Persistence is an optimal hedging strategy for bacteria in volatile environments

Alexander P Browning[a,b,1], Jesse A Sharp[a,b], Tarunendu Mapder[a,b], Christopher M Baker[a], Kevin Burrage[a,b,c], and Matthew J Simpson[a]

[a] *School of Mathematical Sciences, Queensland University of Technology, Australia*
[b] *ARC Centre of Excellence for Mathematical and Statistical Frontiers, QUT, Australia*
[c] *Department of Computer Science, University of Oxford, UK*

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Abstract

Bacteria invest in a slow-growing subpopulation, called *persisters*, to ensure survival in the face of uncertainty. This hedging strategy, which we term *cellular hedging*, is remarkably similar to financial hedging where diversifying an investment portfolio protects against economic uncertainty. In this work, we provide a new foundation for understanding cellular hedging by unifying the study of biological population dynamics and the mathematics of financial risk management. Our approach explicitly incorporates environmental volatility as a stochastic process, and we seek to find the persister strategy that maximizes the expected per-capita growth rate by formulating a stochastic optimal control problem. The analysis demonstrates that investing in persister production is only advantageous in the presence of environmental volatility, suggesting a stochastic model is essential to elucidate the phenomenon. Analytical and simulation results probe the optimal persister strategy, revealing results that are consistent with experimental observations and suggest new opportunities for experimental investigation. Overall, we provide a new way of modeling cellular decision making by unifying previously disparate theory from mathematical biology and finance.

Keywords: persisters, bet hedging, optimal control, financial mathematics, antibiotic resistance, stochastic growth

Shortly after the clinical introduction of penicillin, Bigger noticed a resistance in a small subpopulation of *Staphylococcal pyogenes* [1]. The resistant cells, termed *persisters*, are a genetically identical, slow-growing, phenotypic variant. Bacteria invest in persisters to ensure survival: persisters are less proliferative than regular cells in a nutrient rich environment but can withstand adversity [2]. These strategies are akin to financial hedging, where diversifying an investment portfolio protects against economic uncertainty [3–5]. This type of strategy in biology and ecology is commonly referred to as *bet-hedging* [6].

Bacterial persistence poses a significant clinical challenge and is highly advantageous to bacteria. For example, persisters can be resistant to antibiotic treatment [7, 8], are undetectable in routine clinical tests [9], and are thought to be responsible for the incurability of many infections [10]. Antimicrobial treatments will, therefore, benefit from an understanding of how persisters arise and function [11]. The similarity between bacterial persistence and financial strategies suggests a new pathway to investigate persister dynamics. In this study, we provide a novel, quantitive understanding of persister strategies using techniques from financial mathematics and stochastic optimal control theory. Persister production is known to depend on the environment [7, 12, 13] and our mathematical modeling approach allows us to unearth how various optimal persister strategies depend upon environmental volatility.

Persisters can be revealed experimentally by disinfecting a population of *Escherichia coli* (*E. coli*) with ampicillin, which targets proliferative cells [7]. The initially rapid reduction in colony size eventually slows, revealing a small subpopulation that is less sensitive to the treatment. Experimental investigations are complicated by the extreme scarcity of persister cells: typically less than 1 in 10^5 cells are persisters in wild type *E. coli* [14]. For subpopulations of this size, stochastic effects are significant [15, 16].

[1] Corresponding author. E-mail: ap.browning@qut.edu.au

In producing persisters, bacteria allocate resources to hedge against environmental volatility for the purpose of survival. Our hypothesis is that these processes occur in much the same way that a financial investor hedges a portfolio to protect from, and take advantage of, economic volatility. The archetypal example of portfolio diversification is Merton's portfolio problem (MPP) [3]. Here, an investor allocates a fraction of their wealth in a high-yield volatile asset, such as stocks; and a low-yield stable asset, such as government bonds. Stochastic optimal control theory [17] is used to maximize the investors wealth by modeling the proportion allocated to each asset as a *control*. Merton's work reveals that it is not advantageous to possess the low-yield asset in the absence of uncertainty, or when the growth of the high-yield asset is deterministic. MPP revolutionized the field of mathematical finance, and many of the ideas in MPP formed the basis of Merton's later work on options pricing [18, 19] that led to the the 1997 Nobel Prize in Economics.

In this work, we develop a new framework for studying cellular hedging by unifying two previously disparate fields: biological population dynamics and financial mathematics. We describe bacteria growth using a stochastic model that explicitly incorporates environmental volatility. Previous mathematical studies have successfully explained why organisms that employ bet-hedging strategies like bacterial persistence have an advantage over those that do not [4–6, 20]. We apply stochastic optimal control theory [17, 21] to probe the persister strategy that maximizes the per-capita growth of a bacteria colony under various types of stochastic environment. A fundamental result of our study indicates that investing in persisters is only advantageous in the presence of environmental volatility, suggesting study of bacterial persistence warrants a stochastic model. Our new model and approach leads to mathematical results that are consistent with observations from several key experimental studies, and provides new insights into bacteria dynamics in the face of environmental volatility.

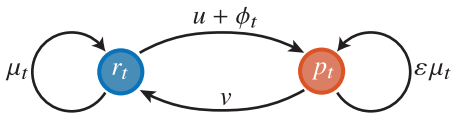

Fig. 1. Schematic of the bacteria growth model with regular cells, r_t, and persisters, p_t. Switching from persister to regular cells is constant, v. Switching from regular cells to persisters is taken to be the sum of a constant rate, u, and a variable rate, ϕ_t, that depends upon environmental volatility.

Stochastic model of bacteria growth

To capture intrinsic noise associated with small subpopulation sizes, we model bacteria dynamics with a system of Itô stochastic differential equations (SDEs) driven by Wiener noise [16, 17]. This choice of model can be thought of as a bridge between discrete Markov models [15], and continuous ordinary differential equation (ODE) models [7]. In addition, we model the dynamics of bacteria in a volatile environment by coupling the growth rate to a stochastic process representing the environment [22].

We model the population of regular (non-persister) cells, r_t, and persisters, p_t. Here, a subscript t indicates that each stochastic processes depends on time, t. The growth of regular cells is described by an Itô SDE such that the net growth rate, μ_t, is subject to stochastic fluctuations characterized by a Wiener process scaled by an intensity σ [16]. The quiescence of persisters manifests slow metabolic activity, so we assume the growth rate of persisters is some small proportion, $\varepsilon < 1$, that of regular cells. The net persister growth rate is subject to stochastic fluctuations characterized by a Wiener process scaled by an intensity $\eta = \varepsilon\sigma$.

The current literature classifies two types persister production in a population: variable and environment dependent, commonly referred to as Type I; or constant, commonly referred to as Type II [7] (Fig. 1). It is understood that constant production of persisters, which results in an approximately constant proportion in a growing population, may be regulated by stochastic gene fluctuation on a single cell level [23–25]. In addition, a growth feedback mechanism [26] — possibly regulated by quorum sensing [27] and intracellular signalling [28] — may enable cells to respond and vary the persister production rate. In our model, regular cells switch to persisters at a rate of $u+\phi_t$, where: $u \geq 0$ is the constant rate [7]; and $\phi_t > 0$ is the variable, environment-dependent, rate. Persisters revert to regular cells at a constant rate $v \geq 0$. Cells in our model, therefore, may use both Type I and Type II strategies.

These dynamics give rise to a system of Itô SDEs

$$\mathrm{d}r_t = \mu_t r_t\,\mathrm{d}t + \sigma r_t\,\mathrm{d}W_t^{(1)} - \big[(u+\phi_t)r_t - v p_t\big]\,\mathrm{d}t\,, \quad (1a)$$

$$\mathrm{d}p_t = \underbrace{\varepsilon\mu_t p_t\,\mathrm{d}t + \eta p_t\,\mathrm{d}W_t^{(2)}}_{\text{Subpopulation growth}} + \underbrace{\big[(u+\phi_t)r_t - v p_t\big]\,\mathrm{d}t}_{\text{Phenotype switching}}\,. \quad (1b)$$

Here, $W_t^{(1)}$ and $W_t^{(2)}$ are two independent Wiener processes that satisfy $W_{t+h}^{(i)} - W_t^{(i)} \sim \mathcal{N}(0,h)$, $h > 0$, $i = 1, 2$ where $\mathcal{N}(0,h)$ denotes a normal distribution with mean zero and variance h. Such a system of equations is said to be in the form of multiplicative Wiener noise [21]. We note that in removing variability from the model by setting $\sigma = \eta = 0$, we recover the ODE model of Balaban *et al.* [7].

We find it natural to consider the variable transformation

$$n_t = r_t + p_t, \quad \theta_t = p_t \,/\, n_t, \quad (2)$$

such that n_t represents the colony size and θ_t represents the proportion of persisters. Following Itô's lemma [17], the transformed state equations are

$$\begin{aligned}\mathrm{d}n_t &= \big[1-\theta_t(1-\varepsilon)\big]\mu_t n_t\,\mathrm{d}t \quad (3a)\\ &\quad + \sigma n_t(1-\theta_t)\,\mathrm{d}W_t^{(1)} + \eta n_t\theta_t\,\mathrm{d}W_t^{(2)}\,,\\ \mathrm{d}\theta_t &= \big[(1-\theta_t)(u+\phi_t) - \theta_t v - (1-\theta_t)\theta_t\big((1-\varepsilon)\mu_t \quad (3b)\\ &\quad - \sigma^2(1-\theta_t) + \eta^2\theta_t\big)\big]\,\mathrm{d}t\\ &\quad - (1-\theta_t)\theta_t\sigma\,\mathrm{d}W_t^{(1)} + \eta(1-\theta_t)\theta_t\,\mathrm{d}W_t^{(2)}\,,\end{aligned}$$

revealing that the dynamics of the persister proportion, θ_t, are independent of the colony size, n_t.

Stochastic model environmental volatility. We model environmental volatility by assuming $\mu_t = m(\zeta_t)$, where ζ_t is a stochastic process that represents a volatile environment. There are many appropriate choices for $m(\cdot)$ and ζ_t, but we focus our analysis on three environments, samples paths of each shown in Fig. 2*a*–*c*. These are

1. *Constant.* (Fig. 2*a*) We set $m(\zeta) \in \{\mu_G, \mu_S\}$, such that μ_t is constant. Here μ_G represents a colony during growth; and μ_S represents a colony under stress or specifically, antimicrobial treatment. For numerical results, we choose $\mu_G = 2\,\mathrm{h}^{-1}$ and $\mu_S = -2\,\mathrm{h}^{-1}$ to match experimental data for *E. coli* during growth and ampicillin treatment [7].

2. *Monod.* (Fig. 2*b*) Monod kinetics are commonly used to model the growth of bacteria [29–31], and feature dynamics with an asymptotic upper bound on the growth rate, but no lower bound. We describe this environment by a mean-reverting Ornstein-Uhlenbeck process [17] and couple the growth rate using a Monod equation [29],

$$m(\zeta_t) = \frac{\mu_{\max}\zeta_t}{K_\zeta + \zeta_t} - \delta, \quad (4a)$$

$$\mathrm{d}\zeta_t = \xi(K_\zeta - \zeta_t)\,\mathrm{d}t + \kappa\,\mathrm{d}W_t^{(3)}\,, \quad (4b)$$

where $\mathrm{d}W_t^{(3)}$ is a Wiener process independent of both $\mathrm{d}W_t^{(1)}$ and $\mathrm{d}W_t^{(2)}$. Here, ζ_t experiences fluctuations proportional to κ and a reversion force to the state $\zeta_t = K_\zeta$ of strength proportional to ξ. The Monod coupling features an asymptotic upper-bound on the growth rate of $\mu_t = \mu_{\max}$ and no lower bound. In effect, unfavourable environmental changes have a larger effect on the growth rate than those that are favorable, and the growth rate experiences reversion to a growth rate $\mu_t = \mu_{\max}/2 - \delta$, where δ represents the natural death rate. For numerical results, we choose $\xi = 0.1\,\mathrm{h}^{-1}$, $K_\zeta = 1$, $\zeta_0 = 1$, $\kappa = 0.3$, $\mu_{\max} = 8\,\mathrm{h}^{-1}$ and $\delta = 2\,\mathrm{h}^{-1}$. To deal with the discontinuity at $\zeta = -K_\zeta$, we truncate Eq. 4a so that $m(\zeta) = m(-0.5)$ for $\zeta < -0.5$.

3. *Poisson.* (Fig. 2*c*) Kussell *et al.* [15] model environmental variability by assuming alternating periods of growth and stress, of durations τ_G and τ_S, respectively. We reproduce this type of environment in a stochastic model by assuming that the environment switches between growth and stress according to a Poisson process. We consider that $m(\zeta) = \mu_G$ for $\zeta \geq 0$; $m(\zeta) = \mu_S$ for $\zeta < 0$; and model the environment as the Poisson process

$$\mathrm{d}\zeta_t = \rho(\zeta_t)\,\mathrm{d}P_t\,, \quad \mathrm{d}P_t \sim \mathrm{Po}(\mathrm{d}t\,\lambda(\zeta_t)), \quad (5a)$$

$$\text{where } \rho(\zeta_t) = \begin{cases} -2, & \zeta_t = +1,\\ +2, & \zeta_t = -1,\end{cases} \quad (5b)$$

$$\text{and } \lambda(\zeta_t) = \begin{cases} 1/\tau_G, & \zeta_t = +1,\\ 1/\tau_S, & \zeta_t = -1.\end{cases} \quad (5c)$$

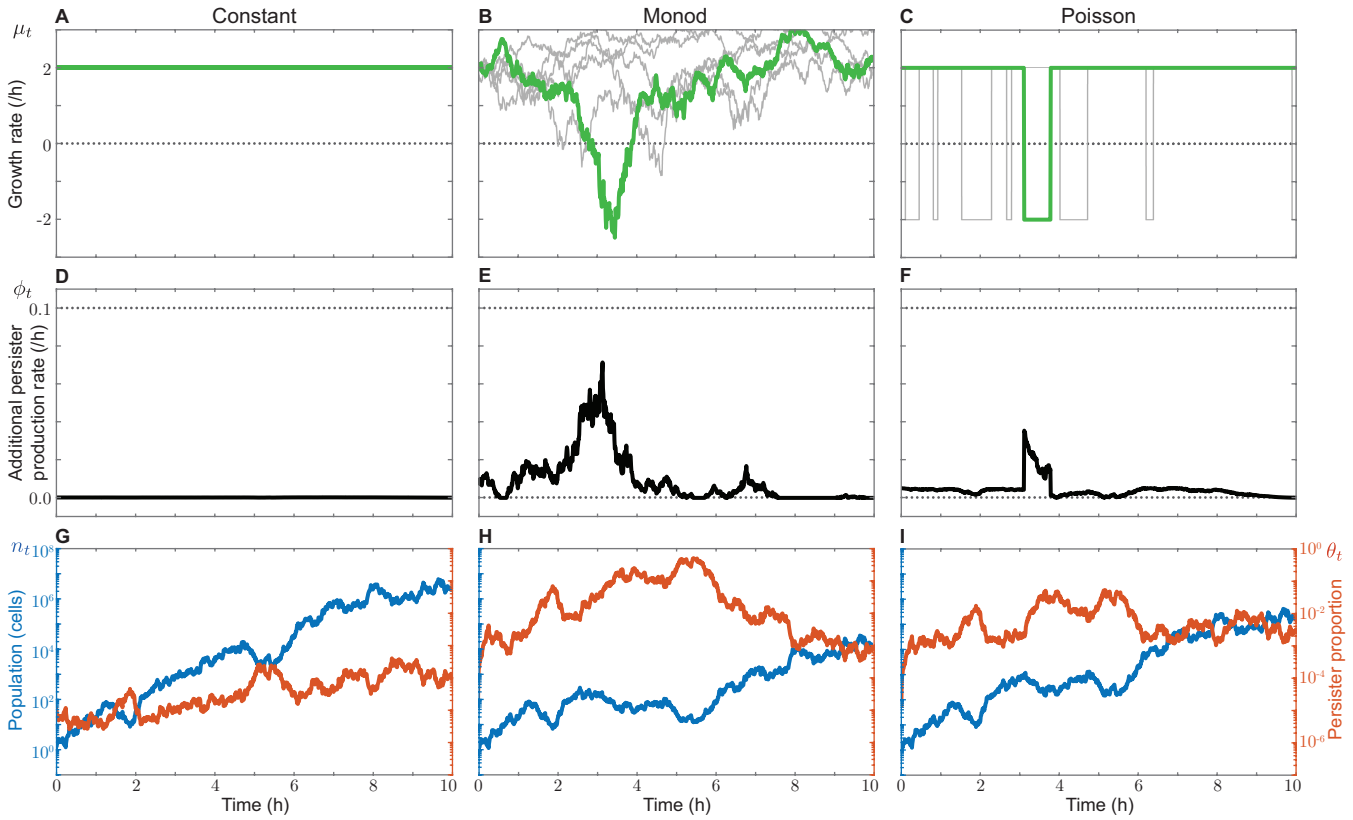

Fig. 2. Optimal persister production in cell populations under various types of environment. (a)-(c) Growth rates, μ_t, sampled from each environment to simulate bacteria growth in the rest of the figure. Also shown are and five additional, independent, realisations (light grey); (d)-(f) The variable persister production rate, ϕ_t^*, for a single realisation of the model. (g)-(i) The population, n_t, (blue, left scale) and persister proportion, θ_t, (red, right scale), for a single realisation of the model. The seeds used to generate the Wiener processes $W^{(1)}$ and $W^{(2)}$ are identical for all environments.

This formulation allows for a mean time of τ_G in the growth phase, where $\mu_t = \mu_G$, and a mean time of τ_S in the stress phase, where $\mu_t = \mu_S$. For numerical results in this study, we choose $\tau_G = 9.5\,\mathrm{h}$, $\tau_S = 0.5\,\mathrm{h}$, $\zeta_0 = 1$, $\mu_G = 2\,\mathrm{h}^{-1}$ and $\mu_S = -2\,\mathrm{h}^{-1}$.

These choices represent environments in which changes happen gradually (Monod environment) or abruptly (Poisson environment), as demonstrated in Fig. 2*b* and Fig. 2*c*, respectively. For comparative purposes we choose similar realizations of each environment and choose the initial condition for all environments such that $\mu_0 = \mu_G = 2\,\mathrm{h}^{-1}$. In the supporting material, we explore results for two other models of environmental volatility: (4) an Ornstein-Uhlenbeck process without a Monod coupling (Fig. S1d) and, (5) a Duffing oscillator (Fig. S1e).

Optimal persister production strategies

Our ideas of *cellular hedging* suggest that a cell population chooses a persister production strategy $\mathcal{S} = \{u, v, \phi_t\}$ that is optimal in some way. Mathematically, we define optimality as the strategy that maximizes some *fitness measure*, which we now construct. We assume that there is no explicit cost to producing persisters with constant rates u and v; but that there is a quadratic running cost to produce persisters with a variable rate ϕ_t. The cost of applying a non-zero ϕ_t accounts for the sensing mechanisms that cells must use to respond to the environment.

We choose a fitness measure, commonly referred to as a *payoff* in optimal control theory, as

$$J_{\mathcal{S}} = \mathbb{E}\Big[\underbrace{\int_0^T \alpha\phi_t^2\,\mathrm{d}t}_{\text{Sensing}} + \underbrace{\log_{\mathrm{e}}(n_T)}_{\text{Growth}}\Big],\ \alpha < 0. \tag{6}$$

Here T denotes a terminal time, so that the maximization is carried out on the interval $t \in [0, T]$; and, α characterizes a trade-off between growth and operating the sensing mechanisms required to vary the persister production rate. Maximizing the logarithmic term in Eq. 6 can be interpreted as cells maximizing their per-capita growth rate over the interval $t \in [0, T]$, since

$$\log_{\mathrm{e}}(n_T) = \log_{\mathrm{e}}(n_0) + \int_0^T \frac{1}{n_t}\frac{\mathrm{d}n_t}{\mathrm{d}t}\,\mathrm{d}t\,,$$

and $\log_{\mathrm{e}}(n_0)$ is constant [32]. Our choice of fitness measure corresponds to evolutionary mechanisms that perpetuate highly productive populations [15].

Together with the state equations (Eq. 3), maximizing Eq. 6 corresponds to an optimal control problem. We now review essential elements of Hamilton-Jacobi-Bellman (HJB) optimal control theory.

Optimal control theory. A stochastic control problem may be stated as

$$\max_{\omega_t \in \mathcal{U}} \mathbb{E}\left(\int_0^T \mathcal{L}(\mathbf{x}_t, \omega_t, t)\,\mathrm{d}t + \Phi(\mathbf{x}_T)\right), \tag{7}$$

where ω_t is the control from the set of allowable controls, $\mathcal{U}$. The state equations are given by the q-dimensional stochastic

process, $\mathbf{x}_t$, governed by

$$\mathrm{d}\mathbf{x}_t = \mathbf{M}(\mathbf{x}_t, \omega_t, t)\,\mathrm{d}t + \mathbf{\Sigma}(\mathbf{x}_t, \omega_t, t)\,\mathrm{d}\mathbf{W}_t\,, \tag{8}$$

where $\mathbf{M} \in \mathbb{R}^q$ gives the drift of the process; $\mathrm{d}\mathbf{W}_t \in \mathbb{R}^q$ is a Wiener process of the same dimension as $\mathbf{x}_t$ with independent constituants; and, $\mathbf{\Sigma}\mathbf{\Sigma}^{\mathrm{tr}} \in \mathbb{R}^{q\times q}$ describes the covariance of the Wiener process (superscript $^{\mathrm{tr}}$ denotes the matrix transpose). In this study, we also study a form of $\mathbf{x}_t$ that includes Poisson jump noise, and we include the derivation of the HJB equations for this case in the supporting material.

To solve the optimal control problem we define the value function, $V(\mathbf{x}, s)$, as the optimal payoff obtainable starting at $\mathbf{x}$ at time s and continuing to terminal time T. That is,

$$V(\mathbf{x}, s) = \max_{\omega_t \in \mathcal{U}} \mathbb{E}\left(\int_s^T \mathcal{L}(\mathbf{x}_t, \omega_t, t)\,\mathrm{d}t + \Phi(\mathbf{x}_T)\right), \quad \text{subject to } \mathbf{x}_s = \mathbf{x}. \tag{9}$$

Hamilton-Jacobi-Bellman (HJB) theory describes V as the solution of a partial differential equation (PDE) [17,21], given by

$$0 = \max_{\omega_t \in \mathcal{U}} \left(\mathcal{A}V + \mathcal{L}(\mathbf{x}_t, \omega_t, t)\right), \tag{10}$$

where $\mathcal{A}$ is the stochastic generator for the process governing $\mathbf{x}_t$. For the q-dimensional system given by Eq. 8, the stochastic generator is given by

$$\mathcal{A}V = \frac{\partial V}{\partial s} + \sum_{i=1}^{q}\left(\mathbf{M}_i \frac{\partial V}{\partial x_i} + \frac{1}{2}\sum_{j=1}^{q}(\mathbf{\Sigma}\mathbf{\Sigma}^{\mathrm{tr}})_{ij}\frac{\partial^2 V}{\partial x_i \partial x_j}\right) \tag{11}$$

where $\mathbf{M}_i$ denotes the ith element of $\mathbf{M}(\mathbf{x}_t, \omega_t, t)$, and $(\mathbf{\Sigma}\mathbf{\Sigma}^{\mathrm{tr}})_{ij}$ denotes the element in the ith row and jth column of $\mathbf{\Sigma}\mathbf{\Sigma}^{\mathrm{tr}}$, for $\mathbf{\Sigma} = \mathbf{\Sigma}(\mathbf{x}_t, \omega_t, t)$. Substituting the terminal time $s = T$ into Eq. 9 provides the terminal condition $V(\mathbf{x}, T) = \Phi(\mathbf{x}_T)$.

In certain cases, the argmax$(\mathcal{A}V + \mathcal{L})$ term in Eq. 10 may be found analytically through differentiation. This yields both an expression for the optimal control, ω_t^*; and a q-dimensional, non-linear, PDE coupled to a terminal condition. In the case of MPP, Eq. 10 has an analytical solution [3], however in most cases we are required to solve Eq. 10 numerically. To obtain an optimal trajectory starting at state $\mathbf{x}_0$, we solve the state equations (Eq. 8), coupled to the solution of the HJB PDE (Eq. 10), forward in time using the Euler-Maruyama algorithm [33]. Full details of the numerical techniques and code used in this work are provided as supporting material.

Constant persister production. We first examine a cell colony that can only produce persisters at a constant rate, so we fix the variable rate, $\phi_t = 0$. In this case, we only assume the constant environment so $\mu_t = \mu_G$.

As u and v only appear in the optimal control problem linearly, the solution is not finite unless a bound is enforced on u and v [34]. The unbounded problem corresponds to a cell population that is able to move itself instantaneously to anywhere in the state space. To address this, we assume that the constant switching rates u and v are chosen by the population to control the steady-state persister proportion, which we denote $\hat{\theta}_t \in [0, 1]$. Taking the drift term in the state equation for θ_t (Eq. 3b) to be zero, we see that u and v are related to $\hat{\theta}_t$ by

$$u = \hat{\theta}_t\left(\frac{v}{1-\hat{\theta}_t} + \eta^2\hat{\theta}_t - (1-\hat{\theta}_t)\sigma^2 + (1-\varepsilon)\mu_G\right). \tag{12}$$

At this point, we note that Eq. 12 is only consistent if $\hat{\theta}_t$ is constant, and we address this shortly.

Allowing $\hat{\theta}_t$ to be a control, the state equation for n_t (Eq. 3a) becomes

$$\frac{\mathrm{d}n_t}{n_t} = (1-\hat{\theta}_t(1-\varepsilon))\mu_G\,\mathrm{d}t + \sqrt{(1-\hat{\theta}_t)^2\sigma^2 + \hat{\theta}_t^2\eta^2}\,\mathrm{d}W_t\,, \tag{13}$$

since $a\,\mathrm{d}W_t^{(1)} + b\,\mathrm{d}W_t^{(2)}$ can be considered as $\sqrt{a^2+b^2}\,\mathrm{d}W_t$. Assuming that $\hat{\theta}_t$ is chosen to maximize the fitness measure (Eq. 6) reveals a problem closely related to MPP [3]. The problems differ intrinsically in that an investor is able to reallocate their portfolio instantaneously and without cost — though variations of MPP address this — whereas the cell colony consists of finitely many cells that act heterogeneously using finite switching rates to produce persisters.

We apply stochastic optimal control theory [17,21] and exploit the similarity between this formulation of the persister problem and MPP [3] to obtain an analytical solution, in which the optimal control, denoted $\hat{\theta}_t^* \in [0, 1]$, is given by

$$\hat{\theta}_t^* = \max\left[0, \min\left(1, \frac{\sigma^2 - (1-\varepsilon)\mu_G}{\eta^2 + \sigma^2}\right)\right]. \tag{14}$$

Full details of this analytical solution are given in the supporting material. As is the case with MPP, $\hat{\theta}_t^*$ is constant, so the strategy is independent of both the time and system state. Solving Eqs. 12 and 14 simultaneously gives $\hat{\theta}_t^* = 1.19 \times 10^{-5}$ with $\sigma = 1.414222$ which matches experimental observations for wild type *E. coli* bacteria where $\mu_G = 2\,\mathrm{h}^{-1}$, $\varepsilon = 0$, $\eta = \varepsilon\sigma = 0$, $u = 1.2 \times 10^{-6}\,\mathrm{h}^{-1}$ and $v = 0.1\,\mathrm{h}^{-1}$ [7]. For numerical results in the rest of this study, we fix σ, ε, η, u and v to these values, and set the initial persister proportion $\theta_0 = 1.19 \times 10^{-5}$.

Variable persister production. We now consider a cell colony that is able vary persister production in response to their environment. In this case, $\phi_t \in \mathbb{R}^+$, can be thought of as a Markovian control [17], or a control that is chosen based on information that includes the current state and time, but does not carry a memory about the past state. For numerical stability, we place an upper-bound on the control such that $\phi_t \leq 0.1\,\mathrm{h}^{-1}$.

To solve the control problem, we define the value function, V, by

$$V(x, y, z, s) = \max_{\phi_t} \mathbb{E}\left[\int_s^T \alpha\phi_t^2\,\mathrm{d}t + \log_{\mathrm{e}}(n_T)\right], \tag{15a}$$

$$\text{where} \quad (n_s, \theta_s, \zeta_s) = (x, y, z). \tag{15b}$$

HJB optimal control theory describes V by the PDE, given by Eq. 10. In our case, we can express the PDE as

$$0 = \max_{\phi_t}\left(\alpha\phi_t^2 + \frac{\partial V}{\partial y}\phi_t + \mathcal{H}_\star\right), \tag{16}$$

where $\mathcal{H}_\star$ represents a collection of terms independent of the control ϕ_t. As Eq. 16 is quadratic in ϕ_t, we can carry out the maximization by setting the derivative to zero to find the optimal control, denoted $\phi_t^* \in [0, 0.1]$, as

$$\phi_t^* = \max\left[0, \min\left(0.1, -\frac{1}{2\alpha}\frac{\partial V}{\partial y}(1-y)\right)\right]. \tag{17}$$

Here, $y = \theta_t$ is the current persister proportion. Since $y \in [0, 1]$, an interpretation of Eq. 17 is that the population uses ϕ_t^* to steer the population toward the optimal persister proportion, where $\partial V/\partial y = 0$. As environmental triggers only create persisters (the quiescent state of persisters means they are unable to react to the environment), $\phi_t^* > 0$, and so ϕ_t^* is only active if the current proportion is less than the optimal proportion. Finally, $\phi_t^* \to 0$ as $\alpha \to -\infty$, which

represents the sensing mechanisms becoming prohibitively expensive.

Substituting Eq. 17 into the HJB equation leads to a nonlinear PDE for V that must be solved backward in time. Full details of this equation are given in the supporting material. In summary, the variable transformation $x \to \log_e(x)$ removes all terms containing the independent variable x from the equation. Therefore, we find that the ansatz

$$V(x, y, z, s) = \Psi(y, z, s) + \log_e(x), \tag{18}$$

is consistent with the system. This result reveals that the optimal control (Eq. 17) is independent of colony size since $\partial V/\partial y = \partial \Psi/\partial y$. This gives an optimal strategy of the form $\phi_t^* = \phi^*(\theta_t, \zeta_t)$, which the cells may implement using only information about the proportion of persisters (through, for example, quorum-sensing [27] or intracellular signalling [28]), and the environment (through, for example, a growth feedback mechanism [26]).

In Fig. 2 we show dynamics for cell colonies in all three environments, for $\alpha = -100\,\mathrm{h}$. For these results, we solve for Ψ numerically. As expected, we find that unfavorable environments trigger variable persister production (Fig. 2*e*,*f*), without any additional persister production for the constant environment (Fig. 2*d*). This suggests that a constant persister production strategy is sufficient for an environment with a constant expected growth rate. On the other hand, additional persister production is seen in response to environmental cues for more volatile environments (Fig. 2*e*,*f*). An interesting result in Fig. 2*e* is that the peak variable production rate under the Monod environment lies before the minimum growth rate. This suggests that the model implicitly incorporates mechanisms such that cells understand the environment to the extent that they can anticipate environmental changes, perhaps based on past events or sudden changes in the environment.

Behaviour under unfamiliar environments and antimicrobial treatment. By assuming that cells monitor their growth rate, μ_t, we can explore how a persister strategy that is optimal under one type of environment behaves under another, unfamiliar environment. We couple the growth rate to the optimal control by considering $\phi_t^* = \phi^*(\theta_t, \zeta_t)$ where $\zeta_t = m^{-1}(\cdot)$ denotes the inverse of the growth rate coupling function $m(\zeta)$. In other words, the cells measure the current environment state using the growth rate.

Persisters are revealed experimentally by exposing the population to antibiotics and monitoring the number of viable cells [7]. We simulate this process to examine how cells which have evolved under each environment behave when continuously exposed to a constant expected decay rate. Simulation results in Fig. 3*a*, for a population that can only produce persisters at a constant rate, are similar to experimental results for wild-type *E. coli* [7]. In comparison, we find that colonies that implement a strategy optimal under a more volatile environment, such as the Poisson and Monod environments, produce persisters using the variable rate (Fig. 3*c*) reach saturation of persisters more quickly (Fig. 3*b*) and have a higher long term population (Fig. 3*a*).

Discussion

In this work, we model persister dynamics using a stochastic model and introduce the idea of *cellular hedging* to study an optimal persister production strategy. By applying cross-disciplinary ideas from mathematical finance to the persister problem, we provide several analytical and simulation results to elucidate bacterial persistence under different forms of environmental volatility.

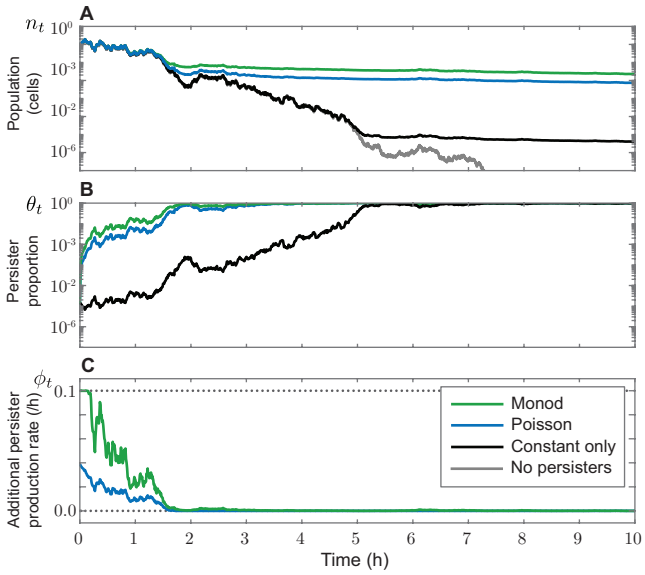

Fig. 3. Single realizations of the model showing the behaviour of a cell population under antibiotic treatment. (a) Shows the population, n_t; (b) shows the persister proportion, θ_t; and, (c) shows the variable persister production rate, ϕ_t^*. In all cases, the cells experience a growth rate $\mu_t = -2\,\mathrm{h}^{-1}$ and implement: the variable strategy optimal under either the Monod environment (green) or the Poisson environment (red); constant switching only optimal under the constant environment (black); or do not produce persisters (grey).

Our model suggests that it is detrimental to produce persisters in the absence of volatility. In examining the optimal strategy for constant persister production, we reveal the proportion of persisters that can be maintained to maximize the expected growth of the population (Eq. 14). Removing volatility from the model by using constant expected growth rate and setting $\sigma = 0$, recovers the deterministic model of Balaban *et al.* [7] and reveals an optimal persister proportion of zero. Therefore, our model predicts that producing persisters in the absence of environmental volatility is suboptimal and, therefore, leads to a lower expected per-capita growth rate. In contrast, our straightforward assumption of Wiener noise in the growth rate reveals a small proportion of persisters (approximately 1 in 10^5) — consistent with experimental observations of Balaban *et al.* [7] — that maximizes the expected per-capita growth rate. This result contrasts with previous mathematical studies that assume external factors, such as antibiotic treatment, to explain persistence [15].

Compared to financial hedging, cellular hedging differs significantly in the complexity of the strategy that can be implemented. A key result is our direct comparison of bacterial persistence and MPP revealing a constant optimal proportion of persisters that should be maintained in an environment with a constant expected growth rate. This result is significant for the persister problem as, unlike in MPP, the cell population cannot directly control the proportion of persisters. However, the population can maintain a constant *expected* population, regulated by constant switching propensities, u and v.

The mechanisms that enable cells to implement cellular hedging strategies that respond to the environment are unknown [35], but they must come at a cost to the cells in terms of additional genetic machinery and rely on limited information available to individual cells. Our study reveals an optimal variable persister production strategy that depends only on the time, persister proportion and the environment, but not on the colony size. Additional results in the

supporting material (Fig. S6) also indicate that these strategies become time-independent far from the terminal time T. Furthermore, the strategy itself is not complex. Rather, cells increase persisters production when the persister proportion for a given growth rate is less than optimal. These results are surprising as optimal control theory only provides the strategy that maximizes the payoff, and does not necessarily enforce any level of complexity in the cellular hedging strategy.

We demonstrate the distinct advantage persisters afford bacteria against antimicrobial treatment (Fig. 3) [7, 9, 11]. Our stochastic model of persister production could be used to improve the efficacy of antimicrobial treatments using optimal control from an optimal treatment perspective [34,36]. Results from our modeling framework imply new ways to design treatment strategies that could be verified experimentally. Additional results (Fig. S6) suggest that ϕ_t decreases monotonically with the growth rate, for all environments considered. Temporarily exposing the cell colony to favorable conditions may decrease the variable persister production rate predicted by the model, lowering the proportion of persisters and thereby making the population more susceptible to antibiotics. This is consistent with experimental observations where exposing a colony to a fresh growth medium before applying antibiotics decreases persistence [37].

Rapid evolution of cellular hedging strategies is experimentally reproducible [38]: Van den Bergh *et al.* [9] found that exposing *E. coli* to daily antibiotic treatments increased survival by between three and 300-fold after just three treatments, and Rodriguez-Beltran *et al.* [39] study induced evolutionary changes in bacteria behaviour through repeated application of antibiotics. Our modeling framework offers an opportunity to investigate an emergent persister strategy from repeated exposure of bacteria to a fluctuating environment. Additionally, financial mathematics techniques may be applied to study the evolutionary process as it occurs. For example, policy adjustment models [40] quantify the cost of strategy change, and can be compared to mutation costs in a rapidly-evolving cell colony.

Conclusions

We provide new insight into cellular hedging by unifying the study of biological population dynamics and the mathematics of financial risk management. A fundamental result of our study is that investing in persisters is only advantageous in the presence of environmental volatility, suggesting that the study of this phenomenon must take a stochastic perspective. We present a new stochastic model of bacteria growth in a volatile environment and apply optimal control theory to probe the persister strategy that maximizes the per-capita growth rate. The framework we develop offers an opportunity for future generalizations to explore cellular decision making in a broader context. Many seemingly complex cellular phenomena from bet-hedging in cancers [20,41], herpes viruses [42] and HIV [43] to decision making in the epithelial-mesenchymal transition [44] could be modeled using ideas from cellular hedging and mathematical finance.

Materials and Methods

We integrate the SDE models using the Euler-Maruyama algorithm [33] with time step $h = 2 \times 10^{-3}$. To compare results between environments, we fix the seeds used to generate the Weiner processes $W_t^{(1)}$ and $W_t^{(2)}$ when simulating different models.

Code used to produce the numerical results is available on GitHub at `github.com/ap-browning/persisters`. Full details of the numerical methods, and the full form of the HJB equation for each environment, are given in the supporting material.

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