Mediterranean Diet Reduces Monocyte Inflammatory Gene Expression and Influences

Social Behavior in Nonhuman Primates

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Abstract: Western diet consumption is associated with inflammation, cardiometabolic disease,

and mortality in humans, while Mediterranean diet consumption confers protective effects. One

likely pathway for this association is through environmentally induced changes in monocyte

function, yet the underlying mechanisms remain elusive. We conducted the first randomized,

long-term diet manipulation in a non-human primate model to determine whether Western- or

Mediterranean-like diets alter monocyte polarization and health. Monocyte gene expression

profiles differed markedly between the two diet groups, with significant differences in over 40%

of expressed genes. The Western diet induced a more proinflammatory monocyte phenotype

overall and upregulated specific monocyte polarization genes. Diet also disrupted the

coexpression of numerous gene pairs, including small RNAs and transcription factors associated

with metabolism and adiposity in humans. Diet altered affiliative and anxiety-associated

behaviors and mediation analysis showed that the diet-altered behaviors contributed significantly

(~50% of the effect of diet on gene expression) to 25% of the differentially expressed genes,

suggesting that diet effects on central mechanisms also modulate monocyte gene expression.

Together, these results identify both behavioral and molecular mechanisms underlying the health

benefits of a Mediterranean diet regimen.

Significance Statement: Some of our largest public health burdens are driven by dietary

changes associated with industrialization, but we still know very little about the molecular

mechanisms underlying this link. Characteristic "Western diets" have been associated with

increased risk for diseases related to chronic inflammation, while Mediterranean diets have anti-

inflammatory benefits. Here, we identify causal effects of diet on inflammatory gene expression

where consumption of the Mediterranean diet reduced inflammatory gene expression in

monocytes. Additionally, our diet manipulation induced behavioral changes associated with

anxiety and social integration, where Mediterranean-fed animals exhibited more positive

affiliative behaviors and reduced anxiety. These behaviors were associated with 25% of the diet-

affected genes, suggesting an important behavioral route through which diet can impact immune

function.

[Main Text]

Introduction

Modern human diets profoundly impact our health and survival, and vary across geography, cultures, and socioeconomic strata. In general, the Western diet derives most of its protein and fat from animal sources, and is high in simple sugars and saturated and n-6 fatty acids. These constituents can arouse the sympathetic nervous system, increase oxidative stress, and elevate levels of inflammatory markers[1–6], and are thus associated with increased risk for metabolic syndrome[7], type II diabetes[8], cardiovascular disease[7,9], autoimmune disorders[10], depression[11], and increased mortality[12]. By contrast, Mediterranean diets are richer in protein and fat from vegetable sources, raw fruits and vegetables, and are higher in monounsaturated and n-3 fatty acids. These latter components have been associated with an anti-inflammatory phenotype[13], reduced incidence of chronic disease, and increased longevity[14–17]. Despite these associations, the casual nature of these links and mechanisms through which these diets induce their effects remain largely unknown.

To date, attempts to understand how Western versus Mediterranean diets affect health through changes in immune phenotypes have relied on (i) correlational analyses of self-reported diet in humans, (ii) limited and short-term dietary interventions in humans, or (iii) experimental manipulations of single nutrients in animal models[18–22]. Approaches (i) and (ii) are limited in their ability to address causality[23,24], and approach (iii) cannot address the potentially important synergistic effects of multiple nutrients. Further, few studies have probed the molecular mechanisms through which diet can alter immune function–data that are critical for understanding the immunological consequences of diet and identifying targets of future therapies

and interventions.

Circulating monocytes are likely to play a key role in modulating the effects of diet and other

factors on health[3–6,25]. Monocytes are important mediators of inflammation, sensitive to local and

systemic factors such as diet and stress, and may provide a key nexus for understanding stress

effects as well as novel targets for therapies. Monocytes and monocyte-derived macrophages are

innate immune cells that vary phenotypically along a spectrum which ranges broadly from

proinflammatory (M1-like) to regulatory/reparative (M2-like). An appropriate balance of

monocyte phenotypes is essential for a healthy immune system. Classically-activated "M1"

monocytes respond to proinflammatory cytokines such as tumor necrosis factor (TNF)-α and

interferon (IFN)-γ by becoming macrophages which propagate the inflammatory response to

infection[26]. In contrast, M2 activated monocytes mobilize tissue repair processes and release

anti-inflammatory cytokines in response to interleukin (IL)-4, IL-13, and transforming growth

factor (TGF)-β[26]. Diet may alter disease propensity by reprogramming the balance between these

proinflammatory and anti-inflammatory monocyte subsets, but this hypothesis remains to be

tested[25].

In addition to altering the regulation of immune cells directly, diet may affect inflammatory

phenotypes indirectly by altering social behaviors, which are known to shape gene expression

programs in immune cells. In particular, multiple sources of social adversity, such as low social

status and poor social integration, have been shown to increase the expression of inflammatory

genes in primary white blood cells in humans and other animals[27–32]. Given that some food

contituents can directly alter social behaviors themselves[33–37], it is therefore possible that diet

effects on immune cell regulation may, to some degree, be mediated by changes in social

environmental conditions. However, because no detailed studies of diet, social behavior, and

immune cell phenotypes have been conducted, it remains unclear how these factors are linked

and how they ultimately scale up to affect health.

To address these gaps, we conducted a whole-diet manipulation to directly compare the effects

of Mediterranean and Western diets on behavior, monocyte gene expression, and physiological

outcomes related to metabolic health in nonhuman primates. By implementing a randomized

preclinical trial design, we were able to identify causal effects of realistic complex diet patterns.

After 15 months of dietary manipulation, cardio-metabolic phenotypes were significantly worse

and proinflammatory gene expression was significantly higher in animals fed a Western diet

relative to a Mediterranean diet. Diet also affected monocyte polarization, altered gene co-

expression patterns, and influenced behavior. Western-fed monkeys became more socially

isolated and exhibited more anxiety-associated behaviors, and these behavioral changes mediated

some of the effects of diet on monocyte gene expression. These behavioral effects imply that the

diet altered monocyte gene expression in part via the central nervous system. Together, these

results suggest both direct and behaviorally-mediated effects of diet on monocyte polarization

may contribute to chronic inflammatory diseases, and identify potential mechanisms by which

Mediterranean-like diets may lead to health benefits.

Results

Diet induced alterations in health indices

Adult female cynomolgus macaques were fed either a Western-like (hereafter, "Western") or a

Mediterranean-like (hereafter, "Mediterranean") diet for 15 months (the equivalent of ~4 years in

a human lifespan; Fig. 1A). The experimental diets were nutritionally matched with respect to

caloric content of macronutrients and formulated to model human diet patterns, as previously

described[38]. Protein and fat were derived primarily from animal sources in the Western diet and

plant sources in the Mediterranean diet. Consequently, the two diets differed in their composition

of key micronutrients, including fatty acids, polyunsaturated fatty acid ratios, fiber, and sodium

(Fig. 1B; see methods and Table S1 for a detailed comparison). As previously reported, Western

diet significantly increased body weight, caloric intake, body fat, insulin resistance, and

hepatosteatosis relative to the Mediterranean diet[38] (Fig. 1C).

Figure 1. Experimental design and diet effects on body weight. **A)** Monkeys were housed in groups of 3-4

animals (*n* = 35 monkeys) and fed standard monkey chow diet for 8 months before being fed experimental diets.

Behavioral data were collected during the last 6 weeks of the baseline phase and the during months 1-14 of the

experimental phase. Body weight measurements reported are from 5 months prior to, and 14 months after the start

of the experimental phase. Monocytes were isolated from blood collected 15 months after the start of the

experimental phase. **B)** Experimental diets were isocaloric with respect to macronutrients, but differed in food

sources and relative amounts of micronutrients. Orange bars indicate nutrients with higher concentration in the

Western diet formulation, while blue bars indicate higher levels of a given nutrient in the Mediterranean diet. See

Table S1 for diet compositions. **C)** Percent change in body weight from baseline after 14 months on the diet ($t_{(23.0)}$

$= 3.02, p = 0.0023$).

Diet induced major shifts in monocyte gene expression

To test how diet affected the phenotypes of circulating monocytes, we used RNA sequencing to

measure genome-wide gene expression of purified CD14+ monocytes after 15 months on the

experimental diets. Diet had a strong effect on monocyte gene expression: the first principal

component of gene expression, which explained 59.2% variance, was significantly associated

with diet ($t_{(25.1)} = 4.41, p = 1.7$ x 10^{-4}; Fig. 2A), and 40% of the 12,240 expressed genes (Table

S2A) were significantly differentially expressed between the two diets ($n = 4,900$ genes, FDR <

0.05; Table S2B). The number of diet-responsive genes was roughly balanced between those that

were more highly expressed in monkeys fed the Mediterranean diet ($n = 2,664$; hereafter

"Mediterranean genes") and those that were more highly expressed in monkeys fed the Western

diet ($n = 2,236$; hereafter "Western genes"). While balanced in direction, the distributions of

effect sizes in these two sets of genes differed significantly (one sided Kolmogorov-Smirnov test,

$D = 0.33, p = 5.2$ x 10^{-112}) and the effect size of diet on Western genes was, on average, 1.6-fold

larger than on Mediterranean genes (Mann-Whitney $U = 4.1$ x $10^6, p = 6.1$ x 10^{-117}; Fig. 2B).

Thus, the strongest effects are seen in genes that are either activated by a Western diet or

suppressed by a Mediterranean diet.

Figure 2. Diet effects on monocyte gene expression. A) Diet significantly predicts the first principal component of gene expression (59.3% variance explained, $t_{(25.0)} = 4.41$, $p = 1.72 \times 10^{-4}$). **B)** The average effect size of diet on Western genes was 1.6-fold larger than the effect size of diet on Mediterranean genes (Mann-Whitney $U = 4.1 \times 10^6$, $p = 6.1 \times 10^{-117}$). **C)** $\log_2$ fold enrichment of proinflammatory (top) and regulatory (bottom) genes in Western genes (orange) and Mediterranean genes (blue). Western genes contained more M1 genes than expected by chance, indicating that the Western diet induced a shift towards a proinflammatory monocyte phenotype. Western genes were enriched for proinflammatory (M1-like) genes (fold-enrichment = 1.27, *95% CI* = 1.09, 1.46), while Mediterranean genes were depleted of these same M1-like genes (fold-enrichment = 0.74, *95% CI* = 0.61, 0.88). Regulatory (M2-like) genes were also under-represented in Mediterranean genes (fold-enrichment = 0.67, *95% CI* = 0.40, 0.97), but not in Western genes (fold-enrichment = 0.95, *95% CI* = 0.60, 1.35).

Monocytes in animals fed the Western diet had higher expression of a number of well-known

inflammatory-related genes, including interleukin-6 ($\beta_{\text{diet}} = 1.66$, FDR = 8.9×10^{-3}; Fig. 2B),

interleukin-1α ($\beta_{\text{diet}} = 1.22$, FDR = 0.03; Table S2B), and two subunits of the NF-κB protein

(*NFKB1* β_{diet} = 0.30, FDR = 0.017; *NFKB2* β_{diet} = 0.42, FDR = 0.012; Table S2B). Western genes

were significantly more likely to be involved in replication and metabolic cellular processes,

including response to growth factor (GO:0070848, weighted Fisher's Exact Test (FET) p =

4.6×10^{-3}) and response to insulin (GO:0032868, weighted FET p = 4.0×10^{-4}; Table S3A),

suggesting that the Western diet also reprogrammed oxidative metabolic aspects of monocyte

gene regulation. Conversely, Mediterranean diet monocyte expression patterns indicated

enhanced oxidation-reduction processes (GO:0055114, weighted FET p = 6.0×10^{-3}; Table S3B),

a critical function in muting proinflammatory monocytes.

We next conducted a more targeted analysis of monocyte polarization by focusing on genes that

were previously reported to be differentially expressed between induced proinflammatory (M1)

and regulatory (M2) monocyte polarization[39] (see Table S2C for polarization categories).

Western genes were enriched in M1-associated genes (*n* = 162 genes, fold-enrichment = 1.27,

95% CI = 1.09 - 1.46; Fig. 2C), but not M2-associated genes (*n* = 24 genes, fold-enrichment =

0.95, *95% CI* = 0.60 - 1.35). Conversely, both M1-associated genes (*n* = 112 genes, fold-

enrichment = 0.74, *95% CI* = 0.61 - 0.88) and M2-associated genes (*n* = 20 genes, fold-

enrichment = 0.67, *95% CI* = 0.40 - 0.97) were underrepresented among Mediterranean genes.

Together, these observations indicate that a Western diet induces a more proinflammatory (M1-

like) phenotype.

Next, to identify putative upstream gene regulatory mechanisms, we examined whether diet-

induced changes in gene expression were associated with *cis*-regulatory transcription factor

binding sites. We identified 34 distinct transcription factor-binding motifs enriched within 2

kilobases of the transcription start sites of Mediterranean genes and one that was enriched near

the transcription start sites of Western genes (FDR < 0.05; Fig. 3, Table S4). Diet significantly

altered expression of the genes encoding for seven of these 35 transcription factors, including

IRF3, IRF8, MEF2C, and SP1, which drive monocyte fate and polarization in response to

extracellular signals[40–44]. Thus, some of the diet-associated changes in monocyte gene regulation

may be mediated by changes in the expression and *cis*-regulatory binding of key transcription

factors.

Figure 3. Transcription factor (TF) binding motifs correlated with diet effects on gene expression. The log-odds ratio of TF binding motif enrichment in Western genes (orange) or Mediterranean genes (blue) are depicted on the x-axis. The y-axis shows the effect size of diet on the expression of the gene that encodes for the TF. Only TFs with binding motifs significantly enriched in either gene set and that were detectably expressed in our samples are shown, with those significantly effected by diet outlined and labeled.

Diet alters gene co-expression patterns

Next we asked whether diet altered the magnitude or direction of pairwise gene expression

correlations among the most strongly diet-affected genes, as such effects could reveal key gene

regulatory networks that are altered by diet, that may themselves be regulated by key upstream

targets[45,46]. Drawing on a newly developed approach, "correlation by individual level product"

(CILP)[47], we identified 445 gene pairs that exhibited significant changes (FDR < 20%) in their

correlation between the Mediterranean- and Western-fed monkeys (Table S5A; Fig. 4A). The

majority (97%) of these gene pairs exhibited positive associations in one diet and negative

associations in the other, suggesting that diet can completely reverse the co-expression

relationship between two genes (Figure 4A). We further identified 16 "hub" genes that exhibited

differential correlations with partner genes more so than expected by chance (Fig. 4B, Table

S5B). These hub genes were enriched for genes encoding transcription factors (OR = 7.40, FET

$p = 7.0 \times 10^{-3}$), including SOX4 (essential for normal insulin secretion and glucose tolerance)

and NR4A2 (involved in lipid, carbohydrate, and energy metabolism[48,49]), suggesting immune

and metabolic reprogramming by the diet manipulation. Interestingly, the hub gene involved in

the greatest number of differentially-correlated gene pairs was *RF00283, aka SCARNA18,* a non-

coding RNA that has been associated with BMI, HDL cholesterol, and aging in human genome-

wide association studies[50–53] (Fig. 4B-D), identifying it as a key regulatory RNA that is altered

by diet and has a cascading effect on other genes and pathways.

Figure 4. Diet affects monocyte gene co-expression. A) The Pearson correlation between each pair of genes within each of the experimental diets. Gene pairs that are significantly differently correlated between diets are highlighted in blue (*n* = 445 significant pairs, FDR < 20%). **B)** Of the genes involved in significant pairs, some were paired with more genes than expected by chance (*n* = 16 "hub" genes; dotted black line is the maximum number of significant pairs expected by chance). The strongest hub gene was the non-coding RNA *RF00283*. **C)** Residual normalized expression of *RF00283* is significantly greater in Western- than Mediterranean-fed monkeys (β_{diet} = 0.507, FDR = 2.3 x 10^{-6}). **D)** Example of a differential correlation involving *RF00283*. Residual normalized expression of *RF00283* is plotted against expression of *KLF11*, a differentially-expressed transcription factor that regulates insulin and has been associated with type II diabetes in humans[54]. The two genes were more highly expressed in Western monocytes, and were positively correlated with one another in Western-fed monkeys (*r* = 0.61, *p* < 0.005) and negatively correlated in Mediterranean-fed monkeys (*r* = -0.63, *p* < 0.01).

Diet altered social behavior

There were no differences in behavior during the baseline phase (all $p > 0.1$; Fig. S1A, B). While

on the experimental diets, monkeys fed the Mediterranean diet spent significantly more time in

body contact (Mann-Whitney $U = 280$, Holm-Bonferroni adjusted p (p_{HB}) = 1.2 x 10^{-5}) and

resting ($U = 267$, $p_{HB} = 1.6$ x 10^{-3}), while those fed the Western diet spent significantly more

time alone ($U = 48$, $p_{HB} = 4.7$ x 10^{-3}; Fig. 5A). All other measured behaviors did not pass our

stringent p-value threshold after multiple hypothesis testing correction (Fig. S1C,D), although

two additional behaviors differed at an uncorrected p-value < 0.05 (percent of time attentive and

rate grooming self). Therefore, to increase our ability to identify diet-affected suites of behaviors,

we leveraged the fact that many behaviors co-occurred (Fig. S2) by conducting a principal

component analysis[55,56]. Behaviors associated with dominance interactions—including

aggression, submission, and being groomed—all loaded heavily onto the first principal

component, which explained 32.2% of the overall variance in behavior and did not differ

between diets (Welch-Satterthwaite $t_{(30.3)} = 0.323$, $p = 0.75$; Fig. S3, Table S6A). The first

principal component was significantly correlated with dominance rank (Fig. S4, Note S1).

The second principal component, which explained 18.8% of the variance in behavior, differed

significantly between the two diets ($t_{(26.8)} = -4.02$, $p = 4.2$ x 10^{-4}; Fig. 5B), and thus represented a

composite of diet-altered behaviors (hereafter, DAB). No other principal component was

significantly correlated with diet and thus PC2 captures the primary behavioral component

causally affected by diet (Fig. 5C, Table S6B). PC2 captured a number of anxiety and social

behaviors (Fig. S5, Table S6A). Specifically, body contact is indicative of social integration and

was positively correlated with PC2 loading (hereafter, DAB score), which was higher in

Mediterranean fed animals. Conversely, behaviors related to social isolation and anxiety[57–62]

(e.g., percent of time alone, rate of grooming self, rate of scratching) were associated with lower

DAB scores, and hence more prevalent in animals fed the Western diet (Fig. 5C). Thus, PC2

captured a measure of social integration associated with consuming a Mediterranean-like diet,

and social isolation and anxiety associated with consuming a Western-like diet.

Figure 5. Diet alters behavioral phenotype. A) Three behaviors were significantly different between the two

diet groups. Monkeys fed the Mediterranean diet spent more time in body contact (p_{HB} = 1.2 x 10^{-5}) and resting

(p_{HB} = 1.6 x 10^{-3}) than Western-fed monkeys. Monkeys eating the Western diet spent more time alone than

Mediterranean-fed monkeys (p_{HB} = 4.7 x 10^{-3}). **B)** Composite measures of diet-altered behavior (DAB scores)

were significantly higher in Mediterranean diet compared to Western diet animals ($t_{(32.0)}$= 5.30, p = 8.2 x 10^{-6}). **C)**

Principal component 2 (PC2) explained 18.8% of the variance in behavior and was the only PC significantly

correlated with diet (see Table S6B for correlation between diet and other PCs). **D)** Six of the 21 behaviors

observed are significantly correlated with DAB score (Benjamini-Hochberg adjusted p < 0.05). Here, significant

correlations with DAB score in which behaviors are more frequent in Mediterranean diet or Western diet

monkeys are indicated with blue or orange points, respectively.

Diet-altered behaviors mediate expression of 25% of differentially expressed monocyte genes

Given the strong effects of diet on both behavior and monocyte gene expression, we tested if the

effect of diet on monocyte gene expression was mediated by the diet-induced changes in

behavior. Of the 4,900 diet-affected genes, 29% were also significantly associated with DAB

score in a univariate model ($n = 1,418$, FDR < 0.05). Of these, DAB score significantly mediated

the effect of diet on the expression of 1220 genes (25% of all diet-associated genes, $p < 0.05$;

Fig. 6A). DAB score mediation accounted for significantly more of the effect of diet in DAB-

mediated Western genes ($\mu = 51.1$ %, $\delta = 12.4\%$), than DAB-mediated Mediterranean genes ($\mu =$

44.2 %, $\delta = 10.0\%$; Mann-Whitney $U = 2.4$ x 10^5, $p = 7.5$ x 10^{-23}; Fig. 6B). These DAB-

mediated genes were also significantly more likely to be Western genes than Mediterranean

genes ($n = 741$ Western genes, 61%, two-sided binomial test $p = 6.3$ x 10^{-14}), and were enriched

in regulation of inflammatory response (GO:0050727, weighted FET $p = 2.9$ x 10^{-3}; Table S7A-

C). Together, this shows that the effect of diet on monocyte gene regulation may partially be due

to diet-induced changes in key social behaviors.

In support of this mediation effect, we compared expression of a well-studied set of social

adversity-responsive genes known as the "conserved transcriptional response to adversity"

(CTRA)[28] in the Western- and Mediterranean-fed animals in our study. Animals fed a Western

diet exhibited significantly higher expression of pro-inflammatory genes included in the CTRA

(Mann-Whitney $U = 222$, $p = 0.016$) and lower expression of antiviral- and antibody-related

CTRA genes (Mann-Whitney $U = 82$, $p = 0.023$; Table S2C, Fig. S6).

We also tested the hypothesis that diet could alter behavior through its changes on peripheral

immune cell gene expression. We tested this in the 28% of genes for which monocyte gene

expression significant predicted DAB in a univariate model ($n = 1,353$, FDR < 0.05), and found

that gene expression significantly mediated the effect of diet on DAB score in 940 genes (19% of

all diet-associated genes, $p < 0.05$; Fig. 6A). Almost all of these genes (99.5%; 936/940) were

significantly mediated by diet-induced changes in DAB. As with DAB score mediating gene

expression, the genes that mediated the effect of diet on DAB score were more likely to be

Western genes than Mediterranean genes ($n = 558$ Western genes, 59%, two-sided binomial test

$p = 1.0 \times 10^{-8}$). Unlike DAB score mediating the effect of diet on gene expression, the portion of

the effect of diet that was accounted for by gene expression did not vary between Western ($\mu =$

27.5 %, $\delta = 5.4\%$) and Mediterranean genes ($\mu = 27.3$ %, $\delta = 4.6\%$; Mann-Whitney $U = 1.1 \times$

10^5, $p = 0.75$; Fig. 6B).

Figure 6. Behavior partially mediates the effect of diet on gene expression for 25% of diet-associated genes.

A) Diet-altered behavior (DAB) mediated the effect of diet on gene expression for 25% (n = 1220) of genes for which diet had an effect (DEG). For 19% of differentially expressed genes (DEG), gene expression mediated the effect of diet on DAB score. **B)** DAB score mediated 24-97% of the effect of diet on gene expression in 1220 genes (n = 741 Western genes, orange; n = 479 Mediterranean genes, blue). DAB score mediated a greater number of Western genes than Mediterranean genes (p = 6.3 x 10^{-14}) and accounted for a greater portion of the effect size of diet (p = 7.5 x 10^{-23}) in Western genes. **C)** In gene-by-gene models of DAB score as a function of diet + gene expression, gene expression mediated 15-51% of the effect of diet on DAB in 940 genes (n = 558 Western genes; n = 382 Mediterranean genes). Gene expression mediated a greater number of Western genes than Mediterranean genes (p = 1.0 x 10^{-8}), although expression of these genes did not account for more of the effect of diet on DAB score than Mediterranean genes (Mann-Whitney U = 1.1 x 10^5, p = 0.75).

Western diet induces mosaic response

Western diet induced substantial variation in multiple phenotypes, including body weight, gene

expression, and behavior; consistent with previous studies demonstrating that some individuals

may be more resistant (or susceptible) to the effects of a Western diet[63], presumably due to

genetic variation or past environmental exposures. However, we were unable to identify any

consistencies in individual responsiveness across the phenotypes (Fig. S7). For instance,

monkeys that exhibited a strong gene regulatory response to the Western diet did not exhibit a

large increase in body weight or a strong negative DAB score (all $p > 0.2$). Furthermore, change

in body weight did not significantly predict the expression of any genes at an FDR < 20%.

Western diet fed individuals thus exhibited a mosaic response to diet across multiple phenotypes,

presumably involving interactions between diet, environment, and the genome.

Discussion

This study shows, for the first time, that a whole-diet manipulation exerted profound effects on

monocyte function and social behavior in a primate. Forty percent of expressed genes were

differentially expressed between monkeys fed Western or Mediterranean diets, indicating that

diet dramatically altered monocyte programming. Relative to Western group monocytes,

Mediterranean group monocytes exhibited reduced proinflammatory gene expression and

regulatory gene expression. Our findings recapitulate and extend previous studies, such as a

randomized human cross-over trial that demonstrated that peripheral blood monocytes from

elderly individuals consuming a Mediterranean like diet enriched in olive oil had reduced

proinflammatory gene expression relative to diets more enriched in saturated fat (butter)[64].

Beyond mean differences in gene expression levels, we also identified differences in gene co-

expression and enrichment of transcription factor binding motifs, suggesting that diet exerts a

strong effect on gene regulatory networks.

We identified enrichment of binding motifs for numerous transcription factors that appear to be

involved in diet-regulated gene expression. Of note, members of the E26 transformation-specific

(ETS), specificity protein (Sp)/Krüppel-like family (KLF), myocyte-specific enhancer factor

(MEF), and interferon-regulatory factor (IRF) families of transcription factors, which have all

been linked to myeloid differentiation[40–43], were overrepresented in regulatory regions of genes

with higher expression in the Mediterranean diet group ("Mediterranean genes). IRF-1 and IRF-8

are linked to M1 monocyte polarization, while IRF-3 is associated with M2 polarization, and all

three transcription factors had binding motifs enriched in Mediterranean genes. The sole

transcription factor with binding sites enriched in Western diet-associated genes, ATF2, is a key

mediator of inflammatory pathways and diseases, including response to bacterial endotoxin,

atherosclerosis, and obesity[65–67]. Interestingly, Western genes were enriched for activation of the

MAPKK pathway, which lies upstream of ATF2[68], supporting its putative role in monocyte gene

regulation. Transcription factors were also overrepresented in the pairs of differentially co-

expressed genes, indicating that diet may be altering the networks through which inflammatory

genes are regulated. Broadly, this suggests that the two experimental diets differentially affect

transcriptional networks involved in monocyte differentiation and polarization. It is also worth

noting that the M1/M2 paradigm of monocyte polarization is a simplification of the more

complex heterogeneity of monocytes.[69,70] For example, there are at least 3 classes of monocytes

in the circulation, classical, intermediate, and non-classical, which individually have different

phenotypes. We did not assess proportions of these or try to isolate individual monocyte subsets

in the current study, thus the patterns of gene expression observed could represent altered

proportions of these subsets and well as shifts in monocyte polarization within subsets[71,72].

Diet induced changes in behavior, as monkeys consuming the Western diet exhibited more

behaviors related to anxiety and social isolation, a phenotype remarkably similar to that observed

in juvenile Japanese macaques born to mothers consuming a high-fat Western diet[73]. In that

study, offspring behavior was associated with maternal levels of macrophage-derived chemokine

(MDC), which showed higher expression in Western-diet fed animals in our study (β_{diet} = 0.243,

FDR = 0.059). Our findings suggest that a Western diet may also exert similar behavioral effects

in adulthood.

We observed that for a subset (25%) of genes, the diet-altered behavior (DAB) score mediated

the effect of diet on monocyte gene expression. This observation suggest involvement of

mechanistic pathways in which diet first impacts the brain, which in turn impacts monocyte

function. Monocytes have been shown to be responsive to social isolation[29] and anxiety[28]. Social

isolation and anxiety, produced by Western diet consumption, may be accompanied by increased

sympathetic outflow and increased hypothalmic-pituitary adrenal production of cortisol, both of

which modulate monocyte intracellular processes governing inflammatory molecule

production[74–76]. Supporting the involvement of these systems, we previously reported that the

Western diet group had increased sympathetic activity, and increased cortisol concentrations[77].

Therefore, it is possible that a Western diet contributes to inflammation by producing a more

socially isolated or anxious animal with increased sympathetic and hypothalamic pituitary

adrenal activity, which in turn alters monocyte function. Higher expression of genes in the

conserved transcriptional response to adversity support this pathway[28,29].

There are numerous pathways through which diet may affect behavior. Diet may induce changes

in the central nervous system by altering gut microbiota which alters vagal input to the brain[78].

We previously showed in these NHPs that diet had a strong effect on the gut microbiome[79], and

that compared to the Mediterranean group, Western diet NHPs had lower parasympathetic

(vagal) activity at the time the monocyte transcriptome was assessed[77]. Taken together these

observations suggest that diet-induced changes in vagal tone in the gut-brain axis may be one

pathway through which diet impacted brain function, potentially affecting behavior.

We also observed that for some genes (19%), diet-induced changes in monocyte gene expression

significantly mediated the effect of diet on behavior (DAB). This observation suggests

underlying mechanisms which first impact peripheral monocyte function, which in turn impacts

brain function. Western diet may disrupt the blood-brain barrier, increasing infiltration of

Western-diet induced cytokines, chemokines, and myeloid cells from the periphery[80,81]. Once in

the brain these molecules can alter BDNF production, neurotransmitter systems, and

hypothalamic-pituitary-adrenal function[80]. Western diet induced inflammatory molecules also

may effect the brain through direct effects on the afferent vagus nerve[82], activation of glial

cells[83], and alter neuronal membrane lipid composition affecting neurotransmission[84], whereas a

Mediterranean diet may have direct anti-inflammatory actions by increasing n-3 fatty acids in the

brain[85]. These results support both mediation pathways, suggesting that multiple mechanistic

pathways contributed to these observations.

The behavioral analysis also showed that the first principal component described dominance-

related behaviors. While the dominance component accounted for the largest proportion of the

variance in behavior, it was notably unaffected by the diet manipulation. From a socio-biological

perspective this suggests that dominance-related behavior is resistant to perturbation, which is

consistent with known stability of dominance hierarchies in female cynomolgus monkeys[33]. The

second principal component that captured affiliation, anxiety, and social isolation was

significantly affected by the two diets, suggesting that these behaviors are susceptible to dietary

interventions.

In summary, we found that diet significantly alters behavior and monocyte polarization. The

Western diet promoted a proinflammatory monocyte phenotype relative to a Mediterranean diet,

which supports the role of monocyte polarization in diet-associated chronic inflammatory

diseases. Thus, avoiding a Western-style diet and/or consuming a Mediterranean-style diet could

be beneficial in preventing or treating chronic inflammation and disease. The majority of the

effects of diet are presumably mediated through direct or combined actions of

saturated/polyunsaturated fats, n-6:n-3 ratios, pro- and anti-antioxidant characteristics, and other

unique features of the protein, carbohydrate, and fat constituents in the two diets. Monocyte

reprogramming was also partially mediated by the diet-induced changes in behavior, although

the mechanisms by which this occurred are unknown. Ongoing and future work will address

interactions between social behavior (e.g., social status) and diet to further understand how

environmental stressors may impact inflammation in the periphery and in the central nervous

system.

Materials and Methods

Subjects

Forty-three adult (age: mean = 9.0, range = 8.2-10.4 years, estimated by dentition), female

cynomolgus macaques (*Macaca fascicularis*), were obtained (Shin Nippon Biomedical

Laboratories, USA SRC, Alice, TX) and housed at the Wake Forest School of Medicine Primate

Center (Winston-Salem, NC) as previously described [38]. Briefly, the monkeys were socially

housed in groups of 3-4 and consumed standard monkey chow (Table S1) during an eight-month

baseline phase, after which pens were assigned to receive either the Western (5 groups, $n = 21$)

or Mediterranean (6 groups, $n = 22$) diet, balanced on pretreatment characteristics that reflected

overall health, including body weight, body mass index, and plasma triglyceride concentrations

([38]; Fig. 1A). Two monkeys did not tolerate the experimental diet, and were switched to standard

monkey chow, three animals died during the course of the study, and three samples were

removed for insufficient CD14 purification (see "Removal of Batch Effects" below), resulting in

a final sample size of 35 animals (Western $n = 20$, Mediterranean $n = 15$). All animal

manipulations were performed according to the guidelines of state and federal laws, the US

Department of Health and Human Services, and the Animal Care and Use Committee of Wake

Forest School of Medicine.

Experimental Diets

Experimental diets (Table S1) were formulated to be isocaloric with respect to protein, fat, and

carbohydrates, and identical in cholesterol content (~ 320mg / 2000 kilocalories (Cals)/day) as

previously described[38]. The Western diet was formulated to be similar to that consumed by

American women age 40-49 as reported by the US Dept. Agriculture, with protein and fat

derived mainly from animal sources. The Western diet was relatively high in saturated fat and

sodium, and low in monounsaturated fat and n-3 fatty acids. The Mediterranean diet was

formulated to mimic key aspects of the traditional Mediterranean diet, with an n-6:n-3 fatty acid

ratio similar to a traditional hunter-gatherer type diet[12,86,87]. Protein and fats were derived mainly

from plant sources, fish and dairy, and monounsaturated fatty acids were relatively high.

Mediterranean diet contained more complex carbohydrates and fiber, and less sodium and

refined sugars than Western diet. Key ingredients included English walnut powder and extra-

virgin olive oil which were the primary components provided to participants in the PREDIMED

study, a landmark dietary intervention study that illustrated the role of the Mediterranean diet in

cardiovascular disease prevention[88].

Behavioral Characterization

Behavioral data were collected weekly during two 10-minute focal observations, randomly

ordered and balanced for time of day, for 6 weeks during the baseline phase (2 hours/monkey

total) and for 14 months during the experimental phase (17.7 hours/monkey total). Behaviors

were collected as previously described[89], and combined into summary behaviors (e.g.,

"aggression" was a combination of all total, noncontact, contact aggressive events). No

significant differences in behavioral variables were observed between the diet groups which

consuming the baseline standard monkey chow diet. In order to quantify the overall impact of

diet on behavior, we conducted a principal component analysis using the R package *FactoMineR*

[90].

Blood Sample Collection

The monkeys were trained to run out of their social groups on voice command. Blood was drawn

via venipuncture within 9 minutes of entering the building,. Blood was collected into EDTA-

containing tubes, mixed with an equal amount of PBS without calcium or magnesium, and

overlaid on a 90% Ficoll-Paque Plus/10% PBS solution in LeucoSep tubes followed by

centrifugation at 800 x g for 20 min. Isolated PBMCs were then immediately used for the

collection of CD14+ monocytes by positive selection using a Miltenyi bead-based protocol

following manufacturer's instructions (Miltenyi Biotec, Bergisch Gladbach, Germany). After

assessing cell viability and numbers, CD14+ monocytes were stored in 85% FBS, 15% DMSO

sterile freezing media at -80°C and transferred to liquid nitrogen for storage until RNA

extraction.

RNA extraction and sequencing

RNA was extracted from monocytes using the AllPrep DNA/RNA Mini Kit (Qiagen, Inc.,

Hilden, Germany), and quantified using a NanoDrop spectrophotometer and Agilent 2100

Bioanalyzer with RNA 6000 Nano chips (Agilent Technology, Inc., Santa Clara, CA). RNA

libraries were prepared for sequencing by the Cancer Genomics Shared Resource (Wake Forest

School of Medicine, Winston-Salem, NC) using the TruSeq-stranded total RNA kit (Illumina),

which includes a ribosomal depletion step. The RNA-seq libraries were then sequenced using

single-end 76-bp reads on an Illumina NextSeq 500 to an average read depth of 34.5 million

reads per sample (range 25.9 – 41.6 million reads). Reads were mapped to the *Macaca*

fascicularis reference genome (Macaca_fascicularis_5.0, v 93, Ensembl)[91,92] using HiSat2[93] and

then converted to a sample-by-gene read count matrix using featureCounts[94] (median = 38.0%;

range 24.5 - 50.4% of reads mapped to exons).

Read Count Normalization and Removal of Batch Effects

First, we removed genes with low expression (median reads per kilobase per million reads

mapped < 1), which resulted in 12,240 genes for downstream analyses. We normalized read

counts using the *voom* function of the R package *limma*[95]. While investigating monocyte purity,

three samples differed in CD3 gene expression from the rest by several orders of magnitude. We

concluded that these samples were contaminated with CD3+ cells (i.e., inefficient CD14

purification, see Fig. S8) and excluded them from all analyses, leaving a final sample size of 35

monkeys ($n = 20$ fed the Western diet, $n = 15$ Mediterranean diet). To control for batch effects

related to RNA quality and monocyte purity, we calculated the residual gene expression from a

model of normalized gene expression as a function of CD14 expression, CD3 expression, RNA

integrity, and RNA concentration. These residual gene expression values were used for all

subsequent analyses.

Modeling Effect of Diet on Gene Expression

In order to determine which genes were significantly affected by diet, we modeled the residual

expression of each gene as a function of diet using a linear mixed effects model controlling for

relatedness among monkeys using the R package *EMMREML*[96]. Relatedness was estimated using

the ngsRelate program[97] with SNP genotypes inferred from the RNA-seq reads using bcftools

mpileup[98]. We calculated an empirical false discovery rate (FDR) for each gene using a

permutation-based approach[30]. Genes that passed a threshold of FDR < 0.05 were considered

differentially expressed between the two diets. To examine global patterns of variation in gene

expression, we conducted principal component analysis on the correlation matrix of normalized

residual gene expression using the *prcomp* function in R.

Enrichment analyses

Gene ontology (GO) enrichment analyses were conducted using Fisher's Exact Tests and the

weight01 algorithm to test for enrichment implemented in the R package *topGO*[99]. For a more

targeted analysis of M1 and M2 specific genes, we identified a set of differentially expressed

genes in our data set that were previously found to be involved in monocyte polarization[39] (638

proinflammatory and 138 regulatory), which we used to explore monocyte polarization in the

current study. We calculated the proportion of genes more highly expressed in the

Mediterranean- and Western-fed animals in each polarization category and tested for

significance using a permutation test ($n = 100,000$ permutations).

Transcription Factor Binding Site Analysis

We tested for enrichment of transcription factor binding motifs within 2 kb (upstream or

downstream) of the transcription start sites of differentially expressed "Western genes" or

"Mediterranean genes" (FDR < 0.05) using the program HOMER[100] and equivalent regions

around the transcription start sites of all genes expressed in these data as the background set for

enrichment testing. We searched for known vertebrate transcription factor binding motifs and

report the TF motifs passing a threshold of FDR < 0.05.

Gene-gene co-expression analysis

In addition to testing whether diet led to mean differences in gene expression between Western

and Mediterranean animals, we also tested whether diet impacted the correlation structure among

expressed genes (i.e., gene co-expression). Specifically, we used 'correlation by individual level

product' (CILP)[47], to test whether diet affected the magnitude or direction of pairwise gene

expression correlations among the top 140 most differentially expressed genes ($n = 9730$ gene-

gene pairs tested, equivalent to $_{140}C_2$). To test whether a given pair of genes was differentially

co-expressed as a function of diet, we first obtained a vector of products for each gene pair by

multiplying the normalized gene expression values for two genes together. Normalization was

performed by scaling expression values to mean 0 and unit variance within Mediterranean and

Western subsets of the data respectively, to ensure that distributional differences between sample

groups did not bias our results, following previously described procedures[47]. Each of these

vectors of products were used as the outcome variable in a linear mixed effects model

implemented in the R package *EMMREML*[96], which included a fixed effect of diet and a random

effect to control for genetic relatedness. To assess significance, we extracted the p-value

associated with the diet effect for all 9730 gene pairs. We then repeated each linear mixed effects

model 100 times after permuting diet, extracted the p-value associated with the diet effect, and

used these values to calculate an empirical FDR distribution[30].

Using this approach, we identified 445 gene pairs that were significantly differentially co-

expressed as a function of diet at a 20% empirical FDR. Next, we performed two follow up

analyses to understand their biological import. First, we tested for the existence of 'hub genes',

defined as genes that displayed differential co-expression to their tested partner genes more so

than expected by chance. To define the null distribution for identifying hub genes, we randomly

sampled 445 gene pairs from the set of all 9730 tested gene pairs 1000 times and calculated the

number of partners a focal gene had in each sample; we considered a gene to be a significant

'hub gene' if it fell outside the 95th percentile of this distribution, which was equivalent to a focal

gene that displayed significant differential co-expression with 13 or more of its tested partner

genes. Second, we asked whether the set of 'hub genes' we identified were enriched for

transcription factors, relative to the background set of all 140 genes tested for differential co-

expression. We performed this analysis because many of the proposed mechanisms to generate

large scale changes in gene co-expression patterns involve changes in transcription factor

function or activity[45,46]. To implement the enrichment analysis, we used the TRRUST database

of known mammalian transcription factors for annotation [101] paired with hypergeometric tests.

Mediation

To explore relationships between DAB score and differential gene expression, we conducted

mediation analyses using a bootstrapping approach involving 10,000 bootstrap iterations of two

models: (Model 1) the expression of each gene as a function of diet, and (Model 2) the

expression of each gene as a function of diet and DAB score[102]. For each bootstrap iteration, we

then calculated the mediation effect (i.e., the indirect effect) of DAB score as the difference

between the effect size of diet in Model 1 (β_{diet}) and Model 2 (β'_{diet}). We considered there to be a

mediation effect when the 90% confidence interval for the indirect effect ($\beta_{diet}-\beta'_{diet}$) did not

include zero.

A similar method was used to calculate the mediation of gene expression on DAB, testing the

difference between the effect size of diet in two models: (Model 3) DAB as a function of diet,

and (Model 4) DAB as a function of diet and the expression of each gene.

Supplementary Materials

Fig. S1. Diet manipulation altered behavior.

Fig. S2. Behaviors exhibit significant correlations with one another.

Fig. S3. Correlation of observed behaviors with PC1.

Fig. S4. The first PC of all behavioral data captures dominance rank.

Fig. S5. Correlation of observed behaviors with diet-altered behavior measure (DAB; PC2).

Fig. S6. Expression of genes in the conserved transcriptional response to adversity (CTRA[28])

indicate inflammatory effects of a Western diet that parallel the effects of social adversity.

Fig. S7. Greater phenotypic variability in Western diet fed monkeys does not show consistency

in individual responsiveness across phenotypes.

Fig. S8. Quality control of cell purity by CD14 and CD3 expression levels: three samples were

excluded due to lower CD14 and high CD3 – possible T cell contamination.

Fig. S9. RNA Integrity was correlated with both uncorrected gene expression and relative rank.

Table S1. Nutritional Contents of Human and Nonhuman Primate Diets

Table S2. Effects of Diet on Gene Expression

Table S3A. Biological Processes Enriched in Western Genes Compared to Other Measured

Genes

Table S3B. Biological Processes Enriched in Mediterranean Genes Compared to Other Measured

Genes

Table S4. Transcription Factor Binding Site Motif Enrichment

Table S5A. Gene Pair Correlations Across and Within Diet Groups

Table S5B. Differentially Correlated Genes

Table S6A. Behavior Loadings onto Principal Components 1 and 2 and Correlation with Diet

and Rank

Table S6B. Behavior Principal Components and Correlation with Diet

Table S7A. Biological Processes Enriched in Behavior-Mediated Differentially-Expressed Genes

(DEG)

Table S7B. Biological Processes Enriched in Behavior-Mediated Western Genes

Table S7C. Biological Processes Enriched in Behavior-Mediated Mediterranean Genes

Note S1. Regarding rank and RNA integrity (RIN).

References

1.   Giugliano, D., Ceriello, A. & Esposito, K. The effects of diet on inflammation: emphasis on

the metabolic syndrome. *J. Am. Coll. Cardiol.* **48**, 677–685 (2006).

2.   Lopez-Garcia, E. *et al.* Major dietary patterns are related to plasma concentrations of

markers of inflammation and endothelial dysfunction. *Am. J. Clin. Nutr.* **80**, 1029–1035

(2004).

3.   Drescher, H. K. *et al.* The Influence of Different Fat Sources on Steatohepatitis and Fibrosis

Development in the Western Diet Mouse Model of Non-alcoholic Steatohepatitis (NASH).

Front. Physiol. **10**, (2019).

4.   Holt, E. M. *et al.* Fruit and vegetable consumption and its relation to markers of

inflammation and oxidative stress in adolescents. *J. Am. Diet. Assoc.* **109**, 414–421 (2009).

5.   Nanri, A., Moore, M. A. & Kono, S. Impact of C-reactive protein on disease risk and its

relation to dietary factors. *Asian Pac. J. Cancer Prev. APJCP* **8**, 167–177 (2007).

6.   Nettleton, J. A. *et al.* Dietary patterns are associated with biochemical markers of

inflammation and endothelial activation in the Multi-Ethnic Study of Atherosclerosis

(MESA). *Am. J. Clin. Nutr.* **83**, 1369–1379 (2006).

7.   Drake, I., Sonestedt, E., Ericson, U., Wallström, P. & Orho-Melander, M. A Western dietary

pattern is prospectively associated with cardio-metabolic traits and incidence of the

metabolic syndrome. *Br. J. Nutr.* **119**, 1168–1176 (2018).

8. Smyth, S. & Heron, A. Diabetes and obesity: the twin epidemics. *Nat. Med.* **12**, 75–80

(2006).

9. Smil, V. Coronary Heart Disease, Diet, and Western Mortality. *Popul. Dev. Rev.* **15**, 399

(1989).

10. Manzel, A. *et al.* Role of 'Western diet' in inflammatory autoimmune diseases. *Curr. Allergy*

Asthma Rep. **14**, 404 (2014).

11. Jacka, F. N. *et al.* Association of Western and traditional diets with depression and anxiety in

women. *Am. J. Psychiatry* **167**, 305–311 (2010).

12. Cordain, L. *et al.* Origins and evolution of the Western diet: health implications for the 21st

century. *Am. J. Clin. Nutr.* **81**, 341–354 (2005).

13. O'Keefe, J. H., Gheewala, N. M. & O'Keefe, J. O. Dietary strategies for improving post-

prandial glucose, lipids, inflammation, and cardiovascular health. *J. Am. Coll. Cardiol.* **51**,

249–255 (2008).

14. Farchi, G., Fidanza, F., Mariotti, S. & Menotti, A. Is diet an independent risk factor for

mortality? 20 year mortality in the Italian rural cohorts of the Seven Countries Study. *Eur. J.*

Clin. Nutr. **48**, 19–29 (1994).

15. Osler, M. & Schroll, M. Diet and mortality in a cohort of elderly people in a north European

community. *Int. J. Epidemiol.* **26**, 155–159 (1997).

16. Romagnolo, D. F. & Selmin, O. I. Mediterranean Diet and Prevention of Chronic Diseases.

Nutr. Today **52**, 208–222 (2017).

17. Trichopoulou, A. *et al.* Diet and overall survival in elderly people. *BMJ* **311**, 1457–1460

(1995).

18. Hu, F. B. Dietary pattern analysis: a new direction in nutritional epidemiology. *Curr. Opin.*

Lipidol. **13**, 3–9 (2002).

19. Whelton, P. K. *et al.* The Effects of Nonpharmacologic Interventions on Blood Pressure of

Persons With High Normal Levels: Results of the Trials of Hypertension Prevention, Phase

I. *JAMA* **267**, 1213–1220 (1992).

20. Kimmig, L. M. & Karalis, D. G. Do Omega-3 Polyunsaturated Fatty Acids Prevent

Cardiovascular Disease? A Review of the Randomized Clinical Trials. *Lipid Insights* **6**,

LPI.S10846 (2013).

21. Ohlow, M. J., Sohre, S., Granold, M., Schreckenberger, M. & Moosmann, B. Why Have

Clinical Trials of Antioxidants to Prevent Neurodegeneration Failed? - A Cellular

Investigation of Novel Phenothiazine-Type Antioxidants Reveals Competing Objectives for

Pharmaceutical Neuroprotection. *Pharm. Res.* **34**, 378–393 (2017).

22. Steinhubl, S. R. Why Have Antioxidants Failed in Clinical Trials? *Am. J. Cardiol.* **101**, S14–

S19 (2008).

23. Stice, E. & Durant, S. Elevated objectively measured but not self-reported energy intake

predicts future weight gain in adolescents. *Appetite* **81**, 84–88 (2014).

24. Suchanek, P., Poledne, R. & Hubacek, J. A. Dietary intake reports fidelity--fact or fiction?

Neuro Endocrinol. Lett. **32 Suppl 2**, 29–31 (2011).

25. Devêvre, E. F. *et al.* Profiling of the Three Circulating Monocyte Subpopulations in Human

Obesity. *J. Immunol.* **194**, 3917–3923 (2015).

26. Mosser, D. M. & Edwards, J. P. Exploring the full spectrum of macrophage activation. *Nat.*

Rev. Immunol. **8**, 958–969 (2008).

27. Cole, S. W. Social regulation of human gene expression: mechanisms and implications for

public health. *Am. J. Public Health* **103 Suppl 1**, S84-92 (2013).

28. Cole, S. W. *et al.* Loneliness, eudaimonia, and the human conserved transcriptional response

to adversity. *Psychoneuroendocrinology* **62**, 11–17 (2015).

29. Cole, S. W. The Conserved Transcriptional Response to Adversity. *Curr. Opin. Behav. Sci.*

28, 31–37 (2019).

30. Snyder-Mackler, N. *et al.* Social status alters immune regulation and response to infection in

macaques. *Science* **354**, 1041–1045 (2016).

31. Snyder-Mackler, N. & Lea, A. J. Functional genomic insights into the environmental

determinants of mammalian fitness. *Curr. Opin. Genet. Dev.* **53**, 105–112 (2018).

32. Tung, J. & Gilad, Y. Social environmental effects on gene regulation. *Cell. Mol. Life Sci.*

CMLS **70**, 4323–4339 (2013).

33. Kaplan, J. R., Manuck, S. B. & Shively, C. The effects of fat and cholesterol on social

behavior in monkeys.: *Psychosom. Med.* **53**, 634–642 (1991).

34. Warden, C. H. & Fisler, J. S. Comparisons of Diets Used in Animal Models of High-Fat

Feeding. *Cell Metab.* **7**, 277 (2008).

35. Kougias, D. G. *et al.* Effects of Perinatal Exposure to Phthalates and a High-Fat Diet on

Maternal Behavior and Pup Development and Social Play. *Endocrinology* **159**, 1088–1105

(2018).

36. Hollis, F., Mitchell, E. S., Canto, C., Wang, D. & Sandi, C. Medium chain triglyceride diet

reduces anxiety-like behaviors and enhances social competitiveness in rats.

Neuropharmacology **138**, 245–256 (2018).

37. Kasprowska-Liśkiewicz, D. *et al.* The ketogenic diet affects the social behavior of young

male rats. *Physiol. Behav.* **179**, 168–177 (2017).

38. Shively, C. A. *et al.* Mediterranean versus Western Diet Effects on Caloric Intake, Obesity,

Metabolism, and Hepatosteatosis in Nonhuman Primates. *Obes. Silver Spring Md* **27**, 777–

784 (2019).

39. Schmidl, C. *et al.* Transcription and enhancer profiling in human monocyte subsets. *Blood*

123, e90-99 (2014).

40. Schuler, A. *et al.* The MADS transcription factor Mef2c is a pivotal modulator of myeloid

cell fate. *Blood* **111**, 4532–4541 (2008).

41. Scott, E., Simon, M., Anastasi, J. & Singh, H. Requirement of transcription factor PU.1 in

the development of multiple hematopoietic lineages. *Science* **265**, 1573–1577 (1994).

42. Zhang, D. E. *et al.* Sp1 is a critical factor for the monocytic specific expression of human

CD14. *J. Biol. Chem.* **269**, 11425–11434 (1994).

43. Chistiakov, D. A., Myasoedova, V. A., Revin, V. V., Orekhov, A. N. & Bobryshev, Y. V.

The impact of interferon-regulatory factors to macrophage differentiation and polarization

into M1 and M2. *Immunobiology* **223**, 101–111 (2018).

44. Günthner, R. & Anders, H.-J. Interferon-Regulatory Factors Determine Macrophage

Phenotype Polarization. *Mediators Inflamm.* **2013**, 1–8 (2013).

45. de la Fuente, A. From 'differential expression' to 'differential networking' - identification of

dysfunctional regulatory networks in diseases. *Trends Genet. TIG* **26**, 326–333 (2010).

46. Gaiteri, C., Ding, Y., French, B., Tseng, G. C. & Sibille, E. Beyond modules and hubs: the

potential of gene coexpression networks for investigating molecular mechanisms of complex

brain disorders: Beyond modules and hubs. *Genes Brain Behav.* **13**, 13–24 (2014).

47. Lea, A. *et al.* Genetic and environmental perturbations lead to regulatory decoherence. *eLife*

8, (2019).

48. Goldsworthy, M. *et al.* Role of the Transcription Factor Sox4 in Insulin Secretion and

Impaired Glucose Tolerance. *Diabetes* **57**, 2234–2244 (2008).

49. Pearen, M. A. & Muscat, G. E. O. Minireview: Nuclear Hormone Receptor 4A Signaling:

Implications for Metabolic Disease. *Mol. Endocrinol.* **24**, 1891–1903 (2010).

50. Davis, J. P. *et al.* Common, low-frequency, and rare genetic variants associated with

lipoprotein subclasses and triglyceride measures in Finnish men from the METSIM study.

PLOS Genet. **13**, e1007079 (2017).

51. Kanai, M. *et al.* Genetic analysis of quantitative traits in the Japanese population links cell

types to complex human diseases. *Nat. Genet.* **50**, 390–400 (2018).

52. Tachmazidou, I. *et al.* Whole-Genome Sequencing Coupled to Imputation Discovers Genetic

Signals for Anthropometric Traits. *Am. J. Hum. Genet.* **100**, 865–884 (2017).

53. Dluzen, D. F. *et al.* Extracellular RNA profiles with human age. *Aging Cell* **17**, e12785

(2018).

54. Neve, B. *et al.* Role of transcription factor KLF11 and its diabetes-associated gene variants

in pancreatic beta cell function. *Proc. Natl. Acad. Sci. U. S. A.* **102**, 4807–4812 (2005).

55. Benito, X., Fritz, S. C., Steinitz-Kannan, M., Vélez, M. I. & McGlue, M. M. Lake

regionalization and diatom metacommunity structuring in tropical South America. *Ecol.*

Evol. **8**, 7865–7878 (2018).

56. Seltmann, M. W., Helle, S., Adams, M. J., Mar, K. U. & Lahdenperä, M. Evaluating the

personality structure of semi-captive Asian elephants living in their natural habitat. *R. Soc.*

Open Sci. **5**, 172026 (2018).

57. Shively, C. A., Register, T. C., Appt, S. E. & Clarkson, T. B. Effects of long-term sertraline

treatment and depression on coronary artery atherosclerosis in premenopausal female

primates. *Psychosom. Med.* **77**, 267–278 (2015).

58. Maestripieri, D., Schino, G., Aureli, F. & Troisi, A. A modest proposal: displacement

activities as an indicator of emotions in primates. *Anim. Behav.* **44**, 967–979 (1992).

59. Troisi, A. *et al.* Affect regulation in alexithymia: an ethological study of displacement

behavior during psychiatric interviews. *J. Nerv. Ment. Dis.* **188**, 13–18 (2000).

60. Troisi, A. Displacement activities as a behavioral measure of stress in nonhuman primates

and human subjects. *Stress Amst. Neth.* **5**, 47–54 (2002).

61. Schino, G., Perretta, G., Taglioni, A. M., Monaco, V. & Troisi, A. Primate displacement

activities as an ethopharmacological model of anxiety. *Anxiety* **2**, 186–191 (1996).

62. Coleman, K., Robertson, N. D. & Bethea, C. L. Long-term ovariectomy alters social and

anxious behaviors in semi-free ranging Japanese macaques. *Behav. Brain Res.* **225**, 317–327

(2011).

63. Shively, C. A., Register, T. C. & Clarkson, T. B. Social Stress, Visceral Obesity, and

Coronary Artery Atherosclerosis in Female Primates. *Obesity* **17**, 1513–1520 (2009).

64. Camargo, A. *et al.* Expression of proinflammatory, proatherogenic genes is reduced by the

Mediterranean diet in elderly people. *Br. J. Nutr.* **108**, 500–508 (2012).

65. Fledderus, J. O. *et al.* Prolonged shear stress and KLF2 suppress constitutive

proinflammatory transcription through inhibition of ATF2. *Blood* **109**, 4249–4257 (2007).

66. Miyata, Y., Fukuhara, A., Otsuki, M. & Shimomura, I. Expression of Activating

Transcription Factor 2 in Inflammatory Macrophages in Obese Adipose Tissue. *Obesity*

(2012) doi:10.1038/oby.2012.154.

67. Reimold, A. M., Kim, J., Finberg, R. & Glimcher, L. H. Decreased immediate inflammatory

gene induction in activating transcription factor-2 mutant mice. *Int. Immunol.* **13**, 241–248

(2001).

68. Herlaar, E. & Brown, Z. p38 MAPK signalling cascades in inflammatory disease. *Mol. Med.*

Today **5**, 439–447 (1999).

69. Martinez, F. O. & Gordon, S. The M1 and M2 paradigm of macrophage activation: time for

reassessment. *F1000Prime Rep.* **6**, (2014).

70. Nahrendorf, M. & Swirski, F. K. Abandoning M1/M2 for a Network Model of Macrophage

Function. *Circ. Res.* **119**, 414–417 (2016).

71. Wolf, S. A., Boddeke, H. W. G. M. & Kettenmann, H. Microglia in Physiology and Disease.

Annu. Rev. Physiol. **79**, 619–643 (2017).

72. Michalson, K. T. *et al.* Monocyte Polarization is Altered by Total-Body Irradiation in Male

Rhesus Macaques: Implications for Delayed Effects of Acute Radiation Exposure. *Radiat.*

Res. **192**, 121–134 (2019).

73. Thompson, J. R. *et al.* Maternal Diet, Metabolic State, and Inflammatory Response Exert

Unique and Long-Lasting Influences on Offspring Behavior in Non-Human Primates. *Front.*

Endocrinol. **9**, (2018).

74. Cacioppo, J. T., Cacioppo, S., Capitanio, J. P. & Cole, S. W. The neuroendocrinology of

social isolation. *Annu. Rev. Psychol.* **66**, 733–767 (2015).

75. Holwerda, S. W. *et al.* Relative burst amplitude of muscle sympathetic nerve activity is an

indicator of altered sympathetic outflow in chronic anxiety. *J. Neurophysiol.* **120**, 11–22

(2018).

76. Juruena, M. F., Eror, F., Cleare, A. J. & Young, A. H. The Role of Early Life Stress in HPA

Axis and Anxiety. *Adv. Exp. Med. Biol.* **1191**, 141–153 (2020).

77. Shively, C. A. *et al.* Mediterranean diet, stress resilience, and aging in nonhuman primates.

Neurobiol. Stress **13**, 100254 (2020).

78. Bonaz, B., Bazin, T. & Pellissier, S. The Vagus Nerve at the Interface of the Microbiota-Gut-

Brain Axis. *Front. Neurosci.* **12**, (2018).

79. Nagpal, R. *et al.* Gut Microbiome Composition in Non-human Primates Consuming a

Western or Mediterranean Diet. *Front. Nutr.* **5**, 28 (2018).

80. Raison, C. L., Capuron, L. & Miller, A. H. Cytokines sing the blues: inflammation and the

pathogenesis of depression. *Trends Immunol.* **27**, 24–31 (2006).

81. Yang, H. *et al.* Transcriptome profiling of brain myeloid cells revealed activation of Itgal,

Trem1, and Spp1 in western diet-induced obesity. *J. Neuroinflammation* **16**, (2019).

82. Maier, S. F. & Watkins, L. R. Cytokines for psychologists: Implications of bidirectional

immune-to-brain communication for understanding behavior, mood, and cognition. *Psychol.*

Rev. **105**, 83–107 (1998).

83. Graham, L. C. *et al.* Chronic consumption of a western diet induces robust glial activation in

aging mice and in a mouse model of Alzheimer's disease. *Sci. Rep.* **6**, (2016).

84. Du, J. *et al.* The Role of Nutrients in Protecting Mitochondrial Function and

Neurotransmitter Signaling: Implications for the Treatment of Depression, PTSD, and

Suicidal Behaviors. *Crit. Rev. Food Sci. Nutr.* **56**, 2560–2578 (2016).

85. Layé, S., Nadjar, A., Joffre, C. & Bazinet, R. P. Anti-Inflammatory Effects of Omega-3 Fatty

Acids in the Brain: Physiological Mechanisms and Relevance to Pharmacology. *Pharmacol.*

Rev. **70**, 12–38 (2018).

86. Bédard, A., Riverin, M., Dodin, S., Corneau, L. & Lemieux, S. Sex differences in the impact

of the Mediterranean diet on cardiovascular risk profile. *Br. J. Nutr.* **108**, 1428–1434 (2012).

87. Kafatos, A., Verhagen, H., Moschandreas, J., Apostolaki, I. & Van Westerop, J. J.

Mediterranean diet of Crete: foods and nutrient content. *J. Am. Diet. Assoc.* **100**, 1487–1493

(2000).

88. Estruch, R. *et al.* Retraction and Republication: Primary Prevention of Cardiovascular

Disease with a Mediterranean Diet. N Engl J Med 2013;368:1279-90. *N. Engl. J. Med.* **378**,

2441–2442 (2018).

89. Shively, C. A. Social subordination stress, behavior, and central monoaminergic function in

female cynomolgus monkeys. *Biol. Psychiatry* **44**, 882–891 (1998).

90. Lê, S., Josse, J. & Husson, F. **FactoMineR** : An *R* Package for Multivariate Analysis. *J. Stat.*

Softw. **25**, (2008).

91. Kersey, P. J. *et al.* Ensembl Genomes 2018: an integrated omics infrastructure for non-

vertebrate species. *Nucleic Acids Res.* **46**, D802–D808 (2018).

92. Kinsella, R. J. *et al.* Ensembl BioMarts: a hub for data retrieval across taxonomic space.

Database J. Biol. Databases Curation **2011**, bar030 (2011).

93. Kim, D., Langmead, B. & Salzberg, S. L. HISAT: a fast spliced aligner with low memory

requirements. *Nat. Methods* **12**, 357–360 (2015).

94. Liao, Y., Smyth, G. K. & Shi, W. featureCounts: an efficient general purpose program for

assigning sequence reads to genomic features. *Bioinforma. Oxf. Engl.* **30**, 923–930 (2014).

95. Ritchie, M. E. *et al.* limma powers differential expression analyses for RNA-sequencing and

microarray studies. *Nucleic Acids Res.* **43**, e47 (2015).

96. Akdemir, D. & Godfrey, O. U. *EMMREML: Fitting Mixed Models with Known Covariance*

Structures. (2015).

97. Hanghøj, K., Moltke, I., Andersen, P. A., Manica, A. & Korneliussen, T. S. Fast and accurate

relatedness estimation from high-throughput sequencing data in the presence of inbreeding.

GigaScience **8**, (2019).

98. Li, H. *et al.* The Sequence Alignment/Map format and SAMtools. *Bioinforma. Oxf. Engl.* **25**,

2078–2079 (2009).

99. Alexa, A. & Rahnenfuhrer, J. *topGO: Enrichment Analysis for Gene Ontology.* (2019).

100. Heinz, S. *et al.* Simple combinations of lineage-determining transcription factors prime cis-

regulatory elements required for macrophage and B cell identities. *Mol. Cell* **38**, 576–589

(2010).

101. Han, H. *et al.* TRRUST v2: an expanded reference database of human and mouse

transcriptional regulatory interactions. *Nucleic Acids Res.* **46**, D380–D386 (2018).

102. Preacher, K. J. & Hayes, A. F. SPSS and SAS procedures for estimating indirect effects in

simple mediation models. *Behav. Res. Methods Instrum. Comput. J. Psychon. Soc. Inc* **36**,

717–731 (2004).

Acknowledgments: We thank Beth Uberseder, Maryanne Post, JD Bottoms, Edison Floyd,

Jason Lucas, Joshua Long, Diane Wood, and Sherri Samples for technical support. We thank

Nicholas Lozier, Tiffany Pan, Marina Watowich, and Jenny Tung for their helpful feedback on

previous versions of this manuscript.

Funding: This work was funded by R01HL087103 (CAS), R01HL122393 (TCR),

U24DK097748 (TCR) from NIH and intramural funding from the Department of Pathology,

Wake Forest School of Medicine (CAS). NSM was supported by R00AG051764 and

R01AG060931 from NIH, and AJL was supported by a postdoctoral fellowship from the Helen

Hay Whitney Foundation. The Wake Forest Comprehensive Cancer Center Cancer Genomics

Shared Resource is supported by P30CA012197 and by a NIH Shared Instrumentation Grant

S10OD023409 to GAH.

Author Contributions: Conceptualization, C.A.S. and T.C.R.; Methodology, C.A.S., T.C.R.,

N.S.-M., and C.S.C.J.; Formal Analysis – C.A.S., T.C.R., N.S.-M., A.J.L., and C.S.C.J;

Investigation, M.E., A.N.V., N.A.V., S.C.P., and S.Y.W.; Writing – Original Draft, C.A.S.,

T.C.R., N.S.-M., and C.S.C.J.; Writing – Review & Editing, all authors; Visualization – N.S.-M.,

A.J.L., and C.S.C.J; Funding Acquisition, C.A.S. and T.C.R.; Supervision, C.A.S., T.C.R., and

N.S.-M.

Competing Interests: The authors declare no competing interests.

Data Availability

All data and code used to complete these analyses can be found at

https://github.com/cscjohns/diet_behavior_immunity. The raw data can be accessed from the

gene expression omnibus repository from accession # GSE144314.