

Climate change will drive novel cross-species viral transmission

Colin J. Carlson^{1,2,†}, Gregory F. Albery^{1,3,†}, Cory Merow⁴, Christopher H.
Trisos⁵, Casey M. Zipfel¹, Evan A. Eskew³, Kevin J. Olival³, Noam Ross³, and
Shweta Bansal¹

¹*Department of Biology, Georgetown University, Washington, D.C., USA.*

²*Center for Global Health Science & Security, Georgetown University, Washington, D.C., USA.*

³*EcoHealth Alliance, New York, NY, USA.*

⁴*Ecology and Evolutionary Biology, University of Connecticut, Storrs, CT, USA.*

⁵*African Climate and Development Initiative, University of Cape Town, Cape Town, South
Africa.*

[†] *These authors share equal authorship.*

January 24, 2020

Abstract

Between 10,000 and 600,000 species of mammal virus are estimated to have the potential to spread in human populations, but the vast majority are currently circulating in wildlife, largely undescribed and undetected by disease outbreak surveillance^{1,2,3}. In addition, changing climate and land use drive geographic range shifts in wildlife, producing novel species assemblages and opportunities for viral sharing between previously isolated species^{4,5}. In some cases, this will inevitably facilitate spillover into humans^{6,7}—a possible mechanistic link between global environmental change and emerging zoonotic disease⁸. Here, we map potential hotspots of viral sharing, using a phylogeographic model of the mammal-virus network, and projections of geographic range shifts for 3,870 mammal species under climate change and land use scenarios for the year 2070. Shifting mammal species are predicted to aggregate at high elevations, in biodiversity hotspots, and in areas of high human population density in Asia and Africa, sharing novel viruses between 3,000 and 13,000 times. Counter to expectations, holding warming under 2°C within the century does not reduce new viral sharing, due to greater range expansions—highlighting the need to invest in surveillance even in a low-warming future. Most projected viral sharing is driven by diverse hyperreservoirs (rodents and bats) and large-bodied predators (carnivores). Because of their unique dispersal capacity, bats account for the majority of novel viral sharing, and are likely to share viruses along evolutionary pathways that could facilitate future emergence in humans. Our findings highlight the urgent need to pair viral surveillance and discovery efforts with biodiversity surveys tracking range shifts, especially in tropical countries that harbor the most emerging zoonoses.

38 Main Text

39 In the face of rapid environmental change, survival for many species depends on moving
40 to track shifting climates. Even in a best case scenario, many species are projected
41 to shift a hundred kilometers or more in the next century^{9,10}. In the process, many
42 animals will bring their parasites and pathogens into new environments^{4,11}, creating new
43 evolutionary opportunities for host jumps⁸. Most conceptual frameworks for cross-species
44 transmission revolve around how these host jumps facilitate the spillover of new zoonotic
45 pathogens into humans^{12,13,14}, but viral evolution is an undirected process¹⁵, in which
46 humans are only one of over 5,000 mammal species with over 12 million possible pairwise
47 combinations¹⁶. Despite their indisputable significance, zoonotic emergence events are
48 just the tip of the iceberg; almost all cross-species transmission events will occur among
49 wild mammals, largely undetected and mostly inconsequential for public health.

50 Of the millions of possible pairwise viral exchanges, the vast majority are biologically
51 implausible, as host species' geographic ranges currently do not overlap. However, as
52 ranges shift, a small fraction of possible interactions will occur, of which a subset will
53 lead to viral establishment in a novel host. Which subset results in establishment de-
54 pends on *opportunity* and *compatibility*^{14,17,18}, analogous to exposure and susceptibility
55 within populations, and both dimensions pose an important predictive challenge. The
56 ability of species to track shifting habitats in a changing climate will determine which
57 pairs of species encounter each other for the first time^{4,19}. Habitat selection and be-
58 havioral differences can further limit contact, even if species are nominally sympatric¹⁹.
59 Some viruses may spread environmentally between spatially-proximate species with no
60 direct behavioral contact²⁰, but generally, sharing is more likely among species with more
61 ecological overlap²¹. Even among species in close contact, most spillovers are still a dead
62 end; progressively smaller subsets of viruses can infect novel host cells, proliferate, and
63 transmit onward in a new host¹⁸. Their ability to do so is determined by compatibility
64 between viral structures, host cell receptors, and host immunity⁶. Because closely related
65 species share both ecological and immunological traits through identity by descent, phy-
66 logeny is a strong predictor of pathogen sharing^{17,22}, as well as susceptibility to invasion
67 by new viruses^{23,24,25}. In a changing world, these factors should continue to mediate the
68 impact of ecosystem turnover on the mammalian virome.

69 Although several studies have mapped current hotspots of emerging diseases^{3,26,27},
70 few have modeled them in the context of global change. With the global reassortment
71 of animal biodiversity, it is unknown whether bats and rodents will still play a central
72 role in viral emergence^{3,28} (ED Figure 1), or whether hotspots of viral emergence will
73 stay in tropical rainforests^{27,29} which currently harbor most undiscovered viruses^{3,30}.
74 Here, by projecting geographic range shifts and applying fundamental biological rules

for cross-species transmission, we predicted how and where global change could create novel opportunities for viral sharing. We built species distribution models for 3,870 mammal species, and projected geographic range shifts based on four paired scenarios of climate change (representative concentration pathways, RCPs) and land use change (shared socioeconomic pathways, SSPs) by 2070. We treated dispersal potential as an additional layer of biological realism, inferring these limits for species based on allometric scaling³¹, and compared predictions with and without dispersal constraints. We used these projections to identify where novel range overlap among unfamiliar species (“first encounters”) could happen, and used a recently-developed model to predict the probability of viral sharing based on geographic overlap and host phylogenetic similarity¹⁷. (ED Figure 2) This model framework allows powerful inference based on the ~1% of the global mammalian virome that has been described^{1,3,17}. Using this approach, we tested the hypothesis that environmental change should drive biotic homogenization of mammal communities, exposing mammals to novel viruses, and altering the structure of mammal-virus interactions.

Most mammals are projected to undergo rapid range shifts in the next half century¹⁰. If range shifts can keep pace with the velocity of climate change³², we predict that the vast majority of mammal species (89%–98%) will overlap with at least one unfamiliar species somewhere in their future range, regardless of emissions scenario. At the global level, community turnover would permit almost 300,000 novel species interactions (ED Figure 3). These “first encounters” between mammal species will occur everywhere in the world, but are concentrated in tropical Africa and southeast Asia (ED Figure 4). This result was surprising, and counter to our expectation that species might aggregate at higher latitudes, given that most research has focused on poleward range shifts^{33,34,35}, and previous work has anticipated a link between climate change, range shifts, and parasite host-switching in the Arctic^{36,37}. However, our findings show that communities tend to shift along latitudinal gradients together, with species rarely encountering new conspecifics³⁸. In contrast, species will track thermal optima along elevational gradients and aggregate in novel combinations in mountain ranges, especially in tropical areas with the highest baseline diversity, matching prior predictions³⁹.

This global re-organization of mammal assemblages is projected to dramatically impact the structure of the mammalian virome. Accounting for geographic opportunity and phylogenetic compatibility, we projected that a total of 279,427 first encounters in RCP 2.6 would lead to nearly 12,000 novel sharing events. Assuming that spillover will be localized to areas of novel host overlap, we mapped expected viral sharing events, and found again that most sharing should occur in high-elevation, species-rich ecosystems in Africa and Asia (Figure 1A). If species survive a changing climate by aggregating in high elevation refugia, this suggests emerging viruses may be an increasing problem for

113 their conservation^{40,41}. Across scenarios, the spatial signal of expected sharing events is
 114 nearly identical, and dominated more by the extent of range shifts than by underlying
 115 community phylogenetic structure (ED Figure 5); at least in our framework, opportunity
 116 drives spatial patterns more than compatibility.

117 Species' dispersal capacity is likely to constrain range shifts, and therefore to limit
 118 novel viral exchange. We limited the dispersal potential of flightless species further to the
 119 restrictions placed on the SDM projections, based on an established allometric scaling
 120 with body size, trophic rank, and generation time⁴². Dispersal limits caused significant
 121 reductions in range expansions across all scenarios, especially warmer ones, and therefore
 122 drove a reduction in novel interactions. Even in RCP 2.6 (the mildest scenario), limiting
 123 dispersal reduced the number of first encounters by 60%, and reduced the associated viral
 124 sharing events by 69%—to a still-staggering 3,600–3,800 projected viral sharing events.
 125 Because trophic position and body size determine dispersal capacity, carnivores account
 126 for a disproportionate number of first encounters, while ungulates and rodents have
 127 slightly fewer first encounters than expected at random (ED Figure 6) Spatial patterns
 128 also changed dramatically when dispersal constraints were added, with the majority of
 129 first encounters and cross-species viral transmission events occurring in southeast Asia
 130 (Figure 1B, ED Figures 4, 5). This viral sharing hotspot is driven disproportionately
 131 by bats, because their dispersal was left unconstrained; we made this choice given their
 132 exclusion from the original study³¹, genetic evidence that flight allows bats—and their
 133 viruses—to circulate at continental levels^{43,44}, and data suggesting that bat distributions
 134 are already undergoing disproportionately rapid shifts⁴⁵. Bats account for 87% of first
 135 encounters after constraining dispersal, and dominate the spatial pattern, with most of
 136 their first encounters restricted to southeast Asia (Figure 2).

137 Bats' unique capacity for flight could be an important and previously unconsidered
 138 link between climate-driven range shifts and future changes in the mammal virome.
 139 Even non-migratory bats can regularly travel hundreds of kilometers within a lifetime,
 140 far exceeding what small mammals might be able to cover in 50 years; half of all bat
 141 population genetic studies have failed to find any evidence for isolation by distance⁴⁶.
 142 This unique dispersal capacity has inevitable epidemiological implications, with recent
 143 evidence suggesting that continental panmixia may be common for zoonotic reservoirs,
 144 and allow viral circulation at comparable scales^{43,44,47}. We found that a staggering
 145 number of studies have also identified ongoing rapid range expansions in bat species
 146 around the world^{45,48,49,50,51,52,53,54,55}, with little mention in the broader climate change
 147 or emerging disease literature. If flight does allow bats to undergo more rapid range
 148 shifts than other mammals, we expect they should drive the majority of novel cross-
 149 species viral transmission, and likely bring zoonotic viruses into new regions. This could
 150 add an important new dimension to ongoing debate about whether bats are “special”

151 due to their higher viral richness, higher proportion of zoonotic viruses, and potentially
152 unique immune adaptations^{3,56,57,58,59}.

153 More broadly, climate-driven changes in the mammalian virome are likely to cascade
154 in future emergence of zoonotic viruses. Among the tens of thousands of expected viral
155 host jumps, some of the highest-risk zoonoses or potential zoonoses are likely to find
156 new hosts. This may pose a threat to human health down the road: the same general
157 rules for cross-species transmission explain spillover patterns for emerging zoonoses^{60,61},
158 and the viral species that make successful jumps across wildlife species have the highest
159 propensity for zoonotic emergence^{3,7,28}. Just as simian immunodeficiency virus emer-
160 gence in chimpanzees and gorillas facilitated the origin of HIV, or SARS-CoV spillover
161 into civets allowed a bat virus to reach humans, these wildlife-to-wildlife host jumps may
162 be evolutionary stepping stones for the ~10,000 to 600,000 potentially zoonotic viruses
163 that are currently circulating in mammal hosts¹.

164 To illustrate this problem, we constructed a sub-network of 13 possible Zaire ebolavirus
165 hosts in Africa, and projected possible first encounters involving these species (Figure
166 3A-C). We project these 13 species to encounter 3,604 new mammals in RCP 2.6, with
167 a modest reduction to 2,586 species by dispersal limits. These first encounters are pre-
168 dicted to produce 87 new viral sharing events that might include ZEBOV, and which
169 cover a much broader part of Africa than the current zoonotic niche of Ebola⁶². Hu-
170 man spillover risk aside, this could expose several new wildlife species to a deadly virus,
171 historically responsible for sizable primate die-offs⁶³. Moreover, for zoonoses like Zaire
172 ebolavirus without known reservoirs, future host jumps would only complicate urgent
173 efforts to trace the source of spillover and anticipate future emergences^{64,65}. Ebola is
174 far from unique: with 5,762–11,122 first encounters between bats and primates alone
175 leading to an expected 57–181 new viral sharing events across scenarios (Figure 3D),
176 many potential zoonoses are likely to experience new evolutionary opportunities because
177 of climate change.

178 Future hotspots of novel assemblages and viral evolution are projected to coincide
179 areas of high human population density, further increasing vulnerability to potential
180 zoonoses. First encounters are disproportionately likely to occur in areas that are pro-
181 jected to be either human settled or used as cropland, and surprisingly less likely to
182 occur in forests, which current literature highlights as producing most emerging diseases
183 (Figure 4)²⁷. This finding is consistent for bats and non-bats, and may be an accident
184 of geography, but more likely represents the tendency of human settlements to aggre-
185 gate on continental edges and around biodiversity hotspots⁶⁶. Regardless of mechanism,
186 we predict that tropical hotspots of novel viral sharing will broadly coincide with high
187 population density areas in 2070, especially in the Sahel, the Ethiopian highlands and
188 the Rift Valley, India, eastern China, Indonesia, and the Philippines (Figure 4). Some

European population centers also land in these hotspots; recent emergences in this region like Usutu virus⁶⁷ highlight that these populations can still be vulnerable, despite greater surveillance and healthcare access. If range-shifting mammals create ecological release for undiscovered zoonoses, populations in these areas are likely to be the most vulnerable.

Whereas most studies agree that climate change mitigation through reducing greenhouse gas emissions will prevent extinctions and minimize harmful ecosystem impacts, our results suggest that mitigation cannot reduce the likelihood of climate-driven viral sharing. Instead, the mildest, slowest scenarios for biotic homogenization appear likely to produce the most cross-species viral transmission: when climate velocity is lowest, species can successfully track shifting climate optima, leading to more range expansion, and more first encounters. Accounting for dispersal limits, species gained an average of 75% range in the mildest pathway (RCP 2.6); in comparison, only 28% of species experienced a net expansion in the most extreme pathway (RCP 8.5), for an average of 21% range gain. (ED Figure 3A) In fact, in the warmest scenario, up to 326 species lost their entire range, with 168 attributable to dispersal limits alone. As a result, there were 5% fewer first encounters in RCP 8.5 compared to RCP 2.6, and unexpectedly, a 2% reduction in the connectivity of the future global sharing network. (ED Figure 3B,D) Overall, our results indicate that a mild perturbation of the climate system could create thousands of new eco-evolutionary opportunities for viruses. We caution that this does not imply a possible upside to catastrophic warming, which will be accompanied by mass defaunation, devastating disease emergence, and unprecedented levels of human displacement and global instability. Rather, our results highlight the urgency of better wildlife surveillance systems and health infrastructure as a form of climate change adaptation, even if mitigation efforts are successful and global temperatures stay under +2°C.

Our study establishes a macroecological link between climate change and cross-species viral transmission. In practice, the patterns we describe are likely to be complicated by several ecological factors, including the temperature sensitivity of viral host jumps⁶⁸; the possibility that defaunation especially at low elevations might interact with disease prevalence through biodiversity dilution and amplification effects, not captured by our models⁶⁹; or temporal heterogeneity in exposure (hosts might exchange viruses in passing but not overlap by 2070, especially in warmer scenarios). Future work can also expand the scope of our findings to other host-parasite systems; our novel approach, which combines viral sharing models with massive species distribution modeling pipelines, is readily applied to other datasets. Birds have the best documented virome after mammals, and changing migration targets in a warming world may be especially important targets for prediction. With amphibians facing disproportionately high extinction rates due to a global fungal panzootic, and emerging threats like ranavirus causing conservation

227 concern, viral exchange among amphibians may be especially important information for
 228 conservation practitioners⁷⁰. Finally, marine mammals are an important target given
 229 their exclusion here, especially after a recent study implicating reduced Arctic sea ice in
 230 viral sharing among sympatric pinnipeds and sea otters—a result that may be the first
 231 proof of concept for our proposed climate-disease link⁷¹.

232 Because hotspots of cross-species transmission are predictable, our study provides
 233 the first template for how surveillance could target *future* hotspots of viral emergence in
 234 wildlife. In the next decade alone, over a billion dollars could be spent on a proposed
 235 global effort to identify zoonotic threats before they spread from wildlife reservoirs into
 236 human populations². These efforts are being undertaken during the greatest period
 237 of global ecological change recorded in human history, and in a practical sense, the
 238 rapid movement of species and formation of no-analog communities poses an unexpected
 239 challenge for virological research. While several studies have addressed how range shifts
 240 in zoonotic reservoirs might expose humans to novel viruses, few have considered the fact
 241 that most new exposures will be among mammal species. Tracking spillover into humans
 242 is paramount, but so is tracking of viral sharing in wildlife, and targeting surveillance in
 243 hotspots of future sharing may help researchers identify host jumps early on.

Methods

In this study, we develop global maps for terrestrial mammals that model their ecological niche as a function of climate and habitat use. We project these into paired climate-land use futures for 2070, with dispersal limitations set by biological constraints for each species. We predict the probability of viral sharing among species pairs using a model of the mammalian viral sharing network that is trained on phylogenetic relatedness and current geographic range overlaps. With that model, we map the projected hotspots of new viral sharing in different futures. All analysis code is available at github.com/cjcarlson/iceberg.

Mapping species distributions

We developed species distribution models for a total of 3,870 species in this study, divided into two modeling pipelines based on data availability (ED Figures 8, 9).

Data Collection

We scraped the Global Biodiversity Informatics Facility (GBIF) for mammal occurrence records, and developed species distribution models for all 3,870 species with at least 3 unique terrestrial presence records on a 25 km by 25 km grid (one unique point per grid cell). This grain was chosen based on the availability of future land use projections (see below). Spatial and environmental outliers were removed based on Grubb outlier tests (p-value of $1e-3$)⁷².

Poisson point process models

For 3,088 species with at least 10 unique presence records, Poisson point process models (closely related to Maxent) were fit using regularized downweighted Poisson regression⁷³ with 20,000 background points fit with the R package `glmnet`^{74,75,74}. The spatial domain of predictions was chosen based on the continent(s) where a species occurred in their IUCN range map. We trained species distribution models on current climate data using the WorldClim 2 data set⁷⁶, using mean annual temperature, mean diurnal temperature range, annual precipitation, precipitation seasonality, and precipitation in warmest quarter/ (precipitation in warmest quarter + precipitation in coldest quarter). These predictors were chosen based on having global correlations <0.7 among one another. These candidate predictors were further filtered on a species-by-species basis, retaining the maximum number of predictors with correlation <0.7 within the domain where the model was fit.

Models were fit with 5-fold cross validation, where folds were assigned based on spatial clusters to remove the influence of spatial autocorrelation on cross-validation performance statistics. Linear (all species), quadratic (species with >100 records), and product (species with >200 records) features were used. The regularization parameter was determined based on 5-fold cross-validation with each fold, choosing a value 1 standard deviation below the minimum deviance⁷⁷. This resulted in five models per species which were then combined in an unweighted ensemble. Continuous predictions of the ensemble were converted to binary presence/absence predictions by choosing a threshold based on the 5th percentile of the ensemble predictions at training presence locations.

When models were projected into the future, we limited extrapolation to 1 standard deviation beyond the data range of presence locations for each predictor. This decision balances a small amount of extrapolation based on patterns in a species niche with limiting the influence of monotonically increasing marginal responses, which can lead to statistically unsupported (and likely biologically unrealistic) responses to climate.

Range bagging models

For an additional 783 rare species (3 to 9 unique points on the 25 km grid), we produced species distribution models with a simpler range bagging algorithm, a stochastic hull-based method that can estimate climate niches from an ensemble of underfit models^{78,79}, and is therefore well suited for smaller datasets. From the full collection of presence observations and environmental variables range-bagging proceeds by randomly sampling a subset of presences (proportion p) and a subset of environmental variables (d). From these, a convex hull around the subset of points is generated in environmental space. The hull is then projected onto the landscape with a location considered part of the species range if its environmental conditions fall within the estimate hull. The subsampling is replicated N times, generating N ‘votes’ for each cell on the landscape. One can then choose a threshold for the number of votes required to consider the cell as part of the species’ range to generate the binary map used in our downstream analyses. Based on general guidelines in⁷⁸ we chose $p = 0.33$, $d = 2$, and $N = 100$. We then chose the voting threshold to be 0.165 ($=0.33/2$) because this implies that the cell is part of the range at least half the time for each subsample. Upon visual inspection, this generally lead to predictions that were very conservative about inferring that unsampled locations were part of a species distribution. The same environmental predictors and ecoregion-based domain selection rules were used for range bagging models as were used for the point process models discussed above. This hull-based approach is particularly valuable for poorly sampled species which may suffer from sampling bias because bias within niche limits has little effect on range estimates.

Model validation

PPM models performed well, with a mean test AUC under 5 fold cross-validation (using spatial clustering to reduce inflation) of 0.77 (s.d. 0.13). The mean partial AUC evaluated over a range of sensitivity relevant for SDM (0.8-0.95) was 0.8 (s.d. 0.08). The mean sensitivity of binary maps used to assess range overlap (based on the 5% training threshold used to make a binary map) was 0.89 (s.d. 0.08). Range bagging models were difficult to meaningfully evaluate because they were based on extremely small sample sizes (3-9). The mean training AUC (we did not perform cross-validation due to small sample size) was 0.96 (s.d. 0.09). The binary maps had perfect sensitivity (1) because the threshold used to make them was chosen sufficiently low to include the handful of known presences for each species. One way to assess how much we inferred the range for these species is to quantify how much of the range was estimated based on out models, based on the number of (10km) cells predicted to be part of the species range even when it was not observed there. The mean number of cells inferred to contain a presence was 253 (s.d. 448); however, the distribution is highly right skewed with a median of 94. This indicates that the range bagging models were typically relatively conservative about inferring ranges for poorly sampled species.

Habitat range and land use

We used the Land Use Harmonization version 2.0 (LUH2) gridded dataset to capture global patterns in land cover for the present and future⁸⁰. These data are derived from an integrative assessment model that pairs land use scenarios with representative concentration pathways. For the current models, we used historical land-use maps (LUH2 v2h), which are intended for use over the period 850 to 2015 C.E.⁸¹. To capture species' habitat preference, we collated data for all 3,870 mammal species from the IUCN Habitat Classification Scheme version 3.1. We then mapped 104 unique IUCN habitat classifications onto the eight land use types present in the LUH dataset. For 962 species, no habitat data was available, or no correspondence existed between a land type in the IUCN scheme and our land use data; for these species, land use filters were not used. Filtering based on habitat was done conservatively: species were allowed in current and future ranges to exist in a pixel if any non-zero percent was assigned a suitable habitat type; almost all pixels contain multiple habitats. In some scenarios, human settlements cover at least some of a pixel for most of the world, allowing synanthropic species to persist throughout most of their climatically-suitable range. For those with habitat data, the average reduction in range from habitat filtering was 7.6% of pixels.

Refining the dataset

Of the 3,870 species for which we generated distribution models, 103 were aquatic mammals (cetaceans, sirenians, pinnipeds, and sea otters), and 382 were not present in the mammalian supertree that we used for phylogenetic data⁸². These species were excluded. Aquatic species were removed using a two-filter approach, by cross-referencing with Pantheria⁸³. These results were verified by checking no species only had marine habitat use types (see ‘Habitat range and land use’). We also excluded 246 monotremes and marsupials because the shape of the supertree prevented us from fitting satisfactory GAMM smooths to the phylogeny effect, leaving 3,139 non-marine Eutherian mammals with associated phylogenetic data.

Predicting future species distributions

We modeled a total of 16 possible futures, produced by four paired climate-land use change pathways and two optional filters on species ranges (habitat preferences and dispersal limits). The full matrix of possible scenarios captures a combination of scenario uncertainty about global change and epistemological uncertainty about how best to predict species’ range shifts. By filtering possible future distributions based on climate, land use, and dispersal constraints, we aimed to maximize realism; our predictions were congruent with extensive prior literature on climate- and land use-driven range loss^{84,85,86}.

Climate and land use futures

Species distribution models were projected for 2070 using climate models, and then spatially filtered by land use projections. Climate and land-use future pathways are coupled by the Land Use Harmonization 2.0 integrative assessment model^{87,81}, such that every future has a representative concentration pathway (RCP) for climate and a shared socioeconomic pathway (SSP) for land use. For climate we used the HadGEM2 Earth System Model projections for 2070, with the four standard RCPs: 2.6, 4.5, 6.0, and 8.5 (where the values represent added W/m² of solar radiation by the end of the century due to greenhouse gas emissions). These were respectively paired with SSP 1 (“Sustainability”); SSP 2 (“Middle of the Road”); SSP 4 (“Inequality”); and SSP 5 (“Fossil-Fueled Development”).

These pairings can be thought of as a gradient of scenarios of global change with different levels of severity and sustainability. Not all scenarios are possible; the four we selected are drawn as some of the most representative from an underlying “scenario matrix” that includes every possible parameterization, some of which are entirely incompatible⁸⁸. (For example, in the vast majority of integrative assessment models, decarbonization cannot

be achieved fast enough in the SSP 5 scenario to achieve RCP 2.6.) As pairs, SSP-RCP narratives can be merged to create overall narratives about how global change could look. For example, in SSP 1-RCP 2.6, a global transition to renewable energy and mitigation of climate change corresponds to sustainable population growth and economic development. Driven by international cooperation on climate agreements, afforestation and bioenergy cropland become major land uses, while tropical deforestation is strongly reduced. In contrast, in SSP 5-RCP 8.5, business-as-usual development leads to catastrophic levels of warming, unsustainable population growth and increasing poverty, and massive land conversion^{89,90}.

Limiting dispersal capacity

Not all species can disperse to all environments, and not all species have equal dispersal capacity—in ways likely to covary with viral sharing properties. We follow a rule proposed by Schloss *et al.*³¹, who described an approximate formula for mammal range shift capacity based on body mass and trophic position. For carnivores, the maximum distance traveled in a generation is given as $D = 40.7M^{0.81}$, where D is distance in kilometers and M is body mass in kilograms. For herbivores and omnivores, the maximum is estimated as $D = 3.31M^{0.65}$.

We used mammalian diet data from the EltonTraits database⁹¹, and used the same cutoff as Schloss to identify carnivores as any species with 10% or less plants in their diet. We used body mass data from EltonTraits in the Schloss formula to estimate maximum generational dispersal, and converted estimates to annual maximum dispersal rates by dividing by generation length, as previously estimated by another comprehensive mammal dataset⁹². We multiply by 50 years and use the resulting distance as a buffer around the original range map, and constrain possible range shifts within that buffer. For 420 species with missing data in one of the required sources, we interpolated dispersal distance based on the closest relative in our supertree with a dispersal velocity estimate.

Qualified by the downsides of assuming full dispersal⁹³, we excluded bats from the assumed scaling of dispersal limitations. The original study by Schloss *et al.*³¹ chose to omit bats entirely, and subsequent work has not proposed any alternative formula. Moreover, the Schloss formula performs notably poorly for bats: for example, it would assign the largest bat in our study, the Indian flying fox (*Pteropus giganteus*), a dispersal capacity lower than that of the gray dwarf hamster (*Cricetulus migratorius*). Bats were instead given full dispersal in all scenarios: given significant evidence that some bat species regularly cover continental distances^{43,44}, and that isolation by distance is uncommon within many bats' ranges⁴⁶, we felt this was a defensible assumption for modeling purposes. Moving forward, the rapid range shifts already observed in many bat species

(see main text) could provide an empirical reference point to fit a new allometric scaling curve (after standardizing those results for the studies' many different methodologies). A different set of functional traits likely govern the scaling of bat dispersal, chiefly the aspect ratio (length:width) of wings, which is a strong predictor of population genetic differentiation⁴⁶. Migratory status would also be important to include as a predictor although here, we exclude information on long-distance migration for all species (due to a lack of any real framework for adding that information to species distribution models in the literature).

Explaining spatial patterns

To explore the geography of novel assemblages, we used linear models which predicted the number of first encounters (novel overlap of species pairs) at the 25km level ($N = 258,539$ grid cells). Explanatory variables included: richness (number of species inhabiting the grid cell in our predicted current ranges for the given scenario); elevation in meters (derived from the US Geological Service Global Multi-resolution Terrain Elevation Data 2010 dataset); and the predominant land cover type for the grid cell. We simplified the classification scheme for land use types into five categories for these models (human settlement, cropland, rangeland and pasture, forest, and unforested wildland), and assigned pixels a single land use type based on the maximum probability from the land use scenarios. We fitted a model for each scenario and pair of biological assumptions; because of the large effect bats had on the overall pattern, we retrained these models on subsets of encounters with and without a bat species involved. To help model fitting, we log($x+1$)-transformed the response variable (number of overlaps in the pixel) and both continuous explanatory variables (meters of elevation above the lowest point and species richness). Because some elevation values were lower than 0 (i.e., below sea level), we treated elevation as meters above the lowest terrestrial point rather than meters above sea level to allow us to log-transform the data.

Viral sharing models

Generalized Additive Mixed Models

We used a previously-published model of the phylogeography of viral sharing patterns to make predictions of future viral sharing¹⁷. This model was based on an analysis of 510 viruses shared between 682 mammal species³, and predicted the probability that a pair of mammal species will share a virus given their geographic range overlap and phylogenetic relatedness. The original study uncovered strong, nonlinear effects of spatial overlap and phylogenetic similarity in determining viral sharing probability, and simu-

lating the unobserved global network using these effect estimates capitulated multiple macroecological patterns of viral sharing.

In the original study, a Generalized Additive Mixed Model (GAMM) was used to predict virus sharing as a binary variable, based on (1) geographic range overlap; (2) phylogenetic similarity; and (3) species identity as a multi-membership random effect. The phylogeographic explanatory variables were obtained from two broadly available, low-resolution data sources: pairwise phylogenetic similarity was derived from a mammalian supertree previously modified for host-pathogen studies^{82,3}, with similarity defined as the inverse of the cumulative branch length between two species, scaled to between 0 and 1. Geographic overlap was defined as the area of overlap between two species' IUCN range maps, divided by their cumulative range size⁹⁴.

We first retrained the GAMMs from¹⁷ on the pairwise overlap matrix of species distribution models generated for this study, so that present predictions would be comparable with future distributions. Of the 3,139 species in our reduced dataset, 544 had viral records in our viral sharing dataset and shared with at least one other mammal, and were used to retrain the GAMM from¹⁷. To check the performance of the GAMM, we predicted sharing patterns with a) only random effects, b) only fixed effects, and c) with both. Although species-level random effects had a mean effect of ~ 0 , excluding them entirely resulted in a substantial underestimation of the mean viral sharing rates across the network (mean sharing ≈ 0.02 compared to ≈ 0.06). Therefore to ensure that the model recapitulated traits of the observed network, we simulated 1,000 binary sharing networks when predicting with only fixed effects, randomly drawing species-level random effects in each iteration. The mean sharing value across these iterations closely approximated observed sharing probability (~ 0.06).

Model validation and limits

Compared to the current viral sharing matrix, the model performs well with only fixed effects (AUC = 0.80) and extremely well with both fixed and random effects (AUC = 0.93). The model explained a very similar proportion of the deviance in viral sharing to that in Albery *et al.*¹⁷ (44.5% and 44.8% respectively).

In practice, several unpredictable but confounding factors could affect the reliability of this model as a forecasting tool, including temperature sensitivity of viral evolution in host jumps⁶⁸, or increased susceptibility of animals with poorer health in lower-quality habitat or unfavorable climates. Moreover, once viruses can produce an infection, their ability to transmit *within* a new species is an evolutionary race between mutation and recombination rates in viral genomes, host innate and adaptive immunity, virulence-related mortality, and legacy constraints of coevolution with prior hosts and vectors^{60,61}.

486 But data cataloging these precise factors are hardly comprehensive for the hundreds of
487 zoonotic viruses, let alone for the thousands of undescribed viruses in wildlife. Moreover,
488 horizontal transmission is not necessary for spillover potential to be considered significant;
489 for example, viruses like rabies or West Nile virus are not transmitted within human
490 populations but humans are still noteworthy hosts.

491 **Mapping opportunities for sharing**

492 We used the GAMM effect estimates to predict viral sharing patterns across the 3,139
493 mammals with associated geographic range and phylogenetic data, for both the present
494 and future scenarios. By comparing current and future sharing probabilities for each of
495 the four global change scenarios, we estimated which geographic and taxonomic patterns
496 of viral sharing would likely emerge. We separately examined patterns of richness, pat-
497 terns of sharing probability, and their change (i.e., future sharing probability - current
498 sharing probability, giving the expected probability of a novel sharing event).

499 A subset of the mammals in our dataset were predicted to encounter each other for the
500 first time during range shifts. For each of these pairwise first encounters, we extracted the
501 area of overlap in every future scenario, and assigned each overlap a probability of sharing
502 from the mean GAMM predictions and mapped the mean and cumulative probability of
503 a new sharing event happening in a given geographic pixel.

504 **Case study on Zaire ebolavirus**

505 For a case study in possible significant cross-species transmission, we compiled a list
506 of known hosts of Zaire ebolavirus (ZEBOV), a zoonosis with high host breadth that
507 has been known to cause wildlife die-offs, but has no known definitive reservoir. Hosts
508 were taken from two sources: the training dataset on host-virus associations³, and an
509 additional dataset of filovirus testing in bats³⁰. In the latter case, any bats that have
510 been reported antibody positive or PCR-positive for ZEBOV were included. A total
511 of 13 current “known hosts” in Africa were used to predict current possible hosts, and
512 first encounters in all scenarios. We restricted our analysis to Africa because there is
513 no published evidence that Zaire ebolavirus actively circulates outside Africa; although
514 some bat species outside Africa have tested positive for antibodies to ZEBOV, this is
515 likely due to cross-reactivity with other undiscovered filoviruses^{95,96,30}.

516 **Overlap with human populations**

517 To examine the possibility that hotspots of cross-species transmission would overlap with
518 human populations, we used SEDAC’s global population projections version 1.0 for the

519 year 2070⁹⁷. We aggregated these to native resolution, for each of the four SSP paired
520 with the native RCP/SSP pairing for the species distribution models. In Figure 4 we
521 present the population projections for SSP 1, which pairs with RCP 2.6.

522 Acknowledgements

523 This paper is the culmination of several years of idea development and owes special thanks
 524 to many people, including the entire Bansal Lab, Laura Ward Alexander, Kevin Burgio,
 525 Eric Dougherty, Romain Garnier, Wayne Getz, Peta Hitchens, Christine Johnson, and
 526 Isabel Ott. We especially thank Laura Alexander for sharing bat filovirus testing sources
 527 used to compile the Ebola sub-network. Thanks are also extended to José Hidasi-Neto for
 528 publicly-available data visualization code. This research was supported by the George-
 529 town Environment Initiative and the National Socio-Environmental Synthesis Center
 530 (SESYNC) under funding received from the National Science Foundation DBI-1639145.
 531 C.M. acknowledges funding from National Science Foundation grant DBI-1913673.

532 Author Contributions

533 CJC and GFA conceived the study. CM, CJC, and CHT developed species distribution
 534 models; GFA, EAE, KJO, and NR developed the generalized additive models. CJC, GFA,
 535 and CMZ integrated the predictions of species distributions and viral sharing patterns
 536 and designed visualizations. All authors contributed to the writing of the manuscript.

537

Figures

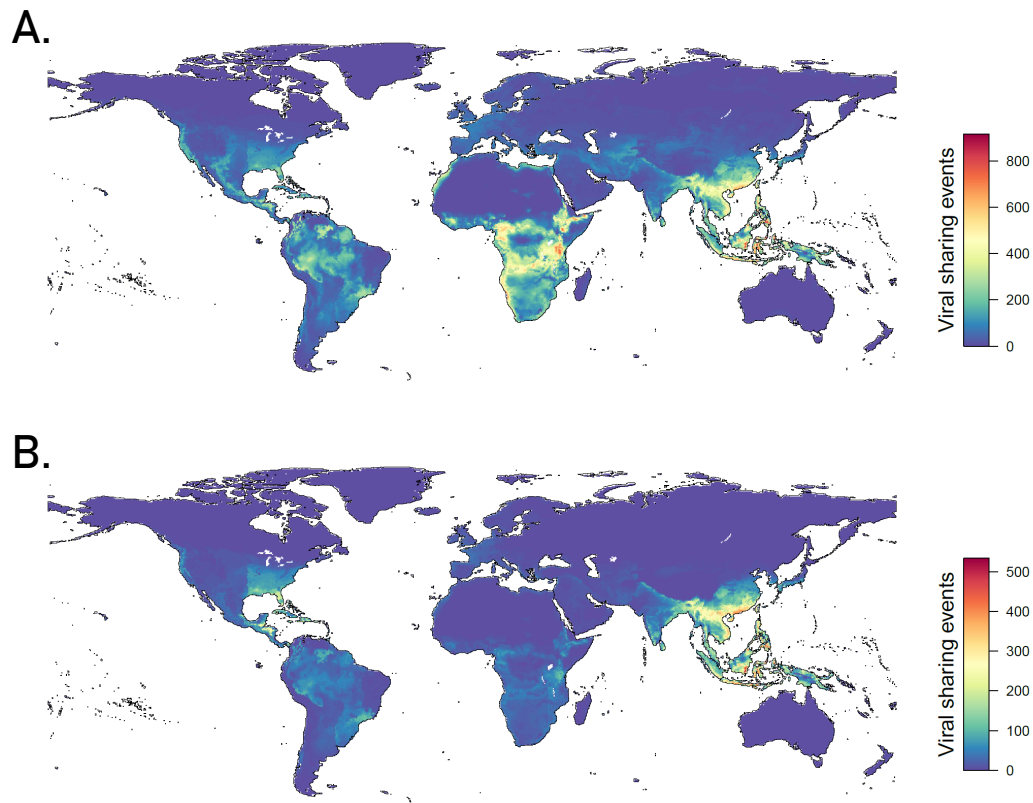


Figure 1: Climate change will drive novel viral sharing among mammal species. The projected number of novel viral sharing events among mammal species in 2070 based on host species geographic range shifts from climate change (RCP 2.6) and land-use change (SSP 1), without dispersal limits (A) and with dispersal limitation (B).

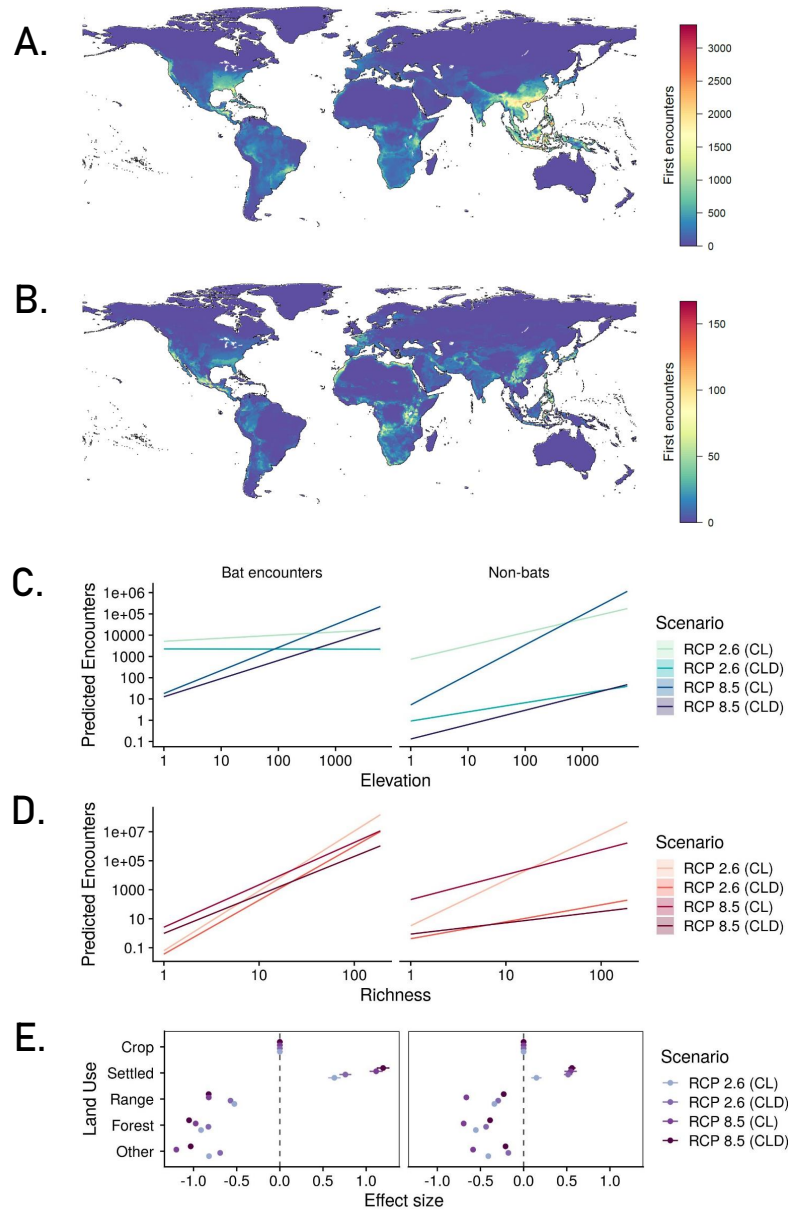


Figure 2: Bats disproportionately drive future novel viral sharing. The spatial pattern of first encounters differs among range-shifting mammal pairs including bat-bat and bat-nonbat encounters (A) and only encounters among non-bats (B). Using a linear model, we show that elevation (C), species richness (D), and land use (E) together explain 57.7% of deviance in new overlaps for bats, and 25.8% for non-bats. Slopes for the elevation effect were generally steeply positive: a \log_{10} -increase in elevation was associated with between a 0.4-1.41 \log_{10} -increase in first encounters.

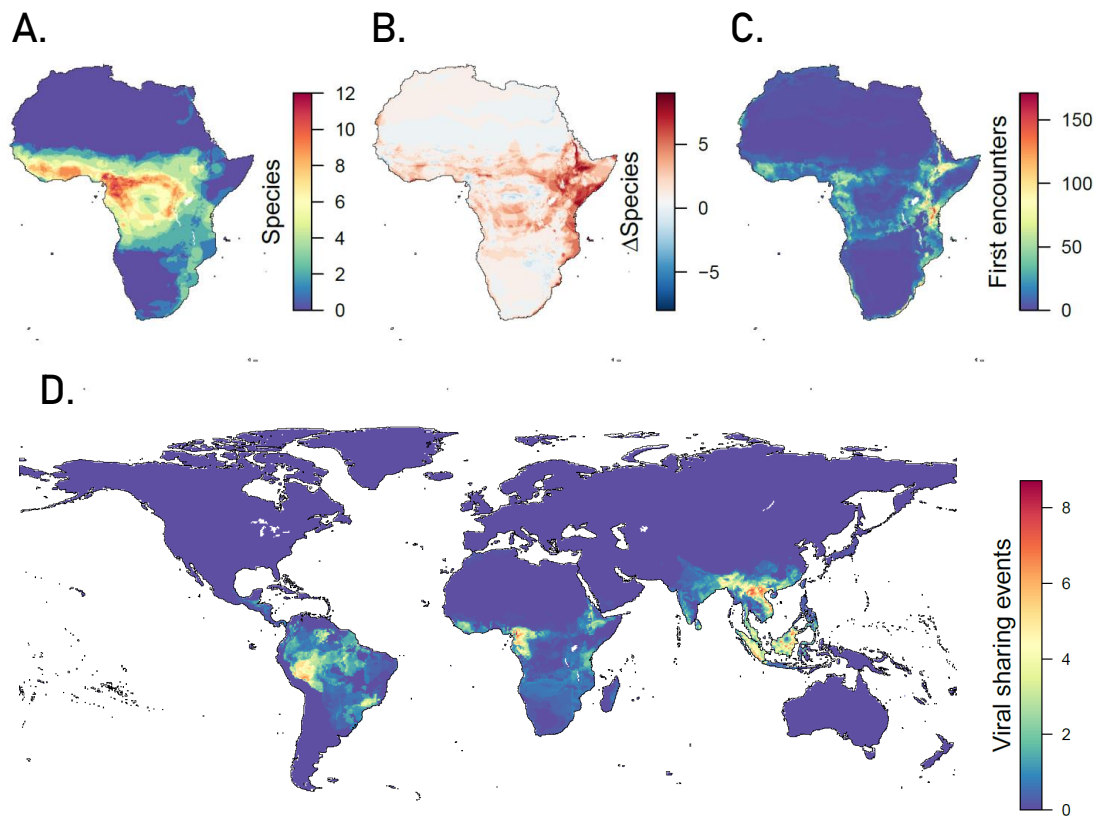


Figure 3: **Range expansions will expose naive hosts to zoonotic reservoirs.** (A) The predicted distribution of known African hosts of Zaire ebolavirus. (B) The change in richness of these hosts as a result of range shifts. (C) Projected first encounters with non-Ebola hosts. (D) Bat-primate first encounters are projected to occur globally, producing novel sharing events.

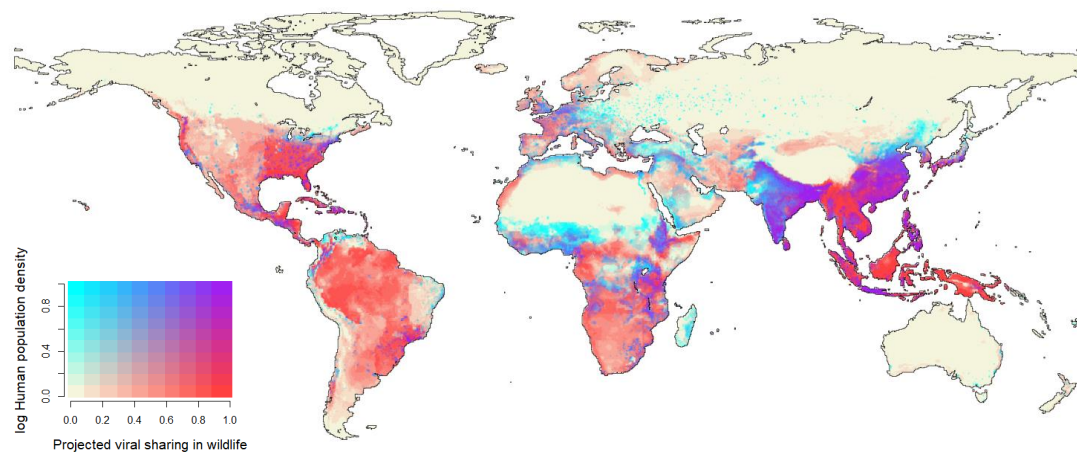
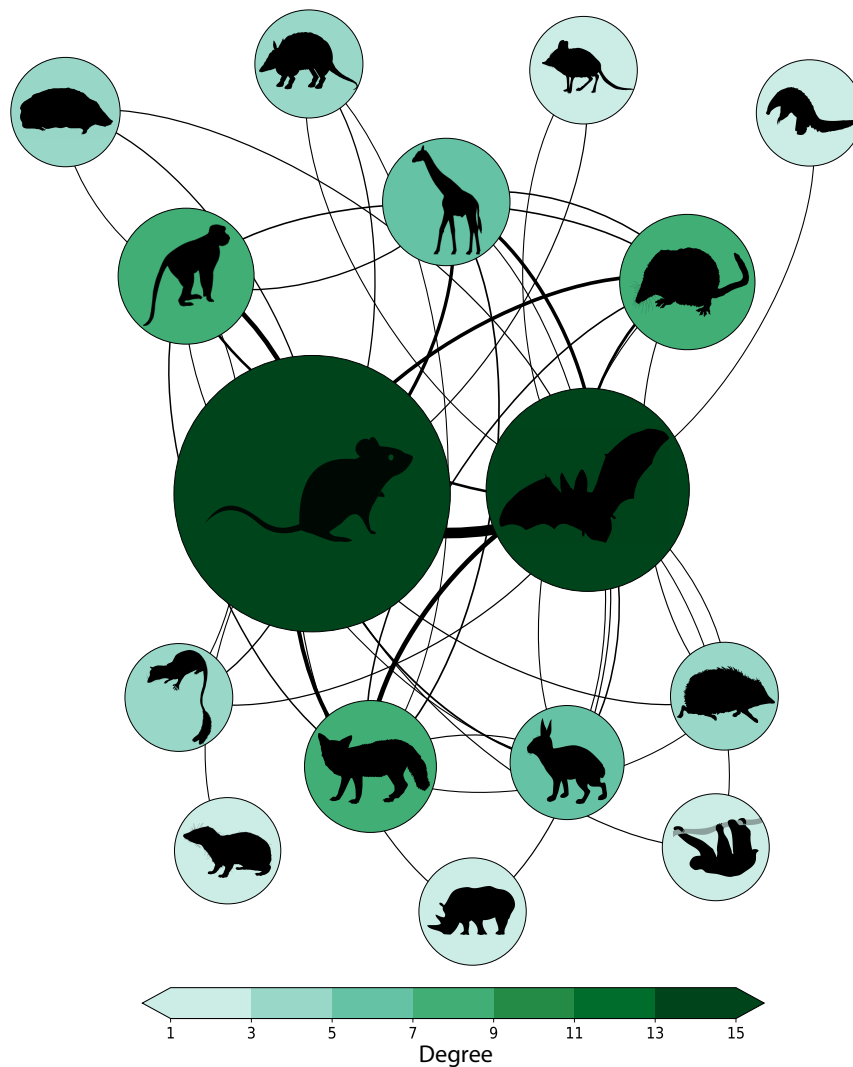
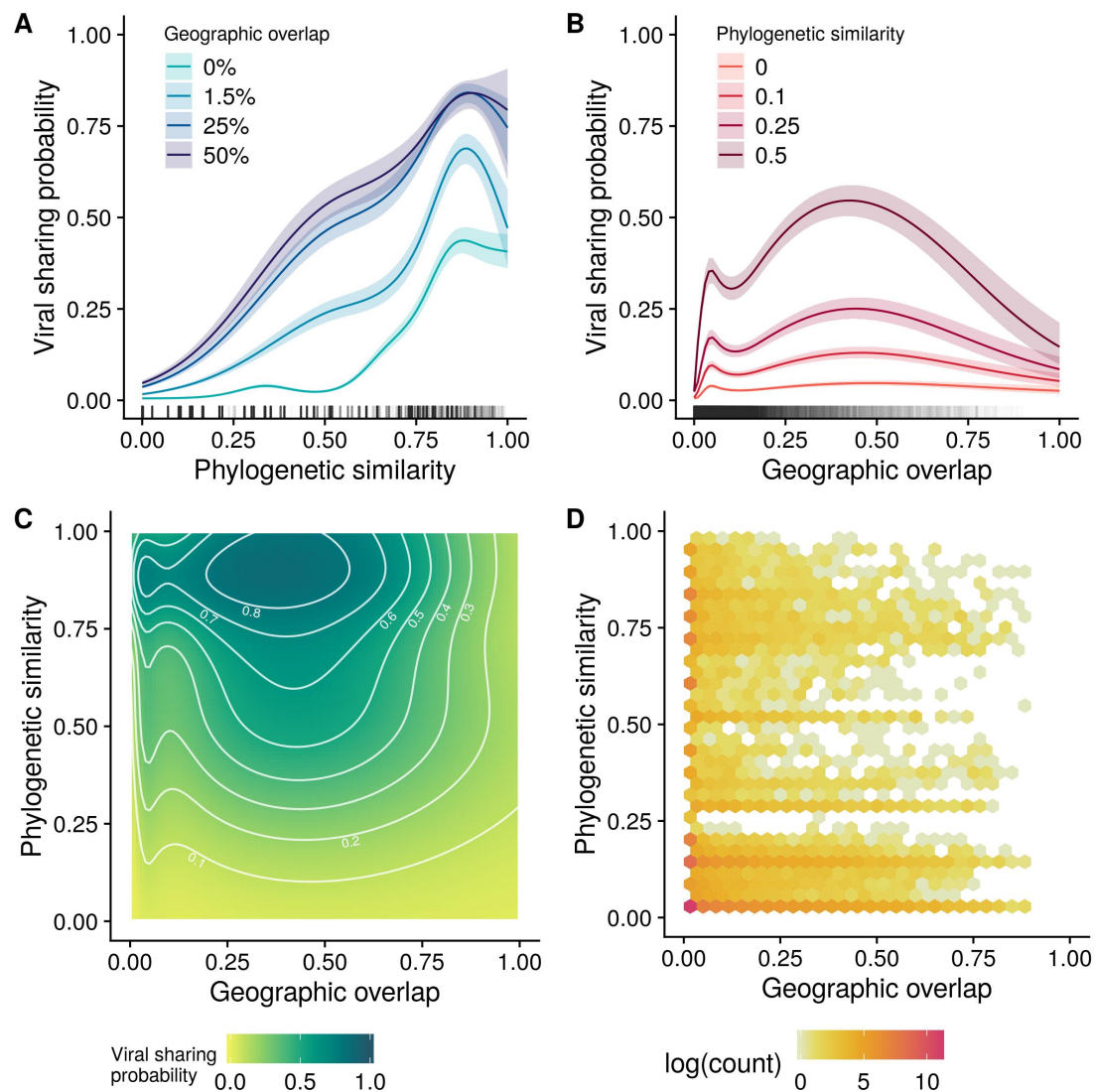


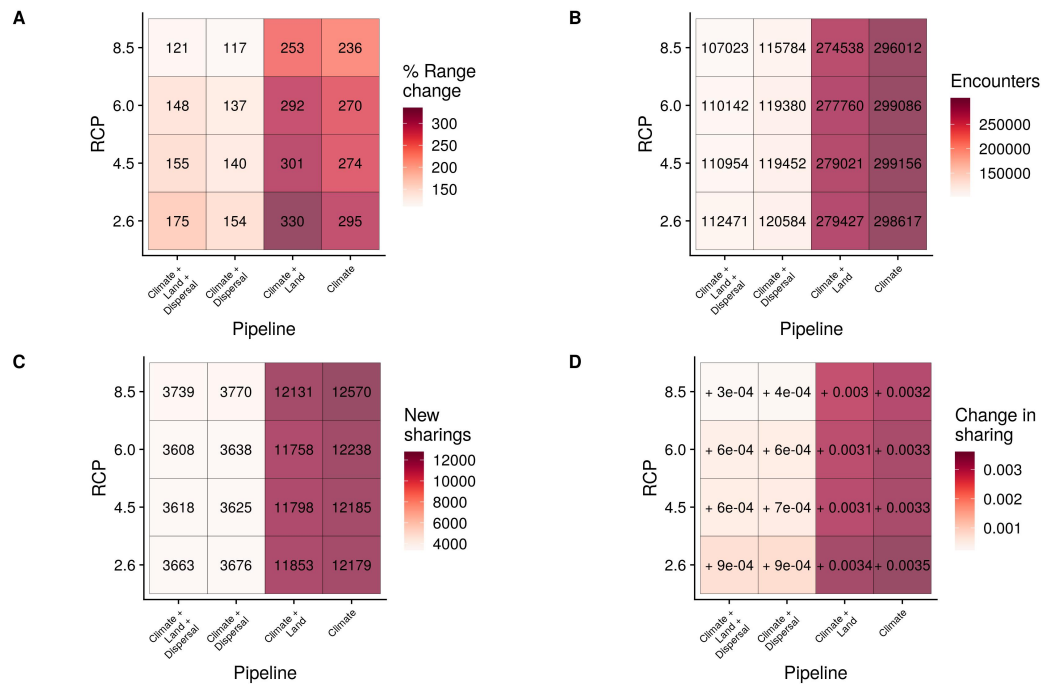
Figure 4: Novel viral sharing events coincide with population centers. In 2070 (RCP 2.6; climate only), human population centers in equatorial Africa, south China and southeast Asia will overlap with projected hotspots of cross-species viral transmission in wildlife. (Both variables are linearly rescaled to 0 to 1.)



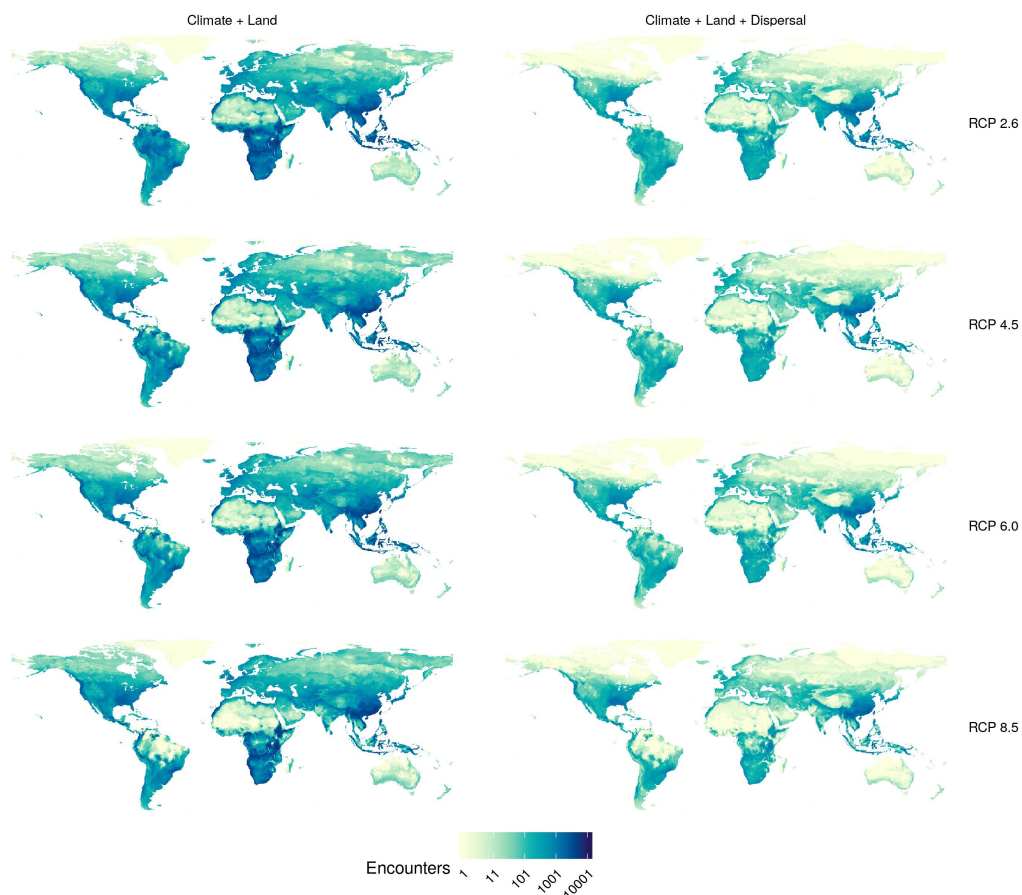
Extended Data Figure 1: **The mammal-virus network.** The present-day viral sharing network by mammal order inferred from modeled pairwise predictions of viral sharing probabilities. Edge width denotes the expected number of shared viruses (the sum of pairwise species-species viral sharing probabilities), with most sharing existing among the most speciose and closely-related groups. Edges shown in the network are the top 25% of links. Nodes are sized by total number of species in that order in the host-virus association dataset, color is scaled by degree.



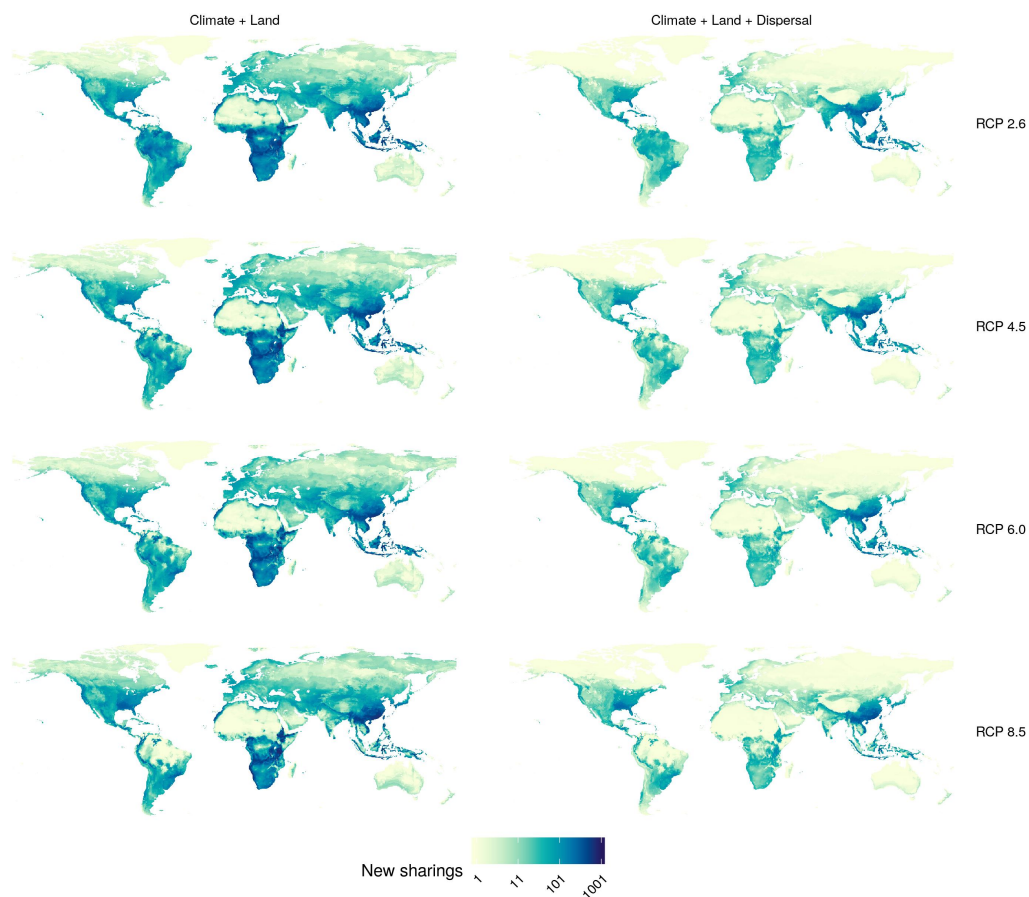
Extended Data Figure 2: **Predicted phylogeographic structure of viral sharing.** Phylogeographic prediction of viral sharing using a generalized additive mixed model. Viral sharing increases as a function of phylogenetic similarity (A) and geographic overlap (B), fit together as a tensor interaction (C). White contour lines denote 10% increments of sharing probability. Declines at high values of overlap may be an artefact of model structure and low sampling in the upper levels of geographic overlap, shown in a hexagonal bin chart for raw data (D).



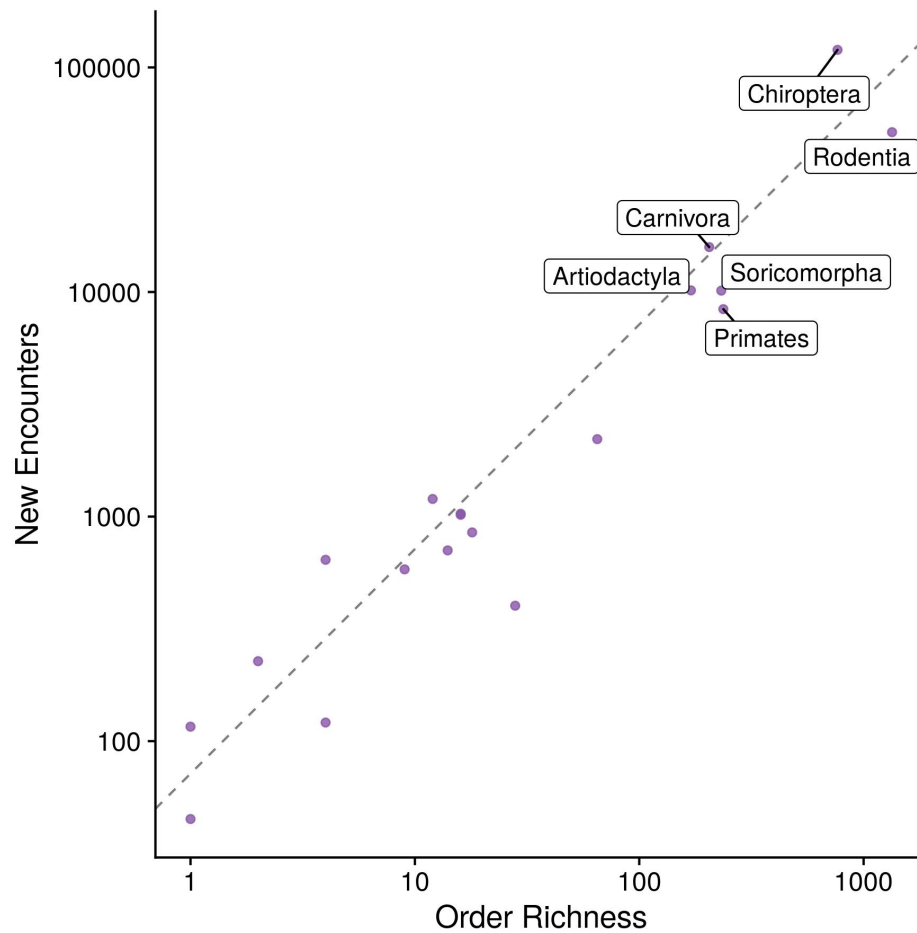
Extended Data Figure 3: **Outcomes by model formulation and climate change scenario.** Heatmaps displaying predicted changes across model formulations. (A) Range expansions were highest in non-dispersal-limited scenarios and in milder RCPs. (B) The number of predicted first encounters was higher in non-dispersal-limited scenarios and in milder RCPs. (C) The number of expected new viral sharing events was higher in non-dispersal-limited scenarios and in more severe RCPs. (D) The overall change in sharing probability (connectance) across the viral sharing network between the present day and the future scenarios; absolute change is minimal but positive across all scenarios, being greatest in non-dispersal-limited scenarios and in milder RCPs.



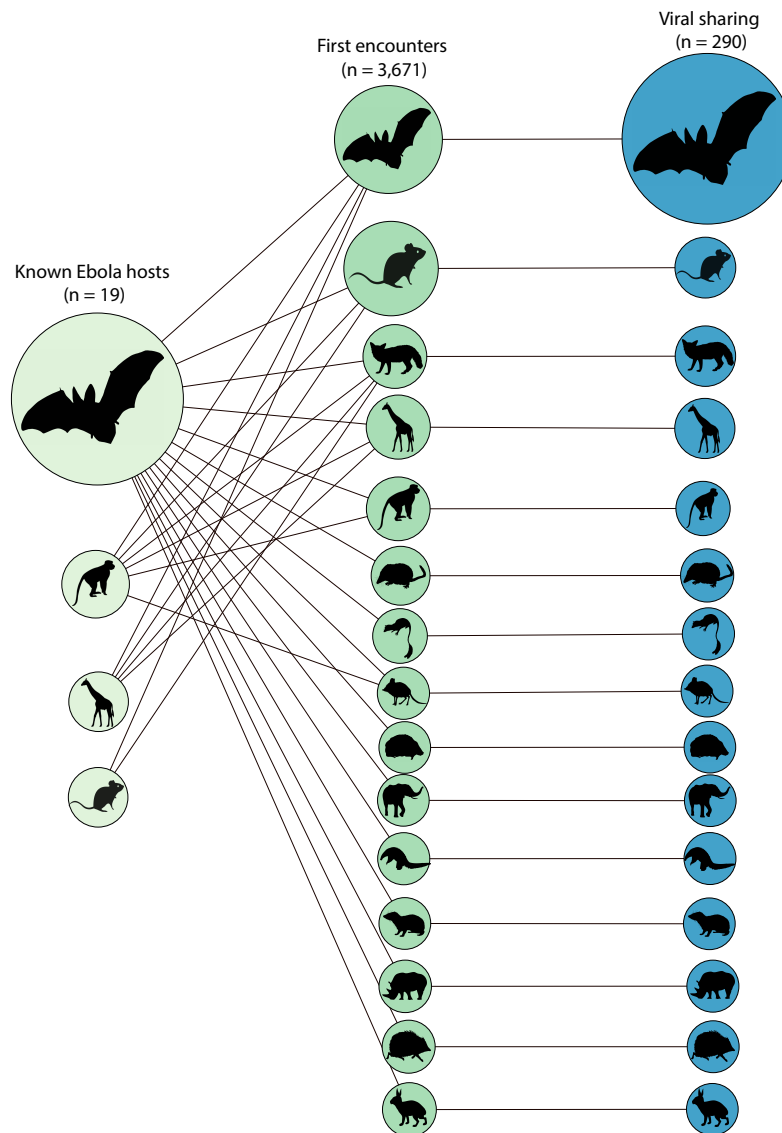
Extended Data Figure 4: **Geographic distribution of first encounters.** Predictions were carried out for four representative concentration pathways (RCPs), accounting for climate change and land use change, without (left) and with dispersal limits (right). Darker colours correspond to greater numbers of first encounters in the pixel.



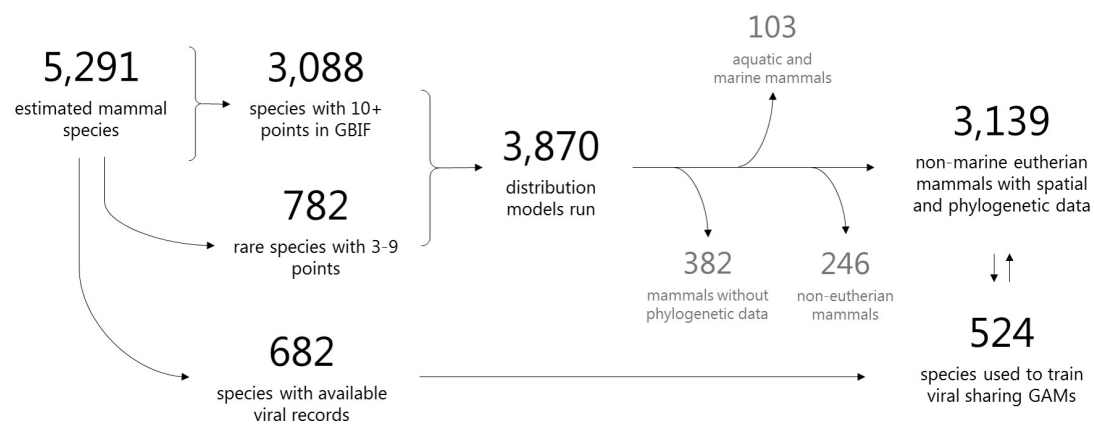
Extended Data Figure 5: **Geographic distribution of expected viral sharing events from first encounters.** Predictions were carried out for future distributions for four representative concentration pathways (RCPs), accounting for climate change and land use change, without (left) and with dispersal limits (right). Darker colours correspond to greater numbers of new viral sharing events in the pixel. Probability of new viral sharing was calculated by subtracting the species pair's present sharing probability from their sharing probability that our viral sharing GAMMs predicted. This probability was projected across the species pair's range intersection, and then summed across all novel species pairs in each pixel.



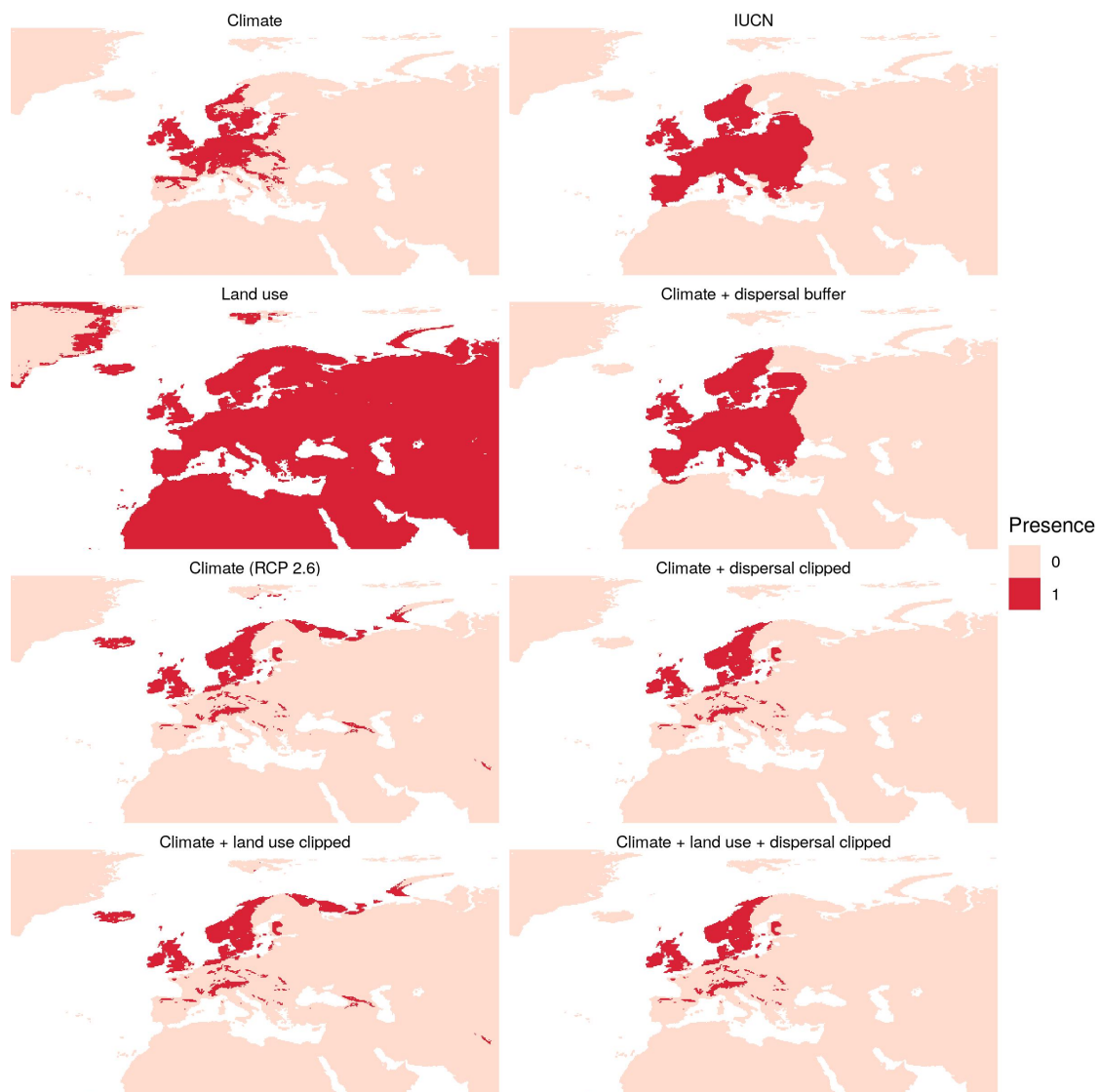
Extended Data Figure 6: **Order-level heterogeneity in first encounters.** Dispersal stratifies the number of first encounters (RCP 2.6 with all range filters), where some orders have more than expected at random, based on the mean number of first encounters and order size (line).



Extended Data Figure 7: **Projected viral sharing from suspected Ebola reservoirs is dominated by bats.** Node size is proportional to (left) the number of suspected Ebola host species in each order, which connect to (middle) first encounters with potentially naive host species; and (right) the number of projected viral sharing events in each receiving group. (Node size denotes proportions out of 100% within each column total.) While Ebola hosts will encounter a much wider taxonomic range of mammal groups than current reservoirs, the vast majority of viral sharing will occur disproportionately in bats.



Extended Data Figure 8: **Data processing workflow.** Summary of species inclusion across the modeling pipeline for species distributions and viral sharing models. The final analyses in the main text use 3,139 species of Eutherian mammals across all scenarios.



Extended Data Figure 9: **Species distribution modeling workflow for a single species.** A focal species (the European red deer, *Cervus elaphus*) is displayed as an illustrative example. The present day climate prediction (top left) was clipped to the same continent according to the IUCN distribution (top right). This was then clipped according to *Cervus elaphus* land use (second row, left). The known dispersal distance of the red deer was used to buffer the climate distribution (second row, right). The future distribution predictions (RCP 2.6 shown as an example) are displayed in the bottom four panels, for each of the four pipelines: only climate (third row, left); climate + dispersal clip (third row, right); climate + land use clip (bottom row, left) and climate + land use + dispersal clip (bottom row, right). The four distributions clearly display the limiting effect of the dispersal filter (bottom right panels) in reducing the probability of novel species interactions (bottom left panels). The land use clip had little effect on this species as the entire distribution area was habitable for the red deer.

References

538

- 539 1. Carlson, C. J, Zipfel, C. M, Garnier, R, & Bansal, S. (2019) Global estimates
540 of mammalian viral biodiversity accounting for host sharing. *Nature Ecology and*
541 *Evolution* **3**, 1070–1075.
- 542 2. Carroll, D, Daszak, P, Wolfe, N. D, Gao, G. F, Morel, C. M, Morzaria, S, Pablos-
543 Méndez, A, Tomori, O, & Mazet, J. A. (2018) The global virome project. *Science*
544 **359**, 872–874.
- 545 3. Olival, K. J, Hosseini, P. R, Zambrana-Torrel, C, Ross, N, Bogich, T. L, & Daszak,
546 P. (2017) Host and viral traits predict zoonotic spillover from mammals. *Nature*
547 **546**, 646.
- 548 4. Altizer, S, Ostfeld, R. S, Johnson, P. T, Kutz, S, & Harvell, C. D. (2013) Climate
549 change and infectious diseases: from evidence to a predictive framework. *Science*
550 **341**, 514–519.
- 551 5. Cleaveland, S, Haydon, D, & Taylor, L. (2007) Overviews of pathogen emergence:
552 which pathogens emerge, when and why? *Wildlife and Emerging Zoonotic Diseases:*
553 *The Biology, Circumstances and Consequences of Cross-Species Transmission* pp.
554 85–111.
- 555 6. Parrish, C. R, Holmes, E. C, Morens, D. M, Park, E.-C, Burke, D. S, Calisher, C. H,
556 Laughlin, C. A, Saif, L. J, & Daszak, P. (2008) Cross-species virus transmission and
557 the emergence of new epidemic diseases. *Microbiology and Molecular Biology Reviews*
558 **72**, 457–470.
- 559 7. Woolhouse, M. E, Haydon, D. T, & Antia, R. (2005) Emerging pathogens: the
560 epidemiology and evolution of species jumps. *Trends in Ecology & Evolution* **20**,
561 238–244.
- 562 8. Hoberg, E. P & Brooks, D. R. (2015) Evolution in action: climate change, biodi-
563 versity dynamics and emerging infectious disease. *Philosophical Transactions of the*
564 *Royal Society B: Biological Sciences* **370**, 20130553.
- 565 9. Burrows, M. T, Schoeman, D. S, Richardson, A. J, Molinos, J. G, Hoffmann, A,
566 Buckley, L. B, Moore, P. J, Brown, C. J, Bruno, J. F, Duarte, C. M, et al. (2014)
567 Geographical limits to species-range shifts are suggested by climate velocity. *Nature*
568 **507**, 492.

- 569 10. Chen, I.-C, Hill, J. K, Ohlemüller, R, Roy, D. B, & Thomas, C. D. (2011) Rapid
570 range shifts of species associated with high levels of climate warming. *Science* **333**,
571 1024–1026.
- 572 11. Carlson, C. J, Burgio, K. R, Dougherty, E. R, Phillips, A. J, Bueno, V. M, Clements,
573 C. F, Castaldo, G, Dallas, T. A, Cizauskas, C. A, Cumming, G. S, et al. (2017) Par-
574 asite biodiversity faces extinction and redistribution in a changing climate. *Science*
575 *Advances* **3**, e1602422.
- 576 12. Wolfe, N. D, Dunavan, C. P, & Diamond, J. (2007) Origins of major human infectious
577 diseases. *Nature* **447**, 279.
- 578 13. Lloyd-Smith, J. O, George, D, Pepin, K. M, Pitzer, V. E, Pulliam, J. R, Dobson,
579 A. P, Hudson, P. J, & Grenfell, B. T. (2009) Epidemic dynamics at the human-animal
580 interface. *Science* **326**, 1362–1367.
- 581 14. Plowright, R. K, Parrish, C. R, McCallum, H, Hudson, P. J, Ko, A. I, Graham,
582 A. L, & Lloyd-Smith, J. O. (2017) Pathways to zoonotic spillover. *Nature Reviews*
583 *Microbiology* **15**, 502.
- 584 15. Gould, S. J & Lewontin, R. C. (1979) The spandrels of San Marco and the Panglos-
585 sian paradigm: a critique of the adaptationist programme. *Proceedings of the Royal*
586 *Society of London. Series B: Biological Sciences* **205**, 581–598.
- 587 16. Alexander, K. A, Carlson, C. J, Lewis, B. L, Getz, W. M, Marathe, M. V, Eubank,
588 S. G, Sanderson, C. E, & Blackburn, J. K. (2018) The ecology of pathogen spillover
589 and disease emergence at the human-wildlife-environment interface. *The Connections*
590 *Between Ecology and Infectious Disease* pp. 267–298.
- 591 17. Albery, G. F, Eskew, E. A, Ross, N, & Olival, K. J. (2019) Predicting the global
592 mammalian viral sharing network using phylogeography. *bioRxiv* p. 732255.
- 593 18. Faria, N. R, Suchard, M. A, Rambaut, A, Streicker, D. G, & Lemey, P. (2013) Simul-
594 taneously reconstructing viral cross-species transmission history and identifying the
595 underlying constraints. *Philosophical Transactions of the Royal Society B: Biological*
596 *Sciences* **368**, 20120196.
- 597 19. Dougherty, E. R, Seidel, D. P, Carlson, C. J, Spiegel, O, & Getz, W. M. (2018)
598 Going through the motions: incorporating movement analyses into disease research.
599 *Ecology Letters* **21**, 588–604.
- 600 20. Sun, B, Jia, L, Liang, B, Chen, Q, & Liu, D. (2018) Phylogeography, transmission,
601 and viral proteins of Nipah virus. *Virologica Sinica* **33**, 385–393.

- 602 21. Huang, S, Bininda-Emonds, O. R, Stephens, P. R, Gittleman, J. L, & Altizer, S.
603 (2014) Phylogenetically related and ecologically similar carnivores harbour similar
604 parasite assemblages. *Journal of Animal Ecology* **83**, 671–680.
- 605 22. Davies, T. J & Pedersen, A. B. (2008) Phylogeny and geography predict pathogen
606 community similarity in wild primates and humans. *Proceedings of the Royal Society*
607 *B: Biological Sciences* **275**, 1695–1701.
- 608 23. Farrell, M. J & Davies, T. J. (2019) Disease mortality in domesticated animals is
609 predicted by host evolutionary relationships. *Proceedings of the National Academy*
610 *of Sciences* **116**, 7911–7915.
- 611 24. Longdon, B, Brockhurst, M. A, Russell, C. A, Welch, J. J, & Jiggins, F. M. (2014)
612 The evolution and genetics of virus host shifts. *PLoS Pathogens* **10**, e1004395.
- 613 25. Streicker, D. G, Turmelle, A. S, Vonhof, M. J, Kuzmin, I. V, McCracken, G. F,
614 & Rupprecht, C. E. (2010) Host phylogeny constrains cross-species emergence and
615 establishment of rabies virus in bats. *Science* **329**, 676–679.
- 616 26. Jones, K. E, Patel, N. G, Levy, M. A, Storeygard, A, Balk, D, Gittleman, J. L, &
617 Daszak, P. (2008) Global trends in emerging infectious diseases. *Nature* **451**, 990.
- 618 27. Allen, T, Murray, K. A, Zambrana-Torrel, C, Morse, S. S, Rondinini, C, Di Marco,
619 M, Breit, N, Olival, K. J, & Daszak, P. (2017) Global hotspots and correlates of
620 emerging zoonotic diseases. *Nature Communications* **8**, 1124.
- 621 28. Johnson, C. K, Hitchens, P. L, Evans, T. S, Goldstein, T, Thomas, K, Clements,
622 A, Joly, D. O, Wolfe, N. D, Daszak, P, Karesh, W. B, et al. (2015) Spillover and
623 pandemic properties of zoonotic viruses with high host plasticity. *Scientific Reports*
624 **5**, 14830.
- 625 29. Han, B. A, Kramer, A. M, & Drake, J. M. (2016) Global patterns of zoonotic disease
626 in mammals. *Trends in Parasitology* **32**, 565–577.
- 627 30. Han, B. A, Schmidt, J. P, Alexander, L. W, Bowden, S. E, Hayman, D. T, & Drake,
628 J. M. (2016) Undiscovered bat hosts of filoviruses. *PLoS Neglected Tropical Diseases*
629 **10**, e0004815.
- 630 31. Schloss, C. A, Nuñez, T. A, & Lawler, J. J. (2012) Dispersal will limit ability of
631 mammals to track climate change in the western hemisphere. *Proceedings of the*
632 *National Academy of Sciences* **109**, 8606–8611.

- 633 32. Loarie, S. R, Duffy, P. B, Hamilton, H, Asner, G. P, & Field, C. B. (2009) The
634 velocity of climate change. *Nature* **462**, 1052–1055.
- 635 33. Hickling, R, Roy, D. B, Hill, J. K, Fox, R, & Thomas, C. D. (2006) The distributions
636 of a wide range of taxonomic groups are expanding polewards. *Global Change Biology*
637 **12**, 450–455.
- 638 34. Mason, S. C, Palmer, G, Fox, R, Gillings, S, Hill, J. K, Thomas, C. D, & Oliver,
639 T. H. (2015) Geographical range margins of many taxonomic groups continue to
640 shift polewards. *Biological Journal of the Linnean Society* **115**, 586–597.
- 641 35. Bebber, D. P, Ramotowski, M. A, & Gurr, S. J. (2013) Crop pests and pathogens
642 move polewards in a warming world. *Nature Climate Change* **3**, 985.
- 643 36. Davidson, R, Simard, M, Kutz, S. J, Kapel, C. M, Hamnes, I. S, & Robertson, L. J.
644 (2011) Arctic parasitology: why should we care? *Trends in Parasitology* **27**, 239–245.
- 645 37. Hoberg, E. P, Cook, J, Agosta, S, Boeger, W, Galbreath, K, Laaksonen, S, Kutz, S,
646 & Brooks, D. (2017) Arctic systems in the quaternary: ecological collision, faunal
647 mosaics and the consequences of a wobbling climate. *Journal of Helminthology* **91**,
648 409–421.
- 649 38. Trisos, C. H, Merow, C, & Pigot, A. L. (2020) Timing and abruptness of ecological
650 disruption from climate change. *Nature* p. in press.
- 651 39. Colwell, R. K, Brehm, G, Cardelús, C. L, Gilman, A. C, & Longino, J. T. (2008)
652 Global warming, elevational range shifts, and lowland biotic attrition in the wet
653 tropics. *Science* **322**, 258–261.
- 654 40. Pauchard, A, Milbau, A, Albiñ, A, Alexander, J, Burgess, T, Daehler, C, Englund,
655 G, Essl, F, Evengård, B, Greenwood, G. B, et al. (2016) Non-native and native
656 organisms moving into high elevation and high latitude ecosystems in an era of
657 climate change: new challenges for ecology and conservation. *Biological Invasions*
658 **18**, 345–353.
- 659 41. Atkinson, C. T & LaPointe, D. A. (2009) Introduced avian diseases, climate change,
660 and the future of hawaiian honeycreepers. *Journal of Avian Medicine and Surgery*
661 **23**, 53–63.
- 662 42. Schloss, A. L, Kicklighter, D. W, Kaduk, J, Wittenberg, U, & Intercomparison,
663 T. P. O. T. P. N. M. (1999) Comparing global models of terrestrial net primary
664 productivity (NPP): comparison of NPP to climate and the Normalized Difference
665 Vegetation Index (NDVI). *Global Change Biology* **5**, 25–34.

- 666 43. Peel, A. J, Sargan, D. R, Baker, K. S, Hayman, D. T, Barr, J. A, Cramer, G, Suu-
667 Ire, R, Broder, C. C, Lembo, T, Wang, L.-F, et al. (2013) Continent-wide panmixia
668 of an African fruit bat facilitates transmission of potentially zoonotic viruses. *Nature*
669 *Communications* **4**, 2770.
- 670 44. Riesle-Sbarbaro, S. A, Amponsah-Mensah, K, De Vries, S, Nicolas, V, Lalis, A, Suu-
671 Ire, R, Cunningham, A. A, Wood, J. L, & Sargan, D. R. (2018) The Gambian
672 epauletted fruit bat shows increased genetic divergence in the Ethiopian highlands
673 and in an area of rapid urbanization. *Ecology and Evolution* **8**, 12803–12820.
- 674 45. Wu, J. (2016) Detection and attribution of the effects of climate change on bat
675 distributions over the last 50 years. *Climatic Change* **134**, 681–696.
- 676 46. Olival, K. (2012) Evolutionary and ecological correlates of population genetic struc-
677 ture in bats. *Evolutionary history of bats fossils, molecules and morphology* pp.
678 267–316.
- 679 47. Olival, K. J, Latinne, A, Islam, A, Epstein, J. H, Hersch, R, Engstrand, R. C, Gurley,
680 E. S, Amato, G, Luby, S. P, & Daszak, P. (2019) Population genetics of fruit bat
681 reservoir informs the dynamics, distribution and diversity of Nipah virus. *Molecular*
682 *Ecology* p. in press.
- 683 48. Ancillotto, L, Santini, L, Ranc, N, Maiorano, L, & Russo, D. (2016) Extraordi-
684 nary range expansion in a common bat: the potential roles of climate change and
685 urbanisation. *The Science of Nature* **103**, 15.
- 686 49. Ancillotto, L, Budinski, I, Nardone, V, Di Salvo, I, Della Corte, M, Bosso, L, Conti,
687 P, & Russo, D. (2018) What is driving range expansion in a common bat? hints
688 from thermoregulation and habitat selection. *Behavioural Processes* **157**, 540–546.
- 689 50. Geluso, K, Mollhagen, T. R, Tigner, J. M, & Bogan, M. A. (2005) Westward expan-
690 sion of the eastern pipistrelle (*Pipistrellus subflavus*) in the United States, including
691 new records from New Mexico, South Dakota, and Texas. *Western North American*
692 *Naturalist* **65**, 12.
- 693 51. Kurta, A, Winhold, L, Whitaker, J. O, & Foster, R. (2007) Range expansion and
694 changing abundance of the eastern pipistrelle (Chiroptera: Vespertilionidae) in the
695 central Great Lakes region. *The American Midland Naturalist* **157**, 404–412.
- 696 52. Lundy, M, Montgomery, I, & Russ, J. (2010) Climate change-linked range expan-
697 sion of Nathusius’ pipistrelle bat, *Pipistrellus nathusii* (Keyserling & Blasius, 1839).
698 *Journal of Biogeography* **37**, 2232–2242.

- 699 53. McCracken, G. F, Bernard, R. F, Gamba-Rios, M, Wolfe, R, Krauel, J. J, Jones,
700 D. N, Russell, A. L, & Brown, V. A. (2018) Rapid range expansion of the Brazilian
701 free-tailed bat in the southeastern United States, 2008–2016. *Journal of Mammalogy*
702 **99**, 312–320.
- 703 54. Roberts, B. J, Catterall, C. P, Eby, P, & Kanowski, J. (2012) Latitudinal range shifts
704 in Australian flying-foxes: A re-evaluation. *Austral Ecology* **37**, 12–22.
- 705 55. Uhrin, M, Hüttmeir, U, Kipson, M, Estók, P, Sachanowicz, K, Bücs, S, Karapandža,
706 B, Paunović, M, Presetnik, P, Bashta, A.-T, et al. (2016) Status of Savi's pipistrelle
707 *Hypsugo savii* (Chiroptera) and range expansion in Central and south-eastern Eu-
708 rope: a review. *Mammal Review* **46**, 1–16.
- 709 56. Brook, C. E & Dobson, A. P. (2015) Bats as 'special' reservoirs for emerging zoonotic
710 pathogens. *Trends in Microbiology* **23**, 172–180.
- 711 57. Luis, A. D, Hayman, D. T, O'Shea, T. J, Cryan, P. M, Gilbert, A. T, Pulliam,
712 J. R, Mills, J. N, Timonin, M. E, Willis, C. K, Cunningham, A. A, et al. (2013) A
713 comparison of bats and rodents as reservoirs of zoonotic viruses: are bats special?
714 *Proceedings of the Royal Society B: Biological Sciences* **280**, 20122753.
- 715 58. Olival, K. J, Weekley, C. C, & Daszak, P. (2015) Are bats really 'special' as viral
716 reservoirs? what we know and need to know. *Bats and Viruses* pp. 281–294.
- 717 59. Wang, L.-F, Walker, P. J, & Poon, L. L. (2011) Mass extinctions, biodiversity and
718 mitochondrial function: are bats 'special' as reservoirs for emerging viruses? *Current*
719 *Opinion in Virology* **1**, 649–657.
- 720 60. Geoghegan, J. L, Senior, A. M, Di Giallonardo, F, & Holmes, E. C. (2016) Virological
721 factors that increase the transmissibility of emerging human viruses. *Proceedings of*
722 *the National Academy of Sciences* **113**, 4170–4175.
- 723 61. Walker, J. W, Han, B. A, Ott, I. M, & Drake, J. M. (2018) Transmissibility of
724 emerging viral zoonoses. *PloS One* **13**, e0206926.
- 725 62. Pigott, D. M, Millea, A. I, Earl, L, Morozoff, C, Han, B. A, Shearer, F. M, Weiss,
726 D. J, Brady, O. J, Kraemer, M. U, Moyes, C. L, et al. (2016) Updates to the zoonotic
727 niche map of Ebola virus disease in Africa. *Elife* **5**, e16412.
- 728 63. Bermejo, M, Rodríguez-Teijeiro, J. D, Illera, G, Barroso, A, Vilà, C, & Walsh, P. D.
729 (2006) Ebola outbreak killed 5000 gorillas. *Science* **314**, 1564–1564.

- 730 64. Hayman, D. T. (2019) African primates: Likely victims, not reservoirs, of
731 ebolaviruses. *The Journal of Infectious Diseases*.
- 732 65. Goldstein, T, Anthony, S. J, Gbakima, A, Bird, B. H, Bangura, J, Tremeau-Bravard,
733 A, Belaganahalli, M. N, Wells, H. L, Dhanota, J. K, Liang, E, et al. (2018) The
734 discovery of Bombali virus adds further support for bats as hosts of ebolaviruses.
735 *Nature Microbiology* **3**, 1084.
- 736 66. Williams, J. N. (2013) Humans and biodiversity: population and demographic trends
737 in the hotspots. *Population and Environment* **34**, 510–523.
- 738 67. Cheng, Y, Tjaden, N. B, Jaeschke, A, Lühken, R, Ziegler, U, Thomas, S. M, &
739 Beierkuhnlein, C. (2018) Evaluating the risk for Usutu virus circulation in Europe:
740 comparison of environmental niche models and epidemiological models. *International*
741 *Journal of Health Geographics* **17**, 35.
- 742 68. Roberts, K. E, Hadfield, J. D, Sharma, M. D, & Longdon, B. (2018) Changes in
743 temperature alter the potential outcomes of virus host shifts. *PLoS Pathogens* **14**,
744 e1007185.
- 745 69. Faust, C. L, Dobson, A. P, Gottdenker, N, Bloomfield, L. S, McCallum, H. I, Gille-
746 spie, T. R, Diuk-Wasser, M, & Plowright, R. K. (2017) Null expectations for disease
747 dynamics in shrinking habitat: dilution or amplification? *Philosophical Transactions*
748 *of the Royal Society B: Biological Sciences* **372**, 20160173.
- 749 70. Cunningham, A. (2018) *Infectious disease threats to amphibian conservation*. (Glas-
750 gow Natural History Society), Vol. 27.
- 751 71. VanWormer, E, Mazet, J, Hall, A, Gill, V, Boveng, P, London, J, Gelatt, T, Fadely,
752 B, Lander, M, Sterling, J, et al. (2019) Viral emergence in marine mammals in the
753 North Pacific may be linked to Arctic sea ice reduction. *Scientific Reports* **9**, 1–11.
- 754 72. Grubbs, F. E et al. (1950) Sample criteria for testing outlying observations. *The*
755 *Annals of Mathematical Statistics* **21**, 27–58.
- 756 73. Renner, I. W, Elith, J, Baddeley, A, Fithian, W, Hastie, T, Phillips, S. J, Popovic,
757 G, & Warton, D. I. (2015) Point process models for presence-only analysis. *Methods*
758 *in Ecology and Evolution* **6**, 366–379.
- 759 74. Friedman, J, Hastie, T, & Tibshirani, R. (2010) Regularization paths for generalized
760 linear models via coordinate descent. *Journal of Statistical Software* **33**, 1.

- 761 75. Phillips, S. J, Anderson, R. P, & Schapire, R. E. (2006) Maximum entropy modeling
762 of species geographic distributions. *Ecological Modelling* **190**, 231–259.
- 763 76. Fick, S. E & Hijmans, R. J. (2017) WorldClim 2: new 1-km spatial resolution climate
764 surfaces for global land areas. *International Journal of Climatology* **37**, 4302–4315.
- 765 77. Hastie, T, Tibshirani, R, & Friedman, J. H. (2009) *The elements of statistical learn-*
766 *ing: data mining, inference, and prediction*. (Springer-Verlag, New York), 2nd edi-
767 tion.
- 768 78. Drake, J. M. (2015) Range bagging: a new method for ecological niche modelling
769 from presence-only data. *Journal of The Royal Society Interface* **12**, 20150086–9.
- 770 79. Drake, J. M & Richards, R. L. (2018) Estimating environmental suitability. *Ecosphere*
771 **9**, e02373.
- 772 80. Hurtt, G. C, Chini, L. P, Froking, S, Betts, R, Feddema, J, Fischer, G, Fisk, J,
773 Hibbard, K, Houghton, R, Janetos, A, et al. (2011) Harmonization of land-use
774 scenarios for the period 1500–2100: 600 years of global gridded annual land-use
775 transitions, wood harvest, and resulting secondary lands. *Climatic Change* **109**, 117.
- 776 81. Hurtt, G, Chini, L, Sahajpal, R, Froking, S, Bodirsky, B, Calvin, K, Doelman, J,
777 Fisk, J, Fujimori, S, Goldewijk, K, et al. (2018) *LUH2: Harmonization of global*
778 *land-use scenarios for the period 850-2100*.
- 779 82. Fritz, S. A, Bininda-Emonds, O. R, & Purvis, A. (2009) Geographical variation in
780 predictors of mammalian extinction risk: big is bad, but only in the tropics. *Ecology*
781 *Letters* **12**, 538–549.
- 782 83. Jones, K. E, Bielby, J, Cardillo, M, Fritz, S. A, O'Dell, J, Orme, C. D. L, Safi,
783 K, Sechrest, W, Boakes, E. H, Carbone, C, et al. (2009) PanTHERIA: a species-
784 level database of life history, ecology, and geography of extant and recently extinct
785 mammals: Ecological Archives E090-184. *Ecology* **90**, 2648–2648.
- 786 84. Jetz, W, Wilcove, D. S, & Dobson, A. P. (2007) Projected impacts of climate and
787 land-use change on the global diversity of birds. *PLoS Biology* **5**, e157.
- 788 85. Pecl, G. T, Araújo, M. B, Bell, J. D, Blanchard, J, Bonebrake, T. C, Chen, I.-C,
789 Clark, T. D, Colwell, R. K, Danielsen, F, Evengård, B, et al. (2017) Biodiversity
790 redistribution under climate change: Impacts on ecosystems and human well-being.
791 *Science* **355**, eaai9214.

- 792 86. Powers, R. P & Jetz, W. (2019) Global habitat loss and extinction risk of terrestrial
793 vertebrates under future land-use-change scenarios. *Nature Climate Change* **9**, 323.
- 794 87. Hurtt, G. C, Chini, L. P, Frohking, S, Betts, R. A, Feddema, J, Fischer, G, Fisk, J. P,
795 Hibbard, K, Houghton, R. A, Janetos, A, Jones, C. D, Kindermann, G, Kinoshita,
796 T, Klein Goldewijk, K, Riahi, K, Shevliakova, E, Smith, S, Stehfest, E, Thomson,
797 A, Thornton, P, van Vuuren, D. P, & Wang, Y. P. (2011) Harmonization of land-
798 use scenarios for the period 1500–2100: 600 years of global gridded annual land-use
799 transitions, wood harvest, and resulting secondary lands. *Climatic Change* **109**,
800 117–161.
- 801 88. van Vuuren, D. P, Riahi, K, Calvin, K, Dellink, R, Emmerling, J, Fujimori, S, Kc, S,
802 Kriegler, E, & O'Neill, B. (2017) The Shared Socio-economic Pathways: Trajectories
803 for human development and global environmental change. *Global Environmental*
804 *Change* **42**, 148–152.
- 805 89. Popp, A, Calvin, K, Fujimori, S, Havlik, P, Humpenöder, F, Stehfest, E, Bodirsky,
806 B. L, Dietrich, J. P, Doelmann, J. C, Gusti, M, et al. (2017) Land-use futures in the
807 shared socio-economic pathways. *Global Environmental Change* **42**, 331–345.
- 808 90. Riahi, K, van Vuuren, D. P, Kriegler, E, Edmonds, J, O'Neill, B. C, Fujimori, S,
809 Bauer, N, Calvin, K, Dellink, R, Fricko, O, Lutz, W, Popp, A, Cuaresma, J. C, Kc,
810 S, Leimbach, M, Jiang, L, Kram, T, Rao, S, Emmerling, J, Ebi, K, Hasegawa, T,
811 Havlik, P, Humpenöder, F, Da Silva, L. A, Smith, S, Stehfest, E, Bosetti, V, Eom,
812 J, Gernaat, D, Masui, T, Rogelj, J, Streffer, J, Drouet, L, Krey, V, Luderer, G,
813 Harmsen, M, Takahashi, K, Baumstark, L, Doelman, J. C, Kainuma, M, Klimont, Z,
814 Marangoni, G, Lotze-Campen, H, Obersteiner, M, Tabeau, A, & Tavoni, M. (2017)
815 The Shared Socioeconomic Pathways and their energy, land use, and greenhouse gas
816 emissions implications: An overview. *Global Environmental Change* **42**, 153–168.
- 817 91. Wilman, H, Belmaker, J, Simpson, J, de la Rosa, C, Rivadeneira, M. M, & Jetz,
818 W. (2014) Eltontraits 1.0: Species-level foraging attributes of the world's birds and
819 mammals: Ecological archives e095-178. *Ecology* **95**, 2027–2027.
- 820 92. Pacifici, M, Santini, L, Di Marco, M, Baisero, D, Francucci, L, Marasini, G. G,
821 Visconti, P, & Rondinini, C. (2013) Generation length for mammals. *Nature Con-*
822 *servation* **5**, 89.
- 823 93. Bateman, B. L, Murphy, H. T, Reside, A. E, Mokany, K, & VanDerWal, J. (2013)
824 Appropriateness of full-, partial-and no-dispersal scenarios in climate change impact
825 modelling. *Diversity and Distributions* **19**, 1224–1234.

- 826 94. Araújo, M. B, Rozenfeld, A, Rahbek, C, & Marquet, P. A. (2011) Using species co-
827 occurrence networks to assess the impacts of climate change. *Ecography* **34**, 897–908.
- 828 95. Olival, K. J, Islam, A, Yu, M, Anthony, S. J, Epstein, J. H, Khan, S. A, Khan, S. U,
829 Cramer, G, Wang, L.-F, Lipkin, W. I, et al. (2013) Ebola virus antibodies in fruit
830 bats, Bangladesh. *Emerging Infectious Diseases* **19**, 270.
- 831 96. Yang, X.-L, Zhang, Y.-Z, Jiang, R.-D, Guo, H, Zhang, W, Li, B, Wang, N, Wang,
832 L, Waruhiu, C, Zhou, J.-H, et al. (2017) Genetically diverse filoviruses in *Rousettus*
833 and *Eonycteris* spp. bats, China, 2009 and 2015. *Emerging Infectious Diseases* **23**,
834 482.
- 835 97. Gao, J. (2017) Downscaling global spatial population projections from 1/8-degree
836 to 1-km grid cells, (NCAR technical note NCAR/TN-537+ STR. Boulder, Colorado:
837 National Center for Atmospheric Research), Technical report.