Quantifying the importance of an inference model in Bayesian phylogenetics

in Bayesian phylogenetics

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Summary

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¹⁰   **1.**   Phylogenetic trees are currently routinely reconstructed from an
¹¹ alignment of character sequences (usually nucleotide sequences). Bayesian
¹² tools, such as MrBayes, RevBayes and BEAST2, have gained much popu-
¹³ larity over the last decade, as they allow joint estimation of the posterior
¹⁴ distribution of the phylogenetic trees and the parameters of the underlying
¹⁵ inference model. An important ingredient of these Bayesian approaches
¹⁶ is the species tree prior. In principle, the Bayesian framework allows
¹⁷ for comparing different tree priors, which may elucidate the macroevolu-
¹⁸ tionary processes underlying the species tree. In practice, however, only
¹⁹ macroevolutionary models that allow for fast computation of the prior
²⁰ probability are used. The question is how accurate the tree estimation
²¹ is when the real macroevolutionary processes are substantially different
²² from those assumed in the tree prior.

²³ **2.**   Here we present `pirouette`, a free, libre and open-source R package
²⁴ that assesses the inference error made by Bayesian phylogenetics for a
²⁵ given macroevolutionary diversification model. `pirouette` makes use of
²⁶ BEAST2, but its philosophy applies to any Bayesian phylogenetic infer-
²⁷ ence tool.

²⁸ **3.**   We describe `pirouette`'s usage and the biological scientific question
²⁹ it can answer, including full examples.

³⁰ **4.**   Last, we discuss the results obtained by the examples and their in-
³¹ terpretation.

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³³ **Keywords:** Bayesian model selection, BEAST2, computational biology,
³⁴ evolution, phylogenetics, R, tree prior

1 Introduction

The development of new powerful Bayesian phylogenetic inference tools, such as BEAST [Drummond & Rambaut 2007], MrBayes [Huelsenbeck & Ronquist 2001] or RevBayes [Höhna *et al.* 2016a] has been a major advance in constructing phylogenetic trees from character data (usually nucleotide sequences) extracted from extant (but also extinct, where possible) organisms, and hence in our understanding of the main drivers and modes of diversification.

BEAST [Drummond & Rambaut 2007] is a typical Bayesian phylogenetics tool, that needs both character data and priors to infer a posterior distribution of phylogenies. Specifically, for the species tree prior - which describes the process of diversification - BEAST has built-in priors such as the Yule [Yule 1925] and (constant-rate) birth-death [Nee *et al.* 1994] models. These simple tree priors are the most commonly used, as they represent some biologically realistic processes (i.e. branching at per species constant rates), while being computationally fast. BEAST's successor, BEAST2 [Bouckaert *et al.* 2019], has a package manager, that allows third-party users to extend existing functionalities. For example, one can add novel diversification models by writing a BEAST2 package that contains the likelihood formula of a phylogeny under the novel diversification model, i.e. the prior probability of a species tree. Many diversification models (and their associated probability algorithms) have been developed, e.g., models in which diversification is time-dependent [Nee *et al.* 1994; Rabosky & Lovette 2008], or diversity-dependent [Etienne *et al.* 2012], or where diversification rates change for specific lineages and their descendants [Etienne & Haegeman 2012; Rabosky 2014; Alfaro *et al.* 2009]. Other models treat speciation as a process that takes time [Rosindell *et al.* 2010; Etienne & Rosindell 2012; Lambert *et al.* 2015], or where diversification rate depends on one or more traits [Maddison *et al.* 2007; FitzJohn 2012]. Despite such a rich theoretical landscape, only a

few of these diversification models are available as tree priors in BEAST2.

When a novel diversification model is introduced, its performance in in-
ference should be tested. Part of a model's performance is its ability to re-
cover parameters from simulated data with known parameters (e.g. [Etienne
et al. 2014]), where ideally the estimated parameter values closely match the
known/true values. Even when a diversification model passes this procedure,
it is not necessarily used as tree prior in Bayesian inference. Bayesian phy-
logenetic inference often requires that the prior probability of the phylogeny
according to the diversification model has to be computed millions of times.
Therefore, biologically interesting but computationally expensive tree priors are
often not implemented, and simpler priors are used instead. This is not neces-
sarily problematic, when the data are very informative, as this will reduce the
influence of the tree prior. However, the assumption that tree prior choice is of
low importance must first be verified.

There have been multiple attempts to investigate the importance of tree
prior choice. For example, Sarver and colleagues, [Sarver *et al.* 2019] showed
that the choice of tree prior does not substantially affect phylogenetic inferences
of diversification rates. However, they only compared current diversification
models to one another, and thus this does not inform us on the importance of
a new tree prior.

Here we introduce a method to quantify the importance of a novel tree
prior. The method starts with a phylogeny generated by the new model. Next,
nucleotide sequences are simulated that follow the evolutionary history of the
given phylogeny. Then, using BEAST2's built-in tree priors, a Bayesian poste-
rior distribution of phylogenies is inferred. We then compare the inferred with
the original phylogenies. How to properly perform this comparison forms the
heart of our method. Only new diversification models that result in a large

discrepancy between inferred and simulated phylogenies will be worth the effort and computational burden to implement a species tree prior for in a Bayesian framework.

Our method is programmed as an R package [R Core Team 2013] called `pirouette`. `pirouette` is built on `babette` [Bilderbeek & Etienne 2018], which calls BEAST2 [Bouckaert *et al.* 2019].

2   Description

The goal of `pirouette` is to quantify the importance of a tree prior. It does so by measuring the inference error made for a given reconstructed phylogeny, simulated under a (usually novel) diversification model. We refer to the model that has generated the given tree as the 'generative tree model' p_G. We define the birth-death (BD) model [Nee *et al.* 1994] as the standard tree model, as many (non-standard) tree models have a parameter setting for which they reduce to this model. One such example is the diversity-dependent (DD) diversification model [Etienne & Haegeman 2020; Etienne *et al.* 2012]: the DD model is a BD model with a speciation or extinction rate that depends on the number of species and a clade-level carrying capacity. For a carrying capacity of infinity, the DD model reduces to the BD model. When benchmarking a novel tree model, one will typically construct phylogenies for different combinations of the diversification model's parameters, to assess under which scenarios the inference error cannot be neglected. While we recommend many replicate simulations when assessing a novel tree prior, our examples contain only one replicate, as the goal is to show the workings of `pirouette`, instead of doing an extensive analysis. The supplementary material, however, does show the results of replicated runs under multiple settings.

`pirouette` allows the user to specify a wide variety of custom settings. These

settings can be grouped in macro-sections, according to how they operate in the pipeline. We summarize them in Table 1 and Table 2.

2.1   pirouette's pipeline

The pipeline to assess the error BEAST2 makes in inferring this phylogeny contains the following steps:

1. The user supplies one or (ideally) more phylogenies from a new diversification model.

2. From the given phylogeny an alignment is simulated under a known alignment model A.

3. From this alignment, according to the specified inference conditions C, an inference model I is chosen (which may differ from the generative model).

4. The inference model and the alignment are used to infer a posterior distribution of phylogenies.

5. The phylogenies in the posterior are compared with the given phylogeny to estimate the error made, according to the error measure E specified by the user.

The pipeline is visualized in Fig. 1. There is also the option to generate a 'twin tree', that goes through the same pipeline. The utility of this twin tree will be explained below.

The first step simulates an alignment from the given phylogeny (Fig. 1, 1a $\rightarrow$ 2a). For the sake of clarity, here we will assume the alignment consists of DNA sequences, but one can also use other heritable material such as amino acids. The user must specify a root sequence, a mutation rate and a site model. The root sequence is the DNA sequence of the shared common ancestor of all species,

Sub-argument	Description	Possible values
tree_prior	Macroevolutionary diversification model	BD, CBS, CCP, CEP, Yule
clock_model	Clock for the DNA mutation rates	RLN, strict
site_model	Nucleotide substitution model	GTR, HKY, JC, TN
mutation_rate	Pace at which mutation occurs	mutation_rate $\in \mathbb{R}_{>0}$
root_sequence	DNA sequence at the root of the tree	any combination of a, c, g, t
model_type	Criterion to select an inference model	Generative, Candidate
run_if	Condition under which an inference model is used	Always, Best candidate
do_measure_evidence	Sets whether or not the evidence of the model must be computed	TRUE, FALSE
error_fun	Specifies how to measure the error	nLTT, $|\gamma|$
burn_in_fraction	Specifies the percentage of initial posterior trees to discard	burn_in_fraction $\in [0,1]$

Table 1: Most important parameter options. BD = birth death [Nee *et al.* 1994], CBS = coalescent Bayesian skyline [Drummond *et al.* 2005], CCP = coalescent constant-population, CEP = coalescent exponential-population, Yule = pure birth model [Yule 1925], RLN = relaxed log-normal clock model [Drummond *et al.* 2006], strict = strict clock model [Zuckerkandl & Pauling 1965], GTR = Generalized time-reversible model [Tavaré 1986], HKY = Hasegawa, Kishino and Yano [Hasegawa *et al.* 1985], JC = Jukes and Cantor [Jukes *et al.* 1969], TN = Tamura and Nei [Tamura & Nei 1993].

Symbol	Macro-argument	Description
G	Generative model	The full setting to produce BEAST2 input data. Its core features are the tree prior p_G, the clock model c_G and the site model s_G.
A	Alignment model	Specifies the alignment generation, such as the clock model c_G, site model s_G and root sequence.
X_i	i-th candidate experiment	Full setting for a Bayesian inference. It is made by a candidate inference model I_i and its inference conditions C_i.
I	Inference model	The assumed phylogenetic inference model, of which the main components are the tree prior p_I, assumed clock model c_I and assumed site model s_I.
C	Inference conditions	Conditions under which I is used in the inference. They are composed of the model type, run condition and whether to measure the evidence.
E	Error measure parameters	Errors measurement setup that can be specified providing an error function to measure the difference between the original phylogeny and the inferred posterior. The first iterations of the MCMC chain of the posterior may not be representative and can be discarded using a burn-in fraction.

Table 2: Definitions of terms and relative symbols used in the main text and in Fig 1. To run the pipeline A, X and E must be specified.

Figure 1: `pirouette` pipeline. The pipeline starts from a phylogeny (1a) simulated by the generative tree model p_G. The phylogeny is converted to an alignment (2a) using the generative alignment model $A = (c_G, s_G)$, composed of a clock model and a site model. The user defines one or more experiments. For each candidate experiment X_i (a combination of inference model I_i and condition C_i), if its condition C_i is satisfied (which can depend on the alignment), the corresponding inference model $I = I_i$ is selected to be used in the next step. The inference models (3a) of the selected experiments use the alignment (2a) to each create a Bayesian posterior of (parameter estimates and) phylogenies (4a). Each of the posteriors' trees is compared to the true phylogeny (1a) using the error measure E, resulting in an error distribution (5a). Optionally, for each selected inference model a twin pipeline can be run. A twin phylogeny (1b) can be generated from the original phylogeny (1a) using the twin tree model p_t, selected among standard diversification models; the default option is the standard BD model, with parameters estimated from the original phylogeny. A twin alignment (2b) is then simulated from the twin phylogeny using clock model c_G and site model s_G imported from the generative model. The twin pipeline follows the procedure of the main pipeline, resulting in a twin error distribution (5b).

₁₃₉ and is set to four different equally-sized mononucleotide blocks by default, which

₁₄₀ helps interpreting the resulting alignment. Supported nucleotide substitution

₁₄₁ models (part of a DNA site model) are JC, HKY, TN and GTR (see Table 1 for

₁₄₂ the meaning of these abbreviations). JC is the default nucleotide substitution

₁₄₃ model (NSM), in which the nucleotide substitution rates between all nucleotides

₁₄₄ are equal and constant over time.

₁₄₅ The second step (Fig. 1, 3a) selects one or more inference model(s) I from a

₁₄₆ set of standard inference models $I_1, \ldots, I_n$. For example, if the generative model

₁₄₇ is known and standard, one can specify the inference model to be the same as

₁₄₈ the generative model. If the tree model is unknown or non-standard - which is

₁₄₉ the primary motivation for this paper -, one can pick a standard inference model

₁₅₀ which is considered to be closest to the true tree model. Also, if we want to run

₁₅₁ only the inference model that fits best to an alignment from a set of candidates,

₁₅₂ one can specify these inference models as well (see section 'Candidate models').

₁₅₃ The third step infers the posterior distributions, using the simulated align-

₁₅₄ ment (Fig. 1, 2a → 4a), and the inference models that were selected in the

₁₅₅ previous step (3a). For each selected experiment a posterior distribution is in-

₁₅₆ ferred, using the `babette` [Bilderbeek & Etienne 2018] R package which makes

₁₅₇ use of BEAST2. This step usually takes up most of the pipeline's computation

₁₅₈ time.

₁₅₉ The fourth step quantifies the inference error made. First the burn-in frac-

₁₆₀ tion is removed, i.e. the first phase of the Markov chain Monte Carlo (MCMC)

₁₆₁ run, which samples an unrepresentative part of paraneter and tree space. By

₁₆₂ default, `pirouette` removes the first 10% of the posterior. From the remaining

₁₆₃ posterior, `pirouette` creates an error distribution, by measuring the difference

₁₆₄ between the true tree and each of the posterior trees (Fig. 1, 4a → 5a). The user

₁₆₅ can specify a function to quantify the differences between the true and poste-

 rior trees. By default, the package uses the nLTT statistic [Janzen *et al.* 2015],

which is the absolute difference between the normalized lineages-through-time

plots of two trees. The nLTT statistic is chosen, as it can operate on any two

trees (regardless of their crown ages and number of taxa) and its results have

a clear range from zero to one. This normalized result makes it possible to

compare trees from a distribution of trees from any tree model.

2.2   Twinning

Similar to lab experiments, `pirouette` allows to perform a control measurement,

by use of a process we call twinning. In this context, this control results in

an error distribution that is the baseline error of the pipeline. The difference

between the 'true' and 'twin' error distributions is caused only by the mismatch

between the generative tree model and the assumed tree prior.

The twinning process, T, encompasses two steps: T_1, that generates a 'twin

tree' (Fig. 1, 1b) and T_2, which generates a 'twin alignment' (Fig. 1, 2b). Both

twin tree and alignment will be analyzed in the same way as the true tree and

alignment.

We define a phylogeny τ as the combination of branching times $\vec{t}$ and topol-

ogy ψ, and denote as τ_G the phylogeny produced by a (possibly non-standard)

generative diversification model, having branching times $\vec{t}_G$ and topology ψ_G.

The first step (T_1) of the twinning process creates a tree τ_T with branching

times $\vec{t}_T$ while preserving the original topology ψ_G:

$$\tau_G = (\vec{t}_G, \psi_G) \xrightarrow{T_1} \tau_T = (\vec{t}_T, \psi_G) \tag{1}$$

The default option for the diversification model p_T is the standard BD model. It

is then possible to use the likelihood function L_T for this diversification model

to find the parameters θ_T^* (e.g. speciation and extinction rates, in case of a BD

model) that maximize this likelihood applied to the true tree, conditioned on its number of tips n_G:

$$\max[L_T(\theta_T | \tau_G, n_G)] \rightarrow \theta_T^*. \tag{2}$$

We use θ_T^* to simulate a number $n_T = n_G$ of branching times $\vec{t}_T$ for the twin tree τ_T, under the process p_T, while preserving the topology. We simulate the new branching times using the TESS package [Höhna 2013; Höhna *et al.* 2016b].

The second step (T_2) of the twinning process simulates the twin alignment with the same clock model, site model and mutation rate used to simulate the original alignment. The twin alignment can be simulated in any user-defined way. `pirouette` supplies the option to have it simulated with the same mutation rate as the true alignment. By default, however, not only the same mutation rate is used, but also the total number of mutations matches the true alignment. The total number of mutations is defined as the amount of different nucleotides between the (known) root sequence compared to the sequences at the tips.

2.3 Candidate models

The user has to specify exactly one standard inference model, but may be unsure which one to pick. To account for this, the user can specify a set of candidate inference models. Each of these candidate inference models is run in an initial, relatively short, analysis; the candidate model with the highest evidence (i.e., marginal likelihood) will then be used in another, longer, inference run, resulting in another error distribution. The evidence for an inference model is estimated by nested sampling [Russel *et al.* 2019], using the `NS BEAST2` package.

If twinning is used, a candidate model that has the highest evidence for the twin alignment is also used to create another error distribution.

2.4 Stochasticity caused by simulating phylogenies

Finally, if the goal is to evaluate BEAST2's performance on a non-standard tree model, one must also consider the last source of stochasticity: the different phylogenies a tree model generates. A single phylogeny cannot be considered as fully representative of the model. For this reason multiple phylogenies must be considered. If the number of considered phylogenies is high enough, the comparison between the main pipeline's aggregated error distribution and its twin counterpart leads to a fair evaluation of the new tree model with respect to the baseline error.

3 Usage

We show the usage of `pirouette` on an tree generated by the non-standard diversity-dependent (DD) tree model [Etienne & Haegeman 2020; Etienne *et al.* 2012], which is a BD model with a speciation rate that is dependent on the number of species.

The code to reproduce our results can be found at `https://github.com/richelbilderbeek/pirouette_example_30` and a simplified version is shown here for convenience:

```
library(pirouette)

# Create phylogeny
set.seed(314)
phylogeny <- create_exemplary_dd_tree(
  n_taxa = 6,
  crown_age = 10
)

```

```
232  # Setup pirouette
233  pir_params <- create_std_pir_params(
234     folder_name = "example_30"
235  )
236
237  # Do the runs
238  pir_out <- pir_run(
239     phylogeny = phylogeny ,
240     pir_params = pir_params
241  )
242
243  # Save
244  library(ggplot2)
245  pir_outs <- list()
246  pir_outs[[1]] <- pir_out
247  pir_plots(pir_outs) +
248     ggtitle("pirouette example 30") +
249     ggsave("errors.png", width = 7, height = 7)
250
251  pir_save(
252     phylogeny = phylogeny ,
253     pir_params = pir_params ,
254     pir_out = pir_out ,
255     folder_name = "example_30"
256  )
```

The DD tree that is generated by this code is as shown in Figure 2, which has an arbitrarily chosen crown age of ten time units and six taxa.

Figure 2: The example tree resulting from a diversity-dependent (DD) simulation.

²⁵⁹ Using the default `pirouette` settings, the error distribution shown in Figure
²⁶⁰ 3 is produced.

Figure 3: The inference error made for an assumed generative tree model and best candidate model, compared with the error obtained for the twin tree. The assumed generative tree model is the model assumed to be closest to the actual generative tree model that generated the true tree. The twin distributions show the baseline inference error. Vertical dashed lines show the median error value per distribution.

₂₆₁ In the upper panel of Figure 3, we can see that the error distributions of ₂₆₂ the assumed generative model differ strongly between the true and twin tree.

This difference shows the extent of the mismatch between the actual tree model (which is DD) and the (BD) tree prior used. Because these distributions are distinctively different, the inference error we make when using an incorrect (that is, BD) tree prior on a DD tree is profound.

Comparing the upper and lower panel of Figure 3, we can see that the best candidate model is only slightly better at inferring the true tree, than the assumed generative model, thereby showing that they cannot compensate for the true generative tree model not being among the inference models.

The candidate model that had highest evidence given the simulated alignment, was JC, RLN, BD (see Table 1 for the meaning of these abbreviations). JC was indeed the site model used when simulating the alignments. The RLN clock model is an unexpected choice: the RLN clock model assumes that mutation rates differ between lineages, whereas the alignment was simulated with equal mutation rates for all lineages. The BD model is indeed closer to the DD model than Yule, as both BD and DD model allow for extinction, while Yule does not.

4 Discussion

We showed how to use `pirouette` to quantify the importance of a tree prior in Bayesian phylogenetics, assuming the simplest standard tree model possible. In principle any other standard tree model can be assumed, but we chose to provide the simplest example.

Figure 3 illustrates the primary result of our pipeline: it shows the error distributions for the true tree and the twin tree when either the (assumed) generating model or the best candidate model is used in inference. The clear difference between the error distributions for the true tree and the twin tree suggests that the choice of tree prior does matter. We note, however, that only

one tree from a novel tree model is not enough to determine the impact of using an incorrect tree prior. Instead, a distribution of multiple trees, generated by the novel tree model, should be used. In the supplementary material we have provided some examples.

Like most phylogenetic experiments, the setup of `pirouette` involves many choices. A prime example is the length of the simulated DNA sequence. One expects that the inference error decreases for longer DNA sequences. We investigated this superficially and confirmed this prediction (see the supplementary materials). However, we note that for longer DNA sequences, the assumption of constant substitution rates may become less realistic and hence longer sequences may require more parameters. Hence, simply getting longer sequences will not always lead to a drastic reduction of the influence of the species tree prior. Fortunately, `pirouette` provides a pipeline that works for all choices.

Interpreting the results of `pirouette` is up to the user; `pirouette` does not answer the question whether the inference error is too large to trust the inferred tree. The user is encouraged to use different statistics to measure the error. The nLTT statistic is a promising starting point, as it can compare any two trees and results in an error distribution of known range, but one may also explore other statistics. In principle, `pirouette` allows for this, but in our example we used a diversification model (DD) that only deviates from the Yule and BD models in the temporal branching pattern, not in the topology.

As noted in the introduction, Duchêne and colleagues [Duchene *et al.* 2018], also developed a method to assess the adequacy of a tree model on empirical trees. They simulate trees from the posterior distribution of the parameters and then compare this to the originally inferred tree using tree statistics, to determine whether the assumed tree model in inference indeed generates the tree as inferred. This is useful if these trees match, but when they do not, this does not

mean that the inferred tree is incorrect; if sufficient data is available the species tree prior may not be important, and hence the inference may be adequate even though the assumed species tree prior is not. In short, the approach is applied to empirical trees and compares the posterior and prior distribution of trees (with the latter generated with the posterior parameters!). `pirouette` aims to identify when assuming standard priors for the species tree leads to incorrect inference if one believes more complex diversification models are operating than can be currently accommodated in inference. In short, our approach applies to simulated trees and compares the posterior distributions of trees generated with a standard and non-standard model, but inferred with a standard one. The two methods therefore complement one another.

However, we note that the `pirouette` pipeline is not restricted to exploring the effects of a new species tree model. The pipeline can also be used to explore the effects of non-standard clock or site models, such as relaxed clock models with a non-standard distribution, correlated substitutions on sister lineages, or elevated substitutions rates during speciation events. It is, however, beyond the scope of this paper to discuss all these options in more detail.

In conclusion, `pirouette` can show the errors to be expected when the model assumed in inference is different from the actual generative model. The user can then judge whether or not this new model should be implemented in our Bayesian phylogenetic tool.

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6 Data Accessibility

All code is archived at `http://github.com/richelbilderbeek/pirouette_article`, with DOI `https://doi.org/12.3456/zenodo.1234567`.

7 Author contributions

RJCB, GL and RSE conceived the idea for the package. RJCB created, tested and revised the package. GL provided major contributions to the package. RJCB wrote the first draft of the manuscript, GL and RSE contributed to revisions.

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