Impacts of predator-mediated interactions along a climatic gradient

on the population dynamics of an alpine bird

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Abstract

According to classic theory, species' population dynamics and distributions are less influenced by species interactions under harsh climatic conditions compared to under more benign climatic conditions. In alpine and boreal ecosystems in Fennoscandia, the cyclic dynamics of rodents strongly affect many other species, including ground-nesting birds such as ptarmigan. According to the 'alternative prey hypothesis' (APH), the densities of ground-nesting birds and rodents are positively associated due to predator-prey dynamics and prey-switching. However, it remains unclear how the strength of these predator-mediated interactions change along a climatic harshness gradient in comparison with the effects of climatic variation. We built a hierarchical Bayesian model to estimate the sensitivity of ptarmigan populations to interannual variation in climate and rodent occurrence across Norway during 2007–2017. Ptarmigan abundance was positively linked with rodent occurrence, consistent with the APH. Moreover, we found that rodent dynamics had stronger effects on ptarmigan in colder regions. Our study highlights how species interactions play an important role for the population dynamics of species at higher latitudes and suggests that they can become even more important in the most climatically harsh regions.

Keywords: Boreal ecosystems; Climate change; Indirect interactions; Population cycles; Predator-prey interactions

Introduction

Climatic variability and species interactions are two key drivers influencing the spatial and temporal patterns in the distribution and abundance of organisms [1, 2]. An old hypothesis, originally proposed by Darwin, posits that climate is the main determinant of species' range limits in harsh abiotic regions whereas species interactions are a more important determinant under benign abiotic conditions (hereafter the 'classic hypothesis') [3]. Range limits are determined by local population growth rates; hence, this hypothesis also implies that population growth rates should be more sensitive to climate variability in climatically harsh regions but more sensitive to species interactions in more climatically benign regions. Previous studies have examined changes in demographic and population processes along elevational and latitudinal climatic gradients [4-7], but few studies have been able to tease apart the sensitivity of population growth to climate and species interactions over large spatial and temporal scales.

In the cold environments of alpine and boreal areas, vertebrate species co-exist in relatively simple food-webs but biotic interactions still exert strong impacts on species' population dynamics [8]. One particular characteristic of northern alpine and boreal ecosystems in Fennoscandia is the rodent population dynamics, which are characterized by large inter-annual cyclic fluctuations in abundance, with peak years coming at regular intervals [9, 10]. Similar cycles are seen with other species in other parts of the world e.g., the snowshoe hare *(Lepus americanus)* in the boreal ecosystems of North America [11]. Although the mechanisms are still contested, predator-prey interactions are generally regarded as the most parsimonious explanation underlying the cyclic dynamics of rodents [12, 13], especially due to specialist predators, such as mustelids [14].

The population dynamics of many species of alpine and boreal communities, including the avian community, often covary with the phase of the rodent cycle [15, 16]. One hypothesised reason for this covariation is that ground-nesting birds (such as ptarmigan and grouse, *Tetraonidae*) and rodents share a similar set of generalist predators, including the red fox (*Vulpes vulpes*) and corvids [17, 18]. Hagen [19] proposed the 'alternative-prey hypothesis' (APH), stating that ground-nesting birds are less vulnerable to predation in peak rodent years because generalist predators switch to feeding on rodents. By contrast, ground-nesting birds suffer greater predation in rodent decline/crash years because there are fewer prey options available to hunting predators. Other indirect interactions based on shared predators, such as apparent competition, are typically predicted to lead to negative correlations in the abundance of alternative prey species [20]. Here, predator-mediated interactions between rodents and ground-nesting birds are hypothesized to lead to a positive association, such that ground-nesting birds are more abundant in peak rodent years.

Similar indirect effects of rodent population dynamics have been examined in a range of ecosystems,

and across a range of alternative prey species [16, 21, 22].

Previous studies have found partial support for the main predictions of the APH [16, 23];

however, the APH also makes secondary predictions about how the relative importance of predator-

mediated interactions should change along ecological gradients. In particular, Angelstam [15]

predicted that interactions should be stronger in regions with fewer prey species because the

predators become functionally more specialist, causing tighter linkages between predators and prey.

By contrast, in regions where predators have access to a more diverse array of prey species,

fluctuations in rodent populations should have weaker effects on the dynamics of predator

populations. Indeed, the reduced amplitude of the rodent population cycle at lower latitudes is

probably due to a stabilising effect of more generalist predators [24-26]. Climatic conditions,

especially snow cover, has also been linked with characteristics of the rodent cycles, with more cyclic

behaviour in regions with greater snow cover [16, 27, 28]. Based on these arguments, ground-nesting

birds might be less affected by the rodent cycle in less climatically harsh areas, where there is a more

diverse set of prey resources and weaker linkage between rodents and predators.

Alpine birds in Fennoscandia are reported to be declining in abundance over the last decades

[29], but the underlying causes for the widespread declines remain poorly understood. Willow

ptarmigan (*Lagopus lagopus*) are a resident species of montane habitats. Previous studies of

population dynamics have shown that the abundance of willow ptarmigan typically fluctuate in

synchrony with the small rodent dynamics [16, 30]. In addition, recruitment rates are affected by

weather conditions during the breeding season [30]. Thus, ptarmigan are a good model species for

testing alternative hypotheses about the relative importance of climate forcing and species

interactions on population dynamics. In this study, we compared the sensitivity of the population

dynamics of willow ptarmigan to temporal variability in both climate and rodent populations using a

multi-year (2007–2017) dataset covering a large geographic gradient across Norway. We assumed no

direct effect of rodents on ptarmigan but rather we expected their population dynamics to be linked

due to prey-switching of shared predators as predicted by the APH. Therefore, we used the statistical

relationship between rodent abundance and ptarmigan population dynamics as a signal of predator-

mediated interactions (Fig. 1). Although we use the term "harsh" to describe the coldest climatic

conditions within our study sites, we note that they are not necessarily harsh for the ptarmigan, a

species well-adapted to cold conditions. We tested three key predictions that emerge based on the

classic hypothesis about the relative importance of climate vs species interactions, and the

alternative prey hypothesis (Fig. 1):

1) Under the classic hypothesis, we predicted that predator-mediated interactions, and hence rodent population dynamics, would more strongly affect ptarmigan in warmer regions. By contrast, under the APH, we predicted that predator-mediated interactions, and hence rodent population dynamics, would more strongly affect ptarmigan in colder regions, where there are fewer prey sources for predators and stronger trophic linkages between predators and their prey populations.

2) Under both hypotheses, we predicted that ptarmigan population growth would be more sensitive to inter-annual climatic variability in colder regions, because they are closer to the edge of their physiological (climate) and ecological (food availability) niche, which may render them more sensitive to climatic variability.

3) Last, we predicted that the standardized effect sizes of rodents on the population growth rate of ptarmigan would be generally greater than the effect of climate because ptarmigan are well-adapted for life in cold environments and predation is a major determinant of reproductive success.

Fig. 1 (A) The hypothesized direct (solid) and indirect (dashed) relationships between ptarmigan and rodents as alternative prey, and their generalist predators, such as the red fox. In peak rodent years, ptarmigan are mostly ignored by predators; by contrast, in crash rodent years, ptarmigan are more frequently depredated. Together, these processes lead to a positive effect of rodent abundance on ptarmigan abundance. (B) Predicted effects of interannual climatic variability (grey lines) and rodent abundance (orange lines) on ptarmigan population dynamics according to the classic hypothesis (bottom) versus the 'alternative prey hypothesis' (top).

Methods

Population abundance data

We used a dataset of line-transect surveys covering almost the full latitudinal extent of Norway for an 11-year period (2007–2017). The surveys come from a structured citizen science program, based on coordinated collection by local and regional initiatives. Volunteer surveyors were instructed to follow common field procedures for estimation of ptarmigan densities using distance-sampling methods [31]. At a national or regional scale, however, the program does not allow for a random selection of study sites, because site selection is focused on the alpine habitats used by ptarmigan and is contingent on landowners establishing surveys on private and public lands. During the surveys, a team of two persons followed pre-defined transect lines, using trained pointing dogs to search both sides of the transect line, usually during August. For each detection of a cluster of ptarmigan (a group or an individual), surveyors recorded the perpendicular distances of the ptarmigan from the transect, the number of birds, the GPS coordinates and the date and time of day. In addition, length (and geographical position) of the transect line is recorded, as well as some other auxiliary data [32]. Since 2013, all data management, development of field procedures and planning of study design has been coordinated by the Hønsefuglportalen project (http://honsefugl.nina.no). Transects are spatially clustered into survey regions (median of 17 line-transects per survey region), reflecting data that are reported to the same survey coordination node. We used all data collected since 2007, when the monitoring was substantially expanded, until 2017. The total effort varied across the period, and not all study sites were surveyed each year. We subsetted our analysis to 708 line-transects that were visited in at least 6 years (a median of 10 years), each with a mean length of 3.7 km. We excluded observations made at distances greater than 200 m from the transect line, as well as detections by the surveyor, and not by the dog, at distances > 10 m away from the transect line because those observations were likely due to the surveyor flushing birds when moving towards the pointing dog which were observed only because the surveyor left the transect line.

Covariate data

Rodent data: During the line surveys, surveyors recorded whether rodents were observed along the transect as a binary variable (yes/no). The surveyors were not asked to distinguish among rodent species, but the five main species in the study area that show cyclic dynamics include: Norwegian lemming (*Lemmus lemmus*), tundra vole (*Microtus oeconomus*), field vole (*M. agrestis*), bank vole (*Myodes glareolus*) and the grey red-backed vole (*M. rufocanus*). Rodent occurrence is denoted by R in subsequent statistical models.

Climatic data: We obtained high-resolution climatic data from the Norwegian Meteorological institute (MET Norway: www.met.no). Data were based on an interpolation from local weather stations, with a spatial resolution of 1x1 km and are available on a daily basis. For each line-transect, we applied a buffer of 200 m and matched our transect locations with the climate data. With the compiled data, we characterised the spatio-temporal variation in climatic conditions during spring (prior to and at the start of the breeding period) and winter (marking the end of the breeding period). Specifically, we calculated:

1) Spring onset - first year day with a rolling 7-day mean snow depth of zero. We use the term 'spring' loosely since snow cover did not completely disappear until summer in some regions.
2) Spring temperature - average daily mean temperature during March, April and May
3) Winter onset - first year day with a rolling 7-day mean snow depth above zero
4) Winter temperatures – average daily mean temperature during December, January and February

For each climatic variable, we calculated (1) the average climatic conditions for each transect between 2007 and 2017, which defined the spatial climatic gradient (denoted as *CS*) and (2) the difference between the climatic value of each year at each transect and the spatial variable of each transect, which defined the temporal variation (i.e., anomalies, denoted as *CT*). Hence, we considered the effects of different characterisations of the climatic harshness gradient.

Harvesting data: We also compiled available data on harvest bags on ptarmigan but this data was only available at a coarse scale of administrative units and not at the line-transect scale (see SOM A for further details – no effect of harvesting on ptarmigan was found in our subsequent analysis but the covariate was retained in our model regardless).

Statistical analysis

Ptarmigan dynamics

We modelled ptarmigan dynamics using a hierarchical Bayesian model that included two linked models: a model for the observation process along each transect line (i.e. the observation model) and a model that described the ecological dynamics including the effect of climate and rodents on ptarmigan density as the latent variable (i.e. the state model).

(1) Observation model: Estimation of effective strip width of the ptarmigan line-transects

We fitted a distance-sampling detection model to estimate the effective strip width of each transect

[31]. The central assumption of distance-sampling theory is that the detection probability decreases

with increasing distance from the line-transect. We modelled the perpendicular distances of

observation events (one or more birds in a group) from the transect as a half-normal distribution. On

the transect line, we assumed perfect detection – a common assumption in distance-sampling [31].

We modelled sigma - the parameter of this half-normal distribution that reflects the rate of distance-

decay of detections - to be dependent on group size (i.e. the number of birds in each observation).

Mean group size was modelled with a Poisson mixed-effects model with line and survey region,

interacting with year (as a factor). To allow for any spatial variation in sigma, we additionally

included random effects for transect line (L, in equation below) and survey region (SR), allowing

some differences in how detectability declines with distance due to landscape or habitat features.

We also considered fixed effects of forest cover or precipitation during the survey but found no

effects (95% credible intervals overlapped zero) and thus did not include these factors in the model.

The final model for sigma was:

$$log(sigma_{i,t}) = b_0 + b_{GS} GroupSize_{i,t} + L_i + SR_i$$

201 The effective strip width of each transect (*i*) in each year (*t*) was calculated from sigma based on the

202 following equation:

$$ESW_{i,t} = \sqrt{\frac{(\pi * sigma_{i,t}{}^2)}{2}}$$

205 *(2) State model: Testing the effect of climate and rodent on ptarmigan populations*

206 We then used the estimated effective strip width (ESW) and data on transect length (TL) to relate

207 the total number of individuals observed along each transect (N, as following a negative binomial

208 distribution with constant dispersion parameter, *r*) to the latent variable, ptarmigan density, D,

209 (abundance per km^2) for each year *t* and each transect *i*:

$$Ptarmigan\_Obs_{i,t} \sim Negative\ Binomial(N_{i,t}, r)$$

$$N_{i,t} = D_{i,t} \times TL_{i,t} \times ESW_{i,t} \times 2$$

Ptarmigan density ($D_{i,t}$) was modelled in two different ways. In a first model ("random-effects

model"), we modelled the ptarmigan dynamics using a series of random effects that reflected the

spatial and temporal structure of the data. The random terms included year (as a factor), transect

line, survey region and survey region-year. We used this model to make predictions of the realized

ptarmigan dynamics, without explicitly specifying the underlying ecological covariates.

In a second set of models ("mixed-effects model"), we tested the additive effects of the two main

ecological variables of interest: rodent occurrence and climate as well as the interaction between

rodent occurrence and the spatial climatic variable. We tested the four climatic variables (winter

temperature, winter onset, spring temperature, spring onset) in separate models.

Density in year 1 was modelled using spatial random effects among line-transects. Density in

subsequent years were modelled as spatial (i) and temporal (t) variation in the population growth

rate as:

$$\ln D_{i,t} = \ln D_{i,t-1} + r_{i,t}$$

where the growth rate, r, was decomposed into:

$$r_{i,t} = b_0 + b_A \ln D_{i,t-1} + L_i + SR_i + Year_t + b_{CS} \, CS_i + b_{CT} \, CT_{i,t} + b_R \, R_{i,t} + b_{RL} \, R_{i,t-1} +$$
$$b_h \, H_{i,t-1} + b_{int\_CT} \, CS_i \, CT_{i,t} + b_{int\_R} \, CS_i \, R_{i,t}$$

where b_A assumes a Gompertz form of density-dependence, an assumption made by previous

ptarmigan studies [33, 34]. L and SR were random effects to account for the spatial grouping of data

into line-transects (L) and survey region (SR). A year (as a factor) random effect was included to

account for any additional causes of temporal variability.

CS and CT were covariates for the spatial gradient in climate and temporal anomalies for the climatic

variable, respectively. Two coefficients (b_R and b_{RL}) for rodents (R) tested the effects of rodent

occurrence immediately preceding the ptarmigan survey (during the same year, t) and a lagged effect

of rodent occurrence during the previous year (RL, *t-1*). We also accounted for the potential effect of

harvesting (*H*) on ptarmigan growth. All covariates were centered and scaled to units of 1 standard

deviation prior to analysis.

The critical tests of the classic versus the alternative prey hypotheses were based on the interaction

terms of our model. The interaction terms were: (1) between CS and CT to test whether the effect of

annual climatic anomalies changed along the climate gradient or (2) between CS and R (of the same

year) to test whether the effect of rodents changed along the climatic gradient.

In preliminary analyses, we explored additional random effects (a coarser spatial term) but they had

little effect on our results so we retained the simpler model.

Table 1. Table of model parameters on ptarmigan density and their meanings.

Term of model	Type	Statistical meaning
b_0	Fixed	Intercept
b_A	Fixed	Density-dependence
b_{CS}	Fixed	Spatial climatic variable
b_{CT}	Fixed	Temporal climatic anomalies
b_R	Fixed	Rodent occurrence (same year)
b_{RL}	Fixed	Lagged effect of rodent occurrence (previous year)
b_H	Fixed	Harvesting during the previous hunting season
b_{int_CT}	Fixed	Interaction term between temporal and spatial climate
b_{int_R}	Fixed	Interaction term between rodents and spatial climate
L	Random	Line-transect effect
SR	Random	Survey region effect
$Year$	Random	Year effect

Rodent dynamics

249   We included a submodel for rodent occurrence (presence/absence, modelled as a Bernoulli

250   distribution) within our hierarchical model for ptarmigan. This submodel was based on a series of

251   random intercept terms (for survey region [SR], year [Y], as a factor] and their interaction) to flexibly

252   predict the interannual variability in rodent dynamics. Hence, the rodent submodel was:

253
$$Rodent\_Occ_{i,t} \sim Bernoulli(R_{i,t})$$

254
$$logit(R_{i,t}) = b_0 + Y_t + SR_i + Y_t\, SR_i$$

Modelling rodent occurrence allowed us to impute missing data in the observed rodent dataset.

Additionally, the model predicted the probability of rodent occurrence at the level of the survey

region rather than at the transect-level, which mean meant smoothing over local reporting and

ecological variation. This choice was based on the assumptions that many predators (especially birds

of prey) forage over reasonable distances and are not likely affected by fine-scale transect-level

variation. We also considered a more complicated autoregressive model of order 2 for the rodent

dynamics, but similar cyclic dynamics were obtained, and similar effects on ptarmigan were found

(SOM B), so we proceeded with the simpler assumption-free random-effects model.

In further models, we also investigated how rodent dynamics changed along the climatic gradient. In

Bayesian mixed-effects models with binomial errors, we tested the effects of the spatial climatic

variables and temporal climatic anomalies variables on the probability of rodent occurrence. Line-

transect and survey regions were also included as random intercepts. Each of the four types of

climatic variables (winter temperature, winter onset, spring temperature, spring onset) were tested

in separate models.

Models were fitted using JAGS within R version 3.6.0. We used three MCMC chains, ran for 50,000

iterations, discarding the first 25,000 iterations as a burn-in period. We used uninformative or weakly

informative priors for all parameters, but a narrower prior for the density-dependence term (a

uniform prior between -2 and 0) to help convergence. We assessed model convergence using MCMC

chain traceplots and the Gelman-Rubin R-hat statistic (all <1.1). To assess the fit of the model, we

used the DHARMa package [35] that uses simulations from the fitted model to create scaled

residuals, which were then checked against the expected distribution using a quantile-quantile plot

and against the model predictions to check for any systematic deviations in fit. Covariate effects

were inferred when the 95% credible intervals did not overlap zero. Code for the fitted model is

available in the SOM C.

Results
Data description

The 708 line-transects (Fig. 2a) were clustered into 36 survey regions (Fig. 2b) distributed along a

wide latitudinal and climatic gradient across Norway. The warmest regions were those nearest the

coast and the coldest regions were inland and upland (Fig. 2b). Along each transect, a median of 2

groups were observed (range = 0–18), with a median of 4 (1–35) individuals seen per group. As

expected by distance-sampling theory, the frequency of observations declined with increasing

distance from the line-transect (Fig. 3). The average effective strip width was 107 m (interquartile

range = 100–114 m). Sigma of the half-normal distribution was positively affected by group size, with

the estimated effective strip width being 95 m for the smallest group size and 126 m for the largest

group size (Fig. 3).

Fig. 2 (a) Location of the 708 line-transects across Norway are shown by red lines. (b) The location of each of the 36 survey regions (a survey region is a cluster of neighbouring line-transects), coloured by their mean winter temperature during 2007–2017. Fig. S1 shows the same maps for the other climatic variables.

Fig. 3. Distance-dependent decay of ptarmigan detectability along the line-transects. The black line is the prediction for the median ptarmigan group size; the lower and upper dashed red lines are the predictions for the minimum and maximum ptarmigan group size.

 Climatic, rodent and ptarmigan population variation

Climate and rodent occurrence both showed large and generally spatially synchronized fluctuations

(Fig. 4, note: this figure shows the climatic gradient split into simplified quartiles, but climate was

treated as a continuous variable in all analyses). Our 11-year dataset spanned c. 3 rodent cycles. The

probability of rodent occurrence increased along the climatic harshness gradient, driven by higher

occurrence probability in peak years within colder regions (Table S1). For instance, in the first peak

rodent years, during 2010/2011, mean rodent occurrence across transects was 38% in the region of

the warmest quantile (based on winter temperature) but 64% in the coldest. In the second peak

rodent years, during 2014/2015, rodent occurrence was 18% in the warmest and 37% in the coldest.

Fig. 4 Time-series of winter temperature (°C), probabilities of rodent occurrence and willow

ptarmigan densities in August (birds per km^2) as predicted by the random-effects models. In our

analysis, the spatial climatic gradient was treated as a continuous variable. Here, for presentation

purposes we divided here the spatial climatic gradient into quartiles with equal numbers of transects

within each. Lines shown are means for each of the 36 survey regions (averaged over the transects

within each). Fig. S2 shows the time-series for the other climatic variables and Fig. S3 shows the

locations of each quartile.

Mean effects of climatic and rodent fluctuations on ptarmigan populations

Ptarmigan growth was higher in more climatically benign regions (positive spatial effects of climate

shown in Fig. 5), described by warmer springs and winters, and later winters and earlier springs.

Similarly, temporally, ptarmigan growth increased in years with warmer spring temperatures but was

less affected by the timing of seasonal onsets and winter temperatures (temporal effects of climate

shown in Fig. 5). Ptarmigan density was positively associated with rodent occurrence of the same

year, but negatively associated with rodent occurrence in the previous year (Fig. 5). Effects sizes of

rodents were larger than the effect sizes of the temporal variability in climate.

Season temperature

Season onset

Rodents

Fig. 5 Effects of seasonal temperatures, seasonal onsets, and rodent occurrence on changes in ptarmigan population density. Climate variables are split into a spatial variable (mean values across years) and a temporal variable (annual anomalies around the mean spatial variation). Covariates were standardized to units of standard deviation to facilitate comparison of effect sizes. Violins represent the full posterior distributions. Point and ranges represent mean effects and 95% credible intervals of the distributions. The dashed vertical red line is the line of no effect. The sign of the effects of spring onset were flipped so that larger values represent lower climatic harshness (i.e., earlier springs), consistent with the other climatic variables.

Changes in the effects of climatic and rodent fluctuations along the climatic harshness gradient

The statistical effects of rodent occurrence probability on ptarmigan densities were more positive in more climatically harsh areas compared with in more climatically benign areas (Fig. 6, left). In fact, the predicted effects of rodent occurrence were close to zero in regions with the warmest and earliest springs but strongly positive in the coldest regions with earliest winters and latest springs. The effects of temporal climatic variability also tended to become more positive along the climatic harshness gradient, when described in terms of spring temperature (Fig. 6, right) but the 95% credible intervals of the interaction terms overlapped zero (Fig. S4).

Fig. 6 Changes in the effect sizes of temporal variability in climate and rodent occurrence and along each climatic spatial gradient on ptarmigan density. X-axes are transformed so that left to right is an increasing climatic harshness gradient (i.e., the values towards the right represent colder temperatures, earlier winters and later spring). Shown are the mean standardized effect size and 95% credible interval. Each x-axis is centered around the median values for each variable (oC for temperatures and days for onset). 95% credible intervals for the interaction terms are shown in Fig. S4.

Discussion

Understanding how fluctuations in climatic conditions and species interactions affect species'
population dynamics is central to current research in population ecology. We outlined two different
hypotheses about how the strength of species interactions might vary along a climatic gradient, and
tested our predictions using a large-scale dataset on the willow ptarmigan. Our results suggest that
the impacts of predator-mediated interactions on willow ptarmigan increase along the climate
gradient, indicated by more strongly positive signals of rodent dynamics in colder areas. Hence, our
results were consistent with predictions stemming from the alternative prey hypothesis (APH) rather
than the classical view of how species interactions change with increasing climatic harshness.

Our results support our main prediction based on the APH: a stronger link between rodent
fluctuations and ptarmigan dynamics in colder regions. According to the APH, shared predators are
expected to prey-switch towards rodents and away from ptarmigan, when rodents are more
abundant [15, 19, 23]. Ptarmigan had higher growth rates during years with more rodents, which
would be consistent with lower predation pressure. Hence, our results are consistent with the APH,
previously tested over smaller-scales [23, 25, 36]. While the APH primarily considers the immediate
functional response of predators, the contrasting negative lagged effects of rodents is consistent
with a numerical response of predators due to greater prey density, leading to negative effects of
rodents on ptarmigan in the subsequent year. The weaker effect of rodents, and hence predator-
mediated interactions, on ptarmigan density in warmer regions could be caused by greater prey
diversity and more generalist predators, weakening the trophic links among the dynamics of
predators and prey species [8, 15].

Previous research has shown how the role of species interactions can increase or decrease in
strength along an ecological gradient due to different biotic and abiotic processes [5, 37-39].
Probably the most common reason for changing strengths of interactions over a geographic range is
changes in community assemblages and the abundance of interacting partners [3, 40, 41]. Our results
appear to be at odds with the classic prediction of stronger species interactions at the warmer end of
a species range [3], but the prediction arises from the assumption that the abundance of interacting
partners is greater in warmer regions. The assumption may hold in many scenarios, but in our case
rodents showed greater peaks in occurrence in colder regions. Indeed, other studies have shown an
increase in the amplitude of rodent dynamics from southern to northern Fennoscandia [8]. Hence,
our results are consistent with species interactions being important in areas where potential
interacting partners are more common, at least periodically.

Another factor likely to affect the changing importance of species interactions is whether the

predators are specialist or generalist species. When interactions involve specialists, they can be

important over the whole distributional range of an organism [42]. Although none of the interactions

between ptarmigan, rodents and predators are intrinsically specialist, they may be regarded as

functionally specialist in cold regions due to the relatively low number of species in the food web.

Since interaction networks at high latitudes tend to be often more specialised [43], other type of

indirect interactions may also become more important in more northerly regions.

We found weaker support for interannual climatic variability having a greater effect on

ptarmigan populations in colder regions. Climatic effects may be direct and related to species'

physiology, or more likely indirect and related to climate-caused changes in the availability of

resources or suitable habitat. Harsh conditions may expose an organism to the edge of its niche

space, hence climatic variation at the colder edge of a species' range should be more likely to affect

its population growth [44]. However, the ptarmigan is a well-adapted alpine species able to inhabit

regions with harsher climates than those in our study area (e.g., in Alaska and Siberia). Our study

does not therefore address the geographic range limits of the ptarmigan. However, other studies

have found clearer evidence that species' sensitivity to climate increases towards the range edge [45,

46]. Further work might consider the non-linearity in the relationship between climatic conditions

and ptarmigan density to examine in more depth the effects of climatic variation at the climatic

extremes.

High-altitude or -latitude organisms are among those most strongly exposed to climate

change [47]. Our findings help understand the main pathways through which climate change may

affect the population dynamics of ptarmigan, including the direct effects of warming versus indirect

effects mediated through predators [48, 49]. Ptarmigan benefit from warmer springs, hence, climate

change, at least in the short-term, may have some positive effects [50]. However, our results suggest

that changes in predation pressure mediated via rodents are likely to be more important. One

predicted consequence of climate change is a dampening of rodent cycles [16, 51]. Dampening cycles

could mean no or less frequent years of high rodent abundance, which offer temporal refuges from

predation that yield 'boom' years with high ptarmigan productivity. Hence, a warming climate may

lead to a more constant rate of predation pressure on ptarmigan, lowering mean population growth

rates. However, dampening of the cycles might also reduce the long-term abundance of predators,

lowering predation pressure on the ptarmigan [12]. In particular, more frequent years with low vole

abundance is expected to directly affect predator reproduction in the spring [51].

Our analysis has several limitations. First, the rodent data was only collected as a binary

presence/absence variable along each line-transect and hence may not reflect the full changes in the

relative abundances of rodents among years. Moreover, our rodent data was collected in the

autumn, but rodent abundance in spring and early summer is probably the key parameter that drives

predation pressure on ptarmigan and other ground-nesting birds. However, no comparable spring

data for rodents are available spanning the range of our study system. Usually, we could expect

spring and autumn rodent densities of the same year to be correlated [16] but that might vary with

climatic conditions [51]. Overall, the patterns in the proportion of rodents seen within our data

showed the expected large-scale synchronised annual cycles or rodents reported elsewhere [52].

Second, although we can reasonably infer the interaction between ptarmigan and rodents to be

caused by shared predators, data on predators would also help better understand the links between

predator abundance, prey preference, and alternative prey availability [23]. Last, our analysis only

focused on changes in the strength of one specific type of species interaction. Willow ptarmigan may

be involved in various other types of species interactions such as competition with other herbivorous

species, including ungulates, which differ in their strength along the climate harshness gradient.

Hence, cumulatively, species interactions could still be more important at the warmer edge of its

distribution.

Our analyses suggest that predator-mediated interactions become even more important in

the colder regions of boreal ecosystems, contrary to the classic view that species interactions are

more important at the warmer edge of species' distributions. The role of predator-mediated

interactions along a climatic gradient are more generally likely to depend on the diet breadth of

predators and availability of different prey types. Rodent cycles – regarded as the heartbeat of boreal

ecosystems - cause changes in prey availability that lead to predator-mediated interactions for

alternative prey species. Long-term dampening of the rodent cycles that is predicted to arise due to

climate change is likely to have widespread repercussions for the dynamics of many species in the

boreal, especially ground-nesting birds such as the willow ptarmigan.

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Data availability

The ptarmigan dataset (for different geographic regions) is freely available via GBIF [53-55]. The subset used for the analysis of this paper, along with the rodent data, are available at Dryad (link here XXX).

Competing interests

The authors have no competing interested to declare.

Author contributions

EBN and HCP designed the line-transect sampling protocols and helped to develop and maintain the Hønsefuglportalen system. DB developed the research questions, with contributions from EBN. DB conducted the statistical analyses. DB wrote the first draft of the manuscript and all coauthors helped to refine the hypotheses and develop the manuscript.

References

1.   Pearson R.G., Dawson T.P. 2003 Predicting the impacts of climate change on the distribution of species: are bioclimate envelope models useful? *Glob Ecol Biogeogr* **12**(5), 361-371. (doi:10.1046/j.1466-822X.2003.00042.x).
2.   Mutshinda C.M., O'Hara R.B., Woiwod I.P. 2011 A multispecies perspective on ecological impacts of climatic forcing. *J Anim Ecol* **80**(1), 101-107. (doi:10.1111/j.1365-2656.2010.01743.x).
3.   Louthan A.M., Doak D.F., Angert A.L. 2015 Where and When do Species Interactions Set Range Limits? *Trends Ecol Evol* **30**(12), 780-792. (doi:10.1016/j.tree.2015.09.011).
4.   Ettinger A.K., Ford K.R., HilleRisLambers J. 2011 Climate determines upper, but not lower, altitudinal range limits of Pacific Northwest conifers. *Ecology* **92**(6), 1323-1331. (doi:10.1890/10-1639.1).
5.   Kozlov M.V., Stekolshchikov A.V., Soderman G., Labina E.S., Zverev V., Zvereva E.L. 2015 Sap-feeding insects on forest trees along latitudinal gradients in northern Europe: a climate-driven patterns. *Glob Change Biol* **21**(1), 106-116. (doi:10.1111/gcb.12682).
6.   Lim J.Y., Fine P.V.A., Mittelbach G.G. 2015 Assessing the latitudinal gradient in herbivory. *Glob Ecol Biogeogr* **24**(10), 1106-1112. (doi:10.1111/geb.12336).
7.   Roslin T., Hardwick B., Novotny V., Petry W.K., Andrew N.R., Asmus A., Barrio I.C., Basset Y., Boesing A.L., Bonebrake T.C., et al. 2017 Higher predation risk for insect prey at low latitudes and elevations. *Science* **356**(6339), 742-744. (doi:10.1126/science.aaj1631).
8.   Hanski I., Henttonen H., Korpimaki E., Oksanen L., Turchin P. 2001 Small-rodent dynamics and predation. *Ecology* **82**(6), 1505-1520. (doi:10.1890/0012-9658(2001)082[1505:srdap]2.0.co;2).
9.   Krebs C.J. 1996 Population cycles revisited. *J Mammal* **77**(1), 8-24. (doi:10.2307/1382705).
10.   Elton C. 1942 *Voles, Mice and Lemmings - Problems in Population Dynamics*. Oxford, Clarendon press.
11.   Krebs C.J. 2011 Of lemmings and snowshoe hares: the ecology of northern Canada. *Proceedings of the Royal Society B-Biological Sciences* **278**(1705), 481-489. (doi:10.1098/rspb.2010.1992).

12.   Ims R.A., Fuglei E. 2005 Trophic interaction cycles in tundra ecosystems and the impact of
climate change. *Bioscience* **55**(4), 311-322. (doi:10.1641/0006-3568(2005)055[0311:ticite]2.0.co;2).
13.   Myers J.H. 2018 Population cycles: generalities, exceptions and remaining mysteries.
Proceedings of the Royal Society B-Biological Sciences **285**(1875). (doi:10.1098/rspb.2017.2841).
14.   Hanski I., Hansson L., Henttonen H. 1991 Specialist predators, generalist predators, and the
microtine rodent cycle. *J Anim Ecol* **60**(1), 353-367. (doi:10.2307/5465).
15.   Angelstam P., Lindstrom E., Widen P. 1984 Role of predation in short-term population
fluctuations of some birds and mammals in Fennoscandia. *Oecologia (Berlin)* **62**(2), 199-208.
(doi:10.1007/bf00379014).
16.   Kausrud K.L., Mysterud A., Steen H., Vik J.O., Ostbye E., Cazelles B., Framstad E., Eikeset A.M.,
Mysterud I., Solhoy T., et al. 2008 Linking climate change to lemming cycles. *Nature* **456**(7218), 93-
U93. (doi:10.1038/nature07442).
17.   Moss R., Watson A. 2001 Population cycles in birds of the grouse family (Tetraonidae). In
Advances in Ecological Research, Vol 32 (ed. Caswell H.), pp. 53-111.
18.   Valkama J., Korpimaki E., Arroyo B., Beja P., Bretagnolle V., Bro E., Kenward R., Manosa S.,
Redpath S.M., Thirgood S., et al. 2005 Birds of prey as limiting factors of gamebird populations in
Europe: a review. *Biological Reviews* **80**(2), 171-203. (doi:10.1017/s146479310400658x).
19.   Hagen Y. 1952 *Rovfuglene og viltpleien*. Oslo, Gyldendal Norsk forlag.
20.   Holt R.D., Grover J., Tilman D. 1994 Simple rules for interspecific dominance in systems with
exploitative and apparent competition. *Am Nat* **144**(5), 741-771. (doi:10.1086/285705).
21.   McKinnon L., Berteaux D., Bety J. 2014 Predator-mediated interactions between lemmings
and shorebirds: A test of the alternative prey hypothesis. *Auk* **131**(4), 619-628. (doi:10.1642/auk-13-
154.1).
22.   Zarybnicka M., Riegert J., Kouba M. 2015 Indirect food web interactions affect predation of
Tengmalm's Owls Aegolius funereus nests by Pine Martens Martes martes according to the
alternative prey hypothesis. *Ibis* **157**(3), 459-467. (doi:10.1111/ibi.12265).
23.   Breisjoberget J.I., Odden M., Wegge P., Zimmermann B., Andreassen H. 2018 The alternative
prey hypothesis revisited: Still valid for willow ptarmigan population dynamics. *Plos One* **13**(6).
(doi:10.1371/journal.pone.0197289).
24.   Bjornstad O.N., Falck W., Stenseth N.C. 1995 Geographic gradient in small rodent density-
fluctuations – a statistical modeling approach. *Proceedings of the Royal Society B-Biological Sciences*
262(1364), 127-133. (doi:10.1098/rspb.1995.0186).
25.   Kurki S., Helle P., Linden H., Nikula A. 1997 Breeding success of black grouse and capercaillie
in relation to mammalian predator densities on two spatial scales. *Oikos* **79**(2), 301-310.
(doi:10.2307/3546014).
26.   Linden H. 1988 Latitudinal gradients in predator-prey interactions, cyclicity and synchronism
in voles and small game populations in Finland. *Oikos* **52**(3), 341-349. (doi:10.2307/3565208).
27.   Hansson L., Henttonen H. 1985 Gradients in density variation of small rodents – the
important of latiude and snow cover. *Oecologia* **67**(3), 394-402. (doi:10.1007/bf00384946).
28.   Ims R.A., Yoccoz N.G., Killengreen S.T. 2011 Determinants of lemming outbreaks. *Proc Natl
Acad Sci U S A* **108**(5), 1970-1974. (doi:10.1073/pnas.1012714108).
29.   Lehikoinen A., Green M., Husby M., Kalas J.A., Lindstrom A. 2014 Common montane birds are
declining in northern Europe. *J Avian Biol* **45**(1), 3-14. (doi:10.1111/j.1600-048X.2013.00177.x).
30.   Kvasnes M.A.J., Pedersen H.C., Storaas T., Nilsen E.B. 2014 Large-scale climate variability and
rodent abundance modulates recruitment rates in Willow Ptarmigan (Lagopus lagopus). *Journal of
Ornithology* **155**(4), 891-903. (doi:10.1007/s10336-014-1072-6).
31.   Buckland S.T., Anderson D.R., Burnham K.P., Laake J.L., Borchers D.L., Thomas L. 2001
Introduction to distance sampling: estimating abundance of biological populations. London, Oxford
University Press.
32.   Nilsen E.B., Vang R., Asbjørnsen E. 2018 Tetraonid line transect surveys from Norway: Data
from Finnmarkseiendommen (FeFo). Version 1.2. . (ed. dataset N.I.f.N.R.S.e.).

33.   Aanes S., Engen S., Saether B.E., Willebrand T., Marcstrom V. 2002 Sustainable harvesting
strategies of Willow Ptarmigan in a fluctuating environment. *Ecol Appl* **12**(1), 281-290.
(doi:10.1890/1051-0761(2002)012[0281:shsowp]2.0.co;2).
34.   Pedersen H.C., Steen H., Kastdalen L., Broseth H., Ims R.A., Svendsen W., Yoccoz N.G. 2004
Weak compensation of harvest despite strong density-dependent growth in willow ptarmigan.
Proceedings of the Royal Society B-Biological Sciences **271**(1537), 381-385.
(doi:10.1098/rspb.2003.2599).
35.   Hartig F. 2020 DHARMa: Residual Diagnostics for Hierarchical (Multi-Level / Mixed)
Regression Models. R package version 0.3.2.0. https://CRAN.R-project.org/package=DHARMa. (
36.   Francksen R.M., Whittingham M.J., Ludwig S.C., Roos S., Baines D. 2017 Numerical and
functional responses of Common Buzzards Buteo buteo to prey abundance on a Scottish grouse
moor. *Ibis* **159**(3), 541-553. (doi:10.1111/ibi.12471).
37.   Didion M., Kupferschmid A.D., Wolf A., Bugmann H. 2011 Ungulate herbivory modifies the
effects of climate change on mountain forests. *Clim Change* **109**(3-4), 647-669. (doi:10.1007/s10584-
011-0054-4).
38.   Fussmann K.E., Schwarzmuller F., Brose U., Jousset A., Rall B.C. 2014 Ecological stability in
response to warming. *Nature Climate Change* **4**(3), 206-210. (doi:10.1038/nclimate2134).
39.   Vucic-Pestic O., Ehnes R.B., Rall B.C., Brose U. 2011 Warming up the system: higher predator
feeding rates but lower energetic efficiencies. *Glob Change Biol* **17**(3), 1301-1310.
(doi:10.1111/j.1365-2486.2010.02329.x).
40.   Vazquez D.P., Poulin R., Krasnov B.R., Shenbrot G.I. 2005 Species abundance and the
distribution of specialization in host-parasite interaction networks. *J Anim Ecol* **74**(5), 946-955.
(doi:10.1111/j.1365-2656.2005.00992.x).
41.   Fort H., Vazquez D.P., Lan B.L. 2016 Abundance and generalisation in mutualistic networks:
solving the chicken-and-egg dilemma. *Ecol Lett* **19**(1), 4-11. (doi:10.1111/ele.12535).
42.   Neuschulz E.L., Merges D., Bollmann K., Gugerli F., Bohning-Gaese K. 2018 Biotic interactions
and seed deposition rather than abiotic factors determine recruitment at elevational range limits of
an alpine tree. *Journal of Ecology* **106**(3), 948-959. (doi:10.1111/1365-2745.12818).
43.   Dalsgaard B., Schleuning M., Maruyama P.K., Dehling D.M., Sonne J., Vizentin-Bugoni J.,
Zanata T.B., Fjeldsa J., Bohning-Gaese K., Rahbek C. 2017 Opposed latitudinal patterns of network-
derived and dietary specialization in avian plant-frugivore interaction systems. *Ecography* **40**(12),
1395-1401. (doi:10.1111/ecog.02604).
44.   Oedekoven C.S., Elston D.A., Harrison P.J., Brewer M.J., Buckland S.T., Johnston A., Foster S.,
Pearce-Higgins J.W. 2017 Attributing changes in the distribution of species abundance to weather
variables using the example of British breeding birds. *Methods Ecol Evol* **8**(12), 1690-1702.
(doi:10.1111/2041-210x.12811).
45.   Mills S.C., Oliver T.H., Bradbury R.B., Gregory R.D., Brereton T., Kuhn E., Kuussaari M.,
Musche M., Roy D.B., Schmucki R., et al. 2017 European butterfly populations vary in sensitivity to
weather across their geographical ranges. *Glob Ecol Biogeogr* **26**(12), 1374-1385.
(doi:10.1111/geb.12659).
46.   Myers-Smith I.H., Elmendorf S.C., Beck P.S.A., Wilmking M., Hallinger M., Blok D., Tape K.D.,
Rayback S.A., Macias-Fauria M., Forbes B.C., et al. 2015 Climate sensitivity of shrub growth across the
tundra biome. *Nature Climate Change* **5**(9), 887-+. (doi:10.1038/nclimate2697).
47.   Burrows M.T., Schoeman D.S., Buckley L.B., Moore P., Poloczanska E.S., Brander K.M., Brown
C., Bruno J.F., Duarte C.M., Halpern B.S., et al. 2011 The Pace of Shifting Climate in Marine and
Terrestrial Ecosystems. *Science* **334**(6056), 652-655. (doi:10.1126/science.1210288).
48.   Cahill A.E., Aiello-Lammens M.E., Fisher-Reid M.C., Hua X., Karanewsky C.J., Ryu H.Y., Sbeglia
G.C., Spagnolo F., Waldron J.B., Warsi O., et al. 2013 How does climate change cause extinction?
Proceedings of the Royal Society B-Biological Sciences **280**(1750). (doi:10.1098/rspb.2012.1890).
49.   Cahill A.E., Aiello-Lammens M.E., Fisher-Reid M.C., Hua X., Karanewsky C.J., Ryu H.Y., Sbeglia
G.C., Spagnolo F., Waldron J.B., Wiens J.J. 2014 Causes of warm-edge range limits: systematic review,

proximate factors and implications for climate change. *J Biogeogr* **41**(3), 429-442.
(doi:10.1111/jbi.12231).
50.   Wegge P., Rolstad J. 2017 Climate change and bird reproduction: warmer springs benefit
breeding success in boreal forest grouse. *Proceedings of the Royal Society B-Biological Sciences*
284(1866). (doi:10.1098/rspb.2017.1528).
51.   Cornulier T., Yoccoz N.G., Bretagnolle V., Brommer J.E., Butet A., Ecke F., Elston D.A.,
Framstad E., Henttonen H., Hornfeldt B., et al. 2013 Europe-Wide Dampening of Population Cycles in
Keystone Herbivores. *Science* **340**(6128), 63-66. (doi:10.1126/science.1228992).
52.   Framstad E. 2017 Ground vegetation, epiphytes, small mammals and birds. Summary of
results In *Terrestrial Ecosystems Monitoring in 2016* (NINA Report 1376).
53.   Nilsen E.B., Vang R., Asbjørnsen E. 2020 Tetraonid line transect surveys from Norway: Data
from Finnmarkseiendommen (FeFo). Version 1.5. Norwegian Institute for Nature Research. Sampling
event dataset https://doi.org/10.15468/s7c8qd
54.   Nilsen E.B., Vang R., Breisjøberget J.I. 2002 Tetraonid line transect surveys from Norway:
Data from Statskog. Version 1.6. Norwegian Institute for Nature Research. Sampling event dataset
https://doi.org/10.15468/q2ehlk.
55.   Nilsen E.B., Vang R., Kjønsberg M., Kvasnes M.A.J. 2020 Tetraonid line transect surveys from
Norway: Data from Fjellstyrene. Version 1.3. Norwegian Institute for Nature Research. Sampling
event dataset https://doi.org/10.15468/975ski.