Aberrant pathogenic GM-CSF+ T cells and inflammatory CD14+CD16+ monocytes in severe pulmonary syndrome patients of a new coronavirus

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Pathogenic human coronavirus infections, such as severe acute respiratory syndrome CoV (SARS-CoV) and Middle East respiratory syndrome CoV (MERS-CoV), cause high morbidity and mortality.1,2 Recently, a severe pneumonia-associated respiratory syndrome caused by a new coronavirus was reported at December 2019 (2019-nCoV) in the city Wuhan, Hubei province, China3,5, which was also named as pneumonia-associated respiratory syndrome (PARS)6. Up to 9th of February 2020, at least 37, 251 cases have been reported with 812 fatal cases according to the report from China CDC. However, the immune mechanism that potential orchestrated acute mortality from patients of 2019-nCoV is still unknown. Here we show that after the 2019-nCoV infection, CD4+ T lymphocytes are rapidly activated to become pathogenic T helper (Th) 1 cells and generate GM-CSF etc. The cytokines environment induces inflammatory CD14+CD16+ monocytes with high expression of IL-6 and accelerates the inflammation. These aberrant and excessive immune cells may enter the pulmonary circulation in huge numbers and play an immune damaging role to causing lung functional disability and quick mortality. Our results demonstrate that excessive non-effective host immune responses by pathogenic T cells and inflammatory monocytes may associate with severe lung pathology. Therefore, we suggest that monoclonal antibody that targets the GM-CSF or interleukin 6 receptor may potentially curb immunopathology caused by 2019-nCoV and consequently win more time for virus clearance.

Coronavirus, including SARS and MERS, has caused two large-scale pandemic in the last two decades1,2. Although viral evasion of host immune responses and virus-induced cytopathic effects are believed to be critical in disease severity, studies from humans who died of SARS and animal models suggested that an excessive and aberrant host immune response resulting in an exuberant immunopathology and lethal disease7,9. Similarly, patients infected with 2019-nCoV, that have been reported recently, have increased plasma concentrations of
inflammation related cytokines, including interleukins (IL) 2, 7, and 10, granulocyte-colony
stimulating factor (G-CSF), interferon-\(\gamma\)-inducible protein 10 (IP10), monocyte
chemoattractant protein 1 (MCP1), macrophage inflammatory protein 1 alpha (MIP1A), and
tumour necrosis factor \(\alpha\) (TNF-\(\alpha\)), especially in moribund patients\(^{10}\). Importantly, 2019-nCoV
infected patients have developed characteristic pulmonary ground glass changes on imaging
and lymphocytes decreasing\(^{11,12}\). These phenomena suggest severe pulmonary inflammation
and cytokine storm also exist in 2019-nCoV infection. At present, symptomatic treatments
with organ support to moribund patients are the mainstays of clinical managements. It is
urgent to identify the immunopathology mechanism to delay the pulmonary immune injury.

In patients infected with SARS-CoV, it has been reported that the severity of pulmonary
immune injury correlated with extensive infiltration of neutrophils and macrophages in the
lungs\(^{13,14}\), accompanied with increased numbers of neutrophils and monocytes and lower
CD8\(^{+}\) and CD4\(^{+}\) T cell counts in the peripheral blood samples\(^{15-17}\). To identify the immune
characteristic of patients infected with 2019-nCoV, peripheral blood samples from patients
with severe pneumonia were collected for immune analysis. Consistent with previous clinical
characteristics reports\(^{18}\), these hospitalized patients with confirmed 2019-nCoV infection
involved from The First Affiliated Hospital of University of Science and Technology of China
commonly have fever symptoms. The patients in intensive care unit (ICU) have significantly
decreased concentrations of haemoglobin and albumin, but increased concentrations of
C-reactive protein, alanine aminotransferase, aspartate aminotransferase and lactate
dehydrogenase (Extended Data Table 1). The number of total leukocytes in peripheral blood
had no significant differences between patients of 2019-CoV and healthy controls, whereas
the number of lymphocytes decreased significantly in ICU patients. Specifically, monocytes
from both ICU and non-ICU patients significantly decreased compared with healthy controls.
The number of T cells also significantly decreased from both ICU and non-ICU patients. The
CD4\(^{+}\) T cells from both patients in ICU and non-ICU decreased remarkably, whereas CD8\(^{+}\) T
cells decreased more significantly in ICU patients. Other kinds of leukocytes, including
granulocyte, B cells and NK cells have no significantly change in numbers between patients
of 2019-nCoV and healthy controls (Extended Data Figure. 1).

To demonstrate the status of these aberrant altered T cells, several lymphoid antigens have
been analyzed on T cells. These CD4\(^{+}\) T cells in patients infected with 2019-nCoV have
higher expression of CD69, CD38, and CD44 compared with healthy controls (Fig.1a, b),
indicating their activated status. OX40 have been reported to play a major role in promoting
clonal expansion and inducing production of several cytokines in T cells\(^{19}\). In patients
infected with 2019-nCoV, OX40 expression increased remarkably on CD4\(^{+}\) T cells, especially
in severe ICU patients (Fig.1a, b). CD8\(^{+}\) T cells in patients infected with 2019-nCoV also
showed activated phenotype with higher expression of CD69, CD38 and CD44 (Fig.1c, d).
41BB (CD137; TNFRS9) is an activation-induced co-stimulatory molecule, which is
important to priming immune responses of cytotoxic CD8\(^{+}\) T cells\(^{20}\). In ICU patients infected
with 2019-nCoV, the expression of 41BB increased significantly compared to healthy controls
(Fig.1c, d). It has been reported that co-expression of Tim-3 and PD-1 may represent a subset
of T cells with more severe exhaustion in virus infections\(^{21,22}\). It is worth noting that much
higher percentage of co-expression Tim3⁺PD-1⁺ T subset exist both in CD4⁺ and CD8⁺ T cells from patients of 2019-nCoV (Fig.1e-h), especially in ICU patients, suggesting the exhausted status in T cells in these patients infected 2019-CoV.

To further identify the key pathogenic cytokines and the main source of these cytokines, interferon-γ (IFNγ), TNF-α, granulocyte-macrophage colony-stimulating factor (GM-CSF) and IL-6 have been selected to analyzed through intracellular cytokine staining, for these inflammatory mediators have been proven to be critical as the primary cause of inflammatory cytokine storm in patients infected with SARS-CoV or MERS-CoV. Without re-stimulation with PMA or incubation with monensin, high percentage of GM-CSF⁺ and IL-6⁺ expressions could be found in CD4⁺ T cells from patients infected with 2019-nCoV in both ICU and non-ICU patients compared to healthy controls (Fig.2a, c). ICU patients with more severe pneumonia showed correlated higher percentage of GM-CSF⁺ and IL-6⁺CD4⁺ T cells (Fig.2a, c). Pathogenic Th1 cells with both IFN-γ and GM-CSF expression have been reported in central nervous system inflammation. Importantly, aberrant pathogenic Th1 cells with co-expressing IFNγ and GM-CSF exist only in ICU patients infected 2019-nCoV, whereas little was found in non-ICU patients and healthy controls, indicating this pathogenic Th1 cells which have correlative evidence from patients with severe disease, play a critical role for hyper-inflammatory responses in 2019-nCoV pathogenesis (Fig.2b, d). Meanwhile, TNF-α were not significant up-regulated in CD4⁺ T cells from patients of 2019-nCoV (Extended Data Figure 2a-c). CD8⁺ T cells from ICU patients also showed expression of GM-CSF compared to those from non-ICU patients and healthy controls. IL-6 and TNF-α were not found in CD8⁺ T cells (Extended Data Figure 2d, e). Neither NK cells nor B cells were the secreting source of GM-CSF and IL-6 (Extended Data Figure 2f-i).

GM-CSF has been recently been implicated in the pathogenesis of inflammatory and autoimmune diseases, in a mechanism that controls diverse pathogenic capabilities of inflammatory myeloid cells. Among these myeloid cells, monocyte is the pathogenic GM-CSF responsive cells that require GM-CSF to initiate tissue damage in both mouse and human. To identify whether inflammatory monocyte exist in patients infected 2019-nCoV, phenotype and subpopulation of monocytes have been analysis. There was little CD14⁺CD16⁺ inflammatory monocyte subset in healthy controls. By contrast, significant higher percentage of CD14⁺CD16⁺ inflammatory monocyte exist in peripheral blood of patient infected 2019-nCoV. The percentage of CD14⁺CD16⁺ monocyte was much higher in severe pulmonary syndrome patients from ICU (Fig.3a, c). Moreover, these monocyte from patients infected 2019-nCoV also showed capability to secrete GM-CSF. Importantly, significantly higher expression of IL-6 secreted from these inflammatory monocyte especially in ICU patients, which let the cytokine storm even worse (Fig.3b, d). Meanwhile, the number of GM-CSF⁺ monocytes and IL-6⁺ monocytes increased rapidly (Fig.3e), suggesting the potential high risk of inflammatory cytokine storm caused by monocytes that may migrate to the lung and further derive into macrophage or monocyte derived dendritic cells. Thus, in patients infected with 2019-nCoV, GM-CSF potentially links the severe pulmonary syndrome-initiating capacity of pathogenic Th1 cells (GM-CSF⁺IFNγ⁺) with the inflammatory signature of monocytes (CD14⁺CD16⁺ with high expression of IL-6) and their progeny. These activated
immune cells may enter the pulmonary circulation in large numbers and played an immune
damaging role in severe pulmonary syndrome patients (Fig.4).

The study provides the detailed immunopathology report on 2019-nCoV, suggesting excessive
activated immune response caused by pathogenic GM-CSF$^+$ Th1 cells and inflammatory
CD14$^+$CD16$^+$ monocytes may connect pulmonary immunopathology leading to deleterious
clinical manifestations and even acute mortality after 2019-nCoV infections. Consistent with
the situation with SARS-CoV or MERS-CoV$^{12,28}$, it is remarkable that children always
experience mild-moderate clinical illness, elderly individuals exhibit worse outcomes after
infection with 2019-nCoV, further indicating that mature excessive immune response towards
these pathogenic human coronavirus infections play a key role in inducing severe pulmonary
syndrome and even organ failure. However, many urgent questions remain to be answered.
Evidence from alveolar washing fluid and lung autopsy from patients infected 2019-nCoV are
further needed to verify whether and how these aberrant pathogenic immune cells play a fatal
immune damage to cause organ functional disability and mortality. Specific new drugs
targeted 2019-nCoV may take long time to evaluate and develop. At this critical moment,
several marketed drugs to target cytokine storm and reduce immunopathology could be
considered$^{29}$. Blocking inflammatory cytokines may temporarily weaken the anti-infection
immunity, yet such strategy is already the lesser of the evils. Other strategies towards
blocking the over-activated immune response, such as glucocorticoid treatment showed more
side-effect and disappointed outcome towards 2019-CoV$^{18}$. Therefore, we suggest that
monoclonal antibody that targets the GM-CSF or interleukin 6 receptor may potentially
prevent or curb immunopathology caused by 2019-nCoV and consequently win more time for
virus clearance.

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Figure 1. Activated T cells in severe pulmonary syndrome patients of 2019-nCoV.
(a, b) Representative density plots and MFI statistics calculated for CD69, CD38, CD44 and OX40 expressions in gated CD45^+CD3^+CD4^+ T cells (Gating strategy showing in Extended Data Figure 2a) isolated from peripheral blood in healthy controls, ICU and non-ICU patients of 2019-nCoV. (c, d) Representative density plots and MFI statistics calculated for CD69, CD38, CD44 and 41BB expressions in gated CD45^+CD3^+CD8^+ T cells isolated from peripheral blood in healthy controls, ICU and non-ICU patients of 2019-nCoV. (e, f) Representative density plots and percentage statistics calculated for Tim-3 and PD-1 co-expressions in gated CD45^+CD3^+CD4^+ T cells isolated from peripheral blood in healthy controls, ICU and non-ICU patients of 2019-nCoV. (g, h) Representative density plots and percentage statistics calculated for Tim-3 and PD-1 co-expressions in gated CD45^+CD3^+CD8^+ T cells isolated from peripheral blood in healthy controls, ICU and non-ICU patients of 2019-nCoV. Data represent the mean ± SEM. One-way ANOVA. P<0.05 was considered statistically significant.
Figure 2. Pathogenic Th1 cells with high expression of GM-CSF in severe pulmonary syndrome patients of 2019-nCoV.

(a) Representative density plots showing an analysis of GM-CSF and IL-6 expressions in gated CD45⁺CD3⁺CD4⁺ T cells (Gating strategy showing in Extended Data Figure 1a) isolated from peripheral blood in healthy controls, ICU and non-ICU patients of 2019-nCoV. (b) Representative density plots showing an analysis of co-expression of GM-CSF and IFN-γ in gated CD45⁺CD3⁺CD4⁺ T cells isolated from peripheral blood in healthy controls, ICU and non-ICU patients of 2019-nCoV. (c) Statistics calculated by the percentage of GM-CSF⁺ or IL-6⁺ cells from CD4⁺ T cells. (d) Statistics calculated by the percentage of GM-CSF and IFN-γ co-expressing CD4⁺ T cells. Data represent the mean ± SEM. One-way ANOVA. P<0.05 was considered statistically significant.
Figure 3. Inflammatory monocytes with high expression of IL-6 in severe pulmonary syndrome patients of 2019-nCoV.

(a) Representative density plots showing an analysis of CD14 and CD16 expressions in gated CD45+ monocytes (Gating strategy showing in Extended Data Figure 1a) isolated from peripheral blood in healthy controls, ICU and non-ICU patients of 2019-nCoV. (b) Representative density plots showing an analysis of GM-CSF and IL-6 expressions in gated CD45+CD14+ monocyte cells isolated from peripheral blood in healthy controls, ICU and non-ICU patients of 2019-nCoV. (c) Statistics calculated by the percentage of CD14+CD16+ subsets from monocytes. (d) Statistics calculated by the percentage of GM-CSF+ or IL-6+ cells from CD14+ monocytes. (e) Statistics calculated by the cell number of GM-CSF+ CD14+ or IL-6+CD14+ monocytes. Data represent the mean ± SEM. One-way ANOVA. *P*<0.05 was considered statistically significant.
Figure 4. Pathogenic Th1 cells and inflammatory monocytes have positive correlations with severe pulmonary syndrome in patients infected 2019-nCoV.

Pathogenic CD4+Th1 (GM-CSF+IFN-γ+) cells were rapidly activated to produce GM-CSF and other inflammatory cytokines to form a cascade signature of inflammatory monocytes (CD14+CD16+ with high expression of IL-6) and their progeny. These activated immune cells may enter the pulmonary circulation in large numbers and played an immune damaging role in severe pulmonary syndrome patients. The monoclonal antibodies that targets the GM-CSF or interleukin 6 receptor may potentially prevent or curb immunopathology caused by 2019-nCoV.