- 1 **Title:** Climate explains geographic and temporal variation in mosquito-borne disease
- 2 dynamics on two continents

- 4 Jamie M. Caldwell¹, A. Desiree LaBeaud², Eric F. Lambin^{3,4}, Anna M. Stewart-Ibarra^{5,6},
- 5 Bryson A. Ndenga⁷, Francis M. Mutuku⁸, Amy R. Krystosik², Efraín Beltrán Ayala⁹,
- 6 Assaf Anyamba¹⁰, Mercy J. Borbor-Cordova¹¹, Richard Damoah¹², Elysse N. Grossi-
- 7 Soyster², Froilán Heras Heras¹³, Harun N. Ngugi^{14,15}, Sadie J. Ryan¹⁶⁻¹⁸, Melisa M.
- 8 Shah¹⁹, Rachel Sippy^{13,20,21}, Erin A. Mordecai¹
- 10 Department of Biology, Stanford University, 371 Serra Mall, Stanford, California, USA
- ² Department of Pediatrics, Division of Infectious Diseases, Stanford University, 300
- 12 Pasteur Drive, Stanford, California, USA
- 13 School of Earth, Energy & Environmental Sciences, and Woods Institute for the
- Environment, Stanford University, Stanford, California 94305, USA.
- ⁴ Georges Lemaître Earth and Climate Research Centre, Earth and Life Institute,
- 16 Université catholique de Louvain, 1348 Louvain-la-Neuve, Belgium.
- ⁵ Department of Medicine and Department of Public Health and Preventative Medicine,
- SUNY Upstate Medical University, Syracuse, NY, USA
- 19 ⁶ InterAmerican Institute for Global Change Research (IAI), Montevideo, Uruguay
- ⁷ Centre for Global Health Research, Kenya Medical Research Institute, Kisumu, Kenya
- 21 ⁸ Department of environment and health sciences, technical university of Mombasa,
- Mombasa, Kenya
- ⁹ Technical University of Machala, Machala, Ecuador

¹⁰ Universities Space Research Association and NASA Goddard Space Flight Center, 24 25 Greenbelt, MD, USA. ¹¹ Facultad de Ingeniería Marítima y Ciencias del Mar, Escuela Superior Politécnica del 26 27 Litoral, ESPOL, Guayaquil, Ecuador ¹² Morgan State University and NASA Goddard Space Flight Center, Greenbelt, MD, 28 29 USA. ¹³ Center for Research SUNY-Upstate-Teófilo Dávila Hospital, Machala, Ecuador 30 ¹⁴ Department of Biological Sciences, Chuka University, Chuka, Kenya 31 ¹⁵ Department of Zoology, School of Biological Sciences University of Nairobi, Nairobi, 32 33 Kenya ¹⁶ Emerging Pathogens Institute, University of Florida, Gainesville, Florida 34 ¹⁷ Quantitative Disease Ecology and Conservation (QDEC) Lab, Department of 35 36 Geography, University of Florida, Gainesville, Florida; ¹⁸ School of Life Sciences, University of KwaZulu, Natal, South Africa 37 ¹⁹ Department of Medicine, Division of Infectious Diseases, Stanford University, 300 38 39 Pasteur Drive, Stanford, California, USA ²⁰ Institute for Global Health and Translational Science, SUNY-Upstate Medical 40 41 University, Syracuse, NY, USA ²¹ Department of Medical Geography, University of Florida, Gainesville, FL, USA 42 43 44 45 46

Abstract:

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

Climate drives population dynamics, but when the underlying mechanisms are unresolved, studies can lead to seemingly context-dependent effects of climate on natural populations. For climate-sensitive vector-borne diseases such as dengue, chikungunya, and Zika, climate appears to have opposing effects in different contexts. In this study, our objective was to test the extent to which a mathematical model, parameterized with climate-driven mosquito physiology measured in laboratory studies, predicts observed vector and disease dynamics in the field across ecologically and culturally distinct settings in Ecuador and Kenya. The model incorporates different rainfall functions and time lags. We show that the climate-driven model captures three key epidemic characteristics across settings: the number, timing, and duration of outbreaks. In addition, the model generates a range of disease dynamics consistent with observations of Aedes aegypti abundances and laboratory-confirmed arboviral incidence with varying levels of accuracy (28 - 85%) for vector dynamics, 44 - 88% for human disease dynamics). Further, we find that the model predicted vector dynamics better in sites with a smaller proportion of young children in the population, lower mean temperature, and a larger proportion of homes with piped water and made of cement. A mechanistic model with limited calibration to local data that robustly captures the influence of climate on viruses transmitted by Aedes aegypti provides critical information to help guide future intervention efforts and improve disease projections associated with climate change.

Introduction:

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

Climate is a major driver of species interactions and population dynamics, but the mechanisms underlying the ecological effects of climate are often poorly understood and rarely tested in the field [1]. One of the primary ways that climate impacts populations is through its effects on species' vital rates [2]. However, the effects of climate on population dynamics may appear context dependent in the field because multiple climate variables can act synergistically, with each climate variable potentially affecting multiple vital rates, and their impacts may be nonlinear, changing direction and relative importance across a gradient of conditions [3,4]. Therefore, paradoxically, while climate is thought to be one of the most pervasive drivers of ecological processes, its directional and dynamical effects on systems are often poorly understood and difficult to predict. Vector-borne diseases provide an interesting case study to test whether climate sensitive traits measured in controlled, laboratory settings can reproduce the wide range of dynamics observed in the field. For example, transmission of mosquito-borne viral (arboviral) diseases such as dengue, chikungunya, and Zika occur along a spectrum from low levels of year-round endemic transmission [5] to large seasonal or interannual outbreaks [6]. We hypothesize that important features of these differing dynamics arise due to regional or seasonal differences in climate, where the magnitude and direction of the effects of climate on vector and disease dynamics differ [7–12]. Understanding the mechanisms that drive disease dynamics can help address two critically important research priorities for arboviruses like dengue, chikungunya, and Zika: assessing intervention strategies and projecting climate change impacts on disease

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

dynamics. While phenomenological models often replicate arboviral disease dynamics remarkably well [13], mechanistic models that do not rely on local data for calibration and capture mosquito population dynamics and interactions between mosquitoes and humans will provide more realistic projections for epidemic dynamics across a broad range of transmission settings. With no widely available vaccine, vector control (e.g., larvicides, Wolbachia-infected mosquito releases) remains the primary method for preventing arboviral disease transmission, and, like other vector-borne diseases with complex transmission dynamics, model simulations can help guide effective intervention efforts [14,15]. Further, mechanistic models are better suited to predict how climate change will impact future disease burden and distribution, as projected climate conditions are outside the current arboviral climate niche space [16]. Despite the potential usefulness of mechanistic approaches, validation with vector and disease data are limited, raising an important question about which epidemic characteristics, if any, we should expect a model to capture when the model was parameterized with data that is on different scales (e.g., individuals versus populations) and independent from the transmission system we wish to predict. Thus, because we cannot study epidemic dynamics in every possible transmission setting, it becomes important to understand the extent to which models derived from fundamental and laboratory-measured traits explain disease dynamics across diverse settings. We hypothesize that a climate-driven mechanistic model with limited calibration should capture many important characteristics of disease dynamics for dengue, chikungunya, and Zika because of the ecology of Aedes aegypti, the primary disease vector. Ae. aegypti are

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

anthropophilic, globally distributed mosquitoes that breed in artificial containers with standing water [17,18]. All mosquito and parasite traits that are important for transmission and linked to metabolism, such as reproduction, development, survival, biting rate, and extrinsic incubation period, are temperature dependent with an intermediate thermal optimum [19–21]. Humidity is positively associated with mosquito survival because the high surface area to volume ratio of mosquitoes exposes them to desiccation [22,23]. Standing water from rainfall provides essential larval and pupal habitat for mosquitoes, but the relationship is complex because heavy rainfall can flush away breeding habitats [24–26] and water storage practices during drought can increase water availability, mosquito abundance, and contact between mosquitoes and people [27– 29]. A previous simulation study predicted that in settings with suitable climate for transmission throughout the year (e.g., mean temperature = 25° C; range = $20 - 30^{\circ}$ C), temperature drives the timing and duration of outbreaks, but not the maximum number of infections or final epidemic size [30]. This finding suggests that a model that incorporates temperature-dependent vector traits should capture some important epidemic characteristics. In this study, our goal was to test the extent to which climate-driven mosquito traits drive disease dynamics across two geographically distinct regions and to characterize additional climatological, ecological, and social factors that may mediate the effects of climate on disease dynamics. We built on previous mechanistic and semi-mechanistic models that incorporate the Aedes mosquito life cycle and human disease dynamics [30-35] by combining a suite of temperature, humidity, and rainfall dependent trait functions

140

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

into one epidemiological model. We validated the model with Ae. aegypti abundances and laboratory-confirmed dengue, chikungunya, and Zika cases from two equatorial countries with distinct socioeconomic, geographic, cultural, and disease transmission settings: Ecuador and Kenya (Fig. 1, Table 1). The study sites within each country were distributed across a gradient of temperature, humidity, and rainfall. Previous studies have found that Ae. aegypti and dengue were positively associated with warm and wet conditions in Ecuador and Kenya [6,36–38], although other Ae. aegpyti-vectored arboviruses in Kenya such as chikungunya have been associated with warm and dry conditions [39]. Both countries have all four dengue serotypes circulating and have recently experienced outbreaks of chikungunya; yet, arboviral transmission dynamics differ in each country. In Ecuador, dengue is a re-emerging disease with large seasonal epidemics that frequently result in severe dengue [6]; by contrast, in Kenya, dengue is transmitted at low levels year-round [5] and intermittent self-limiting outbreaks often go undetected [40]. Further, compared with South America, severe dengue is rare in sub-Saharan Africa, perhaps because African strains of Ae. aegpyti have lower susceptibility to all four dengue serotypes [41], and/or because people of African ancestry are less susceptible to severe dengue [42].

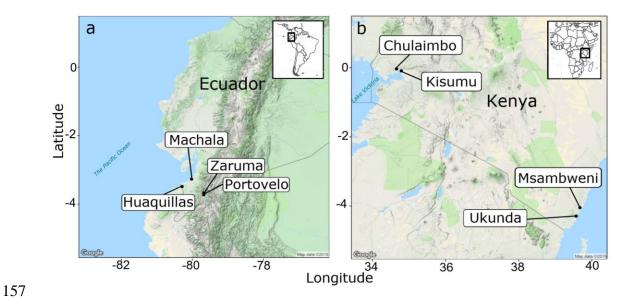


Figure 1: Study sites within two equatorial countries: (a) Ecuador in South America and (b) Kenya in East Africa.

Table 1: Study sites differ geographically, climatologically, and socioeconomically.

¹Mean annual normalized difference vegetation index (NDVI) is a proxy for photosynthesis and measured as a difference in spectral reflectance in the visible and near-infrared regions from NASA/NOAA MODIS (MOD13A1) [43]. ²Dominant land cover type is measured and classified from spectral and temporal features from NASA/NOAA MODIS (MCD12Q1) [44]. Land cover types include (9) Tree cover 10 - 30%, (10) Dominated by herbaceous annuals, (13) >30% impervious surface area, and (14) 40 - 60% mosaics of small-scale cultivation. Bed net use represents availability of and/or willingness to adopt intervention strategies for preventing infection rather than a

direct adaptive response to preventing infection by day-biting Ae. aegpyti mosquitoes.

	I	I	1	1		I	1	1
	Huaquillas, Ecuador	Machala, Ecuador	Portovelo, Ecuador	Zaruma, Ecuador	Chulaimbo, Kenya	Kisumu, Kenya	Msmabweni, Kenya	Ukunda, Kenya
Site characteristics	Site characteristics							
Elevation (m)	15	6	645	1,155	1,328	1,100	4	8
Location	Coastal	Coastal	Inland	Inland	Inland	Inland	Coastal	Coastal
Mean annual NDVI ¹	0.22	0.12	0.61	0.57	0.63	0.35	0.33	0.52
Dominant land cover type ²	13	13	9	10	14	13	13	10
Climate			•	•	•		•	
Mean temperature (°C)	26	26	25	22	24	26	28	28
Mean relative humidity (%)	81	84	81	86	69	50	76	78
Mean annual rainfall (mm)	317	669	500	1115	1125	810	1048	922
Demographics								
Human population size	57,366	279,887	13,673	25,615	7,304	491,893	15,371	80,193
Population <5 years (%)	10	9	9	8	12	12	13	14
Population of African ancestry (%)	5.1	6.0	3.3	2.9	100.0	100.0	100.0	100.0
Housing quality (% houses)	•	•	•	•	•		•	
Piped water inside home	90	91	100	96	2	4	3	11
No screens on windows	7	60	91	99	74	78	43	21
House materials (cement/mud/wood)	87/5/0	87/8/5	95/0/5	93/1/1	29/70/0	77/17/0	38/62/0	51/47/0
Exposure, vulnerability, and	adaptive ca	pacity						
Arboviruses present	dengue, chikungunya, Zika				>200 documented including dengue,			
					chikungunya, Yellow fever, Rift Valley			
					fever, West Nile fever, O'nyong-nyong			
Insecticide use (% houses)	19	28	46	37	0	0	11	55
Bednet use (% houses)	77	55	15	21	93	92	0	96
Other vector control	Ultra-low volume fumigation with				Mosquito coils			
strategies used	malathion (organophosphate) and							
	community mobilization to eliminate							
	larval habitats			1.1				
Annual gross domestic	\$177 billion USD			\$85.98 billion USD				
product by country (2018)								

Results:

172

173

174

176

177

178

Capturing key epidemic characteristics

175 The dynamic susceptible, exposed, infectious – susceptible, exposed, infectious, removed

(SEI-SEIR) compartmental model parameterized with temperature-, humidity-, and

rainfall-dependent mosquito life history traits (Fig. 2) reproduced three key

characteristics of epidemics: number of outbreaks, timing of outbreak peak, and duration

of outbreaks. We defined an outbreak as a continuous time period with peak cases exceeding the mean number of cases (predicted or observed) plus one standard deviation within a site. Across all sites, the number of outbreaks predicted by the model closely matched the number of outbreaks observed ($R^2 = 0.79$, p < 0.01; Fig. 3a). Supporting our *a priori* expectations based on a previous simulation study [30], we found that the climate-driven model predicted peak timing of outbreaks ($R^2 = 0.71$, p < 0.01; Fig. 3b) and outbreak duration ($R^2 = 0.51$, p < 0.01; Fig. 3c) well but did not predict the final outbreak size (Fig. 3d) or maximum number of infections (Fig. 3e) across sites. Overall, the model predicted four outbreaks that were not observed and did not predict five outbreaks that occurred. The model may miss an outbreak (i.e., false negatives) when, for example, suitable climate occurs but the pathogen is not introduced or the susceptible population is depleted from previous outbreaks.

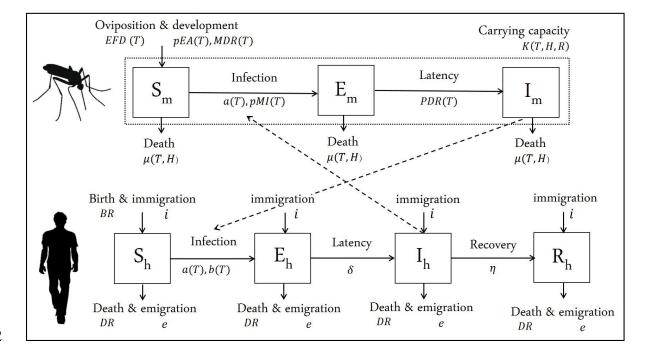


Figure 2: SEI-SEIR epidemiological model framework. The mosquito population is split among susceptible (S_m) , exposed (E_m) , and infectious (I_m) compartments (squares) and the human population is split among susceptible (S_h) , exposed (E_h) , infectious (I_h) , and recovered (R_h) compartments. Solid arrows indicate the direction individuals can move between classes and dashed arrows indicate the direction of transmission. Transitions among compartments are labeled by the appropriate processes and corresponding rate parameters (see Methods for parameter definitions and more detail). Rate parameters with a T, H, and R are temperature-, humidity-, and rainfall-dependent, respectively. The total adult mosquito population $(S_m, E_m,$ and I_m compartments; dotted rectangle) is maintained at an abundance less than or equal to the mosquito carrying capacity.

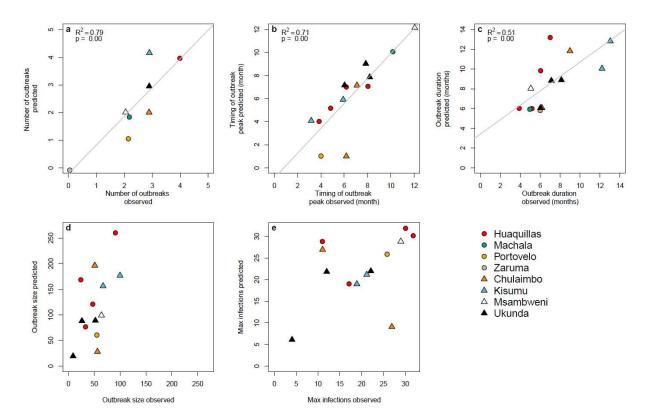


Figure 3: Model predictions for the number, timing, and duration of arboviral outbreaks closely matched field observations. Scatterplots show model predictions versus observations for different epidemic characteristics. (a) Number of outbreaks indicates the total number of predicted and observed outbreaks in a site over the study period. (b) Timing of outbreak peak, (c) outbreak duration, (d) outbreak size, and (e) maximum infections (e.g., max I_h during an outbreak) correspond to individual outbreaks where model predictions and observations overlapped in time, therefore, some plots show multiple data points per site. Outbreaks are colored by site with different symbols for Ecuador (circles) and Kenya (triangles). We show regression lines and associated statistics for statistically significant relationships. For visualization purposes, we jittered the data points to show overlapping data and we excluded data from Machala in plots (d)

outbreak size and (e) maximum infections because the magnitude differed substantially from all other sites.

Capturing spatio-temporal disease dynamics across sites

218

219

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

240

The SEI-SEIR model generated mosquito and disease dynamics that better reflected observed dynamics in some sites than others (Fig. 4, Table 2). Model-predicted mosquito abundances were significantly correlated with field-collected observations of mosquito abundances in all eight study sites, explaining 28 - 85% of site-level variation through time based on pairwise correlations with an adjusted p-value for time series data (following [45]). Based on surveys conducted across all vector life stages in Kenya (only adult mosquitoes were collected in the Ecuador surveys), the SEI-SEIR model explained variation in the abundance of adult mosquitoes (28-63%) better than pupae (25-32%), late instars (30-33%), early instars (20-36%), and eggs (33-55%), likely because the model did not explicitly incorporate other mosquito life history stages. Model-predicted disease cases were significantly correlated with laboratory-confirmed arboviral incidence in seven of the eight study sites, explaining 44 - 88% of site-level variation through time (within sites with statistically significant pairwise correlations). We confirmed that the predicted dynamics were stable with sensitivity analyses to initial conditions (see Methods), as emerging diseases can display chaotic dynamics due to a high sensitivity to initial conditions. Overall, the model reproduced disease dynamics slightly better for sites in Ecuador compared with Kenya.

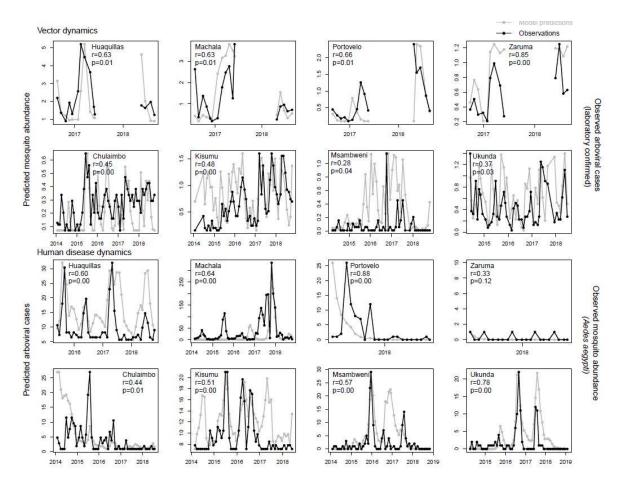


Figure 4: Model predicts vector and human disease dynamics better in some settings than others. Each plot shows the time series of SEI-SEIR model predictions (grey dots connected by grey lines) and field observations (black dots connected by black lines) for vector (top two rows) and human disease (bottom two rows) dynamics for each study site with the pairwise correlation (r) and adjusted p-value (p). We calculated observed mosquito abundances as the mean number of adult *Ae. aegypti* per house, month, year, and site. We calculated observed arboviral cases as the total number of laboratory-confirmed dengue (any serotype), chikungunya, and Zika cases per month, year, and site; six of the eight study sites only included dengue cases (see Methods). The first and third rows show sites in Ecuador and the second and fourth rows show sites in Kenya. We show uncertainty in model predictions in Figs. S1-2.

Table 2: Model predictions reflect a range of observed transmission dynamics when incorporating different rainfall functions and time lags across sites. For each study site, we calculated pairwise correlations between time series of field observations (*Ae. aegypti* abundances or arboviral cases) and time series of model predictions for the SEI-SEIR model with one of three rain functions for mosquito carrying capacity (Brière, Inverse, or Quadratic) and six time lags (0-5 months). This table shows specifications for the model (e.g., rain function and time lag) with the highest pairwise correlation value, r, for each study site and observation type (vectors or human disease cases), as well as the statistical significance of the correlation value (adjusted p-value) based on the Modified Chelton method [45] to account for temporal autocorrelation.

	Vector dynamics				Human disease dynamics			
Site	Rainfall function	r	Adjusted p-value	Lag (months)	Rainfall function	r	Adjusted p-value	Lag (months)
Huaquillas, Ecuador	Quadratic	0.63	0.01	1	Inverse	0.60	0.00	2
Machala, Ecuador	Quadratic	0.63	0.01	0	Brière	0.64	0.00	4
Portovelo, Ecuador	Brière	0.66	0.01	1	Brière	0.88	0.00	3
Zaruma, Ecuador	Inverse	0.85	0.00	1	Inverse	0.33	0.12	0
Chulaimbo, Kenya	Inverse	0.45	0.00	1	Quadratic	0.36	0.02	4
Kisumu, Kenya	Brière	0.48	0.00	0	Quadratic	0.51	0.00	4
Msambweni, Kenya	Inverse	0.28	0.04	0	Inverse	0.57	0.00	3
Ukunda, Kenya	Inverse	0.37	0.03	1	Inverse	0.78	0.00	5

We found evidence that rainfall affects transmission through multiple mechanisms and at different time lags (Table 2). Since the effect of rainfall on mosquito abundances is not well understood, we simulated disease dynamics for each site three times, using one of

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

three hypothesized rainfall relationships (Brière, inverse, and quadratic; Fig. S3). We determined the best rainfall function and time lag for each site based on the highest pairwise correlation value between model predictions and observations. The model with the exponentially decreasing inverse rain function (Fig. S3c), which indicates that mosquito abundances peak when there is no or low rainfall (likely as a result of water storage practices and/or unreliable water sources) described observed mosquito and disease dynamics most often, especially in the Kenya sites (Table 2), where household access to piped water is very low (Table 1). The left-skewed unimodal Brière rainfall function (Fig. S3a), which indicates that mosquito abundances increase with increasing rainfall until some threshold where flushing occurs, described disease dynamics in some settings, particularly in the Ecuador sites. The symmetric unimodal quadratic rainfall function (Fig. S3b), which indicates that mosquito abundances peak with intermediate amounts of rainfall and are reduced with low and high rainfall values, also described disease dynamics in some settings. Interestingly, we did not find a single rainfall function that consistently described dynamics for mosquitoes or arboviral cases across study sites, or for both mosquitoes and arboviral cases within individual study sites (Table 2). In contrast, we did find some consistency with time lags. The model best predicted mosquito abundances in the same month or one month in the future. In more than half of the sites, the model best predicted human disease cases three to four months in the future, and in almost all sites at least two months in the future (the exception is Zaruma, where very few arbovirus cases were reported during the study period and were likely due to importation rather than local transmission). Given that multiple rainfall functions and time lags are supported by field data (even within the same study site), we propose a

conceptual model that incorporates multiple pathways for rainfall to affect disease dynamics along a continuum of rainfall (Fig. 5), in contrast to distinct functional relationships for a given setting, which motivated the approach used in this study.

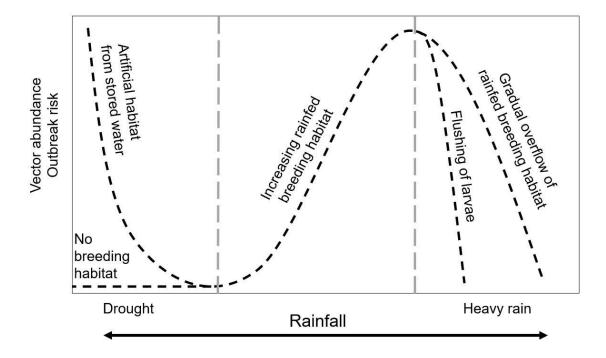


Figure 5: Conceptual model for nonlinear functional relationships between rainfall and vector abundance and arboviral outbreak risk. Dashed lines show multiple potential pathways for rainfall to affect transmission dynamics and include the functional relationships supported in this study. Labels indicate the hypothesized mechanisms along a gradient of rainfall. Adapted from [46].

Factors that mediate disease dynamics predictability

The ability of the model to generate similar dynamics to those found in the field varied with demography, housing quality, and climate. Although the sample size is small (N = 8

sites), we found that the SEI-SEIR model generally predicted vector dynamics better in sites with a smaller proportion of young children in the population (R^2 = 0.89, p < 0.01; Fig. 6a), lower mean temperature (R^2 = 0.63, p < 0.05; Fig. 6c), and a larger proportion of homes with piped water (R^2 = 0.76, p < 0.01; Fig. 6b) and made of cement (R^2 = 0.69, p < 0.05; Fig. 6d; list of all factors we assessed are provided in Table 1). Based on the range of mean temperatures at our study sites ($22 - 28^{\circ}$ C), our findings indicate that vector dynamics become less predictable as temperatures near the optimal temperature for transmission (derived in previous studies as 29°C) following the shape and slope in the R_0 curve (Fig. 7). This complements phenomenological models that have found minimal effects of temperature near the empirically derived thermal optima (Fig. 7). None of the socio-economic factors that we examined in this study (Table 1) explained variability in the pairwise correlations for human disease cases among sites.

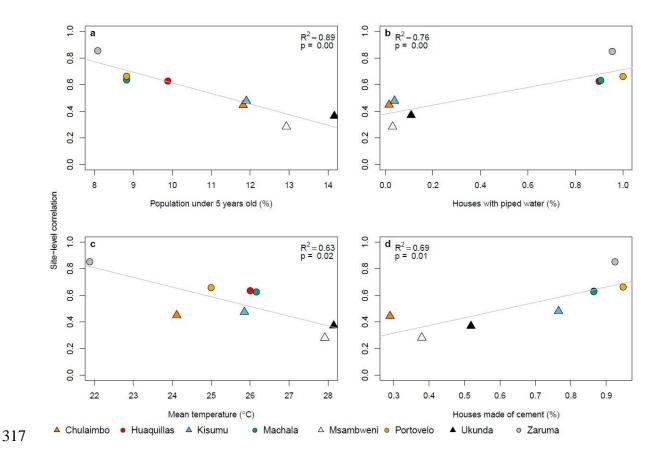
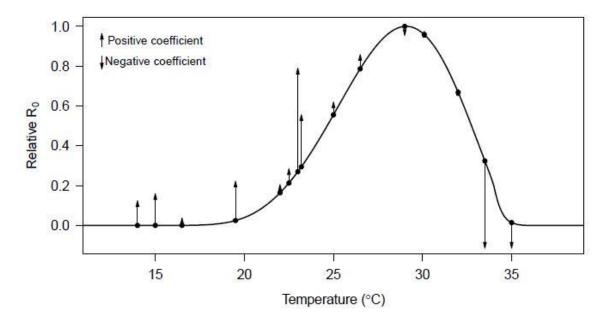


Figure 6: Demography, housing construction, and climate affect model predictive capacity for vectors. Factors that influence the predictability of vector dynamics include (a) proportion of the population under five years of age, (b) proportion of houses with piped water, (c) mean temperature, and (d) proportion of houses made with cement (walls and/or floors). Points indicate the pairwise correlation value for a single site (colors) with different symbols for Ecuador (circles) and Kenya (triangles). Each plot also shows the linear regression lines and associated statistics.



327

328

329

330

331

332

333

334

335

336

337

338

339

340

Figure 7: Independently predicted relative R₀ from a model derived from laboratory studies explains differences in the magnitude and direction of the effects of temperature on dengue transmission in the field across varied settings from **previous studies.** The black line shows the relative basic reproductive number $(R_0,$ normalized to a 0-1 scale) plotted against temperature based on all temperaturedependent traits from [19] used in the SEI-SEIR model presented here. Points indicate mean temperature values from previous field-based statistical analyses that related dengue cases with minimum, maximum, or mean ambient temperature; arrows correspond to the direction (up = positive, down = negative) and relative effect size of the temperature – dengue relationship based on coefficient values from the following studies: [47,48,57,58,49–56]. See Methods and Table S1 for more detail. As expected, the largest observed positive effects of temperature occurred in the rapidly increasing portion of the R₀ curve (~22-25°C; consistent with findings in this study) and the largest observed negative effects occurred well above the predicted optimum, near the upper thermal limit (~33-35°C).

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

Discussion: Directly observing the influence of climate on species interactions and population dynamics is often challenging because of interacting and nonlinear relationships. Here, we directly and quantitatively connect laboratory-based climate relationships to observed mosquito and disease dynamics in the field, supporting the mechanistic role of climate in these disease systems. The trait-based modeling approach captured several key epidemic characteristics and generated a range of disease dynamics along a spectrum of settings with low levels of transmission to seasonal outbreaks, helping to reconcile seemingly context dependent effects (i.e., opposite conclusions about the magnitude and direction of effects; Fig. 7) of climate on arboviral transmission dynamics from the literature [7– 12,47]. The results of this study shed some light on the influence of climate in driving endemic versus epidemic dengue transmission. Although Ecuador typically experiences seasonal epidemics [6] and Kenya typically experiences low levels of year-round transmission [5], the sites within this study suggest that epidemic transmission is more common in settings with clear seasonality (e.g., coastal sites) whereas endemic transmission is more common in settings with more climate variability (e.g., inland sites), regardless of country. Coastal sites experienced more regular seasonal climate cycles, likely because oceans buffer climate variability, and this seasonality corresponded with seasonal epidemics. In contrast, the inland sites experienced more day-to-day climate variability, which resulted in more fluctuations in disease cases. As a result, the occurrence and persistence of

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

suitable temperature, rainfall, and humidity conditions enabling outbreaks were less regular in sites with more climate variability. The ability of the model to detect key epidemic characteristics across endemic and epidemic settings indicates that climate plays a major role in driving when outbreaks occur and how long they last. Using field data on mosquitoes and disease cases from diverse settings and a model parameterized with data from other studies, we identified several key epidemic characteristics that we should (and should not) expect to capture in new settings. While we would never expect a perfect correlation between model predictions and observations, even if the model perfectly captured climate-host-vector dynamics because of the many additional factors that affect transmission in nature, our results indicate that a model with limited calibration can determine the number of outbreaks across settings remarkably well (Fig. 3a). This finding could be particularly useful for prioritizing surveillance or intervention activities across a range of a potential sites that would otherwise appear equal in their propensity for outbreaks (e.g., similar climate conditions). We also show that the model captures the peak timing of outbreaks (Fig. 3b) and outbreak duration (Fig. 3c) but not the final outbreak size (Fig. 3d) or maximum number of infections (Fig. 3e), supporting the hypothesis that the magnitude of disease cases during an outbreak in settings with year-round climate suitability for disease transmission are invariant to temperature, as proposed by [30], likely because the magnitude of disease cases is probably more strongly driven by the availability of susceptible hosts.

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

406

407

408

Given that the model generally did not predict the magnitude of outbreaks, we asked how well the model reproduced vector and human disease dynamics (i.e., variation over time) across sites and whether this relationship varied systematically with different socioeconomic factors. Across sites, the range of temporal correlations between model predictions and observations (N = 8; Fig. 4, Table 2) provides an informative metric for the proportion of true disease dynamics that we might expect to capture in new settings, ranging from 28 - 88%. The correlations varied with demography, housing construction, and climate (Fig. 6). The model may have better explained vector dynamics in locations with a lower proportion of children under five years old for a variety of reasons, including because bottom-heavy demographic pyramids are often associated with lower socioeconomic status and higher mobility throughout the day. In addition to the demographic makeup of sites, housing construction within sites also seems to modify transmission dynamics: vector dynamics were less predictable in sites with more houses with piped water and made of cement (Fig. 6b,d). These results suggest that piped water may prevent additional contact between humans and mosquitoes associated with stored water around the home. In addition, housing materials like cement that lower indoor temperature could artificially decrease climate suitability for mosquitoes, thereby decreasing the probability that mosquitoes will enter and bite people inside their homes. Despite incorporating all known temperature-dependent mosquito traits into the SEI-SEIR model, we still found vector dynamics became less predictable near the empirically derived thermal optima for arboviral transmission (Figs. 6c, 7). This finding may be associated with physiological or behavioral responses of mosquitoes to temperatures near

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

428

429

430

431

their thermal safety margin [59,60] and/or humans modifying their environment (as described above) in locations optimal for transmission. Across the study sites, we found support for three hypothesized relationships between rainfall and mosquito carrying capacity as well as several time lags between model predictions and disease observations. Support for multiple rainfall functions could indicate that the effects of rainfall on immature habitat is highly heterogenous, which has been found in previous research in Ecuador [27] and Kenya [61]. Alternatively, the combination of multiple rainfall relationships and time lags could arise from nonlinear and delayed effects of extreme climate such as droughts and floods. More specifically, we hypothesize that there may be multiple mechanistic relationships for the effects of rainfall on mosquito abundance and arboviral disease dynamics (Fig. 5), and they may act on different time scales. For example, previous research indicated that dengue outbreaks were more likely to occur four to five months after a drought and one month after excessive rainfall and a statistical model that incorporated these duel exposure-lagresponse functions was highly effective at predicting dengue outbreaks in Barbados [62]. Further, if multiple rainfall relationships act in concert across varying time lags, this would help to explain why many different time lags have been observed between rainfall and arboviral dynamics in previous studies [6,27,51,63–65]. Future research can build on this study to improve our understanding of arboviral dynamics across settings. There were several factors that we did not include in this study, such as existing vector control programs, infrastructure, and preexisting immunity in the

population. For instance, in Ecuador, factors such as distance to abandoned properties, interruptions in access to piped water, shaded patios, and use of vector control are documented to influence arbovirus transmission [66], whereas in the study sites in Kenya, factors associated with arboviral transmission are less well studied and there are currently no widely used vector control or local arboviral surveillance programs employed. Future studies could further improve the model by incorporating human immune dynamics associated with interactions among different dengue serotypes [67] or cross-reactivity among viral antibodies [68], differential susceptibility across human age classes [69], and heterogeneity in contact rates between mosquitoes and people based on human behavior and movement [70,71]. Further, as experimental data becomes available for trait estimates specific to chikungunya and Zika, this model could be partitioned to model each arboviral disease individually. This is likely to be an important addition as the different arboviruses tend to peak in different years, possibility due to differences in viral development rates and extrinsic incubation periods among arboviruses. Therefore, validating the model with all three arboviruses combined may oversimplify the complex interannual dynamics that arise due to competition among arboviruses in mosquitoes and humans. There were not enough data for chikungunya and Zika cases in this study to formally test such patterns. This study provides strong evidence that a trait-based model, parameterized independently from field data, can reproduce key epidemic characteristics and a range of spatiotemporal arboviral disease dynamics. Such mechanistic, climatedriven models will become increasingly important to support public health efforts in the face of novel climate regimes emerging due to climate change.

453

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

Materials and Methods:

455

456

457

458

459

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

Climate data We collected in situ measurements of daily mean temperature, relative humidity, and rainfall at each study site and interpolated missing data where necessary. We used temperature and humidity measurements from HOBO loggers and rainfall measurements from rain gauges for sites in Kenya. We used temperature, humidity, and rainfall measurements from automatic weather stations operated by the National Institute of Meteorology and Hydrology in Ecuador. For Kenya, we interpolated missing temperature data from NOAA Global Surface Summary of the Day (Table S2, Fig. S4) and interpolated missing rainfall data from NOAA Climate Prediction Center Africa Rainfall Climatology dataset (Table S2, Fig. S5). For Ecuador, we interpolated missing temperature (Table S2, Fig. S4) and rainfall (Table S2, Fig. S5) data using the nearest study site where possible and otherwise based on long term mean values for the corresponding Julian day. To interpolate missing data, we linearly regressed all measurements taken on the same day in two datasets and then used the linear model to interpolate temperature for the site with missing data based on the climate measurement from the secondary source for the date when the data was missing (Figs. S4-5). For rainfall, we first calculated a moving window of 14-day accumulated rainfall (which is short enough to capture variability and seasonality in rainfall patterns and follows [72]) for each day before interpolation because modeled daily rainfall values are less reliable than accumulated rainfall over a two week period. We interpolated 14-day cumulative rainfall for any day with a missing rainfall value in the prior 14 days. For both Kenya and Ecuador, we interpolated missing relative humidity data based on long term mean values

- 478 for the corresponding Julian day (Table S2). We then calculated the saturation vapor
- pressure deficit (SVPD) from temperature and humidity to use in the humidity function
- because previous research suggests SVPD is a more informative measure of the effect of
- humidity on mosquito survival compared with relative humidity [73]. To calculate
- 482 SVPD, we first calculated the saturation vapor pressure as:

$$SVP = 610.7 * 10^{7.5*T/(273.3+T)}$$
 (1)

where (*T*) is temperature in degrees Celsius. We then calculated SVPD (in kilopascals) as

$$SVPD = 1 - \frac{RH}{100} * SVP \tag{2}$$

- 484 where RH is relative humidity. The final dataset had no missing values for temperature
- 485 (Fig. S6), rainfall (Fig. S7), and humidity (Fig. S8).
- 487 Vector surveys

- We collected, counted, sexed, and classified mosquitoes by species, and aggregated the
- data to mean number of *Aedes aegypti* per house, month, year, and site to account for
- 490 differences in survey effort across months and sites. We collected adult mosquitoes using
- 491 Prokopack aspirators [74]. In Ecuador, we collected mosquitoes from approximately 27
- houses per site (range = 3-57 houses across four sites) every one-to-two weeks during
- 493 three, four-month sampling periods between July 2016 and August 2018 (≈ 37 sampling
- 494 weeks per site) to capture different parts of the transmission season. We aggregated the
- Ecuador vector data to monthly values (≈ 15 sampling months per site) to correspond
- 496 with the temporal resolution of surveys in Kenya. In Kenya, we collected mosquitoes
- from approximately 20 houses per site (range = 1-47 houses across four sites) every
- 498 month between January 2014 and October 2018 (\approx 54 sampling months per site). In

500

501

502

503

504

505

506

507

508

509

510

511

512

513

514

515

516

517

518

519

520

521

Kenya, we also collected pupae, late instars, and early instars from containers with standing water around the home and collected eggs by setting ovitraps for an average of four days in and around each house monthly. We brought pupae, late and early instars, and eggs to the insectary and reared them to adulthood to classify individuals by sex and species. All mosquito traps capture a small portion of the true mosquito population; therefore, using consistent trapping methods at the same locations through time allows us to compare relative mosquito population dynamics across study sites rather than the absolute magnitude of mosquito abundances. Arboviral surveys For Ecuador, we analyzed laboratory-confirmed dengue, chikungunya, and Zika cases provided by the Ministry of Health (MoH) of Ecuador. The MoH collects serum samples from a subset of people with suspected arbovirus infections, and samples are tested at the National Public Health Research Institute by molecular diagnostics (RT-PCR) or antibody tests (IgM ELISA for dengue), depending on the number of days of illness. Results are sent to the MoH Epidemiological Surveillance and Control National Directorate (SIVE Alerta system). Laboratory-confirmed dengue cases were available for all four sites from 2014 to 2018. Laboratory-confirmed chikungunya cases were available for Machala and Huaquillas from 2015 to 2018. Laboratory-confirmed Zika cases were available for Machala from 2016 to 2018. For Kenya, we used laboratory-confirmed dengue cases aggregated by site and month between 2014 and 2018 collected in a passive surveillance study on childhood febrile

523

524

525

526

527

528

529

530

531

532

533

534

535

536

537

538

539

540

541

542

543

544

illness in Kenya (NIH R01AI102918, PI: ADL). The study population consisted of 7,653 children less than 18 years of age with undifferentiated febrile illness. Children with fever enrolled in the study when attending outpatient care in one of the four study sites (Mbaka Oromo Health Centre in Chulaimbo, Obama Children's Hospital in Kisumu, Msambweni District Hospital in Msambweni, and Ukunda/Diani Health Center in Ukunda). Local health officers collected comprehensive clinical and demographic data and phlebotomy at the initial visit. We tested each child's blood for dengue viremia by molecular diagnostics (conventional PCR [75] or targeted multiplexed real-time PCR when available [76]), or serologic conversion between an initial and a follow up visit (IgG ELISA [77]). For arboviral data collection in Ecuador and Kenya, participants provided consent and all local and institutional protocols were followed. SEI-SEIR model We adapted an SEI-SEIR model parameterized for dengue transmission in Ae. aegypti mosquitoes [30] to simulate mosquito abundance and arboviral cases through time based on daily weather conditions in eight study locations. The model (equations 3-9; Fig. 2), created independently from the observed data described above, allows mosquito life history traits and viral development rate to vary with temperature (r) following [30], mosquito carrying capacity to vary with accumulated 14-day rainfall (R) following [72], and mosquito mortality to vary with humidity (i.e., saturation vapor pressure deficit) (H) following [73].

$$\frac{dS_m}{dt} = \varphi(T, H) * \frac{1}{\mu(T, H)} * N_m * \left(1 - \frac{N_m}{K(T, R, H)}\right) - \left(\alpha(T) * pMI(T) * \frac{I_h}{N_h} + \mu(T, H)\right) * S_m$$
(3)

$$\frac{dE_m}{dt} = a(T) * pMI(T) * \frac{I_h}{N_h} * S_m - \left(PDR(T) + \mu(T, H)\right) * E_m$$
(4)

$$\frac{dI_m}{dt} = PDR(T) * E_m - \mu(T, H) * I_m$$
(5)

$$\frac{dS_h}{dt} = -a(T) * b(T) * \frac{I_m}{N_h} * S_h + BR * S_h - DR * S_h + ie * N_h - ie * S_h$$
(6)

$$\frac{dE_h}{dt} = a(T) * b(T) * \frac{I_m}{N_h} * S_h - \delta * E_h - DR * E_h - ie * E_h$$

$$\tag{7}$$

$$\frac{dI_h}{dt} = \delta * E_h - \eta * I_h - DR * I_h - ie * I_h \tag{8}$$

$$\frac{dR_h}{dt} = \eta * I_h - DR * R_h - ie * R_h \tag{9}$$

546 where

$$\varphi(T,H) = EFD(T) * pEA(T) * MDR(T)$$
(10)

The adult mosquito population (N_m) is separated into susceptible (S_m), exposed (E_m), and infectious (I_m) compartments and the human population (N_h) is separated into susceptible (S_h), exposed (E_h), infectious (I_h), and recovered (R_h) compartments (Fig. 2). Climate-independent model parameters (Table 3) include the intrinsic incubation period (s), human infectivity period (η), birth rate (B_R), death rate (D_R), and immigration/emigration rate (D_R). The temperature-dependent SEI-SEIR model was developed by Huber et al. [30] and allows mosquito life history traits and viral development rate to vary according to thermal response curves fit from data derived in laboratory experiments conducted at constant temperatures (Table 3). Although laboratory experiments do not reflect real-world conditions, the physiological responses measured are biologically meaningful. The temperature-dependent traits include eggs laid per female per day (D_R), the probability of egg-to-adult survival (D_R), mosquito development rate (D_R), mosquito mortality rate (lifespan⁻¹; D_R), biting rate (D_R), probability of mosquito infection per bite on an infectious

- host (pm1), parasite development rate (PDR), and probability of mosquito infectiousness given an infectious bite (b). We modified the mosquito mortality rate equation to vary as
- a function of temperature and humidity by fitting a spline model based on a pooled
- survival analysis of Ae. aegypti [73] (Fig. S9):

$$\mu(T,H) = \frac{1}{c * (T - T_0) * (T - T_m)} + (1 - (0.01 + 2.01 * H)) * y \qquad H < 1$$
 (11)

$$\mu(T,H) = \frac{1}{c * (T - T_0) * (T - T_m)} + (1 - (1.22 + 0.27 * H)) * y \qquad H \ge 1$$
 (12)

- where the rate constant (c), minimum temperature (T_0) , and maximum temperature (T_m)
- 565 equal -1.24, 16.63, and 31.85 respectively (Table 4), humidity (H) is the saturation vapor
- pressure deficit, and y is a scaling factor that we set to 0.005 and 0.01, respectively, to
- restrict mosquito mortality rates within the range of mortality rates estimated by other
- studies [19,73]. The linear humidity function has a steeper slope at lower humidity values
- (equation 11) compared with higher humidity values (equation 12) based on previous
- 570 research [73] (Fig. S9).
- We modeled adult mosquito carrying capacity, K, as a modified Arrhenius equation
- 573 following [30,78]:

$$K(T, H, R) = \frac{EFD(T_0) * pEA(T_0) * MDR(T_0) * \mu(T_0, H_0)^{-1} - \mu(T_0, H_0)}{EFD(T_0) * pEA(T_0) * MDR(T_0) * \mu(T_0, H_0)^{-1}} * N_{m.max}$$

$$* e^{\frac{-E_A * (T - T_0)^2}{K_B * (T + 273) * (T_0 + 273)}} * f(R)$$
(13)

- with T_0 and H_0 set to the temperature and humidity where carrying capacity is greatest (i.e.,
- 575 physiological optimal conditions from laboratory experiments; 29°C and 6 kPA), $N_{m.max}$
- set to the maximum possible mosquito abundance in a population (twice the human
- population size following [30]), and the Boltzmann constant, (K_B) , is 8.617 x 10^{-5} eV/K.

We set the activation energy, E_A , as 0.05 based on [30]. Since there were no experimental data from which to derive the functional response of mosquito carrying capacity across a gradient of rainfall values, we tested several functional relationships based on hypothesized biological relationships between freshwater availability and immature mosquito breeding habitat, modeling the effect of rainfall on carrying capacity, f(R), as either:

$$f(R_{\text{Brière}}) = c * R * (R - R_{min}) * \sqrt{(R_{max} - R)} * z$$

$$\tag{14}$$

$$f(R_{Quadratic}) = c * (R - R_{min}) * (R - R_{max}) * z$$
(15)

$$f(R_{\text{Inverse}}) = \frac{1}{R} * z \tag{16}$$

where minimum rainfall (R_{min}) equaled 1 mm and maximum rainfall (R_{max}) equaled 123 mm based on the high probability of flushing [26]. The quadratic function is similar to the rainfall function found in [26] and the inverse function is based on the rainfall function used in [72]. We used rate constants (c) of 7.86e⁻⁵ and -5.99e⁻³ for the Brière and quadratic functions respectively, based on rate constants for other parameters with similar functional forms (Table 4). We also included a scaling factor, z (0. 28, 0.025, and 0.60 respectively), to restrict the maximum carrying capacity to produce model outputs based on a subsample of the total population for comparison with observations. Since the rate constant, c, is multiplied by z, inferring the exact value of c is not necessary because it is scaled by c. The scaling factor could be removed from the model to simulate dynamics in the total population.

Table 3: Values of temperature-invariant parameters used in the model. We derived daily birth and death rates in the model by dividing the per capita birth and death rates by 360 days. The World Bank Open Data can be found at https://data.worldbank.org/.

Parameter	Definition	Value	Source
δ^{-1}	Intrinsic incubation period (days)	5.9	[30]
η^{-1}	Human infectivity period (days)	5.0	[30]
BR	Annual birth rate (per 1000 people)	31.782 (Ecuador) 20.175 (Kenya)	The World Bank Open Data
DR	Annual death rate (per 1000 people)	5.284 (Ecuador) 5.121 (Kenya)	The World Bank Open Data
ie	Immigration/emigration rate	0.01	Expert opinion

Table 4: Fitted thermal responses for Ae. aegypti life history traits. Traits were fit to a Brière $[cT(T-T_0)(T_m-T)^{\frac{1}{2}}]$ or a quadratic $[c(T-T_m)(T-T_0)]$ function where T represents temperature. T_0 and T_m are the critical thermal minimum and maximum, respectively, and c is the rate constant. Thermal responses were fit by [19] and also used in [30]. Parasite development rate was measured as the virus extrinsic incubation rate.

Trait	Definition	Function	С	T_0	$T_{\rm m}$
а	Biting rate (day ⁻¹)	Brière	2.71x10 ⁻⁰⁴	14.67	41.00
EFD	Eggs laid per female per day	Brière	2.08x10 ⁻⁰²	14.06	32.03
pEA	Probability of mosquito egg-to-adult survival	Quadratic	-3.36x10 ⁻⁰³	7.68	38.31
MDR	Mosquito egg-to-adult development rate (day ⁻¹)	Brière	1.49x10 ⁻⁰⁴	15.12	37.67
Lf	Adult mosquito lifespan (days)	Quadratic	-1.24	16.63	31.85
b	Probability of mosquito infectiousness	Brière	9.86x10 ⁻⁰⁴	12.05	32.79
pMI	Probability of mosquito infection	Brière	5.23x10 ⁻⁰⁴	1.51	34.74
PDR	Parasite development rate (day ⁻¹)	Brière	1.04×10^{-04}	11.50	38.97

To initiate the model, we used site-specific values for human population size and randomly selected one set of values for all sites for the proportion of mosquitoes and humans in each compartment. For Ecuador, we used population estimates from official population projections produced by Proyección de la Población Ecuatoriana, por años calendario, según cantones 2010-2020

(https://www.ecuadorencifras.gob.ec/proyecciones-poblacionales/) with population sizes of 57,366, 279,887, 13,673, and 25,615 for Huaquillas, Machala, Portovelo, and Zaruma, respectively, based on 2017 projections. For Kenya, we estimated the population sizes

629

630

631

632

633

634

635

636

637

638

639

640

641

642

643

644

645

646

647

648

649

650

served by each outpatient care facility by creating a polygon around all the geolocations of study participants' homes enrolled at each outpatient care facility and summed population count data from NASA's Socioeconomic Data and Applications Center Gridded Population of the World v4 (https://doi.org/10.7927/H4JW8BX5) within each polygon using ArcGIS v 10.4.1. We estimated population sizes of 7,304, 547,557, 240,698, and 154,048 for Chulaimbo, Kisumu, Msambweni, and Ukunda, respectively. We set the ratio of mosquitoes to humans to two, following [30]. We used the following values as the initial proportion of mosquitoes and humans in each model compartment: $S_m = 0.22$, $E_m = 0.29$, $I_m = 0.49$, $S_h = 0.58$, $E_h = 0.22$, $I_h = 0.00$, and $R_h = 0.20$. We determined that the model was invariant to initial proportion values after a short burn-in period (90 days) based on a sensitivity analysis (Fig. S10); therefore, we randomly selected one set of initial proportion values from the sensitivity analysis for all the model simulations. We also determined that the temporal trajectories of model dynamics did not change when we varied the critical thermal minimum, maximum, and rate constants (Table 4) for *Aedes aegypti* life history traits (Fig. S1-2). We ran all model simulations using the deSolve package in R statistical software v 3.5.3 [79]. Model validation To validate the SEI-SEIR model, we calculated pairwise correlations with an adjusted pvalue to account for autocorrelation for each site. For the pairwise correlations, we used the ccf function in base R [79] to calculate correlations between the two times series of

652

653

654

655

656

657

658

659

660

661

662

663

664

665

666

667

668

669

670

671

672

673

model predictions and observations with 0, 1, 2, 3, 4, and 5-month lags. We then calculated an adjusted p-value using the Modified Chelton method [45] to adjust the null hypothesis test of sample correlation between autocorrelated time series. To assess predictions and observations for vector dynamics for each site, we compared monthly time series of the total predicted mosquito population from the SEI-SEIR model with the monthly time series of mean number of Aedes aegypti (per house). We followed the same procedure to compare model predictions with other mosquito life stages for sites in Kenya. Similarly, to compare predictions and observations for human disease dynamics for each site, we compared monthly times series of predicted infected individuals from the SEI-SEIR model with the monthly time series of total laboratory-confirmed arboviral cases. For subsequent analyses, we used model predictions from the model (e.g., SEI-SEIR model with a specific rainfall function and time lag) with the highest pairwise correlation value. To compare key epidemic characteristics between model predictions and observations and to compare site-specific correlations with socio-economic factors, we used linear regression models using the lm function in that stats package in R [79]. We defined outbreaks as a continuous time period where the peak cases exceeded the mean number of cases (predicted or observed) plus one standard deviation within a site. We then used those outbreak periods to count the total number of outbreaks within each site, and, for predicted and observed outbreaks that overlapped in time, the duration, peak timing total outbreak size, and maximum number of infections. We compared predictions and observations for each of these metrics with linear regression. Since we were interested in

674

675

676

677

678

679

680

681

682

683

684

685

686

687

688

689

690

691

692

693

694

695

696

whether model predictions matched observations for each independent outbreak period, we did not allow varying intercepts or slopes by site. Similarly, we compared the pairwise correlation values (described above) across all sites with each socio-economic factor listed in Table 1 separately using linear regressions. Comparison of R_0 with prior studies We collected effect sizes of temperature on dengue incidence from 12 peer-reviewed studies from the literature (Table S1). We selected studies with mean temperatures across the predicted temperature range where arboviral transmission can occur. We scaled the coefficient values to visualize the relative effect of temperature across studies given that the original analyses were conducted with different temperature metrics and across different temperature ranges. We provide additional information and sources in Table S1. Data availability statement: Climate data, epidemic characteristics data, and socioeconomic data are available at https://github.com/jms5151/SEI-SEIR_Arboviruses. We used this data to create figures 3, 4, and 6 in the main text and supplemental figures 1-3 and 6-10; we provide the data used in figure 7 in supplemental table 1. We can provide vector and arboviral case data upon request with permission from appropriate data providers (e.g., Ecuador Ministry of Health). **Code availability:** Model and analysis codes are available at https://github.com/jms5151/SEI-SEIR_Arboviruses.

697

698

699

700

701

702

703

704

705

706

707

708

709

710

711

712

713

714

715

716

717

718

719

Acknowledgements: JMC, ADL, EFL, and EAM were supported by a Stanford Woods Institute for the Environment – Environmental Ventures Program grant (PIs: EAM, ADL, and EFL). EAM was also supported by a Hellman Faculty Fellowship and a Terman Award. ADL, BAN, FMM, ENGS, MSS, ARK, RD, AA, and HNN were supported by a National Institutes of Health R01 grant (AI102918; PI: ADL). EAM, AMSI, and SJR were supported by a National Science Foundation (NSF) Ecology and Evolution of Infectious Diseases (EEID) grant (DEB-1518681) and AMSI and SJR were also supported by an NSF DEB RAPID grant (1641145). EAM was also supported by a National Institute of General Medical Sciences Maximizing Investigators' Research Award grant (R35GM133439) and an NSF and Fogarty International Center EEID grant (DEB-2011147). We thank Cat Lippi for assistance with formatting household quality survey data from Ecuador. Author contributions: EAM, ADL, EFL, and JMC conceived of project. JMC conducted analyses and wrote manuscript. EAM, ADL, EFL, and AMSI secured funding for the project. BNN, FMM, EBA, AA, MJBC, RD, FHH, RM, and HNN collected data. ENGS and MMS conducted laboratory analyses. ARK, SJR, and RS processed data. All authors revised and approved of the manuscript.

References:

720

- 721 1. Ockendon N, Baker DJ, Carr JA, White EC, Almond REA, Amano T, et al.
- Mechanisms underpinning climatic impacts on natural populations: altered species
- interactions are more important than direct effects. Glob Chang Biol. 2014;20:
- 724 2221–2229. doi:10.1111/gcb.12559
- 725 2. Boggs CL, Inouye DW. A single climate driver has direct and indirect effects on
- 726 insect population dynamics. Ecol Lett. 2012;15: 502–508. doi:10.1111/j.1461-
- 727 0248.2012.01766.x
- 3. Burkett VR, Wilcox DA, Stottlemyer R, Barrow W, Fagre D, Baron J, et al.
- Nonlinear dynamics in ecosystem response to climatic change: Case studies and
- policy implications. Ecol Complex. 2005;2: 357–394.
- 731 doi:10.1016/j.ecocom.2005.04.010
- 732 4. Molnár PK, Sckrabulis JP, Altman KA, Raffel TR. Thermal Performance Curves
- and the Metabolic Theory of Ecology—A Practical Guide to Models and
- Experiments for Parasitologists. J Parasitol. 2017;103. doi:10.1645/16-148
- 735 5. Hortion J, Mutuku FM, Eyherabide AL, Vu DM, Boothroyd DB, Grossi-Soyster
- 736 EN, et al. Acute Flavivirus and Alphavirus Infections among Children in Two
- 737 Different Areas of Kenya, 2015. Am J Trop Med Hyg. 2019;100: 170–173.
- 738 doi:10.4269/ajtmh.18-0297
- 739 6. Stewart-Ibarra AM, Lowe R. Climate and Non-Climate Drivers of Dengue
- 740 Epidemics in Southern Coastal Ecuador. Am J Trop Med Hyg. 2013;88: 971–981.
- 741 doi:10.4269/ajtmh.12-0478
- 742 7. Jury MR. Climate influence on dengue epidemics in Puerto Rico. Int J Environ

- 743 Health Res. 2008;18: 323–334. doi:10.1080/09603120701849836
- 744 8. Campbell KM, Haldeman K, Lehnig C, Munayco C V., Halsey ES, Laguna-Torres
- VA, et al. Weather Regulates Location, Timing, and Intensity of Dengue Virus
- Transmission between Humans and Mosquitoes. Michael E, editor. PLoS Negl
- 747 Trop Dis. 2015;9: e0003957. doi:10.1371/journal.pntd.0003957
- 748 9. Adde A, Roucou P, Mangeas M, Ardillon V, Desenclos J-C, Rousset D, et al.
- Predicting Dengue Fever Outbreaks in French Guiana Using Climate Indicators.
- 750 Scarpino S V., editor. PLoS Negl Trop Dis. 2016;10: e0004681.
- 751 doi:10.1371/journal.pntd.0004681
- 752 10. Dhimal M, Gautam I, Joshi HD, O'Hara RB, Ahrens B, Kuch U, et al. Risk
- Factors for the Presence of Chikungunya and Dengue Vectors (Aedes aegypti and
- Aedes albopictus), Their Altitudinal Distribution and Climatic Determinants of
- 755 Their Abundance in Central Nepal. Turell MJ, editor. PLoS Negl Trop Dis.
- 756 2015;9: e0003545. doi:10.1371/journal.pntd.0003545
- 757 11. Descloux E, Mangeas M, Menkes CE, Lengaigne M, Leroy A, Tehei T, et al.
- 758 Climate-Based Models for Understanding and Forecasting Dengue Epidemics.
- Anyamba A, editor. PLoS Negl Trop Dis. 2012;6: e1470.
- 760 doi:10.1371/journal.pntd.0001470
- 761 12. Aswi A, Cramb SM, Moraga P, Mengersen K. Epidemiology and Infection
- Bayesian spatial and spatio-temporal approaches to modelling dengue fever: a
- 763 systematic review. Epidemiol Infect. 2018;147. doi:10.1017/S0950268818002807
- 764 13. Johansson MA, Apfeldorf KM, Dobson S, Devita J, Buczak AL, Baugher B, et al.
- An open challenge to advance probabilistic forecasting for dengue epidemics. Proc

- 766 Natl Acad Sci. 2019; 201909865. doi:10.1073/pnas.1909865116
- 767 14. Michael E, Singh BK, Mayala BK, Smith ME, Hampton S, Nabrzyski J.
- 768 Continental-scale, data-driven predictive assessment of eliminating the vector-
- borne disease, lymphatic filariasis, in sub-Saharan Africa by 2020. BMC Med.
- 770 2017;15: 176. doi:10.1186/s12916-017-0933-2
- 771 15. Smith T, Maire N, Ross A, Penny M, Chitnis N, Schapira A, et al. Towards a
- comprehensive simulation model of malaria epidemiology and control.
- 773 Parasitology. 2008. pp. 1507–1516. doi:10.1017/S0031182008000371
- 774 16. Ryan SJ, Carlson CJ, Mordecai EA, Johnson LR. Global expansion and
- redistribution of Aedes-borne virus transmission risk with climate change. Han
- 776 BA, editor. PLoS Negl Trop Dis. 2019;13: e0007213.
- 777 doi:10.1371/journal.pntd.0007213
- 778 17. Kraemer MU, Sinka ME, Duda KA, Mylne AQ, Shearer FM, Barker CM, et al.
- The global distribution of the arbovirus vectors *Aedes aegypti* and *Ae. albopictus*.
- 780 Elife. 2015;4. doi:10.7554/eLife.08347
- 781 18. Powell JR, Tabachnick WJ, Powell JR, Tabachnick WJ. History of domestication
- and spread of Aedes aegypti A Review. Mem Inst Oswaldo Cruz. 2013;108: 11–
- 783 17. doi:10.1590/0074-0276130395
- 784 19. Mordecai EA, Cohen JM, Evans M V., Gudapati P, Johnson LR, Lippi CA, et al.
- Detecting the impact of temperature on transmission of Zika, dengue, and
- chikungunya using mechanistic models. Althouse B, editor. PLoS Negl Trop Dis.
- 787 2017;11: e0005568. doi:10.1371/journal.pntd.0005568
- 788 20. Shocket MS, Ryan SJ, Mordecai EA. Temperature explains broad patterns of Ross

- 789 River virus transmission. Elife. 2018; doi:10.7554/eLife.37762.001
- 790 21. Paull SH, Horton DE, Ashfaq M, Rastogi D, Kramer LD, Diffenbaugh NS, et al.
- 791 Drought and immunity determine the intensity of West Nile virus epidemics and
- 792 climate change impacts. Proc R Soc B Biol Sci. 2017;284: 20162078.
- 793 doi:10.1098/rspb.2016.2078
- 794 22. Costa EAP de A, Santos EM de M, Correia JC, Albuquerque CMR de. Impact of
- small variations in temperature and humidity on the reproductive activity and
- survival of Aedes aegypti (Diptera, Culicidae). Rev Bras Entomol. 2010;54: 488–
- 797 493. doi:10.1590/S0085-56262010000300021
- 798 23. Gaaboub IA, El-Sawaf SK, El-Latif MA. Effect of Different Relative Humidities
- and Temperatures on Egg-Production and Longevity of Adults of Anopheles
- 800 (Myzomyia) pharoensis Theob.1. Zeitschrift für Angew Entomol. 2009;67: 88–94.
- 801 doi:10.1111/j.1439-0418.1971.tb02098.x
- 802 24. Koenraadt CJM, Harrington LC. Flushing Effect of Rain on Container-Inhabiting
- Mosquitoes Aedes aegypti and Culex pipiens (Diptera: Culicidae). J Med Entomol.
- 804 2009;45: 28–35. doi:10.1603/0022-2585(2008)45[28:FEOROC]2.0.CO;2
- Paaijmans KP, Wandago MO, Githeko AK, Takken W, Vulule J. Unexpected High
- Losses of Anopheles gambiae Larvae Due to Rainfall. Carter D, editor. PLoS One.
- 807 2007;2: e1146. doi:10.1371/journal.pone.0001146
- 808 26. Benedum CM, Seidahmed OME, Eltahir EAB, Markuzon N. Statistical modeling
- of the effect of rainfall flushing on dengue transmission in Singapore. Reiner RC,
- editor. PLoS Negl Trop Dis. 2018;12: e0006935.
- 811 doi:10.1371/journal.pntd.0006935

812 27. Stewart Ibarra AM, Ryan SJ, Beltrán E, Mejía R, Silva M, Muñoz Á. Dengue 813 Vector Dynamics (Aedes aegypti) Influenced by Climate and Social Factors in 814 Ecuador: Implications for Targeted Control. Mores CN, editor. PLoS One. 2013;8: 815 e78263. doi:10.1371/journal.pone.0078263 816 28. Pontes RJ, Spielman A, Oliveira-Lima JW, Hodgson JC, Freeman J. Vector 817 densities that potentiate dengue outbreaks in a Brazilian city. Am J Trop Med Hyg. 818 2000;62: 378–383. doi:10.4269/ajtmh.2000.62.378 819 29. Anyamba A, Linthicum KJ, Small JL, Collins KM, Tucker CJ, Pak EW, et al. 820 Climate Teleconnections and Recent Patterns of Human and Animal Disease 821 Outbreaks. Zhou X-N, editor. PLoS Negl Trop Dis. 2012;6: e1465. 822 doi:10.1371/journal.pntd.0001465 823 30. Huber JH, Childs ML, Caldwell JM, Mordecai EA. Seasonal temperature variation 824 influences climate suitability for dengue, chikungunya, and Zika transmission. 825 Althouse B, editor. PLoS Negl Trop Dis. 2018;12: e0006451. 826 doi:10.1371/journal.pntd.0006451 827 31. Lourenço J, Recker M. The 2012 Madeira Dengue Outbreak: Epidemiological 828 Determinants and Future Epidemic Potential. Scarpino S V., editor. PLoS Negl 829 Trop Dis. 2014;8: e3083. doi:10.1371/journal.pntd.0003083 830 32. Li R, Xu L, Bjørnstad ON, Liu K, Song T, Chen A, et al. Climate-driven variation 831 in mosquito density predicts the spatiotemporal dynamics of dengue. Proc Natl 832 Acad Sci. 2019;119: 3624–3629. doi:10.1073/PNAS.1806094116 833 33. Wang X, Tang S, Cheke RA. A stage structured mosquito model incorporating 834 effects of precipitation and daily temperature fluctuations. J Theor Biol. 2016;411:

835 27–36. doi:10.1016/j.jtbi.2016.09.015 836 34. Siraj AS, Oidtman RJ, Huber JH, Kraemer MUG, Brady OJ, Johansson MA, et al. 837 Temperature modulates dengue virus epidemic growth rates through its effects on 838 reproduction numbers and generation intervals. Althouse B, editor. PLoS Negl 839 Trop Dis. 2017;11: e0005797. doi:10.1371/journal.pntd.0005797 840 35. Oidtman RJ, Lai S, Huang Z, Yang J, Siraj AS, Reiner RC, et al. Inter-annual 841 variation in seasonal dengue epidemics driven by multiple interacting factors in 842 Guangzhou, China. Nat Commun. 2019;10. doi:10.1038/s41467-019-09035-x 843 36. Stewart-Ibarra AM, Muñoz ÁG, Ryan SJ, Ayala EB, Borbor-Cordova MJ, 844 Finkelstein JL, et al. Spatiotemporal clustering, climate periodicity, and social-845 ecological risk factors for dengue during an outbreak in Machala, Ecuador, in 846 2010. BMC Infect Dis. 2014;14: 610. doi:10.1186/s12879-014-0610-4 847 37. Agha SB, Tchouassi DP, Turell MJ, Bastos ADS, Sang R. Entomological 848 assessment of dengue virus transmission risk in three urban areas of Kenya. Reiner 849 RC, editor. PLoS Negl Trop Dis. 2019;13: e0007686. 850 doi:10.1371/journal.pntd.0007686 851 38. Agha SB, Tchouassi DP, Bastos ADS, Sang R. Dengue and yellow fever virus 852 vectors: seasonal abundance, diversity and resting preferences in three Kenyan 853 cities. Parasit Vectors. 2017;10: 628. doi:10.1186/s13071-017-2598-2 854 39. Chretien J-P, Anyamba A, Bedno SA, Breiman RF, Sang R, Sergon K, et al. 855 Drought-Associated Chikungunya Emergence Along Coastal East Africa. Am J 856 Trop Med Hyg. 2007;76: 405–407. doi:10.4269/ajtmh.2007.76.405 857 40. Vu DM, Mutai N, Heath CJ, Vulule JM, Mutuku FM, Ndenga BA, et al.

- Unrecognized Dengue Virus Infections in Children, Western Kenya, 2014-2015.
- 859 Emerg Infect Dis. 2017;23: 1915–1917. doi:10.3201/eid2311.170807
- 860 41. Gubler DJ, Nalim S, Saroso JS, Saipan H, Tan R. Variation in Susceptibility to
- Oral Infection with Dengue Viruses among Geographic Strains of Aedes Aegypti
- *. Am J Trop Med Hyg. 1979;28: 1045–1052. doi:10.4269/ajtmh.1979.28.1045
- 42. Xavier-Carvalho C, Chester Cardoso C, de Souza Kehdya F, Guilherme Pacheco
- A, Ozório Moraesa M. Host genetics and dengue fever. Infect Genet Evol.
- 865 2017;56: 99–110. doi:10.1016/J.MEEGID.2017.11.009
- 866 43. Didan K, Barreto Munoz A, Solano R, Huete A. MODIS Vegetation Index User's
- Guide (MOD13 Series) [Internet]. Available: http://vip.arizona.edu
- 868 44. Sulla-Menashe D, Friedl MA. User Guide to Collection 6 MODIS Land Cover
- 869 (MCD12Q1 and MCD12C1) Product. 2018; doi:10.5067/MODIS/MCD12Q1
- 870 45. Pyper BJ, Peterman RM. Comparison of methods to account for autocorrelation in
- correlation analyses of fish data. Can J Fish Aquat Sci. 1998;55: 2127–2140.
- 872 doi:10.1139/f98-104
- 873 46. Shocket MS, Anderson CB, Caldwell JM, Childs ML, Han S, Harris M, et al.
- 874 Environmental drivers of vector-borne disease. Population Biology of Vector-
- borne Diseases. Oxford University Press;
- 876 47. Hurtado-Daz M, Riojas-Rodrguez H, Rothenberg S, Gomez-Dantes H, Cifuentes
- E. Impact of climate variability on the incidence of dengue in Mexico. Trop Med
- 878 Int Heal. 2007;12. doi:10.1111/j.1365-3156.2007.01930.x
- 879 48. Colón-González FJ, Bentham G, Lake IR. Climate Variability and Dengue Fever
- in Warm and Humid Mexico. Am J Trop Med Hyg. 2011;84: 757–763.

- 881 doi:10.4269/ajtmh.2011.10-0609
- Wang C, Jiang B, Fan J, Wang F, Liu Q. A Study of the Dengue Epidemic and
- Meteorological Factors in Guangzhou, China, by Using a Zero-Inflated Poisson
- Regression Model. Asia Pacific J Public Heal. 2014;26: 48–57.
- 885 doi:10.1177/1010539513490195
- 886 50. Minh An DT, Rocklöv J. Epidemiology of dengue fever in Hanoi from 2002 to
- 2010 and its meteorological determinants. Glob Health Action. 2014;7: 23074.
- 888 doi:10.3402/gha.v7.23074
- 889 51. Laureano-Rosario AE, Garcia-Rejon JE, Gomez-Carro S, Farfan-Ale JA, Muller-
- Kargera FE. Modelling dengue fever risk in the State of Yucatan, Mexico using
- regional-scale satellite-derived sea surface temperature. Acta Trop. 2017;172: 50–
- 892 57. doi:10.1016/j.actatropica.2017.04.017
- 893 52. Wu P-C, Guoa H-R, Lung S-C, Lin C-Y, Su H-J. Weather as an effective predictor
- for occurrence of dengue fever in Taiwan. Acta Trop. 2007;103: 50–57.
- 895 doi:10.1016/j.actatropica.2007.05.014
- 896 53. Karim MN, Munshi SU, Anwar N, Alam MS. Climatic factors influencing dengue
- cases in Dhaka city: a model for dengue prediction. Indian J Med Res. 2012;136:
- 898 32–9. Available: http://www.ncbi.nlm.nih.gov/pubmed/22885261
- 899 54. Nakhapakorn K, Tripathi N. An information value based analysis of physical and
- climatic factors affecting dengue fever and dengue haemorrhagic fever incidence.
- 901 Int J Health Geogr. 2005;4: 13. doi:10.1186/1476-072X-4-13
- 902 55. Gharbi M, Quenel P, Gustave J, Cassadou S, Ruche G La, Girdary L, et al. Time
- series analysis of dengue incidence in Guadeloupe, French West Indies:

904 Forecasting models using climate variables as predictors. BMC Infect Dis. 905 2011;11: 166. doi:10.1186/1471-2334-11-166 906 56. Sharmin S, Glass K, Viennet E, Harley D. Interaction of Mean Temperature and 907 Daily Fluctuation Influences Dengue Incidence in Dhaka, Bangladesh. Kasper M, 908 editor. PLoS Negl Trop Dis. 2015;9: e0003901. doi:10.1371/journal.pntd.0003901 909 57. Sriprom M, Chalvet-Monfray K, Chaimane T, Vongsawat K, Bicout DJ. Monthly 910 district level risk of dengue occurrences in Sakon Nakhon Province, Thailand. Sci 911 Total Environ. 2010;408: 5521-5528. doi:10.1016/J.SCITOTENV.2010.08.024 912 58. Martínez-Bello D, López-Quílez A, Prieto AT. Spatiotemporal modeling of 913 relative risk of dengue disease in Colombia. Stoch Environ Res Risk Assess. 914 2018;32: 1587–1601. doi:10.1007/s00477-017-1461-5 915 59. Mordecai EA, Caldwell JM, Grossman MK, Lippi CA, Johnson LR, Neira M, et 916 al. Thermal biology of mosquito-borne disease. Byers J (Jeb), editor. Ecol Lett. 917 2019; ele.13335. doi:10.1111/ele.13335 918 60. Carrington LB, Armijos MV, Lambrechts L, Barker CM, Scott TW. Effects of 919 Fluctuating Daily Temperatures at Critical Thermal Extremes on Aedes aegypti 920 Life-History Traits. PLoS One. 2013;8. doi:10.1371/journal.pone.0058824 921 61. Ngugi HN, Mutuku FM, Ndenga BA, Musunzaji PS, Mbakaya JO, Aswani P, et al. 922 Characterization and productivity profiles of Aedes aegypti (L.) breeding habitats 923 across rural and urban landscapes in western and coastal Kenya. Parasit Vectors. 924 2017;10: 331. doi:10.1186/s13071-017-2271-9 925 62. Lowe R, Gasparrini A, Van Meerbeeck CJ, Lippi CA, Mahon R, Trotman AR, et

al. Nonlinear and delayed impacts of climate on dengue risk in Barbados: A

926

927 modelling study. PLoS Med. 2018;15. doi:10.1371/journal.pmed.1002613 928 63. Li C, Wang X, Wu X, Liu J, Ji D, Du J. Modeling and projection of dengue fever 929 cases in Guangzhou based on variation of weather factors. Sci Total Environ. 930 2017;605–606: 867–873. doi:10.1016/j.scitotenv.2017.06.181 931 64. Li CF, Lim TW, Han LL, Fang R. Rainfall, abundance of Aedes aegypti and 932 dengue infection in Selangor, Malaysia. Southeast Asian J Trop Med Public 933 Health. 1985;16: 560–8. Available: http://www.ncbi.nlm.nih.gov/pubmed/3835698 934 65. Johansson MA, Dominici F, Glass GE. Local and Global Effects of Climate on 935 Dengue Transmission in Puerto Rico. Massad E, editor. PLoS Negl Trop Dis. 936 2009;3: e382. doi:10.1371/journal.pntd.0000382 937 Kenneson A, Beltrán-Ayala E, Borbor-Cordova MJ, Polhemus ME, Ryan SJ, Endy 66. 938 TP, et al. Social-ecological factors and preventive actions decrease the risk of 939 dengue infection at the household-level: Results from a prospective dengue 940 surveillance study in Machala, Ecuador. Messer WB, editor. PLoS Negl Trop Dis. 941 2017;11: e0006150. doi:10.1371/journal.pntd.0006150 942 67. Reich NG, Shrestha S, King AA, Rohani P, Lessler J, Kalayanarooj S, et al. 943 Interactions between serotypes of dengue highlight epidemiological impact of 944 cross-immunity. J R Soc Interface. 2013;10: 20130414. 945 doi:10.1098/rsif.2013.0414 946 68. Wen J, Elong Ngono A, Regla-Nava JA, Kim K, Gorman MJ, Diamond MS, et al. 947 Dengue virus-reactive CD8+ T cells mediate cross-protection against subsequent Zika virus challenge. Nat Commun. 2017;8: 1459. doi:10.1038/s41467-017-948 949 01669-z

- 950 69. Rodriguez-Barraquer I, Salje H, Cummings DA. Opportunities for improved
- surveillance and control of dengue from age-specific case data. Elife. 2019;8.
- 952 doi:10.7554/eLife.45474
- 953 70. Stoddard ST, Forshey BM, Morrison AC, Paz-Soldan VA, Vazquez-Prokopec
- GM, Astete H, et al. House-to-house human movement drives dengue virus
- 955 transmission. Proc Natl Acad Sci U S A. 2013;110: 994–999.
- 956 doi:10.1073/pnas.1213349110
- 957 71. Wesolowski A, Qureshi T, Boni MF, Sundsøy PR, Johansson MA, Rasheed SB, et
- al. Impact of human mobility on the emergence of dengue epidemics in Pakistan.
- 959 Proc Natl Acad Sci. 2015;112: 11887–11892. doi:10.1073/pnas.1504964112
- 960 72. Vaidya A, Bravo-Salgado AD, Mikler AR. Modeling climate-dependent
- population dynamics of mosquitoes to guide public health policies. Proc 5th ACM
- Conf Bioinformatics, Comput Biol Heal Informatics BCB '14. 2014; 380–389.
- 963 doi:10.1145/2649387.2649415
- 964 73. Schmidt CA, Comeau G, Monaghan AJ, Williamson DJ, Ernst KC. Effects of
- desiccation stress on adult female longevity in Aedes aegypti and Ae. albopictus
- 966 (Diptera: Culicidae): results of a systematic review and pooled survival analysis.
- 967 Parasit Vectors. 2018;11: 267. doi:10.1186/s13071-018-2808-6
- 968 74. Vazquez-Prokopec GM, Galvin WA, Kelly R, Kitron U. A New, Cost-Effective,
- 969 Battery-Powered Aspirator for Adult Mosquito Collections. J Med Entomol.
- 970 2009;46: 1256–1259. doi:10.1603/033.046.0602
- 971 75. Waggoner JJ, Gresh L, Mohamed-Hadley A, Ballesteros G, Davila MJV, Tellez Y,
- et al. Single-Reaction Multiplex Reverse Transcription PCR for Detection of Zika,

973 Chikungunya, and Dengue Viruses. Emerg Infect Dis. 2016;22: 1295–7. 974 doi:10.3201/eid2207.160326 975 Lanciotti RS, Calisher CH, Gubler DJ, Chang GJ, Vorndam A V. Rapid detection 76. 976 and typing of dengue viruses from clinical samples by using reverse transcriptase-977 polymerase chain reaction. J Clin Microbiol. 1992;30: 545–51. Available: 978 http://www.ncbi.nlm.nih.gov/pubmed/1372617 979 77. Grossi-Soyster EN, Cook EAJ, de Glanville WA, Thomas LF, Krystosik AR, Lee 980 J, et al. Serological and spatial analysis of alphavirus and flavivirus prevalence and 981 risk factors in a rural community in western Kenya. Bingham A, editor. PLoS Negl 982 Trop Dis. 2017;11: e0005998. doi:10.1371/journal.pntd.0005998 983 78. Palamara GM, Childs DZ, Clements CF, Petchey OL, Plebani M, Smith MJ. 984 Inferring the temperature dependence of population parameters: The effects of 985 experimental design and inference algorithm. Ecol Evol. 2014;4: 4736–4750. 986 doi:10.1002/ece3.1309 987 79. Team RC. R: A Language and Environment for Statistical Computing. R Found 988 Stat Comput. 2018; Available: https://www.r-project.org 989