Quantifying the dynamics of pig movements improves targeted disease surveillance and

control plans

Running title: Time-varying networks improve surveillance in pigs

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Summary

Tracking animal movements over time can fundamentally determine the success of disease

control interventions throughout targeting farms that are tightly connected. In commercial pig

production, animals are transported between farms based on growth stages, thus it generates

time-varying contact networks that may influence the dynamics of disease spread. Still, risk-

based surveillance strategies are mostly based on static network. We reconstructed the static and

temporal pig networks of one Brazilian state from 2017 to 2018, comprising 351,519 movements

and 48 million transported pigs. The static networks failed to capture time-respecting movement

pathways. Therefore, we propose a time-dependent network susceptible-infected (SI) model to

simulate the temporal spread of an epidemic over our pig network globally through the temporal

movement of infected animals among farms and locally with a stochastic compartmental model

in each farm, configured to calculate the minimum number of farms needed to achieve effective

disease control. In addition, we propagated disease on the pig temporal network to calculate the

cumulative contacts as a proxy of epidemic sizes and evaluated the impact of network-based

disease control strategies. The results show that targeting the first 1,000 farms ranked by degree

would be sufficient to control disease spread. Our finding also suggested that assuming a worst-

case scenario in which every movement could transmit disease, pursuing farms by degree would

limit the transmission to 29 farms, which is lower than the number of infected farms for random

surveillance, with epidemic sizes of 2,593 farms. Overall, the proposed modeling framework

provides a parsimonious solution for targeted disease surveillance when temporal movement data

is available. Finally, the top 1,000 farms ranked by degree could benefit from enhanced

biosecurity plans and improved surveillance, which constitute important next steps in

strategizing targeted disease control interventions.

Key words: swine movement data, dynamic contact networks, disease surveillance, SI

models.

Introduction

Between-farm animal movement has historically been associated with disease spread (Ortiz-

Pelaez et al., 2006; Smith et al., 2013). Animal movement data has allowed for the initial

development of network-based approach, which are dominated by the use of static network

metrics to identify vulnerable farms. In a contact network, each farm (node) can play a role in the

dynamics of disease spread, and some can be classified as vulnerable because they periodically

receive infected animals (a high "in" degree); therefore, the identification of these farms is more

important in scenarios of the absence or low prevalence of disease (Qi et al., 2019). Such

locations may serve as sentinels. On the other hand, farms that are more likely to spread disease

throughout their contact network could be targeted, by which restricting or monitoring outgoing

movements could effectively reduce disease spread (Cárdenas et al., 2018; Payen et al., 2019).

Ultimately, the use of network-based metrics to select farms for isolation or surveillance schemes

has shown great value (Mohr et al., 2018; Firestone et al., 2019; Payen et al., 2019). This is a

direct consequence of the availability of animal movement data (Büttner et al., 2016; Firestone et

al., 2019); however, methodologies have not yet evolved as expected. Most published

approaches make use of static networks to describe animal movement and subsequently make

inferences about outbreaks and epidemics (Büttner et al., 2013, 2018; Marquetoux et al., 2016;

Motta et al., 2017; Alarcón et al., 2019; Kinsley et al., 2019). However, the temporal anomalies

and seasonality of movements can only be identified if movement paths are aggregated to

smaller time windows (Cárdenas et al., 2018; Augusta et al., 2019; Sterchi et al., 2019), which

may not be sufficient to make inferences about disease dynamics within the contact networks.

Disease transmission is not restricted to the historical record of movement (Firestone et al., 2019;

Qi et al., 2019; Sterchi et al., 2019); namely, when disease transmission actually occurs by the

movement of infected animals is likely to be an event in time that is dependent on when animals

are sent and, more importantly, when they are received at the final destination, and this rationale

has been largely overlooked (Chaters et al., 2019; Sterchi et al., 2019). Nevertheless, static

networks have been used to measure the effect of targeted farm removal, on the network

fragmentation by monitoring giant strongly or weakly connected components, which would

create small islands restricting the overall connectivity among farms (Motta et al., 2017;

Mekonnen et al., 2019). Other studies have proposed the use of the basic reproduction number

(R_0) and have measured the contribution of node removal in disease transmission (Marquetoux et

al., 2016; Alarcón et al., 2019; Passafaro et al., 2019), still based on static views of full networks.

Therefore, it is recognized that the current neglected temporality of real-world animal

movements has been a constant roadblock to the advancement of network epidemiology,

especially in the development of methodologies (Pellis et al., 2015).

Stochastic simulations have been used extensively to measure the impact of disease control

strategies in future disease spread scenarios (Smith et al., 2017; Funk et al., 2018; Kim et al.,

2019; Lanzas et al., 2020). Mathematical models have also been used to experimentally spread

disease over contact networks and reveal important dynamic characteristics of epidemics (Miller,

2017; P. Kim and Lee, 2018; Darbon et al., 2019). For example, compartmental models can be

used to describe the epidemic spreading in temporal contact networks, which provides an

opportunity for tracing outbreaks and contact among nodes over time (Guinat et al., 2016;

Colman et al., 2019; Ferdousi et al., 2019; Sterchi et al., 2019). Here, we used two years of pig

movement data from one Brazilian state to estimate the optimal number of farms for target

surveillance by tracking between-farm contact pathways and inferring epidemic sizes throughout

disease propagation. We first described the complete pig movements and then challenged the use

of static networks against dynamic networks. We proposed a stochastic compartmental model to

simulate disease spread in a two-year empirical networks while accounting for the temporal

order of the movements. Our model formulation allowed the calculation of the minimum number

of farms needed to reduce disease spread and approximate the impact of network-based disease

control strategies (e.g., farm isolation, movement restriction, enhanced biosecurity) on the

expected epidemic sizes. Finally, we applied the proposed model to the time-varying pig

network, ultimately to provide a practical background for the identification of targeted farms,

which were then mapped and described in regard to their biosecurity and infrastructure.

Material and methods

Data collection and entry

The record of pig shipments among all registered farms of one Brazilian state was used to

reconstruct contact networks from 2017 until 2018 (SEAPI-RS, 2018). Information about each

swine farm included farm identification, production types (breeding, certified-swine-breeder,

sow, nursery, wean-to-finisher, finisher, subsistence and others), farms infrastructure (e.g.,

presence of a fence, presence of a cleaning and disinfection station, see table 1 for the full list),

geolocation and farms population capacity. Each pig shipment included date(s), farm of origin

and destination, number of animals transported and purpose of movement (weaning, finishing,

slaughter or other). Movement data with missing information of farm location, production type,

or farm of origin or destination were excluded prior to the analysis. Additionally, farms declared

inactive (no pigs raised in the past two years or out of business) were not considered in the

analysis. Movements from or to other states were also excluded from any analysis. Finally, farms

were classified as commercial if they had active contracts with integrated swine companies and

as noncommercial otherwise. From the total number of movement active pig farms 11,849, a

subset of farms 9,500 had registered geographic coordinates, therefore were used for the

simulation modeling subsections.

Complete and monthly networks reconstruction and analysis

Two directed networks were reconstructed. First, a static network with directed "edges"

represented by pig shipments between two "nodes", where nodes corresponded to pig farms. The

static network, movements from January 2017 to December 2018 were analyzed by the

traditionally method of aggregating movements into one snapshot. Second, networks were

constructed based on a temporal resolution of 30 day windows (Lentz et al., 2016). For both

complete and monthly network, centrality metrics were calculated (Table 1) (Cárdenas et al.,

2018). As slaughterhouses constitute sinks and we were interested in potential disease spread

among living animals, transports to slaughter were excluded from the network analysis, but are

kept for descriptive purposes (see figure 2).

Temporal variability of the network structure

Static networks are considered an approximation of all movement paths for a defined period,

e.g., animal movement between pig farms for one month (Büttner et al., 2013; Lentz et al.,

2016). This can artificially create accessibility paths between nodes that are not always

accessible by all "nodes" in a network, which can lead to an overestimation of the connectivity

among the nodes (Lentz et al., 2016). To quantify the amounts of such static error, the number of

paths in the static view can be compared with the number of paths in the time-series network

(Lentz et al., 2013, 2016). This ratio is known as the causal fidelity "c", where:

$$c = \frac{p(P_n)}{p(P_n)} = \frac{Number\ of\ paths\ in\ the\ temporal\ network}{Number\ of\ paths\ in\ the\ static\ network}. \tag{1}$$

The number of paths is the number of nonzero elements in p or P. A causal fidelity of one means

that the temporal network is well represented by its static network, while networks with lower

causal fidelity values would not represent the temporal paths well because most of the paths are

not causal. Here, we calculated the causal fidelity on timescales that are typically used to

investigate animal movement data: monthly, 6 months and 1 year and for the full two years of

data (Lentz et al., 2016). We also calculated the edge loyalty (θ), which measures the fraction of

preserved edges (θ) of a given premise between two consecutive years, $t - 1$ and t. To

quantify $\theta_i^{t-1,t}$, we define Y_i^{t-1} as the set of edges from the premise i in the time $t - 1$ and Y_i^t as

the set of edges from the premise i at time t. Then, $\theta_i^{t-1,t}$ is given by the Jaccard Index in

equation 2 (Valdano et al., 2015; Schulz et al., 2017).

$$\theta_i^{t-1,t} = \frac{|Y_i^{t-1} \cap Y_i^t|}{|Y_i^{t-1} \cup Y_i^t|} \tag{2}$$

146   For the node loyalty, we use the same Jaccard Index,

$$node\ loyalty\ _i^{t-1,t} = \frac{|V^{t-1} \cap V^t|}{|V^{t-1} \cup V^t|} \tag{3}$$

where V_i^{t-1} is the set of farms that are active in period $t - 1$, and V_i^t is the set of farms that

moved at least one animal at time t (Valdano et al., 2015; Schulz et al., 2017).

The minimum number of farms needed for effective control disease spreading process over

time-varying network

We created a stochastic susceptible-infectious (SI) epidemic model to calculate the minimum

number of farms needed to effectively reduce the number of new cases within the contact

network. This model framework integrates infection dynamics in each subpopulation as

continuous-time Markov chains using the Gillespie stochastic simulation algorithm (Widgren et

al., 2019). Transitions from $S \rightarrow I$ was in a two-stage model with a one-way transition depending

on $\frac{\beta S_i I_i}{S_i + I_i}$ with β as the transmission rate. The completely stochastic simulation can be summarized

as follows: the empirical movement data from 2017 until 2018 was used to simulate disease

spread over a time-varying network; here, we assume a herd prevalence of 10% and a within-

farm prevalence of 0.1%, which could transmit between and within farms. For the within-farm

transmission rate, we use a set of β values of 0.01, 0.05, 0.1 and 0.2, which can also be

considered the product of the contact rates and the transmission probability between animals into

the farms. Another assumption was that the contact rates among pigs were homogenous. For the

between-farm transmission, we considered the observed number of pigs that moved between

farms and the population size at each farm. As initial condition, model simulation started with

950 infected farms (10% prevalence from a total population of 9,500 farms) at time "0". Here,

we proposed two realistic strategies to seed infected nodes at the time "0". First, pig farms were

selected at random from all farms with at least one movement from 2017 to 2018. For the second

strategy, infected farms were chosen proportionally to the number of farms in each production

system (N=17). This approach allows a more realistic scenario in which disease can emerge at

any location of the study area (see Figure 1). The simulated control intervention was based only

on the sequential removal of farms from the time "0" based on the static network described

above. While farms were removed, the number of new cases per 100 susceptible farms was

calculated. We chose an arbitrary number of 2,000 herds to be removed, and at the end of each

simulation scenario the incidence of new cases I was calculated as follows:

$$I = \frac{Number\ of\ new\ infected\ herds\ in\ 2017 - 2018}{\begin{array}{c}(Number\ of\ herds\ that\ traded\ with\ other\ herds\ in\ 2017 - 2018)\\ -\ (Initial\ number\ of\ infected\ herds\ in\ 2017 - 2018\ that\ traded\ with\ other\ herds\ )\end{array}}$$

Infection was seeded into all candidate farms randomly ten times, from which simulations were

run and the median incidence was derived for each run. For a sensitivity analysis, we ran

simulations with a diverse set of β parameters (see supplementary Figure S2). In addition, we ran

simulations for commercial and noncommercial farms separately with the same parameters

described above (see supplementary figures S4 and S5).

Cumulative contacts as a proxy of epidemic sizes

The modeling setup described by (Payen et al., 2019) was adapted to identify the best network-

based metric to be used in the identification of farms to undergo target surveillance. This

approach is based on a classic deterministic SI, spreading model that follows movements

temporality (Dubé et al., 2008; Nöremark et al., 2011; Konschake et al., 2013). Thereafter, we

will refer to this model as "spread cascade", which is a temporal representation of the cumulative

and consecutive contacts among farms, and therefore can be interpreted as the size of an

epidemic (Payen et al., 2019). The model started from each seed farm in the network at a given

time "0", with an infection that was active for one day. The hypothetical disease spread followed

the chronological order of movements over two years. In short, when there was a directed

interaction, including the three dimensions of time t, infected farm i and susceptible farms j,

disease has successfully been transmitted. These events were counted in favor of each farm i

cascade and the triplet t, i, j and added to the temporal links of the stream; see (Payen et al.,

2019) for more details of the methodology. Overall, we recorded the daily spread cascades,

which are represented by averages and $\pm 95\%$ CI, and we also calculated the frequencies of the

spread cascades sizes for each seed node in the network (n = 9,450) as a proxy for the worst-case

scenario in epidemic propagation (Payen et al., 2019). We used the static network to initially

rank all pig farms in descending order of degree, betweenness, PageRank, and closeness and

then removed the top 10, 250, 500, 750 and 1,000 farms at day "0" of the simulation while

recording the spread cascade sizes for the full two years of movement data. For comparison with

usual surveillance practices, we selected the same number of farms at day "0" randomly. The

contribution on containing the epidemic spread by each network-based target surveillance

strategy was evaluated by comparison of the final spread cascade distributions using a

nonparametric Kruskal-Wallis test followed by Dunn's post hoc test, where the p-values below

5% were significant. Once the most efficient network-based surveillance strategy was defined,

the biosecurity, infrastructure and population of the selected farms were described.

Software

The software used for statistical analysis and graphics was R statistical software (versions 3.4.1,

5.1.3) (R Core Team) with R Studio editor using the igraph 1.2.4 (Csardi and Nepusz, 2006),

tidyverse 1.2.1(Wickham, 2017), SimInf 6.3.0 (Widgren et al., 2019), sf 0.5-3 (Pebesma, 2018),

and brazilmaps 0.1.0 (Siqueira, 2019) packages. Spyder 3.3.3 software for Python version 3.7

and Adobe Illustrator CC 2018 software were also used.

Results

Description of pig movements

The total number of active pig farms in the complete database included 11,849 farms, 351,519

movements and more than 48 million pigs transported. The majority, 66%, were identified as

noncommercial farms, while the number of farms per commercial system varied from 10 to 465

from a total of 17 production systems. Among those farms included 6,555 (55.32%) finisher

farms, 1,816 (15.33%) nurseries, 470 (3.97%) wean-to-finisher farms, 433 (3.65%) breeding

farms, 122 (1.03%) certified swine breeder farm and 250 (2.11%) other farms, which included

insemination stations and isolation units. Finally, among the noncommercial farms, we identified

2,101 (17.73%) as small pig holders (backyard farms or for subsistence) and 102 (0.86%)

slaughterhouses. Most commercial and noncommercial pig farms were located at the northern

and central east of the state (see Figure 1). While we have described movements to

slaughterhouses, they were not accounted in any network analysis thereafter (see Figure 2).

Figure 1. Geographical location of commercial and noncommercial pig farms

More than 51% of movements were from finisher farms to slaughterhouses, with a monthly

median of 7,657 shipments. The second largest number of movements from nurseries to finisher

farms was 13.51%, with a monthly median of 1,992 transfers. Breeding farms made up for

4.65% of the movements to finisher farms and 4.3% of movements to the nurseries, with 673

monthly median movements. Finally, wean-to-finisher farms sent a small but significant amount

of pigs to slaughter (3.5%), while certified swine breeder farms accounted for 2.8% of

movements to slaughter.

Figure 2. Depiction of pig movements among all farm types from January 2017 to December

2018. The arrow size indicates the percentage of movements, and the color is defined by the

origin of the movements. The size of the circles represents the total number of farms.

Monthly network analysis

From the two years of movement data, we reconstructed the monthly static networks. The

number of farms (nodes), movements (edges) and animals varied substantially with a steady

increase from July 2017 until October of the same year (see Figure 3). The network diameter

revealed a positive increasing trend of 33% from April 2018 through December 2018, December

2018, which decreased thereafter; this pattern was different from the monthly values from the

previous year. Regarding the giant network components, from a general view using the static

networks, the giant weakly connected component (GWCC) was formed by 7,563 farms, and the

giant strong connected component (GSCC) had 863 farms; these sizes corresponded to 80.03%

and 9.13% of the total number of farms in the network. In addition, the monthly analysis showed

a positive trend of the GSCC in 2017 and a negative trend in 2018, and the GWCC had a more

stable pattern with two consecutive peaks in July and October of 2018.

Figure 3. Monthly variations in the network metrics of pig movements from January 2017

through December 2018.

Loyalty and causal fidelity

The node loyalty of 2018 compared with 2017 was 0.65, while edge loyalty was 0.19. This

means that only 65% of the farms that traded pigs in 2017 were preserved in 2018, and 19% of

the edges were preserved between 2017 and 2018. On the other hand, the monthly causal fidelity

varied from 0.33 in July 2018 and reached the highest value of 0.55 in July 2017 (see

supplementary Figure S1). For the network that evaluated time periods of six months, the causal

fidelity was 0.22 for the first half of 2017 and 0.47 for the second half of 2018, and the annual

evaluation was in 2017, 0.18 in 2018, and 0.33 for the entire static network. The causal fidelity

demonstrated a discrepancy between static and temporal network views. The concept of time

does not per se exist in these static networks; thus, the use of traversal network models here

would reduce the accuracy toward overestimating disease outbreaks, which would likely mislead

the selection of farms to undergo network-based surveillance (Lentz et al., 2016). In addition, we

calculated a causal error from the causal fidelity, which can be approximated by the inverse of

"c" and represents the overall overestimation of disease outbreaks if static networks are used.

The first six months of 2017 had the worst results, with overestimations at 4.54 and 1.81 for the

July 2018 network.

Number of farms for targeted surveillance and epidemic sizes

The stochastic runs showed that the removal of 2,000 farms did not completely control the

disease from spreading, but it was reduced markedly to a maximum of 1.95 new cases per 100

farms when β of 0.2 was used. Since this study may be used to guide decisions in the field, we

consulted with the local stakeholders before deciding on the final number of farms to be on the

surveillance hot-list (personal communications with Dr. Lopes, Chair of the Animal Movement

and Surveillance Department of the state of Rio Grande do Sul). Based on local capacities given

that the average number of new cases was reduced to as low as 7.88 cases per 100 farms with a β

of 0.2 when 1,000 farms were removed from the network, the former was set to be the maximum

number of feasible farms to be targeted. Furthermore, we compared two infectious seed

scenarios: a) infections started at random farms and b) stratified random infections forced to start

in proportion to the number of commercial and noncommercial farms. Both have very similar

results which reinforced the importance of non-commercial farms in the role of spreading

diseases into commercial pig farms and vice versa (Figure 4, also see supplementary figure S2,

S4 and S5).

Figure 4. Simulated disease spread dynamics within the swine contact network. Simulations

assumed two seed infection scenarios a) random, where 950 pig farms (10% prevalence) were

infected at day "0", and b) stratified, in which the proportion of the 950 pig farms (10%

prevalence) was equally divided between commercial and noncommercial farms. The

simulations assumed a farm prevalence of 0.1%.

Finally, based on deterministic runs (Payen et al., 2019), epidemic sizes were significantly

smaller even if 500 farms were removed from the full network. The average spread cascade

immediately decreased to below 40 after only 250 farms were removed, which was independent

of network metrics, except by random removals (Figure 5). In the 1,000-node removal scenario,

there were two very tight removal strategies: degree and betweenness. The smallest outbreak size

of 29 pig farms was calculated when the first 1,000 farms with the highest degree were removed

(Figure 6). Even though the cascades spread of both surveillance scenarios was similar, the

degree was significantly lower than betweenness (Figure 6, also see supplementary table 2). As

expected, random removals resulted in a much larger epidemic sizes, reaching up to 2,593 farms

(Figure 6, also see supplementary figure S3).

Figure 5. Epidemic size by node removal. The y-axis shows the cumulative size of the spread

cascades from January 2017 through December 2018 for five scenarios. On day "0", 10, 250,

500, 750 or 1,000 farms were removed from the simulation, while the temporal network metrics

were calculated for all remaining farms. The lines is the average values and shaded ± 95% CIs.

Figure 6. Final spread cascades sizes by each target-network surveillance strategy. In light blue,

the estimated spread cascade, which had a maximum value of 29 farms while targeting farms at

random, resulted in a cascade of 2,593 farms (green).

Descriptive analysis of biosecurity, infrastructure and population profile of the targeted

farms.

The geographic locations of the 1,000 target farms across the study area are shown in Figure 7.

Most vulnerable farms were commercial (73.9%), with one production system contribution with

138 farms. From the comprehensive list of biosecurity and infrastructure characteristics, farms

had a median of 2 barns (IQR-1-11), capacities for housing were approximately 1,100 pigs

(IQR=162-9,925), the median number of piglets was 1,122 (IQR-16-7,229), breeding farms had

4 (IQR=1-18) boars, and the gilts population was 544 (IQR-11-3,044). More details about the

farms' biosecurity and infrastructures can be found in Table 2.

Figure 7. The geographic locations of the top 1,000 farms ranked by logarithm of degree.

Discussion

Static networks have been widely used in the identification of farms with many contacts, often

referred to as hubs. The removal of such highly connected nodes from networks has been one of

the most prevalent approaches for guiding target activities such as enhancing surveillance;

overall, removing these nodes has had some success in reducing disease transmission (Rautureau

et al., 2012; Büttner et al., 2016; Passafaro et al., 2019). However, static network suffer from a

number of drawbacks that often lead to overestimate the size of disease outbreaks (Rautureau et

al., 2012; Büttner et al., 2016; Passafaro et al., 2019). Static networks very often cannot represent

what is truly a transient event (Dubé et al., 2008; Fielding et al., 2019), such as when a farm

receives a shipment, forming an edge between farms, whereas as soon as animals are unloaded,

the edge no longer exists. Our findings emphasize that if static networks were to be used here,

the size of disease outbreaks would be substantially overestimated. To address this issue, we

developed models that can account for the temporal order of movements and provide a more

realistic estimate of expected epidemic sizes. Our model formulation allows for the calculation of

the minimum number of farms that need to be removed to reduce disease spread and

approximate the impact of network-based target disease control strategies (e.g., farm isolation,

movement restriction, enhancement of biosecurity) on the expected epidemic sizes.

The volume and types of movements of this study well represent the expectations of a modern

swine industry in which farms are specialized and integrated (Augusta et al., 2019; Passafaro et

al., 2019). As expected, the majority of the movements were from finisher farms to

slaughterhouses and from nurseries to finishers (Lee et al., 2017; Alarcón et al., 2019; Augusta et

al., 2019; Machado et al., 2019). We also identified shipments between commercial and

noncommercial farms (see supplementary table S1 for more details). These movements may

represent important concern regarding foreign animal diseases. While we assume that most

integrated pig systems in developed countries would not receive pig shipment from

noncommercial farms, information about movement from external sources has not been reported

widely in the literature (Sterchi et al., 2019). However, a more recent study in Iowa described the

presence of such pig flows that were not considered in their analysis (Passafaro et al., 2019).

Therefore, there is a need to capture movements between commercial and noncommercial pig

farms and vice-versa. Gathering this information becomes more important as the pig industry

around the world prepares for the introduction of ASF, which will require tracking animal

movements among all pig farms (Brown and Bevins, 2018; Jurado et al., 2018).

In this study, we did not examine feed-truck or other vehicle movements or sharing. Such

information is absent from the majority of the current literature (Lentz et al., 2016), but it has

been included in some studies (Augusta et al., 2019; Sterchi et al., 2019). It is critical to

acknowledge that the movement of feed trucks and short-distance transportation of breeding

replacement animals or culled sows contributed to disease transmission (Silva et al., 2019).

Indeed, before these additional contacts are not taken into consideration, our understanding of

disease spread pathways will likely continue to be limited.

The number of active pig farms is seasonal and skewed with a variation of plus or minus 300

farms over the two-year period. A similar trend was observed in the number of animals and

overall monthly shipments. In contrast to our observations, the number of active pig farms varied

largely compared with other pig populations in Germany (Büttner et al., 2016), France

(Rautureau et al., 2012) and Canada (Thakur et al., 2016), even though it is difficult to compare

distinct networks (Miele et al., 2019). Some factors that have contributed to the variations in our

study may be associated with hog prices, which tends to be more volatile than in other countries;

thus, producers may choose to place more animals in a favorable economic scenario. In addition,

the monthly network diameter almost doubled in the middle of 2018, which seems to be a highly

abnormal pattern in the network. A possible explanation for this may be related to the global

configuration of the network because the nodes were connected in a way that increased the

length of the shortest paths between them, which was accompanied by an increase in the GWCC.

Consequently, during this period, we observed an increase in the number of nodes and

connections; therefore, a given node would require more steps to reach another farm, within the

network which could alone reduce the speed of disease transmission (Vidondo and Voelkl,

2018). While the GWCC increased over the years, the GSCC suffered from relevant variations

throughout the months, especially in the second half of 2018, where new paths connected the

GSCC with other minor components and consequently increased the size of the GWCC, which

could have implications for disease transmission and persistence (Schulz et al., 2017;

Marquetoux et al., 2016).

Previous studies have considered static network views to make inferences about disease spread

or to calculate outbreak sizes (Büttner et al., 2013; Lee et al., 2017; Salines et al., 2017).

However, these followed the assumption that movement aggregation would not overestimate or

introduce edge biases (Vidondo and Voelkl, 2018). It has been suggested that to some extent,

static networks provide useful information about disease related risks (Mohr et al., 2018).

However, most static network studies lacked basic statistical support for their decisions for data

aggregation (Büttner et al., 2013; Lee et al., 2017; Salines et al., 2017), some studies have rather

mention that other studies also used similar time windows, which would support their decisions

to data aggregation (Büttner et al., 2013; Lee et al., 2017; Salines et al., 2017). Contact network

aggregation has been a practice, even when the temporal sequences of the movements were

available (Amaku et al., 2015; Cárdenas et al., 2018). In addition, restricting movements from

and to highly connected farms reduced the connectivity between network components (GWCC

and GSCC) in cattle (Motta et al., 2017; Vidondo and Voelkl, 2018) and swine networks (Guinat

et al., 2016; Kinsley et al., 2019; Sterchi et al., 2019), which, to some extent, may contain disease

from spreading throughout the contact network. On the other hand, nodal static network

measures of monthly networks performed poorly in predicting outbreak risk (Vidondo and

Voelkl, 2018), which may reiterate the limitation of static network in explaining the spread of

diseases over time-varying networks.

The causal fidelity test is used to quantify the error of the static representation of a temporal

network (Lentz et al., 2013). Thus, without such verification, it is difficult to decide if and at

which time window inferences or metrics from static networks can be interpreted (Lentz et al.,

2013; Büttner et al., 2016). In this study, we found that July 2017 had the best time-respecting

movement representation; the value was far lower than the causal fidelity reported in the

literature, for example, two different studies in pigs reported values of 92% and 72% for monthly

networks (Büttner et al., 2016) and (Lentz et al., 2013), respectively. However, a study of Swiss

pig transport data demonstrated that only a small fraction of 0.34% of the farms was connected

within one month (Sterchi et al., 2019). The low casual fidelity we identified may be associated

with how producers or companies do business, as a producer can easily start and stop contracting

with production systems because of compensation, price of feed, etc. These heavily compromise

the use of static network metrics when the objective is to identify and direct active surveillance

plans towards highly connected farms. The use of static networks tends to artificially connect a

larger number of farms, which will consequently bias surveillance strategies to less relevant

farms and compromise resource allocation.

Our results show that different network-based intervention schemes successfully reduced the

spread of disease independently from the local β transmission probability rates at the farm. We

show that targeting farms by degree was the most effective intervention in reducing the number

of new cases, followed by targeting farms by betweenness, which required ~ 1,300 farms to be

quarantine. Finally, as anticipated random removal had little impact in controlling disease. A

recent study explored the contribution of existing trade duration on disease transmission

probabilities and the impact on epidemic sizes, suggesting that epidemic sizes were sensitive to

network activity (Farine and Whitehead, 2015; Lebl et al., 2016). Indeed, we explicitly

considered node activity while disease spread was simulated. Two other studies attempted to

account for time-dependent disease spread while aiming to reduce R_0. One study used

noncompartmental model approach and found that the number of farms that needed to be

removed by degree approached 50% of all farms (Kinsley et al., 2019). The second study used a

mechanistic approach in which ASF spread was simulated and found that degree ranking was the

best strategy to contain the disease (Ferdousi et al., 2019). In summary, degree was shown to be

the best strategy in reducing disease burden, while betweenness had the second best

performance. It is worth noting that betweenness is a useful metric when studying disease

diffusion (Farine and Whitehead, 2015), mainly because it is a representation of the shortest path

between nodes. Here noncommercial farms with high betweenness may bridge several

commercial firms; therefore, more details about these unexplored pathways should be

investigated in the future.

Based on the identified minimum number of farms of 1,000 that needed to be removed from the

contact network, we simulated outbreak propagation with a deterministic SI model and identified

the most efficient network-based metric capable of reducing the expected size of spread

cascades.

We showed that degree was the most significant intervention, with no more than 29 pig farms

involved per outbreak (Figure 6). In addition, our results demonstrated that targeting more than

500 farms by degree was efficient in reducing epidemic sizes. Random surveillance was

unexpectedly better than clossenes and PageRank for most of the two years of propagation. One

possible explanation may be that this network is not well connected, and farms that remain to be

removed may be too similar with respect to their clossennes and PageRank values; therefore, the

impact on the propagation is expected to be the one resulting from random removals.

Our results have showed an uneven proportion of commercial and noncommercial pig farms

within the first 1,000 farms with the highest degree. This illustrates the interplay between the two

populations, where infection can arrive at either and propagate in both directions. Here, most

farms of interest lacked important biosecurity features, such as cleaning and disinfection stations

(Dee et al., 2006). A study in Argentina also highlighted the lack of biosecurity on highly

connected farms (Alarcón et al., 2019). On the other hand, in our study, most farms did not

utilize rendering, but preferred to compost dead animals. The former was previously described as

an important risk factor for the spread of porcine reproductive and respiratory syndrome (PRRS)

in the US (Velasova et al., 2012; Silva et al., 2019). In the same study, composting was found to

be the less risky practice when removing cull or dead pigs (Silva et al., 2019). Another relevant

biosecurity point that has been described as a risk for disease introduction is the number of farm

entries; here, we noticed that the majority of the farms had only one main road, which potentially

represents less risk for local disease transmission (Silva et al., 2019). A disproportionate number

of these highly risky farms were nurseries and breeding farms, which has often been proposed as

a desired group to be targeted for surveillance (Dorjee et al., 2013). In addition to the

identification of priority farms, the identification of relevant biosecurity gaps at those farms is

invaluable, especially under the current preparedness for the introduction of foreign animal

diseases such as ASF. One of the best options is the implementation of enhanced biosecurity

plans, such the Secure Pork Supply (SPS) Plan, designed to provide business continuity in the

event of a foreign animal disease outbreak as well as help protect operations from endemic

diseases (Pudenz et al., 2019).

Finally, more comprehensive studies coupling animal movements with local and environmental

routes of transmission while accounting for farms biosecurity and infrastructure are needed to

further explore the mechanisms of pig disease propagation, for which a complete multiscale

mechanistic model is necessary (Qi et al., 2019).

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Considerations

The use of static networks to target key farms to control the disease from spreading should be

interpreted with caution due to several caveats associated with data aggregation, as shown in this

study and elsewhere (Lentz et al., 2016). An obvious issue here is to assume that the same pair of

farms would trade with each other throughout the entire study period. Therefore, we argue that

the causal fidelity of the temporal network should be presented before considering the use of the

static networks in the development of disease surveillance or control strategies (Lentz et al.,

2016; Payen et al., 2019). Recent studies did not account for this issue (Büttner et al., 2013,

2018; Marquetoux et al., 2016; Motta et al., 2017; Alarcón et al., 2019), or considered the

temporal network pathway accessibility of each farm. In summary, these studies used static

networks to disassemble networks into small islands, which possibly overestimates the size of

outbreaks systematically, and therefore, the results should be interpreted with caution.

A likely limitation of this study was how farms were classified as commercial or noncommercial.

Farms without information about production systems were classified as non-commercial.

According to the state veterinarians (SEAPI-RS, 2018), all farms are required by law to inform

yearly whether they are raising pigs for any commercial system, which makes our assumption

fair. However, this could be in some extend associated with the inflated amount of movement

between commercial and non-commercial (see supplementary table S1).

Finally, the modeling framework proposed here can be broadly used to calculate the minimum

number of farms to be targeted for the control of disease spread when temporal contact data are

available and to determine the network metrics that should be preferred for selection of target

farms according to the approximation of epidemic sizes. In addition, the proposed framework

could be used as a decision tool for both local official veterinary services and the private sector

for evaluating animal movements and developing a more risk-oriented surveillance strategies.

The model may also help develop an idea of the amount of personnel and material resources that

would need to be allocated to control the epidemic, which is key for preparedness for future

events (Casal et al., 2019).

Conclusion

One-third of all pig farms traded with the same partners in the course of two years; consequently,

the static networks did not offer reasonable information about the realized temporal movement

pathways. In this scenario, the use of static networks to target vulnerable or high-risk farms by

network metrics would likely overestimate the role of those farms in disease transmission and

limit the accuracy needed to optimize surveillance. Our proposed modeling framework provides

a parsimonious solution to this problem; it handles the temporal order of movements, calculates

the minimum number of farms needed to be targeted and approximates the expected epidemic

size in a worst-case scenario where each movement can effectively transmit disease. The results

from the application of this approach identified priority pig farms that had remarkably limited

biosecurity. This fact imposes important vulnerability to pathogen introduction and spread;

therefore, those farms could benefit from the implementation of enhanced biosecurity plans, such

as what has been promoted and implemented in the US by SPS plan. Finally, the introduction of

risk-based approaches may optimize the costs of routine surveillance and can be used in

preparation for the introduction of FAD into the local pig industry.

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Authors' contributions

GM, FPNL, JV and AARM coordinated movement data collection. GM conceived paper ideas.

GM, JAG and NCC participated in the design of the study. NCC and GM conducted data

processing and cleaning. GM, JAG and NCC wrote and modified computer algorithms and

conducted analysis. GM, JAG and NCC developed simulation models. GM and NCC wrote the

manuscript and JA edited the manuscript. All authors discussed results and commented on the

manuscript.

Conflict of interest

All authors confirm that there are no conflicts of interest to declare

Ethical statement

The authors confirm that the ethical policies of the journal, as noted on the journal's author

guidelines page. Since this work did not involve animal sampling neither questioner data

collection there was no need for ethics permits.

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References

Alarcón, L. V, P. CIPRIOTTI, M. Monterubbianessi, C. Perfumo, E. Mateu, and A. Allepuz,

2019: Network analysis of pig movements in Argentina: identification of key farms in the

spread of diseases and relationship with their biosecurity level. *Front. Vet. Sci.*DOI:

10.3389/conf.fvets.2019.05.00006.

Amaku, M., J.H. de H. Grisi-Filho, R.L. Negreiros, R.A. Dias, F. Ferreira, J.S.F. Neto, R.I.

Cipullo, F.S. Marques, and R. Ossada, 2015: Infectious disease surveillance in animal

movement networks: An approach based on the friendship paradox. *Prev. Vet. Med.*DOI:

10.1016/j.prevetmed.2015.08.002.

Augusta, C., G.W. Taylor, and R. Deardon, 2019: Dynamic contact networks of swine

movement in Manitoba, Canada: Characterization and implications for infectious disease

spread. *Transbound. Emerg. Dis.*DOI: 10.1111/tbed.13220.

Brin, S., and L. Page, 1998: The anatomy of a large scale hypertextual Web search engine

published version. *Comput. Networks ISDN Syst.*DOI: 10.1.1.109.4049.

Brown, V.R., and S.N. Bevins, 2018: A review of African swine fever and the potential for

introduction into the United States and the possibility of subsequent establishment in feral

swine and native ticks. *Front. Vet. Sci.*DOI: 10.3389/fvets.2018.00011.

Büttner, K., J. Krieter, A. Traulsen, and I. Traulsen, 2013: Static network analysis of a pork

supply chain in Northern Germany-Characterisation of the potential spread of infectious

diseases via animal movements. *Prev. Vet. Med.*DOI: 10.1016/j.prevetmed.2013.01.008.

Büttner, K., J. Salau, and J. Krieter, 2016: Quality assessment of static aggregation compared to

the temporal approach based on a pig trade network in Northern Germany. *Prev. Vet.*

*Med.*DOI: 10.1016/j.prevetmed.2016.05.005.

Büttner, K., J. Salau, and J. Krieter, 2018: Effects of data quality in an animal trade network and

their impact on centrality parameters. *Soc. Networks*DOI: 10.1016/j.socnet.2018.01.001.

Cárdenas, N.C., J.O.A. Galvis, A.A. Farinati, J.H.H. Grisi-Filho, G.N. Diehl, and G. Machado,

2018: Burkholderia mallei: the dynamics of networks and disease transmission.

Transbound. Emerg. Dis. **0**, DOI: 10.1111/tbed.13071.

Casal, J., C. Saegerman, S. Bertagnoli, G. Meyer, J.P. Ganière, P. Caufour, K. De Clercq, P.

Jacquiet, C. Hautefeuille, F. Etore, and S. Napp, 2019: A simple method to estimate the

number of doses to include in a bank of vaccines. The case of Lumpy Skin Disease in

France. *PLoS One*DOI: 10.1371/journal.pone.0210317.

Chaters, G.L., P.C.D. Johnson, S. Cleaveland, J. Crispell, W.A. De Glanville, T. Doherty, L.

Matthews, S. Mohr, O.M. Nyasebwa, G. Rossi, L.C.M. Salvador, E. Swai, and R.R. Kao,

2019: Analysing livestock network data for infectious disease control: An argument for

routine data collection in emerging economies. *Philos. Trans. R. Soc. B Biol. Sci.*DOI:

10.1098/rstb.2018.0264.

Colman, E., P. Holme, H. Sayama, and C. Gershenson, 2019: Efficient sentinel surveillance

strategies for preventing epidemics on networks. (Daniela Paolotti, Ed.)*PLOS Comput. Biol.*

15, e1007517, DOI: 10.1371/journal.pcbi.1007517.

Csardi, G., and T. Nepusz, 2006: The igraph software package for complex network research.

InterJournal Complex Syst.

Darbon, A., D. Colombi, E. Valdano, L. Savini, A. Giovannini, and V. Colizza, 2019: Disease

persistence on temporal contact networks accounting for heterogeneous infectious periods.

*R. Soc. Open Sci.*DOI: 10.1098/rsos.181404.

Dee, S.A., J. Deen, and C. Pijoan, 2006: Evaluation of an industry-based sanitation protocol for

602  full-size transport vehicles contaminated with porcine reproductive and respiratory

603  syndrome virus. *J. Swine Heal. Prod.*

604 Dorjee, S., C.W. Revie, Z. Poljak, W.B. McNab, and J. Sanchez, 2013: Network analysis of

605  swine shipments in Ontario, Canada, to support disease spread modelling and risk-based

606  disease management. *Prev. Vet. Med.*DOI: 10.1016/j.prevetmed.2013.06.008.

607 Dubé, C., C. Ribble, D. Kelton, and B. McNab, 2008: Comparing network analysis measures to

608  determine potential epidemic size of highly contagious exotic diseases in fragmented

609  monthly networks of dairy cattle movements in Ontario, Canada. *Transbound. Emerg.*

610  *Dis.*DOI: 10.1111/j.1865-1682.2008.01053.x.

611 Farine, D.R., and H. Whitehead, 2015: Constructing, conducting and interpreting animal social

612  network analysis. *J. Anim. Ecol.*DOI: 10.1111/1365-2656.12418.

613 Ferdousi, T., S.A. Moon, A. Self, and C. Scoglio, 2019: Generation of swine movement network

614  and analysis of efficient mitigation strategies for African swine fever virus. DOI:

615  10.1371/journal.pone.0225785.

616 Fielding, H.R., T.J. McKinley, M.J. Silk, R.J. Delahay, and R.A. McDonald, 2019: Contact

617  chains of cattle farms in Great Britain. *R. Soc. Open Sci.*DOI: 10.1098/rsos.180719.

618 Firestone, S.M., Y. Hayama, R. Bradhurst, T. Yamamoto, T. Tsutsui, and M.A. Stevenson, 2019:

619  Reconstructing foot-and-mouth disease outbreaks: a methods comparison of transmission

620  network models. *Sci. Rep.*DOI: 10.1038/s41598-019-41103-6.

621 Freeman, L.C., 1978: Centrality in social networks conceptual clarification. *Soc. Networks*DOI:

622  10.1016/0378-8733(78)90021-7.

623 Funk, S., A. Camacho, A.J. Kucharski, R.M. Eggo, and W.J. Edmunds, 2018: Real-time

624  forecasting of infectious disease dynamics with a stochastic semi-mechanistic model.

*Epidemics*DOI: 10.1016/j.epidem.2016.11.003.

Guinat, C., A. Relun, B. Wall, A. Morris, L. Dixon, and D.U. Pfeiffer, 2016: Exploring pig trade

patterns to inform the design of risk-based disease surveillance and control strategies. *Sci.*

*Rep.*DOI: 10.1038/srep28429.

Jurado, C., M. Martínez-Avilés, A.D. La Torre, M. Štukelj, H.C. de C. Ferreira, M. Cerioli, J.M.

Sánchez-Vizcaíno, and S. Bellini, 2018: Relevant measures to prevent the spread of African

Swine Fever in the European Union Domestic Pig Sector. *Front. Vet. Sci.*DOI:

10.3389/fvets.2018.00077.

Kim, M., D. Paini, and R. Jurdak, 2019: Modeling stochastic processes in disease spread across a

heterogeneous social system. *Proc. Natl. Acad. Sci. U. S. A.*DOI:

10.1073/pnas.1801429116.

Kim, P., and C.H. Lee, 2018: Epidemic Spreading in Complex Networks with Resilient Nodes:

Applications to FMD. *Complexity*DOI: 10.1155/2018/5024327.

Kinsley, A.C., A.M. Perez, M.E. Craft, and K.L. Vanderwaal, 2019: Characterization of swine

movements in the United States and implications for disease control. *Prev. Vet. Med.*DOI:

10.1016/j.prevetmed.2019.01.001.

Konschake, M., H.H.K. Lentz, F.J. Conraths, P. Hövel, and T. Selhorst, 2013: On the Robustness

of In- and Out-Components in a Temporal Network. *PLoS One*DOI:

10.1371/journal.pone.0055223.

Lanzas, C., K. Davies, S. Erwin, and D. Dawson, 2020: On modelling environmentally

transmitted pathogens. *Interface Focus* **10**, 20190056, DOI: 10.1098/rsfs.2019.0056.

Lebl, K., H.H.K. Lentz, B. Pinior, and T. Selhorst, 2016: Impact of network activity on the

spread of infectious diseases through the german pig trade network. *Front. Vet. Sci.*DOI:

10.3389/fvets.2016.00048.

Lee, K., D. Polson, E. Lowe, R. Main, D. Holtkamp, and B. Martínez-López, 2017: Unraveling

the contact patterns and network structure of pig shipments in the United States and its

association with porcine reproductive and respiratory syndrome virus (PRRSV) outbreaks.

*Prev. Vet. Med.*DOI: 10.1016/j.prevetmed.2017.02.001.

Lentz, H.H.K., A. Koher, P. Hövel, J. Gethmann, C. Sauter-Louis, T. Selhorst, and F.J. Conraths,

2016: Disease spread through animal movements: A static and temporal network analysis of

pig trade in Germany. *PLoS One*DOI: 10.1371/journal.pone.0155196.

Lentz, H.H.K., T. Selhorst, and I.M. Sokolov, 2013: Unfolding Accessibility Provides a

Macroscopic Approach to Temporal Networks. *Phys. Rev. Lett.*DOI:

10.1103/PhysRevLett.110.118701.

Machado, G., C. Vilalta, M. Recamonde-Mendoza, C. Corzo, M. Torremorell, A. Perez, and K.

VanderWaal, 2019: Identifying outbreaks of Porcine Epidemic Diarrhea virus through

animal movements and spatial neighborhoods. *Sci. Rep.* **9**, DOI: 10.1038/s41598-018-

36934-8.

Marquetoux, N., M.A. Stevenson, P. Wilson, A. Ridler, and C. Heuer, 2016: Using social

network analysis to inform disease control interventions. *Prev. Vet. Med.*DOI:

10.1016/j.prevetmed.2016.01.022.

Mekonnen, G.A., G. Ameni, J.L.N. Wood, S. Berg, and A.J.K. Conlan, 2019: Network analysis

of dairy cattle movement and associations with bovine tuberculosis spread and control in

emerging dairy belts of Ethiopia. *BMC Vet. Res.*DOI: 10.1186/s12917-019-1962-1.

Miele, V., C. Matias, S. Robin, and S. Dray, 2019: Nine quick tips for analyzing network data.

(Francis Ouellette, Ed.)*PLOS Comput. Biol.* **15**, e1007434, DOI:

10.1371/journal.pcbi.1007434.

Miller, J.C., 2017: Mathematical models of SIR disease spread with sexual-network and other

transmission mechanisms. *Infect. Dis. Model.*DOI: 10.1016/j.idm.2016.12.003.

Mohr, S., M. Deason, M. Churakov, T. Doherty, and R.R. Kao, 2018: Manipulation of contact

network structure and the impact on foot-and-mouth disease transmission. *Prev. Vet.*

*Med.*DOI: 10.1016/j.prevetmed.2018.05.006.

Motta, P., T. Porphyre, I. Handel, S.M. Hamman, V. Ngu Ngwa, V. Tanya, K. Morgan, R.

Christley, and B.M.D. Bronsvoort, 2017: Implications of the cattle trade network in

Cameroon for regional disease prevention and control. *Sci. Rep.*DOI: 10.1038/srep43932.

Nöremark, M., N. Håkansson, S.S. Lewerin, A. Lindberg, and A. Jonsson, 2011: Network

analysis of cattle and pig movements in Sweden: Measures relevant for disease control and

risk based surveillance. *Prev. Vet. Med.*DOI: 10.1016/j.prevetmed.2010.12.009.

Ortiz-Pelaez, A., D.U. Pfeiffer, R.J. Soares-Magalhães, and F.J. Guitian, 2006: Use of social

network analysis to characterize the pattern of animal movements in the initial phases of the

2001 foot and mouth disease (FMD) epidemic in the UK. *Prev. Vet. Med.*DOI:

10.1016/j.prevetmed.2006.04.007.

Passafaro, T.L., A.F.A. Fernandes, B.D. Valente, N.H. Williams, and G.J.M. Rosa, 2019:

Network analysis of swine movements in a multi-site pig production system in Iowa, USA.

*Prev. Vet. Med.*DOI: 10.1016/J.PREVETMED.2019.104856.

Payen, A., L. Tabourier, and M. Latapy, 2019: Spreading dynamics in a cattle trade network:

Size, speed, typical profile and consequences on epidemic control strategies. *PLoS One*DOI:

10.1371/journal.pone.0217972.

Pebesma, E., 2018: Sf: Simple Features for R. R package version 0.5☐3. .

Pellis, L., F. Ball, S. Bansal, K. Eames, T. House, V. Isham, and P. Trapman, 2015: Eight

challenges for network epidemic models. *Epidemics*DOI: 10.1016/j.epidem.2014.07.003.

Pudenz, C.C., L.L. Schulz, and G.T. Tonsor, 2019: Adoption of secure pork supply plan

biosecurity by U.S. Swine producers. *Front. Vet. Sci.*DOI: 10.3389/fvets.2019.00146.

Qi, L., G. Beaunée, S. Arnoux, B.L. Dutta, A. Joly, E. Vergu, and P. Ezanno, 2019:

Neighbourhood contacts and trade movements drive the regional spread of bovine viral

diarrhoea virus (BVDV). *Vet. Res.*DOI: 10.1186/s13567-019-0647-x.

Rautureau, S., B. Dufour, and B. Durand, 2012: Structural vulnerability of the French swine

industry trade network to the spread of infectious diseases. *Animal*DOI:

10.1017/S1751731111002631.

Salines, M., M. Andraud, and N. Rose, 2017: Pig movements in France: Designing network

models fitting the transmission route of pathogens. *PLoS One*DOI:

10.1371/journal.pone.0185858.

Schulz, J., A. Boklund, T.H.B. Halasa, N. Toft, and H.H.K. Lentz, 2017: Network analysis of pig

movements: Loyalty patterns and contact chains of different holding types in Denmark.

*PLoS One*DOI: 10.1371/journal.pone.0179915.

SEAPI-RS, 2018: SEAPI, Secretaria da Agricultura, Pecuária e arrigação [Online] Available at

http://www.agricultura.rs.gov.br/inicial.

Silva, G.S., G. Machado, K.L. Baker, D.J. Holtkamp, and D.C.L. Linhares, 2019: Machine-

learning algorithms to identify key biosecurity practices and factors associated with

breeding herds reporting PRRS outbreak. *Prev. Vet. Med.*DOI:

10.1016/j.prevetmed.2019.104749.

Siqueira, P., 2019: brazilmaps: Brazilian Maps from Different Geographic Levels. .

Smith, R.P., A.J.C. Cook, and R.M. Christley, 2013: Descriptive and social network analysis of

pig transport data recorded by quality assured pig farms in the UK. *Prev. Vet. Med.* **108**,

167–177, DOI: 10.1016/j.prevetmed.2012.08.011.

Smith, T.A., N. Chitnis, M. Penny, and M. Tanner, 2017: Malaria Modeling in the Era of

Eradication. *Cold Spring Harb. Perspect. Med.* **7**, a025460, DOI:

10.1101/cshperspect.a025460.

Sterchi, M., C. Faverjon, C. Sarasua, M.E. Vargas, J. Berezowski, A. Bernstein, R. Grütter, and

H. Nathues, 2019: The pig transport network in Switzerland: Structure, patterns, and

implications for the transmission of infectious diseases between animal holdings. *PLoS*

*One*DOI: 10.1371/journal.pone.0217974.

Thakur, K.K., C.W. Revie, D. Hurnik, Z. Poljak, and J. Sanchez, 2016: Analysis of Swine

Movement in Four Canadian Regions: Network Structure and Implications for Disease

Spread. *Transbound. Emerg. Dis.*DOI: 10.1111/tbed.12225.

Valdano, E., C. Poletto, A. Giovannini, D. Palma, L. Savini, and V. Colizza, 2015: Predicting

Epidemic Risk from Past Temporal Contact Data. *PLoS Comput. Biol.*DOI:

10.1371/journal.pcbi.1004152.

Velasova, M., P. Alarcon, S. Williamson, and B. Wieland, 2012: Risk factors for porcine

reproductive and respiratory syndrome virus infection and resulting challenges for effective

disease surveillance. *BMC Vet. Res.*DOI: 10.1186/1746-6148-8-184.

Vidondo, B., and B. Voelkl, 2018: Dynamic network measures reveal the impact of cattle

markets and alpine summering on the risk of epidemic outbreaks in the Swiss cattle

population. *BMC Vet. Res.*DOI: 10.1186/s12917-018-1406-3.

Wasserman, S., and K. Faust, 1994: Social Network Analysis: Methods and Applications :

Cambridge University Press.

Wickham, H., 2017: Tidyverse: Easily install and load "tidyverse" packages. .

Widgren, S., P. Bauer, R. Eriksson, and S. Engblom, 2019: Siminf: An R package for data-driven

stochastic disease spread simulations. *J. Stat. Softw.*DOI: 10.18637/jss.v091.i12.

Table 1. Description of network terminology and definition applied to pig movements.

Parameter	Definition	References
Nodes	A pig farms.	Wasserman and Faust (1994)
Edge	A link between two nodes.	Wasserman and Faust (1994)
Degree (k)	The number of unique contacts to and from a farm. When direction was taken into account, the ingoing and outgoing contacts were separated: the "out" degree is the number of contacts originating from a specific farm, and the "in" degree is the number of contacts directed toward a specific premise.	Wasserman and Faust (1994)
PageRank	Measure of the Google PageRank, a link analysis algorithm that produces a ranking importance of all the farms from 0 to 1.	Brin and Page (1998)
Closeness centrality	Closeness centrality measures how many steps are required to access every other vertex from a given node. This measure can be directed as incoming steps or outgoing steps.	Freeman (1978)
Betweenness	The frequency at which a node is located on the shortest path between any two pairs of nodes in the network.	Freeman (1978)
Giant weakly connected	The proportion of nodes that are connected in the largest component when directionality of movement is ignored.	Wasserman and Faust

component (GWCC)		(1994)
Giant strongly connected component (GSCC)	The proportion of nodes that are connected in the largest component when directionality of movement is considered.	Wasserman and Faust (1994)

750
751 **Table 2.** Description of biosecurity and infrastructure of the targeted farms.

Variable	Overall proportion
Presence of isolation barn(s)	No=77.6%
	Yes=22.4%
Fence around the farm	No=54.7%
	Yes=45.3%
Cleaning and disinfection station(s)	No=72.7%
	Yes=27.3%
Vegetation barrier around the farm	No=86.7%
	Yes=13.3%
Single entry point	No=72.4%
	Yes=27.6%
Bird nets	No=59.6%
	Yes=40.4%
Pig loading and unloading outside the farm's designated clean area	No=61.3%
	Yes=38.7%
Special clothing dedicated for farm use (shower in)	No=77.7%
	Yes=22.3%
Feed bins inside the designated clean area while loading is external	No=60.3%
	Yes=39.7%
*Vulnerability score ***	
Low	0.21%
Medium	47.4%
High	17.8%
Not informed	34.8%
Veterinary care	
From commercial system	63.5%
Independent veterinary	4.5%
Absent	1.1%
Not informed	30.9%
Farm type	
Nursery	32.23%
Breeding farm	20.2%
Finisher farm	15.1%
Certified swine breeder farm	5.65%
Wean-to-finisher	1.49%
Not informed	25.33%
Water sources	
Pond	42.8%
Natural water fountain	22.2%
Public source	9.9%
Lake	1.5%
Water cistern	0.3%
Not informed	23.3%
Feed source	

From production system	68.7%
Own feed mill	7.8%
Not informed	23.5%
Destination of dead animals	
Compost	76.0%
Incinerator	0.23%
Bury	0.23%
Septic pit	0.24%
Not informed	23.3%
Manure destination	
Lagoon	28.9%
Septic pit	2.48%
Not informed	68.62%
Barn infrastructure	
Concrete or bricks	51.9%
Wood	1.29%
Mixed of wood and concrete	23.4%
Not informed	23.5%

752 ***This measures the level of vulnerability the introduction of pathogen into certified-swine-

753 breeder-farms, briefly is a combination of risks measurements form distance to farms to presence

754 of biosecurity and infrastructure of the farm.

Figure Legend

Figure 1. Geographical location of commercial and non-commercial pig farms

Figure 2. Description of pig movements between all farms types from January 2017 to

December 2018. The arrows size shows the percentage of movements and the color is given by

the origin of the movements. The size of the circles represented the total number of farms.

Figure 3. Description of the monthly variation of the network metrics of the pig movements

from January 2017 until December 2018.

Figure 4. Simulated disease spread dynamics within the swine contact network. Simulations

assumed two seed infection scenarios a) random, where 950 pig farms (10% prevalence) were

infected at day "0", and b) stratified, in which the proportion of the 950 pig farms (10%

prevalence) was equally divided between commercial and noncommercial farms. The

simulations assumed a farm prevalence of 0.1%.and non-commercial farms. The simulations

assumed within farm prevalence of 0.1%.

777
Figure 5. Epidemic size by node removal. Y-axis show the cumulative size of the spread

cascades from January 2017 until December 2018 for five scenarios. From 10, 250, 500, 750 to

1,000 farms were removed on day "0" of the simulation while temporal network metrics were

calculated for all remaining farms. Line are the average values and shade 95% CI.

Figure 6. Final spread cascade sizes by each target-network surveillance strategy. In light-blue

the estimate spread cascade which had maximum value of 29 farms while targeting farms at

random resulted in cascade of 2,593 farms (green).

786
787   **Figure 7.** The geographic locations of the top 1,000 farms ranked by logarithm of degree.