Phylogeographic analysis of Goodman's mouse lemur reveals historical interconnectivity

of Madagascar's Central Highlands and eastern rainforests

George P. Tiley[1,†], Marina B. Blanco[1,2], José M. Ralison[3], Rodin M. Rasoloarison[3,4],

Amanda R. Stahlke[5], Paul A. Hohenlohe[5], Anne D. Yoder[1]

[1]Department of Biology, Duke University, Durham, NC 27708, USA

[2]Duke Lemur Center, Duke University, Durham, NC 27705, USA

[3]Département de Biologie Animale, Université d'Antananarivo, BP 906, Antananarivo 101,

Madagascar

[4]Behavioral Ecology and Sociobiology Unit, German Primate Centre, 37077 Göttingen,

Germany

[5]Institute for Bioinformatics and Evolutionary Studies, Department of Biological Sciences,

University of Idaho, Moscow, ID 83844, USA

[†]Author for correspondence: george.tiley@duke.edu

Keywords: Central Highland Savannah, Coalescent, Historical Demography, Fragmentation,

RAD sequencing, Mouse Lemur

Abstract

The Central Highland Plateau of Madagascar is largely composed of savannah, interspersed

with patches of closed-canopy forests. The origins of this landscape are now accepted as long

preceding the arrival of humans in Madagascar, but whether savannah vegetation has remained

static or expanded over time remains a topic of debate. By analyzing multiple populations of

Goodman's mouse lemur (*Microcebus lehilahytsara*), a small-bodied nocturnal primate, we

reconstruct its phylogeographic and demographic history to serve as a proximal estimate of the

time at which the Central Highland and eastern forests became discontinuous. We applied

coalescent methods to RADseq data to infer phylogeographic relationships, population

structure, and migration corridors among sampling sites. Analyses revealed that forest

fragmentation occurred rapidly and affected both the Central Highlands and eastern forests

during a period of decreased precipitation near the last glacial maximum. There was clear

structure between Central Highland populations versus those of the eastern rainforests, but

there has also been historical migration across this presumed gene flow barrier. Findings

support origins of the Central Highland savannah that predate human arrival and demonstrate

that this natural landscape has long impacted the population dynamics of Goodman's mouse

lemur.

Introduction

Madagascar is considered to be a biodiversity hotspot with high levels of species diversity and endemism [1]. Much of this diversity is at risk due to deforestation and forest fragmentation [2-4], which is due at least in part to human activity and agricultural practices [5,6]. But the timing of and degree to which humans have transformed these closely-canopy forests and have affected their connectivity has been a point of contention. Much of central Madagascar is covered with savannah, which was originally thought to be the product of fires from slash-and-burn agriculture and cattle farming [7,8]. The best available archeological evidence places humans in Madagascar between 2 thousand years ago (KYA) and 10 KYA [9-11]. This timeline does coincide with an increased frequency of fire based on charcoal records from sediment cores during the late to mid Holocene [12-14]. Pollen abundances from these same sediment cores imply the Central Highland vegetation transitioned from woody taxa to grasses at this time as well, although the transition likely began following Pleistocene glaciations [13,15] and similar shifts in plant composition were not limited to the Central Highlands [16,17].

Juxtaposed to the hypothesis of rapid human-mediated deforestation, ecological and phylogenetic studies of Madagascar's grasses suggest the Central Highland Savannah (CHS) originated well before the Holocene. Levels of endemism observed in the CHS are comparable to the putatively natural grasslands of Africa [18,19]. Although few C_4 lineages such as *Aristida* and *Loudetia* dominate the CHS [20-22], diversity among endemic C_4 grass clades are more consistent with global diversification of C_4 grasses during the Miocene [e.g 23] rather than many independent migration events [24]. Thus, we are left with a complex natural history where the CHS has ancient origins as a savannah, but has likely experienced deforestation and fragmentation associated with Pleistocene climate change and early human activity.

Elucidating the natural history of Madagascar's CHS is necessary for understanding patterns

and the processes of speciation in many of Madagascar's endemic lineages. For example,

biogeographic barriers to gene flow formed during historical climate change may explain recent

radiations in many of Madagascar's endemic clades [25]. Barriers can be rivers, or open

grasslands in the case of taxa restricted to forested environments, such as mouse lemurs [26].

Demographic studies of mouse lemurs may help develop a more accurate reconstruction of pre-

human conditions in Madagascar and inform conservation research goals. A range of forest

fragmentation hypotheses have been used to evaluate population declines in sifaka populations

throughout the Holocene [27,28], but recognizing a more ancient fragmentation scenario into the

Pleistocene would warrant consideration of additional historic demographic models and the

need to explore data and methods more appropriate for older timescales [29]. Although losses

in genetic diversity of endangered species putatively due to human action are observable [30-

33], a better reconstruction of pre-human conditions can reveal if some populations were

already vulnerable, especially given that some populations have been robust to recent habitat

loss and fragmentation [34].

To evaluate hypotheses regarding scale of fragmentation for the Central Highlands, we leverage

the unique distribution of a mouse lemur, *Microcebus lehilahytsara*. *M. lehilahytsara* is found in

eastern montane forests but is considered a highland specialist that occurs in isolated forest

fragments in the CHS [35,36]. It is the only mouse lemur species known to inhabit forest

fragments in the CHS. The extended range of *M. lehilahytsara* may be due to the ability to enter

a state of prolonged torpor [36]; capable of reducing their metabolism during extreme low

temperatures in the CHS during the winter season. While prolonged torpor or hibernation is

obligatory in *Cheirogaleus*, another member of the family Cheirogaleidae [37,38], mouse

lemurs vary in their expression of daily torpor or hibernation, with some species reported to

display daily torpor (less than 24 hours) exclusively and unable to deposit fat stores to sustain

prolonged torpor [38,39]. Recently, species-level analyses of molecular data that included *M.*

lehilahytsara from the CHS diverged from its sister species *M. mittermeieri* in the eastern

rainforests near 50 KYA [40]; the species status of *M. mittermeieri* has recently been called into

question [41]. Here, we sampled multiple populations of *M. lehilahytsara* from sites in the CHS

and eastern forests to further investigate the natural history of Madagascar's CHS with RADseq

data.

Methods

Study sites and sampling

Microcebus lehilahytsara individuals were captured at five field sites. Three of the locations,

from north to south, Riamalandy, Ambatovy, and Tsinjoarivo are mid-elevation rainforests,

situated along the eastern escarpment. Forests are fragmented along the corridor; although, the

distance between patches has been exacerbated by deforestation since the 1950s [4]. Annual

rainfall is ~1400 mm and daily temperatures vary between ~14 and 23 °C at Riamalandy, while

Tsinjoarivo and Ambatovy, receive 1700-2000 mm of rainfall with temperatures between 15 and

23 °C [42]. The other two locations, Ambohitantely and Ankafobe, are located within the Central

Highlands. Annual rainfall is around 1460 mm, and daily temperatures are between 12.5 and 23

°C [42]. Samples were collected between 2004 and 2016. Mouse lemurs were captured using

live Sherman traps and released back to the forest later the same day [see 36]. Detailed

descriptions of sampling sites are given in the Supplementary Methods (Supplementary

Material).

RADseq library preparation and sequencing

Microcebus lehilahytsara samples were gathered from existing collections and a combination of

genomic DNA from available tissue and previous whole-genome amplifications were used in

order to maximize sampling of these rare and vulnerable species. Library preparations for RAD

sequencing followed Ali et al. [43], using a single-digest protocol with the restriction enzyme

SbfI. A median insert size of 380 bp was selected for paired-end sequencing at the Duke

Sequencing Core on an Illumina Hi-Seq 4000 with an anticipated 15x coverage per individual.

Assembly and orthology evaluation

Data were assembled using STACKS v2.0b [44,45] using *de novo* methods. Final loci output by

STACKS were filtered such that at least three of the 5 sample geographic locations were

represented with at least half of the taxa present. RAD loci were supported by a minimum depth

of three reads; all other assembly and orthology details are the Supplementary Methods

(Supplementary Material).

Phylogenetic analyses

We estimated phylogenies of all *M. lehilahytsara* individuals or species trees of sampling sites

using four approaches: 1) concatenated ML analysis with RAxML v8.2.11 [46], 2) topological

invariants of quartets approximated with SVDquartets [47] implemented through PAUP* v4.0a

build 165 [48], 3) an implementation of the multispecies coalescent (MSC) model for SNP data,

SNAPP v.1.3.0 [49], through BEAST2 v2.4.8 [50], and 4) the MSC with joint gene tree

estimation using BPP v4.0 [51,52]. Phylogenetic hypothesis testing with RAxML and SNAPP

were performed with an approximately unbiased (AU) test [53] and Bayes factors respectively.

Complete details for phylogenetic analyses are in the Supplementary Methods (Supplementary

Material).

Delimiting population structure and testing migration hypotheses

To elucidate migration patterns between *M. lehilahytsara* populations in the CHS and eastern

forests, we developed competing migration hypotheses and tested models with Bayes factors.

We analyzed biallelic SNPs with STRUCTURE v. 2.3.4 [54], using the admixture model and

default priors (Supplementay Material). Pairwise fixation index (F_{ST}) were calculated between

populations in ARLEQUIN v. 3.5.2.2 [55] and an AMOVA analysis [56] was performed for

Central Highland versus eastern populations. Isolation-by-distance was investigated with a

Mantel test. Results were used to develop testable hypotheses that were evaluated with Bayes

factors for isolation-with-migration models in MIGRATE v4.2.14 [57].

Notably, we did not use the default uniform prior for nucleotide diversity (θ) estimates. Instead,

we used independent effective population size (N_e) estimates from three mouse lemur species

[30] to develop a prior distribution on θ (Supplementary Material). θ was rescaled N_e to using a

mutation rate of 1.64×10^{-8}, which is based on a direct germline estimate for mouse lemurs [58].

Two sets of analyses were conducted on phased RAD loci. First, we used all individuals binned

into either a CHS or eastern population. We were limited in the number of loci computationally

feasible for obtaining marginal likelihood estimates, and thus analyzed 100 jackknifed data sets

of 100 RAD loci. The second analysis attempted to differentiate between east-to-west migration

and west-to-east migration among Ambatovy, Ambohitantely, and Ankafobe, and did not allow

for missing data. The three-population analyses used four diploid individuals per population.

Estimating divergence time of eastern and Central Highland mouse lemurs

We used the MSC model for divergence time estimation among *M. lehilahytsara* lineages with

BPP v4.0 [51,52]. We fixed the topology recovered in the majority of analyses here and used

several lines of evidence to support rooting with Riamalandy (Supplementary Material),

including a mitochondrial phylogeny of *M. lehilahytsara* individuals and outgroups (Table S2). A

partitioned ML analysis and bootstrapping was conducted with PAUP* v4.0a build 163 [48].

Phased haploid sequences were used and at least two individuals per population were required

for each locus. We ran eight chains of 2500 samples and evaluated convergence. Final MSC

parameter estimates were based on combined chains. Coalescent branch lengths were

rescaled to absolute time using a germline mutation rate estimate [60] and generation time

estimates from wild [59] and captive [60] populations. Robustness of divergence times to prior

choice was also evaluated (Supplementary Material).

Characterizing population size changes during Pleistocene glaciations

We estimated change in population size over time with extended Bayesian skyline plots [61] in

BEAST2 [50]. For all sampling locations with four or more individuals, we jackknifed 100 RAD

loci eight times, with no missing data. Sequences were treated as phased and haploid. Each

jackknife replicate was run twice to check convergence. Large posteriors were required for

agreement on the number of change points and population size estimates between replicates;

each chain ran for 500 million generations after a 100 million generation burnin while sampling

every 10000 generations. Operators for increasing the sampling of change points followed

recommendations by Trucchi et al. [62]. We used linear piecewise reconstructions of population

size changes.

Results

Circumscribing orthologous RAD loci

All individuals included in our analyses had an average depth of coverage greater than 10x per

locus. In total, 92889 loci were recovered across all taxa with an average of 49201 loci per

individual (Table S1). On average, 31% of loci were shared between pairs of taxa (Figure S1).

The number of loci shared between individuals are largely not due to phylogenetic signal, as

there was only a slight negative correlation between evolutionary distance and the proportion of

shared loci (pearson's r = -0.19; p = 0.001). Rather, this pattern was driven by the number of

reads for each individual (pearson's r = 0.86; p = 3.54e-08). Only 54018 loci had at least one

biallelic SNP.

Support for Central Highland and eastern Microcebus lehilahytsara

All phylogenetic analyses recovered a well-supported split between *M. lehilahytsara* from

Central Highland localities (Ambohitantely and Ankafobe) and eastern forests (Riamalandy,

Ambatovy, and Tsinjoarivo) (Figure 1). But the relationships among eastern sampling locations

was contentious across analyses. The ML, SNAPP, and BPP analyses recovered Ambatovy as

sister to Tsinjoarivo (H_{TA}; Figures S2- S5); although, the posterior probability from SNAPP was

weak. SVDquartets strongly conflicted the H_{TA} topology, favouring Riamalandy sister to

Tsinjoarivo (H_{RT}) when analyzing all variable sites (Figure S6) or complete RAD loci (Figure S7).

However, the H_{RT} topology was sensitive to the underlying data. Analyses of biallelic SNPs

alone with SVDquartets recovered a third well-supported topology (H_{RA}; Figure S8). Although in

one out of ten resampled datasets, when treating all individuals as independent lineages,

Ambatovy was not recovered as a monophyletic clade. Topological hypothesis testing

differentiated among the three recovered phylogenetic hypotheses: H_{TA}, H_{RT}, and H_{RA}.

AU tests rejected H_{RT}, but could not reject H_{RA} despite H_{TA} being present in the majority of ML

bootstrap trees (Figure S9). Bayes factors from SNAPP provided insights into the prevalence of

both H_{RA} and H_{TA} hypotheses across analyses. SNAPP supported lumping Riamalandy and

Ambatovy together as opposed to lumping Ambatovy and Tsinjoarivo (Table 1). This result is

surprising, considering H_{TA} was the maximum clade credibility tree from the posterior sample in

the unconstrained analyses (Figures S3 and S4). All phylogenetic evidence suggested a shared

common ancestor for Ambatovy and Tsinjoarivo, but Tsinjoarivo may be more evolutionarily

distant than expected. Phylogenetic analyses showed strong support for both CHS and eastern

clades, and at least no very recent gene flow as evident by monophyly of our five sampling

locations. However, such analyses do not rule out historical gene flow between the CHS and

eastern forests.

Migration between eastern and Central Highland forests

There was admixture between eastern and Central Highland *M. lehilahytsara* as well as

population structure within the eastern forests that mirrored SNAPP Bayes factors (Figure 1).

STRUCTURE analyses recovered an optimal *k* = 3 in 19 out of 20 jackknifed datasets (Figures

S10-S12). The remaining replicate recovered an optimal *k* = 2 (Figure S11). All analyses

supported a clustering of Central Highland and eastern *M. lehilahytsara*, with some degree of

admixture between the two. The Ambohitantely site largely clusters with Ankafobe, but

individuals may share between 10% and 40% of their alleles with Ambatovy and Riamalandy.

Tsinjoarivo often forms its own unique cluster, except in the one jackknife replicate where *k* = 2.

Admixture between Central Highland and eastern *M. lehilahytsara* was also supported by

analyses of genetic variation with F_{ST} statistics. Pairwise F_{ST} revealed similar levels of genetic

divergence between Ambatovy and Ambohitantely to that of Ambatovy and Tsinjoarivo, despite

the more contiguous forest among eastern sampling locations (Table S3). The geographic

structure between Central Highland and eastern *M. lehilahytsara* explains a small fraction of

genetic variation, with most variation explained within populations (Table S4). A Mantel test

revealed little evidence for isolation-by-distance (pearson's *r* = 0.09; *p* = 0.39), suggesting

barriers to gene flow aside distance alone (Figure S13).

Bayes factors were computed for different isolation-with-migration models to further investigate

the patterns of admixture observed between the Ambatovy, Ambohitantely, and Ankafobe

genetic clusters. The *n-islands* model was unambiguously the best model for both analyses that

included all individuals modeled as two populations (Figure S14; Table S5) or three populations

(Figure 2). There were no large discrepancies in the estimated migration rates; however, the

selected model suggested a lower N_e in Ankafobe and Ambohitantely compared to Ambatovy.

Assuming a mutation rate of 1.64 x 10^{-8}, MIGRATE estimated contemporary population sizes to

be 12652 for Ankafobe and Ambohitantely, and 17226 for Ambatovy. Their N_e estimates,

although influenced by the prior, are not driven by the prior alone (Figures S15-S17).

Additionally, a higher N_e in Ambatovy is supported across all models regardless of the migration

and divergence scenarios (Figure 2). Because of Ambatovy's higher effective population size,

the number of migrants leaving towards Ambohitantely or Ankafobe per generation is

approximately 1.2, compared to 0.8 effective migrants per generation leaving Ambohitantely or

Ankafobe towards Ambatovy.

Dating Population Divergences

Using the H_{TA} topology supported by Bayes factor analyses, divergence times were estimated

using the MSC model. The topology was rooted with Riamalandy based on the analysis of

mitochondrial data from *Microcebus lehilahytsara* and outgroup taxa (Figure S18) and inference

of clock-like trees from BPP (Figure 3). Most MSC parameters converged across eight chains

with potential scale reduction factors approaching 1 (Table S6). A few parameters still retained

high variation between chains for both θ and τ, but this discrepancy was largely due to one or

two chains with inefficient mixing for different prior combinations (Figure S19-S26). Divergence

times were not strongly influenced by the priors (Figure 3) and implied that divergence between

the CHS and eastern forest, as well as the forest patches within them, occurred almost

simultaneously. Absolute estimates inferred that divergence among our sampled lineages

occurred near 31 KYA (Table 2).

Population declines in the CHS

Population sizes experienced some slight contractions near 25 KYA (Figure 4) that roughly

coincide with the divergence of sampling locations (Table 2). Analyses also suggested an uptick

in population size through the Holocene until the present. Ambatovy had consistently higher N_e

estimates across jackknife replicates. Ankafobe had the lowest N_e estimates. Intermediate N_e

estimates were observed for individuals at Ambohitantely and Tsinjoarivo, despite that

Ambohitantely is a more fragmented habitat in the CHS. There is high uncertainty in N_e based

on comparisons across jackknife replicates (Figure S27), but individual analyses tended to

converge (Figures S28-S31).

Discussion

Population structure of M. lehilahytsara predates human colonization of Madagascar

Our results provide additional evidence in favour of the now well-accepted view that

Madagascar's Central Highlands are an ancient mosaic of grassland and wooded savannah

[18,19], while elucidating the evolution of a uniquely distributed mouse lemur. Phylogenetic

analyses revealed an unambiguous split between CHS and eastern forest sampling locations

(Figure 1; Figures S2-S8), which was also supported by analyses of genetic structure (Figure 1;

Figure S12). Three competing topological hypotheses were uncovered concerning the

relationships among our eastern rainforest sampling locations, but the northern-most sampling

location, Riamalandy, is likely sister to more southern sites. Phylogenetic evidence suggested

that *M. lehilahytsara* dispersed as a ring around the higher elevation regions of the CHS, and

that the CHS acted to some extent as a barrier to gene flow during a period of vegetational

change and aridification through the LGM [16,63]. The northern end of this barrier may have

remained permeable since watersheds are hypothesized to have buffered the region from

aridification more evident in the southern extent of the CHS [25,41,63,64].

Tsinjoarivo is a population isolated by biogeographic barriers from Ambatovy and Riamalandy.

M. lehilahytsara from Tsinjoarivo was previously recognized as genetically distinct based on a

study of few mitochondrial and nuclear loci, although Bayes factors did not indicate Tsinjoarivo

to be an independent lineage under the MSC model compared to other *M. lehilahytsara* [65].

The findings of Hotaling et al. [65] were recapitulated here, with little to no admixture between

Tsinjoarivo and Ambatovy or Riamalandy (Figure 1; Figure S12). The isolation of Tsinjoarivo

from other eastern sampling locations is plausible, because it is separated by the Mangoro

River. Our BPP analyses date the separation of Tsinjoarivo from Ambatovy near 27 KYA, which

again coincides with a time of cool and dry conditions during the LGM [66,67]. Interpreting

absolute dates is tenuous at best, especially given these ages contradict the hypothesis that

riverine barriers formed during times of increased precipitation [25], but Madagascar is

ecologically complex and there is likely no single causal factor that shaped the distribution of *M.*

lehilahytsara. Our divergence time estimates may be indicative of more widespread forest

fragmentation and vegetative turnover linked to paleoclimatic oscillations.

The mosaic of grassland and wooded savannah was not impermeable by mouse lemurs

Three lines of evidence support that *M. lehilahytsara* had a large ancestral distribution that

covered both the CHS and eastern rainforest escarpment, and that the two were

interconnected. This large ancestral distribution underwent some contraction and developed

population substructure during a period coincident with the LGM. One is the successive

divergence times of the CHS and eastern forest populations (Figure 3 and Table 2). The

divergences between mouse lemurs from Ankafobe and Ambohitantely, Ambatovy and

Tsinjoarivo, and their common ancestors occurred near-simultaneously, and this result is not an

artifact of prior choice (Figures S19-S25). The pattern of divergence and population structure

formed by reduced gene flow is reflected in θ estimates from BPP. MSC estimates suggested a

large population size proceeding divergence events (Table 2), which likely explains why a larger

prior on θ resulted in a higher posterior probability and reduced uncertainty in the divergence

time for the MRCA of CHS and eastern *M. lehilahytsara*.

The strong preference for an *n-islands* model from MIGRATE (Figure 2; Figure S13; Table S5)

also supported the scenario of populations that diverged simultaneously from a single larger

distribution. There was little discernable direction in gene flow, which may explain why models

that incorporated divergence did not perform as well as the *n-islands* model. The migration rates

inferred for our data are arguably high, between 0.8 and 1.2 migrants exchanged per

generation. Such migration rates could explain the admixture in Ambohitantely (Figure 1; Figure

S12) and low F_{ST}, despite the many generations that must have passed (Tables S3 and S4).

Although, parameter identifiability is a concern for our parameter-rich models [57], especially

given that our short RAD loci individually contain very few informative coalescent events

(Figures S15-S17). That caveat in mind, the *n-islands* model agreed with the MSC results in that

eastern *M. lehilahytsara* has a higher N_e than *M. lehilahytsara* in the CHS. Analyses of genetic

variation with F_{ST} also suggested that the mosaic of grassland and wooded savannah in the

Central Highlands at least does not pose a more significant barrier to gene flow than the

Mangoro river (Table S3). Even with its close proximity to Ambohitantely and historical gene

flow across the Central Highlands, Ankafobe is genetically distant compared to eastern forest

populations (Figure S13). The increased genetic distance is possibly explained by the small

amount of suitable habitat (less than 1 Km^2) and inbreeding.

The third piece of evidence is that analyses of population size change over time share some

patterns with the MSC and isolation-with-migration model analyses (Figure 4; Figures S27-S31).

We found similar demographic trends among the four populations analyzed with EBSP

methods, and all displayed a slight population decrease before the LGM followed by a recovery.

These patterns roughly reflect N_e estimates from MSC analyses and the similar demographic

histories among populations would also agree with our hypothesis that *M. lehilahystara* had a

single widespread population that experienced fragmentation. Although, there are large

disparities in N_e estimates between EBSP and other methods, which raises concern for

interpreting absolute population size values from molecular data.

Limitations of RADseq and demographic interpretations

Scrutiny is needed in reconciling our results from various coalescent analyses, as we are

analyzing short RAD loci that individually carry little information about demographic history

(Figures S14-S17). And notably, no migration is assumed after divergence in the MSC model

implemented here; however, we expect estimation errors from unaccounted migration to make

divergence times older [40,68,69]. Therefore, the processes that shaped existing *M.*

lehilahytsara population structure must have occurred before human arrival and mid-Holocene

droughts. Although charcoal records [12,15], stratigraphic pollen occurrence [13,70], and

megafauna subfossils [11] all point to human-mediated large-scale ecological changes in the

Central Highlands [14], our analyses implied that *M. lehilahytsara* population divergences

coincide with Pleistocene climate change near the LGM. A similar narrative has been recovered

in other analyses of lemur populations [27,28] and an endemic olive tree [71]. Therefore, these

analyses represent first steps in revealing the demographic history of *M. lehilahytsara*, while

also contributing the evolving natural history of Madagascar's Central Highlands [18,19,22,42].

Making conservation biology predictions from models of molecular evolution

It is difficult to evaluate our N_e estimates in the context of conservation biology since comparable

numbers to other mouse lemur species are not available, and here we provided a range of N_e

estimates that are sensitive to model (i.e. isolation-with-migration, MSC, Extended Bayesian

Skyline) choice. The most recent IUCN evaluation of *M. lehilahytsara* considers the species to

be vulnerable [72] while geographic neighbors in the eastern rainforest are considered

endangered [31-33]. If *M. lehilahytsara* has adaptive traits that allowed the species to expand its

range into the CHS while related species were restricted to lowland forests, *M. lehilahytsara*

may have been more robust to Pleistocene climate change and even more recent

anthropogenic disturbances. We hypothesize that torpor in *M. lehilahytsara* [36] has allowed its

increased range, compared to other mouse lemurs. Although, *M. lehilahytsara* populations in

the CHS have maintained smaller population sizes compared to those in the eastern forests

(Table 2; Figure 4; Figure S27). These differences in population size are expected, given that

the connectivity and area of available habitat is reduced. For example, the Ankafobe forest

patch is less than 1 Km^2. CHS areas are also subjected to frequent wildfires, which easily

spread across secondary grasslands and destroy isolated forest fragments [42].

M. lehilahytsara in the CHS is not simply a sink population from an eastern forest source. CHS

populations have been evolutionarily independent from the eastern escarpment for

approximately 5000 generations considering the lower end of the divergence time 95% HPD

intervals for nodes A and B (Table 2). After divergence, CHS populations still exchanged

migrants (Figure 2) with the eastern forests despite the historic mosaic of grassland and

wooded savannah [22]. The geographic isolation with occasional migration to and from the east

may have been important for sustaining large N_e despite the relatively small habitable areas.

Our best migration model estimated an N_e of 13000 for both Ankafobe and Ambohitantely and

MSC estimates are 19000 and 31000 respectively. There are discrepancies in the absolute N_e,

but our results suggest that N_e has remained sufficiently large such that the strength of selection

was strong enough to overcome fixation of deleterious alleles by drift alone [73]. The sustained

lower N_e in the CHS compared to the eastern forests does imply a negative consequence of

habitat loss and fragmentation, and that a loss of CHS populations could put *M. lehilahytsara* at

risk, similar to closely related species in the eastern rainforest [40].

Mouse lemurs are iconic of Madagascar's endemism and diversity, but they can also be a

valuable system for reconstructing the island's complex natural history. Molecular data from *M.*

lehilahytsara has bracketed population divergences, which we interpret as at least some forest

loss and fragmentation in the CHS and eastern rainforest escarpment, to late Pleistocene. We

interpret the phylogenetic hypothesis, demographic inference, and dating as rapid fragmentation

during the LGM. Additional comparative work among populations of multiple species may further

refine our reconstruction of the CHS biogeography while also informing the demographic history

of mouse lemurs and their conservation.

Ethics

Research was conducted under permits N 0233/07/MINENV.EF/SG/DGEF/DPSAP/SSE, N

229/14/MEEF/SG/DGF/DCB.SAP/SCB, and N225/15/MEEMF/SG/DGF/DAPT/SCBT issued by

the Ministry of Environment, Water and Forests of the Malagasy Government.

Data Accessibility

New sequence data is available through NCBI BioProject PRJNA560399. Individual SRA

identifiers are in Table S7. Alignments and configuration files are available from the Dryad

Digital Repository: XXXX

Author Contributions

GPT and ADY conceived the study. MBB, JMR, and RMR performed the fieldwork. ARS and

PAH constructed RADseq libraries. GPT performed analyses. GPT, MBB, and ADY wrote the

manuscript.

Competing Interests

The authors declare no competing interests.

Funding

The research was supported by Duke Tropical Conservation Initiative Grant to ADY, CI/Primate

Action Fund and Duke Lemur Center/SAVA Conservation research funds to MBB, and Duke

University research funds to ADY. ADY would also like to acknowledge support from the John

Simon Guggenheim Memorial Foundation and the Alexander von Humboldt Foundation.

Acknowledgements

This is Duke Lemur Center publication #XXXX.

References

1. Myers N, Mittermeier RA, Mittermeier CG, da Fonseca GAB, Kents J. 2000 Biodiversity hotspots for conservation priorities. *Nature* **403**:853-858.

2. Green GM, Sussman RW. 1990. Deforestation history of the eastern rain forests of Madagascar from satellite images. *Science* **248**:212-215.

3. Harper GJ, Steininger MK, Tucker CJ, Juhn D, Hawkins F. 2007. Fifty years of deforestation and forest fragmentation in Madagascar. *Environmental Conservation* **34**:325-333.

4. Vieilledent G, Grinand C, Rakotomalala FA, Ranaivosoa R, Rakotoarijaona J-R, Allnutt TF, Achard F. 2018 Combining global tree cover loss data with historical national forest cover maps to look at six decades of deforestation and forest fragmentation in Madagascar. *Biological Conservation* **222**:189-197.

5. Farris ZJ, Gerber BD, Valenta K, Rafaliarison R, Razafimahaimodison JC, Larney E, Rajaonarivelo T, Randriana Z, Wright PC, Chapman CA. 2017 Threats to a rainforest carnivore community: A multi-year assessment of occupancy and co-occurrence in Madagascar. *Biological Conservation* **210**:116-124.

6. Styger E, Rakotondramasy HM, Pfeffer MJ, Fernandes ECM, Bates DM. 2007 Influence of slash-and-burn farming practices on fallow succession and land degradation in the rainforest region of Madagascar. *Agriculture, Ecosystems, and Environment* **119**:257-269.

7. Perrier de la Bâthie H. 1921 La vegetation Malgache. *Annales du Musée Colonial de Marseille* **9**:1-226.

8. Humbert H. 1927 La destruction d'une flore insulaire par le feu. Principaux aspects de la vegetation à Madagascar. *Mémoires de L'Académie Malgache* **5**:1-78.

9. Dewar RE, Wright HT. 1993 The culture history of Madagascar. *Journal of World Prehistory* **7**:417-466.

10. Dewar RE, Radimilahy C, Wright HT, Jacobs Z, Kelly GO, Berna F. 2013 Stone tools and foraging in northern Madagascar challenge Holocene extinction models. *Proc Natl Acad Sci USA.* **110**:12583-12588.

11. Hansford J, Wright PC, Rasoamiaramanana A, Pérez VR, Godfrey LR, Errickson D, Thompson T, Turvey ST. 2018 Early Holocene human presence in Madagascar evidenced by exploitation of avian megafauna. *Science Advances* **4**:eaat6925.

12. Burney DA. 1987a Late Holocene vegetational change in central Madagascar. *Quatern Res.* **28**:130-143.

13. Gasse F, Van Campo E. 1998 A 40,000-yr pollen and diatom record from Lake Tritrivakely, Madagascar, in the southern tropics. *Quatern Res.* **49**:299-311.

14. Virah-Sawmy M, Willis KJ, Gillson L. 2010 Evidence for drought and forest declines during the recent megafaunal extinctions in Madagascar. *J Biogeogr.* **37**:506-519.

15. Burney DA. 1987b Late Quaternary stratigraphic charcoal records from Madagascar.
Quatern Res. **28**:274-280.

16. Burney DA, Burney LP, Godfrey LR, Jungers WL, Goodman SM, Wright HT, Jull AJT. 2004
A chronology for late prehistoric Madagascar. *Journal of Human Evolution* **47**:25-63.

17. Virah-Sawmy M, Willis KJ, Gillson L. 2009 Threshold response of Madagascar's littoral
forest to sea-level rise. *Global Ecology and Biogeography* **18**:98-110.

18. Bond WJ, Silander Jr JA, Ranaivonasy J, Ratirarson J. 2008 The antiquity of Madagascar's
grasslands and the rise of C_4 grassy biomes. *J Biogeogr.* **35**:1743-1758.

19. Vorontsova MS, Besnard G, Forest F, Malakasi P, Morat K, Clayton WD, Ficinski P, Savva
GM, Nanjarisoa OP, Razanatsoa J, et al. 2016 Madagascar's grasses and grasslands:
anthropogenic or natural? *Proc R Soc B.* **283**:20152226.

20. Koechlin J. 1993 Natural grasslands: eastern hemisphere and resume. In: Coupland RT
editor. Grasslands of Madagascar. Amsterdam: Elsevier. p. 291-301.

21. Kull CA. 2003 Fire and the management of highland vegetation. In: Goodman SM,
Benstead JP, editors. The natural history of Madagascar. Chicago: The University of Chicago
Press. p. 153-157.

22. Solofondranohatra CL, Vorontsova MS, Hackel J, Besnard G, Cable S, Williams J,
Jeannoda V, Lehmann CER. 2018 Grass functional traits differentiate forest and savanna in the
Madagascar Central Highlands. *Frontiers in Ecology and Evolution* **6**:184.

23. Edwards EJ, Smith SA. 2010 Phylogenetic analyses reveal the shady history of C_4 grasses.
Proc Natl Acad Sci USA. **107**:2532-2537.

24. Hackel J, Vorontsova M, Nanjarisoa O, Hall R, Razanatsoa J, Malakasi P, Besnard G. 2018
Grass diversification in Madagascar: *in situ* radiation of two large C_3 shade clades and support
for a Miocene to Pliocene origin of C_4 grassy biomes. *J Biogeogr.* **45**:750-761.

25. Wilmé L, Goodman SM, Ganzhorn JU. 2006 Biogeographic evolution of Madagascar's
microendemic biota. *Science* **312**:1063-1065.

26. Olivieri G, Zimmerman E, Randrianambinina B, Rasoloharijaona S, Rakotondravony D,
Guschanski K, Radespiel U. 2007. The ever increasing diversity in mouse lemurs: Three new
species in north and northwestern Madagascar. *Mol Phylogenet Evol.* **43**:309-327.

27. Quémére E, Amelot X, Pierson J, Crouau-Roy B, Chikhi L. 2012 Genetic data suggest a
natural prehuman origin of open habitats in northern Madagascar and question the deforestation
narrative in this region. *Proc Natl Acad Sci USA.* **109**:13028-13033.

28. Salmona J, Heller R, Quémére E, Chikhi L. 2017. Climate change and human colonization
triggered habitat loss and fragmentation in Madagascar. *Mol Ecol.* **26**:5203-5222.

29. Yoder AD, Poelstra JW, Tiley GP, Williams RC. 2018 Neutral theory is the foundation of
conservation genetics. *Mol Biol Evol.* **35**:1322-1326.
530

531   30. Olivieri GL, Sousa V, Chikhi L, Radespiel U. 2008 From genetic diversity and structure to
532   conservation: Genetic signature of recent population declines in three mouse lemur species
533   (*Microcebus* spp.). *Biological Conservation* **141**:1257-1271.
534
535   31. Andriaholinirina N, Baden A, Blanco M, Chikhi L, Cooke A, Davies N, Dolch R, Donati G,
536   Ganzhorn J, Golden C, et al. 2014 *Microcebus danfossi. The IUCN Red List of Threatened*
537   *Species* 2014a: e.T136347A16112905. http://dx.doi.org/10.2305/IUCN.UK.2014-
538   1.RLTS.T136347A16112905.en. Downloaded on 25 November 2018.
539
540   32. Andriaholinirina N, Baden A, Blanco M, Chikhi L, Cooke A, Davies N, Dolch R, Donati G,
541   Ganzhorn J, Golden C, et al. 2014 *Microcebus bongolavensis. The IUCN Red List of*
542   *Threatened Species* 2014b: e.T136537A16112834. http://dx.doi.org/10.2305/IUCN.UK.2014-
543   1.RLTS.T136537A16112834.en. Downloaded on 25 November 2018.
544
545   33. Baden A, Blanco M, Chikhi L, Cooke A, Davies N, Dolch R, Donati G, Ganzhorn J, Golden
546   C, Groeneveld LF, et al. 2014 *Microcebus ravelobensis. The IUCN Red List of Threatened*
547   *Species* 2014: e.T39751A16113536. http://dx.doi.org/10.2305/IUCN.UK.2014-
548   1.RLTS.T39751A16113536.en. Downloaded on 25 November 2018.
549
550   34. Sgarlata GM, Salmona J, Alexio-Pais I, Rakotonanahary A, Sousa AP, Kun-Rodrigues C,
551   Ralantoharijaona T, Jan F, Zaranaina R, Rasolondraibe E, Zaonarivelo JR, Andriaholinirina NV,
552   Chikhi L. 2018 Genetic differentiation and demographic history of the northern rufous mouse
553   lemur (*Microcebus tavaratra*) across a fragmented landscape in northern Madagascar.
554   *International Journal of Primatology* **39**:65-89.
555
556   35. Radespiel U, Ratsimbazafy JH, Rasoloharijaona S, Raveloson H, Andriaholinirina N,
557   Rakotondravony R, Randrianarison RM, Randrianambinina B. 2012 First indications of a
558   highland specialist among mouse lemurs (*Microcebus* spp.) and evidence for a new mouse
559   lemur species from eastern Madagascar. *Primates* **53**:157-170.
560
561   36. Blanco MB, Andriantsalohimisantatra AA, Rivoharison TV, Andriambeloson J-B. 2017
562   Evidence of prolonged torpor in Goodman's mouse lemurs at Ankafobe forest, central
563   Madagascar. *Primates* **58**:31-37.
564
565   37. Dausmann KH, Glos J, Ganzhorn JU, Heldmaier G. 2005 Hibernation in the tropics: lessons
566   from a primate. *Journal of Comparative Physiology B*. **175**:147-155.
567
568   38. Blanco MB, Dausmann KH, Faherty SL, Yoder AD. 2018. Tropical heterothermy is "cool":
569   The expression of daily torpor and hibernation in primates. *Evolutionary Anthropology* **27**: 147-
570   161.
571
572   39. Schmid J. 2001. Daily torpor in free-ranging gray mouse lemurs (*Microcebus murinus*) in
573   Madagascar. International *Journal of Primatology* **22**:1021-1031.
574
575   40. Shußler D, Salmona J, Blanco MB, Poelstra J, Tiley GP, Andriambeloson JB, Bouchez O,
576   Campbell CR, Etter PD, Iribar A, et al. 2019 Complex processes of cryptic speciation in mouse
577   lemurs from a micro-endemism hotspot in Madagascar. *bioRxiv* doi:
578   https://doi.org/10.1101/742361.
579
41. Yoder AD, Campbell CR, Blanco MB, dos Reis M, Ganzhorn JU, Goodman SM, Hunnicutt
KE, Larsen PA, Kappeler PM, Rasoloarison RM, et al. 2016 Geogenetic patterns in mouse

lemurs (genus *Microcebus*) reveal the ghosts of Madagascar;s forests past. *Proc Natl Acad Sci*
USA. **113**:8049-8056.

42. Goodman SM, Raherilalao MJ, Wohlauser S. 2018 The terrestrial protected areas of
Madagascar: Their history, description, and biota. Association Vahatra. Antananarivo,
Madagascar.

43. Ali OA, O'Rourke SM, Amish SJ, Meek MH, Luikart G, Jeffres C, Miller MR. 2016 RAD
Capture (Rapture): Flexible and efficient sequence-based genotyping. *Genetics* **202**:389-400.

44. Catchen J, Amores A, Hohenlohe P, Cresko W, Postlethwait J. 2011 Stacks: building and
genotyping loci de novo from short-read sequences. *G3.* **1**:171-182.

45. Catchen J, Hohenlohe P, Bassham S, Amores A, Cresko W. 2013 Stacks: an analysis tool
set for population genomics. *Mol Ecol.* **22**:3124-3140.

46. Stamatakis A. 2014 RAxML version 8: a tool for phylogenetic analysis and post-analysis of
large phylogenies. *Bioinformatics* **30**:1312-1313.

47. Chifman J, Kubatko L. 2014 Quartet inference from SNP data under the coalescent model.
Bioinformatics **30**:3317-3324.

48. Swofford DL. 2003 PAUP*. Phylogenetic Analysis Using Parsimony (*and Other Methods).
Version 4. Sinauer Associates, Sunderland, Massachusetts.

49. Bryant D, Bouckaert R, Felsenstein J, Rosenberg N, RoyChoudhury A. 2012 Inferring
species trees directly from biallelic genetic markers: bypassing gene trees in a full coalescent
analysis. Mol Biol Evol. 29:1917-1932.

50. Bouckaert RR, Heled J, Kuehnert D, Vaughan TG, Wu C-H, Xie D, Suchard MA, Rambaut
A, Drummond SJ. 2014 BEAST 2: A software platform for Bayesian evolutionary analysis. *PLoS*
Comput Biol. **10**:e1003537. doi: 10.1371/journal.pcbi.1003537.

51. Rannala B, Yang Z. 2003 Bayes estimation of species divergence times and ancestral
population sizes using DNA sequences from multiple loci. *Genetics* **164**:1645-1656.

52. Flouri T, Jiao X, Rannala B, Yang Z. 2018 Species tree inference with BPP using genomic
sequences and the multispecies coalescent. *Mol Biol Evol.* **35**:2585-2593.

53. Shimodaira H. 2004 Approximately unbiased tests of regions using multistep-multiscale
bootstrap resampling. *Annals of Statistics* **32**:2616-2641.

54. Pritchard JK, Stephens M, Donnelly P. 2000 Inference of population structure using
multilocus genotype data. *Genetics* **155**:945-959.

55. Excoffier L, Lischer HEL. 2010. Arlequin suite ver 3.5: A new series of programs to perform
population genetics analyses under Linux and Windows. *Mol Ecol Resour.* **10**: 564-567.

56. Excoffier L, Smouse PE, Quattro JM. 1992. Analysis of molecular variance inferred from
metric distances among DNA haplotypes: Application to human mitochondrial DNA restriction
data. *Genetics* **131**:479-491.

633
634 57. Beerli P, Palczewski M. 2010 Unified framework to evaluate panmixia and migration
635 direction among multiple sampling locations. *Genetics* **185**:313-326.
636
637 58. Campbell CR, Tiley GP, Poelstra JW, Hunnicutt KE, Larsen PA, dos Reis M, Yoder AD.
638 2019. Pedigree-based measurement of the do novo mutation rate in the gray mouse lemur
639 reveals a high mutation rate, few mutations in CpG sites, and a weak sex bias. *bioRxiv* doi:
640 https://doi.org/10.1101/724880.
641
642 59. Radespiel U., Lutermann H., Schmelting B., Zimmermann E. 2019 An empirical estimate of
643 1253 the generation time of mouse lemurs. *Am. J. Primatol.* **81**:e23062.
644
645 60. Zohdy S, Gerber BD, Tecot S, Blanco MB, Winchester JM, Wright PC, Jernvall J. 2014
646 Teeth, sex, and testosterone: aging in the world's smallest primate. *PLoS One* **9**:e109528.
647
648 61. Heled J, Drummond AJ. 2008 Bayesian inference of population size history from multiple
649 loci. *BMC Evol Biol.* **8**:289.
650
651 62. Trucchi E, Gratton P, Whittington JD, Cristofari R, Le Maho Y, Stenseth NC, C Le Bohec.
652 2014 King penguin demography since the last glaciation inferred from genome-wide data. *Proc*
653 *R Soc B.* **281**:20140528.
654
655 63. Samonds KE, Crowley BE, Rasolofomanana TRN, Andriambelomanana MC,
656 Andrianavalona HT, Ramihangihajason TN, Rakotozandry R, Nomenjanahary ZB, Irwin MT,
657 Wells NA, et al. 2019 A new late Pleistocene subfossil site (Tsaramody, Sambaina basin,
658 central Madagascar) with implications for the chronology of habitat and megafaunal community
659 change on Madagascar's Central Highlands. *Journal of Quaternary Science* **34**:379-392.
660
661 64. Muldoon KM, Crowley BE, Godfrey LR, Rasoamiaramanana A, Aronson A, Jernvall J,
662 Wright PC, Simons EL. 2012. Early Holocene fauna from a new subfossil site: A first
663 assessment from Christmas River, south central Madagascar. *Madagascar Conservation and*
664 *Development* **7**:23-29.
665
666 65. Hotaling S, Foley ME, Lawrence NM, Bocanegra J, Blanco MB, Rasoloarison R, Kappeler
667 PM, Barrett MA, Yoder AD, Weisrock DW. 2016 Species discovery and validation in a cryptic
668 radiation of endangered primates: coalescent-based species delimitation in Madagascar's
669 mouse lemurs. *Mol Ecol.* **25**:2029-2045.
670
671 66. Scholz CA, Cohen AS, Johnson TC, King J, Talbot MR, Brown ET. 2011 Scientific drilling in
672 the great rift valley: The 2005 Lake Malawi scientific drilling project – an overview of the past
673 145,000 years of climate variability in southern hemisphere East Africa. *Palaeogeography,*
674 *Palaeoclimatology, Palaeoecology* **303**:3-19.
675
676 67. Clark PU, Dyke AS, Shakun JD, Carlson AE, Clark J, Wohlfarth B, Mitrovica JX, Hostetler
677 SW, McCabe AM. 2009 The last glacial maximum. *Science* **325**:710-714.
678
679 68. Cruickshank TE, Hahn MW. 2014. Reanalysis suggests that genomic islands of speciation
680 are due to reduced diversity, not reduced gene flow. *Mol Ecol.* **23**:3133-3157.
681
682 69. Leaché AD, Harris RB, Rannala B, Yang Z. 2014. The influence of gene flow on species
683 tree estimation: a simulation study. *Syst Biol.* **63**:17-30.

684
685 70. Burns SJ, Godfrey LR, Faina P, McGee D, Hardt B, Ranivoharimanana L, Randrianasy J.
686 2016. Rapid human-induced landscape transformation in Madagascar at the end of the first
687 millennium of the common era. *Quaternary Science Reviews* **134**:92-99.
688
689 71. Salmona J, Olofsson JK, Hong-Wa C, Razanatsoa J, Rakotonasolo F, Ralimanana H,
690 Randriamboavonjy T, Suescun U, Vorontsova MS, Besnard G. 2020. Late Miocene origin and
691 recent population collapse of the Malagasy savanna olive tree. *Biol J Linn Soc.* **129**:227-243.
692
693 72. Andriaholinirina N, Baden A, Blanco M, Chikhi L, Cooke A, Davies N, Dolch R, Donati G,
694 Ganzhorn J, Golden C, et al. 2014. *Microcebus lehilahytsara*. *The IUCN Red List of Threatened
695 Species* 2014c: e.T136199A16113109. http://dx.doi.org/10.2305/IUCN.UK.2014-
696 1.RLTS.T136199A16113109.en. Downloaded on 28 November 2018.
697
698 73. Lynch M, Lande R. 1998. The critical effective size for a genetically secure population.
699 Animal Conservation 1:70-72.

700   **Tables**

Table 1- Marginal likelihood estimates and model selection with SNAPP

Steps	Topological Hypothesis	Marginal Likelihood	Bayes Factor[†]	Model Probability
20	RA	-41399.18	0.00	1
	TA	-41847.70	-897.05	1.61×10^{-195}
	RT	-41689.27	-580.19	1.03×10^{-126}
40	RA	-41531.26	0.00	1
	TA	-41848.94	-635.35	1.08×10^{-138}
	RT	-42060.55	-1058.58	1.36×10^{-230}
60	RA	-41328.81	0.00	1
	TA	-41618.87	-580.12	1.07×10^{-126}
	RT	-42057.39	-1457.16	0

[†]Bayes factors computed by marginal log-likelihoods with respect to the best model as 2 * ($\ln L_i$ - $\ln L_{Max}$)

Table 2 - Absolute divergence times and population sizes estimated[†] from BPP.

Node	N_e (x 10^3)	95% HPD	Divergence time (kya)	95% HPD
Ankafobe	19.2	17.3, 25.7	-	-
Ambohitantely	31.2	27.1, 40.9	-	-
Tsinjoarivo	23.1	14.0, 26.1	-	-
Ambatovy	55	33.5, 63.0	-	-
Riamalandy	204.5	193.4, 216.0	-	-
A	60.2	10.6, 201.3	26.7	15.9,30.4
B	22	15.1, 34.2	19	16.9, 25.2
C	875.1	776.9, 955.0	31	25.4, 42.7
D	108.3	106.6, 110.1	194.1	189.0, 198.8

[†]Estimates are based on combined posterior for priors θ = 0 and τ = 1.

Figure Legends

Figure 1 – Well-supported phylogenetic lineages of *Microcebus lehilahytsara* reflect genetic structure. The maximum clade credibility tree from SNAPP. Node support represents ML bootstrap, SVDquartets bootstrap, and posterior probabilities from SNAPP respectively. An asterisk represents 100% support. Branch lengths are median coalescent units with 95% highest posterior densities from SNAPP. STRUCTURE results are shown for an optimal k = 3. GPS coordinates for each sampling site are shown on the map.

Figure 2 – Migration models reveal gene flow between the Central Highlands and eastern forests. Bayes factors were used to test migration hypotheses between the eastern rainforest, Ambatovy (grey), and two fragmented forest patches in the Central Highlands, Ambohitantely (pink and grey) and Ankafobe (pink). These three locations were selected because of admixture detected in Ambatovy based on STRUCTURE analyses. θ parameters are scaled as $4N_e\mu$ and mutation-scaled migration parameters are shown near arrows. Divergence times for relevant models are in coalescent units.

Figure 3 – Divergence time estimates with BPP. Posterior distributions of MSC parameters for the combined posterior of prior choice 01. Branch lengths are in coalescent units and error bars represent 95% highest posterior density intervals around the median node height. Violin plots are the posterior distributions of node heights for prior choices 1 though 4 to show that while divergence time estimates are sensitive to prior choice, our biological conclusions are not.

Figure 4 – Population size change over time at sampling locations. Posterior distributions of θ at discretized bins of τ determined by the number of change points. Only Jackknife replicate one is shown for each population. The blue box shows the duration of the LGM.

Riamalandy   */*/*

Ambatovy   */*/*

Tsinjoarivo   */*/*

                          99/0/0.86

Ambohitantely   */*/*

                //*

                          //*

Ankafobe

                //*

Model	Marginal Likelihood	Bayes Factor*	Model Probability
0A – East to west	-408766.23	NA	0
0B – West to east	-408169.12	1194.22	0
1 – Stepping stone	-404252.47	9027.52	0
2 – N-island	-402558.14	12416.18	1
3 – Divergence and isolation	-415036.4	-12540.34	0
4 – Divergence and migration	-407777.86	1976.74	0

*Bayes factors computed with natural logarithms of marginal likelihoods with respect to model 0A as 2 * (lnL$_i$ – lnL$_{0A}$)
†θ values are presented as 10^{-3}