# Title

The open bar is closed: restructuration of a native parasitoid community following successful control of an invasive pest.

# Authors

David Muru[1], Nicolas Borowiec[1], Marcel Thaon[1], Nicolas Ris[1], Elodie Vercken[1].
Corresponding author: david.muru@inrae.fr

# Affiliations

[1]Institut Sophia Agrobiotech - INRAE, Université Côte d'Azur, CNRS, ISA, France.

# Keywords

*Dryocosmus kuriphilus*, *Torymus sinensis*, community structure, biocontrol, co-occurrence, niche displacement, exploitative competition, native parasitoids.

# Abstract

The rise of the Asian chestnut gall wasp *Dryocosmus kuriphilus* in France has benefited the native community of parasitoids originally associated with oak gall wasps by becoming a trophic subsidy. However, the successful biological control of this pest has then led to significant decreases in populations densities. Here we investigate how the invasion of the Asian chestnut gall wasp *Dryocosmus kuriphilus* in France and its subsequent control by the exotic parasitoid *Torymus sinensis* has impacted the newly-formed community of native parasitoids.

We explored multi-year native community dynamics during the rise and fall of the invasive pest. Community dynamics were monitored in 26 locations during 5 years after the *T. sinensis* introduction. In an attempt to understand how some mechanisms such as local extinction or competition come into play, we analyzed how the patterns of co-occurrence between the different native parasitoid species changed through time.

Our results demonstrate that native parasitoid communities experienced increased competition as the *D. kuriphilus* levels of infestation decreased. During the last year of the survey, two alternative patterns were observed depending on the sampled location: either native parasitoid communities were almost inexistent, or they were dominated by one main parasitoid: *Mesopolobus sericeus*. We observe that the two patterns correlate with the habitat and that they may be explained by environmental features such as differences in the natural reservoirs for native parasitoids. These results highlight how the "boom-and-bust" dynamics of an invasive pest followed by successful biological control can deeply alter the structure of native communities of natural enemies.

## Introduction

Biological invasions are defined as the introduction, establishment and expansion of populations outside of their native area. Apart from their well-known effects on agricultural production (see Mc Michael and Bouma 2000 for a review), biological invasions are also identified as major drivers of global changes in biodiversity worldwide. More generally, invasions are known to have a diversity of direct and indirect negative effects on native ecosystems (see McGeoch et al 2015 for a review of environmental impacts caused by invasion). In particular, they can deeply alter interspecific interactions and restructure native communities, often with negative consequences (Ricciardi and Isaac 2000, Ricciardi 2001, Carroll 2007). For instance, invasive predators, parasitoids or pathogens were proven to drastically reduce the size of resident prey or host populations (Daszak et al 2000) or to compete with other species from the same trophic level (Hamilton et al 1999, Grosholz 2002, see David et al 2017 for a review of the impacts of invasive species on food webs).

While much research has focused on invasive top-consumers (predators, parasitoids, etc), a smaller amount of literature examines the impact of primary-consumer invasive species as a new resource for the native community (Carlsson et al 2009). The situation where a successful invasive species becomes a resource for native species of a higher trophic level is referred to as a form of facilitation and the invasive species acts as a trophic subsidy (Rodriguez 2006). Examples of trophic subsidies include some invasive macroalgae (Olabarria et al 2009, Rossi et al 2010, Suarez-Jimenez et al 2017), some phytophagous insects (Barber et al 2008, Girardoz et al. 2008, Jones et al 2014, Haye et al 2015, Herlihy et al 2016, Noyes 2017), *Drosophila suzukii* (Mazzetto et al 2016) or four invasive gall wasp species (Schonrogge and Crawley 2000). Hence, trophic subsidies, possibly in pair with changes in population densities of local populations (Eveleigh et al. 2007) may change the established dynamics within recipient communities by creating, for example, apparent competition (Settle and Wilson 1990, Holt and Bonsall 2017) or niche displacement (Mallon et al 2007). Even when the invasion is only transient and therefore the trophic subsidy finally disappears, the consequences on native communities can be lasting on the recipient community. Indeed, recovery from the removal of the alien species is not systematic and may thus lead to further disequilibrium within the community (Courchamp et al. 2003). For example, Mallon et al. (2017) have recently reported a permanent niche displacement of native species caused by a failed invasion by *Escherichia coli* in soil microcosms, and referred to it as a "legacy effect". Such lasting effects of transient invasions observed by Mallon et al. (2017), for instance on niche breadth or space, may occur as well for community structure, in terms of overall community restructuration (e.g. forcing species to modify the way they exploit available resources). However, empirical data on the response of native community dynamics to transient invasion remain scarce, as there is a lack of studies exploring multi-year community dynamics during the rise and fall of an invasive pest.

Classical biological control – i.e. the deliberate introduction of an exotic biological control agent to durably regulate a target (usually exotic) pest (Eilenberg et al. 2001) - can provide valuable empirical data on the dynamics of communities disturbed by two successive invaders, the pest and its introduced natural enemy. Indeed, during a biological control program, the dynamics of local communities are disturbed twice consecutively. Firstly, the arrival of the invasive pest decreases the level of primary resource and can alter

the abundances of its native competitors and natural enemies (Jones et al 2014, Haye et al 2015, Herlihy et al 2016). Then, the establishment of the biological control agent will again modify the community structure by strongly reducing the abundance of the invasive pest and potentially interacting with native species. Long term direct and indirect impacts of either the pest or its natural enemy on the recipient community have been documented (Henneman et Memmott 2001, also see Louda et al 2003 for a review of 10 case studies with quantitative data). Interactions such as those existing between parasitoids and their hosts may also be impacted by the dynamics inherent to classical biological control. If they are able to, native non-specialist parasitoids may be displaced from their native hosts to an invasive one. However, the native community of parasitoids can be outcompeted from the exotic pest by the exotic parasitoid (Naranjo 2017) that has been chosen for its efficiency exploiting the exotic pest. Firstly, this could lead to local extinction of native populations of parasitoids, especially if the exotic parasitoid outcompetes them on the native host(s) as well (Bengtsson 1989). On the other hand, the introduction of an exotic parasitoid to control an exotic pest often leads to a displacement of the native community of parasitoids that have become associated with the exotic pest (Bennett 1993, Lynch and Thomas 2000, van Lenteren et al. 2006). This happens logically when the introduced parasitoid is specialized on the exotic pest and is a superior competitor or more adapted to find and exploit the pest than its native counterparts (Naranjo 2017). The resulting displacement might only be a step backwards, bringing the system back to the previous pattern of host-parasitoid dynamics (before the pest invaded the area), or a novel state might emerge, depending on the resilience of the native species.

However, the temporal dynamics and spatial variability of these processes remain poorly understood and empirical data are greatly lacking at this point with, to our knowledge, no reports of such non-intentional effect in the context of biological control. Therefore, here we use successful classical biological control of an invasive pest as a framework to properly investigate how these two subsequent invasions impact the structure of native communities.

The Asian chestnut gall wasp *Dryocosmus kuriphilus* (Hymenoptera:Cynipidae), native to China, was accidentally introduced in Italy in 2002 (Brussino et al 2002) and is now distributed throughout Italy and other European countries (EPPO, 2014). *D. kuriphilus* is a specialist attacking only chestnut trees. In absence of competitors (Bernardo et al., 2013) and specialized antagonists, D. *kuriphilus* was able to proliferate quickly and massively. Therefore, it became a trophic subsidy for several native parasitoids previously associated to gall wasps from other plants/trees (Matosevic & Melika 2013, Panzavolta et al. 2013, Francati et al 2015, Noyes 2019). In response to damage observed on chestnut production and also apiculture, classical biological control programs were quickly implemented in newly infested countries. *Torymus sinensis* was chosen as a biological control agent due to its high specificity of parasitism (Quacchia et al 2014, Ferracini et al 2017) and its previous effective control of the target pest outside Europe (Gyoutoku and Uemura 1985, Cooper and Rieske 2007, 2011). In France, *T. sinensis* has been proven established with fast and significant impacts on the targeted pest in the subsequent years (Borowiec et al 2018). This thus led to the quite unique opportunity to investigate how local communities evolve with regard to the deprivation of their trophic subsidy whereas most scientific work usually studies the recruitment of native parasitoids by the exotic biological control agents and its impact on food webs (Henneman & Memmott 2001, Eveleigh et al. 2007, Barber et al. 2008, Girardoz et al. 2008, David et al 2017).

## Methodology

**Biological control introductions**

In France, first isolated spots of *Dryocosmus kuriphilus* were observed from 2005 close to the Italian border but its pervasive presence in South of France was only patent from 2010. *Torymus sinensis* was introduced in France between 2011 and 2014, on a total of 59 sites (chestnut orchards) separated by at least 4 km. In each site, the monitoring of native parasitoids started one year prior to *T. sinensis* release. The introductions covered a wide geographical area (920 km from North to South, 1030 km from East to West) in metropolitan France including Corsica. Two propagule sizes (100 and 1000 individuals) of *T. sinensis* were introduced in separated sites but *T. sinensis* established itself in all sites whatever the initial propagule size was (see Borowiec et al. 2018 for more details). For this study we kept only the 26 sites for which at least five consecutive years of monitoring were available (Figure 1).

*Figure 1 - Map of the survey. Red points correspond to sites within an agricultural landscape (mainly apple orchards) whereas green points correspond to sites within a semi-natural landscape (mainly forests).Torymus sinensis was released in 2009 (Am1, Am2), in 2011 (Ard1, Cor1, Cor2, Cor3, Dro1, Dro2, Dro3, Var1) and in 2012 (Ard1T, Ard2, Ard3,Ard4, ArdX6, Cor3T, Cor6, Cor7, Corz1, Dord2, Gard1, Her1, Lot1, Lot2, Lot5, VarX).*

**Sampling of insect communities associated with chestnut galls**

149
150
151    -    Estimation of *D. kuriphilus* levels of infestation
152
Ten chestnut trees per site per year were sampled and, for each of them, ten totally random
twigs were selected at human height and inspected for galls as explained in Table 1. The
number of galls in a twig can therefore be zero. From these, the infestation levels of *D.*
*kuriphilus* were estimated, by combining information on the mean percentage of buds with
at least one gall and on the mean number of galls per bud as shown in Table 1. Because of
the difficulty of this task (geographical cover, meteorological contingencies, staff's
availability and skills), infestations were finally not available for some sites and/or dates
160

| | | MEAN NUMBER OF GALLS PER BUD | | |
|---|---|---|---|---|
| | | <1 | [1-2[ | 2≤ |
| **MEAN % OF BUDS WITH AT LEAST ONE GALL** | [0-33%] | 1 | 2 | 3 |
| | [34-66%] | 2 | 3 | 4 |
| | [67-100%] | 3 | 4 | 5 |

161    *Table 1 -Table showing how the classes of infestation by D. kuriphilus were determined*

162
163    -    Diversity and abundance of associated parasitoids
164
As detailed in Borowiec *et al.* 2018, both the exotic *Torymus sinensis* and the native
parasitoids (see table 2) were counted from winter dry galls which are easily distinguished as
they are brown and dry. In France, the asian chestnut gall wasp is the only gall wasp on
chestnut trees, therefore confusion with other species was impossible. We collected 2000 to
5000 galls per site during the two first years and then only 500 to 2000. Galls were gathered
on several trees. Once collected, galls were put in hermetic boxes placed outdoors from
January to October so that parasitoids could develop and emerge in natural conditions. Each
box referred to one site and contained 500 galls which is the good compromise between the
number of galls and the ability of parasitoids to emerge and get into the collecting vial
placed at the extremity of the boxes. All emerged insects were collected and then stored in
96° ethanol. In addition to the exotic and ubiquitous *T. sinensis*, nine main native parasitoids
were identified (Table 2). All identifications were based on morphological characters.
*Eupelmus* species were identified using the latest descriptions of the *Eupelmus urozonus*
complex (Al Khatib et al 2014 and 2016). Other species were identified by using an
unpublished key to chalcidoid parasitoids in oak cynipid galls (Askew and Thuroczy,
unpublished). DNA Barcoding using part of the mitochondrial coding gene *Cytochrome*
*Oxidase I* was used for a representative set of each morphologically-described genus and/or
species to ascertain their identifications (Warot & Viciriuc, unpublished).
183
184

| Native species | Host range | References |
|---|---|---|
| *Aulogymnus* **spp.** | Hymenoptera > Cynipidae | 1 |
| *Eupelmus azureus* | Hymenoptera > Cynipidae | 2 |
| *Eupelmus kiefferi* | Coleoptera Diptera Hemiptera Hymenoptera Lepidoptera | 2 |
| *Eupelmus urozonus* | Diptera Hymenoptera Neuroptera | 2 |
| *Eurytoma setigera* | Hymenoptera > Cynipidae > Cynipinae | 1, 3, 4 |
| *Megastigmus dorsalis** | Hymenoptera > Cynipidae | 1 |
| *Mesopolobus sericeus* | Hymenoptera > Cynipidae > Cynipinae > Cynipini | 1 |
| *Torymus auratus* | Hymenoptera > Cynipidae > Cynipinae | 1, 4 |
| *Sycophila biguttata* | Hymenoptera > Cynipidae | 1 |

*Table 2 - List of native species of native parasitoids with their known host range.*

*\* This species may be a complex of cryptic parasitoid species but all specialized on Cynipidae.*
*References : 1: Askew and Thuroczy, unpublished – 2: Al Khatib et al 2014 & 2016 – 3: Murakami et al. 1994 – 4: Noyes 2019.*

**Statistical analysis**

<u>Co-occurrence analyses</u>

To assess the patterns of co-occurrence between parasitoid species, and their evolution over time, we used the C-score (Stone and Roberts 1990) from each annual matrix of presence-absence of the nine native species. As *T. sinensis* was always present and here we only consider species occurrences (presence/absence), it was excluded of the analysis.

The C-score is a measure of average pairwise species segregation. It measures the mean number of checkerboard units between all pairs of species in a data matrix. The number of checkerboard units for each pair of species *i* and *k* is calculated as follows:

$$Cscore_{ik} = (S_i - Q)(S_k - Q) \qquad (1)$$

where Q is the number of shared sites, $S_i$ and $S_k$ are the number of sites in which species *i* and *k* are respectively found. In equation (1) the C-score will be equal to zero if species *i* and *k* share all sites. Conversely, C-score will be equal to one if species species *i* and *k* are never found together. The overall C-score for the community is then calculated as follows:

$$Overall\_Cscore = \sum \frac{(S_i - Q)(S_k - Q)}{\frac{R(R-1)}{2}} \qquad (2)$$

where R is the number of rows (=species) in the matrix (Stone and Roberts 1990, Gotelli
2000). When compared to other co-occurrence indices such as CHECKER (Diamond 1975), V-
ratio (Robson 1972, Schluter 1984) and COMBO (Pielou and Pielou 1968), C-score has the
smallest probability of type I and II errors (Gotelli 2000).
A low value of C-score is indicative of an aggregative pattern, while a high value is indicative
of an exclusion pattern. However, because the value of the C-score depends on the
frequency of occurrence of the species, inter-annual comparisons cannot be performed
directly. We thus used the co-occurrence null model from the EcoSimR package (Gotelli
2015) of R (R Development Core Team 2018) to create null assemblages based on our
observed presence-absence species matrices. This was done by randomizing (by transposing
sub-matrices) species occurrences but keeping row and columns totals fixed (Gotelli 2000).
Thus, differences between sites are maintained, making this method appropriate to detect
patterns of species interactions (Gotelli 2000). Each randomization produces one matrix in
which a 'simulated' C-score is calculated. Such randomization is replicated ten thousand
times. The significance of the observed C-scores was computed as the proportion of
simulated values equal to or more extreme than the observed C-score value.
In order to graphically compare each year, all c-score values were normalized by using:
230

$$Adjusted\ C\_score_t = \frac{x_t - \mu(Simulated\ C\_score)_t}{\sigma(Simulated\ C\_score)_t}$$

231
232  where $x$ takes the values of observed and simulated C-score and t refers to the year after
233  release of *T. sinensis* (from 1 to 5). The higher the observed value compared to the simulated
234  values, the more the community is structured by competitive exclusion patterns. Conversely,
235  the lower the observed value, the more the community structure relies on association
236  patterns.
237
238  <u>Native parasitoids community structure</u>
239
We described the community structure each year after the release of *T. sinensis* by using the
R package 'pheatmap' (Raivo 2019). We created clustered heatmaps with the 'pheatmap'
function to visualize how communities of native parasitoids are structured during the survey.
Sites were clustered depending on their native parasitoid absolute abundance using
aggregative clustering. As a distance measure between clusters x and y we used the
Euclidean distance which is calculated as follows:
246

$$d_{euc}(x, y) = \sqrt{\sum_{i=1}^{n} (x_i - y_i)^2}$$

247
248  As a linkage function, we chose the complete linkage which takes the maximum distance
249  between the two clusters:
250

$$f = \max(d(x, y))$$

Aggregative clustering starts by computing a distance between every pair of units to be
clustered and creating a distance matrix. The two clusters with the smallest value (e.g. A and
B) are then merged into a new cluster (e.g. AB). The matrix is then recalculated and as we
use the complete linkage the distance between the new cluster and the other clusters (e.g. C
and D). The distance between AB and C will be equal to the maximum value between C and
A and between C and B.
258
259          Landscaped context
260
In order to try to evaluate the potential role of habitat on the community structure during
the last year of survey, we used a Principal Component Analysis (PCA) considering the
abundances of each native species using the 'FactoMineR' package of R (Husson et al. 2019).
In other words sites were plotted in the first dimension of the PCA depending on the
abundance of native parasitoids in each of them. We then plotted the two main categories
of habitat: (i) orchards located within an agricultural landscape with a poor amount of semi-
natural habitat; (ii) orchards located within semi-natural habitats (forest, hill, mountain,
etc.). The habitat categorization was made based on qualitative estimates first during field
work and confirmed later on satellites views.
270
271 # Results
272
**Control of *Dryocosmus kuriphilus* by *Torymus sinensis***
A fast increase of the *Torymus sinensis*' relative frequency was observed during the 5 years
of survey, 90% of galls being finally parasitized by *T. sinensis* (Figure 2). This was easily
monitored as each gall can only contain one *T. sinensis* inside. In parallel, the infestation
levels of *D. kuriphilus* decreased markedly, based on the sites in which infestation data was
available for each year of the survey (Am1, Am2, Ard1, Gard2, Var1).
279

280
281  *Figure 2 - Infestation levels of D. kuriphilus and Relative frequency of T. sinensis in galls each year of the survey. Here are*
282  *only shown the sites were the infestation was consistently measured for all years of the survey.*

**Diversity and abundances of native species**

Taken as a whole, 71494 specimens of *T. sinensis* and 12016 specimens of native parasitoids were obtained from 284425 galls from the 26 sites during the 5 years of the survey. Galls from all locations were monitored for emerging parasitoids at our laboratory.
In terms of abundance, the native species were ordered as follows: *Mesopolobus sericeus* (n=3792, 31.6%), *Eupelmus urozonus* (n=2069, 17.2%), *Megastigmus dorsalis* (n=1877, 15.6%), *Eupelmus azureus* (n=1752, 14.6%), *Eurytoma setigera* (n=586, 4.9%), *Eupelmus kiefferi* (n=491, 4.1%), *Aulogymnus* spp. (n=403, 3.4%), *Sycophyla biguttata* (n=116, 1%), *Torymus auratus* (n=19, 0.1%). 911 (7,5%) individuals remained undetermined using both morphological and molecular identification and were thus discarded from the analysis.

In terms of occurrence, *Torymus sinensis* was observed in the 130 possible site-by-year combinations. In comparison, the results for native species were as follows: *Eupelmus urozonus* (n=111), *Eurytoma setigera* (n=86), *Eupelmus kiefferi* (n=74), *Megastigmus dorsalis* (n=59), *Mesopolobus sericeus* (n=50), *Eupelmus azureus* (n=49), *Aulogymnus* spp. (n=33) *Sycophyla biguttata* (n=26), *Torymus auratus* (n=7).

The mean abundances of all nine native parasitoids are given for each year in Figure S1 (*Supplemental Figure S1*). Taken as a whole, they peaked during the second and/or third years of the survey.

A potential pitfall with such survey could be to miss some other relevant species because of some insufficient sampling effort. However, we are quite confident that this is not the case insofar as, excepted for the rarest species (*T. auratus*), all other species were found at least common or even very common in some site-by-date combination (Figure 3).

*Figure 3 - Frequencies of observations the different native species.*
*Site-by-date-by-species combinations were sorted according to four classes of abundance: Rare (only 1 individual of the species), Uncommon (From 2 to 10 individuals), Common (From 11 to 100 individuals) and Very common (From 101 to max abundance sampled).*

**Co-occurrence null model analyses**

Figure 4 shows the temporal evolution of the observed C-score (red bar) with regard to the related distribution of simulated C-score values (histograms in blue, 95% confidence interval in-between the dotted black bars). Here, a clear trend is observed towards an overall competitive exclusion the fifth year (p.value < 0.05), beginning from the third year of the survey (p.value = 0.05) and being higly significant during the fifth year (p.value < 0.05). This means that the competition for *D. kuriphilus* increased, probably due both to its decline and the increase of *T. sinensis*. Therefore, as years passed, there was a decreasing chance of observing co-occurrence of native species by sampling *D. kuriphilus* galls. More details about which pairs of species were co-occuring more frequently across the years can be extracted from the heatmaps made for each year of the survey (supplementary material Figure S2-S5).

331
332 *Figure 4 - C-score values for the native community of parasitoids for the first (A), third (B) and fifth year (C) after the release*
333 *of T. sinensis. Blue histogram represents the simulated values, the red bar represents the observed C-score and the dotted*
334 *lines represent the 95% confidence interval.*

335
336

**Inter-site variability**

The overall tendency to competitive exclusion seems to result from two alternative patterns
(Figure 4): (i) in 19 sites, community of native parasitoids is poor (e.g. Gard1, Ard1T, etc.) or
even sometimes non-existent (e.g. Lot5, Corz1, etc.) during the 5th year of survey (Figure 4,
top cluster); (ii) in the remaining 7 sites, the community is dominated by *Mesopolobus*
*sericeus* (Figure 4, bottom cluster).

**Role of the environment**

The Principal Component Analysis on native parasitoid abundances confirms that, five years after the release of *T. sinensis*, communities are mostly structured by the local presence of *M. sericeus* (Figure 5A). The analysis of the projection of the different sites highlights that the abundance of *M. sericeus* is correlated with the type of habitat, semi-natural orchards being more likely to host this particular species (Figure 5B). As the PCA suggested an effect driven mostly by the response of the species *M. sericeus*, we planned on testing with a generalized linear model whether the abundance of *M. sericeus* was significantly different in agricultural and semi-natural habitats. However, given the numerous zeros within the agricultural category, we settled with a generalized linear model testing whether the occurrence (modelled as binomial) of *M. sericeus* was significantly different between the two habitats. *M. sericeus* is indeed occurring more frequently within semi-natural orchards (p.value=0.0396).

Figure 6 – A: Variables correlation plot of the PCA built from the abundance of native parasitoids Colors represent the contribution of the different species to the structure of the variation within each axis. B: Projection of the different geographical sites on two principal component axis of the PCA. The colors discriminate the two main habitat: Agricultural (red) and Semi-natural (green).

375    **Analysis without *Mesopolobus sericeus***
376

To verify whether the strong pattern driven by the species *M. sericeus* might have obscured
smaller responses in other species, we also did the analysis without *M. sericeus*. We found
that the native community still evolves towards a segregation pattern, although it is only
significant the fifth year of the survey (supplementary material Figure S6).
The clustering was significantly different with virtually no difference between sites. Here all
sites (excepted Cor1) are grouped in the same big cluster (supplementary material Figure
S7). In the same vein, the PCA showed a weaker differentiation between our two types of
habitat (supplementary material Figure S8).
385
386
387

## Discussion

Our work showed the restructuration of the native communities following the successful control of *Dryocosmus kuriphilus* by the biological control agent *Torymus sinensis*. In fact, communities appear to evolve differently depending on the landscape context of the sampling site. The following discussion will be structured in three main parts. We will first discuss the sampling effort of the study, then we will discuss the restructuration of the native community and finally we will interpret the influence of the landscaped context.

*Sampling*

Our sampling effort was strong enough so that we can be confident we did not miss any relevant species. Even if a single individual of a given species was sampled relatively frequently (about 20% of all data), all species except the most rare one were also sampled at high density in some cases. This implies that no species was so rare that the probability to have by chance completely missed another species with comparable abundance is close to zero. With regard to the most rare species, *Torymus auratus,* it is already known to be rare in winter dry galls but more abundant in spring fresh galls, so that our sampling method might have underestimated its abundance (Kos et al 2015, Ferracini et al 2018). Furthermore, species rankings according to abundance and occurrence are not the same, which confirms that our sampling method is robust enough to accurately detect species even when their abundance is low. These results, taken together, support our claim that the probability that we missed another species abundant or frequent enough to have a significant impact on community structure is indeed extremely low.

*Structure of the native community*

The nine native parasitoids that appeared to use *D. kuriphilus* as a trophic subsidy in our survey are related to oak gall wasps, although their degree of generalism is highly variable (Table 2). Some species such as *Eupelmus kiefferi* and *E. urozonus* are indeed extremely polyphagous whereas, in contrast, *Mesopolobus sericeus* is substantially more specific, being specialized on only one tribe of Cynipids (Noyes 2019). The main result of the co-occurrence model analyses is the increasing exclusive competition through time (Figure 4). This means that the native community undergoes a significant restructuration, which is correlated in time with the rarefaction of *D. kuriphilus*. The first parasitoids actually exploiting *D. kuriphilus* were *E. urozonus*, *E. azureus* and *M. dorsalis* (Figure S1), those species combining (i) a wide host range, (ii) a known affinity with several Cynipidae and (iii) pre-existing resource in such kinds of habitats (Al khatib et al. 2014, 2016). They were thus therefore likely to shift and more successful in colonizing invasive host (Cornell and Hawkins 1993, Hawkins 2005). However, in the following years, *Mesopolobus sericeus*, which was not detected during the first year of survey (Figure S1), was the sole species able to markedly increase in abundance and to persist on this trophic subsidy (Figure 5). And this, despite the fact that it was not detected once at the beginning of the survey in any site (Figure S1). It seems to be the same for *Aulogymnus* spp. except that they are much less abundant and this includes all species from the *Aulogymnus* genus. The examples of native species displaced by invasive species are numerous (e.g. Rowles and O'Dowd 2007, Bohn et al 2008, Inoue et al

2008, Sebastian et al 2015) therefore we are not surprised to observe such outcome for
most of our native species, which are outcompeted by the introduced specialist *T. sinensis*.
Only *Mesopolobus sericeus* appears to be able to coexist at significant levels with *Torymus*
*sinensis* on *Dryocosmus kuriphilus*. Furthermore, the most generalist parasitoids of our
system probably exhibit a switching behavior (Murdoch 1969). This behavior refers to a
situation where a predator (or here a parasitoid) exhibits adaptive, flexible choices between
all available preys (or hosts). This choice entails positively frequency-dependent predation.
In our study abundance of other hosts are unknown. It is therefore possible that *D.*
*kuriphilus* eventually becomes rarer than other hosts, forcing generalists to switch to more
abundant hosts (Pelletier 2000). However, switching preys can have stabilizing or
destabilizing effects, even possibly causing extinction of the predator (Van Leeuwen et al
2007).
447
448    *Influence of landscape context*
449
Increasing landscape complexity (mostly defined as the increase of the proportion of semi-
natural habitat) is generally associated with increases in natural enemy abundance and/or
diversity (Bianchi et al 2006, Schmidt et al 2008, Gardiner et al, 2009a, 2009b). In our study
sites enclosed within large amounts of semi-natural habitats contained the most diverse and
abundant native parasitoid communities (Figure 5). In particular, the persistence of *M.*
*sericeus,* the most specialist native parasitoid, was modulated by the local environment
(Figure 6B). Indeed, *M. sericeus* was the native parasitoid that seems to be the best at
exploiting *D. kuriphilus*, although only within semi-natural habitat. Among the seven sites
showing a marked domination of *M. sericeus* (Figure 5)*,* six of which are located in the island
of Corsica (Figure 5). Islands are home to plant and animal communities with relatively little
diversification, simplified trophic webs and high rates of endemism (Williamson 1981,
Chapuis et al., 1995). These three points in addition of their smaller size (Cassey 2003)
partially explain that oscillations (or perturbations) within the resident community are more
incline to destructive outcomes such as extinctions (Elton 1958). Nonetheless, five
continental sites (i.e. ArdX6, Dro3, Ard1T, Ard4, Ard1; assigned with stars in Figure 5) also
exhibit a slightly less marked but similar increase of *M. sericeus* (Figure 5), four of them
being in semi-natural landscapes (Figure 1). We thus think that the final dominance of *M.*
*sericeus* towards other native species is rather explained by differences in the landscape
rather than a "mainland *versus* island" dichotomy. In fact, most of the known hosts of *M.*
*sericeus* are oak gall wasps (Noyes 2019), oak trees being rarer in agricultural landscapes
than in semi-natural ones. Large populations of *M. sericeus* acting like sources for the
colonization of chestnut orchards are consequently more likely to be sustained in this latter
habitat.
473
474    *Limitations and strengths of the study*
475
In classical biological control, the ecological impacts of a biological control agent are usually
explained by the use of non-target hosts/preys (Louda et al 2003). Although *Torymus*
*sinensis* has shown a slight host range expansion it appeared to be with minimal impact and
no effect expected on distribution and abundance of non-target hosts (Ferracini et al 2017).
Thus its impact on native competitors is mediated by its successful control of the shared
host.

We need to point out that although our study contains insightful information on how our
native parasitoid community structure evolves, species dynamics we observed the last year
of the survey are not fixed but quite the opposite. Species dynamics are most probably still
evolving towards a, yet unknown, state of equilibrium. We are still not in measure to predict
with certainty what will happen when *D. kuriphilus* will become even rarer. Maybe *T.*
*sinensis* will remain the dominant species or maybe because of the presence of its native
hosts, *M. sericeus* will outperform *T. sinensis* at least in semi-natural habitats. Furthermore,
although we evidenced a successful host range expansion (by newly including *T. sinensis* into
their diet) from the majority of these native parasitoids, nothing is known about how the
populations dynamics evolved on their native hosts.
492
493    *Conclusion*
494
Classical biological control offers an exciting frame to investigate real-time population
dynamics during invasive processes. Yet, the opportunities remain rare because of various
reasons including (i) the quite high rate of establishment's failure, (ii) the temporal frame
required for the observation of significant patterns and (iii) the lack of funding for post-
release surveys. With regard to this context, the deliberate introduction of *T. sinensis* against
*D. kuriphilus* in France thus was a quite unique opportunity. Our work sheds a new light on
how the "boom-and-bust" (here defining the situation in which a period of great prosperity
is abruptly followed by one of decline) dynamics of an invasive pest can impact the structure
of native communities of potential antagonists. Our results evidence a site-specific scenario
where a sole native species, *M. sericeus*, dominates the native community on the trophic
subsidy and is able to co-exist with the exotic and specialized competitor, *T. sinensis*. *M.*
*sericeus* is now able to exploit both the native gall wasps and *D. kuriphilus*. This extended
host range may have lasting impacts on *T. sinensis* populations, all the more so *D. kuriphilus*
will reach a low density at a global scale. In turn, the rarefaction of *D. kuriphilus* and the
competition with *M. sericeus* might constrain *T. sinensis* to exploit new hosts. Therefore it
would be of particular interest to study the long-term evolution of these two species as the
ideal expected outcome of classical biological control is the everlasting control of the host
with no unintentional negative impact on the recipient community. Another open
perspective of this work is to analyze how the structure of native parasitoids evolves within
the oak gall wasp's community.

## References

Al Khatib, F. *et al.* An integrative approach to species discrimination in the *Eupelmus urozonus* complex (Hymenoptera, Eupelmidae), with the description of 11 new species from the Western Palaearctic. *Syst. Entomol.* **39**, 806–862 (**2014**).

Al khatib, F. *et al.* Multilocus phylogeny and ecological differentiation of the 'Eupelmus urozonus species group' (Hymenoptera, Eupelmidae) in the West-Palaearctic. *BMC Evol. Biol.* **16**, (2016).

Barber, N. A., Marquis, R. J. & Tori, W. P. Invasive prey impacts the abundance and distribution of native parasitoids. *Ecology* **89**, 2678–2683 (**2008**).

Baudrot, V., Perasso, A., Fritsch, C., Giraudoux, P. & Raoul, F. The adaptation of generalist predators' diet in a multi-prey context: Insights from new functional responses. *Ecology* **97**, 1832–1841 (**2016**).

Bengtsson, J. Interspecific competition increases local extinction rate in a metapopulation system. *Nature* **340**, 713–715 (**1989**).

Bennett, F. D. Do Introduced Parasitoids Displace Native Ones ? *Florida Entomol.* **76**, 54–63 (**1993**).

Bernardo, U. *et al.* Description of *Synergus castaneus* n. sp. (Hymenoptera: Cynipidae: Synergini) Associated With an Unknown Gall on *Castanea* spp. (Fagaceae) in China. *Ann. Entomol. Soc. Am.* **106**, 437–446 (**2013**).

Bianchi, F. J. J. A., Booij, C. J. H. & Tscharntke, T. Sustainable pest regulation in agricultural landscapes: A review on landscape composition, biodiversity and natural pest control. *Proc. R. Soc. B Biol. Sci.* **273**, 1715–1727 (**2006**).

Blackburn, T. M., Cassey, P., Duncan, R. P., Evans, K. L. & Gaston, K. J. Avian extinction and mammalian introductions on oceanic islands. *Science (80-. ).* **305**, 1955–1958 (**2004**).

Bøhn, T., Amundsen, P. A. & Sparrow, A. Competitive exclusion after invasion? *Biol. Invasions* **10**, 359–368 (**2008**).

Borowiec, N. *et al.* Early population dynamics in classical biological control: establishment of the exotic parasitoid *Torymus sinensis* and control of its target pest, the chestnut gall wasp *Dryocosmus kuriphilus*, in France. *Entomol. Exp. Appl.* **166**, 367–379 (**2018**).

Brussino, G. *et al.* Pericoloso insetto esotico per il castagno europeo. *Avversità delle piante.* 59–62 (**2002**).

Carroll, S. P. Natives adapting to invasive species: Ecology, genes, and the sustainability of conservation. *Ecol. Res.* **22**, 892–901 (**2007**).

Cassey, P. A comparative analysis of the relative success of introduced land birds on islands.
*Evol. Ecol. Res.* **5**, 1011–1021 (**2003**).

Chapuis, J.L., Barnaud, G., Bioret, F., Lebouvier, M. & Pascal, M. L'éradication des espèces
introduites, un préalable à la restauration des milieux insulaires. Cas des îles françaises.
*Natures Sci. Sociétés* **3**, s51–s65 (**1995**).

Cooper, W. R. & Rieske, L. K. Community Associates of an Exotic Gallmaker, *Dryocosmus*
*kuriphilus* (Hymenoptera: Cynipidae), in Eastern North America. *Ann. Entomol. Soc.*
*Am.* **100**, 236–244 (**2007**).

Cooper, W. R. & Rieske, L. K. A native and an introduced parasitoid utilize an exotic gall-
maker host. *BioControl* **56**, 725–734 (**2011**).

Cornell, H. V & Hawkins, B. A. Accumulation of Native Parasitoid Species on Introduced
Herbivores⬜: A Comparison of Hosts as Natives and Hosts as Invaders. *Am. Soc. Nat.*
**141**, 847–865 (**1993**).

Courchamp, F., Chapuis, J. L. & Pascal, M. Mammal invaders on islands: Impact, control and
control impact. *Biol. Rev. Camb. Philos. Soc.* **78**, 347–383 (**2003**).

Cox, J. G. & Lima, S. L. Naiveté and an aquatic-terrestrial dichotomy in the effects of
introduced predators. *Trends Ecol. Evol.* **21**, 674–680 (**2006**).

Crawley, M. J. & Schonrogge, K. Quantitative webs as a means of assessing the impact of
alien insects. *J. Anim. Ecol.* **69**, 841–868 (**2000**).

Daszak, P., Cunningham, A. A. & Hyatt, A. D. Emerging infectious diseases of wildlife -
Threats to biodiversity and human health. *Science,* **287**, 443–449 (**2000**).

David, P. *et al.* Impacts of Invasive Species on Food Webs: A Review of Empirical Data.
*Adv. Ecol. Res.* **56**, 1–60 (**2017**).

Diamond, J. Assembly of species communities. in *Ecology and Evolutioin of Communities*
(eds. Cody, M. & Diamond., J.) 342–444 (**1975**).

Eilenberg, J., Hajek, A. & Lomer, C. Suggestions for unifying the terminology in biological
control. *BioControl* **46**, 387–400 (**2001**).

Elton, C. The ecology of invasions by plants and animals. (**1958**).

Eveleigh, E. S. *et al.* Fluctuations in density of an outbreak species drive diversity cascades in
food webs. *Proc. Natl. Acad. Sci. U. S. A.* **104**, 16976–16981 (**2007**).

Ferracini, C., Ferrari, E., Pontini, M., Hernandez Nova, L. K., Saladini, M. A., Alma, A. Post-
release evaluation of non-target effects of *Torymus sinensis*, the biological control agent of
*Dryocosmus kuriphilus* in Italy. *BioControl* **62**, 445-456 (**2017**).

Ferracini, C., Bertolino, S., Bernardo, U., Bonsignore, C. P., Faccoli, M., Ferrari, E., Lupi, D.,
Maini, S., Mazzon, L., Nugnes, F., Rocco, A., Santi, F., & Tavella, L. Do *Torymus sinensis*

(Hymenoptera: Torymidae) and agroforestry system affect native parasitoids associated with the Asian chestnut gall wasp? *Biological Control* **121**, 36-43 (**2018**).

Francati, S., Alma, A., Ferracini, C., Pollini, A. & Dindo, M. L. Indigenous parasitoids associated with *Dryocosmus kuriphilus* in a chestnut production area of Emilia Romagna (Italy). *Bull. Insectology* **68**, 127–134 (**2015**).

Gardiner, M. M., Landis, D. A., Gratton, C., Schimdt, N. P., O'Neal, M., Mueller, E. E., Chacon, J. M., Heimpel, G. E., DiFonzo, C. D. Landscape composition influences patterns of native and exotic lady beetle abundance. *Divers. Distrib.* **15**, 554–564 (**2009a**).

Gardiner, M. M., Landis, D. A., Gratton, C., DiFonzo, C. D., O'Neal, M., Chacon, J. M., Wayo, M. T., Schimdt, N. P., Mueller, E. E., Heimpel, G. E. Landscape diversity enhances biological control of an introduced crop pest in the north-central USA. *Ecol. Appl.* **19**, 143–154 (**2009b**).

Girardoz, S., Kenis, M. & Quicke, D. L. J. Recruitment of native parasitoids by an exotic leaf miner, Cameraria ohridella: Host-parasitoid synchronization and influence of the environment. *Agric. For. Entomol.* **8**, 49–56 (**2006**).

Gotelli, N. Null model analysis of species co-occurrence patterns. *Ecology* **81**, 2606–2621 (**2000**).

Gotelli, N., Hart, E. & Ellison, A. Package ' EcoSimR '. *R Packag.* (**2015**).

Grosholz, E. Ecological and evolutionary consequences of coastal invasions. *Trends Ecol. Evol.* **17**, 22–27 (**2002**).

Gyotoku, Y. & Uemura, M. Ecology and biological control of the chestnut gall wasp, *Dryocosmus kuriphilus* Yasumatsu (Hymenoptera: Cynipidae), 1: Damage and parasitization in Kumamoto prefecture [Japan]. *Proc. Assoc. plant Prot. kyushu* (**1985**).

Hamilton, J. G., Holzapfel, C. & Mahall, B. E. Coexistence and interference between a native perennial grass and non-native annual grasses in California. *Oecologia* **121**, 518–526 (**1999**).

Hawkins, B. Pattern and process in host-parasitoid interactions. (**2005**).

Haye, T., Fischer, S., Zhang, J. & Gariepy, T. Can native egg parasitoids adopt the invasive brown marmorated stink bug, *Halyomorpha halys* (Heteroptera: Pentatomidae), in Europe? *J. Pest Sci. (2004).* **88**, 693–705 (**2015**).

Henneman, M. L. & Memmott, J. Infiltration of a hawaiian community by introduced biological control agents. *Science.* **293**, 1314–1316 (**2001**).

Herlihy, M. V., Talamas, E. J. & Weber, D. C. Attack and Success of Native and Exotic Parasitoids on Eggs of *Halyomorpha halys* in Three Maryland Habitats. *PLoS One* **11**, e0150275 (**2016**).

Howarth, F. G. Classical biocontrol: panacea or Pandora's Box. *Proc. Hawaiian Entomol.*
*Soc.* **24**, 239–244 (**1983**).

Howarth, F. G. Environmental Impacts of Classical Biological Control. *Annu. Rev. Entomol.*
**36**, 485–509 (**1991**).

Husson, A. F., Josse, J., Le, S., Mazet, J. & Husson, M. F. Package ' FactoMineR '. *R*
*Packag.* (**2019**).

Inoue, M. N., Yokoyama, J. & Washitani, I. Displacement of Japanese native bumblebees by
the recently introduced *Bombus terrestris* (L.) (Hymenoptera: Apidae). *J. Insect*
*Conserv.* **12**, 135–146 (**2008**).

Jones, A. L., Jennings, D. E., Hooks, C. R. R. & Shrewsbury, P. M. Sentinel eggs
underestimate rates of parasitism of the exotic brown marmorated stink bug,
*Halyomorpha halys. Biol. Control* **78**, 61–66 (**2014**).

Keitt, B. S., Wilcox, C., Tershy, B. R., Croll, D. A. & Donlan, C. J. The effect of feral cats on
the population viability of black-vented shearwaters (*Puffinus opisthomelas*) on
Natividad Island, Mexico. *Anim. Conserv.* **5**, 217–223 (**2002**).

Kolde, R. Package ' pheatmap '. *R Packag.* 1–8 (**2019**).

Kos, K., Kriston, E., & Melika, G. Invasive chestnut gall wasp *Dryocosmus kuriphilus*
(Hymenoptera: Cynipidae), its native parasitoid community and association with oak
gall wasps in Slovenia. *Eur. J. Entomol.* **112**(4), 698-704 (**2015**).

Lewi, S. & Alan, R. The checkerboard score and species distributions. *Oecologia* **85**, 74–79
(**1990**).

Louda, S. M., Pemberton, R. W., Johnson, M. T. & Follett, P. A. NONTARGET E FFECTS
—THE ACHILLES ' HEEL OF BIOLOGICAL CONTROL□? Retrospective
Analyses to Reduce Risk Associated with Biocontrol Introductions. *Annu. Rev.*
*Entomol.* **48**, 365–396 (**2003**).

Lynch, L. D. & Thomas, M. B. Nontarget effects in the biocontrol of insects with insects,
nematodes and microbial agents: the evidence. *Biocontrol News Inf.* **21**, 117N – 130N
(**2000**).

Mallon, C. A. *et al.* The impact of failure: Unsuccessful bacterial invasions steer the soil
microbial community away from the invader's niche. *ISME J.* **12**, 728–741 (**2018**).

Matošević, D. & Melika, G. Recruitment of native parasitoids to a new invasive host: First
results of *Dryocosmus kuriphilus* parasitoid assemblage in Croatia. *Bull. Insectology*
**66**, 231–238 (**2013**).

McGeoch, M. A., Lythe, M. J., Henriksen, M. V. & McGrannachan, C. M. Environmental
impact classification for alien insects: A review of mechanisms and their biodiversity
outcomes. *Curr. Opin. Insect Sci.* **12**, 46–53 (**2015**).

Murakami, Y., Hiramatsu, T. & Maeda, M. Parasitoid complexes of the chestnut gall wasp
(Hymenoptera: Cynipidae) in two localities before introduction of *Torymus*
(*Syntomaspis*) *sinensis* (Hymenoptera: Torymidae), with special reference to prediction
of results after release of the parasitoid. *Jap. Jour. Appl. Zool. Entomol.* **38**, 29-41
(**1994**).

Naranjo, S. E. Displacement of Native Natural Enemies by Introduced Biological Control
Agents in Agro- Ecosystems: A Serious Non-target Effect or Not? in *Proceedings of*
*the 5th International Symposium on Biological Control of Arthropods* 46–49 (**2017**).

Noyes, J.S. Universal Chalcidoidea Database. World Wide Web electronic publication.
http://www.nhm.ac.uk/chalcidoids (**2019**, last access 2019-11).

Olabarria, C., Incera, M., Garrido, J., Rodil, I. F. & Rossi, F. Intraspecific diet shift in Talitrus
saltator inhabiting exposed sandy beaches. *Estuar. Coast. Shelf Sci.* **84**, 282–288
(**2009**).

Panzavolta, T. *et al.* Native parasitoids associated with Dryocosmus kuriphilus in Tuscany,
Italy. *Bull. Insectology* **66**, 195–201 (**2013**).

Pelletier, J. D. Are large complex ecosystems more unstable? A theoretical reassessment with
predator switching. *Math. Biosci.* **163**, 91–96 (**2000**).

Pielou, D. P. & Pielou, E. C. Association among species of infrequent occurrence: The insect
and spider fauna of *Polyporus betulinus* (Bulliard) Fries. *J. Theor. Biol.* **21**, 202–216
(**1968**).

Quacchia, A., Moriya, S., Askew, R., Schönrogge, K. *Torymus sinensis*: biology, host range
and hybridization. *Acta Hort* **1043**, 105–112 (**2014**).

Ricciardi, A. & Macisaac, H. J. Recent mass invasion of the North American Great Lakes by
Ponto–Caspian species. **15**, 62–65 (**2000**).

Ricciardi, A. Facilitative interactions among aquatic invaders: Is an 'invasional meltdown'
occurring in the great lakes? *Can. J. Fish. Aquat. Sci.* **58**, 2513–2525 (**2001**).

Robson, D. Appendix: statistical tests of significance. *J. Theor. Biol.* **34**, 350–352 (**1972**).

Rodriguez, L. F. Can invasive species facilitate native species? Evidence of how, when, and
why these impacts occur. *Biol. Invasions* **8**, 927–939 (**2006**).

Rossi, F., Olabarria, C., Incera, M. & Garrido, J. The trophic significance of the invasive
seaweed *Sargassum muticum* in sandy beaches. *J. Sea Res.* **63**, 52–61 (**2010**).

Rowles, A. D. & O'Dowd, D. J. Interference competition by Argentine ants displaces native
ants: Implications for biotic resistance to invasion. *Biol. Invasions* **9**, 73–85 (**2007**).

Sax, D. F. & Gaines, S. D. Species invasions and extinction: The future of native biodiversity
on islands. *Proc. Natl. Acad. Sci.* **105**, 11490–11497 (**2008**).

Schluter, D. A Variance Test for Detecting Species Associations , with Some Example
Applications. *Ecology* **65**, 998–1005 (**1984**).

Sebastián, O. S., Navarro, J., Llorente, G. A. & Richter-Boix, Á. Trophic strategies of a non-
native and a native amphibian species in shared ponds. *PLoS One* **10**, (**2015**).

Steadman, D. W. Extinction and biogeography of tropical Pacific birds. (University of
Chicago Press, **2006**).

Suárez-Jiménez, R. *et al.* Importance of the invasive macroalga *Undaria pinnatifida* as
trophic subsidy for a beach consumer. *Mar. Biol.* **164**, 1–9 (**2017**).

Suckling, D. M. & Sforza, R. F. H. What magnitude are observed non-target impacts from
weed biocontrol? *PLoS One* **9**, (**2014**).

Van Leeuwen, E., Jansen, V. A. A. & Bright, P. W. How population dynamics shape the
functional response in a one-predator-two-prey system. *Ecology* **88**, 1571–1581 (**2007**).

van Lenteren, J. C., Bale, J., Bigler, F., Hokkanen, H. M. T. & Loomans, A. J. M. Assessing
Risks of Releasing Exotic Biological Control Agents of Arthropod Pests. *Annu. Rev.*
*Entomol.* **51**, 609–634 (**2006**).

Williamson, M. Island populations. 300 (**1984**).