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The Essential Facts of Wuhan Novel Coronavirus Outbreak in China and Epitope-based Vaccine Designing against 2019-nCoV

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Abstract

Wuhan Novel Coronavirus (2019-nCoV) outbreak has become global pandemic which has raised the concern of scientific community to deign and discover a definitive cure against this deadly virus which has caused deaths of numerous infected people upon infection and spreading. To date, there is no antiviral therapy or vaccine is available which can effectively combat the infection caused by this virus. This study was conducted to design possible epitope-based subunit vaccines against the 2019-nCoV using the approaches of reverse vaccinology and immunoinformatics. Upon continual computational experimentation three possible vaccine constructs were designed and one vaccine construct was selected as the best vaccine based on molecular docking study which is supposed to effectively act against the Wuhan Novel Coronavirus. Later, molecular dynamics simulation and in silico codon adaptation experiments were carried out in order to check biological stability and find effective mass production strategy of the selected vaccine. Hopefully, this study will contribute to uphold the present efforts of the researches to secure a definitive treatment against this nasty virus.

Keywords: 2019-nCoV; Coronavirus; China; Reverse Vaccinology; Vaccine; Wuhan

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1. Introduction

1.1. Origin of Coronavirus, Their Morphology, Pathology and Others

Coronaviruses (CoVs), is enveloped positive-sense RNA viruses, which is surrounded by crown-shaped, club-like spikes projection on the outer surface [1][2]. Coronaviruses spike protein are glycoprotein that are embedded over the viral envelope. This spike protein attaches to specific cellular receptors and initiates structural changes of spike protein, and causes penetration of cell membranes which results in the release of the viral nucleocapsid into the cell [3]. The viral spike protein includes N-terminal, which is crucial for identification of Coronaviruses [4]. Coronaviruses have a large RNA genome in the size ranging from 26 to 32 kilobases and capable of obtaining distinct ways of replication [5]. Like all other RNA viruses, coronavirus under goes replication of genome and transcription of mRNAs upon infection. Synthesis of a full-length negative-strand RNA serves as template for full-length genomic RNA [3][6]. Coronaviruses are a large family of viruses belonging to the family Coronaviridae and the order Nidovirales [7]. that are common in many different species of animals, including camels, cattle, cats, and bats [8], This group of viruses can cause wide varieties of respiratory illness in mammals and birds. In human, it causes common cold, leading to severe illness pneumonia to elderly people and immunocompromised people, like hospital patients [9]. This viral pathogen was responsible for the Middle East Respiratory Syndrome (MERS) and Severe Acute Respiratory (SARS), and 2019 Novel coronavirus (2019-nCoV) in China outbreaks [10]. Coronavirus in form of SARS, MERS and Novel coronavirus are lethal leading to large number of deaths. Coronavirus subfamily is classified into four genera, the alpha, beta, gamma and delta coronaviruses. Human CoV infections are caused by alpha- and beta-CoVs. CoVs are common human pathogens, and 30% to 60% of the Chinese population is positive for anti-CoV antibodies [11]. These viral infections caused by coronaviruses are generally

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associated the upper respiratory tract to lower respiratory tract. Immunocompromised people like elderly people and infants are more vulnerable and susceptible to this group of viruses [12].

For many decades HCoVs are well adapted to humans and it had been prevalent among Human races. In 1960s, first identification of Human coronaviruses (HCoVs) was made in patients with common cold. Following that incident more HCoVs has been detected, that involved the two major outbreak SARS and MERS, two pathogens that, upon infection, can cause fatal respiratory disease in humans [13].

The most common coronaviruses among humans are 229E, NL63, OC43, and HKU1 and some can evolve and cause human diseases, becoming new human coronaviruses. Three recent examples of these are 2019-nCoV, SARS-CoV, and MERS-CoV [14]. SARS-CoV was the causal agent of the severe acute respiratory syndrome outbreaks in 2002 and 2003 in Guangdong Province, China. 6-8 MERS-CoV was the pathogen responsible for severe respiratory disease outbreaks in 2012 in the Middle East [15].

Chinese scientists were able to sequence the viral genome using next genome sequencing from the collected samples and identified the cause as a Sars-like coronavirus. The genetic sequence is now available for scientist all around the world which would aid faster diagnosis of further cases [2] [16][17]. Based on full genome sequence data on the Global Initiative on Sharing All Influenza Data [GISAID] platform, the genetic sequence of the 2019 novel coronavirus (2019-nCoV) ensuring faster development of point-of-care real-time RT-PCR diagnostic tests specific for 2019-nCoV [18]. Association of genome sequence with other evidences demonstrated that 2019 nCoV is 75 to 80% identical to the SARS-CoV and even more closely related to several bat coronaviruses. 2019-nCoV can be cultured in the same cells that are ideal for growing SARS-CoV

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and MERS-CoV, unlike SARS-CoV or MERS-CoV, 2019-nCoV grows better in primary human airway epithelial cells than in standard tissue-culture cells [19]. Another group of scientists have investigated the origin of the 2019- nCoV, using comprehensive sequence analysis and comparison in conjunction with relative synonymous codon usage (RSCU) within different animal species and sequences of newly identified coronavirus 2019-nCoV. The evidence and analysis suggest that the 2019-nCoV appears to be a recombinant virus between the bat coronavirus and an origin-unknown coronavirus. the viral spike glycoprotein recombined within themselves, and recognizes cell surface receptor. Scientist are speculating that the 2019-nCoV originated form snakes based on Homologous recombination within the spike glycoprotein could be a probable way of transmission in between snake and humans [20].

1.2. Severity of China Outbreak of Wuhan Novel Coronavirus (2019-nCoV)

China is in its extremity due to the outbreak of a respiratory virus, called coronavirus and considered as related to SARS virus [21][22]. It is reported to be originated in the animal and seafood market of Wuhan, Hubei province, China – one of the largest cities in China which has 11M residents thus entitled as Wuhan coronavirus or more commonly 2019-nCoV. [21] [23] World health organization (WHO) is investigating the virus since 31st December, 2019 after being reported. [24][25] 2019-nCOV is a deadly virus and Chinese authority reported WHO a pneumonia like case on 31st December, 2019 and the infection has spread destructively since then [26]. First death of a 61 years old man was reported on 11th January by Chinese health authorities in Wuhan, Hubei province, China [25][27][28]. Second death of a 69 years old man was reported on 15th January in the central city of Wuhan, Hubei province, China who had symptoms of pneumonia [29][30]. In Thailand, first emerging international case outside the China was reported on 13th January and the first emerging case outside the Wuhan city, China was reported on 19th January

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[25][31]. On January 20, it was reported that 2019-nCOV virus can be transmitted from human to human which cracked the idea that it can only be transmitted through infected animals [32]-[35] and on the same day China Health authority reported the third death [36]. It was recorded that the number of infections rose upto 10 fold between 17th-20th January [37]. United States confirmed first infected case in Washington state on 21st January who had recently been returned from china [38]. As of 23rd January China reported 830 infected cases and the death toll rose upto 25 in China [39][40]. On the same date several confirmed infected cases were reported worldwide, Taiwan (1 case), Macao(2 cases), Hong Kong(2cases),Vietnam (2cases) ,Thailand (3cases), Japan (2cases), South Korea(2 cases),Singapore (1case) ,United States (1case) [41]. On 24 January the health minister of France Agnes Buzyn announced three confirmed infected cases, two in Paris and 1 case in Bordeaux. France is the first European country to be announced as infected [42]-[44]. Chinese authority banned public and air transport in Wuhan on 23 January and the residents of the country were kept under quarantine [45]. National Health Commission of China reported on 25th January that as of 24th January the virus took 41 lives and 1300 infected cases were reported worldwide [46]-[48]. As of January 25, China confirmed 2000 infected cases worldwide, among these 1975 cases were reported in China with rising death toll to 56 [49]-[51]. Chinese health authority announced on January 27 that the coronavirus death toll jumped to 81 and 2744 infected cases in china were reported [52]-[54]. German authorities confirmed first infected case of coronavirus and it is reported as second European country to be infected. The health authority said that the patient is in the good condition and is kept under medical observation [55][56]. Health and Human Services Secretary of U.S. said on 28th January that the infected cases in U.S. remained five with no deaths, while France reported forth infected case. In china 106 deaths with more than 4520 infected cases were reported on 28th January [57][58]. Alarming situation due to rising

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confirmed cases outside China made governments of other countries to start evacuation of their citizens from Wuhan, China and these include U.S., France, Japan, Australia, Germany, New Zealand, France [59]. Approximately, 170 deaths and 7,711 confirmed cases were reported with 1,737 new cases by Chinese National health commission on 29th January [60]. The U.S. government evacuated 240 Americans, mostly diplomats and their family members from the epicenter of the virus outbreak on 29th January by a chartered airplane [61]. Another report stated that on the same day chartered ANA airplane by Japanese government landed in Heneda Airport after evacuating 206 Japanese nationals [62]. The World Health Organization (WHO) had a meeting at the WHO headquarter in Geneva on 30th January and WHO declared the outbreak as Public Health Emergency and it is also said by the committee that the declaration was made giving priority the other countries outside China [63]. On 30th January the U.S. state department advised their citizens not to travel to China as the death toll jumped to 204 with 9,692 infected cases worldwide [64]. In China 213 deaths and globally 9,826 confirmed cases were reported on 31st January by Chinese national health authorities, among which 9,720 cases in China and 106 cases in 19 countries outside China [65][66]. On 1st February the death toll reached to 258 with 11,948 confirmed cases worldwide stated by Hubei Provincial Health Commission [67][68]. On the same day German Ministry of Health said that a military airplane landed at Frankfurt airport after evacuating 128 German citizens from Wuhan who will be quarantined till mid of February and the health minister also stated that Germany military aircraft delivered aid and 1000 protective suits to Chinese authorities requested by China [69]. First known fatality outside China reported of a 44 year old man in Philippines by Philippine health officials on 2nd February as the novel coronavirus outbreak took 362 lives and 17,205 cases were reported according to Chinese and World Health Organization data [70][71].The Nation's flagship carrier, Qantas , was chartered by Australian

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government and the airplane landed in Learmonth, Western Australia after evacuating 243 citizens including 89 children on 3rd February from where the citizens would be sent by another flight to Christmas Island to be quarantined [72]. New 1000-bed facility Huoshenshan Hospital in Wuhan is built in only 10 days to mitigate the pressure of patients in other medical hospitals in Wuhan. A crew of more than 7000 workers started working around the clock on 24 January which covers 60000 square meters with special quarantine area and opened its door to receive patients on 3rd February [73][74]. Kerala government announced the third confirmed case in India on 3rd February who is a student [75] just like the 1st case (reported on 30th January in Kerala) and 2nd case (reported on 2nd February in Kerala) according to the Ministry of Health and Family welfare of India [76][77]. A British pharmaceutical company, GlaxoSmithKline (GSK) stated on 3rd February that they would like to collaborate with Coalition for Epidemic Preparedness Innovations (CEPI) that works for developing vaccines. GSK said that they are ready to work with their adjuvant technology to increase immunity to the patients as well as to develop vaccine for coronavirus outbreak Health authorities in China stated on 4th February that as end of 3rd February the coronavirus fatalities jumped to 425 in China and 427 worldwide as the second fatality announced in Hong Kong of a 39 year old man reported on same day in the early morning [75][78]. The authority also said that the confirmed cases reached to 20,438 worldwide [79][80].

To date, there is no effective antiviral therapies that can combat the Coronavirus infections and hence the treatments are only supportive. Use of Interferons in combination with Ribavirin is somewhat effective. However, the effectiveness of combined remedy needs to be further evaluated [81].

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This experiment is carried out to design novel epitope-based vaccine against four proteins of Wuhan novel coronavirus (2019-nCoV) i.e., nucleocapsid phosphoprotein which is responsible for genome packaging and viral assembly [82]; surface glycoprotein that is responsible for membrane fusion event during viral entry [83][84]; ORF3a protein that is responsible for viral replication, characterized virulence, viral spreading and infection [85] and membrane glycoprotein which mediates the interaction of virions with cell receptors [86] using the approaches of reverse vaccinology.

Reverse vaccinology refers to the process of developing vaccines where the novel antigens of a virus or microorganism or organism are detected by analyzing the genomic and genetic information of that particular virus or organism. In reverse vaccinology, the tools of bioinformatics are used for identifying and analyzing these novel antigens. These tools are used to dissect the genome and genetic makeup of a pathogen for developing a potential vaccine. Reverse vaccinology approach of vaccine development also allows the scientists to easily understand the antigenic segments of a virus or pathogen that should be given more emphasis during the vaccine development. This method is a quick, cheap, efficient, easy and cost-effective way to design vaccine. Reverse vaccinology has successfully been used for developing vaccines to fight against many viruses i.e., the Zika virus, Chikungunya virus etc. [87][88].

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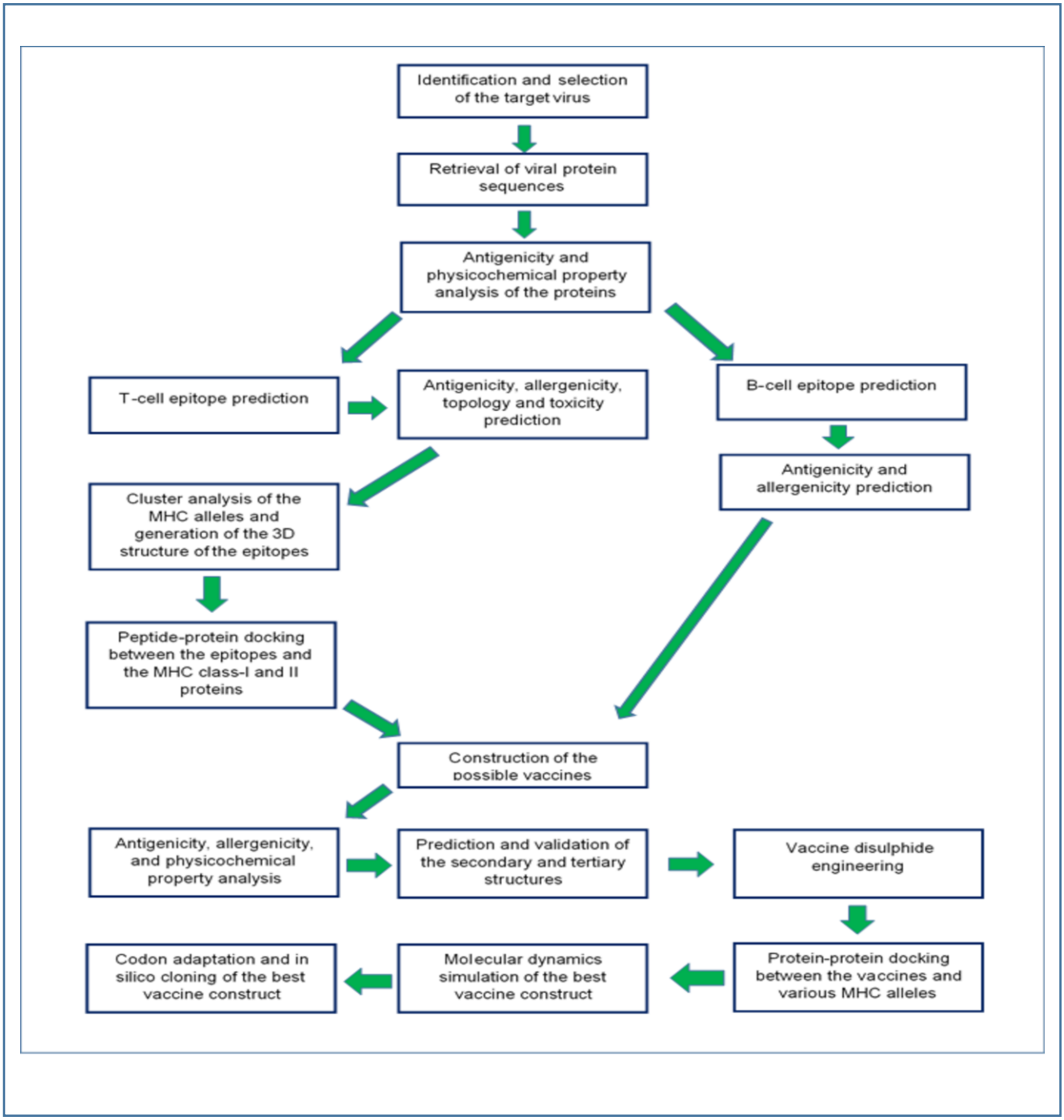

Figure 01. Strategies employed in the overall study

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2. Materials and Methods:

The current study was conducted to develop potential vaccines against the Wuhan novel coronavirus (strain 2019-nCoV) (Wuhan seafood market pneumonia virus) exploiting the strategies of reverse vaccinology (Figure 01).

2.1. Strain Identification and Selection

The strain of the Wuhan novel coronavirus 2019 was selected by reviewing various entries of the online database of National Center for Biotechnology Information or NCBI (https://www.ncbi.nlm.nih.gov/).

2.2. Retrieval of the Protein Sequences

Four protein sequences i.e., nucleocapsid phosphoprotein (PubMed accession no: QHD43423.2), membrane glycoprotein (PubMed accession no: QHD43419.1), ORF3a protein (PubMed accession no: QHD43417.1) and surface glycoprotein (PubMed accession no: QHD43416.1) were downloaded from the NCBI (https://www.ncbi.nlm.nih.gov/) database in fasta format.

2.3. Antigenicity Prediction and Physicochemical Property Analysis of the Protein Sequences

The antigenicity of protein sequences was determined using an online tool called VaxiJen v2.0 (http://www.ddg-pharmfac.net/vaxijen/VaxiJen/VaxiJen.htm). During the antigenicity prediction, the threshold was set at 0.4 in the tumor model [89]-[91]. Only the protein sequences that were found to be highly antigenic, were selected for further analysis. Next, the selected antigenic protein sequences were analyzed by ExPASy's online tool and ProtParam (https://web.expasy.org/protparam/) to determine their physicochemical properties [92]. Different physicochemical properties i.e., the number of amino acids, molecular weight, theoretical pI,

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extinction co-efficient, estimated half-life, aliphatic index and grand average of hydropathicity (GRAVY) were then determined.

2.4. T-cell and B-cell Epitope Prediction

The online epitope prediction server Immune Epitope Database or IEDB (https://www.iedb.org/) was used for T-cell and B-cell epitope prediction. The database contains a huge collection of experimental data on T-cell epitopes and antibodies. These experimental data are collected from various experiments that are conducted on human, non-human primates and other animals [93]. The NetMHCpan EL 4.0 prediction method was used for MHC class-I restricted CD8+ cytotoxic T-lymphocyte (CTL) epitope prediction for HLA-A*11-01 allele and the MHC class-II restricted CD4+ helper T-lymphocyte (HTL) epitopes were predicted for HLA DRB1*04-01 allele, using the Sturniolo prediction method. Ten of the top twenty MHC class-I and MHC class-II epitopes were randomly selected based on their antigenicity scores (AS) and percentile scores. For, the B-cell lymphocytic epitopes (BCL), with amino acid number of more than ten, were selected and predicted using Bipipered linear epitope prediction method.

2.5. Transmembrane Topology and Antigenicity Prediction of the Selected Epitopes

The epitopes selected in the previous step were then used in the transmembrane topology experiment using the transmembrane topology of protein helices determinant, TMHMM v2.0 server (http://www.cbs.dtu.dk/services/TMHMM/) [94]. The antigenicity of the epitopes were determined again by the online VaxiJen v2.0 (http://www.ddg-pharmfac.net/vaxijen/VaxiJen/VaxiJen.htm) server and the threshold for the tumor model was kept at 0.4 for this time also.

2.6. Allergenicity and Toxicity Prediction of the Epitopes

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The allergenicity of the selected epitopes were predicted using two online tools i.e., AllerTOP v2.0 (https://www.ddg-pharmfac.net/AllerTOP/) as well as AllergenFP v1.0 (http://ddg-pharmfac.net/AllergenFP/). During the selection after the allergenicity prediction, the results predicted by the AllerTOP server were given more priority because the server has better accuracy (88.7%) than AllergenFP server of 87.9% [95][96]. The toxicity prediction of the selected epitopes was carried out using ToxinPred server (http://crdd.osdd.net/raghava/toxinpred/) using SVM (support vector method) based method, keeping all the parameters default. After the antigenicity, allergenicity and toxicity tests, the epitopes that were found to be antigenic, non-allergenic and non-toxic, were considered as the best possible epitopes and the sequences that were found to be highly antigenic as well as non-allergenic, were considered as B-cell epitopes and as a result used in the next phases of this experiment.

2.7. Cluster Analysis of the MHC Alleles

The cluster analysis of the MHC alleles was carried out to identify the alleles of the MHC class-I and class-II molecules with similar binding specificities. The cluster analysis of the MHC alleles were carried out using online tool MHCcluster 2.0 (http://www.cbs.dtu.dk/services/MHCcluster/) [97]. During the analysis, the number of peptides to be included was kept at 50,000, the number of bootstrap calculations were set to 100 and all the HLA supertype representatives (MHC class-I) as well as the HLA-DR representatives (MHC class-II) were also selected. The NetMHCpan-2.8 prediction method was used for analyzing the MHC class-I alleles. The server depicts the results in the forms of MHC specificity tree and MHC specificity heat-map.

2.9. Generation of the 3D Structures of the Selected Epitopes

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The 3D structures of the epitopes were generated using PEP-FOLD3 (http://bioserv.rpbs.univ-paris-diderot.fr/services/PEP-FOLD3/) server. Only the best selected epitopes from previous experiments were used for 3D structure generation [98]-[100].

2.10. Molecular Docking of the Selected Epitopes

The molecular docking study of the epitopes were conducted using PatchDock (https://bioinfo3d.cs.tau.ac.il/PatchDock/php.php) server against HLA-A*11-01 allele (PDB ID: 5WJL) and HLA DRB1*04-01 (PDB ID: 5JLZ). PatchDock tool is an online docking tool that works on specific algorithms. The algorithms divide the connolly dot surface representations of the compounds into concave and convex patches as well as flat patches. Next, the complementary patches are matched for generating candidate transformations as well as potential solutions. Finally, an RMSD (root mean square deviation) score is applied to the candidate solutions. Thereafter, the redundant solutions are discarded based on analyzing the RMSD score. The top scored solutions are considered as the top ranked solutions by the server. After successful docking by PatchDock, the refinement and re-scoring of the docking results were carried out by the FireDock server (http://bioinfo3d.cs.tau.ac.il/FireDock/php.php). After refinement of the docking scores, the FireDock server generates global energies for the best solutions and ranks them based on the generated global energies and the lowest global energy is always considered as the best docking score [101]-[104]. The best results were visualized using Discovery Studio Visualizer [105].

2.11. Vaccine Construction

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Three possible vaccines were constructed against the selected Wuhan novel coronavirus. The predicted CTL, HTL and BCL epitopes were joined together by linkers for vaccine construction. The vaccines were constructed maintaining the sequence i.e., adjuvant, PADRE sequence, CTL epitopes, HTL epitopes and BCL epitopes. Three different adjuvants i.e., beta defensin, L7/L12 ribosomal protein and HABA protein (*Mycobacterium tuberculosis*, accession number: AGV15514.1), were used and the three vaccines differ from each other by their adjuvant sequence. Beta-defensin adjuvant acts as agonist and stimulate the activation of the toll like receptors (TLRs): 1, 2 and 4. Studies have showed that L7/L12 ribosomal protein and HABA protein also activate TLR-4. During the vaccine construction, EAAAK linkers were used to connect the adjuvant with the PADRE sequence, GGGS linkers were used to attach the PADRE sequence with the CTL epitopes and the CTL epitopes with the other CTL epitopes, GPGPG linkers were used to conjugate the CTL epitopes with the HTL epitopes and also the HTL epitopes themselves. The KK linkers were used to attach the HTL epitopes with the BCL epitopes. Studies have showed that, the PADRE sequence stimulates the cytotoxic T-lymphocyte response of the vaccines that contain it [106]-[116].

2.11. Antigenicity, Allergenicity and Physicochemical Property Analysis of the Constructed Vaccines

The antigenicity of the constructed vaccines was predicted by the online server VaxiJen v2.0 (http://www.ddg-pharmfac.net/vaxijen/VaxiJen/VaxiJen.htm) using the tumor model where the threshold was kept at 0.4. The allergenicity of the predicted vaccines were determined by two online tools i.e., AlgPred (http://crdd.osdd.net/raghava/algpred/) and AllerTop v2.0 (https://www.ddg-pharmfac.net/AllerTOP/). The AlgPred server predicts the possible allergens

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based on similarity of the known epitope of any of the known region of a protein [117]. MEME/MAST motif prediction was used to predict the allergenicity of the vaccines by AlgPred. Later, the online server ProtParam (https://web.expasy.org/protparam/) was used to predict different physicochemical properties of the constructed vaccines.

2.12. Secondary and Tertiary Structure Prediction of the Vaccine Constructs

The secondary structure generation of the vaccine constructs were performed using online server PRISPRED (http://bioinf.cs.ucl.ac.uk/psipred/), which is a simple and easy secondary structure generating tool and can predict the transmembrane topology, transmembrane helix, fold and domain recognition with certain degrees of accuracy[118][119]. The secondary structures of the vaccine constructs were predicted using the PRISPRED 4.0 prediction method. Next, another online tool, NetTurnP v1.0 (http://www.cbs.dtu.dk/services/NetTurnP/) was used to predict the β-sheet structure of the vaccines [120]. The 3D structures of the vaccines were then generated using online server RaptorX (http://raptorx.uchicago.edu/) [121]-[123].

2.13. 3D Structure Refinement and Validation

The 3D structures of the constructed vaccines were refined using online protein refinement tool, 3Drefine (http://sysbio.rnet.missouri.edu/3Drefine/) [124]. For each of the vaccine, the refined model 1 was downloaded for validation. The validation was then performed by analyzing the Ramachandran plots which were generated using the PROCHECK (https://servicesn.mbi.ucla.edu/PROCHECK/) server [125][126].

2.14. Vaccine Protein Disulfide Engineering

The vaccine protein disulfide engineering was carried out with the aid of Disulfide by Design 2 v12.2 (http://cptweb.cpt.wayne.edu/DbD2/) server which predicts the potential sites within a

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protein structure that have higher possibility of undergoing disulfide bond formation [127]. During disulfide engineering, the intra-chain, inter-chain and C_β for glycine residue were selected and the χ_3 Angle was kept -87° or +97° ± 5 and C_α-C_β-S_γ Angle was kept 114.6° ±10.

2.15. Protein-Protein Docking

In protein-protein docking, the constructed Wuhan novel coronavirus 2019 vaccines were analyzed by docking against some MHC alleles as well as the toll like receptor (TLR) and then one best vaccine was selected based on the docking scores. During viral infections, the MHC complex recognize the viral particles as potent antigens. The various portions of the MHC molecules are encoded by different MHC alleles. For this reason, the constructed vaccines should have very good binding affinity with these segments of the MHC complex that are encoded by different alleles [128]. In this experiment, all the vaccines constructs were docked against DRB1*0101 (PDB ID: 2FSE), DRB3*0202 (PDB ID: 1A6A), DRB5*0101 (PDB ID: 1H15), DRB3*0101 (PDB ID: 2Q6W), DRB1*0401 (PDB ID: 2SEB), and DRB1*0301 (PDB ID: 3C5J) alleles. Moreover, it has been proved that TLR-8 is responsible for mediating the immune responses against the RNA viruses and TLR-3 is responsible for mediating immune responses against the DNA viruses [129][130]. The coronavirus is an RNA virus [131]. For this reason, the vaccine constructs of Wuhan novel coronavirus were docked against TLR-8 (PDB ID: 3W3M). The protein-protein docking study was conducted using different online docking tools to improve the prediction accuracy of the docking study. At first, the docking was performed by ClusPro 2.0 (https://cluspro.bu.edu/login.php). The server ranks the docked complexes based on their center scores and lowest energy scores. However, these scores do not correspond to the actual binding affinity of the proteins with their targets [132]-[134]. The binding affinity (ΔG in kcal mol^{-1}) of the docked complexes were later generated and analyzed by PRODIGY tool of HADDOCK

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webserver (https://haddock.science.uu.nl/). The lower binding energy represents the higher binding affinity and vice versa [135]-[137]. After that, the docking was again carried out by PatchDock (https://bioinfo3d.cs.tau.ac.il/PatchDock/php.php) server and later refined and re-scored by FireDock server (http://bioinfo3d.cs.tau.ac.il/FireDock/php.php). The FireDock server ranks the docked complexes based on their global energy and the lower global energy represents the better result. Finally, the docking was conducted using HawkDock server (http://cadd.zju.edu.cn/hawkdock/). At this stage, the Molecular Mechanics/Generalized Born Surface Area (MM-GBSA) study was also calculated by the HawkDock server which works on specific algorithm that depicts that, the lower score and lower energy represent the better scores [138]-[141]. For each of the vaccines and their respective targets, the score of model 1 was taken for analysis as well as the model 1 of every complex was taken into consideration for MM-GBSA study. From the docking experiment, one best vaccine was selected. The docked structures were visualized by PyMol tool [142].

2.16. Molecular Dynamic Simulation

The molecular dynamics (MD) simulation study was carried out for only the best selected vaccine. The online MD simulation tool, iMODS (http://imods.chaconlab.org/) was used for molecular dynamics simulation study. iMODS server is a fast, online, user-friendly and effective molecular dynamics simulation server. The deformability, B-factor (mobility profiles), eigenvalues, variance, co-variance map and elastic network of the protein complex, are predicted quite efficiently by the server. For a protein complex, the deformability depends on the ability to deform at each of its amino acid. The eigenvalue represents the motion stiffness of the protein complex and the energy that is required to deform the structure. The lower eigenvalue represents easy deformability of the complex. The server is a fast and easy tool for determining and measuring the protein flexibility

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[143]-[147]. For analysing the molecular dynamics simulation, the CV-1-TLR-8 docked complex was used.

2.17. Codon Adaptation and In Silico Cloning

The best predicted vaccine from the previous steps, was reverse transcribed to a possible DNA sequence which is supposed to express the vaccine protein. The reverse transcribed DNA sequence was adapted according to the target organism, so that the cellular machinery of that organism could use the codons of the newly adapted DNA sequence efficiently for producing the desired vaccine. Codon adaptation is a necessary step of in silico cloning since an amino acid can be encoded by different codons in different organisms (codon biasness) and codon adaptation predicts the best codon for a specific amino acid that should work properly in a particular organism. The predicted protein sequence of the best selected vaccines were used for codon adaptation by the Java Codon Adaptation Tool or JCat server (http://www.jcat.de/). Eukaryotic *E. coli* strain K12 was selected at the JCat server and rho-independent transcription terminators, prokaryotic ribosome binding sites and SgrA1 and SphI cleavage sites of restriction enzymes, were avoided. The protein sequence was reverse translated to the optimized possible DNA sequence by the JCat server. The optimized DNA sequence was then taken and SgrA1 and SphI restriction sites were attached to the N-terminal and C-terminal sites, respectively. Finally, the SnapGene restriction cloning module was used to insert the newly adapted DNA sequence between the SgrA1 and SphI restriction sites of pET-19b vector [148]-[150].

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3. Results

3.1. Identification, Selection and Retrieval of Viral Protein Sequences

The Wuhan novel coronavirus (Wuhan seafood market pneumonia virus) was identified from the NCBI database (https://www.ncbi.nlm.nih.gov/). Four protein sequences i.e., Nucleocapsid Phosphoprotein (accession no: QHD43423.2), Membrane Glycoprotein (accession no: QHD43419.1), ORF3a Protein (accession no: QHD43417.1) and Surface Glycoprotein (accession no: QHD43416.1) were selected for possible vaccine construction and retrieved from the NCBI database in fasta format.

>QHD43423.2 nucleocapsid phosphoprotein [Wuhan seafood market pneumonia virus]
MSDNGPQNQRNAPRITFGGPSDSTGSNQNGERSGARSKQRRPQGLPNNTASWFTALTQ
HGKEDLKFPRGQGVPINTNSSPDDQIGYYRRATRRIRGGDGKMKDLSPRWYFYYLGTG
PEAGLPYGANKDGIIWVATEGALNTPKDHIGTRNPANNAAIVLQLPQGTTLPKGFYAEG
SRGGSQASSRSSSRSRNSSRNSTPGSSRGTSPARMAGNGGDAALALLLLDRLNQLESKM
SGKGQQQQGQTVTKKSAAEASKKPRQKRTATKAYNVTQAFGRRGPEQTQGNFGDQEL
IRQGTDYKHWPQIAQFAPSASAFFGMSRIGMEVTPSGTWLTYTGAIKLDDKDPNFKDQV
ILLNKHIDAYKTFPPTEPKKDKKKKADETQALPQRQKKQQTVTLLPAADLDDFSKQLQQ
SMSSADSTQA

>QHD43419.1 membrane glycoprotein [Wuhan seafood market pneumonia virus]
MADSNGTITVEELKKLLEQWNLVIGFLFLTWICLLQFAYANRNRFLYIIKLIFLWLLWPV
TLACFVLAAVYRINWITGGIAIAMACLVGLMWLSYFIASFRLFARTRSMWSFNPETNILL
NVPLHGTILTRPLLESELVIGAVILRGHLRIAGHHLGRCDIKDLPKEITVATSRTLSYYKLG
ASQRVAGDSGFAAYSRYRIGNYKLNTDHSSSSDNIALLVQ

>QHD43417.1 ORF3a protein [Wuhan seafood market pneumonia virus]
MDLFMRIFTIGTVTLKQGEIKDATPSDFVRATATIPIQASLPFGWLIVGVALLAVFQSASK
IITLKKRWQLALSKGVHFVCNLLLLFVTVYSHLLLVAAGLEAPFLYLYALVYFLQSINFV
RIIMRLWLCWKCRSKNPLLYDANYFLCWHTNCYDYCIPYNSVTSSIVITSGDGTTSPISE
HDYQIGGYTEKWESGVKDCVVLHSYFTSDYYQLYSTQLSTDTGVEHVTFFIYNKIVDEP
EEHVQIHTIDGSSGVVNPVMEPIYDEPTTTTSVPL

>QHD43416.1 surface glycoprotein [Wuhan seafood market pneumonia virus]
MFVFLVLLPLVSSQCVNLTTRTQLPPAYTNSFTRGVYYPDKVFRSSVLHSTQDLFLPFFS
NVTWFHAIHVSGTNGTKRFDNPVLPFNDGVYFASTEKSNIIRGWIFGTTLDSKTQSLLIV
NNATNVVIKVCEFQFCNDPFLGVYYHKNNKSWMESEFRVYSSANNCTFEYVSQPFLMD

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LEGKQGNFKNLREFVFKNIDGYFKIYSKHTPINLVRDLPQGFSALEPLVDLPIGINITRFQT
LLALHRSYLTPGDSSSGWTAGAAAYYVGYLQPRTFLLKYNENGTITDAVDCALDPLSET
KCTLKSFTVEKGIYQTSNFRVQPTESIVRFPNITNLCPFGEVFNATRFASVYAWNRKRISN
CVADYSVLYNSASFSTFKCYGVSPTKLNDLCFTNVYADSFVIRGDEVRQIAPGQTGKIA
DYNYKLPDDFTGCVIAWNSNNLDSKVGGNYNYLYRLFRKSNLKPFERDISTEIYQAGST
PCNGVEGFNCYFPLQSYGFQPTNGVGYQPYRVVVLSFELLHAPATVCGPKKSTNLVKN
KCVNFNFNGLTGTGVLTESNKKFLPFQQFGRDIADTTDAVRDPQTLEILDITPCSFGGVS
VITPGTNTSNQVAVLYQDVNCTEVPVAIHADQLTPTWRVYSTGSNVFQTRAGCLIGAEH
VNNSYECDIPIGAGICASYQTQTNSPRRARSVASQSIIAYTMSLGAENSVAYSNNSIAIPT
NFTISVTTEILPVSMTKTSVDCTMYICGDSTECSNLLLQYGSFCTQLNRALTGIAVEQDK
NTQEVFAQVKQIYKTPPIKDFGGFNFSQILPDPSKPSKRSFIEDLLFNKVTLADAGFIKQY
GDCLGDIAARDLICAQKFNGLTVLPPLLTDEMIAQYTSALLAGTITSGWTFGAGAALQIP
FAMQMAYRFNGIGVTQNVLYENQKLIANQFNSAIGKIQDSLSSTASALGKLQDVVNQN
AQALNTLVKQLSSNFGAISSVLNDILSRLDKVEAEVQIDRLITGRLQSLQTYVTQQLIRAA
EIRASANLAATKMSECVLGQSKRVDFCGKGYHLMSFPQSAPHGVVFLHVTYVPAQEKN
FTTAPAICHDGKAHFPREGVFVSNGTHWFVTQRNFYEPQIITTDNTFVSGNCDVVIGIVN
NTVYDPLQPELDSFKEELDKYFKNHTSPDVDLGDISGINASVVNIQKEIDRLNEVAKNLN
ESLIDLQELGKYEQYIKWPWYIWLGFIAGLIAIVMVTIMLCCMTSCCSCLKGCCSCGSCC
KFDEDDSEPVLKGVKLHYT

3.2. Antigenicity Prediction and Physicochemical Property Analysis of the Protein Sequences

Two proteins: nucleocapsid phosphoprotein and surface glycoprotein, were identified as potent antigens and hence used in the next phases of the experiment (**Table 01**). The physicochemical property analysis was conducted for these two selected proteins. Nucleocapsid phosphoprotein had the highest theoretical pI of 10.07, however, surface glycoprotein had the highest extinction co-efficient of 148960 M^{-1} cm^{-1}. Both of them were found to have similar half-life of 30 hours. However, surface glycoprotein had the highest aliphatic index and grand average of hydropathicity (GRAVY) value among the two proteins (**Table 02**).

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Table 01. The antigenicity determination of the nine selected proteins.

Name of the protein	**Antigenicity (threshold = 0.4; tumor model)**
Nucleocapsid phosphoprotein	Antigenic (0.709)
Membrane glycoprotein	Non-antigenic (0.166)
ORF3a protein	Non-antigenic (0.372)
Surface glycoprotein	Antigenic (0.534)

Table 02. The antigenicity and physicochemical property analysis of the selected viral proteins.

Name of the protein sequence	**Total amino acids**	**Molecular weight**	**Theoretical pI**	**Ext. coefficient (in M^{-1} cm^{-1})**	**Est. half-life (in mammalian cell)**	**Aliphatic index**	**Grand average of hydropathicity (GRAVY)**
Nucleocapsid phosphoprotein	419	45625.70	10.07	43890	30 hours	52.53	-0.971
Surface glycoprotein	1273	141178.47	6.24	148960	30 hours	84.67	-0.079

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3.3. T-cell and B-cell Epitope Prediction and their Antigenicity, Allergenicity and Topology Determination

The MHC class-I and MHC class-II epitopes were determined for potential vaccine construction. The IEDB (https://www.iedb.org/) server generates a good number of epitopes. However, based on the antigenicity scores, ten epitopes were selected from the top twenty epitopes. Later, the epitopes with high antigenicity, non-allergenicity and non-toxicity were selected for vaccine construction. The B-cell epitopes were also determined using IEDB (https://www.iedb.org/) server and based on their antigenicity, non-allergenicity and length (the sequences with more than 10 amino acids) best predicted epitopes were selected for vaccine construction. The topology of the epitopes were also determined and the transmembrane topology experiment revealed that most of the epitopes might remain inside of the cell membrane. Three MHC class-I epitopes i.e., AGLPYGANK, AADLDDFSK and QLESKMSGK, five MHC class-II epitopes QELIRQGTDYKH, LIRQGTDYKHWP, RLNQLESKMSGK, LNQLESKMSGKG and LDRLNQLESKMS of nucleocapsid phosphoprotein were selected based on the selection criteria. Agains four MHC class-I and three MHC class-II epitopes i.e., SVLNDILSR, GVLTESNKK, RLFRKSNLK, QIAPGQTGK, TSNFRVQPTESI, SNFRVQPTESIV and LLIVNNATNVVI were selected for surface glycoprotein.

Table 03 and **Table 04** list the potential T-cell epitopes of nucleocapsid phosphoprotein and **Table 05** and **Table 06** list the potential T-cell epitopes of surface glycoprotein. **Table 07** lists the predicted B-cell epitopes of the two proteins.

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Table 03. MHC class-I epitope prediction and topology, antigenicity, allergenicity and toxicity analysis of the epitopes of nucleocapsid phosphoprotein. AS: Antigenic Score.

Epitope	Start	End	Topology	AS	Percentile scores	Antigenicity (tumor model, threshold= 0.4)	Allergenicity	Toxicity
AGLPYGANK	119	127	Outside	0.547	0.28	Antigenic	Non-allergenic	Non-toxic
KTFPPTEPK	361	369	Outside	0.967	0.01	Non-antigenic	Non-allergenic	Non-toxic
AADLDDFSK	397	405	Outside	0.235	0.96	Antigenic	Non-allergenic	Non-toxic
KSAAEASKK	249	257	Inside	0.670	0.17	Antigenic	Allergenic	Non-toxic
TQALPQRQK	379	387	Inside	0.349	0.60	Non-antigenic	Allergenic	Non-toxic
SSRGTSPAR	201	209	Inside	0.166	1.40	Antigenic	Allergenic	Non-toxic
ASWFTALTQ	50	58	Inside	0.115	1.80	Antigenic	Allergenic	Non-toxic
QLESKMSGK	229	237	Inside	0.084	2.20	Antigenic	Non-allergenic	Non-toxic
KDQVILLNK	347	355	Inside	0.082	2.30	Antigenic	Allergenic	Non-toxic
GTTLPKGFY	164	172	Outside	0.169	1.30	Non-antigenic	Allergenic	Non-toxic

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Table 04. MHC class-II epitope prediction and topology, antigenicity, allergenicity and toxicity analysis of the epitopes of nucleocapsid phosphoprotein. AS: Antigenic Score.

Epitope	Start	End	Topology	AS	Percentile scores	Antigenicity (tumor model, threshold= 0.4)	Allergenicity	Toxicity
QELIRQGTDYKH	289	300	Inside	2.90	3.60	Antigenic	Non-allergenic	Non-toxic
ELIRQGTDYKHW	290	301	Inside	2.90	3.60	Antigenic	Allergenic	Non-toxic
DQELIRQGTDYK	288	299	Inside	2.90	3.60	Antigenic	Allergenic	Non-toxic
SRIGMEVTPSGT	318	329	Inside	2.60	4.80	Non-antigenic	Non-allergenic	Non-toxic
LIRQGTDYKHWP	291	302	Inside	2.90	3.60	Antigenic	Non-allergenic	Non-toxic
RLNQLESKMSGK	226	237	Inside	2.00	8.10	Antigenic	Non-allergenic	Non-toxic
WFTALTQHGKED	52	63	Inside	1.70	11.0	Antigenic	Allergenic	Non-toxic
LNQLESKMSGKG	227	238	Outside	2.00	8.10	Antigenic	Non-allergenic	Non-toxic
LDRLNQLESKMS	224	235	Inside	2.00	8.10	Antigenic	Non-allergenic	Non-toxic
WFTALTQHG	49	60	Outside	1.70	11.0	Antigenic	Allergenic	Non-toxic

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Table 05. MHC class-I epitope prediction and topology, antigenicity, allergenicity and toxicity analysis of the epitopes of surface glycoprotein. AS: Antigenic Score.

Epitope	Start	End	Topology	AS	Percentile scores	Antigenicity (tumor model, threshold= 0.4)	Allergenicity	Toxicity
GVYFASTEK	89	97	Inside	0.938	0.01	Non-antigenic	Non-allergenic	Non-toxic
ASANLAATK	1020	1028	Inside	0.911	0.01	Non-antigenic	Allergenic	Non-toxic
SVLNDILSR	975	983	Inside	0.849	0.04	Antigenic	Non-allergenic	Non-toxic
GVLTESNKK	550	558	Inside	0.731	0.12	Antigenic	Non-allergenic	Non-toxic
SSTASALGK	939	947	Outside	0.779	0.09	Antigenic	Allergenic	Non-toxic
GTHWFVTQR	1099	1107	Inside	0.776	0.09	Non-antigenic	Allergenic	Non-toxic
EILPVSMTK	725	733	Inside	0.773	0.09	Non-antigen	Allergen	Non-toxic
ALDPLSETK	292	300	Outside	0.679	0.16	Antigenic	Allergenic	Non-toxic
RLFRKSNLK	454	462	Inside	0.677	0.16	Antigenic	Non-allergenic	Non-toxic
QIAPGQTGK	409	417	Inside	0.674	0.16	Antigenic	Non-allergenic	Non-toxic

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Table 06. MHC class-II epitope prediction and topology, antigenicity, allergenicity and toxicity analysis of the epitopes of surface glycoprotein. AS: Antigenic Score.

Epitope	Start	End	Topology	AS	Percentile scores	Antigenicity (tumor model, threshold= 0.4)	Allergenicity	Toxicity
FLGVYYHKNNKS	140	151	Inside	4.40	0.45	Non-antigenic	Allergenic	Non-toxic
TSNFRVQPTESI	315	326	Inside	5.30	0.08	Antigenic	Non-allergenic	Non-toxic
VYYHKNNKSWME	143	154	Inside	4.40	0.45	Non-antigenic	Allergenic	Non-toxic
NFRVQPTESIVR	317	328	Inside	5.30	0.08	Antigenic	Allergenic	Non-toxic
GVFVSNGTHWFV	1093	1104	Outside	4.10	0.78	Non-antigenic	Allergenic	Non-toxic
SNFRVQPTESIV	316	327	Inside	5.30	0.08	Antigenic	Non-allergenic	Non-toxic
LLIVNNATNVVI	117	128	Inside	4.30	0.58	Antigenic	Non-allergenic	Non-toxic
EGVFVSNGTHWF	1092	1103	Outside	4.10	0.78	Non-antigenic	Allergenic	Non-toxic
VFVSNGTHWFVT	1094	1105	Outside	4.10	0.78	Non-antigenic	Non-allergenic	Non-toxic
IVNNATNVVIKV	119	130	Inside	4.30	0.58	Antigenic	Allergenic	Non-toxic

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Table 07. B-cell epitope prediction and topology, antigenicity, allergenicity and toxicity analysis of the epitopes of nucleocapsid phosphoprotein and surface glycoprotein.

Nucleocapsid phosphoprotein			**Surface glycoprotein**		
Epitope	**Antigenicity (tumor model, threshold= 0.4)**	**Allergenicity**	**Epitope**	**Antigenicity (tumor model, threshold= 0.4)**	**Allergenicity**
MSDNGPQNQRNAPRITFGGPSDSTGSNQNGERSGARSKQRRPQGLPNNTAS	Antigenic	Non-allergenic	RTQLPPAYTNS	Non-antigenic	Allergenic
RIRGGDGKMKDL	Antigenic	Non-allergenic	LTPGDSSSGWTAG	Antigenic	Non-allergenic
TGPEAGLPYGANK	Antigenic	Non-allergenic	VRQIAPGQTGKIAD	Antigenic	Non-allergenic
GTTLPKGFYAEGSRGGSQASSRSSSRSRNSSRNSTPGSSRGTSPARMAGNGGD	Antigenic	Non-allergenic	SGTNGTKRFDN	Antigenic	Allergenic
QHGKEDLKFPRGQGVPINTNSSPDDQIG	Non-antigenic	Non-allergenic	YQAGSTPCNGV	Antigenic	Non-allergenic
SKMSGKGQQQQGQTVTKKSAAEASKKPRQKRTATKAYN	Antigenic	Non-allergenic	QTQTNSPRRARSV	Antigenic	Non-allergenic
KTFPPTEPKKDKKKKADETQALPQRQKKQQ	Antigenic	Non-allergenic	ILPDPSKPSKRS	Antigenic	Non-allergenic
AFGRRGPEQTQGNFG	Non-antigenic	Allergenic	YGFQPTNGVGYQ	Non-antigenic	Allergenic
-	-	-	RDIADTTDAVRDPQ	Antigenic	Allergenic
-	-	-	VYDPLQPELDSF	Antigenic	Allergenic
-	-	-	KNHTSPDVDLG	Non-antigenic	Non-allergenic

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3.4. Cluster Analysis of the MHC Alleles

The online tool MHCcluster 2.0 (http://www.cbs.dtu.dk/services/MHCcluster/), was used for the cluster analysis of the possible MHC class-I and MHC class-II alleles that may interact with the selected epitopes during the immune responses. The tool illustrates the relationship of the clusters of the alleles in phylogenetic manner. **Figure 02** depicts the result of the cluster analysis where the red zone indicates strong interaction and the yellow zone corresponds to weaker interaction.

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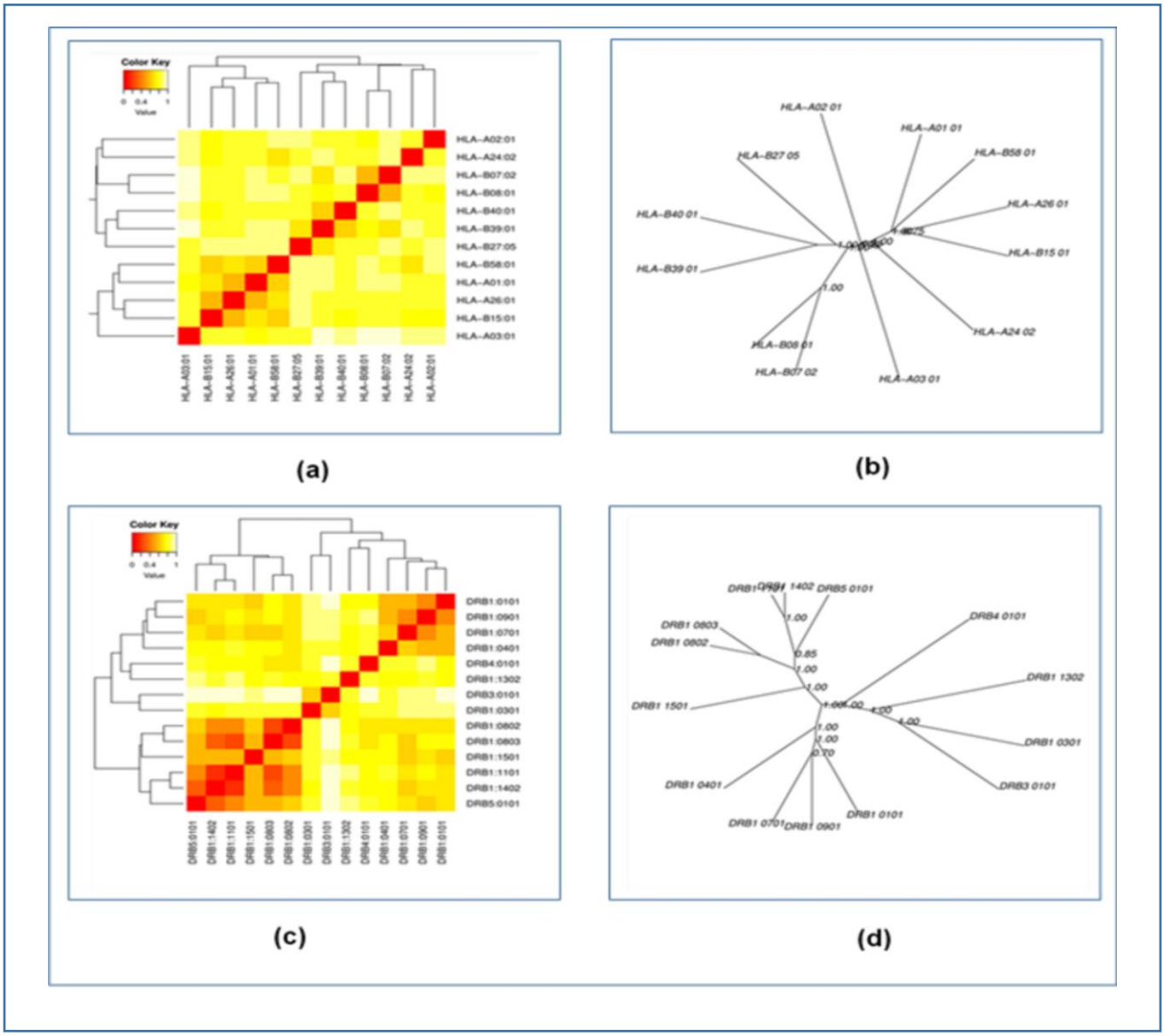

Figure 02. The results of the MHC cluster analysis. Here, (a) is the heat map of MHC class-I cluster analysis, (b) is the tree map of MHC class-I cluster analysis, (c) is the heat map of MHC class-II cluster analysis, (d) is the tree map of MHC class-II cluster analysis.

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3.5. Generation of the 3D Structures of the Epitopes and Peptide-Protein Docking

The peptide-protein docking was conducted to find out, whether all the epitopes had the ability to bind with the MHC class-I as well as MHC class-II molecules or not. The HLA-A*11-01 allele (PDB ID: 5WJL) was used as the receptor for docking with the MHC class-I epitopes and HLA-DRB1*04-01 (PDB ID: 5JLZ) was used as the receptor for docking with the MHC class-II epitopes. Among the MHC class-I epitopes of nucleocapsid phosphoprotein, QLESKMSGK showed the best result with the lowest global energy of -53.28. Among the MHC class-II epitopes of nucleocapsid phosphoprotein, LIRQGTDYKHWP generated the lowest and best global energy score of 16.44. GVLTESNKK generated the best global energy score of -34.60 of the MHC class-I epitopes of surface glycoprotein. Among the MHC class-II epitopes of surface glycoprotein, TSNFRVQPTESI generated the best global energy score of -2.28 (**Table 08 & Figure 03**).

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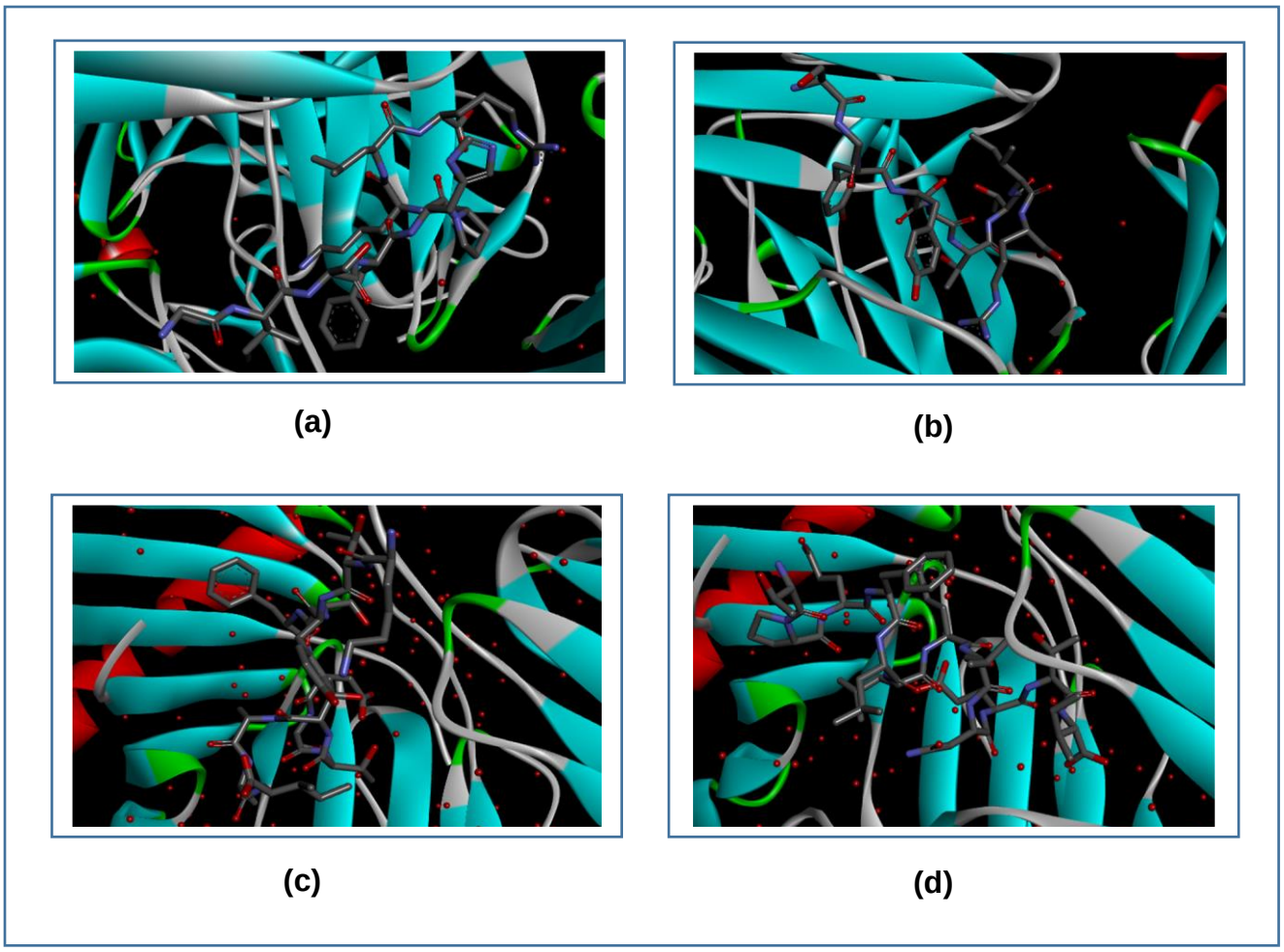

Figure 03. The best poses of interactions between the selected epitopes from the two proteins and their respective receptors. Here, (a) is the interaction between QLESKMSGK and MHC class-I, (b) is the interaction between GVLTESNKK and MHC class-I, (c) is the interaction between LIRQGTDYKHWP and MHC class-II, (d) is the interaction between TSNFRVQPTESI and MHC class-II. The interactions were visualized by Discovery Studio Visualizer.

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Table 08. Results of molecular docking analysis of the selected epitopes.

Name of the protein	Epitope	MHC allele	Global energy	Hydrogen bond energy	Epitope	MHC allele	Global energy	Hydrogen bond energy
Nucleocapsid phosphoprotein	AGLPYGANK	HLA-A*11-01 allele (PDB ID: 5WJL)	-38.28	-1.53	QELIRQGTDYKH	HLA DRB1*04-01 (PDB ID: 5JLZ)	14.62	-1.09
	AADLDDFSK		-15.60	-2.69	LIRQGTDYKHWP		-16.44	-1.66
	QLESKMSGK		-53.28	-4.32	RLNQLESKMSGK		-12.34	-3.41
	-		-	-	LNQLESKMSGKG		14.37	-9.20
	-		-	-	LDRLNQLESKMS		-1.35	0.00
Surface glycoprotein	SVLNDILSR		-27.72	-3.74	TSNFRVQPTESI		-2.28	0.00
	GVLTESNKK		-34.60	-3.64	SNFRVQPTESIV		1.82	0.00
	RLFRKSNLK		-27.48	-4.77	LLIVNNATNVVI		1.38	0.00
	QIAPGQTGK		-26.86	-0.89	-		-	-

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3.7. Vaccine Construction

After successful docking, three vaccines were constructed using the selected epitopes which is supposed to be directed to fight against the Wuhan Novel Coronavirus. To construct the vaccines, three different adjuvants were used i.e., beta defensin, L7/L12 ribosomal protein and HABA protein. PADRE sequence was also used in the vaccine construction process and added after the initial adjuvant sequence. Different linkers i.e., EAAAK, GGGS, GPGPG and KK linkers were used at their appropriate positions and each of the vaccine constructs was ended by an additional GGGS linker. The newly constructed vaccines were designated as: CV-1, CV-2 and CV-3 (**Table 09**).

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Table 09. The three constructed Wuhan Novel Coronavirus vaccine constructs. In the vaccine sequences, the linkers are bolded for easy visualization.

Name of the vaccines	Vaccine constructs	Number of amino acids
Coronavirus vaccine-1 (CV-1)	**EAAAK**GIINTLQKYYCRVRGGRCAVLSCLPKEEQIGKCSTRGRKCCRRKK**EAAAK**AKFVAAWTLKAAA**GGGS** AGLPYGANK**GGGS**AADLDDFSK**GGGS**QLESKMSGK**GGGS**SVLNDILSR**GGGS**GVLTESNKK**GGGS**RLFRKS NLK**GGGS**QIAPGQTGK**GPGPG**QELIRQGTDYKH**GPGPG**LIRQGTDYKHWP**GPGPG**RLNQLESKMSGK**GPGP** **G**LNQLESKMSGKG**GPGPG**LDRLNQLESKMS**GPGPG**TSNFRVQPTESI**GPGPG**SNFRVQPTESIV**GPGPG**LLIVN NATNVVI**KK**MSDNGPQNQRNAPRITFGGPSDSTGSNQNGERSGARSKQRRPQGLPNNTAS**KK**RIRGGDGKMK DL**KK**TGPEAGLPYGANK**KK**GTTLPKGFYAEGSRGGSQASSRSSSRSRNSSRNSTPGSSRGTSPARMAGNGGD**K** **K**SKMSGKGQQQQGQTVTKKSAAEASKKPRQKRTATKAYN**KK**KTFPPTEPKKDKKKKADETQALPQRQKKQQ **KK**LTPGDSSSGWTAG**KK**VRQIAPGQTGKIAD**KK**YQAGSTPCNGV**KK**QTQTNSPRRARSV**KK**ILPDPSKPSKRS **KK**AKFVAAWTLKAAA**GGGS**	596
Coronavirus vaccine-2 (CV-2)	**EAAAK**MAKLSTDELLDAFKEMTLLELSDFVKKFEETFEVTAAAPVAVAAAGAAPAGAAVEAAEEQSEFDVILE AAGDKKIGVIKVVREIVSGLGLKEAKDLVDGAPKPLLEKVAKEAADEAKAKLEAAGATVTVK**EAAAK**AKFVA AWTLKAAA**GGGS**AGLPYGANK**GGGS**AADLDDFSK**GGGS**QLESKMSGK**GGGS**SVLNDILSR**GGGS**GVLTESN KK**GGGS**RLFRKSNLK**GGGS**QIAPGQTGK**GPGPG**QELIRQGTDYKH**GPGPG**LIRQGTDYKHWP**GPGPG**RLNQ LESKMSGK**GPGPG**LNQLESKMSGKG**GPGPG**LDRLNQLESKMS**GPGPG**TSNFRVQPTESI**GPGPG**SNFRVQPTE SIV**GPGPG**LLIVNNATNVVI**KK**MSDNGPQNQRNAPRITFGGPSDSTGSNQNGERSGARSKQRRPQGLPNNTAS**K** **K**RIRGGDGKMKDL**KK**TGPEAGLPYGANK**KK**GTTLPKGFYAEGSRGGSQASSRSSSRSRNSSRNSTPGSSRGTSP ARMAGNGGD**KK**SKMSGKGQQQQGQTVTKKSAAEASKKPRQKRTATKAYN**KK**KTFPPTEPKKDKKKKADET QALPQRQKKQQ**KK**LTPGDSSSGWTAG**KK**VRQIAPGQTGKIAD**KK**YQAGSTPCNGV**KK**QTQTNSPRRARSV**K** **K**ILPDPSKPSKRS**KK**AKFVAAWTLKAAA**GGGS**	681
Coronavirus vaccine-3 (CV-3)	**EAAAK**MAENPNIDDLPAPLLAALGAADLALATVNDLIANLRERAEETRAETRTRVEERRARLTKFQEDLPEQFI ELRDKFTTEELRKAAEGYLEAATNRYNELVERGEAALQRLRSQTAFEDASARAEGYVDQAVELTQEALGTVAS QTRAVGERAAKLVGIEL**EAAAK**AKFVAAWTLKAAA**GGGS**AGLPYGANK**GGGS**AADLDDFSK**GGGS**QLESK MSGK**GGGS**SVLNDILSR**GGGS**GVLTESNKK**GGGS**RLFRKSNLK**GGGS**QIAPGQTGK**GPGPG**QELIRQGTDYK H**GPGPG**LIRQGTDYKHWP**GPGPG**RLNQLESKMSGK**GPGPG**LNQLESKMSGKG**GPGPG**LDRLNQLESKMS**GP** **GPG**TSNFRVQPTESI**GPGPG**SNFRVQPTESIV**GPGPG**LLIVNNATNVVI**KK**MSDNGPQNQRNAPRITFGGPSDST GSNQNGERSGARSKQRRPQGLPNNTAS**KK**RIRGGDGKMKDL**KK**TGPEAGLPYGANK**KK**GTTLPKGFYAEGSR GGSQASSRSSSRSRNSSRNSTPGSSRGTSPARMAGNGGD**KK**SKMSGKGQQQQGQTVTKKSAAEASKKPRQKRT ATKAYN**KK**KTFPPTEPKKDKKKKADETQALPQRQKKQQ**KK**LTPGDSSSGWTAG**KK**VRQIAPGQTGKIAD**KK** YQAGSTPCNGV**KK**QTQTNSPRRARSV**KK**ILPDPSKPSKRS**KK**AKFVAAWTLKAAA**GGGS**	710

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3.8. Antigenicity, Allergenicity and Physicochemical Property Analysis of the Vaccine Constructs

The results of the antigenicity, allergenicity and physicochemical property analysis are listed in **Table 10**. All the three vaccine constructs were found to be antigenic as well as non-allergenic. CV-3 had the highest molecular weight, extinction co-efficient and aliphatic index of 74505.61, 36900 M^{-1} cm^{-1} and 54.97 respectively. All of them had in vivo half-life of 1 hours and CV-2 had the highest GRAVY value of -0.830 among the three vaccines.

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Table 10. The antigenicity, allergenicity and physicochemical property analysis of the vaccine constructs. MW: Molecular Weight

Name of the vaccine constructs	**Total amino acids**	**Antigenicity (tumor model, threshold= 0.4)**	**Allergenicity**	**MW**	**Theoretical pI**	**Ext. coefficient (in M^{-1} cm^{-1})**	**Est. half-life (in mammalian cell)**	**Aliphatic index**	**Grand average of hydropathicity (GRAVY)**
CV-1	596	Antigenic	Non-allergenic	62038.16	10.69	35785	1 hours	46.26	-1.041
CV-2	681	Antigenic	Non-allergenic	70317.45	10.23	32430	1 hours	55.86	-0.830
CV-3	710	Antigenic	Non-allergenic	74505.61	10.31	36900	1 hours	54.97	-0.941

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3.9. Secondary and Tertiary Structure Prediction of the Vaccine Constructs

From the secondary structure analysis, it was determined that, the CV-1 had the highest percentage of the amino acids (67.1%) in the coil formation as well as the highest percentage of amino acids (8%) in the beta-strand formation. However, CV-3 had the highest percentage of 37.8% of amino acids in the alpha-helix formation (**Figure 04** and **Table 11**). CV-1 and CV-2 vaccines had 02 domains, whereas, CV-3 had only one domain. CV-3 had the highest p-value of 2.36e-04. The results of the 3D structure analysis are listed in **Table 12** and illustrated in **Figure 05.**

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Table 11. Results of the secondary structure analysis of the vaccine constructs.

Name of the vaccine	Alpha helix (percentage of amino acids)	Beta sheet (percentage of amino acids)	Coil structure (percentage of amino acids)
CV-1	25%	8%	67.1%
CV-2	31.6%	6.8%	61.6%
CV-3	37.8%	5%	57.2%

Table 12. Results of the tertiary structure analysis of the vaccine constructs.

Name of the vaccine	Number of the domains	p-value	PDB Id of the est. matched template
CV-1	02	2.37e-04	1kj6A
CV-2	02	6.35e-05	1dd3A
CV-3	01	2.36e-04	6cfeA

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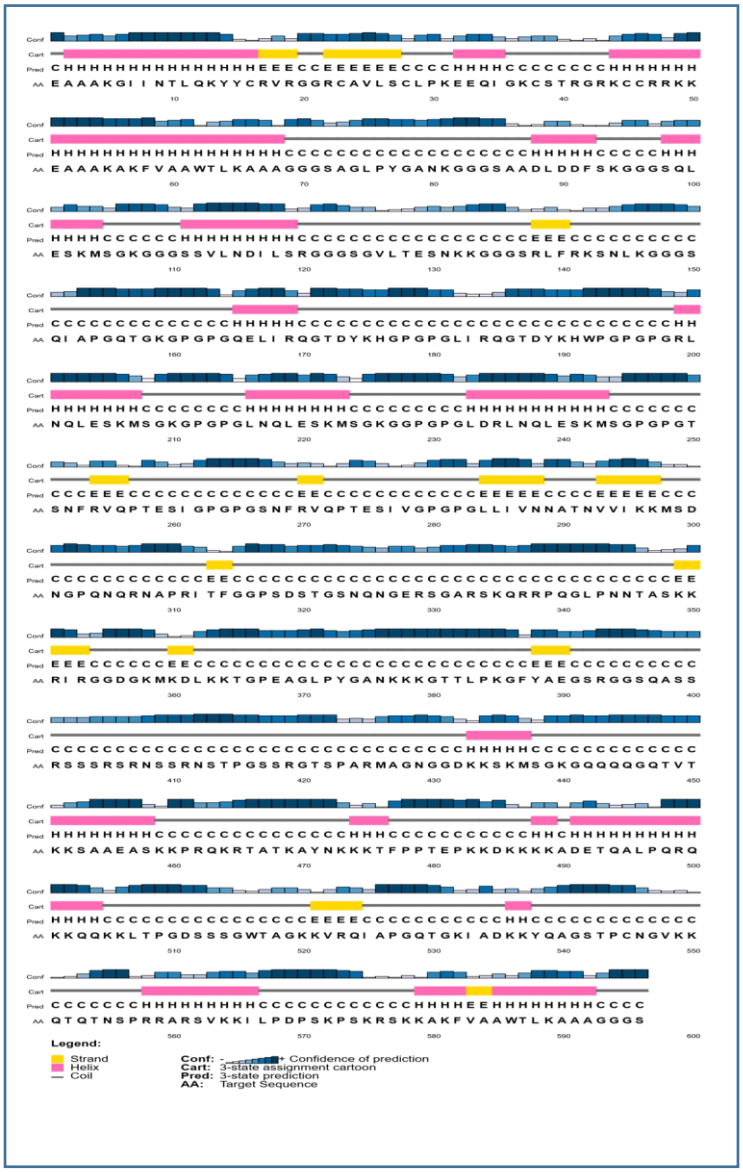
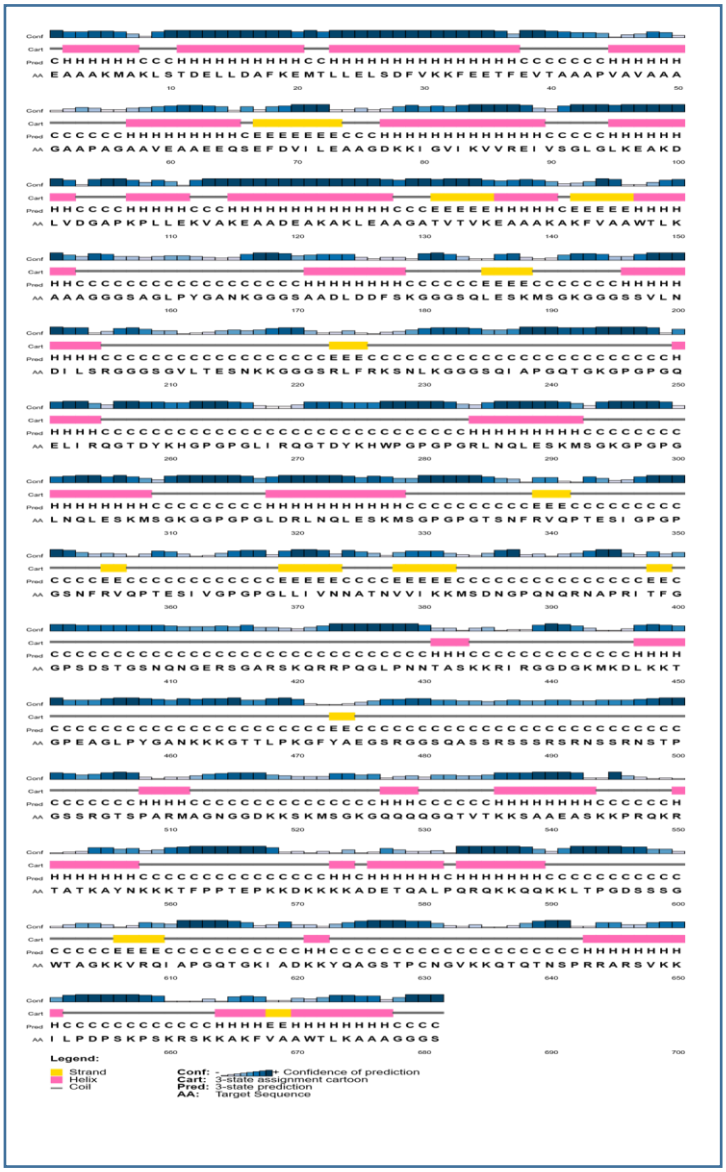


(a)



(b)

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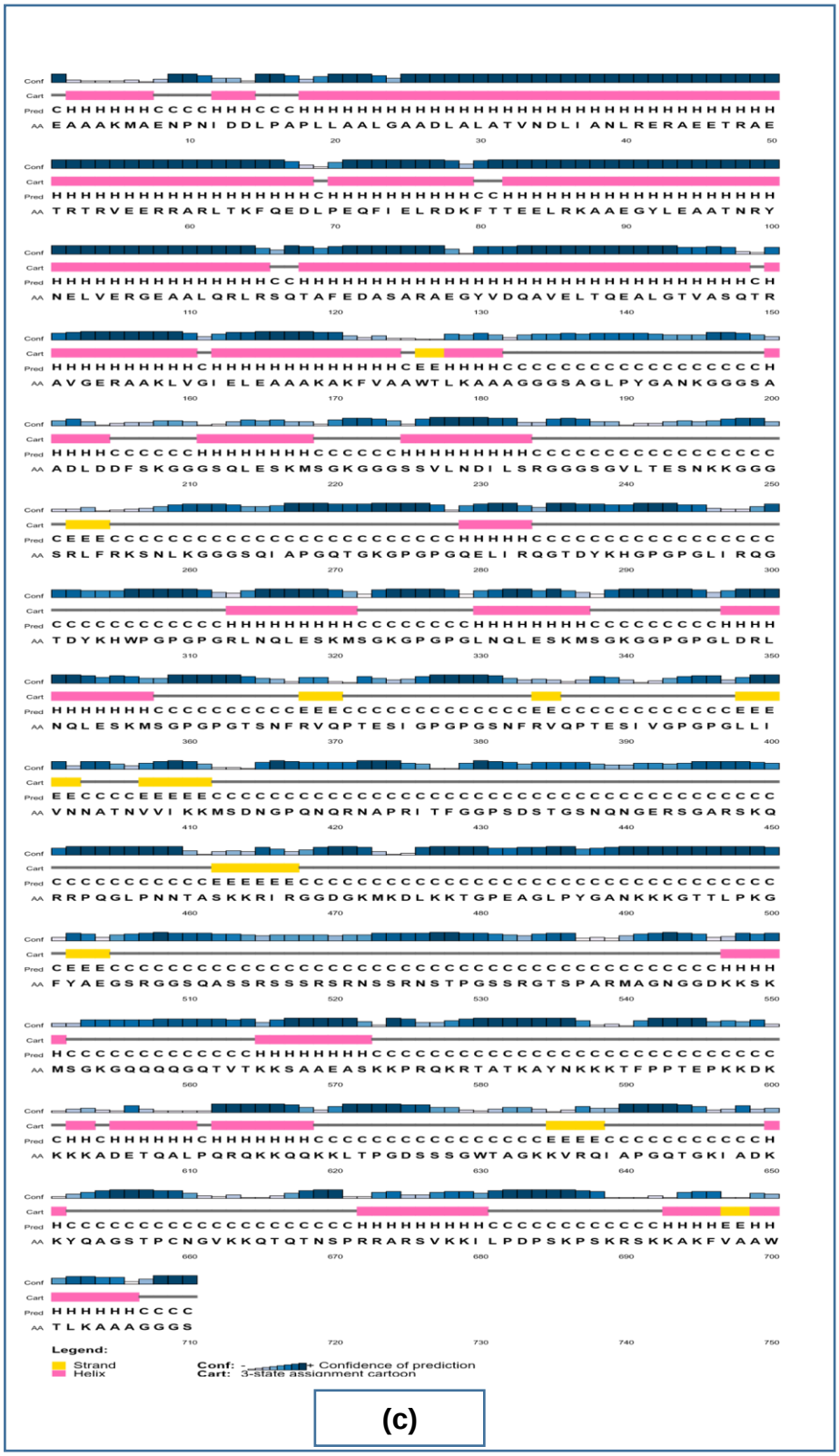

Figure 04. Results of the secondary structure prediction of the three vaccine constructs. Here, (a) is the CV-1 vaccine, (b) is the CV-2 vaccine, (c) is the CV-3 vaccine.

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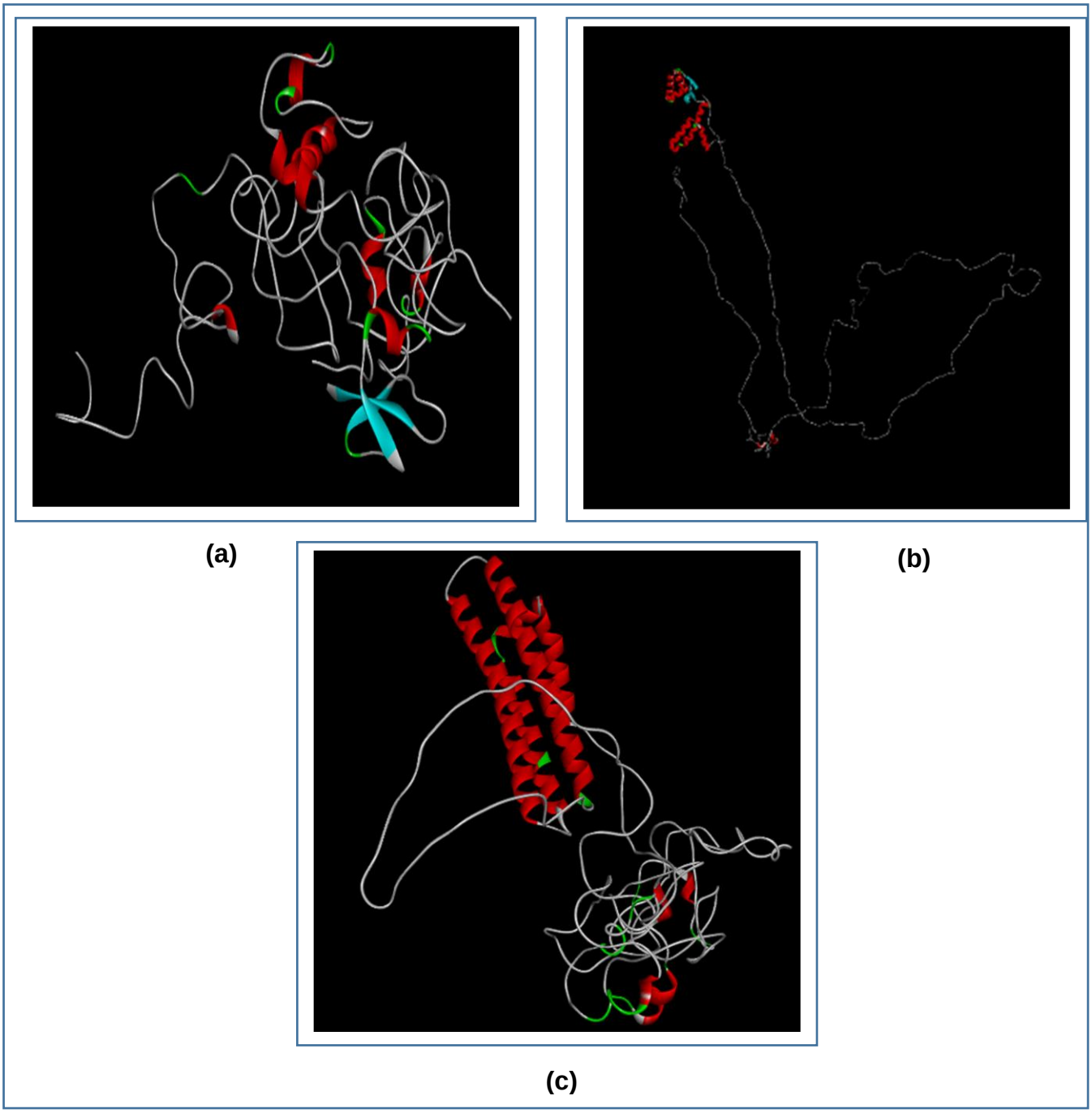

Figure 05. 3D structures of the three predicted vaccine constructs. Here, (a) is CV-1, (b) is CV-2, (c) is CV-3.

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3.10. 3D Structure Refinement and Validation

The 3D protein structures generated in the previous step were refined for further analysis and validation. The refined structures were refined with the aid of the Ramachandran Plots. The analysis showed that CV-1 vaccine had excellent percentage of 94.3% of the amino acids in the most favored region, 4.4% of the amino acids in the additional allowed regions, 0.0% of the amino acids in the generously allowed regions and 1.3% of the amino acids in the disallowed regions. The CV-2 vaccine had 90.0% of the amino acids in the most favored regions, 8.3% of the amino acids in the additional allowed regions, 0.6% of the amino acids in the generously allowed regions and 1.1% of the amino acids in the disallowed regions. The CV-3 vaccine showed the worst result with 77.4% of the amino acids in the most favored regions, 20.9% of the amino acids in the additional allowed regions, 1.4% of the amino acids in the generously allowed regions and 0.3% of the amino acids in the disallowed regions (**Figure 06).**

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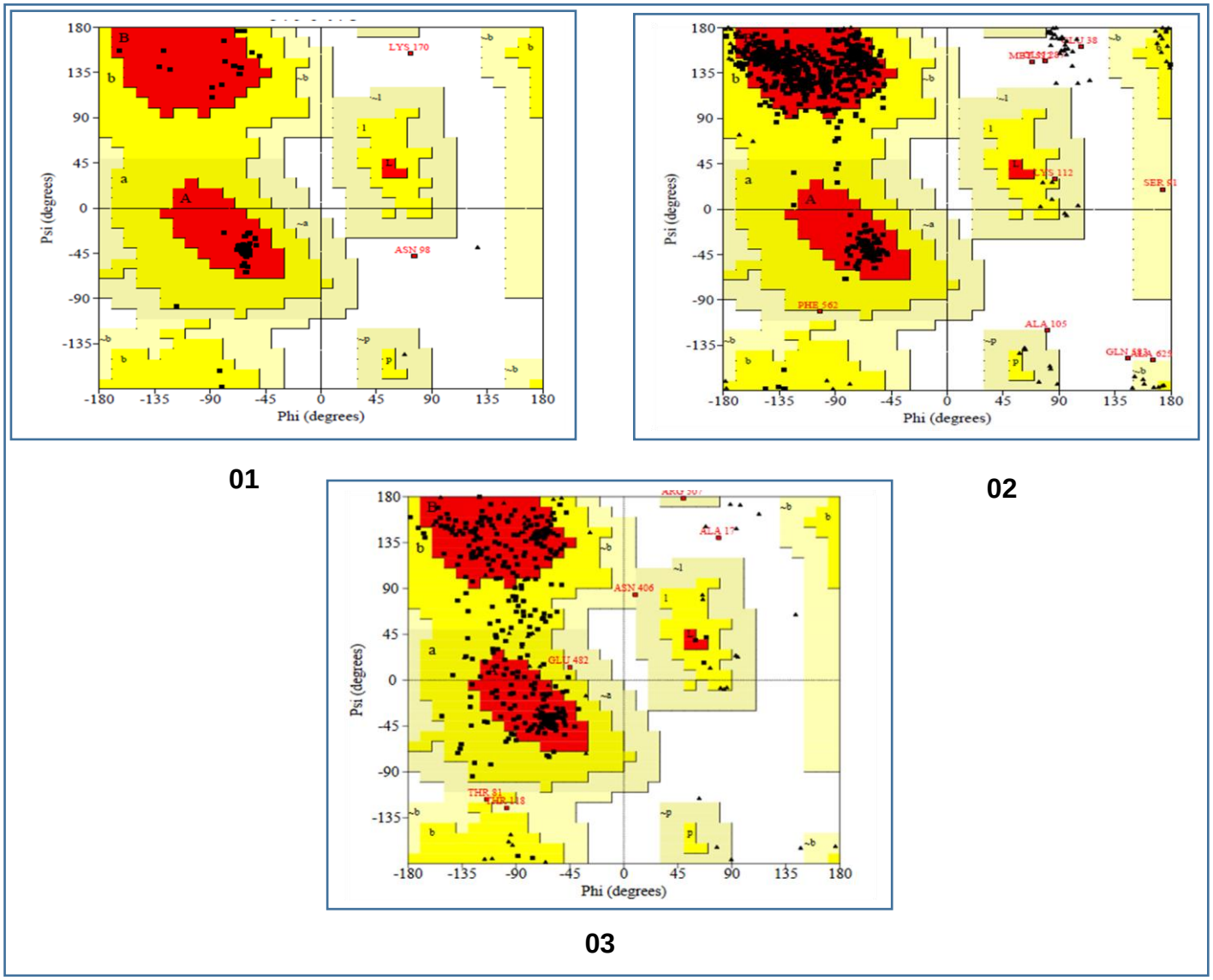

Figure 06. The results of the Ramachandran plot analysis of the three coronavirus vaccine constructs. Here, 01. CV-1 vaccine, 02. CV-2 vaccine, 03. CV-3 vaccine.

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3.11. Vaccine Protein Disulfide Engineering

In protein disulfide engineering, disulfide bonds were generated within the 3D structures of the vaccine constructs. In the experiment, the amino acid pairs that had bond energy value less than 2.00 kcal/mol were selected. The CV-1 generated 10 amino acid pairs that had the capability to form disulfide bonds. However, only one pair was selected because they had the bond energy, less than 2.00 kcal/mol: 276 Ser-311 Arg. However, CV-2 and CV-3 generated 04 and 05 pairs of amino acids, respectively, that might form disulfide bonds and no pair of amino acids showed bond energy less than 2.00 Kcal/mol. The selected amino acid pairs of CV-1 formed the mutant version of the original vaccines (**Figure 07**).

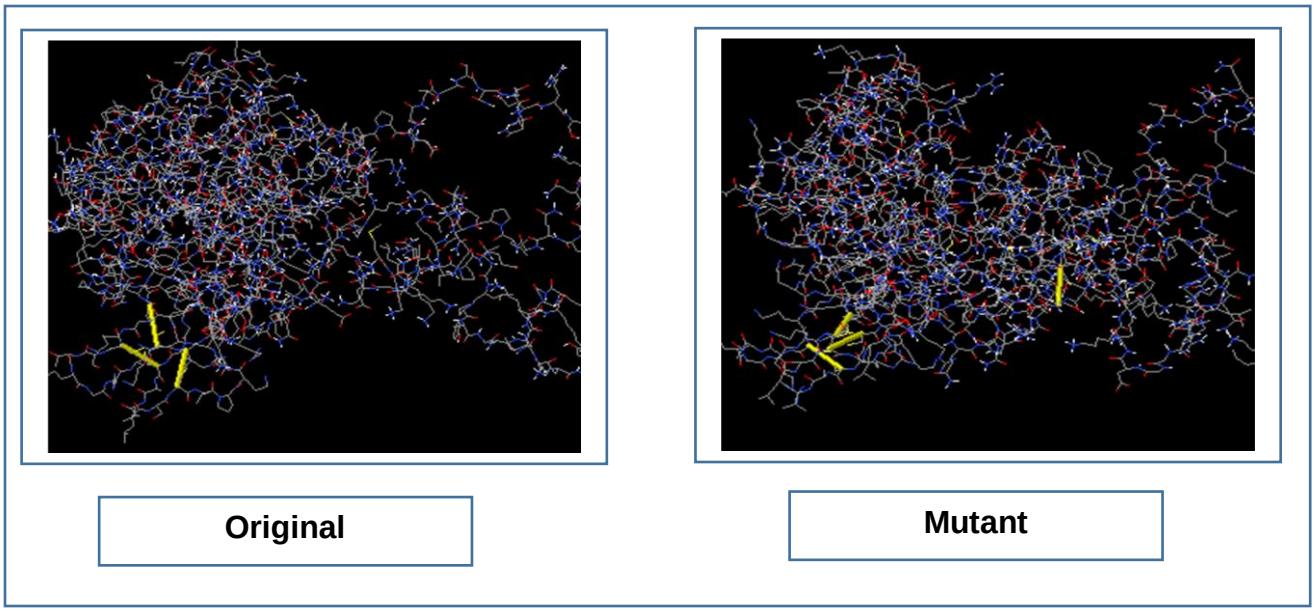

Figure 07. The disulfide engineering of CV-1. The original form is illustrated in the left side and the mutant form is illustrated in the right side.

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3.12. Protein-Protein Docking Study

The protein-protein docking study was carried out to find out the best constructed Wuhan coronavirus vaccine. According to docking result it was found that CV-1 was the best constructed vaccine. CV-1 showed the best and lowest scores in the docking as well as in the MM-GBSA study. However, CV-2 showed the best binding affinity (ΔG scores) with DRB3*0202 (-18.9 kcal/mol) and DRB1*0301 (-18.5 kcal/mol) when analyzed with ClusPro 2.0 and the PRODIGY tool of HADDOCK server. Moreover, when analyzed with PatchDock and FireDock servers, CV-3 showed best global energy scores with most of the MHC alleles i.e., DRB5*0101 (-10.70), DRB5*0101 (-19.59), DRB1*0101 (-17.46) and DRB3*0101 (-12.32). Since CV-1 showed the best results in the protein-protein docking study, it was considered as the best vaccine construct among the three constructed vaccines (**Figure 08** & **Table 13**). Later, the molecular dynamics simulation study and in silico codon adaptation studies were conducted only on the CV-1 vaccine.

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Table 13. Results of the docking study of all the vaccine constructs.

Name of the vaccines	Name of the Targets	PDB IDs of the targets	Binding affinity, ΔG (kcal mol^{-1})	Global energy	HawkDock score (the lowest score)	MM-GBSA (binding free energy, in kcal mol^{-1})
CV-1	DRB3*0202	1A6A	-17.2	-4.22	-6436.60	-55.56
	DRB5*0101	1H15	-19.9	-4.92	-6669.84	-141.66
	DRB1*0101	2FSE	-19.1	4.31	-7297.17	-148.58
	DRB3*0101	2Q6W	-19.2	-7.20	-7581.70	-138.4
	DRB1*0401	2SEB	-21.4	-11.58	-6758.33	-98.56
	DRB1*0301	3C5J	-17.7	-9.09	-5201.43	-114.35
	TLR8	3W3M	-23.2	-23.12	-6514.36	-52.06
CV-2	DRB3*0202	1A6A	-18.9	-10.32	-3477.55	1.01
	DRB5*0101	1H15	-17.7	-10.46	-3761.37	-106.17
	DRB1*0101	2FSE	-16.9	1.58	-3531.12	-106.13
	DRB3*0101	2Q6W	-19.1	16.68	-3707.86	-90.89
	DRB1*0401	2SEB	-20.0	-10.01	-4766.17	-45.76
	DRB1*0301	3C5J	-18.5	-1.87	-3561.16	-18.72
	TLR8	3W3M	-21.1	-18.91	-2945.44	-54.79
CV-3	DRB3*0202	1A6A	-16.9	-10.70	-4023.68	-9.2
	DRB5*0101	1H15	-18.9	-19.59	-4556.87	-12.38
	DRB1*0101	2FSE	-17.1	-17.46	-4602.08	-10.54
	DRB3*0101	2Q6W	-18.4	-12.32	-4767.21	-27.71
	DRB1*0401	2SEB	-20.1	6.55	-3571.79	8.74
	DRB1*0301	3C5J	-17.8	5.35	-4001.56	-12.38
	TLR8	3W3M	-22.8	-10.92	-5008.23	-19.83

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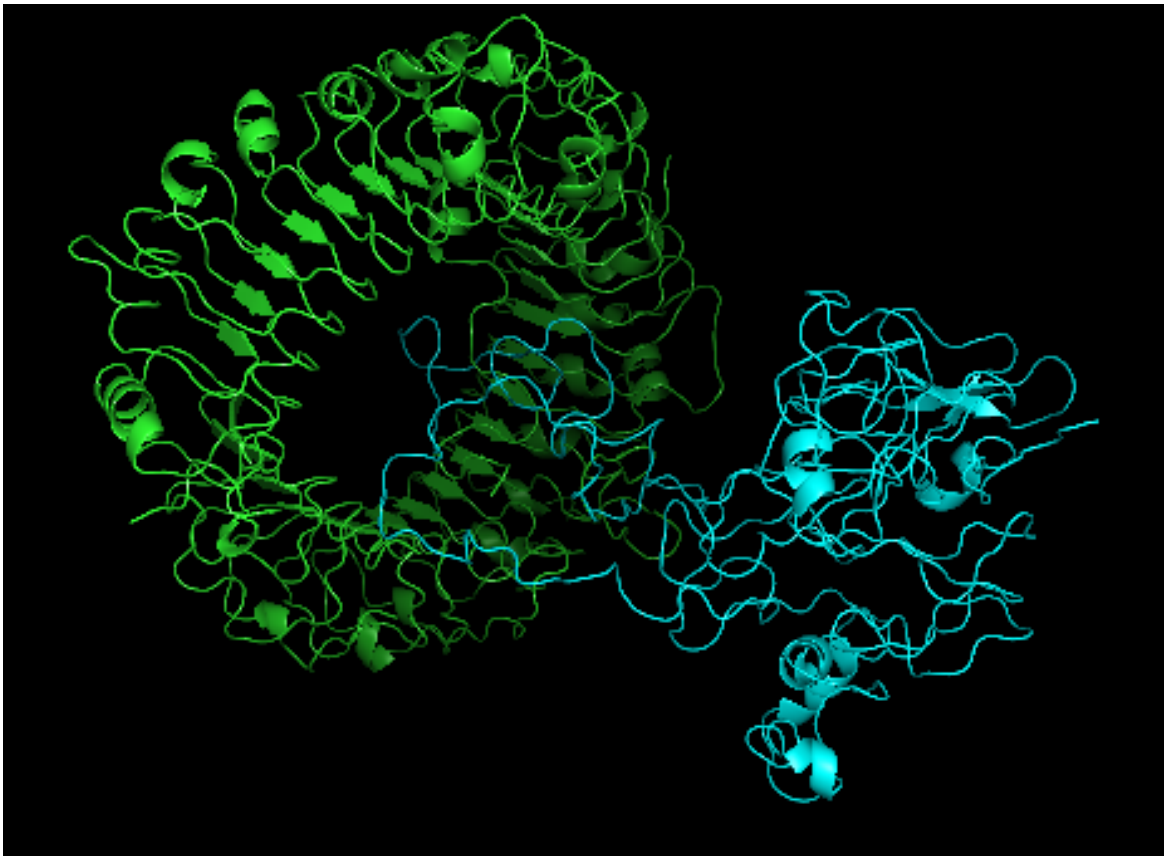

Figure 08. The interaction between TLR-8 (in green color) and CV-1 vaccine construct (in light blue color). The interaction was visualized with PyMol.

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3.13. Molecular Dynamics Simulation

The results of molecular dynamics simulation of CV-1-TLR-8 docked complex is illustrated in **Figure 09**. The deformability graph of the complex illustrates the peaks representing the regions of the protein with high degree of deformability (**Figure 09b**). The B-factor graph of the complex gives easy visualization and comparison between the NMA and the PDB field of the docked complex (**Figure 09c)**. The eigenvalue of the docked complex is depicted in **Figure 09d**. CV-1 and TLR8 docked complex generated eigenvalue of 3.817339e-05. The variance graph illustrates the individual variance by red colored bars and cumulative variance by green colored bars (**Figure 09e)**. **Figure 09f** depicts the co-variance map of the complex, where the correlated motion between a pair of residues are indicated by red color, uncorrelated motion is marked by white color and anti-correlated motion is indicated by blue color. The elastic map of the complex refers to the connection between the atoms and darker gray regions indicate stiffer regions (**Figure 09g**) [145]-[147].

3.14. Codon Adaptation and In Silico Cloning

Since the CV-1 protein had 596 amino acids, after reverse translation, the number nucleotides of the probable DNA sequence of CV-1 would be 1788. The codon adaptation index (CAI) value of 1.0 of CV-1 indicated that the DNA sequences contained higher proportion of the codons that should be used by the cellular machinery of the target organism *E. coli* strain K12 (codon biasness). For this reason, the production of the CV-1 vaccine should be carried out efficiently [151][152]. The GC content of the improved sequence was 51.34% (**Figure 10**). The predicted DNA sequence of CV-1 was inserted into the pET-19b vector plasmid between the SgrAI and SphI restriction sites. However, the DNA sequence did not have restriction sites for SgrAI and SphI restriction

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enzymes. For this reason, SgrA1 and SphI restriction sites were conjugated at the N-terminal and C-terminal sites, respectively. The newly constructed vector is illustrated in **Figure 11**.

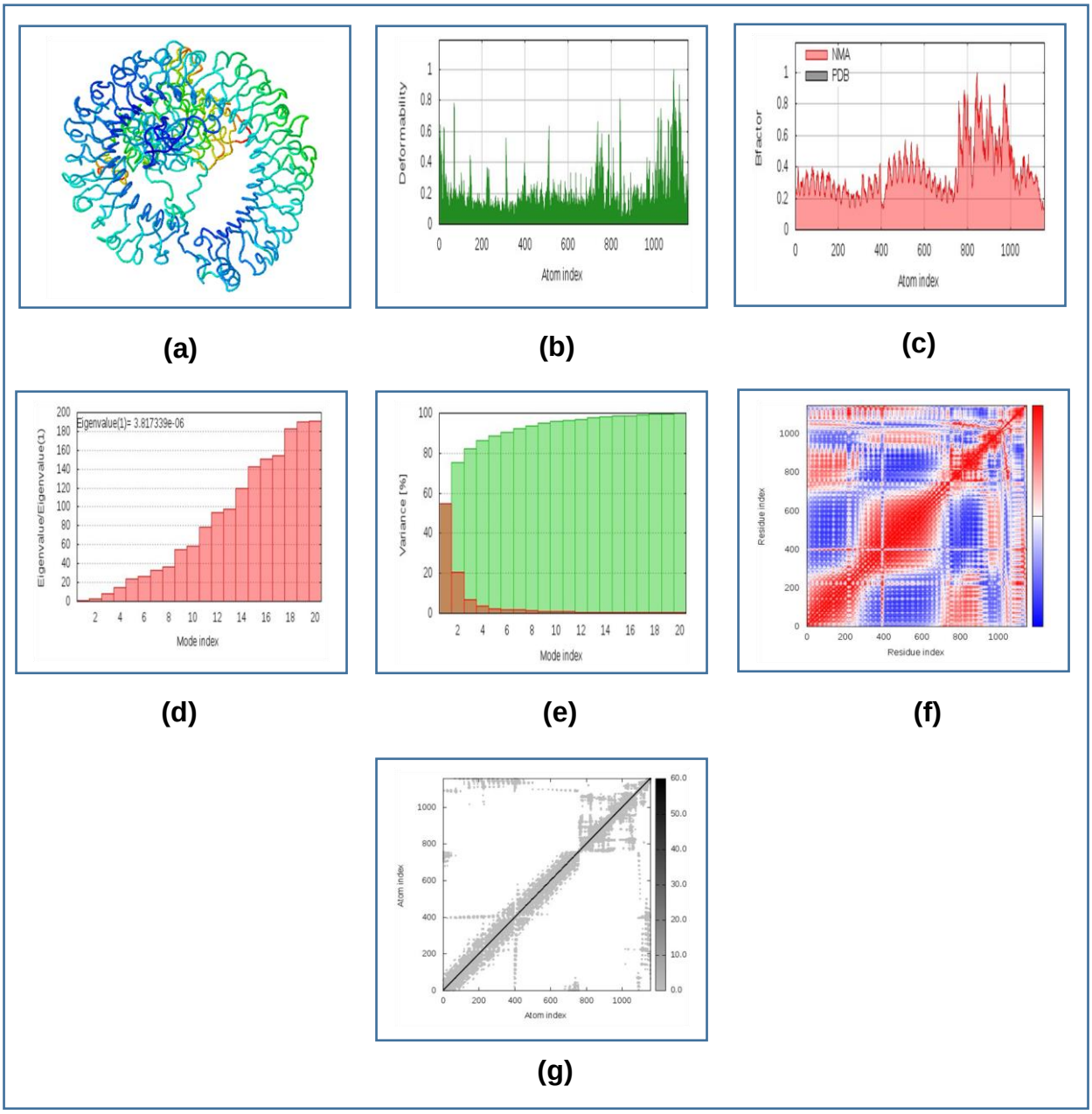

Figure 09. The results of molecular dynamics simulation study of CV-1 and TLR-8 docked complex. Here, (a) NMA mobility, (b) deformability, (c) B-factor, (d) eigenvalues, (e) variance (red color indicates individual variances and green color indicates cumulative variances), (f) co-

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variance map (correlated (red), uncorrelated (white) or anti-correlated (blue) motions) and (g) elastic network (darker gray regions indicate more stiffer regions).

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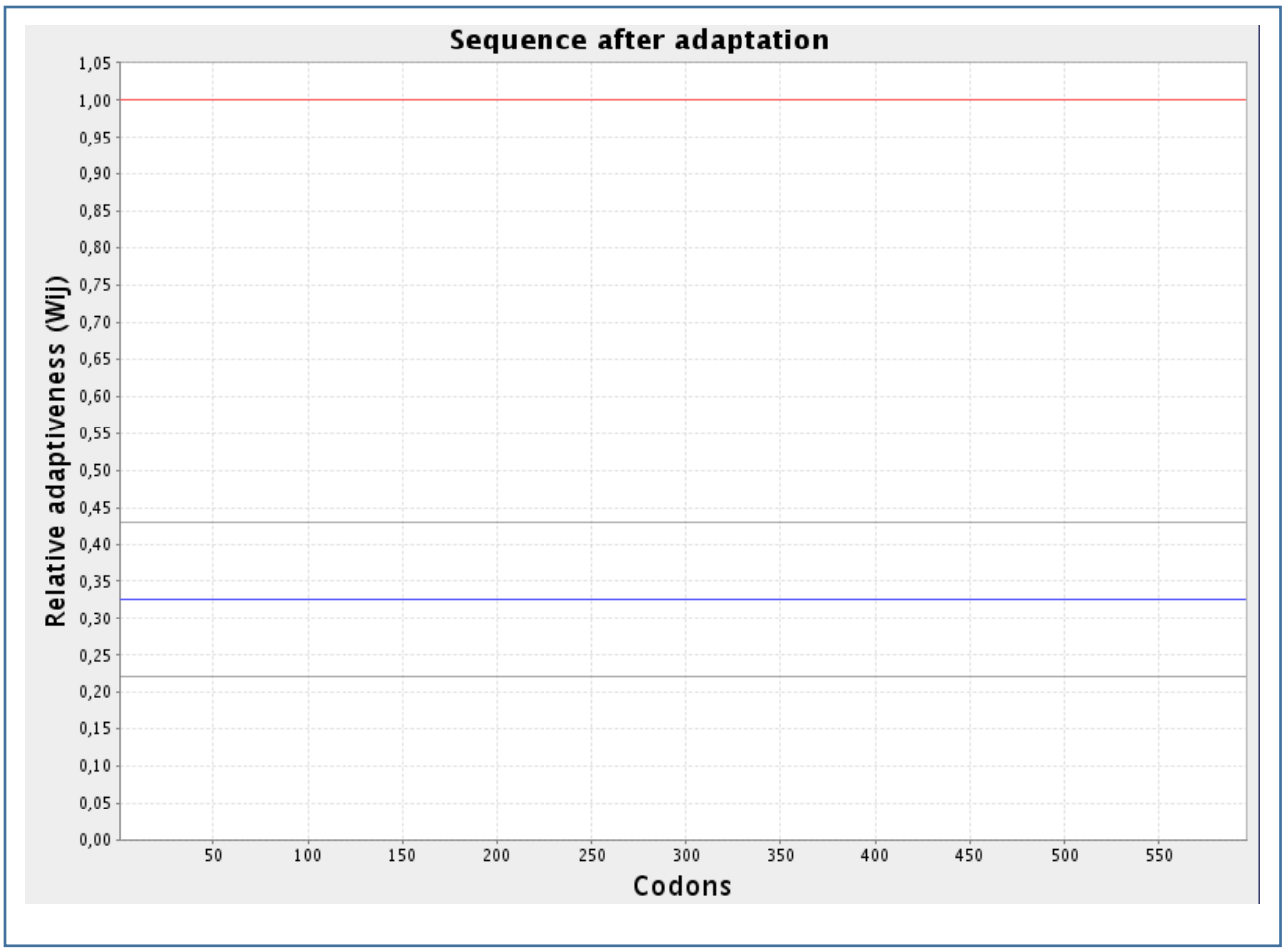

Figure 10. The results of the codon adaptation study of the constructed vaccine, CV-1.

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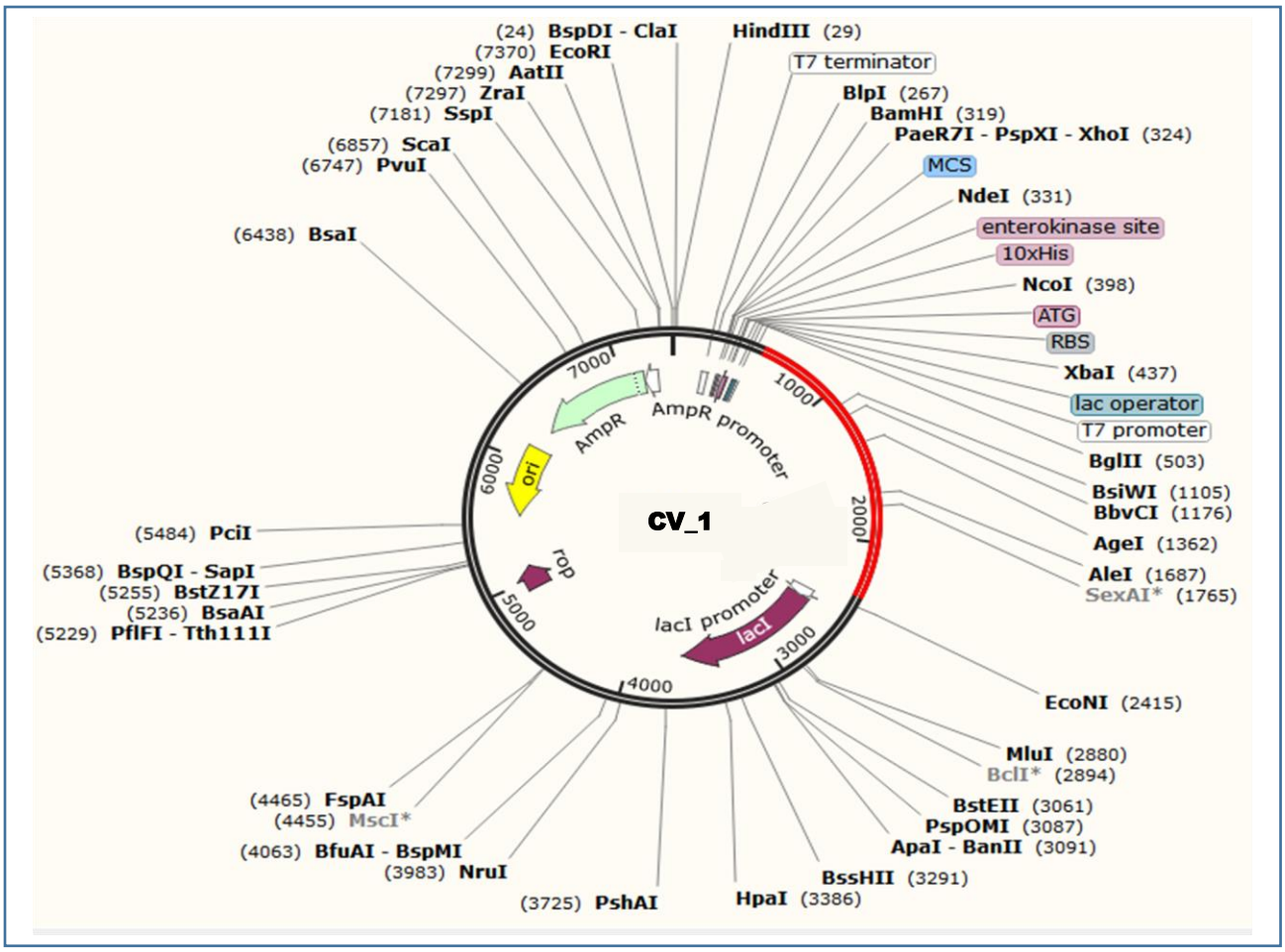

Figure 11. Constructed pET-19b vector with the CV-1 insert (marked in red color).

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4. Discussion

The current study was designed to construct possible vaccines against the Wuhan Novel Coronavirus 2019, which is the cause of the recent outbreak of the deadly viral pneumonia in China. To carry out the vaccine construction, four candidate proteins of the virus were identified and selected from the NCBI database. Only highly antigenic sequences were selected for further analysis since the highly antigenic proteins can induce better immunogenic response [153]. Because the nucleocapsid phosphoprotein and surface glycoprotein were found to be antigenic, they were taken into consideration for vaccine construction.

The physicochemical property analysis was conducted for the two antigenic proteins. The extinction coefficient can be defined as the amount of light that is absorbed by a particular compound at a certain wavelength [154][155]. Surface glycoprotein had the highest extinction co-efficient of 148960 M^{-1} cm^{-1}. The aliphatic index of a protein corresponds to the relative volume occupied by the aliphatic amino acids in the side chains of the protein, for example: alanine, valine etc. [156][157]. Surface glycoprotein had the highest aliphatic index among the two proteins (84.67). For this reason, surface glycoprotein had greater amount of aliphatic amino acids in its side chain than the nucleocapsid phosphoprotein. The grand average of hydropathicity value (GRAVY) for a protein is calculated as the sum of hydropathy values of all the amino acids of the protein, divided by the number of residues in its sequence [158]. Surface glycoprotein also had the highest GRAVY value of -0.079 among the two proteins. However, both of them had the predicted in vivo half-life of 30 hours and nucleocapsid phosphoprotein had the highest theoretical pI of 10.07. Both the proteins showed quite good results in the physicochemical property analysis.

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After the physicochemical analysis of the protein sequences, the T-cell and B-cell epitope prediction was conducted. T-cell and B-cell are the two main types of cells that function in immunity. When an antigen is encountered in the body by the immune system, the antigen presenting cells or APC like macrophage, dendritic cell etc. present the antigen to the T-helper cell, through the MHC class-II molecules on their surface. The helper T-cell contains CD4+ molecule on its surface, for this reason, it is also known as CD4+ T-cell. On the other hand, other type of T-cell, cytotoxic T-cell contains CD8+ molecule on their surface, for which, they are called CD8+ T-cell. MHC class-I molecules present antigens to cytotoxic T-lymphocytes. After activation by the antigen, the T-helper cell activates the B-cell, which starts to produce large amount of antibodies. Macrophage and CD8+ cytotoxic T cell are also activated by the T-helper cell that cause the final destruction of the target antigen [159]-[163]. The possible T-cell and B-cell epitopes of the selected proteins were determined by the IEDB (https://www.iedb.org/) server. Ten MHC class-I and MHC class-II epitopes of the both proteins from the top twenty best predicted epitopes by the server were taken into consideration. The epitopes with high antigenicity, non-allergenicity and non-toxicity were selected to vaccine construction. The B-cell epitopes (predicted by the server) that had more than ten amino acids were taken into consideration and the antigenic and non-allergenic epitopes were selected for vaccine construction. However, most of the epitopes were found to be residing within the cell membrane.

The cluster analysis of the MHC alleles which may interact with the selected epitopes during the immune response, showed quite good interaction with each other. Next the 3D structures of the selected epitopes were generated for peptide-protein docking study. The docking was carried out to find out whether all the epitopes had the capability to bind with their respective MHC class-I and MHC class-II alleles. Since all the epitopes generated quite good docking scores, it can be

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concluded that, all of them had the capability to bind with their respective targets and induce potential immune response. However, among the selected epitopes, QLESKMSGK, LIRQGTDYKHWP, GVLTESNKK and TSNFRVQPTESI generated the best docking scores. After the successful docking study, the vaccine construction was performed. The linkers were used to connect the T-cell and B-cell epitopes among themselves and also with the adjuvant sequences as well as the PADRE sequence. The vaccines, with three different adjuvants, were constructed and designated as: CV-1, CV-2 and CV-3. Since all the three vaccines were found to be antigenic, they should be able to induce good immune response. Moreover, all of them were possibly non-allergenic, they should not be able to cause any allergenic reaction within the body as per in silico prediction. With the highest aliphatic index of 54.97, CV-3 had the highest number of aliphatic amino acids in its side chain. The highest theoretical pI of CV-1 indicated that it requires high pH to reach the isoelectric point. Quite similar results of extinction co-efficient were generated by the three vaccine constructs. The three vaccine construct showed quite good and similar results in the physicochemical property analysis.

The secondary structure of the vaccine constructs revealed that CV-1 had the shortest alpha-helix formation, with 25% of the amino acids in the alpha-helix formation, however, 67.1% of the amino acids in coil formation. For this reason, most of the amino acid acids of CV-1 vaccine was in coil structure, which was also the highest percentage of amino acids in coil structure among the three vaccines. On the other hand, CV-3 had the highest amount of amino acids in the alpha-helix formation (37.8%), among all of the vaccines. However, all the three vaccine constructs had most of their amino acids in their coil structures. In the tertiary structure prediction, all the three vaccine construction showed quite satisfactory results. In the tertiary structure refinement and validation,

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CV-1 vaccine construct generated the best result with 94.3% of the amino acids in the most favored region and 4.4% of the amino acids in the additional allowed regions. CV-2 also showed good result 90.0% of the amino acids in the most favoured region. In the disulfide bond engineering experiment, only CV-1 was found to follow the selection criteria for disulfide bond formation. With the lowest and best results generated by the MM-GBSA study, HawkDock study and ClusPro 2.0 server, CV-1 was considered as the best vaccine construct among the three vaccines. For this reason, CV-1 was selected for molecular dynamics simulation study, codon adaptation and in silico coding study. The molecular dynamics simulation study revealed that the TLR-8-CV-1 docked complex should be quite stable with a good eigenvalue. The complex had less chance of deformation and for this reason, the complex should be quite stable in the biological environment. The **Figure 09f** shows that a good number of amino acids were in the correlated motion that were marked by red color. Finally, codon adaptation and in silico cloning experiments were performed and with 1.0 CAI value, it could be concluded that the DNA sequence should have very high amount of favorable codons that should be able to express the desired amino acids in the target microorganism, *E. coli* strain K12. The DNA sequence also had quite high and good amount of GC content of 51.34%. Finally, the pET-19b vector, containing the CV-1 vaccine insert was constructed which should efficiently and effectively encode the vaccine protein in the *E. coli* cells.

5. Conclusion

The Wuhan Novel Coronavirus 2019 has caused one of the deadliest outbreaks in the recent times. Prevention of the newly emerging Wuhan Novel Coronavirus infection is very challenging as well as mandatory. The potentiality of in silico methods can be exploited to find desired solutions with fewer trials and errors and thus saving both time and costs of the scientists. In this study, potential subunit vaccines were designed against the Wuhan Novel Coronavirus 2019 using various methods

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of reverse vaccinology and immunoinformatics. To design the vaccines, the highly antigenic viral proteins as well as epitopes were used. Various computational studies of the suggested vaccine constructs revealed that these vaccines might confer good immunogenic response. For this reason, if satisfactory results are achieved in various in vivo and in vitro tests and trials, these suggested vaccine constructs might be used effectively for vaccination to prevent the coronavirus infection and spreading. Therefore, our present study should help the scientists to develop potential vaccines and thapeutics against the Wuhan Novel Coronavirus 2019.

Conflict of Interest

Authors declare no conflict of interest regarding the publication of the manuscript.

Data Availability Statement

Authors made all the data generated during experiment and analysis available within the manuscript.

Funding Statement

Authors received no specific funding from any external sources.

Acknowledgements

Authors acknowledge the members of Swift Integrity Computational Lab, Dhaka, Bangladesh, a virtual platform of young researchers for their support during the preparation of the manuscript.

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