**). Moreover, the segments in these shallow-slope clusters have time durations much shorter than typical segments classified as sustained growth and sustained shortening, though typically longer than those classified as brief growth and brief shortening segments (**SuppMat Figure S8**). We term these shallow-slope clusters of segments 'stutters' to convey the idea that these sections of length-history data exhibit high-frequency, low-amplitude fluctuations throughout which the MT length changes little from a macro-level perspective.

Note that stutters are distinguishable from previously identified 'pauses' during which the MT neither grows nor shortens detectably (e.g., (Yenjerla, Lopus, and Wilson 2010; Gierke, Kumar, and Wittmann 2010)). In contrast, MT lengths do indeed change dynamically during stutter periods, though the net rate of change is small. In addition, it is notable that events categorized as pauses are typically described as being rare (<1% of total behavior duration) in the absence of MT stabilizing proteins (e.g., (Walker et al. 1988; Moriwaki and Goshima 2016)). These observations support the conclusion that stutters are different from events previously classified as pauses, though there is likely some overlap, especially in cases where events categorized as pauses are allowed to be short in duration (e.g., (Walker et al. 1988; Guo et al. 2018)). Stutters as described above likely encompass the periods of slowed growth or shortening previously noted (but not quantified or characterized) in recent dynamic instability data acquired at high spatiotemporal resolution (e.g,. (Duellberg, Cade, and Surrey 2016; Rickman et al. 2017); see also (Margolin et al. 2012)). In contrast to previous work, we quantify and consider the role of stutters in DI as part of the procedures included in STADIA.

Together, these characteristics indicate that these transient periods of little length change are clearly distinct from either classical growth or shortening phases. Thus, we assigned these clusters with shallow slopes to a new DI phase class, 'stutters'. The stutter phase consists of 'up stutters' (Positive Slope Cluster 3), 'flat stutters' (Near-zero Slope Cluster) and 'down stutters' (Negative Slope Cluster 1) depending on whether the shallow slopes are positive, near zero, or negative, respectively (**Figure 3 I**).

At this point, every segment of length-history has been classified, and the assignment of individual segments to growth, shortening, and stutter phases can be visualized in the context of the original length-history data as in **Figure 3 G,H**.

MTs spend a significant fraction of time in the stutter phase
We begin to investigate the significance of stutter by first examining the fraction of time that MTs spend in the stutter phase. As one might expect, both *in silico* MTs and physical MTs spend the majority of their time in growth phases. However, in both simulations and experiments, MTs spend a substantial amount of time in stutter phases. Notably, in our *in silico* datasets, the MTs spent more time in stutter (8%) than in shortening (6%) (**Figure 4 A; SuppMat Figures S8,S9**). The *in vitro* MTs spent a substantial amount of the time in the stutter phase (**SuppMat Figures S8,S9**), but direct comparison to time spent in the shortening phase is not conclusive because depolymerizations were not fully captured. Given other

similarities between the simulated and physical MTs, it seems likely the ratio of time in stutter to time in depolymerization would be similar to that observed with simulation MTs. These observations indicate that stutters contribute appreciably to MT behavior as assessed in length-history plots.

Catastrophes are usually preceded by stutters *in silico* and *in vitro*

To investigate the functional significance of stutters, we used STADIA to examine how transitions between phases occur (**Figure 4 B-I; Figure 5**). Specifically, we wished to quantify all examples of transitions to/from growth and shortening, with or without stutter. Considering the chronological ordering of phases, STADIA automatically categorizes the following variations of these phase transitions:

- 'Abrupt Catastrophe': growth ➔ shortening directly (synonymous with classically recognized catastrophe) (**Figure 4 D; Figure 5 D**)
- 'Abrupt Rescue': shortening ➔ growth directly (synonymous with classically recognized rescue) (**Figure 4 E; Figure 5 E**)
- 'Transitional Catastrophe': growth ➔ stutter ➔ shortening (**Figure 4 F; Figure 5 F**)
- 'Transitional Rescue': shortening ➔ stutter ➔ growth (**Figure 4 G; Figure 5 G)**
- 'Interrupted Growth': growth ➔ stutter ➔ growth (**Figure 4 H; Figure 5 H,I**)
- 'Interrupted Shortening': shortening ➔ growth ➔ shortening (**Figure 4 I**)

Note that interrupted shortening is reported only in the simulation data, because only short portions of *in vitro* shortening phases were captured, which prevented observation of interrupted shortening. Similar chronological orderings of phases have previously been considered with pauses (Keller et al. 2008).

Remarkably, when we examined the simulation data, we found that 78% of catastrophes commenced with a stutter, i.e., were transitional (**Figure 4 B**). A related observation is that almost half (44%) of stutters that occurred after a growth segment ended in catastrophe as opposed to returning to growth (**Figure 4 C**). A similar but more dramatic association between stutter and catastrophe was observed in the *in vitro* control data: 86% of catastrophes commenced from a stutter (**Figure 5 A**), and 75% of stutters from growth ended in a catastrophe (**Figure 5 B**).

In contrast to catastrophes, rescues as observed *in silico* rarely occurred with stutter. More specifically, only 5% of *in silico* rescues were transitional (i.e., few rescues initiated from a stutter phase) (**Figure 4 B**), and only 8% of stutters that occurred during depolymerization resulted in a rescue (**Figure 4 C**). Because we do not have data for rescues *in vitro*, we cannot make strong conclusions on the correlation between stutters and rescue in physical MTs. However, these results do suggest that catastrophe and rescue are not simply the mechanistic opposites of each other.

CLASP2γ reduces the frequency of catastrophe by increasing the prevalence of interrupted growth

To further test STADIA's utility in analyzing dynamic instability and to examine both the prevalence and the significance of stutters, we analyzed another, comparable *in vitro* dataset in which the MTs were polymerizing in the presence of the MT binding protein CLASP2γ, which has been previously

characterized as an anti-catastrophe factor (Lawrence and Zanic 2019; Aher et al. 2018). CLASP2 proteins are of interest to the biomedical community because they have been implicated in functions as diverse as kinetochore attachment (Girão et al. 2019), nervous system development (Dillon et al. 2017), and the insulin response (Kruse et al. 2017).

In the presence of CLASP2γ, STADIA again identified separable stutter phases, similar to those identified in the control MTs *in vitro* (**SuppMat Figure S4,S5**). However, dramatic differences between the CLASP2γ data and control *in vitro* data were observed when these data were examined quantitatively by STADIA. First, the frequency of transitional catastrophes in the presence of CLASP2γ was significantly reduced (**Figure 5 A** and **SuppMat Figure S10**). This observation itself is not surprising, given that previous work (e.g., (Sousa et al. 2007; Lawrence et al. 2018; Aher et al. 2018; Majumdar et al. 2018)) has shown that CLASP2γ reduces the frequency of catastrophe (see also **Figure 5 C**). Strikingly, however, CLASP2γ also increased the frequency of interrupted growths (growth-stutter-growth) (**Figure 5 B** and **SuppMat Figure S10**). More specifically, among transitions that begin as growth-to-stutter, CLASP2γ increased the proportion of transitions that are growth-stutter-growth and decreased the proportion of transitions that are growth-stutter-shortening (**Figure 5 B**). Taken together, these data demonstrate that STADIA analysis provides information about CLASP2γ function not supplied by traditional analysis and indicates that CLASP2γ suppresses catastrophe at least in part by enabling stuttering MTs to re-enter the growth phase. This idea is supported by recent reports that MTs can withstand greater growth rate variability without undergoing catastrophe in the presence of CLASP2γ (Lawrence et al. 2018; Lawrence and Zanic 2019) and that CLASP2γ can protect against catastrophe in the presence of lagging protofilaments (Aher et al. 2018).

Mechanisms of stutters and implications for the process of catastrophe
What causes stutters, especially those that disrupt growth, and why are they associated with catastrophe? A fundamental component to answering this question comes from recognizing that when transitioning from growth to stutter, there is a net decrease in the number of subunits that are incorporated into the MT per unit time. This net decrease could occur because new subunits attach to the tip less frequently than during normal growth, or because bound subunits leave the tip more frequently than during growth, or a combination of these two.

While simple stochastic fluctuations in subunit arrival or departure could potentially contribute to stutters, changes in rates of attachment and detachment could also result from changes in tip structure. However, one could argue that the rate of subunit attachment should not vary with tip structure: assuming that longitudinal bonds form first, there are always 13 landing sites for new subunits (Castle and Odde 2013). Therefore, we suggest that stutters following growth segments likely result from a situation where an unusually large fraction of incoming subunits detach from the tip structure without being fully incorporated into the lattice, e.g., because tip taper or other structural features make it difficult for lateral bonds to form. In other words, we suggest that stutters occur when the structure of the tip is such that the subunit detachment rate is unusually high compared to the average detachment rate during growth. This reasoning provides a potential explanation for the correlation between stutter and catastrophe: if fewer subunits are incorporated into the lattice than normal, the stabilizing cap of

GTP-tubulin at the MT end will become smaller, the likelihood of exposing GDP-tubulin subunits will increase, and the possibility of complete cap loss (catastrophe) will rise. At present, these ideas are speculation, but future work may be able to shed light on these hypotheses (see also (McIntosh et al. 2018; Zakharov et al. 2015; VanBuren, Cassimeris, and Odde 2005)).

Relationship to previous work
Differences between stutters and previously identified pauses:
Stutters are similar in time duration to short pauses that have been observed by other authors (e.g., (Dhamodharan and Wadsworth 1995; Dhamodharan et al. 1995; Rusan et al. 2001; Matov et al. 2010)). Pauses can also be significantly longer (e.g., 80 seconds on average in (Rickman et al. 2017)). Pauses have usually been observed *in vivo* (see citations in the Introduction), and are likely caused by MT binding proteins (Moriwaki and Goshima 2016)) and other factors external to the MTs themselves (e.g., reaching the cell edge (Rusan et al. 2001; Komarova, Vorobjev, and Borisy 2002)). In contrast, *in vitro* pauses in the absence of drugs or MTBPs are rare (Walker et al. 1988). The observation that stutters are prevalent in both our *in silico* and *in vitro* datasets suggests that stutters are an intrinsic component of DI itself.

Gierke and colleagues have previously described "*bona fide* pauses" as phases "during which no polymerization or depolymerization occurs." Due to physical detection limits, true pauses would be indistinguishable from periods of very slow polymerization or depolymerization that do not meet the detection threshold (Gierke, Kumar, and Wittmann 2010). Particularly in older datasets with large thresholds (e.g., a length-change threshold of 0.5 microns), most stutters either would have been considered pauses or would not have been separated out from larger growth or shortening phases at all. With newer imaging technology, data can be obtained at high temporal and spatial resolution (e.g., 9.2 fps with 97 nm resolution, (Guo et al. 2018)). In our datasets and others (Maurer et al. 2014; Duellberg et al. 2016; Rickman et al. 2017), the resolution is sufficiently high to observe that length changes do indeed occur during stutters or slow-down episodes.

Thus, we propose the following criteria for distinguishing pauses and stutters: pauses are periods during which no length change occurs (i.e., the absence of dynamics even at the microscopic scale), whereas stutters are dynamic phases during which subunits are attaching to and detaching from the MT but with a smaller net rate of length change as compared to typical growth or shortening phases. In datasets that contain both stutters and pauses, the current version of STADIA would classify *"bona fide* pauses" as 'flat stutters'. However, future versions of STADIA will consider distinguishing pauses as a dedicated phase.

Previously observed behaviors that are similar to particular types of stutters:
Maurer et al. observed short episodes of pause or slow growth prior to catastrophes in experiments with EB1 (Maurer et al. 2014). These pre-catastrophe slow downs are analogous to transitional catastrophes in our terminology. Building beyond previous work, STADIA provides a comprehensive method for detecting and quantifying all types of phase transitions, and distinguishing phase transitions that include stutters from those that do not.

Pauses or slow-downs prior to catastrophe have also been observed in cases where the catastrophe is induced by outside factors such as mechanical force (Janson and Dogterom 2004) or reduction in tubulin concentration (Duellberg et al. 2016; Duellberg, Cade, and Surrey 2016), similar to predictions based on simulations in (Margolin et al. 2012). In contrast, the catastrophes in our datasets occur spontaneously as part of DI; in the *in vitro* datasets, EB1 and CLASP2γ affect the frequency of catastrophe, but the catastrophes still occur stochastically over time, as opposed to being induced by an experimenter at a particular moment.

The episodes of slow growth in Rickman et al. (Rickman et al. 2017) bear some similarity to stutters interrupting growth as identified by STADIA. However, the slow growth episodes of Rickman et al. occurred rarely (2 to 5 occurrences, ~0.26% to ~6.1% of the time analyzed, depending on tubulin concentration). These episodes appear to correspond to the most extreme of our stutters, meaning the stutters with the longest time durations or with the most variability in length during the stutter.

Based on analysis of variability in growth rates in experimental data, Odde et al. proposed a model with "near catastrophes", which are similar to stutters interrupting growth (David J. Odde, Buettner, and Cassimeris 1996). They suggested that the largest of the "near catastrophes" may correspond to previously observed pauses and that the smaller "near catastrophes" would not be easily detected by eye. The time between data points in their analysis was ~ 3 seconds.

Differences between STADIA and previous methods:
Similar to STADIA, many existing time-series analysis methods involve a segmentation step (e.g. (Zaliapin, Gabrielov, and Keilis-Borok 2004)) that is often followed by a classification step (e.g., (Fu 2011)) and have been used in other applications (e.g., identifying runs and pauses in the transport of organelles along MTs by motor proteins (Zaliapin et al. 2005)). To our knowledge, such methods have not been previously applied to MT dynamic instability data. In contrast, most existing DI analysis methods essentially perform classification before segmentation, by beginning with thresholds for classifying growth, shortening, and possibly pause or slow-down periods, and then applying the thresholds to identify segments in the data. Additionally, unlike existing methods that use arbitrary pre-defined thresholds on segment features (length change, time duration, and/or slope), STADIA uses a data-driven approach to identify emergent clusters in the segment feature data.

Although STADIA identifies DI phases at a finer scale than many existing DI analysis methods, STADIA differs from methods that simply consider individual displacements between frames and label them as growth, shortening, or pause using thresholds on the length change (e.g., (Komarova, Vorobjev, and Borisy 2002; Guo et al. 2018)). In contrast to such methods, STADIA identifies larger-scale segments during which a MT exhibits a consistent behavior.

In regard to phase transition analysis, several previous articles grouped their pauses with growth when defining catastrophe and rescue; by their definitions, a catastrophe is a transition from growth or pause to shortening, and a rescue is a transition from shortening to growth or pause (Panda et al. 1996;

Dhamodharan et al. 1995; Dhamodharan and Wadsworth 1995; Rusan et al. 2001; Kamath, Oroudjev, and Jordan 2010; Yenjerla, Lopus, and Wilson 2010; Kiris, Ventimiglia, and Feinstein 2010; Moriwaki and Goshima 2016). By these definitions or analogous definitions with stutter in place of pause, an episode of interrupted shortening would be labeled as a rescue followed by a catastrophe, whereas an interrupted growth would not be distinguished from *un*interrupted growth. STADIA improves upon typical transition analysis by considering all possible transitions between growth, shortening, and stutter (similar to the transitions among growth, shortening, and pause considered in (Keller et al. 2008)). Such transition analysis enables more in-depth investigation of the mechanisms of DI and DI-regulating proteins. For example, the observation that CLASP2γ tends to convert would-be transitional catastrophes into interrupted growths would not have been possible without a method that is able to identify transitional catastrophes and interrupted growths.

CONCLUSIONS

The key results of this work are four-fold: (1) the development of STADIA as an improved analytical tool for quantification of MT behavior; (2) the use of STADIA to more thoroughly examine a previously observed but unquantified phase in MT dynamics, 'stutter', during which MTs undergo little overall length change compared to growth or shortening phases, but do exhibit rapid low-amplitude fluctuations; (3) the observation that stutter is strongly associated with catastrophe *in silico* and *in vitro*; (4) the evidence that the anti-catastrophe factor CLASP2γ reduces catastrophe by increasing the fraction of stutters that return to growth rather than enter shortening phases. Our results concerning the detection of stutters differ from previous work in that STADIA comprehensively and systematically identifies all types of stutters (up stutter, flat stutter, down stutter) across length history data and considers all possible phase transitions containing stutters in its analysis. We suggest that quantification of stutters in future DI analysis through STADIA or similar tools will enable improved analysis of MT dynamics that is more complete, precise, and reproducible. The clearer picture that results from this analysis will facilitate investigation of the mechanisms of catastrophe and rescue and the activities of the MT binding proteins that regulate these transitions.

METHODS

CLASSICAL DI ANALYSIS

For purposes of comparison to STADIA, we use the custom DI analysis program written in MATLAB and described in the Supplemental Methods of (Jonasson et al. 2019), which we refer to as the "peak-valley method" (our implementation of classical DI analysis). Briefly, this method segments growth and shortening phases by first identifying major peaks and valleys in the length-history data using the MATLAB function *findpeaks()*. Then the ascent to each major peak is classified as a growth segment and the descent from the peak is classified as a shortening segment. Each major peak is classified as a catastrophe, where the end of growth and the start of shortening are identified as occurring at the same time point. A major valley is classified as a rescue only if the MT length at the time of the major valley is greater than or equal to a user-defined value called the 'minimum rescue length', in which case the end of shortening and the start of growth are identified as the same point. For a major valley that occurs below the minimum rescue length, the end of shortening can be identified as an earlier point in time

than the start of growth, in which case the time between these points would correspond to a nucleation period near the MT seed (see Supplemental Methods of (Jonasson et al. 2019) for additional details).

For the classical DI analysis in this paper, the minimum prominence for major peaks (minimum height change between a major peak and the nearest major valley) in the peak-valley method was set equal to the maximum height error tolerance in STADIA. The minimum peak height and the minimum rescue length in the peak-valley method were set equal to the sum of the nucleation height threshold plus the maximum height error tolerance in STADIA (see **SuppMat Table S1**).

In the peak-valley method results shown in **Table 1**, the V_{growth} and V_{short} calculations relied on linear regressions fitted to each growth or shortening segment. V_{growth} was calculated as the arithmetic mean of the slopes of the regression lines for all growth segments whose linear regression had an R^2 value of at least 95%. V_{short} was calculated in the same manner using the shortening segments. F_{cat} was calculated as the total number of catastrophes divided by the total time spent in growth phases. Similarly, the F_{res} was calculated as the total number of rescues divided by the total time spent in shortening phases. See **Table 1** for comparison of the DI parameters measured from the peak-valley method and from STADIA.

STATISTICAL TOOL FOR AUTOMATED DYNAMIC INSTABILITY ANALYSIS: <u>STADIA</u>
This section outlines the three major stages of STADIA analysis (Segmentation, Classification, and Phase and Transition Analysis) as well as the parameters used for the inspection of *in silico* and *in vitro* data using STADIA. Refer to **SuppMat Table S1** for a complete list of all STADIA user-defined parameters used for analysis of both *in silico* and *in vitro* MTs.

<u>Segmentation Stage</u>
In the segmentation stage, STADIA takes MT length-history data and generates a continuous piecewise linear approximation of the MT length-history plot. Segmentation also includes a preprocessing step that prepares the user's length-history data for input into STADIA and a post-processing step that prepares the results of the segmentation step for classification.

Justification for segmentation method: To create a more accurate approximation of MT length history data, STADIA employs an iterative approach to create a piecewise linear approximation of the MT length history data satisfying a user-defined maximum error tolerance. We chose this approach because it provides a simple method for identifying points where a change from one sustained MT behavior to another occurs, which may not necessarily be peaks or valleys. Through the maximum error tolerance choice, the user is able to precisely control the accuracy of the linear approximation. Through a minimum timestep choice, the user is able to perform the analysis of MT length history data at the desired timescale. An assumption of performing segmentation in this manner is that MT behavior follows a linear trend at the time scale being analyzed. Finally, this method results in a continuous piece-wise linear approximation, unlike other segmentation methods such as (Zaliapin, Gabrielov, and Keilis-Borok 2003), which produce discontinuous approximations. The choice to produce such an approximation is to address part of the 4th goal for STADIA development: enable comparison to and continuity with previous work. The approximation from STADIA is more comparable to classical DI

analysis methods by extending a more accurate result from the peak-valley method, thus allowing for the conclusions associated with DI behavior intermediate to growth and shortening to be more easily discussed.

Preprocessing: As an initial step, STADIA automatically formats the inputted MT length-history data into a single time series of length-history data points. MT length-history data can be provided either as a long-time observation of a single MT (possible with simulations) or as a series of length histories of multiple MTs (common with experimental data). In the latter case, STADIA automatically connects, or 'stitches', the data from multiple MTs (with separators in between) into a single time series representation of MT length-history data. Note that special treatment of the stitching separator between observations allows the segmentation to avoid misclassification of stitch boundaries as transitions. This preprocessing step allows STADIA to conduct analysis for both simulation data and experimental data in a similar and consistent manner.

In this manuscript, the simulation data were provided as one long time series from an individual MT (no stitching required), while the *in vitro* data (both with and without CLASP2γ) were obtained from multiple MTs over a shorter period of observation (for details, see *Data Acquisition: In Vitro Microtubule Experiments* below). Because long shortening phases were not captured (for technical reasons), the data from a specific MT were broken into samples that typically consisted of a growth phase followed by an initial depolymerization and then termination of that observation. STADIA first placed individual length-history samples for a given MT consecutively into the same time series plot, and then stitched all of the data for all of the MTs within each experiment. Note that the clustering methods used in STADIA require a dataset large enough to display a rich variety of possible DI behaviors. Therefore, instead of analyzing each individual *in vitro* MT for various behaviors, it is necessary to collectively consider multiple MTs from the same experiment so there is enough length-history data to classify. Thus, separately for the control and CLASP2γ *in vitro* datasets this stitching procedure combined all the available behavior into a dedicated single time series representation (one with CLASP2γ and one without).

Segmentation: STADIA takes the single time series graph produced by the preprocessing step and performs segmentation as an adaptive, iterative process according to restrictions provided by user-defined thresholds. The segmentation process begins by identifying major peaks and valleys (i.e., local extrema) in MT length-history data using the *findpeaks()* function in MATLAB. Consecutive extrema are connected by line segments to form an initial linear approximation of the length-history data (**Figure 2 C**). By starting with an initial list of vertices defined by these peaks and valleys, the iterative process seeks to include new vertices to define line segment endpoints. This improves the approximation accuracy by constructing a continuous piecewise linear approximation that satisfies the user-defined parameters of maximum error tolerance and minimum time-step (**Figure 2 D**). Note that the segmentation algorithm implemented in STADIA is similar to the 'Top-down algorithm' described in (Keogh et al. 2001). The segmentation algorithm can be explained in the following steps:

1. Let $\{x^1, x^2, \ldots, x^N\}$ represent these peaks and valleys that serve as the initial list of vertices (i.e. segment endpoints), where x^1 and x^N are the first and last points of the length history data, respectively.

2. For any $i = 1, \dots, N - 1$, define the ith region as the interval between the consecutive pair of initial vertices, x^i and x^{i+1}. Construct a line segment with end points as $x_1^i = x^i$ and $x_2^i = x^{i+1}$ such that the vertices corresponding to the ith region are $\{x_1^i, \dots, x_M^i\}$ (initially $M = 2$, but we seek to grow this list in the following steps).

3. For $j = 1, \dots, M - 1$, consider the jth line segment in the ith region defined by x_j^i and x_{j+1}^i. Calculate the error between this line segment and the corresponding points on the original length-history data.
   o If the maximum error is greater than the user-defined tolerance, then the error condition is not satisfied, and a new data point needs to be included in the vertex list. Proceed to step 4.
   o If the maximum error from this segment is less than the user-defined tolerance, then the error condition is satisfied for the jth line segment in the ith region. Proceed to step 6.

4. Define the data point where the greatest error occurs found in step 3 as x_{new}^i
   o If x_{new}^i violates the user-defined minimum duration constraint, attempt to choose the closest point in the length history data that would satisfy both the error and time-step conditions.

5. Include the newly identified vertex into the list of vertices for the ith region. This will require redefining indices to preserve ordering. For example, in the first step, 1 segment is broken into 2 segments, and the list of vertices corresponding to the ith region is now defined as
$$\{x_1^i, x_2^i, x_3^i\} = \{x_1^i, x_{new}^i, x_2^i\}$$
where the vertex list on the right side are indexed according to the previous iteration, and the updated vertex list on the left side replace the list defined in step 2, such that $x_j^i < x_{j+1}^i$ for all $j = 1, \dots, M - 1$.

6. Repeat steps 3 through 5 until the error condition is satisfied without adding new vertices into the ith region.

7. Repeat steps 2 through 6 for all $i \leq N - 1$.

Note that step 4 in the algorithm is careful not to violate the user-defined minimum time-step constraint by limiting how close two vertices can be to each other, which ensures that the approximation is constructed to the desired time scale. The final result is a continuous, piecewise linear approximation that fits the entire length-history dataset according to a user-defined error threshold (excerpts of the full length-history approximation are illustrated in **Figure 2 D**, and the black lines plotted in **Figure 3 G,H**). The vertices of the piecewise linear approximations provide line segments with endpoints at exact moments where significant changes in MT behavior occur. Thus, the activity covered by each segment between endpoints represents a consistent period of MT length-history behavior that can be identified as belonging to a DI phase in the classification stage.

The following user-defined parameters set the accuracy of the piecewise linear approximations for all datasets considered in this study: minimum duration = 0.5 seconds; error tolerance threshold = 20 subunits (**SuppMat Table S1**).

Post-processing to prepare for classification: Line segments from the piecewise linear approximation each have measured slopes, time durations, and height changes (**Figure 2 E**); this set of measurements provides a 3-D feature space where the segments reside (see below for *Justification for classification feature space*). However, some post-processing is needed before submitting this dataset to the classification process. First, new vertices are added to mark the boundaries between which the MT lengths are below a threshold length, generally chosen to be near the limits of observation in experimental conditions. These periods of time are described as 'nucleation' and are excluded from other analysis. Next, line segments are identified as flat stutters if either their total height change or slope magnitude are below user-defined thresholds. Flat stutters, which are obviously are not characterized as growth nor shortening, are set aside until the end of the classification procedures. The remaining positive and negative sloped segments are considered separately during the next stage (classification).

In this work, line segments containing MT lengths less than 75 subunits were considered nucleation segments. Segments were identified as flat stutters if the absolute value of their total height change was less than 3 subunits or the absolute value of their slopes was less than 0.5 subunits/second.

Justification for classification feature space: Mathematically, knowing the values of only two of time duration, height change, and slope provides sufficient information to calculate the value of the third variable. However, if only two of the three variables were used in the clustering step, then there would be data points that are well separated in the three-dimensional space but that become indistinguishable for all practical purposes when only two of the variables are considered (**Figure S3**). Additionally, which data points become indistinguishable would depend on which pair of variables was used (time duration and height change, time duration and slope, or height change and slope).

The Z = X/Y surface (**Figure S2A**) could be parameterized with only two variables in a way that would maintain the separation from the three-dimensional space. However, one or both of these two new variables would be some combination of the original three variables, and these combinations would not necessarily have clear physical meanings. We therefore chose to use all three of the basic variables (time duration, height change, and slope) to maintain a more direct connection to the biology.

The use of nonlinear combinations of variables is not uncommon in statistics. Additional combinations of our three basic variables as well as other variables may be worth exploring in future work that aims to further dissect MT length history behaviors. For the purposes of the present work, the three basic variables are sufficient for detecting distinct phases within the positive- and negative-slope groups.

Classification Stage
In the classification stage, STADIA takes the results of the segmentation stage (segregated positive and negative sloped line segments that approximate the MT length-history data) and analyzes them using *k*-means clustering, where the number of clusters, the *k*-value, is suggested by the gap statistic.

Justification for using k-means clustering: As noted at the beginning of the Results and Discussion section, the 2nd goal for STADIA development was that it be impartial to the number of behaviors exhibited by MTs, thus removing any assumptions about MT dynamics being restricted to two behaviors (i.e., only growth and shortening). The need for an impartial classification process mandates the use of an unsupervised machine learning method, of which a limited number of techniques are appropriate for this data. Because of its ease of use and interpretability, we chose to use *k*-means to handle classification of MT length-history data. Ideal datasets for *k*-means follow a Gaussian distribution and are regularly (globularly) shaped. Although our data is not Gaussian, *k*-means still provides an objective methodology to find substructures in the overall data structure. The observation that *k*-means enables us to identify and quantify stutters (a behavior that has previously been noted but not quantified) indicates that it provides a useful methodology for categorization and quantification of MT behavior.

Pre-processing: *K*-means clustering uses Euclidean distance (i.e., straight-line distance between two points in 3-D space) as the primary measurement in its algorithm to classify data. Therefore, all features should exist on the same scale so as to give each feature equal weight in the *k*-means classification process. To meet this requirement, the segment features (slope, height change, and time duration values) were transformed by first being log-scaled and then standardized with respect to each feature's statistics (i.e., by subtracting the mean and dividing by the standard deviation). Scaling and standardizing the data in this way is a common practice for analysis utilizing *k*-means clustering and allows for all features to be considered on the same scale to better suit the Euclidean distance used in *k*-means clustering (Hastie, Tibshirani, and Friedman 2009).

Determining appropriate number of clusters for each dataset: The *k*-value (i.e., number of clusters to use in *k*-means) was determined for positive and negative slopes separately in the **Diagnostic Mode** of STADIA. This process utilizes the 'gap statistic', which compares the within-cluster dispersion to a null reference distribution when seeking the optimal number of clusters that best separates the data during *k*-means clustering (i.e., the gap statistic answers the question: 'what number of clusters results in the best separation between the clusters?') (Tibshirani, Walther, and Hastie 2001). The gap statistic is the primary driver in determining the proper *k*-value for clustering the line segment data. However, it is also recommended for the user to check how well the number of clusters suggested by the gap statistic describes the dataset qualitatively. Typically, the optimal *k*-value corresponds to the first local maximum of the gap-statistic plot. In some cases, however, qualitative inspection of the data may suggest that the first local maximum is not optimal, in which case the next local maximum should be used.

For the purposes of informing the optimal *k*-value for use in *k*-means clustering, STADIA repeats clustering procedures for different *k*-values, ranging from 1 through 12, using 100 random starts for each value (*k*-means clustering does not converge to a global maximum so multiple starts are required to determine optimal centroid locations, described below). Simultaneously, STADIA measures the corresponding gap statistic for each value of *k*. As explained above, in our analyses the *k*-value corresponding to the first local maximum of the gap statistic plot was usually chosen as the optimal number of clusters. However, qualitative inspection of the clustering for the positive slopes of the *in*

vitro MTs without CLASP2γ and comparison to the other datasets suggested that the first local maximum greater than $k=1$ (i.e., $k=3$) described the data more appropriately.

k-means clustering: Once the optimal number of clusters is determined for both positive and negative slopes using the **Diagnostic Mode** of STADIA, the user inputs these k-values, and k-means clustering (Lloyd 1982; Macqueen 1967) is performed in the **fully Automated Mode** on the positive and negative slopes separately. Segments from simulation data were clustered using $k=3$ for each of the positive and negative slope groups. Similarly, all experimental data (either with or without CLASP2γ) was clustered using $k=3$ for positive slopes and $k=2$ for negative slopes (as discussed in the Results & Discussion, $k=2$ for negative slopes was appropriate for these datasets because the full depolymerizations were not captured for technical reasons). The final clustering results were obtained from using 500 random starts (again, multiple trials must be performed to determine optimal centroid locations as k-means is not guaranteed to converge to a global optimum); centroid locations that attained the lowest sum of squared distances between the centroids and each point in their respective clusters were chosen for further analysis.

Phase bundling: Following k-means clustering, the resulting positive and negative sloped clusters are collected, along with the 'flat stutters' that were removed prior to clustering, and all are considered together in the 3-D space defined by the segment features (**SuppMat Figure S7**). Additionally, statistics such as average slopes, average time duration, and average height change are calculated for each cluster and are reported (slopes are illustrated in **Figure 3 C,F** and slopes and time durations are illustrated in **SuppMat Figure S8**). Clusters with similar average slopes are bundled together to form larger groups representing DI phase classes (**Figure 3 I**). Clusters with slopes considerably less in magnitude (flatter) are grouped into a newly identified phase called 'stutters' (along with the 'flat stutters' not considered during the clustering process). The remaining clusters with slopes larger in magnitude (i.e., the higher positive valued and lower negative valued slope segments) more closely resembled the classically understood growth and shortening phases. The name 'stutters' suggests that though there may be micro-level fluctuations in the MT length, the net length does not change considerably over the duration of each stutter segment, especially when compared to segments classified as growth or shortening phases.

At this point, every segment identified during the segmentation stage has been classified as belonging to one of the following DI phase classes: nucleation, growth, shortening, or stutter. Applying these phase class labels to each segment in the length-history plot is illustrated in **Figure 3 G,H**. The chronological ordering of phases, recognized as phase transitions, can now be performed.

Phase and Transition Analysis Stage
After classifying segments into phases, classical methods of calculating DI metrics are adapted for use with the newly identified stutter phases.

Phase analysis: The following attributes of each phase class are calculated: percent time spent in each phase ($\frac{total\ time\ spent\ in\ phase}{total\ time} \times 100\%$), total number of segments (counts obtained from the piecewise linear approximation) for each phase, and percent height change corresponding to each phase (**Figure 4 A** and **SuppMat Figure S9**).

Transition analysis: Transition frequencies are calculated in a manner similar to what has been done classically. However, with the newly identified stutter phases, there are additional transitions to consider. In particular, it is necessary to determine whether catastrophes and rescues are (or are not) directly preceded by stutters. Catastrophes and rescues are identified as either abrupt (occurring without detectable stutters) or transitional (occurring via a stutter) (**Figure 4 D,E,F,G and Figure 5 D,E,F,G**). Additionally, our analysis quantifies interrupted growth (growth → stutter → growth) (**Figure 4 H; Figure 5 H,I**) and interrupted shortening (shortening → stutter → shortening) (**Figure 4 I**). Note that MTs shorter than a user-defined threshold are considered to be in 'nucleation' phase (here the threshold used was 75 dimer lengths). Transitions into or out of nucleation phases are not considered here because such MTs would be difficult to detect in experiments, and their behavior might be influenced by the seed.

In agreement with what has been done in classic DI analyses, frequencies of catastrophe and rescue are calculated as the ratio of the number of catastrophe or rescue events to the total time spent in growth or shortening, respectively (**Table 1; SuppMat Figure S9**). Similarly, interruptions are calculated as the ratio of the number of interrupted growths or interrupted shortenings to the total time spent in growth or shortening, respectively. Calculations can be done separately for abrupt and transitional types (e.g., $F_{Abrupt\ Catastrophe} = \frac{\#\ of\ abrupt\ catastrophes}{total\ time\ spent\ in\ growth}$) or collectively by simply adding the frequencies for each type together ($F_{cat} = F_{abrupt\ catastrophe} + F_{transitional\ catastrophe}$) (**SuppMat Figure S9**).

DATA ACQUISITION: *IN SILICO* MICROTUBULE EXPERIMENTS
This section outlines the details regarding the acquisition of simulation MT data including information about both the model and the parameters used.

The computational model: Stochastic model for simulating 13-protofilament (13-PF) MTs
The computational MT model used in this paper to generate the *in silico* length-history data was an updated version of the detailed, stochastic MT model published in (Margolin et al. 2012), which is a kinetic Monte Carlo simulation that tracks the state of individual subunits (representing tubulin dimers) in the entire 13-PF MT structure. Simulation of MTs is done using biochemical rate constants in conjunction with the Gillespie algorithm (Gillespie 1976, 1977) to sample event times and to build a stochastic simulation of individual molecular-level events (e.g., formation/breaking of longitudinal and lateral bonds, and hydrolysis). A key difference between the previous versions and the current computational model is strict adherence to the assumption that only one of the many possible biochemical events occurs at a time. The previous detailed level 13-PF MT model approximated hydrolysis events by allowing several subunits to hydrolyze simultaneously after one of the other four reaction events (lateral bonding/breaking or subunit gain/loss) have occurred. Hydrolysis events are

now considered as a possible event in the same way that the others are handled. This modification resulted in very little change in macro-level behavior of *in silico* MTs, but the ability to output dedicated observations to each dimer-level event is a more accurate representation of MT biochemistry. The overall result of the simulation is *in silico* MTs that exhibit macro-level DI behaviors in agreement with those observed previously (Margolin et al. 2012).

Simulation setup and parameters

The dimer-scale kinetic parameters used in this study to simulate a 13-protofilament MT using the model described above were tuned in (Margolin et al. 2012) based on *in vitro* DI measurements from (Walker et al. 1988); a detailed list of parameters can be found in **SuppMat Table S2**. For the purposes of this analysis, a single non-competing MT was simulated at a constant [free tubulin] of 10 μM for 10 hours of simulation time. The max PF length (i.e., the length of the longest of the 13 protofilaments) was reported as the length of the MT, which was used to generate a length-history plot passed into STADIA. Though the mean PF length could have been used to represent the length of the entire MT, better agreement with the *in vitro* data used here was found using the max PF length instead (see clustering profiles in **SuppMat Figure S4**). Each dimer has a length of 8 nm. The max and mean PF lengths are both reported in units of dimer lengths; this is not the same as the number of dimers in the MT, which would be 13 times the mean PF length.

DATA ACQUISITION: *IN VITRO* MICROTUBULE EXPERIMENTS

This section outlines the details regarding capture of experimental MT data including conditions for a control group (tubulin + EB1) and a group with MTBPs (tubulin + EB1 + CLASP2γ).

Protein preparation

His-CLASP2γ and His-EB1 were purified as previously described (Zanic et al. 2013; Lawrence et al. 2018). Bovine brain tubulin was purified using the high-molarity method (Castoldi and Popov 2003). Tubulin was labeled with TAMRA and Alexa Fluor 488 (Invitrogen) according to the standard protocols, as previously described (Hyman et al. 1991).

TIRF microscopy

Imaging was performed using a Nikon Eclipse Ti microscope with a 100×/1.49 n.a. TIRF objective, NEO sCMOS (complementary metal–oxide–semiconductor) camera; 488- and 561- solid-state lasers (Nikon Lu-NA); Finger Lakes Instruments HS-625 high speed emission filter wheel; and standard filter sets. An objective heater was used to maintain the sample at 35°C. Microscope chambers were constructed as previously described (Gell et al. 2010). In brief, 22 × 22 mm and 18 × 18 mm silanized coverslips were separated by strips of Parafilm to create a narrow channel for the exchange of solution (Gell et al. 2010). Images were acquired using NIS-Elements (Nikon).

Dynamic MT Assay

GMPCPP-stabilized MTs were prepared according to standard protocols (Hyman et al. 1992; Gell et al. 2010). Dynamic MT extensions were polymerized from surface-immobilized GMPCPP-stabilized templates as described previously (Gell et al. 2010). The imaging buffer consisted of BRB80

supplemented with 40 mM glucose, 40 µg/ml glucose oxidase, 16 µg/ml catalase, 0.5 mg/ml casein, 100 mM KCl, 10 mM DTT, and 0.1% methylcellulose. The imaging buffer containing 1 mM GTP and purified proteins was introduced into the imaging chamber. Dynamic MTs were grown with 12 µM Alexa 488-labeled tubulin and 200 nM EB1 with or without 400 nM CLASP2γ and imaged at 2 fps using a 100× objective and an Andor Neo camera (pixel size of 70 nm). Alexa-488-labeled tubulin was used at ratio of 23% of the total tubulin.

In vitro MT length-history data

Length-history data for *in vitro* MTs was obtained from 30 minute-long experiments using both a control group and a group with the stabilizing MTBP, CLASP2γ. Kymographs of dynamic microtubules were generated using the KymographClear macro for ImageJ, and the dynamic MT tip positions as a function of time were determined in KymographClear, using a thresholding-based, edge-detection method that can trace the microtubule tip position in kymographs with subpixel accuracy (Mangeol, Prevo, and Peterman 2016). A subset of this experimental dataset was previously published in (Lawrence et al. 2018). The control group data was acquired from 68 MT seeds, from which 776 individual traces were observed. The group with CLASP2γ was acquired from 29 MT seeds, from which 85 individual traces were observed. After applying the stitching preprocessing step during the STADIA segmentation stage, the control group and the group with CLASP2γ each generated a single stream time series representing length-history data with total time duration over 21 hours and 3.5 hours respectively. The collective consideration of all experimental data samples together meets the needs of the machine learning requirements for reliable clustering results (i.e., the lifetime of a single MT alone would not be a sufficient amount of data for *k*-means clustering). The *in vitro* MT lengths were measured in units of nm, and then divided by 8 nm per dimer length to convert to units of dimer lengths.

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REFERENCES

Aher, Amol, Maurits Kok, Ashwani Sharma, Michel O Steinmetz, Marileen Dogterom, and Anna Akhmanova. 2018. "CLASP Suppresses Microtubule Catastrophes through a Single TOG Domain." *Developmental Cell* 46 (1): 40–58. https://doi.org/10.1016/j.devcel.2018.05.032.

Applegate, Kathryn T., Sebastien Besson, Alexandre Matov, Maria H. Bagonis, Khuloud Jaqaman, and Gaudenz Danuser. 2011. "PlusTipTracker: Quantitative Image Analysis Software for the Measurement of Microtubule Dynamics." *Journal of Structural Biology* 176 (2): 168–84. https://doi.org/10.1016/j.jsb.2011.07.009.

Blackwell, Robert, Oliver Sweezy-Schindler, Christopher Edelmaier, Zachary R. Gergely, Patrick J. Flynn, Salvador Montes, Ammon Crapo, et al. 2017. "Contributions of Microtubule Dynamic Instability and Rotational Diffusion to Kinetochore Capture." *Biophysical Journal* 112 (3): 552–63. https://doi.org/10.1016/j.bpj.2016.09.006.

Castle, Brian T, and David J Odde. 2013. "Brownian Dynamics of Subunit Addition-Loss Kinetics and Thermodynamics in Linear Polymer Self-Assembly." *Biophysical Journal* 105 (11): 2528–40. https://doi.org/10.1016/j.bpj.2013.10.009.

Castoldi, Mirco, and Andrei V. Popov. 2003. "Purification of Brain Tubulin through Two Cycles of Polymerization–Depolymerization in a High-Molarity Buffer." *Protein Expression and Purification* 32 (1): 83–88. https://doi.org/10.1016/S1046-5928(03)00218-3.

Coombes, Courtney E., Ami Yamamoto, Madeline R. Kenzie, David J. Odde, and Melissa K. Gardner. 2013. "Evolving Tip Structures Can Explain Age-Dependent Microtubule Catastrophe." *Current Biology* 23 (14): 1342–48. https://doi.org/10.1016/j.cub.2013.05.059.

Desai, Arshad, and Timothy J. Mitchison. 1997. "Microtubule Polymerization Dynamics." *Annual Review of Cell and Developmental Biology* 13 (1): 83–117. https://doi.org/10.1146/annurev.cellbio.13.1.83.

Dhamodharan, R, M A Jordan, D Thrower, L Wilson, and P Wadsworth. 1995. "Vinblastine Suppresses Dynamics of Individual Microtubules in Living Interphase Cells." *Molecular Biology of the Cell* 6 (9): 1215–29. https://doi.org/10.1091/mbc.6.9.1215.

Dhamodharan, R, and P Wadsworth. 1995. "Modulation of Microtubule Dynamic Instability in Vivo by Brain Microtubule Associated Proteins." *Journal of Cell Science* 108 (Pt 4) (April): 1679–89. http://www.ncbi.nlm.nih.gov/pubmed/7615685.

Dillon, Gregory M., William A. Tyler, Kerilyn C. Omuro, John Kambouris, Camila Tyminski, Shawna Henry, Tarik F. Haydar, Uwe Beffert, and Angela Ho. 2017. "CLASP2 Links Reelin to the Cytoskeleton during Neocortical Development." *Neuron* 93 (6): 1344-1358.e5. https://doi.org/10.1016/j.neuron.2017.02.039.

Doodhi, Harinath, Andrea E Prota, Ruddi Rodríguez-García, Lukas C Kapitein, Anna Akhmanova, and Michel O Steinmetz. 2016. "Termination of Protofilament Elongation by Eribulin Induces Lattice Defects That Promote Microtubule Catastrophes." *Current Biology* 26 (13): 1713–21. https://doi.org/10.1016/j.cub.2016.04.053.

Duellberg, Christian, Nicholas I Cade, David Holmes, and Thomas Surrey. 2016. "The Size of the EB Cap Determines Instantaneous Microtubule Stability." *ELife* 5 (April). https://doi.org/10.7554/eLife.13470.

Duellberg, Christian, Nicholas Ian Cade, and Thomas Surrey. 2016. "Microtubule Aging Probed by Microfluidics-Assisted Tubulin Washout." *Molecular Biology of the Cell* 27 (22): 3563–73. https://doi.org/10.1091/mbc.E16-07-0548.

Fees, Colby P., Cassi Estrem, and Jeffrey K. Moore. 2017. "High-Resolution Imaging and Analysis of Individual Astral Microtubule Dynamics in Budding Yeast." *Journal of Visualized Experiments* 2017 (122). https://doi.org/10.3791/55610.

Fu, Tak Chung. 2011. "A Review on Time Series Data Mining." *Engineering Applications of Artificial Intelligence*. Elsevier Ltd. https://doi.org/10.1016/j.engappai.2010.09.007.

Gell, Christopher, Volker Bormuth, Gary J. Brouhard, Daniel N. Cohen, Stefan Diez, Claire T. Friel, Jonne Helenius, et al. 2010. "Microtubule Dynamics Reconstituted in Vitro and Imaged by Single-Molecule Fluorescence Microscopy." *Methods in Cell Biology* 95 (January): 221–45. https://doi.org/10.1016/S0091-679X(10)95013-9.

Gierke, Sarah, Praveen Kumar, and Torsten Wittmann. 2010. "Analysis of Microtubule Polymerization Dynamics in Live Cells." *Methods in Cell Biology* 97 (C): 15–33. https://doi.org/10.1016/S0091-679X(10)97002-7.

Gildersleeve, R F, A R Cross, K E Cullen, A P Fagen, and R C Williams. 1992. "Microtubules Grow and Shorten at Intrinsically Variable Rates." *The Journal of Biological Chemistry* 267 (12): 7995–8006. http://www.ncbi.nlm.nih.gov/pubmed/1569058.

Gillespie, Daniel T. 1977. "Exact Stochastic Simulation of Coupled Chemical Reactions." *Journal of Physical Chemistry* 81 (25): 2340–61. https://doi.org/10.1021/j100540a008.

Gillespie, Daniel T. 1976. "A General Method for Numerically Simulating the Stochastic Time Evolution of Coupled Chemical Reactions." *Journal of Computational Physics* 22 (4): 403–34. https://doi.org/10.1016/0021-9991(76)90041-3.

Girão, Hugo, Naoyuki Okada, Tony A Rodrigues, Alexandra O Silva, Ana C Figueiredo, Zaira Garcia, Tatiana Moutinho-Santos, et al. 2019. "CLASP2 Binding to Curved Microtubule Tips Promotes Flux and Stabilizes Kinetochore Attachments." *The Journal of Cell Biology* 219 (2). https://doi.org/10.1083/jcb.201905080.

Goodson, Holly V., and Erin M. Jonasson. 2018. "Microtubules and Microtubule-Associated Proteins." *Cold Spring Harbor Perspectives in Biology* 10 (6): a022608. https://doi.org/10.1101/cshperspect.a022608.

Guo, Yuting, Di Li, Siwei Zhang, Yanrui Yang, Jia-Jia Liu, Xinyu Wang, Chong Liu, et al. 2018. "Visualizing Intracellular Organelle and Cytoskeletal Interactions at Nanoscale Resolution on Millisecond Timescales." *Cell* 175 (5): 1430-1442.e17. https://doi.org/10.1016/j.cell.2018.09.057.

Hastie, Trevor, Robert Tibshirani, and Jerome Friedman. 2009. *Springer Series in Statistics: The Elements of Statistical Learning*. *Springer*. https://doi.org/10.1007/b94608.

Hyman, A. A., S. Salser, D. N. Drechsel, N. Unwin, and T. J. Mitchison. 1992. "Role of GTP Hydrolysis in Microtubule Dynamics: Information from a Slowly Hydrolyzable Analogue, GMPCPP." *Molecular Biology of the Cell* 3 (10): 1155–67. https://doi.org/10.1091/mbc.3.10.1155.

Hyman, Anthony, David Drechsel, Doug Kellogg, Steve Salser, Ken Sawin, Pam Steffen, Linda Wordeman, and Tim Mitchison. 1991. "Preparation of Modified Tubulins." *Methods in Enzymology* 196 (January): 478–85. https://doi.org/10.1016/0076-6879(91)96041-O.

Jánosi, Imre M., Denis Chréien, and Henrik Flyvbjerg. 2002. "Structural Microtubule Cap: Stability, Catastrophe, Rescue, and Third State." *Biophysical Journal* 83 (3): 1317–30. https://doi.org/10.1016/S0006-3495(02)73902-7.

Janson, Marcel E., and Marileen Dogterom. 2004. "Scaling of Microtubule Force-Velocity Curves Obtained at Different Tubulin Concentrations." *Physical Review Letters* 92 (24): 248101. https://doi.org/10.1103/PhysRevLett.92.248101.

Jonasson, Erin M., Ava J. Mauro, Chunlei Li, Ellen C. Labuz, Shant M. Mahserejian, Jared P. Scripture, Ivan V. Gregoretti, Mark Alber, and Holly V. Goodson. 2019. "Behaviors of Individual Microtubules and Microtubule Populations Relative to Critical Concentrations: Dynamic Instability Occurs When Critical Concentrations Are Driven Apart by Nucleotide Hydrolysis." *Molecular Biology of the Cell*, October, mbc.E19-02-0101. https://doi.org/10.1091/mbc.e19-02-0101.

Kamath, Kathy, Emin Oroudjev, and Mary Ann Jordan. 2010. *Determination of Microtubule Dynamic Instability in Living Cells*. *Methods in Cell Biology*. Vol. 97. https://doi.org/10.1016/S0091-679X(10)97001-5.

Kapoor, Varun, William G. Hirst, Christoph Hentschel, Stephan Preibisch, and Simone Reber. 2019. "MTrack: Automated Detection, Tracking, and Analysis of Dynamic Microtubules." *Scientific Reports* 9 (1): 1–12. https://doi.org/10.1038/s41598-018-37767-1.

Keller, Philipp J., Francesco Pampaloni, Gianluca Lattanzi, and Ernst H.K. Stelzer. 2008. "Three-Dimensional Microtubule Behavior in Xenopus Egg Extracts Reveals Four Dynamic States and State-Dependent Elastic Properties." *Biophysical Journal* 95 (3): 1474–86. https://doi.org/10.1529/biophysj.107.128223.

Keogh, Eamonn, Selina Chu, David Hart, and Michael Pazzani. 2001. "An Online Algorithm for Segmenting Time Series." *Proceedings - IEEE International Conference on Data Mining, ICDM*, 289–96. https://doi.org/10.1109/icdm.2001.989531.

Kiris, Erkan, Donovan Ventimiglia, and Stuart C. Feinstein. 2010. *Quantitative Analysis of MAP-Mediated Regulation of Microtubule Dynamic Instability in Vitro-Focus on Tau*. *Methods in Cell Biology*. Vol. 95. Academic Press Inc. https://doi.org/10.1016/S0091-679X(10)95024-3.

Komarova, Yulia A., Ivan A. Vorobjev, and Gary G. Borisy. 2002. "Life Cycle of MTs: Persistent Growth in the Cell Interior, Asymmetric Transition Frequencies and Effects of the Cell Boundary." *Journal of Cell Science* 115 (17).

Kruse, Rikke, James Krantz, Natalie Barker, Richard L. Coletta, Ruslan Rafikov, Moulun Luo, Kurt Højlund, Lawrence J. Mandarino, and Paul R. Langlais. 2017. "Characterization of the CLASP2 Protein Interaction Network Identifies SOGA1 as a Microtubule-Associated Protein." *Molecular and Cellular Proteomics* 16 (10): 1718–35. https://doi.org/10.1074/mcp.RA117.000011.

Lawrence, E. J., G. Arpag, S. R. Norris, and M. Zanic. 2018. "Human CLASP2 Specifically Regulates Microtubule Catastrophe and Rescue." *Molecular Biology of the Cell* 29 (10): 1168–77. https://doi.org/10.1091/mbc.E18-01-0016.

Lawrence, E. J., and M. Zanic. 2019. "Rescuing Microtubules from the Brink of Catastrophe: CLASPs Lead the Way." *Current Opinion in Cell Biology* 56 (February): 94–101. https://doi.org/10.1016/j.ceb.2018.10.011.

Lloyd, Stuart P. 1982. "Least Squares Quantization in PCM." *IEEE Transactions on Information Theory* 28 (2): 129–37. https://doi.org/10.1109/TIT.1982.1056489.

Macqueen, J. 1967. "Some Methods for Classification and Analysis." In *Proceedings of the Fifth Berkeley Symposium on Mathematical Statistics and Probability, Volume 1: Statistics*, 233:281–97. http://citeseer.ist.psu.edu/viewdoc/summary?doi=10.1.1.308.8619.

Mahrooghy, Majid, Shantia Yarahmadian, Vineetha Menon, Vahid Rezania, and Jack A. Tuszynski. 2015. "The Use of Compressive Sensing and Peak Detection in the Reconstruction of Microtubules Length Time Series in the Process of Dynamic Instability." *Computers in Biology and Medicine* 65 (October): 25–33. https://doi.org/10.1016/j.compbiomed.2015.07.014.

Majumdar, Shreoshi, Tae Kim, Zhe Chen, Sarah Munyoki, Shih-Chia Tso, Chad A. Brautigam, and Luke M. Rice. 2018. "An Isolated CLASP TOG Domain Suppresses Microtubule Catastrophe and Promotes Rescue." Edited by Thomas Surrey. *Molecular Biology of the Cell* 29 (11): 1359–75. https://doi.org/10.1091/mbc.E17-12-0748.

Maly, Ivan V. 2002. "Diffusion Approximation of the Stochastic Process of Microtubule Assembly." *Bulletin of Mathematical Biology* 64 (2): 213–38. https://doi.org/10.1006/bulm.2001.0265.

Mangeol, Pierre, Bram Prevo, and Erwin J.G. Peterman. 2016. "KymographClear and KymographDirect: Two Tools for the Automated Quantitative Analysis of Molecular and Cellular Dynamics Using Kymographs." *Molecular Biology of the Cell* 27 (12): 1948–57. https://doi.org/10.1091/mbc.E15-06-0404.

Margolin, Gennady, Ivan V Gregoretti, Trevor M Cickovski, Chunlei Li, Wei Shi, Mark S Alber, and Holly V Goodson. 2012. "The Mechanisms of Microtubule Catastrophe and Rescue: Implications from Analysis of a Dimer-Scale Computational Model." *Molecular Biology of the Cell* 23 (4): 642–56. https://doi.org/10.1091/mbc.E11-08-0688.

Matov, Alexandre, Kathryn Applegate, Praveen Kumar, Claudio Thoma, Wilhelm Krek, Gaudenz Danuser, and Torsten Wittmann. 2010. "Analysis of Microtubule Dynamic Instability Using a Plus-End Growth Marker." *Nature Methods* 7 (9): 761–68. https://doi.org/10.1038/nmeth.1493.

Maurer, Sebastian P., Nicholas I. Cade, Gergö Bohner, Nils Gustafsson, Emmanuel Boutant, and Thomas Surrey. 2014. "EB1 Accelerates Two Conformational Transitions Important for Microtubule Maturation and Dynamics." *Current Biology* 24 (4): 372–84. https://doi.org/10.1016/j.cub.2013.12.042.

McIntosh, Richard J., Eileen O'Toole, Garry Morgan, Jotham Austin, Evgeniy Ulyanov, Fazoil Ataullakhanov, and Nikita Gudimchuk. 2018. "Microtubules Grow by the Addition of Bent Guanosine Triphosphate Tubulin to the Tips of Curved Protofilaments." *Journal of Cell Biology* 217 (8): 2691–2708. https://doi.org/10.1083/jcb.201802138.

Mitchison, Tim, and Marc Kirschner. 1984. "Dynamic Instability of Microtubule Growth." *Nature* 312

(5991): 237–42. https://doi.org/10.1038/312237a0.

Moriwaki, Takashi, and Gohta Goshima. 2016. "Five Factors Can Reconstitute All Three Phases of Microtubule Polymerization Dynamics." *The Journal of Cell Biology* 215 (3): 357–68. https://doi.org/10.1083/jcb.201604118.

Odde, D. J., L. Cassimeris, and H. M. Buettner. 1995. "Kinetics of Microtubule Catastrophe Assessed by Probabilistic Analysis." *Biophysical Journal* 69 (3): 796–802. https://doi.org/10.1016/S0006-3495(95)79953-2.

Odde, David J., Helen M. Buettner, and Lynne Cassimeris. 1996. "Spectral Analysis of Microtubule Assembly Dynamics." *AIChE Journal* 42 (5): 1434–42. https://doi.org/10.1002/aic.690420524.

Panda, D, M A Jordan, K C Chu, and L Wilson. 1996. "Differential Effects of Vinblastine on Polymerization and Dynamics at Opposite Microtubule Ends." *The Journal of Biological Chemistry* 271 (47): 29807–12. https://doi.org/10.1074/jbc.271.47.29807.

Patel, Riya J., Kristopher S. Murray, Peter O. Martin, Michael Sinclair, Jared P. Scripture, Holly V. Goodson, and Shant M. Mahserejian. 2020. "Using STADIA to Quantify Dynamic Instability in Microtubules." In *Methods in Cell Biology*, 158:117–43. Academic Press Inc. https://doi.org/10.1016/bs.mcb.2020.03.002.

Pedigo, Susan, and Robley C Williams. 2002. "Concentration Dependence of Variability in Growth Rates of Microtubules." *Biophysical Journal* 83 (4): 1809–19. https://doi.org/10.1016/S0006-3495(02)73946-5.

Pham, D T, S S Dimov, and C D Nguyen. 2005. "Selection of K in K-Means Clustering." *Proceedings of the Institution of Mechanical Engineers, Part C: Journal of Mechanical Engineering Science* 219 (1): 103–19. https://doi.org/10.1243/095440605X8298.

Portran, Didier, Laura Schaedel, Zhenjie Xu, Manuel Théry, and Maxence V. Nachury. 2017. "Tubulin Acetylation Protects Long-Lived Microtubules against Mechanical Ageing." *Nature Cell Biology* 19 (4): 391–98. https://doi.org/10.1038/ncb3481.

Rickman, Jamie, Christian Duellberg, Nicholas I Cade, Lewis D Griffin, and Thomas Surrey. 2017. "Steady-State EB Cap Size Fluctuations Are Determined by Stochastic Microtubule Growth and Maturation." *Proceedings of the National Academy of Sciences of the United States of America* 114 (13): 3427–32. https://doi.org/10.1073/pnas.1620274114.

Rusan, N. M., C. J. Fagerstrom, A. M.C. Yvon, and P. Wadsworth. 2001. "Cell Cycle-Dependent Changes in Microtubule Dynamics in Living Cells Expressing Green Fluorescent Protein-α Tubulin." *Molecular Biology of the Cell* 12 (4): 971–80. https://doi.org/10.1091/mbc.12.4.971.

Sammak, Paul J., and Gary G. Borisy. 1988. "Direct Observation of Microtubule Dynamics in Living Cells." *Nature* 332 (6166): 724–26. https://doi.org/10.1038/332724a0.

Schek, Henry T., Melissa K. Gardner, Jun Cheng, David J. Odde, and Alan J. Hunt. 2007. "Microtubule Assembly Dynamics at the Nanoscale." *Current Biology* 17 (17): 1445–55. https://doi.org/10.1016/j.cub.2007.07.011.

Schulze, Eric, and Marc Kirschner. 1988. "New Features of Microtubule Behaviour Observed in Vivo." *Nature* 334 (6180): 356–59. https://doi.org/10.1038/334356a0.

Shelden, E., and P. Wadsworth. 1993. "Observation and Quantification of Individual Microtubule Behavior in Vivo: Microtubule Dynamics Are Cell-Type Specific." *Journal of Cell Biology* 120 (4): 935–45. https://doi.org/10.1083/jcb.120.4.935.

Smal, Ihor, Ilya Grigoriev, Anna Akhmanova, Wiro J. Niessen, and Erik Meijering. 2010. "Microtubule Dynamics Analysis Using Kymographs and Variable-Rate Particle Filters." *IEEE Transactions on Image Processing* 19 (7): 1861–76. https://doi.org/10.1109/TIP.2010.2045031.

Sousa, Aureliana, Rita Reis, Paula Sampaio, and Claudio E. Sunkel. 2007. "The *Drosophila* CLASP Homologue, Mast/Orbit Regulates the Dynamic Behaviour of Interphase Microtubules by Promoting the Pause State." *Cell Motility and the Cytoskeleton* 64 (8): 605–20.

https://doi.org/10.1002/cm.20208.

Steinley, Douglas. 2006. "K-Means Clustering: A Half-Century Synthesis." *British Journal of Mathematical and Statistical Psychology* 59 (1): 1–34. https://doi.org/10.1348/000711005X48266.

Tibshirani, Robert, Guenther Walther, and Trevor Hastie. 2001. "Estimating the Number of Clusters in a Data Set via the Gap Statistic." *Journal of the Royal Statistical Society: Series B (Statistical Methodology)* 63 (2): 411–23. https://doi.org/10.1111/1467-9868.00293.

Toso, Robert J., Mary Ann Jordan, Kevin W. Farrell, Brian Matsumoto, and Leslie Wilson. 1993. "Kinetic Stabilization of Microtubule Dynamic Instability in Vitro by Vinblastine." *Biochemistry* 32 (5): 1285–93. https://doi.org/10.1021/bi00056a013.

Tran, P. T., R. A. Walker, and E. D. Salmon. 1997. "A Metastable Intermediate State of Microtubule Dynamic Instability That Differs Significantly between plus and Minus Ends." *Journal of Cell Biology* 138 (1): 105–17. https://doi.org/10.1083/jcb.138.1.105.

VanBuren, Vincent, Lynne Cassimeris, and David J. Odde. 2005. "Mechanochemical Model of Microtubule Structure and Self-Assembly Kinetics." *Biophysical Journal* 89 (5): 2911–26. https://doi.org/10.1529/biophysj.105.060913.

Walker, R. A., E. T. O'Brien, N. K. Pryer, M. F. Soboeiro, W. A. Voter, H. P. Erickson, and E. D. Salmon. 1988. "Dynamic Instability of Individual Microtubules Analyzed by Video Light Microscopy: Rate Constants and Transition Frequencies." *The Journal of Cell Biology* 107 (4): 1437–48. https://doi.org/10.1083/jcb.107.4.1437.

Waterman-Storer, Clare M., and E. D. Salmon. 1997. "Actomyosin-Based Retrograde Flow of Microtubules in the Lamella of Migrating Epithelial Cells Influences Microtubule Dynamic Instability and Turnover and Is Associated with Microtubule Breakage and Treadmilling." *Journal of Cell Biology* 139 (2): 417–34. https://doi.org/10.1083/jcb.139.2.417.

Yenjerla, Mythili, Manu Lopus, and Leslie Wilson. 2010. "Analysis of Dynamic Instability of Steady-State Microtubules In Vitro by Video-Enhanced Differential Interference Contrast Microscopy with an Appendix by Emin Oroudjev." *Methods in Cell Biology* 95: 189–206. https://doi.org/10.1016/S0091-679X(10)95011-5.

Zakharov, Pavel, Nikita Gudimchuk, Vladimir Voevodin, Alexander Tikhonravov, Fazoil I. Ataullakhanov, and Ekaterina L. Grishchuk. 2015. "Molecular and Mechanical Causes of Microtubule Catastrophe and Aging." *Biophysical Journal* 109 (12): 2574–91. https://doi.org/10.1016/j.bpj.2015.10.048.

Zaliapin, I., A. Gabrielov, and V. Keilis-Borok. 2003. "Multiscale Trend Analysis." *Fractals* 12 (3): 275–92. https://doi.org/10.1142/S0218348X04002604.

———. 2004. "Multiscale Trend Analysis." *Fractals* 12 (3): 275–92. https://doi.org/10.1142/S0218348X04002604.

Zaliapin, I., I. Semenova, A. Kashina, and V. Rodionov. 2005. "Multiscale Trend Analysis of Microtubule Transport in Melanophores." *Biophysical Journal* 88 (6): 4008–16. https://doi.org/10.1529/biophysj.104.057083.

Zanic, Marija. 2016. "Measuring the Effects of Microtubule-Associated Proteins on Microtubule Dynamics In Vitro." In *The Mitotic Spindle*, 47–61. https://doi.org/10.1007/978-1-4939-3542-0_4.

Zanic, Marija, Per O. Widlund, Anthony A. Hyman, and Jonathon Howard. 2013. "Synergy between XMAP215 and EB1 Increases Microtubule Growth Rates to Physiological Levels." *Nature Cell Biology* 15 (6): 688–93. https://doi.org/10.1038/ncb2744.

Figure 1. Qualitative examples of behavior that does not fit two-state framework in high-resolution simulation (*in silico*) and experimental (*in vitro*) data. (A) An illustration of the classically recognized two-state representation of DI, recognizing behavior as simply growth and shortening phases, with instantaneous transitions known as catastrophe and rescue events. **(B,D)** Zoomed out (two-state representation analogous to (A)) length history plots of simulation data (detailed 13-PF model; see Methods) and experimental data (note that depolymerizations were not tracked in their entirety in these experiments). Black rectangles in B (simulation data) and D (experimental data) indicate the zoomed-in portions shown in C and E, respectively. **(C,E)** Closer inspection of transitions shows ambiguous behavior that cannot clearly be categorized as either growth or shortening. Other behavioral phases need to be considered in order to fully describe MT transitions and identify the exact moments where transitions occur.

Classical Segmentation Method

STADIA Segmentation Method

Figure 2. Continuous piecewise linear approximation of length history data & segment data extraction. *Classical Segmentation Method*: **(A)** Identify major vertices, such that local maxima and minima (blue triangles) are considered as catastrophes and rescues (or nucleation events) respectively. **(B)** Connect major vertices (orange lines) to create a segmentation and approximation of the MT length history data. Then calculate DI metrics from this approximation. *STADIA Segmentation Method*: **(C)** Generate initial approximation by finding major peaks and valleys (local extrema with high prominence) and connecting these with line segments. **(D)** Iteratively include new vertices (blue dots) where the highest error occurs in each segment to produce the final approximation, such that all point-wise errors are less than the user-defined threshold (usually 10 to 25 dimer subunits). Note: this method produces a closer approximation of the data than the classical segmentation method produces. **(E)** Calculate slope, height change, and time duration for each segment, and use for *K*-means clustering during the classification stage of STADIA.

Figure 3. Cluster analysis results, slope metrics for the different clusters across data sets, and subsets of length history data with subsequently labeled DI phases. STADIA performs the cluster analysis using the segment-feature metrics (slope, height change, and time duration) identified in the length history approximation during the segmentation stage. **(A,B)** Log-transformed and standardized segment features extracted from *in silico* and *in vitro* (control, no CLASP2γ) length history data approximations for positive sloped line segments. Three clusters are suggested by the gap statistic for positive slopes in both data sets. **(D,E)** Log-transformed and standardized segment features extracted from *in silico* and *in vitro* (control, no CLASP2γ) length history data approximations for negative sloped line segments. The gap statistic suggested 3 clusters for the *in silico* negative sloped segments and 2 clusters for the *in vitro* negative sloped segments. **(C,F)** Growth rates for the positive sloped segment clusters and shortening rates for negative sloped segment clusters identified in the *in silico* and *in vitro* data sets. For positive slopes in each data set, two clusters (light and dark green) had average growth rates relatively large in magnitude compared to the third (light blue). For negative slopes in each data set, the red (and light red for *in silico* data) labeled shortening rates were on average relatively large in magnitude compared to the purple labeled group. **(G,H)** Previously unlabeled *in silico* and *in vitro* MT length history plots (see Figure 1 A,C) are now labeled according to the subgroups that each line segment fits into. Zoomed-in portions of previously ambiguous length history data (see Figure 1 B,D) are now clearly labeled as well-defined DI phases. **(I)** Examination of the average slopes of the individual clusters indicates that bundling subgroups (clusters) together into larger phase classes based on the average slopes of the individual clusters is appropriate. Clusters with positive and negative slopes relatively larger in magnitude were bundled together into 'Growth' and 'Shortening' phases respectively. The remaining clusters, where significantly less changes in length occur, were bundled together, along with the previously identified 'near zero' slope or flat segments, into a new phase called 'Stutters'. Further, 'Brief' and 'Sustained' sub-classes of the Growth and Shortening phases were characterized by their time durations. The 'Up', 'Flat', and 'Down' sub-classes of the Stutter phase are characterized by the segment slope being positive, near-zero, or negative, respectively.

Figure 4. Phase metrics and transition analysis for *in silico* data (Max Protofilament length). (A) Percent time spent in and percent height (MT length) change occurring during each bundled phase. A large majority of time is spent in the growth phase. Interestingly, *in silico* MTs spend more time in the stutter phase than in the shortening phase, thus making stutters a significant phase worth studying. Most height change occurs during growth and shortening phases, as is expected. Stutters account for a markedly smaller percentage of height change, particularly when considered relative to their percentage of time; this makes sense given that little height change takes place during stutter segments. **(B)** Percentages of transitional vs. abrupt phase changes (transitional = with stutter phase between the growth/shortening phases; abrupt = without stutters) to/from growth and shortening show that catastrophes are primarily transitional, whereas rescues are overwhelmingly abrupt. **(C)** The percentage of phase transitions with stutters are compared separately for growth-to-stutters transitions and shortening-to-stutter transitions. A bit more than half of the transitions entering stutters from a growth phase were observed to return to a growth phase; in other words, stutter phases occur somewhat more commonly in interrupted growth transitions than transitional catastrophes. A vast majority of transitions entering stutters from a shortening phase return to shortening, i.e., stutter phases appear in interrupted shortening transitions much more commonly than transitional rescues. **(D,E,F,G)** Examples of abrupt/transitional catastrophes (D,F) and abrupt/transitional rescues (E,G) for *in silico* MTs. Background colors indicate the subgroup identified by STADIA for various sections of length history data. **(H,I)** Examples of interrupted growth and interrupted shortening for *in silico* MTs, where interruption is defined by MTs in a growth/shortening phase undergoing a transition into a stutter phase, and then returning to growth/shortening (growth-stutter-growth or shortening-stutter-shortening).

Figure 5. Effect of CLASP2γ on the nature of catastrophes and the fate of MTs entering stutter phase. (A) Consistent with what was seen for *in silico* MTs, the majority of catastrophes for *in vitro* MTs without CLASP2γ are transitional. In contrast, introduction of CLASP2γ results in a reduction of stutter phases preceding catastrophes; it is possible that this is due to CLASP2γ promoting growth after stutter phases. **(B)** When MTs transition from growth to stutter phases, they are more likely to transition into a depolymerization phase when CLASP2γ is not present (i.e., without CLASP2γ, transitional catastrophes occur more often than interrupted growth). With CLASP2γ, however, when MTs move from growth to stutter phases, they are more likely to transition back into the growth phase (i.e., with CLASP2γ, interrupted growth occurs more often than transitional catastrophes). These results provide a possible explanation for how CLASP2γ changes the overall makeup of catastrophes for *in vitro* MTs: transitions that would have been transitional catastrophes without CLASP2γ are now interrupted growths with CLASP2γ. **(C)** CLASP2γ decreases the overall frequency of catastrophe without significantly reducing the frequency of transitioning from the growth phase to the stutter phase, indicating that CLASP2γ is preventing catastrophes by promoting growth following stutters without preventing stutters altogether. **(D,E,F,G)** Examples of transitional and abrupt catastrophes, for *in vitro* MTs both with (bottom) and without (top) CLASP2γ, show a clear qualitative distinction between transitional and abrupt catastrophes identified by STADIA. **(H,I)** Examples of interrupted growth exhibited by *in vitro* MTs both with (bottom) and without (top) CLASP2γ shows a transition from growth into stutter and back into growth phase.

Comparison of Full Analysis to k=1 and Classical Method

Simulation Data (Max PF Length)

Method	Total Cat.	Total Res.	F_{cat} (min^{-1})	F_{res} (min^{-1})	V_{growth} (nm/s)	V_{short} (nm/s)
Peak-Valley Method	355	123	0.659	2.483	46.1 ± 5.1	540.0 ± 47.9
Strictly two-state: *k=1 for pos & neg	449	214	0.912	4.391	46.4 ± 18.4	530.4 ± 556.0
Two-state w/ flat stutters: **k=1 for pos & neg	429	195	0.870	4.098	47.2 ± 17.6	547.2 ± 556.8
All phases found by STADIA: k=3 pos, k=3 neg	298	75	0.660	1.944	48.0 ± 7.2 63.2 ± 19.2 22.4 ± 8.8	552.8 ± 87.2 1016.8 ± 717.6 107.2 ± 72.8

Experimental Data (Control)

Method	Total Cat.	Total Res.	F_{cat} (min^{-1})	F_{res}*** (min^{-1})	V_{growth} (nm/s)	V_{short}**** (nm/s)
Peak-Valley Method	802	40	0.717	N.D.	29.5 ± 12.7	330.1 ± 136.5
Strictly two-state: *k=1 for pos & neg	856	83	0.777	N.D.	32.0 ± 24.8	216 ± 199.2
Two-state w/ flat stutters: **k=1 for pos & neg	846	76	0.760	N.D.	32.8 ± 24.8	227.2 ± 198.4
All phases found by STADIA: k=3 pos, k=2 neg	734	18	0.756	N.D.	30.4 ± 7.2 60.0 ± 44.8 15.2 ± 5.6	373.6 ± 143.2 39.2 ± 25.6

Experimental Data (CLASP2γ)

Method	Total Cat.	Total Res.	F_{cat} (min^{-1})	F_{res}*** (min^{-1})	V_{growth} (nm/s)	V_{short}**** (nm/s)
Peak-Valley Method	99	62	0.500	N.D.	43.1 ± 34.4	155.1 ± 77.6
Strictly two-state: *k=1 for pos & neg	142	94	0.720	N.D.	46.4 ± 41.6	96.0 ± 84.0
Two-state w/ flat stutters: **k=1 for pos & neg	131	87	0.676	N.D.	48.0 ± 41.6	108 ± 82.4
All phases found by STADIA: k=3 pos, k=2 neg	86	52	0.498	N.D.	37.6 ± 11.2 100.0 ± 57.6 16.0 ± 7.2	158.4 ± 68.8 32.8 ± 17.6

Legend:
- ▬ Sustained Growth
- ▬ Brief Growth
- ▬ Up Stutter
- ▬ Down Stutter
- ▬ Sustained Shortening
- ▬ Brief Shortening

Table 1. Comparison of DI metrics from classical two-state analysis, STADIA two-state analysis, and STADIA full analysis. The results from the full, automated analysis conducted by STADIA were compared to results from both the classical method (identifying only major peaks and valleys, and connecting line segments to form a course-grained approximation) as well as two-state approaches (a fine-grained approximation was generated by STADIA, but phase classes were restricted to only growth and shortening (*) or growth, shortening, and flat stutters (**)). While there is not one-to-one correspondence between any of the methods, there is general agreement where possible. For the experimental datasets, depolymerizations were not captured in their entirety, so (***) rescue data was not reported and (****) negative slope segments were separated into only two clusters, yielding only two V_{short} measurements. V_{growth} and V_{short} measurements are listed in a mean standard deviation format. Additionally, rescue metrics were not determined (N.D.) for experimental data due to depolymerizations not being captured in their entirety.

Figure S1. Workflow diagram outlining main steps of STADIA.

Figure S2. (A) Data points representing each line segment reside on this Z(X,Y) = X/Y manifold, where Z = slope, X = time, and Y = height. **(B)** Justification for using height, time, and slope is demonstrated using a parallel example in two dimensions where we plot a dataset containing four groups (clusters) of points that fall on the curve determined by the function y = 1/x. Plotting only y (top left) or only x (bottom right) creates the appearance that this dataset contains only three groups of points. In contrast, when the data are plotted in two dimensions (upper right), the data are separated sufficiently to reveal that the dataset actually contains four groups of points. For similar reasons, we need to consider all three variables in our line segment data to properly identify the groups in our dataset.

Figure S3. The segment features (slope, height change, and time duration) for segments identified from the piecewise linear approximation to the *in silico* length history data. Each point corresponds to one line segment from the length history approximation and is colored according to the phase class identified by STADIA after a full analysis. **(A-D)** Multiple perspectives of the same plot, provided to help visualize the 3-D data. **(E)** An illustration of how the segment points lie on the Z=Y/X manifold described in Supplemental Figure S2.

Figure S4. Gap Statistic plots and corresponding clustering profiles for positive slope segments in each log transformed and normalized data set. When using the gap statistic to suggest the best number of clusters to use in *k*-means clustering, the rule of thumb is to use the first *k*-value where the gap statistic plot shows a local maximum. In practice here, we expect the number of clusters to be greater than 1, because the 3-D data structure shows multiple appendages separated by a sparsely populated region of points. Thus, for the case with simulation data using the mean PF length and the control experimental data, the local maximum at *k*=1 is rejected. Taking this into consideration, all data sets indicate that the gap statistic attains the first local maximum greater than one at *k*=3. So, for all positive slope segment data, *k*-means clustering is performed by separating the data into 3 clusters. Furthermore, since the clustering profile of the simulation data using the max PF length more closely resembles the clustering profile of the experimental data, we choose to use the max PF data instead of the mean PF data for presenting the STADIA results in the main text.

Figure S5. Gap Statistic plots and corresponding clustering profiles for negative slope segments in each log transformed and normalized data set. When using the gap statistic to suggest the best number of clusters to use in *k*-means clustering, the rule of thumb is to use the first *k*-value where the gap statistic plot shows a local maximum. The two simulation data sets indicate that the gap statistic attains the first local maximum greater than one at *k*=3, whereas the experimental data sets indicate *k*=2. We attribute this disagreement to the fact that in these experimental datasets, only the beginnings of depolymerization events were captured, thus omitting long time duration shortening segments from the dataset. So, for negative slope segments, we performed *k*-means clustering separating the simulation data into 3 clusters and the experimental data into 2 clusters.

Figure S6. Gap Statistic plots and segment feature plots for an analysis where all slope segments in each data set were considered together (excluding flat segments), not separated into positive and negative slopes as in Figures S4 and S5. Note that the gap statistic plots are monotonically increasing, indicating that the initial dataset was too complex for effective calculation of the gap statistic and that we needed to subdivide it before further analysis. For this reason, the data are not color-coded as in the previous two figures.

Figure S7. Clustering profiles for ALL segments of *in silico* and *in vitro* data. Following separate classification of the positive and negative slopes (see Figures S4 and S5), segment data is un-log-transformed and unstandardized for full view of line segment data in 3-D space. Note that the classification step has already taken place, and these figures are simply for visualizing how the clusters exist in relation to each other in 3-D space.

Figure S8. Box and whisker plots of rates of MT length change (subunits/sec) and time durations for different subgroups across data sets to motivate subsequent bundling (box and whiskers cover the four quartiles of each subset, and the 'X' marks the mean value in the plots). *Note: the first two data sets in A and B are the same as the data presented in Figure 3 C and F, respectively.* **(A)** Growth rates for the positive sloped segment clusters identified in the simulation data sets and the experimental data sets without and with CLASP2γ. In each data set, two clusters (light and dark green) had average growth rates relatively large in magnitude compared to the third (light blue). **(B)** Shortening rates for the negative sloped segment clusters identified in the simulation data and in the experimental data sets without and with CLASP2γ. In each data set, the red-labeled shortening rates were on average relatively large in magnitude compared to the purple-labeled group. **(C)** Time durations for the positive sloped segment clusters show that one subset of segments (light green) represents a longer, more sustained period of consistent behavior than the other two subsets (blue and dark green) for all data sets considered. **(D)** Time durations for negative sloped segment clusters from simulation data also shows that one subset (light red) represents a longer, more sustained period of consistent behavior than the other two subsets (purple and dark red). Since the experimental data did not capture most of the shortening behavior, analysis of longer time duration shortening segments was not possible for the *in vitro* data sets. **(E)** Bundling subgroups together into larger phase classes based on the average slopes of individual clusters. Clusters with positive and negative slopes relatively larger in magnitude were bundled together into 'Growth' and 'Shortening' phases respectively. The remaining clusters, where significantly less changes in length occur, were bundled together, along with the previously identified 'near zero' slope or flat segments, into a new phase called 'Stutters'. Further, 'Brief' and 'Sustained' sub-classes of the Growth and Shortening phases were characterized by their time durations. The 'Up', 'Flat', and 'Down' sub-classes of the Stutter phase are characterized by the segment slope being positive, near-zero, or negative respectively.

Figure S9. Segment statistics for all *in silico* and *in vitro* data sets. *Number of segments*, *percent time*, and *percent height change* for each cluster are recorded for each data set. For the *in silico* data sets where depolymerizations were fully captured, the breakdown of the number of segments, time duration, and height change are representative of the actual time the simulated MT spent in the various phases. As noted throughout the paper, the *in vitro* depolymerizations were not captured in their entirety, and so the number of negative slope segments, percent time, and the percent height change attributed to negatively sloped MT behavior is largely underreported.

Transition Statistics		
	Count	Frequency (s^{-1})
Simulation Data (Mean PF Length)		
Abrupt Catastrophe	20	0.00071
Transitional Catastrophe	238	0.0084
Total Catastrophe	**258**	**0.00911**
Abrupt Rescue	39	0.0164
Transitional Rescue	6	0.0025
Total Rescue	**45**	**0.0189**
Interrupted Growth	75	0.0027
Interrupted Shortening	32	0.0135
Simulation Data (Max PF Length)		
Abrupt Catastrophe	67	0.0025
Transitional Catastrophe	231	0.0085
Total Catastrophe	**298**	**0.0110**
Abrupt Rescue	71	0.0307
Transitional Rescue	4	0.0017
Total Rescue	**75**	**0.0324**
Interrupted Growth	293	0.0107
Interrupted Shortening	48	0.0207
Experimental Data (Control)		
Abrupt Catastrophe	105	0.0018
Transitional Catastrophe	629	0.0108
Total Catastrophe	**734**	**0.0126**
Abrupt Rescue		
Transitional Rescue	N.D.	
Total Rescue		
Interrupted Growth	211	0.0036
Interrupted Shortening	0	0
Experimental Data (CLASP2γ)		
Abrupt Catastrophe	51	0.0049
Transitional Catastrophe	35	0.0034
Total Catastrophe	**86**	**0.0083**
Abrupt Rescue		
Transitional Rescue	N.D.	
Total Rescue		
Interrupted Growth	92	0.0088
Interrupted Shortening	1	0.0043

Figure S10. Detailed transition statistics for each data set. Both *in silico* data sets as well as the control experimental data set demonstrate that a significant majority of catastrophes occur via stutter (i.e., transitional catastrophe), while the CLASP2γ data set shows a shift to MTs exhibiting abrupt catastrophes. We speculate that the shift to abrupt catastrophes is due to CLASP2γ promoting tip extensions (see the Results and Discussion for more information on mechanistic speculation regarding the effects of CLASP2γ). Rescue data for *in silico* MTs indicates clearly that rescues largely occur abruptly. Note that rescue metrics were not determined (N.D.) for the *in vitro* data due to a lack of depolymerizations captured for *in vitro* MTs.

STADIA: User-defined Parameters	
Nucleation height threshold	75 subunits
Minimum time duration of a linear segment	500 ms
Maximum height error tolerance	20 subunits
Maximum height change for near-zero slope segments	3 subunits
Maximum slope magnitude for near-zero slope segments	0.5 subunits/sec
Number of centroids for positive slope segments	3
Number of centroids for negative slope segments (*in silico* data)	3
Number of centroids for negative slope segments (*in vitro* data)	2

Classical Analysis: User-defined Parameters	
Minimum peak height	95 subunits
Minimum rescue length	95 subunits
Minimum Prominence For Major Peaks	20 subunits
Minimum Prominence For Minor Peaks	0.1 subunits
Minimum Regression R^2	0.95

Table S1. User-defined parameters for STADIA and classical analysis.

13-PF Detailed MT Computational Model Parameters	
Number of protofilaments	13
Tubulin concentration	10 µM
Simulation time	10 hours
Seam shift	1.5 subunit lengths
Compete for tubulin	No
Hydrolysis rate	0.7 subunits/sec
HalfMax	200
kgrowT	250
kgrowD	250
kshortT	0.02
kshortD	20
kbondTT	100
kbondTD	100
kbondDT	100
kbondDD	100
kbreakTT	70
kbreakTD	90
kbreakDT	90
kbreakDD	400
SkbondTT	200
SkbondTD	200
SkbondDT	200
SkbondDD	200
SkbreakTT	140
SkbreakTD	180
SkbreakDT	180
SkbreakDD	800

Table S2. Computational model parameters used to generate simulation data. Parameters used are from Margolin et al. 2012.