Drivers of vascular plant species area relationships in British broadleaved woodlands and their

effects on the species area curve

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Summary

1. The loss of plant biodiversity in Great Britain is a major concern, with a fifth of species endangered

or vulnerable according to the latest IUCN Red List. The Government's 25 Year Plan for the

environment [Defra, 2018] aims to halt this loss and build new habitats, including new woodlands. To

ensure that biodiversity loss is halted in existing woodlands and gain is maximised in new ones, we

need to better understand which drivers have been most influential in controlling biodiversity. Here

we focus on vascular plant species' richness.

2. Previous attempts to explain plant species richness have mainly focussed on alpha diversity in a

consistent, fixed unit area. Here, we additionally undertake a novel analysis of the effects of

environmental heterogeneity and abiotic factors on species-area relationships derived from 16

randomly placed quadrats in each of 103 semi-natural, broad-leaved woodlands across Britain.

Species-area relationships were examined at two scales ($4m^2$ to $200m^2$ and $200m^2$ to $3200m^2$) to

explore the relationship between the drivers of species richness and the exponent z, of the canonical

species-area curve, $S = cA^z$. We also explore the use of a new metric ζ_r, based on zeta diversity to

quantify heterogeneity.

3. Habitat heterogeneity increased species richness, as did the exposure of the woodlands to

surrounding natural habitats. Higher levels of soil organic matter and the progression of woodlands

to later successional stages, decreased species richness. Richness was also seen to have a unimodal

response to soil acidity with a peak around pH6. At the smaller scale, heterogeneity elements in the

woodland such as riparian zones or coppicing led to an increase in the value of the exponent of the

species area curve. At the larger scale, species turnover led to an increase in the exponent of the curves

while succession led to a decrease. At both scales, soil organic matter content had a negative effect. ζ_r

was found to be a significant and important variable and to affect both species richness and the slope

of the species accumulation curves at larger scales.

Synthesis. Habitat heterogeneity measures included the presence of coppicing, open areas such as

rides and riparian zones and the difference between species assemblages in different plots in the

woodland. Results suggest that to maximize vascular plant diversity, woodlands should be managed

for heterogeneity. In addition, the increase in richness with exposure to surrounding natural habitats

suggests that woodlands should be large and connected. This implies that if possible current

woodlands should be increased in size, rather than creating new smaller disconnected sites.

Key words: biodiversity, species area curves, species richness, woodlands, zeta diversity.

Introduction

Biodiversity worldwide continues to decline [Butchart et al., 2010, Tittensor et al., 2014] and Britain is

no exception to this trend [Hayhow et al., 2016]. The IUCN Red List for the UK shows 22% of plant taxa

as critically endangered, endangered or vulnerable [Stroh et al., 2014]. Biodiversity is important for

ecosystem function [Gaston & Spicer, 2004], with woodlands performing specific additional functions

including the improvement of water quality, carbon sequestration [Slee and Kyle, n.d], nectar provision

within farmed landscapes [Baude et al 2016] and the provision of habitats for other taxa [Amar et al.,

2006, Fuller & Warren, 1993].

The aims of the new 25-Year Environment Plan [Defra, 2018] include achieving "thriving plants and

wildlife", planting 180,000 hectares of new woodland and ensuring that existing woodlands are better

managed. However, more woodland is not necessarily the answer to biodiversity loss if we cannot

ascertain why our current woodlands are not thriving.

This study was aimed at finding the main drivers of vascular plant species richness in British broad-

leaved semi-natural woodlands to provide research-based evidence on how best to manage, maintain

and increase their plant biodiversity. This research is the only work of its kind that analyses diversity

as the slope of the species area relationship expressed across multiple samples at the national scale.

SUMMARY OF FACTORS THAT HAVE BEEN SHOWN TO AFFECT SPECIES RICHNESS

Table 1 summarises variables that have been shown in the literature to affect plant species richness

and which can also be obtained from the CEH Woodland Survey database [Kirby et al., 1971]. This is

not intended as a comprehensive review and only represents a small selection of the rich literature in

this area. Indeed, the papers cited discuss results for multiple factors affecting species richness. The

reason for this summary is to justify the selection of predictor variables by appealing to prior

knowledge. This is important due to the model averaging procedure used as part of this analysis which

is described in more detail below.

Table 1. Examples of abiotic factors which have been shown by previous authors to affect vascular plant species richness.

Factor	Relationship to species richness	Comments	Reference
pH	positive linear	Studies of beech and hornbeam woodlands report lower species richness with lower pH and increased SOM. This was true for both taxa, suggesting that the effect was not solely the result of the physical barrier created by slowly degrading beech litter.	Koojiman & Cammeraat, [2010]
Soil Organic Matter (SOM)	negative linear		
pH	Unimodal	Greatest richness in woodlands occurred on nutrient rich soils, with a peak around Ellenberg N of 7 for shade loving species.	Cornwell & Grubb, [2003]
Latitude	11% per degree latitude	Authors pointed to altitude as more important in this study on tree species richness. Understory richness was not affected, and this effect may not be evident given the	Ohlemuuler & Wilson, [2000]

		modest change in latitude in our data.	
Isolation	Connected habitat contained 13 – 18% more plant species.	Species were averaged across dispersal mechanism.	Brudvig et al., [2009]
Isolation	Negative trend with increasing isolation	Authors found that plant species richness in agricultural, fragmented landscapes increased with connectivity of landscape elements.	Thiele et al., [2018]
Isolation	Woodland patch area and isolation explained 20% of variation in observed species richness in lowland woodlands	In this study connectivity with wooded linear features indicated lower isolation.	Petit et al., [2004]
Isolation	Correlation coefficient -0.33 to -0.45	Distance to nearest woodlands used as isolation factor and was shown to be negatively correlated with woodland and shrub species richness	Dzwonko & Loster [1988]
Heterogeneity	Plant species richness up to 20% lower in unmanaged forests.	Presence of logging trails used as sign of disturbance in managed forests,	Boch et al., 2013
Heterogeneity	60% change in mean species richness in sites which experienced storm damage	In this work wind throw as a result of the 1987 hurricane generated heterogeneity	Smart et al., 2014

Materials and Methods

HABITAT HETEROGENEITY – THE USE OF ZETA DIVERSITY

Habitat heterogeneity is known to be important for biodiversity [Gardener, 2010], but it is not trivial

to quantify and has been expressed in many ways. Honnay et al., [1999] used length of rides, number

of soil types and a ratio of perimeter and area. Schmidt [2005] used the presence of logging trails, and

Bàldi [2008] used the number of CORINE land cover codes.   In this work several methods were used

to quantify heterogeneity as shown in table 2. However, these methods have some weaknesses. PHI

(Positive Heterogeneity Index, see table 2 for more details) requires additional work to collect the

information and does not indicate the concentration of elements. For example, if a woodland shows

considerable signs of woodland management, only one code is entered, and the PHI increases by 1 for

woodlands with both a large or a small amount of management. In addition, the number of elements

included depend on the additional area of woodland that surveyors cover. A new metric was therefore

generated based on zeta diversity [Hui & McGeoch, 2014] to provide a simple, objective measure.

Zeta diversity is the average number of species shared between multiple plots, [Hui & McGeoch

2014] and can be used to show compositional change in species assemblages, [McGeogh et al., 2017]

and model drivers of species turnover, [Latombe et al., 2017].

ζ_1 is the average number of species in each plot ζ_2, is the average number of species shared between

every combination of two plots. If plots contain the same species, then $\zeta_1 - \zeta_2 = 0$. If all plots contain

different species then $\zeta_1 - \zeta_2 = \zeta_1$.

A homogeneous woodland is more likely to contain species that occur in many plots, whereas a

heterogeneous woodland is more likely to have plots containing different species. This suggests a

method of comparing heterogeneity between woodlands using the ratio,

$\zeta_r = \frac{\zeta_1 - \zeta_2}{\zeta_1}$,

The more homogeneous the woodland, the closer the value of ζ_r to zero. There is sometimes

concern that the use of a diversity metric to predict richness introduces a circularity. This is not the

case when using the ratio ζ_r. ζ_r can take values between 0 and 1. The value depends on the difference

between average numbers of species in each plot to the average number shared between two plots

and not on the richness of the plots. For example, in homogeneous species rich or species poor

woodlands the values of ζ_1 and ζ_2 will be similar, and therefore ζ_r will approach 0. The converse

occurs for a heterogeneous woodland. The ratio ζ_r is not, therefore, a predictor of richness, it can

take values between 0 and 1 for both species rich and species poor woodlands, it is a predictor of

species turnover.

ζ_r can be seen to be like Jaccard's dissimilarity index, with the extension that here the use of ζ

diversity calculations allows us to easily use multiple plots.

The zeta values for each woodland were calculated using the zetadiv function in R, [Latombe et al.,

2018]. The ζ_r coefficient was calculated and added to the set of predictor variables.

DRIVERS OF VASCULAR PLANT SPECIES RICHNESS ACROSS ENTIRE WOODLANDS AND AT TWO

DIFFERENT SCALES.

The drivers of vascular plant species richness were studied in two ways. Firstly, by examining the effect

of a range of variables on the species richness of a fixed area within 103 different woodlands and,

secondly, by looking at the scale dependence of species richness as expressed in the canonical species

area curve (SAC), $S = cA^z$, and the effect of habitat heterogeneity and environmental conditions on the

exponent, z of these curves.

Species accumulated across
the plots, from 200m² to
3200m²

Species area curves
across the nests, from
4m² to 200m²

Figure 1. Two scales of species area dependence. At the nest scale species area curves were calculated for areas from 4m²to 200m². At the plot scales species were accumulated across the 16 plots such that area increased from 200m² to 3200m².

The smallest scale used species accumulated across a concentrically nested set of square quadrats.

(This z value will be referred to as nest z). These are typically referred to as species area curves and z

values of around 0.25 were first proposed by Preston [1960] as being the result of sampling from log-

normal species distributions. Other authors have suggested that the z value at this scale has no

biological meaning [Connor et al, 1983, Tjorve & Tjorve, 2017] but is a result of species distribution.

Using neutral models Cencini et al [2012] predicted that population size affects the shape of the

curve while Sugihara [1979] concludes that the value of 0.25 reflects community structure.

At the larger scale species were accumulated across all the non-contiguous nested quadrats in the site.

(This z value will be referred to as plot z). These curves are referred to as species accumulation curves

and at this scale habitat heterogeneity is likely to play a part in the z value [Storch et al, 2014]. In similar

work, Shen et al [2009] considered the effect of tree density, soil nutrients, pH, slope, elevation and

aspect and heterogeneity and concluded that dispersal and habitat heterogeneity jointly are required

to explain the shape of species area curves. Baldi & Sadler [2017] concluded that habitat heterogeneity,

as quantified by the number of habitat types as shown through CORINE land cover maps, was more

important than area for expressing species richness.

The combined influence of local abiotic factors and woodland heterogeneity on the exponent of curves

constructed over these two different scales, was examined.

DATA

The data were taken from the Centre for Ecology and Hydrology's woodland survey database [Smart

et al., 2013]. One hundred and three woodlands across Great Britain were surveyed between 2000 and

2003. Sixteen plots were randomly located in each woodland, each plot had a total area of 200m^2.

Further, each plot was itself nested with sub-areas of 4m^2, 25m^2, 50m^2, 100m^2 and 200m^2 (see figure

1).

All flora was recorded within each plot. Additional plot data were collected, including diameter at

breast height (DBH) of trees, live basal area (LBA) of trees and shrubs, soil pH, soil organic matter

(SOM) content, National Vegetation Classification (NVC) code, and major soil group (MSG). Additional

woodland information was also recorded, such as whether there were signs of woodland

management, the presence of riparian zones and open areas such as glades or rides. Table 2

summarises the information taken from the dataset and details how it was transformed to create

predictor variables.

The richness for each woodland was calculated to be the unique number of species in the total 16

plots surveyed. The variables within the database were at different scales, plot level and woodland

level. Plot level variables, such as pH, DBH or SOM, were scaled appropriately to the richness by

taking the mean per woodland. Site level variables were buffer, number of NVC codes, number of

major soil groups, area ratio and PHI. The variables fall into two categories; those which represent

heterogeneity and those which describe average site conditions. The bold code H or S in column 2

of table 2 denotes the former and latter respectively.

Table 2 Summary of predictors created from the variables in the dataset. Column 2 denotes either a site level variable, S, where means of plot level values have been taken, or a variable used to represent heterogeneity, H.

Variable	Variable type	Description
Positive Heterogeneity Index (PHI)	H	Codes were recorded for each woodland to show the presence of other elements such as ponds, ditches, buildings, dead wood etc. The sum of codes that corresponded to an element that has been shown by previous authors to be positively correlated with vascular plant species richness. This sum was used as an explanatory variable - PHI. The greater the PHI, the more elements within the woodland that could increase richness by increasing heterogeneity of conditions.
Northing	S	Latitude
Buffer	H	The proportion of land around the wood (to 3500m from the edge) that is not arable, improved grassland or urban. The larger the buffer, the more the surrounding area can be considered as positive for species richness. This variable therefore equates to the inverse of isolation and measures the favourability of the matrix within which each woodland was embedded.
Number of major soil groups (MSG)	S	The number of major soil group codes in each woodland.
Number of National Vegetation Classification (NVC) codes	H	The number of different NVC codes in each woodland. The NVC code represents the assignment of the plant species assemblage to a phyto-sociological classification of British plant communities. The number of different NVC codes therefore describes the number of different plant assemblages present. This is equivalent to the number of CORINE land cover codes used by Baldi [2008]. Although the NVC code might correlate with richness, the number of codes will not, since it is possible that each code could represent a low richness assemblage. However, since NVC codes represent a different species assemblage with different abiotic factors, if there are more codes, it is more likely that the woodland has greater heterogeneity.
pH	S	The pH for the plot
Soil Organic Matter (SOM) content	H	The SOM content for a plot
Live Basal Area (LBA)	S	The LBA of all saplings, trees and shrubs recorded in a plot
Diameter at breast height (DBH)	S	The DBH of all trees recorded in each plot
Tree Density (TD)	S	Count of trees per plot
Area ratio	H	The ratio of area to perimeter of the site. This is a measure of the number of habitats the woodland is exposed to [Honnay et al, 1999].
ζ_r	H	The ratio of ζ_1-ζ_2 to ζ_1, quantifying species turnover between plots.

MODELLING

The modelling was designed to answer three questions as follows:

1) Which variables had most influence on species richness of the entire woodland?

2) Which variables had most influence on the exponent of the species area curve and did this change

with scale?

3) Is the zeta diversity metric ζ_r useful as a measure of heterogeneity?

To answer the first question, model averaging and a gradient boosting machine (GBM) were used to

examine which of the predictor variables had the greatest effect on the species richness of the same

area, 3200m^2, in each of the 103 woodlands. To explore the second, the response of the exponent, z,

of the species area curves to the same set of variables was explored at two scales; 4m^2-200m^2 (nest

level) and 200 – 3200m^2 (plot level) using model averaging. Finally, significance and importance of ζ_r

was observed in the above models as compared to the alternative heterogeneity metrics employed.

MODEL AVERAGING

Model averaging is an ideal method for isolating variables which are most important for predicting a

response when the relationships are linear, [Kimberley et al, 2014; Symonds & Moussalli, 2010]. The

Akaike Information Criteria (AIC) is used to rank a set of competing models, [Burnham, Anderson, &

Huyvaert, 2010]. A subset of these models with the lowest Akaike weights are selected, and

parameters averaged across this set.

Since highly correlated predictor variables are undesirable [Freckleton, 2010] the correlation

coefficients of all variables were examined, and it was found that all variables had Spearman and

Pearson correlation coefficients below 0.54. The variance inflation factors of the models were also

examined and were all below 2.

Once the predictor variables were selected the dataset was standardized as suggested by Gelman,

[2008]. This gives parameter effect sizes that can be compared directly (two standard deviations of

change in the predictor variable will result in the effect size change in the response). A linear model

was created using all the variables. The dredge function in the MuMin package in R [Barton, 2018] was

used to create a set of models containing all permutations of the variable set. These were ranked

according to the AIC. The model with the lowest AIC is considered the model that best represents the

data. This method will always produce a best model, but that model is only "best" relative to all the

other models generated. It is therefore important to ascertain the goodness of fit of the initial linear

model used [Symonds & Moussalli, 2010]. The goodness of fit of the models using all the parameters

was ascertained by examining the fitted values against the residuals and by considering the value of

R^2. None of the models showed any pattern in the residuals. For the species richness models R^2 was

0.47. For nest z, 0.33 and for plot z, 0.47.

All models within an AIC of between two to seven can be considered as equally good [Burnham et al.,

2010] and used to create a model set. The parameter effect sizes are then averaged across this model

set and a 95% confidence interval obtained. If this confidence interval does not cross zero, these

predictors are considered significant, [Kimberley et al., 2014]. In this work, models within AIC of 2 (δ <

2) of the highest ranked model were initially selected. The effect of increasing δ was then examined. If

more variables were introduced which had confidence intervals that crossed zero, the lower value was

used to avoid including redundant models [Grueber et al., 2011]. The relative importance of each effect

size was also obtained, [Burnham, 2015]. Relative importance is a measure of the probability that a

variable is present in the model set. A relative importance of 1 means that a variable appears in every

model set.

GRADIENT BOOSTING MACHINES

Gradient Boosting Machines (GBMs) are ensemble decision tree algorithms where recursive binary splits divide the data into regions, such that the root mean square error (rmse) of the response variable is minimized, [James et al., 2013. In addition, GBMs weight the observations such that those which were difficult to classify after a split are given increased weight. The next tree is then generated using these weighted observations. GBMs are suited to analysing complex ecological datasets because they can fit non-linear relationships and are not sensitive to scale, [De'Ath & Fabricius, 2000; Elith et al., 2008]. They have been shown to be successful, for example, in finding environmental variables which explain species richness of vertebrate groups [Mouchet et al., 2015]. In this analysis the gbm package in R was used, [Ridgeway, 2017].

The data was split into train and test sets using a 75/25 ratio. The GBM also uses a random subset of training data and so automatically provides a validation set. The values that are not selected can be used to give an "out of bag" (OOB) prediction error for the model.

Hyper parameters define the structure of the GBM and must be chosen to optimise the model performance. This was achieved through a hyper parameter tuning grid; a matrix containing a set of permutations of hyper parameters. The model was run across the hyper parameter tuning grid and the hyper parameters which minimized the OOB error were used in the final model.

Over fitting was addressed using the ntrees function of the gbm library. This function looks at the prediction error on the OOB set, and the number of trees is set as the value when this error stops decreasing and starts to increase. The rmse error of the train was half that of the test set, suggesting some overfitting. However, the test set rmse was comparable to that obtained using model-averaging.

To extract the variables which most influence species richness the relative influence (RI) of each variable was used, [Natekin & Knoll, 2013]. The percentage decrease in rmse is averaged for each variable over each time a variable was chosen to make a binary split in the data. The variable which has greatest RI is best at splitting the data and predicting the response.

Decision tree methods such as GBMs sometimes do not give stable results for RI, [Nicodemus et al.,

2007]. Therefore, each model was repeated 100 times and the variables ranked by the average values

of RI. The value of RI is relative and therefore subjective. In this work variables which had RI above 10%

were selected as important for species richness.

The benefit of using these two modelling methods is their ability to detect different responses. Model-

averaging detects linear relationship whereas the GBM can detect more complex patterns in the data,

[James et al, 2013].

SCALE DEPENDENCE OF SPECIES RICHNESS

This part of the work involved calculating species area curves across the nests from $4m^2$ to $200m^2$.

These correspond to type I nested curves as described by Scheiner [2003]. At the larger scale species

were accumulated across the plots from $200m^2$ to $3200m^2$, as shown in figure 1.

At the larger scale, species can be accumulated in 16! alternative ways. The average of a subset of

these accumulation curves is often used to calculate the slope of the species area curve. However,

there are potential problems with this method when the area over which the accumulation occurs is

heterogeneous, [Bacaro et al, 2012; Bevilacqua et al, 2018, Chiarucci et al, 2009]. This method is

effectively selecting all species for accumulation as if they have equal probabilities of occurring in any

plot in the woodland, i.e. the woodland is homogeneous. This is not the case for this data, the species

are grouped into assemblages. Therefore, we took a different approach to maintain this spatial

information. First, species were accumulated in such a way as to give a minimum gradient to the

species accumulation curve. This is equivalent to grouping all similar species assemblages together.

Secondly, species were accumulated to give a maximum gradient. This is equivalent to arranging the

plots in the most heterogeneous manner. Since these two cases represent two extremes in the

woodland (the most homogenous and the most heterogeneous) an average was taken of the two

resulting curves. This spatially explicit method is similar to a type III curve, as described by Scheiner

257 [2003], where mean diversities of adjacent plots are calculated, but here adjacency is in terms of

258 species assemblage's similarity or dissimilarity. In order to check the fit of this method the R^2 value of

259 the log/log transform was calculated and found to be greater than 0.94 for three quarters of the

260 woodlands. At both scales the gradients of the log/log transform of the curves were used as z (see

261 supplementary material for more information). The z values at the two scales were then modelled

262 with the same variables used to model species richness, as listed in table 2.

Results

SPECIES RICHNESS OF FIXED AREA OF 3200M^2

Figure 2 shows the model-averaged effect sizes and 95% confidence intervals for the total richness of

all the 16 200m^2 plots in each woodland. Buffer, number of NVC codes, PHI and ζ_r have a positive effect

on species richness with similar effect sizes. Mean diameter at breast height and mean SOM have a

negative effect, again, with similar effect size. All significant variables are included in every model in

the model set except ζ_r which shows an 80% probability of inclusion. The effect sizes of all significant

variables are similar, with no one parameter clearly having a greater influence on richness than any

other.

Figure 2. Model averaged effect sizes for woodland richness modelled with all variables using δ
< 2 for the model set. Variable importance is in red. The points show the average effect size
and the bars show the 95% confidence intervals.

274 Table 3 shows the parameters with relative influence of 10 or above in the GBM. ζ_r and PHI are again

275 selected as important for predicting richness. The GBM also selects mean LBA, mean pH and Northing.

Table 3. Most influential variables from the gradient boosted machine

Parameter	Relative influence
ζ	16.37
Northing	13.25
meanLBA	11.42
PHI	10.88
mean pH	9.92

SCALE DEPENDENCE OF SPECIES RICHNESS

281 Figure 3 shows the distributions of nest z and plot z at the two scales. At the smaller nest scale the

282 exponents are distributed around 0.25, whilst at the larger scale they are distributed around 0.5.

Figure 3. Distribution of z values at two scales. The pink distribution shows nest scale –
$4m^2$ to $200m^2$. The blue distribution shows plot scale – $200m^2$ to $3200m^2$

Figure 4 shows the model-average effect sizes for nest z. The heterogeneity parameters PHI and ζ_r have

a positive effect on the value of z while mean SOM has a negative effect. The mean SOM is contained

in every model in the model set, but the probability of inclusion of PHI is 82% and for ζ_r only 57%, this

implies that ζ_r is not an important parameter at nest scale. PHI is the most significant parameter, the

effect size double that of mean SOM.

Figure 4. Model averaged effect sizes for nest z modelled with all variables using δ < 1.5 for the model set.

Figure 5 shows the model-average effect sizes for plot z. ζ_r is highly significant and has a positive effect

on plot z. At this larger scale the effect of ζ_r, which quantifies species turnover across the woodland,

has much greater significance than at the smaller nest scale. Mean tree density and mean SOM are

also significant, although less so than ζ_r and their effect sizes are less than a third of ζ_r. Both mean SOM

and mean tree density have a negative effect on plot z.

Figure 5. Model averaged effect sizes for plot z modelled with all the variables using δ < 1.5 for the model set. Variable importance is in red.

Discussion

VARIABLES AFFECTING SPECIES RICHNESS

HETEROGENEITY

Model-averaging and the GBM showed nine variables as affecting richness of the woodlands. Three of

these represent heterogeneity: ζr, number of NVC codes and PHI. Of these, number of NVC codes and

PHI are present in every model in the model set and ζr is present in 80% of the models. The GBM selects

ζ_r as the most influential variable.

The greater PHI, the more evidence there is of woodland management, open areas and riparian zones.

This implies that all these features are required to maximise richness. The number of NVC codes and

ζr quantify the number of different species assemblages. Different species assemblages are more likely

to occur if environmental conditions differ across the woodland – for example, different levels of soil

moisture or different light levels, again stressing the importance of heterogeneity within the

woodlands.

MEAN-DBH AND MEAN-LBA

Model-averaging shows mean DBH to have a negative effect on richness and the GBM selected mean

LBA as influential. This is to be expected, as an increase in either variable implies succession in the

woodland [Packham et al., 2001; Smart et al., 2014] which leads to increased homogeneity because of

a strengthening of the shading filter on the understorey species pool [Keith et al., 2009], again

indicating the importance of woodland management schemes, such as coppicing, to vary the

successional status of the woodland.

BUFFER

Buffer size had a significant positive effect on richness and occurred in every model in the model set.

The size of the buffer relates to the amount of the woodland that is exposed to natural habitats and

therefore has two potential effects. Firstly, the larger the buffer the more connected the woodland

(habitat fragmentation is known to have a negative impact on biodiversity, [Quine, 2011, Watts et al

2008]). Secondly, the larger the buffer, the more protected the woodland from agrochemicals and

airborne environmental pollutants; particularly around the edges [Draaijers, et al., 1988], which are

linked to reduced species richness [Bobbink et al., 1998; Pallet et al., 2016, Simkin et al., 2016; Stevens

et al., 2016]. Previous research has suggested that buffer regions should be included around

woodlands for various reasons, including protection from anthropogenic influences [McWilliam et al.,

2010] and vehicle pollution [Bignal et al., 2008]. It is not possible within this dataset to ascertain the

mechanism for the increase in richness with buffer, but this work agrees with previous authors; larger

more connected woodlands tend to have greater plant species richness.

NORTHING

Northing is selected as an influential variable by the GBM. This is in contradiction to previous authors,

[Gilman et al., 2014; Ohlemuller & Wilson, 2000]. A bivariate plot of Northing with mean TD (figure 6)

shows a negative correlation, suggesting that the increased richness could be due to the addition of

light-demanding species.

The Scottish woodlands might also consist of plant communities which tend to be richer. For example,

if Scottish woodlands were predominantly W6 eutrophic Alder woodlands, they would be richer than

W14 beech woods which are common in the South. However, figure 7 shows that this is not the case.

The distribution of NVC communities is similar in the North and South. What is noticeable, however,

is that for the same community, richness tends to be greater in Scotland (figure 8). This suggests that

other variables, not availabe in this dataset, may be responsible for the increased richness in Scottish

woodlands, or that a combination of positive predictors is more likely to co-occur in the more Northern

woodlands.

Figure 6. Scatter plot of northing with mean tree density suggests that the northern woodlands are more open. This could allow the inclusion of more light-loving species assemblages and hence add to species richness.

Figure 7. Distribution of NVC codes in Scotland and England and Wales. In both areas 30% of communities are W10, Quercus robur woodlands. These communities have one of the widest range of species richnesses of all NVC communities in this dataset.

Figure 8. Density of plot richness for the most common NVC codes in the dataset in Scotland and England and Wales. Given the same NVC community, richness tends to be greater in Scotland for these common communities. Although OV27 is not a woodland community, it was was common in this dataset, demonstrating open areas within the wood.

SOIL ORGANIC MATTER

SOM content was shown to have a significant and important negative effect on species richness in

agreement with Koojiman & Cammeraat [2010]. Organic soils have low pH and low pH soils in

woodlands results in low species richness, [Cornwell & Grubb, 2003]. However, in this data, very few

of the plots were located on organic soils. The six NVC codes shown in table 3 represent 82% of all

plots surveyed and give an indication of the majority of soil types.

Table 3. Soil types for the most common NVC codes in this data

NVC code	% of plots	Soil type
W6 *Alnus glutinosa-Urtica diocica*	5%	Eutrophic moist soil alluvial or on flood fen peat
W8 *Fraxinus excelsior – Acer campestre – Mercurialis perennis*	9.5%	Dry calcareous soils
W21 *Crataegus monogyna – Hedera helix* scrub	10%	Various, base rich
W16 *Quercus* spp. – *Betula* spp. – *Deschampsia flexuosa*	11.5%	Very acidic, oligotrophic, pH<4, sandy podzolic
OV27 *Epilobium angustifolium* Open habitat	14%	Various, fertile, damp
W10 *Quercus robur – Pteridium aquilinum – Rubus fruticosus*	32%	Neutral brown earths

Of these, only W6, 5% of the total, are potentially on peat, but W6 is a high nutrient, high richness community. It is therefore unlikely that the reduced richness with SOM is caused by reduced richness expected on organic soils.

The reduced richness could be due to low pH on other soil types since increased SOM has been shown to decrease pH [Russell, 1960; Williams & McDonald, 1960]. On the other hand, adding SOM can also increase pH, depending on the initial acidity of the soil and the nature of the litter [Ritchie & Dolling, 1985]. In this analysis, increased SOM appears to correlate with either decreased or no change in pH (figure 9).

Figure 10 shows that all the common NVC communities tend to show reduced richness as SOM increases, regardless of soil type. Given also that the NVC code represents a community with similar physical conditions, such as nutrient levels, soil pH and light levels, and while the response to richness with increased SOM is seen across all communities, this finding implies that the variation in richness

374   with SOM is unlikely to be due solely to one of those physical conditions, but perhaps to another factor

375   not measured in this dataset.

Figure 9. Change in plot pH with SOM content. For W10, W16 and W21 plots the pH reduces with increased SOM but this effect is not clear for other plot types

Figure 10. Plot richness with SOM for the six most common NVC communities in this data. The blue boxplots show the range in richness when grouped, the red dots are the raw ungrouped data. The variation in richness is high when SOM is low and vice versa.

Increased nitrogen deposition can both increase SOM [Fang et al., 2014; Zak et al., 2016] and reduce richness [Bobbink et al., 1998; Pitcairn et al., 2002; Simkin et al., 2016; Stevens et al., 2016]. A smaller buffer could allow increased nitrogen deposition, [Draaijers et al., 1988; Kennedy & Pitman, 2004], which might be reflected in the Ellenberg N values [Maskell et al., 2010]. Therefore, if woodlands with smaller buffers showed both increased Ellenberg N values and increased SOM, nitrogen deposition might be the mechanism behind the change in SOM and the reduced richness. In support of this figure 11 shows that while buffer is negatively correlated with Ellenberg N values, but, in

388   contradiction, it is positively correlated with SOM content. The latter association may be confused by

389   the location of plots within the woodland.

Figure 11. Change in mean SOM and Ellenberg N values with Buffer size. If nitrogen deposition was leading to increased SOM, then woodlands with smaller buffers would have greater SOM and larger Ellenberg N. The graph on the right shows that smaller buffers correlates with higher Ellenberg N, but not to greater mean SOM.

In summary, whilst SOM is negatively correlated with species richness, the causal mechanism is not

evident. However, other authors, using modelled estimates of nitrogen deposition rather than

Ellenberg N values with this dataset have shown positive correlation between SOM and modelled

nitrogen deposition, [Kirby, 2005]. It is also interesting to note that nitrogen deposition values were

on average lower in Scotland [RoTaP, 2012], which may be another factor contributing to the increased

richness of the more Northern woodlands.

SOIL pH

The GBM selected mean pH as important for predicting species richness. Figure 12 demonstrates that

in this data plot richness does have a unimodal response to plot pH, in agreement with Cornwell &

Grubb, [2003]

Figure 12. Plot richness response to pH can be seen to be unimodal with a peak around pH6.

EFFECT OF DRIVERS OF SPECIES RICHNESS ON GRADIENT OF SPECIES AREA CURVE AT TWO SCALES.

The fitted nest *z* values were distributed around 0.25. This agrees with the value of z predicted by other

authors [Preston, 1962, Connor et al., 1983; Sugihara, 1976]. At the larger scale, plot z was found to

be distributed around 0.5. These two values correspond to those found by Crawley & Harral [2001] for

vascular plants in the county of Berkshire, South East England.

At smaller scales, PHI has a significant positive effect on nest z while SOM has a negative effect. This

model suggests that biological variables appear to be influencing the slope of the species area curves,

in contradiction to the remarks of other authors [Connor et al., 1983; Sugihara, 1976; Tjorve & Tjorve,

2017], but this is not necessarily the case. Both Sugihara [1979]   and Tjørve & Tjørve [2017] point to

the role of community structure and aggregation in shaping the species area curve. PHI, by introducing

elements into the woodland that increase plant species richness, is likely, on average, to be increasing

the richness of each plot. This will imply a more diverse community over which the curve is

constructed. Similarly, SOM can have a homogenizing effect on the plots due to loss of species richness,

either directly [Kooijmann & Cammeraat, 2010] or because increased SOM indicates increased

nitrogen deposition and eutrophication that favours the abundance of competitive species potentially

suppressing species diversity in the understorey, [Maskell et al, 2010, Pallet et al 2016]. Both PHI and

SOM can be seen therefore, as influencing community structure either positively or negatively and so

effecting nest z. It should also be noted that model-averaged R^2 for nest z was only 0.31, and that of

the full model 0.33 so neither of the variables explained substantial variance in nest z.

At larger scales ζ_r has a significant positive effect on plot z, and mean SOM and mean TD have a

significant negative effect. ζ_r is significant at the larger scale because it represents species turnover

across the woodlands. The greater the species turnover, the greater the expected slope. Similarly,

increasing mean TD and mean SOM are likely to have a homogenising effect on the woodland and

therefore, to reduce plot z. At the larger scale, it is species turnover and heterogeneity that has been

shown to influence the exponent of the species accumulation curves rather than biological factors. At

this scale the model was more successful at explaining the variance in plot z, having R^2 of 0.44 for the

model-averaged model and 0.47 for the full model.

VARIABLES OMITTED

The species richness model explained 47% of the variance in plant richness. Since we chose only those

variables that we both knew were likely to affect richness and that were available within the dataset,

several important potential variables were omitted which may improve the explanatory power of the

model. Woodland age and past land use have been shown to affect species composition [Hermy,

1994] and richness [Dzwonko & Loster, 1989; Peterken & Game 1984]. Ancient woodlands are more

likely to contain long-lived, poorly dispersing perennials whose migration to new woodland sites is

limited [Kimberley, Blackburn & Whyatt, 2015]. This constrains the potential richness of new

woodlands, particularly if they are isolated. Ancient cultivation has also been shown to induce a species

compositional gradient and thereby increase richness [Dupouey et al., 2002].

Plant available phosphorus has been shown to reduce species richness of native plants in alder forests

[Hrivnak et al., 2015]. Phosphorus eutrophication favours competitive species particularly at higher

light levels and this can result in the exclusion of other species [Keersmaeker et al., 2004].

The amount of light reaching the woodland floor influences richness, with greater richness at higher

light levels, [Kenedy & Pitman, 2004]. However, eutrophication together with high light levels

encourages shade-intolerant, competitive species which tend to reduce richness, [Cornwell & Grubb,

2003, Keersmaeker et al., 2004]. Conversely, low pH in conjunction with low light levels can result in

very little ground cover, [Cornwell & Grubb, 2003]

USE OF ζ_r AS HETEROGENEITY METRIC

Results presented here suggest that ζr is a useful way to quantify species turnover. This variable was

selected as the most influential by the GBM and was significant when used in model-averaging. The

fact that ζ_r was a significant parameter when modelling the slope of species accumulation curves

across the wood, but not when modelling the slope of species area curves across plots, demonstrates

that it quantifies species turnover. This metric could be easily generated from species lists as an

indicator to woodland managers of the number of different habitats within the woodlands.

References

Amar, Arjun & M Hewson, C & M Thewlis, R & Smith, Ken & Fuller, Robert & Lindsell, Jeremy &

Conway, Greg & Butler, Simon & A Macdonald, M. (2006). What's Happening to Our Woodland

Birds?

Bacaro, G. *et al.* (2012) 'The spatial domain matters: Spatially constrained species rarefaction in a

Free and Open Source environment', *Ecological Complexity*. doi: 10.1016/j.ecocom.2012.05.007.

Báldi, A. (2008) Habitat heterogeneity overrides the species–area relationship. *Journal of*

Biogeography, 35 (4), 675–681.

Barton, K., (2018). MuMIn: Multi-Model Inference. R package version 1.42.1. https://CRAN.R-

project.org/package=MuMIn

Baselga, A., Jimenez-Valverde, A., Niccolini, G. (2007) A multiple-site similarity measure independent

of richness. *Biology Letters*, 3 (6), 642–645.

Baselga, A. (2013) Multiple site dissimilarity quantifies compositional heterogeneity among several

sites, while average pairwise dissimilarity may be misleading. *Ecography*, 36 (2), 124–128.

Baude, MJ *et al* (2016) Historical nectar assessment reveals the fall and rise of floral resources in

Britain. *Nature* **530**, 85-88.

Bevilacqua, S. *et al.* (2018) 'An approach based on the total-species accumulation curve and higher

taxon richness to estimate realistic upper limits in regional species richness', *Ecology and Evolution*,

8(1), pp. 405–415. doi: 10.1002/ece3.3570.

Bignal, K.L., Ashmore, M.R., Headley, A.D. (2008) Effects of air pollution from road transport on

growth and physiology of six transplanted bryophyte species. *Environmental Pollution*, 156 (2), 332–

340.

Bobbink, R., Hornung, M. & Roelofs, J.G.M. (1998) The effects of air-borne nitrogen pollutants on

species diversity in natural and semi-natural European vegetation. *Journal of Ecology*, 86 (5), 717–

738.

Boch, S., Prati, D., Müller, J., Socher, S., et al. (2013) High plant species richness indicates

management-related disturbances rather than the conservation status of forests. *Basic and Applied*

Ecology, 14 (6), 496–505.

Brudvig, L.A., Damschen, E.I., Tewksbury, J.J., Haddad, N.M., et al. (2009) Landscape connectivity

promotes plant biodiversity spillover into non-target habitats. *Proceedings of the National Academy*

of Sciences, 106 (23), 9328–9332.

Burnham, K.P., Anderson, D.R., Model selection and multimodel inference: a practical information-

theoretic approach. New York: Springer; 2010

Burnham, K.P., Anderson, D.R., Huyvaert, K.P. (2010) AIC model selection and multimodel inference

in behavioral ecology: some background, observations, and comparisons. *Behavioral Ecology and*

Sociobiology, 65 (1), 23–35.

Burnham, K, P., (2015) Multimodel inference: understanding AIC relative variable importance values.

Available from: https://sites.warnercnr.colostate.edu/anderson/wp-

content/uploads/sites/26/2016/11/AICRelativeVariableImportanceWeights-Burnham.pdf. [Accessed

10th August 2018]

Butchart, S. H. M. *et al.* (2010) 'Global biodiversity: Indicators of recent declines', *Science*, 328(5982),

pp. 1164–1168. doi: 10.1126/science.1187512.

Cencini M, Pigolotti S, Muñoz MA. What ecological factors shape species-area curves in

neutral models? *PLoS One*. 2012;7(6):e38232. doi:10.1371/journal.pone.0038232

Chiarucci, A., Bacaro, G., Rocchini, D. & Fattorini, L. (2008) Discovering and rediscovering the

sample-based rarefaction formula in the ecological literature. *Community Ecology*, 9 (1), 121–123.

Chiarucci, A., Bacaro, G., Rocchini, D., Ricotta, C., Palmer, M.W., Scheiner, S.M., (2009), Spatially

constrained rarefaction: incorporating the autocorrelated structure of biological communities into

sample-based rarefaction, *Community Ecology*, 10(2), pp209 – 214

Connor, E. F., McCoy, E. D. and Cosby, B. J. (1983) 'Model discrimination and expected slope values in

species-area studies.', *American Naturalist*, 122(6), pp. 789–796. doi: 10.1086/284171.

Cornwell, W.K., Grubb, P.J. (2003) Regional and local patterns in plant species richness with respect

to resource availability. *Oikos*, 100 (3), 417–428.

Crawley, M.J. (2001) Scale Dependence in Plant Biodiversity. *Science*, 291 (5505), 864–868.

Cutler, D.R., Edwards, T.C., Beard, K.H., Cutler, A., et al. (2007) Random Forests For Classification In

Ecology. *Ecology*, 88 (11), 2783–2792.

De'ath, G., Fabricius, K.E. (2000) Classification and Regression Trees: A Powerful Yet Simple

Technique for Ecological Data Analysis. *Ecology*, 81 (11), 3178.

Defra, (2018), The 25 Year Environment Plan, Available from:

https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file

/693158/25-year-environment-plan.pdf, [Accessed 8th August 2018]

Draaijers, G., Ivens, W., Bleuten, W. (1988) Atmospheric deposition in forest edges measured by

monitoring canopy throughfall. *Water, Air, and Soil Pollution*, 42 (1-2).

Dupouey, J.L., Dambrine, E., Laffite, J.D., Moares, C. (2002) Irreversible Impact of past Land Use on

Forest Soils and Biodiversity. *Ecology*, 83 (11), 2978.

Dzwonko, Z. & Loster, S. (1989) Distribution of Vascular Plant Species in Small Woodlands on the

Western Carpathian Foothills. *Oikos*, 56 (1), 77.

Elith, J., Leathwick, J.R. & Hastie, T. (2008) A working guide to boosted regression trees. *Journal

of Animal Ecology*, 77 (4), 802–813.

Fang, H.J., Cheng, S.L., Yu, G.R., Yang, X.M., et al. (2014) Nitrogen deposition impacts on the amount

and stability of soil organic matter in an alpine meadow ecosystem depend on the form and rate of

applied nitrogen. *European Journal of Soil Science*, 65 (4), 510–519.

Freckleton, R.P. (2010) Dealing with collinearity in behavioural and ecological data: model averaging

and the problems of measurement error. *Behavioural Ecology and Sociobiology*, 65 (1), 91–101

Friedman, J.H., Meulman, J.J., (2003) Multiple additive regression trees with application in

epidemiology. *Statistics in Medicine*, 22 (9), 1365–1381.

Fuller, R.J., Warren, M.S., (1993), Coppiced woodlands: their management for wildlife, Available

from: http://jncc.defra.gov.uk/pdf/pubs93_Coppicedwoodlands.pdf [Accessed 8th August 2018]

Gardener, T. (2010), Monitoring forest biodiversity, Earthscan Publications,UK

Gaston, K.J., Spicer, J.I. (2004) Biodiversity: an introduction. Blackwell.

Gelman, A. (2008) Scaling regression inputs by dividing by two standard deviations. *Statistics in*

Medicine, 27 (15), 2865–2873.

Gillman, L.N., Wright, S.D., Cusens, J., Mcbride, P.D., et al. (2014) Latitude, productivity and species

richness. *Global Ecology and Biogeography,* 24 (1), 107–117.

Grueber, C.E., Nakagawa, S., Laws, R.J. & Jamieson, I.G. (2011) Multimodel inference in ecology

and evolution: challenges and solutions. *Journal of Evolutionary Biology*, 24 (4), 699–711.

Hayhow DB, Burns F, Eaton MA, Al Fulaij N, August TA, Babey L, Bacon L, Bingham C, Boswell J,

Boughey KL, Brereton T, Brookman E, Brooks DR, Bullock DJ, Burke O, Collis M, Corbet L, Cornish N,

De Massimi S, Densham J, Dunn E, Elliott S, Gent T, Godber J, Hamilton S, Havery S, Hawkins S,

Henney J, Holmes K, Hutchinson N, Isaac NJB, Johns D, Macadam CR, Mathews F, Nicolet P, Noble

DG, Outhwaite CL, Powney GD, Richardson P, Roy DB, Sims D, Smart S, Stevenson K, Stroud RA,

Walker KJ, Webb JR, Webb TJ, Wynde R and Gregory RD (2016) State of Nature 2016. The State of

Nature partnership.

Hermy, M. (1994) Effects of former land use on plant species diversity and pattern in European

deciduous woodlands. *Biodiversity, Temperate Ecosystems, and Global Change*, pp123–144.

Honnay, O., Hermy, M., Coppin, P. (1999) Effects of area, age and diversity of forest patches in

Belgium on plant species richness, and implications for conservation and reforestation. *Biological*

Conservation, 87 (1), pp73–84.

Hrivnák, R., Slezák, M., Jarčuška, B., Jarolímek, I., et al. (2015) Native and Alien Plant Species Richness

Response to Soil Nitrogen and Phosphorus in Temperate Floodplain and Swamp Forests. *Forests*. 6 pp

3501-3513

Hubbell, S.P. (1997) A unified theory of biogeography and relative species abundance and its

application to tropical rain forests and coral reefs. *Coral Reefs*, 16 (5).

Hui, C. & Mcgeoch, M.A. (2014) Zeta Diversity as a Concept and Metric That Unifies Incidence-Based

Biodiversity Patterns. *The American Naturalist,* 184 (5), pp684–694.

James, G., Witten, D., Hastie, T., Tibshirani, R., An Introduction to Statistical Learning: with

Applications in R. New York, Springer Verlag; 2013.

Keersmaeker, L.D., Martens, L., Verheyen, K., Hermy, M., et al. (2004) Impact of soil fertility and

insolation on diversity of herbaceous woodland species colonizing afforestations in Muizen forest

(Belgium). *Forest Ecology and Management*, 188 (1-3), pp291–304.

Keith, S.A., Newton, A.C., Morecroft, M.D., Bealey, C.E., et al. (2009) Taxonomic homogenization of

woodland plant communities over 70 years. *Proceedings of the Royal Society B: Biological Sciences*,

276 (1672), pp3539–3544.

Kennedy, F. &Pitman, R. (2004) Factors affecting the nitrogen status of soils and ground flora in

Beech woodlands. *Forest Ecology and Management,* 198 (1-3), pp 1–14.

Kirby, K. J. *et al.* (2005) 'Long term ecological change in British woodland (1971–2001). A re-survey

and analysis of change based on 103 sites in the Nature Conservancy "Bunce 1971" woodland

survey.', *English Nature Research Reports*, 653(May 2014), p. 139.

Kirby, K.J.; Smart, S.M.; Black, H.I.J.; Bunce, R.G.H.; Corney, P.M.; Smithers, R.J.; Shaw, M.W. (2013).

Woodlands survey flora data 1971-2001. NERC Environmental Information Data

Centre. https://doi.org/10.5285/2d023ce9-6dbe-4b4f-a0cd-34768e1455ae

Kimberley, A., Blackburn, G.A., Whyatt, J.D., Kirby, K., et al. (2013) Identifying the trait syndromes of

conservation indicator species: how distinct are British ancient woodland indicator plants from other

woodland species? *Applied Vegetation Science*, 16 (4), pp667–675

Kimberley, A., Blackburn, G.A., Whyatt, J.D., Smart, S.M. (2014) Traits of plant communities in

fragmented forests: the relative influence of habitat spatial configuration and local abiotic

conditions. *Journal of Ecology*, 102 (3), pp 632–640.

Kimberley, A., Blackburn, G.A., Whyatt, J.D., Smart, S.M. (2015) How well is current plant trait

composition predicted by modern and historical forest spatial configuration? *Ecography*,] 39 (1),

pp67–76.

Kooijman, A.M. & Cammeraat, E. (2010) Biological control of beech and hornbeam affects species

richness via changes in the organic layer, pH and soil moisture characteristics. *Functional Ecology*, 24

(2), pp469–477.

Latombe, G., Hui, C., Mcgeoch, M.A. (2017) Multi-site generalised dissimilarity modelling: using zeta

diversity to differentiate drivers of turnover in rare and widespread species. *Methods in Ecology and*

Evolution, 8 (4), pp431–442.

Latombe, G., McGeoch, M.A., Nipperess, D.A., Hui, C., (2018). zetadiv: Functions to Compute

Compositional Turnover Using Zeta Diversity. R package version 1.1.1. https://CRAN.R-

project.org/package=zetadiv

Lawton, J.H., Brotherton, P.N.M., Brown, V.K., Elphick, C., Fitter, A.H., Forshaw, J., Haddow, R.W.,

Hilborne, S., Leafe, R.N., Mace, G.M., Southgate, M.P., Sutherland, W.J., Tew, T.E., Varley, J., &

Wynne, G.R. (2010) Making Space for Nature: a review of England's wildlife sites and ecological

network. Report to Defra.

Liaw, A., Wiener, M., (2002). Classification and Regression by randomForest. R News 2(3),18--22.

Maskell, L.C., Smart, S.M., Bullock, J.M., Thompson, K., Stevens, C.J., (2010) Nitrogen deposition

causes widespread loss of species richness in British habitats, *Global Change in Biology*, 16, pp 671-

679

McGeoch, M.A .,Latombe, G.,Andrew, N.R., Nakagawa,S.,Nipperess, D.A., Roige,M., Marzinelli, E.M.,

Campbell, H.A., Verges,A Thomas, T.,Steinberg,P. D.,Selwood, K. E.,Hui,C.,(2017), The application of

zeta diversity as a continuous measure of compositional change in ecology. Biorxiv, Available from

www.biorxiv.org/content/early/2017/11/09/216580.full.pdf+html Accessed [12th August 2018]

Mcwilliam, W., Eagles, P., Seasons, M. & Brown, R. (2010) The housing-forest interface: Testing

structural approaches for protecting suburban natural systems following development. *Urban*

Forestry & Urban Greening, 9 (2), pp 149–159.

MetOffice, (2018), Weather and climate data, Available from:

https://www.metoffice.gov.uk/climate/uk/regional-climates/ws#rainfall Accessed [12th August

2018]

Mouchet, M., Levers, C., Zupan, L., Kuemmerle, T., et al. (2015) Testing the Effectiveness of

Environmental Variables to Explain European Terrestrial Vertebrate Species Richness across

Biogeographical Scales. *Plos One,* 10 (7).

Natekin, A. Knoll, A. (2013) Gradient boosting machines, a tutorial. *Frontiers in Neurorobotics*.

[Online] 7.

Natural England and RSPB (2014) Climate change adaptation manual, Available from:

http://publications.naturalengland.org.uk/publication/5629923804839936, Accessed [12th August

2018]

Nicodemus, K.K., Wang, W., Shugart, Y. (2007) Stability of variable importance scores and rankings

using statistical learning tools on single-nucleotide polymorphisms and risk factors involved in gene ×

gene and gene × environment interactions. *BMC Proceedings*, 1 (Suppl 1).

Ohlemuller, R., Wilson, J. (2000) Vascular plant species richness along latitudinal and altitudinal

gradients: a contribution from New Zealand temperate rainforests. *Ecology Letters*, 3 (4), pp 262–

266.

O'Brien, E., (1998), Water energy dynamics, climate, and prediction of woody plant species richness:

an interim model, *Journal of Biogeography*, 25, pp 379-398

Oksanen, J., F., Blanchet, G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Minchin, R, P., O'Hara,

O, B., Simpson, L, G., Solymos, P., Stevens, H, H., Szoecs, E., Wagner, H. (2018). vegan: Community

Ecology Package. R package version 2.5-1. https://CRAN.R-project.org/package=vegan

Packham, J.R., Harding, D.J.L., Hilton, G.M., Stuttard, R.A., (2001), Functional ecology of woodlands

and forests, The Netherlands, Kluwer.

Paillet, Y., Bergès, L., Hjältén, J., Ódor, P., Avon,C., Bernhardt-Romermann, M., Bijlsma, R., Bruyn, L.,

Fuhr, M., Grandin, U., Kanka, R., Lundin, L., Luque, S., Magura, T., Matesanz, S., Meszaros, I., Sebastia,

M., Schmidt, W., Standovar, T., Tothmerez B., Uotila, A., Valladares, F., Vellak,K., Virtanen, R., (2010)

Biodiversity Differences between Managed and Unmanaged Forests: Meta-Analysis of Species

Richness in Europe. *Conservation Biology*, 24 (1), pp101–112.

Pallett, D., Pescott, Schäfer, S. (2016) Changes in plant species richness and productivity in response

to decreased nitrogen inputs in grassland in southern England. *Ecological Indicators,* pp 6873–81.

Palpurina, S., Wagner, V., von Wehrden, H., Hájek, M., Horsák, M., Brinkert, A., Hölzel, N., Wesche,

K., Kamp, J. , Hájková, P., Danihelka, J., Lustyk, P., Merunková, K., Preislerová, Z., Kočí, M., Kubešová,

S., Cherosov, M., Ermakov, N., German, D., Gogoleva, P., Lashchinsky, N., Martynenko, V., Chytrý, M.

and Grytnes, J. (2017), The relationship between plant species richness and soil pH vanishes with

increasing aridity across Eurasian dry grasslands. *Global Ecol and Biogeography*, 26: pp 425-434.

Petit, S., Griffiths, L., Smart, S.S., Smith, G.M., et al. (2004) Effects of area and isolation of woodland

patches on herbaceous plant species richness across Great Britain. *Landscape Ecology*, 19 (5), pp463–

472.

Peterken, G.F. & Game, M. (1984) Historical Factors Affecting the Number and Distribution of

Vascular Plant Species in the Woodlands of Central Lincolnshire. *The Journal of Ecology*,72 (1), 155.

Pitcairn, C., Skiba, U., Sutton, M., Fowler, D., et al. (2002) Defining the spatial impacts of poultry farm

ammonia emissions on species composition of adjacent woodland ground flora using Ellenberg

Nitrogen Index, nitrous oxide and nitric oxide emissions and foliar nitrogen as marker variables.

Environmental Pollution, 119 (1), pp 9–21.

Preston, F.W. (1962) The Canonical Distribution of Commonness and Rarity: Part I. *Ecology*, 43 (2),

185.

Quine, C. P., C. Cahalan, A. Hester, J. Humphrey, K. Kirby, A. Moffat, and G. Valatin. (2011). Ch 8

Woodlands. In: R. Watson and S. Albon, editors. UK National Ecosystem Assessment. UNEP & Defra,

pp 242-293

Richerson, P.J. & Lum, K.-L. (1980) Patterns of Plant Species Diversity in California: Relation to

Weather and Topography. *The American Naturalist*, 116 (4), pp 504–536.

Ridgeway, G with contributions from others (2017). gbm: Generalized Boosted Regression Models. R

package version 2.1.3. https://CRAN.R-project.org/package=gbm.

Ritchie, G. &; Dolling, P. (1985) The role of organic matter in soil acidification. *Australian Journal of*

*Soil Research,*23 (4), 569.

RoTAP (2012) *Review of Transboundary Air Pollution: Acidification, Acidification, eutrophication,*

ground level ozone and heavy metals in the UK., *Centre for Ecology & Hydrology on behalf of Defra*

and the Devolved Administrations.

Russell, J. (1960) Soil fertility changes in the long-term experimental plots at Kybybolite, South

Australia. II. Changes in phosphorus. *Australian Journal of Agricultural Research,* 11 (6), 926.

Scheiner, S.M., (2003), Six types of species area curves, *Global Ecology and Biogeography*, 12, pp 441

- 447.

Schmidt, W., (2005), Herb layer species as indicators of biodiversity of managed and unmanaged

beech forests, *Snow and landscape research.* 79 (1,2).

Shen, G., Yu, M., Hu, X.-S., Mi, X., et al. (2009) Species–area relationships explained by the joint

effects of dispersal limitation and habitat heterogeneity. *Ecology,* 90 (11), pp 3033–3041.

Simkin, S.M., Allen, E.B., Bowman, W.D., Clark, C.M., et al. (2016) Conditional vulnerability of plant

diversity to atmospheric nitrogen deposition across the United States. *Proceedings of the National*

Academy of Sciences, 113 (15), pp 4086–4091.

Slee, B., Kyle, C., Biodiversity and Woodland Ecosystems, Available from:

https://www.hutton.ac.uk/sites/default/files/files/Biodiversity%20&%20Woodland%20Ecosystems(1

).pdf., [Accessed 8th August 2018]

Smart, S.M., Ashmore, M.R., Hornung, M., Scott, W.A., et al. (2004) Detecting the Signal of

Atmospheric N Deposition in Recent National-Scale Vegetation Change Across UK. *Water, Air, & Soil*

Pollution, 4 (6), pp 269–278.

Smart, S.M., Kirby, K., Cornway, P., Wood, C.M., (2013), Woodlands survey of Great Britain 1971 -

2001, Available from: http://nora.nerc.ac.uk/id/eprint/504075/1/N504075CR.pdf, [Accessed 10th

August 2018]

Smart, S.M., Ellison, A.M., Bunce, R.G.H., Marrs, R.H., et al. (2014) Quantifying the impact of an

extreme climate event on species diversity in fragmented temperate forests: the effect of the

October 1987 storm on British broadleaved woodlands. *Journal of Ecology*, 102 (5), pp 1273–1287.

Stevens, C.J., Payne, R.J., Kimberley, A. Smart, S.M., (2016) How will the semi-natural vegetation of

the UK have changed by 2030 given likely changes in nitrogen deposition? *Environmental Pollution,*

208 Part B pp 879–889.

Storch, D., Kiel, P, Kuni, W.E., (2014), Scaling communities and biodiversity, available from:

http://www.cts.cuni.cz/~storch/publications/Storch_et_al_2014_Scaling_communities_biodiversity.

pdf, [Accessed 10th August 2018]

Stroh, P.A., Leach, S.J., August, T.A., Walker, K.J., Pearman, D.A., Rumsey, F.J., Harrower, C.A., Fay,

M.F., Martin, J.P., Pankhurst, T., Preston, C.D. & Taylor, I. (2014). A Vascular Plant Red List for

England. Botanical Society of Britain and Ireland, Bristol.

Symonds, M.R.E., Moussalli, A. (2010) A brief guide to model selection, multimodel inference and

model averaging in behavioural ecology using Akaike's information criterion. *Behavioral Ecology and*

Sociobiology. [Online] 65 (1), pp 13–21.

Sugihara, G., (1979), Minimal community structure: an explanation of species abundance patterns,

The American Naturalist, 116(6) pp 770-787.

Tittensor, D., Walpole, M., Hill, S., Boyce, D., Britten, L., Burgess, G., Butchart, H. M., Leadley, W.,

Regan, P., Alkemade, E., Baumung, R., Bellard, R., Bouwman, C., Bowles-Newark, A., Chenery, N.M.,

Christensen, W., Cooper, V., Crowther, H.R., Ye, A., (2014). A mid-term analysis of progress toward

international biodiversity targets. *Science*. 346.

Thiele, J., Kellner, S., Buchholz, S. & Schirmel, J. (2018) Connectivity or area: what drives plant

species richness in habitat corridors? *Landscape Ecology*, 33 (2), 173–181.

Tjørve, E., Tjørve, K.M. (2017) Species-Area Relationship. *eLS*, 1–9.

Watts, K., Quine, C.P., Eycott, A.E., Moseley, D., Humphrey, J.W., Ray, D., (2008) Conserving forest

biodiversity: recent approaches in UK forest planning and management in: Patterns and Processes in

Forest Landscapes, Springer Verlag.

Williams, C. & Donald, C. (1957) Changes in organic matter and pH in a podzolic soil as influenced by

subterranean clover and superphosphate. *Australian Journal of Agricultural Research*, 8 (2), 179.

Zak, D.R., Freedman, Z.B., Upchurch, R.A., Steffens, M., et al. (2016) Anthropogenic N deposition

increases soil organic matter accumulation without altering its biochemical composition. *Global*

Change Biology, 23 (2), pp 933–944.