Running Title: Population structure in Goodman's mouse lemur

Population genomic structure in Goodman's mouse lemur reveals long-standing

separation of Madagascar's Central Highlands and eastern rainforests

George P. Tiley[1,†], Marina B. Blanco[1,2], José M. Ralison[3], Rodin M. Rasoloarison[3,4],

Amanda R. Stahlke[5], Paul A. Hohenlohe[5], Anne D. Yoder[1]

[1]Department of Biology, Duke University, Durham, NC 27708, USA

[2]Duke Lemur Center, Duke University, Durham, NC 27705, USA

[3]Département de Biologie Animale, Université d'Antananarivo, BP 906, Antananarivo 101,

Madagascar

[4]Behavioral Ecology and Sociobiology Unit, German Primate Centre, 37077 Göttingen,

Germany

[5]Institute for Bioinformatics and Evolutionary Studies, Department of Biological Sciences,

University of Idaho, Moscow, ID 83844, USA

[†]Author for correspondence: george.tiley@duke.edu

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Abstract

The Central Highland Plateau of Madagascar is largely composed of grassland savanna,

interspersed with patches of closed-canopy forest. Conventional wisdom has it that these

grasslands are anthropogenic in nature, having been created very recently via human

agricultural practices. Yet, the ancient origins of the endemic grasses suggest that the extensive

savannas are natural biomes, similar to others found around the globe. We use a

phylogeographic approach to compare these two competing scenarios. By sampling multiple

populations of Goodman's mouse lemur (*Microcebus lehilahytsara*), a small-bodied nocturnal

primate, we reconstruct the phylogeographic and demographic history of these "environmental

metronomes" to estimate the time at which their populations diverged, and thus proximally,

when their habitats would have become fragmented. We applied coalescent methods to

RADseq data to infer phylogenetic relationships, population structure, and migration corridors

among sampling sites. These analyses indicate that forest fragmentation occurred rapidly during

a period of decreased precipitation near the last glacial maximum and would have affected both

the Central Highlands and eastern forests. Though there is clear genomic structure separating

the populations of the Central Highland from those of the eastern rainforests, there is also

evidence of historical migration between them. Findings support the hypothesis that the Central

Highland savanna predates human arrival, indicating that it is a natural landscape that has long

impacted the population dynamics of Goodman's mouse lemur, and by extension, other forest-

dwelling organisms in Madagascar.

Introduction

Madagascar is an iconic biodiversity hotspot with high levels of species diversity and endemism (Myers et al., 2000) with the majority of its biodiversity at extreme risk of extinction due to deforestation and forest fragmentation (Greene & Sussman, 1990; Harper, Steininger, Tucker, Juhn, & Hawkins, 2007; Vieilledent et al., 2018). It is widely accepted that the rapid rate of biodiversity loss is due largely to human activity and agricultural practices (Farris et al., 2017; Styger, Rakotondramasy, Pfeffer, Fernandes, & Bates, 2007). Even so, the timing of and degree to which humans have transformed and fragmented Madagascar's closed-canopy forests has been a point of contention. Much of central Madagascar is now covered with savanna; traditionally thought to be strictly the product of fires from slash-and-burn agriculture and cattle farming (Perrier de la Bâthie, 1921; Humbert, 1927; Gade, 1996). The best available archeological evidence places humans in Madagascar between two thousand years ago (KYA) and ten KYA (Dewar & Wright, 1993; Dewar et al., 2013; Hansford et al., 2018; Mitchell, 2019). This timeline coincides with evidence of increased frequency of fire based on isotope records from stalagmites (Railsback et al., 2020) and charcoal records from sediment cores covering the late to mid Holocene (Burney, 1987a; Gasse & Van Campo, 1998; Virah-Sawmy, Willis, & Gillson, 2010). Pollen abundances from these same sediment cores also imply that the Central Highland vegetation transitioned from woody taxa to grasses during this timeframe, thus supporting the hypothesis that the Central Highland Savanna grasslands are the consequence of human activities. Confusingly, however, it appears that the transition to savanna likely began following Pleistocene glaciations (Burney, 1987b; Gass & Van Campo, 1998), long before the arrival of humans, and that similar shifts in plant composition were not limited to the Central Highlands (Burney, 1993; Virah-Sawmy, Willis, & Gillson 2009). This therefore implies that the Central Highland Savanna (CHS) is not strictly the product of human agricultural practices, and instead, may be the consequence of global climate fluctuations, perhaps facilitated by the natural processes of fire ecology (Bowman & Franklin, 2005).

In support of this second scenario, ecological and phylogenetic studies of Madagascar's

grasses suggest the that the CHS originated well before the Holocene, with levels of endemism

comparable to the putatively natural grasslands of Africa (Bond, Silander Jr, Ranaivonasy, &

Ratirarson, 2008; Vorontsova et al., 2016). And though few C_4 grass lineages such as *Aristida*

and *Loudetia* dominate the CHS (Koechlin, 1993; Kull, 2003; Solofondranohatra et al., 2018),

diversity among endemic C_4 grass clades would be more consistent with the global

diversification of C_4 grasses during the Miocene (e.g. Edwards & Smith, 2010) rather than as a

consequence of a large number of independent migration events (Hackel et al., 2018). We are

therefore left with a complex model of Pleistocene climate change, perhaps driving the origins of

the CHS as a savanna, but overlaid with deforestation and fragmentation, likely associated with

early human activity. Consequently, we hypothesize that present-day distributions of forest-

dwelling species, as well as the genetic diversity and connectivity among their populations,

should reflect *both* ancient climactic changes as well as recent human-meditated processes. It

is our goal to examine phylogenomic patterns among populations of Goodman's mouse lemur

(*Microcebus lehilahytsara*) to disentangle the impacts of these two environmental scenarios.

The geographic distribution of *M. lehilahytsara* is uniquely suited for this purpose. Though it is

found in eastern montane forests, it has largely been considered to be a highland specialist

(Radespiel et al., 2012; Blanco et al., 2017). Indeed, it is the only mouse lemur species known

to inhabit the isolated forest fragments in the CHS (Blanco et al., in review). The extended range

of *M. lehilahytsara* may be due to its ability to enter a state of prolonged torpor (Blanco et al.,

2017) thus reducing metabolic demands during the period of extended low temperatures in the

CHS during the winter season. While prolonged torpor or hibernation is obligatory in

Cheirogaleus, another member of the family Cheirogaleidae (e.g. Dausmann, Glos, Ganzhorn,

& Heldmaier, 2005; Blanco et al., 2018), mouse lemurs vary in their expression of daily torpor or

hibernation, with some species reported to display daily torpor (less than 24 hours) exclusively

and unable to deposit fat stores to sustain prolonged torpor (e.g. Schmid, 2001; Blanco et al.,

2018). Although losses in genetic diversity of endangered species putatively due to human

action are observable (Olivieri, Sousa, Chikhi, & Radespiel, 2008; Andriaholinirina et al., 2014a;

Andriaholinirina et al., 2014b; Baden et al., 2014), a better reconstruction of pre-human

conditions can reveal if some populations were already vulnerable, especially given that some

populations have been robust to recent habitat loss and fragmentation (Sgarlata et al., 2018).

A recent phylogeographic study of mouse lemurs that included a very limited representation of

M. lehilahytsara from the CHS suggested that it diverged from its sister species *M. mittermeieri*

in the eastern rainforests near 50 KYA (Yoder et al., 2016). Subsequently, the species status of

M. mittermeieri has been called into question with compelling genetic evidence indicating that it

should be synonymized with *M. lehilahytsara* (Poelstra et al., 2020). This suggests the intriguing

possibility that rather than be a highland specialist, as was originally hypothesized, *M.*

lehilahytsara may instead be a widely distributed generalist, capable of existing in a broad range

of habitat types occurring at varying elevations. Thus, demographic studies of these mouse

lemurs may illuminate pre-human conditions in Madagascar given that they can arguably be

described as "environmental metronomes," expanding and contracting their populations to

match climatically driven pulses in forest distributions.

Here, we sampled populations of putative *M. lehilahytsara* from multiple sites in the CHS as well

as from eastern forests. Using phylogeographic approaches and coalescent methods to analyze

a large RADseq dataset, we explored patterns of genetic connectivity among populations. We

also estimated divergence times and effective population size (N_e) among and between them.

Findings show that population structure formed well-before any human influence, with isolation

of populations in the CHS and eastern forests occurring before the LGM. However, post-

divergence gene flow among the populations suggests that migration was possible between the

CHS and eastern forests, posing no more of a geographic barrier than major rivers. Moreover,

population sizes were largely robust to past climate change, and have actually increased

through the Holocene, even within a 1 Km^2 forest patch.

Methods

Study sites and sampling

Microcebus lehilahytsara individuals were captured at five field sites. Three of the locations,

from north to south, Riamalandy (S16° 17' 06.0" E48° 48' 54.0" 850 m), Ambatovy (S18° 49'

30.15" E48° 19' 06.48" 1100 m) and Tsinjoarivo (S19° 40' 48.5" E47° 46' 14.5" 1609 m) are mid-

elevation rainforests, situated along the eastern escarpment. Forests are fragmented along the

corridor; although, the distance between patches has been exacerbated by deforestation since

the 1950s (Vieilledent et al. 2018). Annual rainfall is 1400 mm and daily temperatures vary

between 14 and 23 °C at Riamalandy (Goodman, Raherilalao, & Wohlauser, 2018), while

Tsinjoarivo and Ambatovy, receive 1700-2000 mm of rainfall with temperatures between 15 and

23 °C (Goodman et al. 2018). At their lowest, temperature can approach freezing conditions at

Tsinjoarivo (Dausmann & Blanco, 2016). The other two locations, Ambohitantely and Ankafobe

(S18° 06' 23.1" E47° 11' 13.2" 1472 m) are located within the Central Highlands. Their

vegetational components are characteristic of moist evergreen forests, although more open and

drier (Goodman et al., 2018). Annual rainfall is around 1460 mm, and daily temperature

between 12.5 and 23 °C, though in winter temperatures can drop substantially, below 6 °C

(Goodman et al., 2018). In addition to generally drier conditions at Ambohitantely and Ankafobe

compared to Ambatovy and Tsinjoarivo, these sites experience greater seasonality of

precipitation too (Supplementary Figure S1). Ambohitantely is a 5600 ha protected area 130 Km

northwest of Antananarivo that incorporates natural forest, savanna, and exotic tree plantations,

but is subject to annual fires and threated by forest loss. Samples were collected in 2004

(Riamalandy), 2006 (Ambohitantely), 2007 (Tsinjoarivo), between 2009 and 2015 (Ambatovy),

and between 2015 and 2016 (Ankafobe). Mouse lemurs were captured using live Sherman

traps and released back to the forest later the same day. For description of capture methods

see Blanco et al. (2017).

RADseq library preparation and sequencing

Microcebus lehilahytsara samples were gathered from existing collections and a combination of

genomic DNA from available tissue and previous whole-genome amplifications were used in

order to maximize sampling of these rare and vulnerable species. Library preparations for RAD

sequencing followed Ali et al. (2016), using a single-digest protocol with the restriction enzyme

SbfI. A median insert size of 380 bp was selected for paired-end sequencing at the Duke

Sequencing Core on an Illumina Hi-Seq 4000 with an anticipated 15x coverage per individual.

Assembly and orthology evaluation

Data were assembled using STACKS v2.0b (Catchen, Amores, Hohenlohe, Cresko, &

Postlethwait, 2011; Catchen, Hohenlohe, Bassham, Amores, & Cresko, 2013) using *de novo*

methods (Supplementary Methods). Final phased loci (we use locus or loci to refer to a

contiguous RAD fragment while sites refer to individual base pairs throughout) output by stacks

were filtered such that at least three of the five sample geographic locations were represented

with a maximum of 50% missing data (Supplementary Methods). These filtering options were

applied in order to maintain computational feasibility of downstream model-based hypothesis

testing procedures in addition to not biasing our analyses due to uneven sampling from

localities. Additionally, most RADseq-based studies rely solely on heuristic measures such as

numbers of individuals and some level of heterozygosity for identifying orthologous clusters; we

use a different approach for evaluating orthology that we argue is less sensitive to any

clustering parameters. We used syntenic evidence from the *Microcebus murinus* reference

genome, with BLASTN (Altschul et al.,1990; Camacho et al., 2009) search of assembled RAD

loci, to identify clusters susceptible to paralogy as well as merge orthologous clusters that were

over-split by STACKS (Supplementary Methods).

An important note regarding our sequence data and library preparation method is that we

expect even depth of coverage for the forward read with the restriction enzyme cut site, but

lowered coverage for the reverse read with the staggered cut. Therefore, while the longer

contigs assembled with the paired-end data are useful for circumscribing orthologous loci, we

are skeptical of our ability to consistently call heterozygous loci with high confidence beyond the

first 151 bp of each locus; many of our downstream hypothesis tests assume we can accurately

call these heterozygous sites. Therefore, for each sequence for each orthologous cluster, we

extracted 145 bp, the first 151 bp minus the 6 bp sequenced overhang from the SbfI restriction

enzyme, for all future analyses. These 145 bp fragments with heterozygous base calls

underwent a final alignment with MUSCLE (Edgar, 2004) v3.8.1551 using default parameters.

Phylogenetic analyses

We estimated a phylogenies of all *M. lehilahytsara* individuals or of sampling sites using four

approaches: 1) concatenated ML analysis with RAxML (Stamatakis, 2014) v8.2.11, 2)

topological invariants of quartets approximated with SVDquartets (Chifman & Kubatko, 2014)

implemented through PAUP* (Swofford, 2003) v4.0a build 165, 3) a Bayesian implementation of

the multispecies coalescent (MSC) model for SNP data (Bryant et al., 2012) through BEAST2

(Bouckaert et al., 2014) v2.4.8, 4) full-likelihood implementation of the MSC with joint gene tree

estimation using BPP v4.0 (Flouri, Jiao, Rannala, & Yang, 2018). Applying these four different

methods allows us to explore uncertainty in phylogeographic history due to model assumptions

and the underlying data used for each method.

RAxML analyses were unpartitioned and used the GTR+Γ model of molecular evolution

(Tavaré, 1986; Yang, 1994). We conducted 20 unconstrained ML searches and performed 100

bootstrap replicates. We also used the ML framework to perform topological hypothesis testing.

Considering that samples from Ambohitantely and Ankafobe are well-supported clade across all

analyses, our phylogenetic question reduces to a single quartet. We then tested three

competing hypotheses regarding the evolutionary history of Riamalandy, Ambatovy, and

Tsinjoarivo. Constrained optimizations of topologies and site likelihoods were performed with

RAxML and we evaluated support among our three competing topologies with an approximately

unbiased (AU; Shimodaira, 2004) test implemented in CONSEL (Shimodaira & Hasegawa,

2001) v0.20.

RADseq data were evaluated using multiple approaches for topological estimation from

invariants. First, we used full-length RAD loci with SVDquartets, which is robust to violations of

independence of loci (Chifman & Kubatko, 2014). Second, we analyzed only variable sites that

represented at least four individuals. To further examine sensitivity of topology estimation with

SVDquartets to our underlying data, we analyzed 10 datasets that only used biallelic SNPS,

randomly sampling one SNP per locus. Analyses sampled all quartets and performed 100

bootstrap replicates. Distributions of bootstrap trees from SVDqaurtets and ML analyses were

compared using unweighted Robinson-Foulds (RF) distances (Robinson & Foulds, 1981)

computed with RAxML.

For SNAPP, we analyzed a single dataset using one biallelic SNP per RAD locus. To reduce the

amount of computing time needed for each analysis, we selected only two individuals per

sampling location. Two replicates were run to evaluate convergence. Mixing and effective

sample sizes were analyzed with TRACER v1.7 (Rambaut et al., 2018), while potential scale

reduction factors were calculated with the R package CODA (Plummer, Best, Cowles, & Vines,

2006). Bayes factor analyses were used to explore competing topological hypotheses,

complementary to our AU tests in the ML framework. We used stepping-stone integration (Xie et

al., 2011) to obtain marginal likelihoods for 20, 40, and 60 steps. It is difficult to predict how

many steps are necessary for marginal likelihood estimation, so we used a range to observe

little change in differences of marginal likelihoods with increasing steps. Because differences in

marginal likelihoods were large, we presented natural log Bayes factors (Kass & Raferty, 1995)

as opposed to computing model probabilities. Prior settings and details regarding MCMC

options are in the Supplementary Methods (Supplementary Material).

Species tree analyses with BPP used fixed species delimitations. We used full-length RAD loci,

considering at least two individuals for sampling location. Eight independent chains collected

2500 posterior samples, which were combined for a posterior of 20000 samples total.

Robustness of species tree estimation to prior choice on common ancestor ages and population

sizes was explored with intentionally inappropriate priors (Supplementary Material).

Delimiting population structure and testing migration hypotheses

To elucidate migration patterns between *M. lehilahytsara* populations in the CHS and eastern

forests, we developed competing migration hypotheses and tested models with Bayes factors.

First, we analyzed biallelic SNPs from RAD loci with STRUCTURE (Pritchard, Stephens, &

Donnelly, 2000) v. 2.3.4, using the admixture model and default priors while treating each site

unlinked and uncorrelated. We created 20 datasets by sampling 10000 unlinked SNPS per

replicate. For each replicate, we ran 20 independent MCMC chains for one to eight clusters

assumed *a priori* (k) in order to calculate Δk for estimating the most likely number of Hardy-

Weinberg groups (Evanno, Regnaut, & Goudet, 2005). Each chain ran for one million

generations after a 100000 generation burn-in.

Population structure was also evaluated with fixation index (F_{ST}) statistics. Pairwise F_{ST} was

calculated between populations as described by Weir & Cockerham (1984) for SNPS with less

than 15% missing data in ARLEQUIN (Excoffier & Lischer, 2010) v. 3.5.2.2. Pairwise F_{ST} were

used to perform an AMOVA analysis (Excoffier, Smouse, & Quattro, 1992) for Central Highlands

(Ambohitantely and Ankafobe) versus eastern (Riamalandy, Ambatovy, and Tsinjoarivo) *M.*

lehilahytsara. A Mantel test was used to test the presence of isolation-by-distance using 9999

permutations. Physical distances between populations were calculated with the Geographic

Distance Matrix calculator v 1.2.3 (http://biodiversityinformatics.amnh.org/open_source/gdmg;

Last Accessed 2 January 2020).

While STRUCTURE analyses can estimate the posterior probabilities of admixture among

Hardy-Weinberg groups, this is not a hypothesis test. As an independent evaluation of gene

flow between *M. lehilahytsara* populations, we computed Bayes factors for isolation-with-

migration models using MIGRATE v4.2.14 (Beerli & Palczewski, 2010). Notably, we did not use

the default uniform prior for nucleotide diversity (θ) estimates. Instead, we used independent

effective population size (N_e) estimates from three mouse lemur species prior to forest

fragmentation (Olivieri et al., 2008) to develop a prior distribution for θ (Supplementary Material).

Two sets of migration hypothesis testing were conducted on full-length RAD loci. First, we used

all individuals binned into either a CHS or eastern population, despite recognizable population

substructure. We were limited in the number of loci computationally feasible for obtaining

marginal likelihood estimates, and thus analyzed 100 data sets of 100 random RAD loci;

missing data was allowed as long as at least two individuals per sampling site were present.

The second analysis attempts to differentiate between east-to-west migration and west-to-east

migration among Ambatovy, Ambohitantely, and Ankafobe, and does not allow for missing data.

The three-population analyses used four diploid individuals per population and compared

isolation-with-migration models to those that also included divergence. Posterior estimates of θ

was rescaled N_e to using a mutation rate of 1.64 x 10^{-8}, which is based on a direct germline

estimate for mouse lemurs (Campbell et al., 2020).

Estimating divergence time of eastern and Central Highland mouse lemurs

We used the multispecies coalescent (MSC) model for divergence time estimation among *M.*

lehilahytsara populations with BPP (Rannala & Yang, 2003; Flouri et al., 2018) v4.0. However,

estimating divergence times and population sizes efficiently requires a fixed and rooted

topology. We used the topology recovered in the majority of analyses here, and used several

lines of evidence to support rooting with Riamalandy (Supplementary Material), including a

phylogeny of *M. lehilahytsara* individuals and outgroups using previously sequenced

mitochondrial data (Table S2). A partitioned ML analysis and bootstrapping was conducted with

PAUP* v4.0a build 163 (Swofford, 2003). Phased haploid sequences were used for BPP and at

least two individuals per population were required for each locus. We ran eight chains of 2500

samples and evaluated convergence. Final MSC parameter estimates were based on combined

chains. Coalescent branch lengths were rescaled to absolute time using a direct germline

mutation rate estimate in mouse lemurs (1.64×10^{-8}; Campbell et al., 2020) and generation time

estimate (3.75 years/generation) based on wild (Radespiel et al., 2019) and captive (Zohdy et

al., 2014) populations. Robustness of divergence times to prior choice was also evaluated

(Supplementary Material).

Characterizing population size changes during Pleistocene glaciations

We estimated change in population size over time with extended Bayesian skyline plots (Heled

& Drummond 2008) with BEAST2 (Bouckaert et al., 2014). Similar analyses have also been

successful in discovering king penguin population increases that coincide with Holocene

warming (Trucchi et al., 2014). For all sampling locations with four or more individuals, we

sampled 100 random RAD loci eight times, with no missing data. Sequences were treated as

phased and haploid. Each sampled dataset was run twice to check convergence. Large

posteriors were required for agreement on the number of change points and population size

estimates between replicates; each chain ran for 500 million generations after a 100 million

generation burnin while sampling every 10000 generations. Operators for increasing the

sampling of change points followed recommendations by Trucchi et al. (2014). We used linear

piecewise reconstructions of population size changes.

Results

Circumscribing orthologous RAD loci

All individuals included in our analyses had an average depth of coverage greater than 10x per

locus assembled with STACKS. In total, 92,889 loci were recovered across all individuals with

an average of 49,201 loci per individual (Supplementary Table S1). On average, 31% of loci

were shared between pairs of individual (Figure 1). The number of loci shared between

individuals are likely not due to phylogenetic signal, as there was only a slight negative

correlation between evolutionary distance and the proportion of shared loci (Pearson's r = -0.19;

p = 0.001). Rather, this pattern was driven, at least in part, by the number of reads, given a

strong positive correlation between the number of loci and the number of reads per individual

(Pearson's r = 0.86; p = 3.54e-08). Although 92,889 RAD loci were recovered, only 54,018 loci

contained at least one biallelic SNP; other loci were completely invariable or consisted of only

singletons.

Support for Central Highland and eastern Microcebus lehilahytsara

All phylogenetic analyses recovered a well-supported split between *M. lehilahytsara* from

Central Highland (Ambohitantely and Ankafobe) and eastern forest (Riamalandy, Ambatovy,

and Tsinjoarivo) localities (Figure 2; Supplementary Figures S2-S8). But the relationships

among eastern sampling locations was contentious across analyses. The ML, SNAPP, and BPP

analyses supported Ambatovy as sister to Tsinjoarivo (H_{TA}; Supplementary Figures S2- S5),

although posterior probabilities from SNAPP revealed uncertainty in the bipartition in question.

SVDquartets strongly conflicted the predominant H_{TA} topology, favoring Riamalandy sister to

Tsinjoarivo (H_{RT}) in all bootstrap replicates when analyzing all variable sites (Supplementary

Figure S6) or complete RAD loci (Supplementary Figure S7). However, the H_{RT} topology, even

with perfect bootstrap support, is sensitive to the underlying data. Analyses of biallelic SNPs

alone with SVDquartets recovered a third topology (H_{RA}); also with high bootstrap support at

bipartitions of sampling locations (Supplementary Figure S8). In one out of ten resampled

datasets, when treating all individuals as independent lineages, Ambatovy was not recovered as

a monophyletic clade. Topological hypothesis testing differentiated among the three recovered

phylogenetic hypotheses: H_{TA}, H_{RT}, and H_{RA}.

AU tests rejected H_{RT}, which came from SVDquartets with all variant sites or full loci. But the AU

test could not reject H_{RA}, despite H_{TA} being present in the majority of ML bootstrap trees and

post burn-in trees (Figure 3). Bayes factors from SNAPP provided insights into the prevalence

of both hypotheses across analyses. SNAPP supported lumping Riamalandy and Ambatovy

together even with increasing step sizes and arguably more precise marginal likelihood

estimates as opposed to lumping Ambatovy and Tsinjoarivo (Table 1). This result is surprising,

considering H_{TA} was the maximum clade credibility tree from the posterior sample in the

unconstrained analyses (Supplementary Figures S3 and S4). All phylogenetic evidence

suggests a shared common ancestor for Ambatovy and Tsinjoarivo, but Tsinjoarivo may be

more isolated due to barriers to gene flow and therefore more evolutionarily distant than

expected. Phylogenetic analyses showed strong support for both CHS and eastern clades, and

at least no very recent gene flow in the last few generations as evident by monophyly of our five

sampling locations. However, such analyses do not rule out gene flow historical gene flow

between the CHS and eastern forests.

Migration between eastern and Central Highland forests

Population genetic analyses revealed admixture between eastern and Central Highland *M.*

lehilahytsara as well as population structure within the eastern rainforest (Tsinjoarivo versus

Ambatovy and Riamalandy) that mirrored findings of SNAPP Bayes factors (Table 1).

STRUCTURE analyses recovered an optimal k = 3 in 19 out of 20 sampled datasets

(Supplementary Figures S9-S11). The remaining replicate recovered an optimal k = 2

(Supplementary Figure S11). All analyses supported a clustering of Central Highland and

eastern *M. lehilahytsara*, with some degree of admixture between the two. The Ambohitantely

site, while phylogenetically sister to Ankafobe, shares about half of its genotypes with Ankafobe

and the other half with Ambatovy/Riamalandy. Tsinjoarivo often forms its own unique cluster,

except in the one replicate where k = 2 and Tsinjoarivo clusters with other eastern forest

populations.

Admixture between Central Highland and eastern *M. lehilahytsara* was also supported by

analyses of genetic variation with F_{ST} statistics. Similar to STRUCTURE analyses, admixture

between the CHS and eastern forests is largely limited to Ambohitantely and Ambatovy,

respectively. Pairwise F_{ST} revealed similar levels of genetic divergence between Ambatovy and

Ambohitantely to that of Ambatovy and Tsinjoarivo (Supplementary Table S3). The geographic

structure between Central Highland and eastern *M. lehilahytsara* explains a small fraction of

genetic variation, with most variation explained within populations (Supplementary Table S4). A

Mantel test revealed little evidence for isolation-by-distance (pearson's r = 0.09; p = 0.39),

suggesting barriers to gene flow aside distance alone (Supplementary Figure S12).

Bayes factors were computed for different isolation-with-migration models to further investigate

the patterns of admixture observed between the Ambatovy, Ambohitantely, and Ankafobe

genetic clusters. The *n-islands* model was unambiguously found to be the best model for both

the analyses that included all individuals modeled as two populations with a subset of data

(Supplementary Figure 13; Supplementary Table 5) or three populations with all of the data

(Figure 4). There were no large discrepancies in the estimated migration rates; however, the

selected model suggested lower effective population sizes in Ankafobe and Ambohitantely

compared to Ambatovy. Assuming a mutation rate of 1.64 x 10^{-8}, MIGRATE estimated

contemporary population sizes to be 12,652 for Ankafobe and Ambohitantely, and 17,226 for

Ambatovy. Their population size estimates, although influenced by the prior, are not driven by

the prior alone (Supplementary Figures S14-S16). Additionally, a higher population size in

Ambatovy is supported across all models regardless of the migration and divergence scenarios

(Figure 4). Because of Ambatovy's higher effective population size, the number of migrants

leaving towards Ambohitantely or Ankafobe per generation is approximately 1.2, compared to

0.8 effective migrants per generation leaving Ambohitantely or Ankafobe towards the east.

Dating Population Divergences

Using the H_{TA} topology supported by Bayes factor analyses, divergence times were estimated

using the MSC model. The topology was rooted with Riamalandy based on the analysis of

mitochondrial data from *Microcebus lehilahytsara* and outgroup taxa (Supplementary Figure

S17) and inference of clock-like trees from BPP (Supplementary Figure 5). Most MSC

parameters converged across eight chains with potential scale reduction factors approaching 1

(Supplementary Table S6). A few parameters still retained high variation between chains for

both θ and τ, but this discrepancy among chains was largely due to one or two chains with

inefficient mixing for different prior combinations (Supplementary Figure S18-S25). Divergence

times were not strongly influenced by the priors (Figure 5) and imply that divergence between

the CHS and eastern forest, as well as the forest patches within them, occurred almost

simultaneously. Absolute estimates inferred that divergence among our sampled lineages

occurred near 25 KYA (Table 2).

Population declines in the CHS

Population sizes experienced some slight contractions until 25 KYA (Figure 6) that coincide with

the divergence of sampling locations (Table 2). Population sizes then increased through the

Holocene and until the present except in Ankafobe. Ambatovy had consistently higher N_e

estimates across replicates. Ankafobe had the lowest N_e estimates. Intermediate N_e estimates

are observed for individuals at Ambohitantely and Tsinjoarivo. There is high uncertainty in N_e

based on comparisons across replicates (Supplementary Figure S26), but individual analyses

tended to converge (Supplementary Figures S27-S30).

Discussion

Population structure of M. lehilahytsara predates human colonization of Madagascar

Our results demonstrated that *M. lehilahytsara* populations have been present in the CHS at

least before the LGM. Phylogenetic analyses strongly supported a split between CHS and

eastern forest sampling locations (Figures 2 and 3; Supplementary Figures S2-S8), although

analyses of genetic structure (Figure 2; Supplementary Figure S11; Supplementary Figure 12;

Supplementary Table S3) and migration rates (Figure 4) implied gene flow after the initial

divergence. If, on the other hand, the divergence of *M. lehilahytsara* in the CHS and eastern

forest had been recent and human mediated, we would expect at least some phylogenetic

uncertainty due to low levels of genetic divergence, similar to Tsinjoarivo, Ambatovy, and

Riamalandy. Three competing topological hypotheses were uncovered concerning the

relationships among the eastern rainforest sampling locations, but the best possible species

tree estimation and hypothesis testing implied the northern-most sampling location, Riamalandy,

is sister to more southern sampling locations. All phylogenetic evidence suggested that *M.*

lehilahytsara expanded as a ring around the higher elevated regions of the CHS and that the

CHS acted to some extent as a barrier to gene flow during a period of vegetational change and

aridification at the LGM (Burney et al., 2004; Samonds et al., 2019). Although, the northern end

of this barrier may have remained permeable even through climate change, watersheds may

have buffered the region from the aridification that is more evident in the southern extent of the

CHS (Wilmé, Goodman, & Ganzhorn, 2006; Muldoon et al., 2012; Yoder et al., 2016; Samonds

et al., 2019).

Rivers are known to function as barriers to gene flow in mouse lemurs (Olivieri et al., 2008), and

our data suggest they are more significant in impeding mouse lemur migration than open

grasslands. Tsinjoarivo likely represents a population isolated from Ambatovy and Riamalandy

by biogeographic barriers. *M. lehilahytsara* from Tsinjoarivo was previously recognized as

genetically distinct based on a study of few mitochondrial and nuclear loci, although Bayes

factors did not indicate Tsinjoarivo to be an independent lineage under the MSC (Hotaling et al.,

2016). The findings of Hotaling et al. (2016) were recapitulated here with our RADseq data, with

little to no allele sharing between Tsinjoarivo and Ambatovy or Riamalandy (Figure 2;

Supplementary Figure S11). The isolation of Tsinjoarivo from other eastern sampling locations

is plausible, because in addition to geographic distance, our sampling locations are separated

by the Mangoro River (Figure. 2). BPP analyses estimate the time of separation between

Tsinjoarivo and Ambatovy to be near 27 KYA, which again coincides with a time of cool and dry

conditions during the LGM (Scholz et al,. 2011; Clark et al., 2009).

The mosaic of grassland and wooded savanna was permeable by mouse lemurs

Three lines of evidence support the hypothesis that *M. lehilahytsara* had a large ancestral

distribution that covered both the CHS and eastern rainforest escarpment. Within this large

geographic area, population substructure would have been formed due to paleoclimatic

oscillations but with the maintenance of some level of post-divergence gene flow. First, based

on our coalescent estimates of divergence times, the initial divergence between the CHS and

eastern forests would have formed during a period coincident with the LGM (Figure 5 and Table

2). Mean ambient temperatures would have been 4°C cooler and precipitation much lower at

this time (Burney et al., 2004), which may have caused some individuals to retreat towards

watershed refugia (Wilmé et al., 2006) while others remained in the CHS. The divergences

between *M. lehilahytsara* from Ankafobe and Ambohitantely, Ambatovy and Tsinjoarivo, and

their common ancestors occurred near-simultaneously, and this result is not an artifact of prior

choice from Bayesian MSC analyses (Supplementary Figures S18-S24). The pattern of

divergence and population structure formed by reduced gene flow is reflected in θ estimates

from BPP. MSC estimates suggested an increased population size proceeding divergence

events (Table 2), which likely explains why a larger prior on θ resulted in a higher posterior

probability and reduced uncertainty in the divergence time for the MRCA of CHS and eastern *M.*

lehilahytsara.

Second, the strong preference for an *n-islands* model from MIGRATE (Figure 4; Supplementary

Figure S12; Supplementary Table S5) also supported the demographic scenario of populations

that diverged simultaneously from a single larger distribution. Although there is genetic

structure, there is little discernable direction in gene flow, which may explain why models that

incorporated divergence did not perform as well as the *n-islands* model. The migration rates

inferred for our data are arguably high, between 0.8 and 1.2 migrants exchanged per

generation. Such migration rates can reduce fixation indices (e.g. Hartl and Clark, 1989) and

explain the relatively high proportion of admixture in Ambohitantely (Figure 2; Supplementary

Figure S11) and low F_{ST} despite the many generations that must have passed since the initial

divergence between the CHS and eastern forest populations (Supplementary Tables S3 and

S4). Although, migration model parameters may be difficult to identify with many migration rate

parameters (Beerli and Palczewski, 2010) and parameter identifiability is a concern for our short

RAD loci (Supplementary Figures S14-S16) that individually contain very few informative

coalescent events, we are nonetheless confident in these results. Given that we acknowledge

the challenges of model testing and parameter estimation, the *n-islands* model agreed with the

MSC results in that eastern *M. lehilahytsara* has a higher N_e than *M. lehilahytsara* in the CHS.

Consistent with our model-based hypothesis testing, analyses of genetic variation with F_{ST}

suggested that the mosaic of grassland and wooded savanna in the Central Highlands may

have posed as much of a barrier to gene flow as the Mangoro river (Supplementary Table S3;

Figure 2). Relatively little of the genetic variation is explained by grouping populations by the

CHS and eastern forest compared to variation within populations (Supplementary Table S4).

The third piece of evidence for *M. lehilahytsara's* large ancestral distribution is that analyses of

population size recovered similar demographic trends at all four sampling locations analyzed

with EBSP methods. Both the CHS and eastern forests reflect a slight population decrease

before the LGM followed by a recovery through the Holocene (Figure 6; Supplementary Figures

S26-S30). Because we do not see changes in the CHS while the eastern forests remained

stable, population size changes are likely due to global or Madagascar-wide effects rather than

smaller-scale regional changes like agriculture. Although, there are large disparities in N_e

estimates between EBSP and other methods, which raises some concern for interpreting

absolute population size values from molecular data, patterns inferred from the different

analyses are consistent.

Making conservation biology predictions from models of molecular evolution

Here we provided a range of present-day N_e estimates that are sensitive to model (i.e. isolation-

with-migration, MSC, Extended Bayesian Skyline) and prior choice, but all analyses suggest *M.*

lehilahytsara populations sizes are healthy (i.e. > 5000; Lynch and Lande, 1998) and have likely

increased through the Holocene. The most recent IUCN evaluation of *M. lehilahytsara* considers

the species to be vulnerable (Andriaholinirina et al., 2014c) while other mouse lemur species in

the eastern rainforest are considered endangered (Andriaholinirina et al., 2014a;

Andriaholinirina et al., 2014b; Baden et al., 2014). If *M. lehilahytsara* has adaptive traits that

allowed the species to expand its range into the CHS while related species were restricted to

lowland forests, *M. lehilahytsara* may have been more robust to Pleistocene climate change and

even more recent anthropogenic disturbances. *M. lehilahytsara* has been regarded as a

highland specialist (Radespiel et al., 2012) and has been documented to be able to enter a state

of prolonged torpor (Blanco et al., 2017; Andriambeloson et al., 2020). We hypothesize that the

capacity for extended torpor in *M. lehilahytsara* is an adaptive trait that has facilitated its

increased geographic range compared to most other mouse lemur species. Although individual

M. lehilahytsara populations in the CHS have maintained smaller population sizes compared to

those in the eastern forests (Table 2; Figure 6; Supplementary Figure S26), there is increasing

evidence that these populations are robust. For example, the Ankafobe forest patch is less than

one Km^2 yet the resident population is thriving (Blanco et al., in review).

We emphasize, however, that *M. lehilahytsara* in the CHS is not simply some sink population

from an eastern forest source. CHS populations have been evolutionarily independent from the

eastern escarpment for approximately 5000 generations if we consider the lower end of the 95%

HPD intervals of divergence times for nodes A and B in our BPP analyses (Table 2). Even so,

CHS populations have exchanged migrants (Figure 4) with the eastern forests despite the

historically open grassland-woodland mosaic (Solofondranohatra et al., 2018). Indeed, the

occasional migration to and from the east may have been important for sustaining large N_e

despite the small range sizes. Our best migration model estimated an N_e of 13,000 for both

Ankafobe and Ambohitantely and MSC estimates are 19,000 and 31,000 respectively. There

are of course discrepancies in the absolute N_e, but our results suggest that N_e has remained

sufficiently large such that the strength of selection has been strong enough to overcome

fixation of deleterious alleles by drift alone (Lynch & Lande, 1998). Nonetheless, the sustained

lower N_e in the CHS compared to the eastern forests may imply a negative consequence of

continued habitat loss and fragmentation if human activities are not mitigated.

The importance of interpreting biodiversity through evolution

The processes that shaped existing *M. lehilahytsara* population structure must have occurred

before human arrival and mid-Holocene droughts. Although charcoal records (Burney,1987a;

Burney 1987b), stratigraphic pollen occurrence (Gasse & Van Campo, 1998; Burns et al., 2016),

and megafauna subfossil extinctions (Hansford et al., 2018) all point to human-mediated large-

scale ecological changes in the Central Highlands (Virah-Sawmy et al., 2010), our analyses

implied that fundamental *M. lehilahytsara* population divergence coincides with Pleistocene

climate change near the LGM – not after. A similar narrative has been recovered in other

analyses of lemur populations (Quémére, Amelot, Pierson, Crouau-Roy, & Chikhi, 2012;

Salmona et al., 2017) and in an endemic olive tree (Salmona et al. 2019). In summary,

phylogeographic analysis of multiple species and localities agree with ecological (Bond et al.

2008) and evolutionary (Vorontsova et al., 2016; Hackel et al., 2018) studies of grasses that

demonstrate that Malagasy savannas are of ancient origin and have long persisted as a mix of

open grassland and woodland. Although fires due to human activity continue to ravage the

Central Highlands (Burney, 1987a; Gasse & Van Campo, 1998; Virah-Sawmy et al., 2010;

Burns et al., 2016; Samonds et al., 2019), leading to devastating forest loss in Madagascar

(Vieilledent et al., 2018), ongoing habitat fragmentation is occurring on a background of habitat

mosaicism that is likely associated with Pleistocene climate change.

Ethics

Research was conducted under permits N 0233/07/MINENV.EF/SG/DGEF/DPSAP/SSE

(Tsinjoarivo), N 229/14/MEEF/SG/DGF/DCB.SAP/SCB (Ambatovy), and

N225/15/MEEMF/SG/DGF/DAPT/SCBT (Ankafobe) issued by the Ministry of Environment,

Water and Forests of the Malagasy Government.

Data Accessibility

All new sequence data has been made available through NCBI BioProject PRJNA560399. All

individual SRA identifiers are available in Supplementary Table S7. Alignments and other data

for analyses are available from the Dryad Digital Repository (Tiley et al., 2020).

Author Contributions

GPT and ADY conceived the study. MBB, JMR, and RMR performed the fieldwork. ARS and

PAH constructed RADseq libraries. GPT performed analyses. GPT, MBB, and ADY wrote the

manuscript. All authors read and approved the final manuscript.

Competing Interests

The authors declare no competing interests.

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Tables

Table 1- Marginal likelihood estimates and model selection with SNAPP

Steps	Topological Hypothesis	Marginal Likelihood	Bayes Factor[†]	Model Probability
20	RA	-41399.18	0.00	1
	TA	-41847.70	-897.05	1.61×10^{-195}
	RT	-41689.27	-580.19	1.03×10^{-126}
40	RA	-41531.26	0.00	1
	TA	-41848.94	-635.35	1.08×10^{-138}
	RT	-42060.55	-1058.58	1.36×10^{-230}
60	RA	-41328.81	0.00	1
	TA	-41618.87	-580.12	1.07×10^{-126}
	RT	-42057.39	-1457.16	0

[†]Bayes factors computed by marginal log-likelihoods with respect to the best model as $2 * (\ln L_i - \ln L_{Max})$

Table 2 - Absolute divergence times and population sizes estimated[†] from BPP.

Node	N_e (x 10^3)	95% HPD	Divergence time (KYA)	95% HPD
Ankafobe	19.2	17.3, 25.7	-	-
Ambohitantely	31.2	27.1, 40.9	-	-
Tsinjoarivo	23.1	14.0, 26.1	-	-
Ambatovy	55	33.5, 63.0	-	-
Riamalandy	204.5	193.4, 216.0	-	-
A	60.2	10.6, 201.3	26.7	15.9,30.4
B	22	15.1, 34.2	19	16.9, 25.2
C	875.1	776.9, 955.0	31	25.4, 42.7
D	108.3	106.6, 110.1	194.1	189.0, 198.8

[†]Estimates are based on combined posterior for priors $\theta = 0$ and $\tau = 1$.

Figure Legends

Figure 1 – Recovery of RAD loci among *Microcebus lehilahytsara*. The diagonal represents

the proportion of total loci recovered for each sample while the upper triangular represents the

proportion of total loci shared between two individuals. The lower triangular represents the

evolutionary distance between two individuals, which is the patristic distance from the

concatenated ML analysis.

Figure 2 – Well-supported phylogenetic lineages of *Microcebus lehilahytsara* reflect

genetic structure. The phylogeny is the maximum likelihood tree with bootstrap support at

nodes. Branch lengths are expected substitutions per site. Metadata for individual identifiers is

available in Table S1. STRUCTURE results for the one SNP per locus dataset is shown with an

optimal k = 3. The GPS coordinates of each sampling site are shown on the map, with

Riamalandy, Ambatovy, and Tsinjoarivo found in the eastern rainforest while Ambohitantely and

Ankafobe are fragmented forest patches in the Central Highland Savanna. Rivers are shown in

blue while forest cover is shown in green. The specific region of Madagascar studied here

shown in the boxed region on the country map. Major ecoregions of Madagascar are shown on

the country map. Green shows the moist forest while brown is dry forest and red is spiny forest.

Shape files of Madagascar's ecoregions were taken from Vieilledent et al. (2016). All *M.*

lehilahytsara sampling locations are considered wet forest.

Figure 3 – Phylogenetic hypothesis testing of eastern Madagascar. Three competing

topological hypotheses were considered using both the AU test with site likelihoods and Bayes

factors from SNAPP. Posterior probabilities of topologies were obtained from marginal

likelihoods for SNAPP analyses and AU tests perform a multiscale bootstrap of site likelihoods

to obtain a p-value that should be interpreted as strength against a topology. No analogous

topology test exists for invariants, as in SVDquartets, but we did examine the RF distances

among bootstrap trees from ML and SVDquartets analyses to summarize phylogenetic

discordance among RAD loci. Pairwise are all bootstrap trees for ML or SVDquartets analyses

compared with themselves. ML are all bootstrap trees, from either ML or SVDquartets analyses,

compared to the ML tree. SVDq are all bootstrap trees, from either ML or SVDquartets

analyses, compared to the SVDquartets bootstrap consensus tree. Although RF distances

among SVDquartets bootstraps are typically higher, they are also closer to being normally

distributed, implying either less uncertainty or potential biases in ML analyses.

Figure 4 – Migration models reveal gene flow between the Central Highlands and eastern

forests. Bayes factors were used to test migration hypotheses between the eastern rainforest,

Ambatovy (grey), and two fragmented forest patches in the Central Highlands, Ambohitantely

(pink and grey) and Ankafobe (pink). These three locations were selected because of admixture

detected in Ambatovy based on STRUCTURE analyses, and geographic locations are colored

as such. θ parameters are scaled as $4N_e\mu$ and mutation-scaled migration parameters are shown

near arrows. Divergence times are shown for relevant models in substitutions per site.

Figure 5 – Divergence time estimates with BPP. Posterior distributions of MSC parameters

for the combined posterior of prior choice 1. Branch lengths are in coalescent units and error

bars represent 95% highest posterior density intervals around the median node height. Violin

plots are the posterior distributions of node heights for prior choices 1 though 4 to show that

while divergence time estimates are sensitive to prior choice, our biological conclusions are not.

Figure 6 – Population size change over time at sampling locations. Posterior distributions

of θ at discretized bins of τ determined by the number of change points. Only the first sampled

dataset is shown for each population. The blue box shows the duration of the LGM.

Model	Marginal Likelihood	Bayes Factor*	Model Probability
0A – East to west	-408766.23	NA	0
0B – West to east	-408169.12	1194.22	0
1 – Stepping-stone	-404252.47	9027.52	0
2 – n-islands	-402558.14	12416.18	1
3 – Divergence and isolation	-415036.4	-12540.34	0
4 – Divergence and migration	-407777.86	1976.74	0

*Bayes factors computed with natural logarithms of marginal likelihoods with respect to model 0A as 2 * (lnL$_i$ – lnL$_{0A}$)

†θ values are presented as 10^{-3}