Temperature-dependence of metabolic rate in tropical and temperate aquatic insects: support for the Climate Variability Hypothesis in mayflies but not stoneflies.

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Running Head:

Aquatic insect sensitivity to climate change

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Abstract:

A fundamental gap in climate change vulnerability research is an understanding of the relative thermal

sensitivity of tropical and temperate organisms. Aquatic insects are vital to stream ecosystem function and

biodiversity, but insufficiently studied with respect to their thermal physiology. With global temperatures

rising at an unprecedented rate, it is imperative that we understand how aquatic insects respond to

increasing temperature and whether these responses vary among taxa, latitudes, and elevations. We

evaluated the thermal sensitivity of standard metabolic rate in stream-dwelling baetid mayflies and perlid

stoneflies across a ~2,000 m elevation gradient in the temperate Rocky Mountains in Colorado, U.S.A.,

and the tropical Andes in Napo, Ecuador. We used temperature-controlled water baths and

microrespirometry to estimate changes in oxygen consumption. Tropical mayflies across all elevations

exhibited greater thermal sensitivity in metabolism compared to temperate species; their metabolic rates

increased more rapidly with temperature and they frequently exhibited behavioral signs of stress. By

contrast, temperate and tropical stoneflies did not clearly differ. Varied responses to temperature among

baetid mayflies and perlid stoneflies may reflect their evolutionary history in the tropics or their different

ecological roles as herbivores and predators, respectively. Our results show that there is physiological

variation across elevations and species and low elevation tropical mayflies may be the most imperiled by

climate warming. However, given such variation among species, broad generalizations about the

vulnerability of tropical ectotherms must be made more cautiously.

Introduction:

52   Metabolism is crucial to all life. It integrates an organism's existing energy supplies to fuel

fitness-related activities like movement, growth, tissue maintenance, and development (Dillon, Wang, &

Huey, 2010; Gillooly, Brown, West, Savage, & Charnov, 2001). In ectotherms, metabolic rates are

sensitive to body temperature, which can influence not only individual fitness, but geographic distribution

and abundance as well (Angilletta, 2009; Terblanche & Chown, 2007; Vannote & Sweeney, 1980). A

major challenge in global change biology is to understand how temperature alters metabolism and energy

budgets of different species, and in turn, impacts the ability of species to cope with climate change

(Colwell, Brehm, Cardelús, Gilman, & Longino, 2008; Parmesan, Yohe, & Andrus, 2003; Walther et al.,

2002). So far, attempts to measure vulnerability to climate change have largely assessed species responses

to temperature challenges, such as lethal and critical thermal limits (Deutsch et al., 2008; Pinsky, Eikeset,

McCauley, Payne, & Sunday, 2019). Despite their utility, these metrics address only the responses of

organisms to extreme temperatures and do not capture the continuous or incremental effects of

temperature on an organism's energy budget. Such effects are particularly important in ectotherms,

because body temperature and metabolic rate increase with environmental temperature, resulting in higher

energy expenditure. This response is likely to trade off against other competing demands such as growth,

reproduction, and locomotor performance (Pörtner et al., 2006). For example, in aquatic ectotherms,

increased metabolic demand resulting from higher temperature can outpace supply of oxygen from the

environment causing decreased performance and lowered tolerance to heat stress (Pörtner, Bock, & Mark,

2017). Thus, understanding how metabolic responses to temperature vary across different thermal

environments can help us predict geographic variation in response to global warming (Dillon et al., 2010).

In this study, we measured metabolic rate profiles in juveniles (nymphs) of two major groups of

stream insects – mayflies and stoneflies – in temperate and tropical streams across a paired elevation

gradient. Mayflies and stoneflies spend most of their life cycle as aquatic nymphs, where they feed, grow,

and develop eggs and sperm packets before emerging to breed as short-lived flying adults (Brittain,

1990). Water temperature is therefore a major driver of life history traits in these taxa. We aimed to

elucidate differences in patterns of species' thermal sensitivity and thus better evaluate potential

vulnerabilities of mayflies and stoneflies to projected climate warming for which, currently, very few data

exist (Chown, Duffy, & Sørensen, 2015).

Much like other types of thermal performance curves (TPCs), oxygen-consumption profiles (used

to measure metabolic rate) vary with increasing temperature and display three identifiable regions

(Pörtner, 2002; Schulte, 2015): 1) an ascending phase; 2) a peak, which indicates the onset of stress (Huey

& Stevenson, 1979; Pörtner & Knust, 2007); and 3) a descending phase of metabolic depression (Guppy

& Withers, 1999). When coupled with other measures of performance and the critical thermal limits,

changes in oxygen consumption provide a more complete picture of how temperature shapes the ecology

of an organism (Angilletta, 2009; Pörtner et al., 2017) (Fig. 1).

Although TPCs for traits like locomotion and metabolic rate are expected to have similar shapes,

a metabolic rate curve is a special type of TPC (Pörtner, 2002; Schulte, Healy, & Fangue, 2011), with

different implications for organismal fitness (Kellermann et al., 2019). The performance traits used in

many TPCs—running speed, feeding rates, or growth—are often interpreted as components of fitness,

such that *more is better*. For these traits, peaks indicate the location of thermal optima (T_{OPT}, Fig. 1) and

suggest that performance is optimized at that temperature (Angilletta, Niewiarowski, & Navas, 2002;

Huey & Kingsolver, 1989; Huey & Stevenson, 1979) or something close to it (Martin & Huey, 2008).

However, for metabolism, more is not necessarily better, and peak rates of standard oxygen consumption

represent elevated minimum costs, typically occurring at warmer temperatures than peak performance

traits (Pörtner, 2001). For example, aerobic scope is maximized where the difference between standard

metabolic rate (SMR) and maximum metabolic rate (MMR) is greatest (Fig. 1A; often the location of

peak performance T_{OPT})—not when SMR itself is at its peak. At higher temperatures, when SMR meets

MMR, a metabolic depression often follows as organismal functions decline (Fig. 1; see also Pörtner,

2002). Thus, the metabolic peak should lie to the right of T_{OPT} (i.e. at higher temperatures than T_{OPT}) and

align with decreasing performance on the TPC (Fig. 1; Pörtner, 2001). We refer to this peak as $T\text{-}MR_{PEAK}$.

The shapes of metabolic rate curves found for different species, populations, and thermal

experiences (e.g., acclimation; DeLong et al., 2018; Havird et al., 2020) could reflect either the acute

effects of temperature on biological rates (Dillon et al., 2010; Payne & Smith, 2017) or the evolutionary

response to past climatic conditions (Dell, Pawar, & Savage, 2011; Deutsch et al., 2008). If the shape of

the metabolic rate TPC simply reflects thermodynamics of biological reactions, we would predict that

organisms of equivalent size experiencing a similar range of temperatures should exhibit similar

metabolic rate curves regardless of their evolutionary history (Payne & Smith, 2017).

Alternatively, the Climate Variability Hypothesis posits that if stable climatic regimes, such as

those found in the tropics, expose organisms to only a narrow range of temperatures and favor the

evolution of thermal specialists (*sensu* Huey & Kingsolver, 1989), then we predict that those organisms

should have lower SMR within the range of temperatures they experience to maximize the allocation of

energy to growth, reproduction, and aerobic performance (Fig.1B). These thermal specialists should also

be more sensitive to temperature, i.e. have metabolic rates that change rapidly in response to temperature

outside their normal range. Climates characterized by large seasonal temperature variation, such as those

found at temperate latitudes, should select for thermal generalists capable of functioning effectively over

a wide range of temperatures (Angilletta et al., 2002; Huey & Kingsolver, 1993). Thermal generalists are

predicted to have relatively low SMRs across a wider range of temperatures (Fig. 1C) and be less

sensitive to increasing temperature to keep their minimum energy demands low. The generality of these

contrasting hypotheses remains unresolved because few comparative studies have measured metabolic

rates in species from temperate and tropical regions under similar experimental conditions.

To test predictions generated by these alternative hypotheses, we examined metabolic rates of

mayflies and stoneflies from temperate and tropical regions and across different elevations. Because

stream temperatures at a given elevation were similar at the time of the experiments, we predicted that the

metabolic rate curves should look similar between temperate and tropical species if they were strictly

determined by thermodynamic effects on metabolic processes (Payne & Smith, 2017). But, if metabolic

rates reflect adaptation to climate, we predicted greater sensitivity to temperature in the tropical species

(Fig. 1B) compared to temperate ones (Fig. 1C; Angilletta et al., 2002; Huey & Kingsolver, 1993).

Figure 1. Expectations for change in standard metabolic rate (oxygen consumption) and performance in
response to temperature in an aquatic ectotherm. The thermal performance curve (TPC, solid green line,
left panel) could refer to any functional performance trait (e.g. swimming performance), where higher
performance is better. T_{OPT}, where performance is highest, reflects the animal's preferred temperature.

The oxygen consumption curve (dashed line) is a special case of TPC. The peak in this case (T-MR$_{PEAK}$)

could represent the temperature at which standard metabolic rate (SMR) is so high that it essentially

equals the maximum metabolic rate (MMR) that the ectotherm can achieve (circle, right panel A). T-

MR$_{PEAK}$ may therefore reflect stress (red color) and aligns with a decline in the functional performance

trait. The general location of behaviorally determined critical thermal limits (CT$_{MIN}$ and CT$_{MAX}$) are also

shown (blue line, left panel). In this study we focus only on SMR. We predict that SMR will rise steeply

in tropical species (right panel B) and be stressful when temperatures exceed the typical narrow range of

temperatures they experience (gray rectangle). In comparison, temperate species should exhibit a gradual

rise in SMR across the same temperatures (right panel C), reflecting the broad range of environmental

temperatures they experience (grey rectangle).

Methods

Site and species descriptions

Temperate sites were located in the Cache La Poudre river drainage of the southern Rocky Mountains in

Colorado, USA, and tropical sites in the Papallacta-Quijos river drainage of the Andes Mountains in Napo

Province, Ecuador. We chose small, relatively pristine streams where the mayflies and stoneflies used in this

study are typically found. Sites at both latitudes were matched by elevation, where high >3000 m, mid = 2500 –

3000 m, and low = 1000 – 2500 m. Streams within each elevation pair were approximately the same temperature

at the time of collection (Table 1). Thus, we were able to make latitudinal comparisons of insect metabolic rate for

each elevation pair under the same thermal conditions.

Ephemeroptera (mayflies) and Plecoptera (stoneflies) do not share genera between tropical and temperate

sites. Therefore, we selected representative species from the same family (i.e. mayfly family Baetidae and

stonefly family Perlidae) at each location to contrast their metabolic rate. Based on our previous DNA-barcoding

work (Gill et al., 2016; Polato et al., 2018; Shah et al., 2017b), we could identify taxa to species using

morphological descriptions of individuals from these sites. Ultimately, we chose these taxa because temperate and

tropical relatives are ecologically and morphologically very similar to each other and because baetid mayflies and

perlid stoneflies tend to be common in streams around the world. Results from our work could therefore have

global implications.

In the Rockies, we collected a total of 313 individual mayflies from the *Baetis* species complex (*Baetis*

bicaudatus at high elevations and *Baetis tricaudatus* at low elevations). We also collected 107 stoneflies

(*Hesperoperla pacifica* at low and mid-elevations). In the Andes, we collected 245 mayflies from the genus

Andesiops. Earlier work has shown cryptic diversity in this genus, such that *Andesiops* collected at each elevation

were putatively different species (Gill et al., 2016). We collected 76 stoneflies from the genus *Anacroneuria*,

which comprises *Anacroneuria rawlinsi* at low elevation and *Anacroneuria guambiana* at mid-elevation.

Although baetid mayflies were easily collected across all elevations, perlid stoneflies were much harder to find

and were altogether absent at high elevation, so latitudinal comparisons between stoneflies are at mid and low

elevations only. We conducted experiments between January and April of 2013 in the Andes and June to August

of 2013 in the Rockies.

Table 1. Average stream temperatures with standard errors and acclimation temperatures for each
temperate (Rocky Mountain) and tropical (Andes) stream from which insects were collected.

Latitude	Site ID	Type	Elevation (m)	Stream temperature (°C)	Acclimation temperature (°C)
Temperate	COP2212	Low	2212	12.5 ± 3.73	14
	COP2590	Mid	2590	10.8 ± 4.68	10
	COP2798	Mid	2798	7.6 ± 1.41	10
	COP3166	High	3166	6.2 ± 3.56	6
Tropical	ECP1845	Low	1845	13.5 ± 0.72	14
	ECP2003	Low	2003	14.0 ± 0.38	14
	ECP2694	Mid	2798	10.0 ± 0.79	10
	ECP3683	High	3683	8.6 ± 0.89	6
	ECP3898	High	3898	6.6 ± 1.74	6

Lab Acclimation

Field-caught nymphs were brought back to the lab (~1600 m at both latitudes) and placed in

containers of filtered stream water from their native streams without food for 48 h. During this time, high

oxygen levels were maintained by bubbling air into the bath, and water was circulated by small pumps,

with temperatures approximating the insects' home stream temperatures (Table 1). Specifically,

acclimation temperatures were chosen to closely match the average temperature of both temperate and

tropical stream pair at each elevation. Temperate and tropical high elevation insects were acclimated to 6

°C, mid-elevation insects to 10 °C, and low elevation insects to 14 °C (Table 1). Acclimation

temperatures did not exactly match average stream temperatures (Table 1) due to the difficultly of

maintaining precise temperatures in large water baths with our heating and cooling equipment, especially

in the tropics. Notwithstanding these limitations, our acclimation temperatures were between 0 - 2.6 °C

from the average stream temperature at any given site. We selected this paired elevation design because

tropical insects failed to acclimate to temperatures outside average temperatures of their native streams

(see also Shah, Funk, & Ghalambor, 2017a). For example, we often detected evidence of stress (e.g.,

jerking movements, unnatural swimming patterns) and even mortality, especially when tropical mayflies

from high and low elevations were warmed outside native stream temperature (i.e., 12 °C and 10 °C,

respectively). Therefore, the use of a single acclimation temperature across all elevations was not

possible. Because acclimation temperatures differed among elevations but were the same for each

elevation pair, we restricted our comparisons to the same elevations between the two latitudes to control

for the background acclimation temperature.

Metabolic Rate

Oxygen consumption rates (metabolic rates, hereafter) were measured using closed glass micro-

respirometry chambers (Unisense A/S, Aarhus, Denmark). After the 48 h acclimation period, we

introduced insects randomly to one of nine test temperatures (5, 7.5, 10, 12.5, 15, 17.5, 20, 22.5, or 25 °C)

by allowing temperature to rise naturally or by slowly cooling (0.5 °C^{-min}) to prevent thermal shock. On

average, 16 individuals per population for mayflies (min = 3, max = 22) and 6 per population for

stoneflies (min = 3, max = 12) were tested at each temperature (Tables S1, S2). Each individual was

tested at only one test temperature. Once at the test temperature, insects were moved individually into

glass chambers filled with stream water that had been treated with UV light to reduce background

microbial respiration and immersed in a water bath held at the test temperature (Fig. S1). Mayflies were

placed in 4 mL chambers, and stoneflies, which are larger, were placed in 50 mL chambers. Chambers

were fitted with a glass-coated magnetic stir bar and a plastic mesh to which the insect could cling (Fig.

S1). We acclimated insects to the chamber space for an additional hour before sealing each chamber with

a glass stopper to prevent further introduction of atmospheric oxygen. Each stopper had a central capillary

hole, allowing us to insert an oxygen microelectrode (500 μm diam.) connected to a picoammeter

(Unisense OXY-Meter, Aarhus, Denmark) to measure change in oxygen concentration. To measure

background respiration rates, a control chamber (without an insect) was included in each experiment. The

microelectrode was first lowered into the control chamber at the start of the experiment. After reading

oxygen concentration continuously for 2 minutes, it was moved to the first chamber that contained an

animal, where data were recorded for another 2 minutes (Fig. S1), and so on. Experiments were

terminated when oxygen levels reached 80% of the air-saturated values because natural dissolved oxygen

levels in our study sites rarely fall below this value. Consequently, we made 3-6 readings for each

chamber – fewer readings at the higher temperatures due to the combined effect of lower oxygen

concentrations and faster oxygen consumption by insects – which lasted around 128 minutes. Because

there were typically 16 chambers running simultaneously, the average time interval between readings was

25 mins. While recording oxygen concentration, we also observed insects inside the chambers, noting any

visible behavioral stress responses or mortality. Insects were then removed from chambers, euthanized in

ethanol, dried, and weighed on a laboratory scale with readability to 0.01mg.

Statistical Analyses

We first calculated average rate of oxygen consumption for each insect or control (blank)

chamber by measuring change in oxygen concentration across all measurements taken in a given chamber

(see Supplemental Materials and Fig. S2 for an explanation of this calculation). We then averaged the

control values for each temperature such that we calculated a control rate for each combination of test

temperature, elevation, and latitude. The mean control rates were subtracted from individual insect

consumption rates to correct for background microbial respiration at that temperature. We conducted all

statistical analyses on these 'control-corrected oxygen consumption rates' (hereafter, oxygen

consumption) in R version 3.4.0 (R Core Team 2017). For each taxon, latitudinal contrasts were

conducted separately for each elevation. Thus, for mayflies, we compared temperate and tropical baetids

in three statistical comparisons: low, mid, and high elevation. Similarly, for stoneflies, we ran two

separate analyses for low and mid elevation.

Because we were interested in the thermal sensitivity of temperate and tropical insect metabolic

rates between elevation pairs (i.e. the thermal reaction norm), we first compared metabolic rates during

the ascending phase of the TPC by fitting a linear model in R with oxygen consumption as the response

variable for each elevation type (i.e. high, mid, and low) separately. For this analysis, we used only the

temperatures 5, 7.5, 10, 12.5, 15 and 17.5 °C because these temperatures were within the ascending phase

of the SMR curves for all populations in this study. The model included fixed effects of test temperature

and latitude, and their interaction, with body mass as a covariate (Packard & Boardman, 1999). A

significant interaction would denote a difference in thermal sensitivity (i.e. slope) between temperate and

tropical species.

We conducted a similar analysis for the descending phase using data from the 17.5 and 20 °C

treatments as these two temperatures encompassed the descending phase of the curve. At the highest

temperatures (22.5 and 25 °C), we observed stress responses (e.g. unnatural movement, frenzied attempts

to escape from the chamber and in some cases elevated metabolic rates) in tropical mayflies. We therefore

did not include SMR at 22.5 and 25 °C in this analysis but instead calculated separate Q_{10} values for these

temperatures (see below).

To compare the maximum SMR at each elevation between temperate and tropical taxa, we

examined latitude x elevation combination individually to find the physiologically relevant peak (i.e.,

before CT_{MAX} and when the first signs of stress were apparent, if observed). This peak was typically at

17.5 °C in our study because in most cases, across both latitudes, SMR was highest at this temperature

even if not always statistically different from 15 °C. We then compared these values using an ANOVA

for each elevation pair using latitude as the fixed effect and body mass as a covariate.

Finally, we calculated Q_{10}, a measure of sensitivity to temperature, using the formula

$$Q_{10} = \left(\frac{R_2}{R_1}\right)^{\left(\frac{10}{T_2 - T_1}\right)}$$

where R_1 and R_2 are reaction (oxygen consumption) rates measured at two different temperatures, T_1 and

T_2, respectively. Q_{10}s were calculated across cooler, 'non-stressful' temperatures ($T_1 = 7.5$ and $T_2 = 15$ °C)

to ensure measurements were within the exponential phase of the metabolic curve and warmer, 'stressful'

temperatures ($T_1 = 15$ and $T_2 = 25$ °C) to capture the relative decline in rates.

Results

Behavioral observations

Tropical mayflies showed visible signs of stress (jerking movements, loss of righting response) at

several test temperatures outside the range typically experienced (e.g., 22.5 and 25 °C; Fig. 2 red bars,

Tables S1; Fig. S3 A); many of the stressed individuals died at the end of the experiment (Table S1; Fig.

S3 B). Low elevation tropical mayflies also experienced some mortality at 5 °C, which is much colder

than the minimum temperatures they encounter in their home streams (Fig. 2F). Temperate mayflies, in

contrast, did not show visible signs of stress at any experimental temperature (Table S2). We also did not

detect any overt signs of stress in any of the tropical or temperate stoneflies in this study.

Thermal sensitivity based on standard metabolic rates and reaction norms

SMR profiles differed between temperate and tropical mayflies and stoneflies at all elevations

(Figs. 2 and 3). In general, metabolic rates of mayflies from the two latitudes were lowest at temperatures

approximating those in their native streams. Outside these temperatures, however, tropical mayflies

exhibited a greater increase in metabolic rate (Fig. 2). In all mayfly populations across both latitudes, a

secondary peak was observed at 22.5 and 25 °C that corresponded with visible stress during experiments.

We denoted the initial peak in SMR as T-MR $_{PEAK}$ (shown as yellow arrows on Figs. 2 and 3) or the onset

of stress, and the secondary peak as an acute stress response because it was followed by death in some

cases. As indicated in the Methods, we excluded these stressful test temperatures from the analyses of the

ascending and descending phases of metabolic rate profiles and dealt with them separately in an analysis

of Q_{10}.

Figure 2. Plots of standard metabolic rate least square means in baetid mayflies from high (A, B), mid
(C,D), and low (E,F) elevations in temperate (left panel) and tropical (right panel) streams. Red bars
indicate temperatures at which we observed stress. T-MR$_{PEAK}$ is shown with yellow arrows for each
population. Gray rectangles represent the approximate stream temperature range measured in each stream.
CT$_{MIN}$ and CT$_{MAX}$ determined in a previous study are shown.

Figure 3. Plots of metabolic rate least square means in perlid stoneflies from mid (A, B) and low (C,D) elevations in temperate (left panel) and tropical (right panel) streams. Stoneflies were not found at high elevation. T-MR$_{PEAK}$ is shown with yellow arrows for each population. Gray rectangles represent the approximate stream temperature range measured in each stream. We did not detect overt stress in stoneflies in this study. CT$_{MIN}$ and CT$_{MAX}$ determined in a previous study are shown.

In the ascending phase, metabolic rates in tropical mayflies changed more rapidly with (i.e., were more sensitive to) temperature increases than were their temperate counterparts when all elevations were combined ($F_{(5,371)} = 8.57$, $p < 0.001$). We also found the same result when testing each elevation separately, where tropical species had greater increases in metabolic rate at high elevation ($F_{(5,97)} = 3.27$, $p = 0.009$, Fig. 4A), mid elevation ($F_{(4,98)} = 4.17$, $p = 0.003$, Fig. 4B), and at low elevation ($F_{(4,152)} = 6.77$, $p < 0.001$, Fig. 4C). In stoneflies, ascending phases did not differ between temperate and tropical populations (Figs. 5A and B)

Temperate mayflies had a steeper decrease in metabolic rate at high elevation (Fig. 4A; $F_{(1,1)} = 4.04$, $p = 0.05$) and at low elevation (Fig. 4C; $F_{(1,1)} = 6.86$, $p = 0.01$). At mid elevation, there was no

significant interaction (Fig. 4B). However, a Student's T-test revealed that oxygen consumption values

were significantly higher in tropical mayflies compared to temperate ones at 17.5°C ($t = 3.946$, $p =$

0.001), but rates were equally low between latitudes at 20°C ($t = -1.006$, $p = 0.33$). This suggests that the

descending phase was steeper for tropical mayflies compared to temperate ones. Descending phases of

stoneflies did not differ between tropical and temperate populations (Figs. 5 A & B).

Figure 4. A comparison of the ascending (left panel) and descending (right panel) phases of metabolic
profiles in baetid mayflies at high (A), mid (B) and low (C) elevations. Values are the least square means
of metabolic rate in mayflies from tropical (white circles) and temperate (black circles) streams, after
accounting for body mass in the model. Vertical bars are standard errors and p values are shown.

Figure 5. A comparison of the ascending (left panel) and descending (right panel) phases of metabolic
profiles in perlid stoneflies at mid (A), and low (B) elevations. Values are the least square means of
metabolic rate in stoneflies from tropical (white circles) and temperate (black circles) streams, after
accounting for body mass in the model. Vertical bars are standard errors and p values are shown.

Finally, the maximum SMR was higher in tropical compared to temperate mayflies at all

elevations (Fig. 6). Stoneflies showed an opposite pattern where tropical species had lower peaks at both

measured elevations, although this was significant only at mid elevation (Fig. 6).

Figure 6. Peak SMRs (i.e. the highest metabolic rate) are compared between temperate and tropical baetid mayflies from high (A), mid (B), and low (C) elevations and perlid stoneflies from mid (D) and low (E) elevations. Vertical bars are standard errors and p values are shown.

Thermal Sensitivity based on Q_{10}

When comparing Q_{10}s across 'stressful' and 'non-stressful' temperatures, we found Q_{10}s for both

taxa to be higher at non-stressful temperatures than at stressful temperatures (Table 2). Across latitude,

tropical mayflies showed consistently greater sensitivity to temperature (higher Q_{10} values) at both non-

stressful and stressful temperatures than their temperate counterparts (Table 2; Fig. 7). Notably, at mid

and low elevations, tropical mayflies were highly sensitive to temperature even at non-stressful

temperatures (Q_{10} = 8.40 and 5.17, respectively). In contrast, tropical stoneflies were less sensitive than

their temperate relatives for stressful and non-stressful temperatures (Fig. 7).

Table 2. A comparison of Q_{10} values calculated for temperate and tropical mayfly metabolic rates
between typically non-stressful temperatures (7.5°C to 15°C) and typically stressful temperatures (15°C
to 25°C).

		Mayflies		*Stoneflies*	
	Elevation	**Temperate**	**Tropical**	**Temperate**	**Tropical**
Non-stressful	High	3.63	3.15	n/a	n/a
	Mid	3.15	5.17	6.76	2.49
	Low	2.36	8.40	6.66	5.78
Stressful	High	1.07	1.46	n/a	n/a
	Mid	0.73	1.59	6.10	0.87
	Low	1.34	1.67	1.25	0.42

Figure 7. Comparison of average Q_{10} for mayflies and stoneflies during the ascending phase of the
metabolic rate curve.

Discussion

Understanding the influence of temperature on the minimum energy demands in ectotherms is

critical to predictions of how global warming will impact different species (Huey & Kingsolver, 2019).

Our study provides a first look at how energy demands of temperate and tropical aquatic insects might

respond to warming streams, by measuring standard metabolic rate. Although we cannot generalize our

results to all temperate and tropical aquatic insect groups, the widespread distribution of the insect

families chosen here and replication of our experiments across elevation provide new insights into the

sources of variation in thermal sensitivity of standard metabolic rate. We observed this variation in the

two different taxa in our study – mayflies and stoneflies – which responded differently to temperature

changes even when they originated from the same streams.

Comparative studies of critical thermal limits have generally found tropical ectotherms to be

adapted to narrower ranges of temperatures compared to their temperate counterparts, suggesting that

tropical species are more vulnerable to global warming (Deutsch et al., 2008; Gutiérrez-Pesquera et al.,

2016; Shah et al., 2017b; Sunday, Bates, & Dulvy, 2011, 2012), a finding consistent with the CVH. Payne

and Smith (2017) recently challenged such conclusions by arguing that narrower thermal breadth may

simply reflect the acute effects of temperature on biological rates of organisms living in warmer

environments. Resolving this debate requires an understanding of the acute effects of temperature on

continuous biological rates like metabolism (Havird et al., 2020). Here, using a paired-elevation design,

we compared the thermal sensitivity of standard metabolic rate (SMR) in closely related temperate and

tropical aquatic insect nymphs. This study design enabled a comparison of temperate and tropical species

under a common range of field, acclimation, and test temperatures. Our results did not support the Payne

and Smith (2016) model, but provided stronger support for the CVH.

Despite originating in streams with similar average environmental temperatures at the time of

collection, tropical and temperate insects had different metabolic rate profiles (Figs. 2, 3). These results

indicate that the variation between temperate and tropical species in our study does not reflect simple,

acute effects of temperature on biological rates, but rather that there are physiological differences between

taxa from these regions. Metabolic rates of tropical baetid mayflies, in particular, were more sensitive to

temperature, and they exhibited stress at lower water temperatures compared to their temperate

counterparts. By contrast, tropical and temperate perlid stoneflies showed little to no divergence in their

thermal sensitivity. The ecological implications of greater thermal sensitivity remain unknown for most

ectotherms, but in the absence of compensatory changes in productivity, elevated metabolic rates could

negatively impact the energy budget and capacity for aerobic performance in these tropical mayflies by

increasing baseline energetic costs (Dillon et al., 2010; Huey & Kingsolver, 2019).

Comparison of metabolic rate profiles

In most cases, metabolic rate rose with increasing temperature, peaked, and then fell. This

pattern, however, varied considerably among taxa and populations and across elevations. As a group, the

tropical mayflies are more thermally sensitive than their temperate counterparts (Figs. 2, 4, 6 and 7). The

metabolic rate curves of tropical mayflies had overall steeper ascending and descending phases, and

consistently higher Q_{10}s, across all three elevations. The particularly high Q_{10} s for mid and low elevation

tropical mayflies were similar to those of North Atlantic stenothermal fishes when experiencing

temperatures away from an optimum (Johnston, Clarke, & Ward, 1991). We note that high elevation

tropical mayflies had similar Q_{10}s to their temperate relatives (Table 2), which may reflect the higher diel

variation (i.e. variation across 24 h) experienced in streams above tree line in the Andes (higher standard

error for stream temperature Table 1). In seasonally stable streams, such diel effects may play a

potentially important role in regulating metabolic activity of aquatic insects (Gilchrist, 1995), but to date,

no studies have explored the relative effects of diel and seasonal thermal variation in freshwater

ectotherms. In general, however, our results support the prediction that tropical mayflies, which

experience more stable stream temperatures year-round (Shah et al., 2017b), are more sensitive to

temperatures outside the ranges they normally experience.

Given that mayflies were paired by elevations having similar mean environmental and

acclimation temperatures, these differences cannot be attributed simply to the acute or passive response of

metabolic rate to higher temperatures (Havird et al., 2020). Instead, these differences appear to reflect

divergence in thermal physiology as shown by the greater thermal sensitivity of metabolism in tropical

mayflies (Figs. 2, 4, 6, 7). By contrast, temperate mayflies exhibited more modest increases in

metabolism in response to temperature and had similar metabolic rate profiles across elevations (Fig. 2).

For both temperate and tropical mayflies, metabolic rates tended to be lower within the range of

environmental temperatures they naturally experienced (Fig. 2), suggesting there may be selection to

reduce the minimum energetic costs associated with maintenance (Artacho & Nespolo, 2009;

Bochdansky, Grønkjær, Herra, & Leggett, 2005; Brown, Marquet, & Taper, 1993). For example, even a

moderate increase in stream temperature outside the normal range (Fig. 2, right edge of grey rectangles) is

associated with peaks in metabolism, which has potentially cascading effects on energy budgets and

possible chronic stress (Fig. 2). If selection acts to minimize SMR under typical stream temperatures

(Brown et al., 1993), these results would support the hypothesis that the narrower and more stable stream

temperatures experienced by tropical mayflies make them more stenothermic and underlie their greater

thermal sensitivity, as predicted by the CVH.

Such an interpretation is consistent with other aspects of temperate and tropical mayfly thermal

physiology. First, thermal breadths (measured as the difference between critical thermal limits) reported

in a parallel study were narrower for tropical (Ecuador) versus temperate (U.S.A) mayfly populations

(Shah et al., 2017b). Second, in this study, we observed visible signs of stress in the metabolic rate

profiles of tropical but not temperate mayflies. Specifically, after the decline in metabolic rate, secondary

peaks occurred at 22.5 °C and 25 °C for most groups, which corresponded to observed signs of behavioral

stress and mortality in tropical but not temperate mayflies. Secondary peaks are relatively rare in

metabolic rate measurements but can occur, often near the critical thermal limits (Havird et al., 2020).

Mayflies showed signs of behavioral stress at these high temperatures, including leg spasms, loss of

righting, release from substrate, and mortality (see Supplemental Information Table S1, Figs. S3 A, B).

Collectively, these results support the idea that temperate mayflies are typically thermal generalists, with

broad thermal breadths and reduced metabolic sensitivity to temperature, whereas tropical mayflies are

thermal specialists, with narrower thermal breadths and increased metabolic sensitivity to temperature

(c.f. Huey & Hertz, 1984).

The differences observed between temperate and tropical mayflies were generally unambiguous

and consistent with prior work in this system. The lack of divergence between temperate and tropical

stoneflies is also consistent with previous research on other physiological traits (Shah et al., 2017a,

2017b). Although there are differences in metabolic rate profiles between temperate and tropical

stoneflies (e.g. higher metabolic rates at warmer temperatures in temperate, but not tropical species), these

differences are far less striking than those observed in mayflies (Fig. 2 vs. 3). Overall, the ascending and

descending phases did not differ between temperate and tropical stoneflies (Figs. 3, 5), but absolute

maximum metabolic rates and Q_{10}s across non-stressful and stressful temperatures were higher in the

temperate species (Fig. 7). Shah et al. (2017a) also found a lack of latitudinal differences in stoneflies

when comparing acclimation abilities. One of the few other studies on the respiration rates of tropical

stoneflies reported similar Q_{10} values to those reported here for non-stressful temperatures in mid-

elevation stoneflies (Jacobsen & Brodersen, 2008), corroborating our conclusions of lower thermal

sensitivity in at least this group of perlid stoneflies. Though many tropical species will be imperiled by

warming (Deutsch et al., 2008; Dillon et al., 2010), the overall mixed support for this prediction in

stoneflies cautions against generalizing about the vulnerability of all tropical species.

Numerous hypotheses could explain the lack of divergence between temperate and tropical

stoneflies. First, stoneflies as a group have a temperate origin and invaded tropical streams relatively

recently (Baumann & Kondratieff, 1991; Stewart, Baumann, & Stark, 1974). Studies suggest that

stoneflies are still adapting to tropical habitats (Polato et al., 2018), which may explain why thermal

physiological traits in present-day temperate and tropical populations are similar.

Second, perlid stoneflies and baetid mayflies have different life histories, which may contribute to

differential sensitivity to temperature. Stoneflies often live much longer in streams as nymphs than

mayflies (which have multiple generations per year), and may experience years of unpredictable variation

in stream temperature (Clifford, 1982). In fact, many stoneflies enter diapause, unlike mayflies (Brittain,

1982; Hynes, 1976), giving them a greater ability to withstand adverse thermal conditions (Brittain,

1990). Once emerged, adult stoneflies often remain close to natal streams, whereas adult mayflies in

tropical (Finn, Encalada, & Hampel, 2016) and temperate (Polato et al., 2018) regions can disperse more

widely. Therefore, stoneflies rely more heavily on the nymph stage for dispersal (Brittain, 1990) and as a

consequence, may experience more thermal variation while moving up- or downstream than juvenile

mayflies. This increased likelihood of encountering thermal variation may favor the evolution of

decreased thermal sensitivity (Muñoz & Bodensteiner, 2019).

Third, perlid stoneflies are important predators of baetid mayflies and midges (Peckarsky, 1980;

Tamaris-Turizo, Turizo-Correa, & del Carmen Zúñiga, 2007) and they actively pursue prey in the wild

(Peckarsky, 1980) and in laboratory settings (Sjöstrom, 1985; A.A.Shah *pers. obs.*). A reliance on aerobic

metabolism and the need to maintain a high metabolic scope for active hunting (Kordas, Harley, &

O'Connor, 2011) could constrain physiological divergence between temperate and tropical stoneflies.

Unfortunately very few comparative studies of metabolic profiles are available to test the generality of

such hypotheses. Nevertheless, higher maximum metabolic rates (Fig. 6) and higher Q_{10}s (Fig. 6) do

suggest some differences between temperate and tropical populations that could have ecological

consequences when pursuing prey across a broad range of temperatures. Because perlid stoneflies and

baetid mayflies interact as predator and prey in temperate and tropical streams (Gamboa, Chacón, &

Segnini, 2009; Peckarsky, 1980), differential sensitivities to temperature may have community-level

consequences (Gilman, Urban, Tewksbury, Gilchrist, & Holt, 2010; Grigaltchik, Ward, & Seebacher,

2012; Kordas et al., 2011; Pincebourde & Casas, 2019; Vucic-Pestic, Ehnes, Rall, & Brose, 2011). Our

results underscore the importance of understanding the physiological responses of key species in order to

assess community-level responses to global warming. Further explicit testing of predator-prey interactions

under different temperatures (Barton & Schmitz, 2009) will provide greater insight into how these

responses differ across latitude (Matassa & Trussell, 2015)

Comparison of metabolic rates and critical thermal maxima as indicators of vulnerability

Some of the results of the present study are surprising because one should expect CT_{MAX}, a

commonly used metric for thermal sensitivity (Diamond et al., 2012; Lutterschmidt & Hutchison, 1997;

Piyaphongkul, Pritchard, & Bale, 2012; Ribeiro, Camacho, & Navas, 2012), to coincide with the

descending limb of the metabolic rate profiles (Fig. 1). Yet, previously measured values of CT_{MAX} in

temperate mayflies occur at much higher temperatures (Fig.1A, B, C). For example, CT_{MAX} was ~29 °C in

high elevation temperate baetid mayflies (Shah et al., 2017b) but they began to show elevated metabolic

rates and behavioral signs of stress at lower temperatures (~17.5 °C). The large difference between

CT_{MAX} and the onset of stress in this study is likely due to methodology, as rapid ramping or high starting

temperatures can inflate estimated values of CT_{MAX} (Rezende, Tejedo, & Santos, 2011; Terblanche,

Deere, Clusella-Trullas, Janion, & Chown, 2007). In addition, the discrepancy could also arise because

CT_{MAX} and metabolic rates are governed by fundamentally different processes. CT_{MAX} is the result of

changes in fluidity of cell membranes and whole-organism physiology in response to heat stress (Cossins,

Friedlander, & Prosser, 1977; Hochachka & Somero, 2002), whereas standard metabolic rates arise from

different activation energies of enzymes and sensitivity of biochemical processes to temperature (Rolfe &

Brown, 1997). Thus, using CT_{MAX} in conjunction with other measures of thermal sensitivity such as

metabolic rate provides a more complete assessment of thermal tolerance.

Climate change in Andean and Rocky Mountain Streams

Mountain stream temperatures are already increasing worldwide (Isaak & Rieman, 2013; Niedrist

& Füreder, 2020). Temperatures of shallow mountain streams, such as those in this study, are also likely

to show concomitant increases with air temperature (Mohseni & Stefan, 1999; Pilgrim, Fang, & Stefan,

1998). Projections of mean air temperature increases of 2-3 °C in the Andes and 3-4 °C in the northern

Rocky Mountains by the 2050s (Nogués-Bravo, Araújo, Errea, & Martínez-Rica, 2007; Rangecroft,

Suggitt, Anderson, & Harrison, 2016) are therefore troubling. However, previous analyses suggest that

aquatic insects of the southern Rocky Mountains are relatively robust to warming because they may

currently occur at suboptimal temperatures (Pyne & Poff, 2017). Moreover, mesocosm and field common

garden experiments showed that temperate baetid mayflies grow faster when subjected to warmer water

(N.L. Poff et al., *unpubl. data*). Given the wider elevation distribution and broad dispersal ability of these

insects (Gill et al., 2016), they may be able to move across the landscape to find thermally suitable

habitats as their environment warms. In contrast to baetid mayflies of the Rockies, Andean mayfly growth

declined with increasing temperature in mesocosms (N.L. Poff et al., *unpubl. data*) further supporting our

conclusion that a warming environment will negatively affect tropical mayfly fitness, especially at low

elevation. Local temperatures may also become higher in some Andean streams where riparian forests are

being cleared for human activities. The resulting exposure to increased solar radiation can alone raise

stream temperature (Moore, Spittlehouse, & Story, 2005).

The tropical mayflies in our study will experience the effects of these increasing temperatures,

which can also lead to a possible mismatch of dissolved oxygen supply and demand (Pörtner, 2001;

Verberk, Bilton, Calosi, & Spicer, 2011; Verberk et al., 2016b). The ability to maintain an adequate rate

of oxygen uptake decreases at higher temperatures for many aquatic insects (Jacobsen & Brodersen,

2008; Rotvit & Jacobsen, 2013), and reduces energy available for activities like movement, growth, and

egg development. Thus, streams need not increase too much in temperature to have a negative impact on

insect metabolism and growth. This is illustrated in a field experiment, where a moderate 2 °C

temperature increase in a stream resulted in declines in abundance of Eurasian mayflies, presumably as a

result of decreased oxygen in the stream (Verberk, Durance, Vaughan, & Ormerod, 2016a).

The thermally sensitive tropical mayflies in our study system may be forced to seek cooler

temperatures at higher elevations (Jacobsen, 2020), but whether they can move upstream is debatable

given the low dispersal rates of aquatic insects in these streams (Polato et al., 2018). Further, it is not

known if aquatic insects have any ability to mitigate the effects of warming through behavior such as

seeking out microhabitats having relatively high oxygen concentrations (e.g., fast-flowing water). Future

work should therefore also focus on whether aquatic insects can move upstream to cooler waters or seek

out high flow microhabitats to counteract the effects of warming in their native streams (Shah, Dillon,

Hotaling, & Woods, 2020; Sheldon, 2012).

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Author contributions

A.A.S. and C.K.G. designed the experiments. A.A.S. conducted fieldwork and experiments, analyzed

results, and wrote the manuscript with contributions from all authors.

Conflicts of Interest

None

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