1	The estrogenic pathway modulates non-breeding female aggression
2	in a teleost fish
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17 Abstract

Aggressive behaviors are widespread among animals and are critical in the competition for 18 19 resources. The physiological mechanisms underlying aggression have mostly been examined in 20 breeding males, in which gonadal androgens, acting in part through their aromatization to 21 estrogens, have a key role. There are two alternative models that contribute to further 22 understanding hormonal mechanisms underlying aggression: aggression displayed in the non-23 breeding season, when gonadal steroids are low, and female aggression. In this study we approach, for the first time, the modulatory role of estrogens and androgens upon non-breeding 24 25 aggression in a wild female teleost fish. We characterized female aggression in the weakly 26 electric fish Gymnotus omarorum and carried out acute treatments 1 h prior to agonistic 27 encounters with either an aromatase inhibitor or an antagonist of androgen receptors. 28 Aromatase inhibition caused a strong distortion of aggressive behavior whereas anti-androgen 29 treatment had no effect on behavior. Territorial non-breeding aggression in female G. omarorum is robust and depended on rapid estrogen actions to maintain high levels of aggression, and ultimately reach conflict resolution from which dominant/subordinate status emerged. Our results taken together with our own reports in males and the contributions from non-breeding aggression in bird and mammal models, suggest a conserved strategy involving fast-acting estrogens in the control of this behavior across species. In addition, further analysis of female non-breeding aggression may shed light on potential sexual differences in the fine tuning of social behaviors.

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38 Keywords

- 39 1. Territorial aggression
- 40 2. Cyproterone acetate
- 41 3. Fadrozole
- 42 4. Gymnotiformes
- 43

44 Highlights

- Female *Gymnotus omarorum* displayed robust territorial aggression in lab settings.
- Acute treatment with aromatase inhibitor lowered aggression levels.
- Aromatase inhibition increased first attack latency and decreased conflict resolution.
- Acute treatment with anti-androgens showed no effects.
- This is the first report of estrogens underlying teleost non-breeding female aggression.

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51 Introduction

- 53 Aggressive behaviors are widespread among animals, and they are key in the competition for
- resources such as food, shelter, and mating opportunities. Males are usually more aggressive
- than females, however robust female aggression is highly prevalent in many species, and not

56 only in the context of maternal aggression. In particular, territorial aggression has been shown 57 to occur in female fish, reptiles, birds, rodents, and non-human primates [1].

The physiological mechanisms underlying aggression have been mostly examined in breeding males, in which the involvement of gonadal androgens has been widely established [2]. In the last 30 years the understanding of the modulation of this complex behavior by sexual hormones has greatly advanced: estrogens have been recognized as additional modulators of aggression and both androgens and estrogens have been shown to have slow and rapid behavioral effects, reflecting genomic and nongenomic mechanisms ([3]; revised in [4]).

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Researchers have incorporated two models which offer valuable opportunities to further 65 understand the physiology of aggression: the very understudied female aggression, which is 66 67 modulated by androgens and estrogens albeit frequently in ways distinct from males, and 68 species in which aggression occurs uncoupled from the breeding season [5-13]. Female aggression has been shown to be promoted by testosterone and at least part of this effect is 69 70 through its aromatization to estrogens [11]. Although estrogens may increase aggression in 71 some species [9,10,14] their effects may differ and brain estrogen receptor subtypes have been 72 shown to mediate opposing effects upon aggressive behavior [15,16]. Some species display 73 aggression uncoupled from the breeding season, when their gonads are regressed and their 74 circulating levels of gonadal androgens are reduced ([17]; revised in [13]). In the non-breeding 75 season, estrogens have a forefront role in the regulation of aggression, mostly through rapid 76 nongenomic mechanisms. Estradiol treatment has been shown to rapidly promote male nonbreeding aggression in mammals and birds [18-23], and acute inhibition of aromatase, the 77 78 enzyme which converts androgens into estrogens, decreases aggression levels [24]. In turn, 79 aggressive interactions between males during the non-breeding season can produce changes 80 of estradiol levels in specific brain areas [25]. In both males and females, non-breeding aggression is linked to modulations of brain estrogen receptors mediating rapid effects [18,26]. 81

Androgens and estrogens can be synthesized locally in the brain either from extragonadal precursors [13,27–29] or *de novo* from cholesterol [30,31]. The study of non-breeding aggression of males and females has opened new avenues of understanding the complexity of the control of aggression and its overall modulation during natural seasonal cycles.

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87 The weakly electric fish *Gymnotus omarorum* is a seasonal breeder, which displays year-long active territorial defense maintaining territories both in the natural habitat [32] and in the lab [33]. 88 89 The non-breeding territorial aggression of wild G. omarorum is robust, elicited in neutral arenas and triggered by the presence of a conspecific [34]. It displays strikingly aggressive encounters, 90 in both intra and intersexual dyads, and males and females show no differences in contest 91 92 outcome, temporal dynamics of the agonistic encounter, levels of aggression, nor submissive 93 signaling [34,35]. Male aggression has been rigorously characterized; contest resolution is 94 biased by body size and once dominant/subordinate status is established; the dominant fish 95 displays a long-lasting exclusion of the subordinate fish from its territory [33]. Male non-breeding aggression is independent of gonadal hormones, as it occurs robustly in fish that have been 96 97 gonadectomized a month prior. In addition, intact fish show circulating androgen (11-98 ketotestosterone) levels unaffected by aggressive encounters. However, aggression is 99 dependent on rapid hormone effects, as the acute inhibition of aromatase distorts contest 100 dynamics and outcome: aggression levels are reduced, and outcome becomes unpredictable 101 [36]. Is fast estrogenic modulation a general strategy underlying non-breeding aggression in this 102 species, independently of sex? This study approaches wild female non-breeding aggression and its control by estrogens and androgens to address this issue. 103

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105 Methods

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107 <u>Animals</u>

108 We used wild adult females of Gymnotus omarorum (Richer-de-Forges et al., 2009) (body-109 length 15 - 26 cm and body weight 9 - 60 g) captured from the field and housed for 4 to 5 weeks in our facilities before experiments. All experiments were carried out during the non-110 breeding season (June to August) [37]. Fish were collected from Laguna del Sauce (34°51'S, 111 112 55°07'W), Maldonado, Uruguay using an electrical detector as previously described [37]. Animals were housed in individual mesh compartments (40x40x60 cm) within large outdoor 113 114 tanks (500 L). These outdoor tanks house aquatic plants brought from the field and were subjected to conditions with natural photoperiod (from LD 10:14 to LD 11:13), temperature 115 (10.41 ± 3.48 °C), and rainfall. To conserve conditions similar to the natural habitat, conductivity 116 117 was maintained under 200 µS/cm [37]. Each fish had a shelter in its compartment and was fed ad libitum with Tubifex tubifex. All experiments were performed according to the regulations for 118 119 the use of animals in research and the experimental protocol was approved by the institutional 120 Ethical Committee of Instituto de Investigaciones Biológicas Clemente Estable (Resolution CEUA IIBCE 004/05/2016). 121

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123 Behavioral set up

124 We observed the agonistic behavior of G. omarorum in dyadic female-female encounters and tested the effect of aromatase inhibition or androgen receptor antagonism during the non-125 126 breeding season. The dyads (7-20% body weight difference between contenders) belonged to one of the following experimental groups: control dyads (n = 8), fadrozole-treated dyads (n = 8) 127 10), or cyproterone acetate-treated dyads (n = 7). We performed the characterization of female 128 agonistic behavior in control dyads. The evaluation of agonistic behavior included engagement 129 in conflict, contest outcome, dynamics, aggression, and submission levels, and these 130 131 parameters were used in the comparison to fadrozole and cyproterone acetate dyads. All 132 experimental groups were composed of fish spanning the same size range and each fish was used only once. Dyads were placed in a behavioral setup (as described in [38]) that allowed 133

simultaneous video and electric recordings, control of photoperiod, water temperature, 134 conductivity, and pH. The setup consisted of 4 experimental tanks (55 × 40 × 25 cm) divided in 135 136 half by a removable glass gate. Due to the nocturnal habits of this species, all experiments were 137 performed at night, in darkness, with infrared LED illumination (Kingbright L- 53F3BT; 940 nm) 138 located above the tanks. Experiments were recorded with an infrared-sensitive video camera 139 (SONY CCDIris, Montevideo, Uruguay) through the glass bottom of the tank. The electric 140 signals of freely moving fish were detected by two pairs of fixed copper wire electrodes connected to two high-input impedance (1 M Ω) amplifiers (FLA-01; Cygnus Technologies Inc., 141 Delaware Water Gap, PA, USA). Images and electric signals were captured by a video card 142 (Pinnacle Systems, PCTV-HD pro stick) and stored in the computer for further analysis. We 143 used a neutral arena protocol with a plain arena (without food or shelter) and simultaneously 144 145 placed each contender in one of the equally sized compartments 2 h prior to the experiment 146 thus providing equal resources (territory and residency) to each individual [34]. Pharmacological manipulations were performed 1 hour before gate removal (see below). The gate was raised 10 147 148 min after sunset, and fish were separated 10 min after conflict resolution. Dyadic contests that 149 did not reach an establishment of dominance/subordination after 20 minutes of interaction were interrupted and labeled as "dyads with engagement without resolution". Dyadic interactions in 150 which there was no engagement during 20 minutes after gate removal were interrupted and 151 considered "dyads with no engagement". 152

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154 <u>Pharmacological manipulations</u>

To analyze the rapid modulation of estrogens and androgens we used acute treatment with different inhibitors. Cyproterone acetate (CA) has been previously reported to effectively block AR in teleost fish [39,40]. Nevertheless, since it had never been used in *G omarorum*, we confirmed the innocuity of its vehicle and the effectiveness of the inhibitor in this species. We performed an experiment blocking a well-known androgenic-dependent trait previously 160 described in non-breeding adults [41]. The electric organ discharge (EOD) of G. omarorum has 161 a multiphasic waveform with four successive components (V1 to V4) [42]. A 15-day treatment 162 with testosterone implants specifically increases the amplitude of the negative component V4 163 which is quantified by the index V4 amplitude/V3 amplitude (AV4/AV3). The reports on the 164 effects of supraphysiological testosterone on EOD waveform were based on mixed groups (non-165 breeding males and females [41]). We subcutaneously implanted 22 animals with testosterone 166 silastic pellets (100 ug/gbw). A stock solution of CA (Sigma, C3412), 2 µg/µl was prepared in mineral oil (Drogueria Industrial Uruguaya), and stored at 4°C. Testosterone implanted animals 167 were divided into a control group (n = 12) which received a daily IP injection of mineral oil for 15 168 169 days, and the treatment group (n = 10) which received a daily IP injection of CA (20 μ g/gbw) for 170 15 days. EOD waveform was recorded following [43]. The testosterone + mineral oil group 171 increased AV4/AV3 amplitude index when comparing day 15 to day 0, as expected (paired t-172 test, p = 0.006, n = 12, Fig. 1A), whereas the group treated with testosterone + cyproterone acetate showed no significant increase in its index (paired t-test, p = 0.8, n = 10, Fig. 1B). This 173 174 result demonstrates cyproterone acetate effectively blocks and rogenic actions in G. omarorum, 175 as shown in other teleost species. To verify the innocuity of mineral oil in the species we 176 compared 6 females injected with PBS to 6 injected with mineral oil (in equal volumes). Fish 177 were recorded individually in the behavioral setup, locomotion was quantified in a 2 minute time window 1 hour after injections. Percentage of time in movement was compared between oil and 178 PBS females, showing no significant difference (Mann-Whitney U test, p = 0.2, $n_{OIL} = 6$, $n_{PBS} = 10^{-1}$ 179 180 6). Basal EOD rate was calculated for each fish 1 hour after injections (see below), and there 181 was no significant difference between oil and PBS females (Mann-Whitney U test, p = 0.57, noil 182 $= 6, n_{PBS} = 6$).

To test the effect of acutely manipulating the androgenic pathway on agonistic behavior, we administered cyproterone acetate to female-female dyads before subjecting them to the neutral arena protocol. One hour before the agonistic encounter, we injected cyproterone acetate (10 µg/gbw, IP) to both individuals, behavioral experiments were performed as described above. To
ensure an effective blocking we used a higher dosis than previously reported for other teleost
species [39].

189 To assess the effect of modulating the estrogenic pathway we used the aromatase inhibitor 190 fadrozole (FAD, Sigma F3806). Fadrozole has been previously reported to effectively block 191 aromatase activity in teleost fish and other vertebrates, including this species [36,44-46]. Before 192 the agonistic encounter, we diluted stock fadrozole (10 μ g/ μ l) in PBS, and IP injected 20 μ g/gbw 193 to both individuals 1 hour prior to gate removal. Behavioral experiments were performed as described above. Control experiments in female-female dyads were carried out injecting PBS (in 194 195 equivalent volume) to both contenders, one hour before the agonistic encounter. Individuals were sexed either by surgical observation 1 month prior, which has been shown has no effect 196 197 upon agonistic behavior in comparison with intact dyads (FAD, control groups following [36]) or 198 by gonadal inspection in euthanized animals after behavioral tests (CA group, euthanization by eugenol solution 8 mg l^{-1}). 199

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201 Data processing

Locomotor and electric displays were analyzed by a researcher blind to the experimental groups 202 203 and treatments. Following [34], we identified the three phases of agonistic encounters: (1) 204 evaluation phase: from time 0 (gate removal) to the occurrence of the first attack; (2) contest 205 phase: from the occurrence of the first attack to conflict resolution (resolution time); and (3) 206 post-resolution phase: 10 min after conflict resolution. Conflict resolution was defined as the 207 moment we observed the third consecutive retreat of one fish without retaliation. This criterion 208 unambiguously defined subordinate status; fish fulfilling this requirement were never observed to change their status in the following 10 min of interaction. To calculate attack rate, we divided 209 210 the number of attacks (bites, nips, nudges) [47] by contest duration time in seconds. We 211 identified previously described transient submissive electric signals: offs (EOD interruptions), 212 chirps (abrupt increases in EOD rate) [34] and electrical submission (stable, post-resolution EOD rate rank) [48]. We calculated off and chirp rate (for contest and post-resolution phases 213 214 together) by dividing the number of offs and chirps produced in both phases by the duration in 215 sec. To calculate electrical submission, we determined mean EOD rates in dominants and subordinates during pre-contest (before gate removal) and post-resolution in 10 - 60 s 216 217 recordings from both phases, using the software Clampfit (Axon, 10.0.0.61). To quantify the difference in EOD rate between contenders, we calculated the subordinate / dominant EOD rate 218 index (S/D rate index). Index values below 1 indicate that the dominant EOD rate is higher than 219 220 the subordinate EOD rate.

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222 Statistics

223 To analyze the effect of cyproterone acetate on EOD waveform we used a paired t-test and 224 compared A V4/ A V3 index in the same individual at day 0 and day 15 of treatment. As behavioral data did not fit a gaussian distribution, they were analyzed with non-parametric tests: 225 226 Wilcoxon Matched-Pairs test (paired variables in the same fish or the same dyad comparing 227 dominant and subordinate), Mann-Whitney U test (independent variables using sets of data from different fish). For this reason, results are expressed as median ± interguartile range 228 229 throughout. Chi square test 2x2 Fisher was used to compare the proportion of dyads that achieved contest resolution in control and FAD groups. 230

- 231
- 232 Results

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234 <u>Female-female non-breeding territorial aggression</u>

236 Female-female dyads of G. omarorum displayed robust agonistic behavior in the neutral arena 237 protocol (Fig. 2). All dyads engaged in agonistic interactions, and all ended in the establishment 238 of stable dominance/subordination relationships. The larger fish became dominant in 6 out of 8 239 contests. Agonistic encounters exhibited characteristic phases previously described for the 240 species: (1) a short evaluation phase (first attack latency = 34.8 ± 8.8 s, n = 8); (2) a contest phase (contest duration, 273 ± 85.6 s, n = 8), and (3) a post-resolution phase (Fig. 2A). The 241 242 contest phase was characterized by overt aggressive displays, higher in dominants compared to subordinates (attack rate Wilcoxon Matched-Pairs test, p = 0.008, n = 8, Fig 2B). In addition, 243 dominant and subordinate attacks were strongly correlated during contests ($R^2 = 0.8$, p = 0.003, 244 n = 8, data not shown). During contest and post-resolution phase subordinates emitted electric 245 signals of submission (off rate 0.02 ± 0.005 , n = 8; and chirp rate 0.025 ± 0.01 , n = 8, data not 246 247 shown). After resolution, EOD rate rank was established, and the acquired status of dominants 248 and subordinates did not reverse (Fig. 2C). In the pre-contest phase contenders did not differ in 249 their basal EOD rates (median S/D rate index = 1.01) whereas after resolution dominants' EOD 250 rates were higher than their counterpart subordinates (median S/D rate index = 0.6; pre-contest 251 vs post-resolution S/D rate index: Wilcoxon Matched-Pairs test, p = 0.016, n = 7, Fig. 2C).

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253 Hormonal modulation of aggression: the analysis of rapid effects through acute treatments.

254 To assess rapid effects of estrogens on the expression of non-breeding female territorial aggression, we acutely treated both fish of the dyad with fadrozole, an inhibitor of the aromatase 255 256 enzyme. The first and foremost effect of aromatase inhibition upon dyadic interaction was a significant decline in overall aggression. As shown in Fig. 3A, 8 out of 8 control dyads engaged 257 in conflict in less than 28 seconds; all of them reached conflict resolution and establishment of 258 259 dominant/subordinate status in less than 156 seconds. Fadrozole-treated dyads showed conflict 260 engagement in 7 of 10 dyads, of which only 5 resolved their conflict (conflict resolution: Chi square test 2x2, Fisher exact Test, p=0.035, $n_{FAD} = 10$, $n_{CTRL} = 8$). The other two dyads which 261

262 engaged in conflict did not achieve resolution in a 20-minute period (Fig. 3A). Of the 5 dyads 263 which resolved the conflict, in 3 the larger contender achieved dominance. The administration of 264 fadrozole increased the latency to the first attack in comparison to control dyads (Mann-Whitney 265 U test, p = 0.014, $n_{FAD} = 7$, $n_{CTRL} = 8$; Fig. 3B) and in the dyads in which conflict was resolved, there 266 was a conspicuous decrease in aggression levels. The attack rates of dominants displayed 267 during contests were significantly lower than control dyads (Mann-Whitney U test, p = 0.019, n_{FAD} 268 = 5, n_{cTRL} = 8; Fig. 3C), as were the attack rates of subordinate fish (Mann-Whitney U test, p = 0.006, $n_{FAD} = 5$, $n_{CTRL} = 8$; Fig. 3D). This striking overall effect upon aggression levels most 269 probably accounts for the lower percentage of conflict resolution. However, it does not generate 270 271 a significant modification in the accompanying electric social signals of submission. 272 Subordinates of the dyads with aromatase inhibition did not differ in off rate (Mann-Whitney U 273 test, p =0.9, $n_{\text{\tiny FAD}} = 7$, $n_{\text{\tiny CTRL}} = 8$), nor chirp rate (Mann-Whitney U test, p =0.3, $n_{\text{\tiny FAD}} = 7$, $n_{\text{\tiny CTRL}} = 8$). In 274 addition, EOD rate rank was established in fadrozole treated dyads just as in control ones, as post-resolution S/D rate index values were lower than before the contest, reflecting a higher rate 275 276 of dominants in comparison to subordinates after resolution (pre-contest vs post-resolution S/D 277 rate index_{FAD}: Wilcoxon Matched-Pairs test, p = 0.06, n = 5, data not shown). Not only did the S/D rate index decrease after resolution as in controls, but the values in themselves did not differ 278 between fadrozole and control dyads, neither during pre-contest phase (Mann-Whitney U test, p 279 = 0.84, n_{FAD} = 5, n_{CTRL} = 7) nor after conflict resolution (Mann-Whitney U test, p = 0.11, n_{FAD} = 5, n_{CTRL} 280 = 7). Fadrozole treated dyads showed no difference compared to controls in locomotor activity 1 281 h after injection, before gate removal (Mann-Whitney U test, p = 0.28, $n_{rad} = 20$, $n_{crrl} = 16$, data 282 283 not shown).

To analyze if, in addition to estrogenic modulation, endogenous androgens have direct rapid effects upon non-breeding aggression, we treated both contenders of a group of dyads with cyproterone acetate, an androgen receptor antagonist. Acutely blocking the androgen receptor function had no effects upon overall aggression dynamics. Neither conflict engagement, latency 288 to first attack, conflict resolution, nor aggression levels of dominant or subordinate fish showed 289 any significant difference in comparison to control dyads (Mann-Whitney U test attack latency, p = 0.56, n_{CA} = 7, n_{CTRL} = 8; Mann-Whitney U test dominant attack rate, p = 0.67, n_{CA} = 7, n_{CTRL} = 290 291 8; Mann-Whitney U test subordinate attack rate, p = 0.09, $n_{CA} = 7$, $n_{CTRL} = 8$, data not shown). 292 Subordinates of the dyads with cyproterone acetate did not differ in off rate emission (Mann-293 Whitney U test, p =0.45, $n_{CA} = 7$, $n_{CTRL} = 8$), nor chirp rate (Mann-Whitney U test, p =0.25, $n_{CA} = 7$) 294 7, n_{CTRL} = 8). EOD rate rank was established in cyproterone acetate treated dyads as well as in control ones (pre-contest vs post-resolution S/D rate index_{CA}: Wilcoxon Matched-Pairs test, p =295 296 0.016, n = 7). Moreover, S/D rate index did not differ between cyproterone acetate and control dyads, neither during pre-contest phase (Mann-Whitney U test, p = 0.38, $n_{CA} = 7$, $n_{CTRL} = 7$) nor 297 after conflict resolution (Mann-Whitney U test, p =, 0.38, $n_{CA} =$ 7, $n_{CTRL} =$ 7). 298

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300 Discussion

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This is the first report on the evaluation of hormonal control of non-breeding female aggression in a teleost species. We show that a. non-breeding females of *Gymnotus omarorum* display robust aggressive territorial behavior, b. this aggression depends on rapid modulation of aromatase, revealing the importance of short-term effects of estrogens, and c. androgens show no rapid modulation upon this behavior.

307

Territorial aggression in *G. omarorum* has previously been reported to occur both in males and females, and be sexually monomorphic [34,35]. Nevertheless, the careful analysis of female aggression separately from male aggression is imperative to approach the hormonal modulation of this behavior. Overtly similar behavior may in fact be based on sexually distinct underlying mechanisms [49]. This is the case of the similar parenting behavior in male and female prairie voles, which are underlain by sexually different vasopressin innervation in key brain areas 314 (reviewed in [49]). In the present study, female G. omarorum engaged, as expected, in highly 315 aggressive and escalated contests during the non-breeding season, competing for space as a 316 resource. After a short evaluation time, contests were initiated and resolved in less than 5 317 minutes. The larger female won most of the fights, submission was signalled by transient social electric signals and dominance was displayed by actively excluding the subordinate fish from 318 319 the acquired territory while establishing an EOD rate rank, as has previously been reported 320 [34,48,50]. Non-breeding territorial behavior in G. omarorum is extremely robust and maintains its features and overall dynamics independently of sex, across controls of many experimental 321 approaches and in surgically sexed animals [33,34,36,48,50,51]. Lab results showing no sexual 322 323 differences in non-breeding territorial aggression complement the data on spacing of this species in the wild, in which males and females own same-sized territories [32]. Non-breeding 324 325 territorial aggression may be related to the defense of foraging patches since electrogeneration 326 has been reported to impose high basal metabolic requirements [52]. Weakly electric fish 327 continuously discharge EODs throughout their life and EOD amplitude is known to be strongly 328 correlated with fish size in G. omarorum [53] and other electric fish [54–56]. Larger fish not only 329 hold larger territories in the wild, regardless of sex [32], but also have a higher chance of 330 winning a contest ([34], this study).

331

Non-breeding aggression in birds and mammals has been reported to be mediated by 332 circulating precursors which are converted into active sex steroids (androgens and estrogens) 333 334 within the brain (reviewed in [12]). As an exploratory step in evaluating the role of sex steroids in non-breeding female aggression in G. omarorum, we pharmacologically manipulated the 335 androgenic pathway. Rapid, nongenomic actions of androgens have been reported to occur in 336 337 various tissues, including the brain, mediated by androgen receptors [57-59]. Cyproterone 338 acetate is an antagonist of androgen receptors, including those mediating fast nongenomic actions [60] and was effective in G. omarorum as it blocked an androgen-induced change in 339

340 EOD waveform (Fig. 1). Short-term blocking of androgen receptors, however, showed no 341 influence upon non-breeding aggression dynamics nor the establishment of 342 dominant/subordinate status. These results suggest that if androgens are directly involved at all 343 in sustaining aggression during the non-breeding season in females, their action may be 344 through genomic mechanisms, and thus be evinced in a longer time frame. In male G. 345 omarorum, non-breeding aggression remains unchanged under long term elimination of gonadal 346 hormones, ruling out their role as modulators [36]. In the year-round territorial fish Stegastes 347 nigricans, non-breeding circulating androgens are low, and remain so in both sexes after an 348 aggressive encounter, although long term androgen receptor blocking decreases aggression in 349 males but not females [17,61]. We have yet to explore if long-term direct effects of androgens, 350 regardless of their source, occur in male and female non-breeding aggression in G. omarorum.

351

352 Estrogens have been put forth as key elements in models of non-breeding aggression. Pioneer 353 studies in birds show long-lasting aromatase inhibition reduces aggression which can be 354 recovered by estradiol treatment [24,62]. We focused on the role of this steroidal pathway in the 355 non-breeding aggression of female G. omarorum using acute aromatase inhibition and showed 356 an important role of estrogens. There was an overall decrease in motivation to display 357 aggression, revealed both by an important delay in initiating overt aggression and a significant 358 decrease in dyads which reached conflict resolution (Fig. 3). These results were strikingly 359 similar to what has been reported for male G. omarorum, in which potential winners failed to 360 either resolve contests or achieve dominance when acutely treated with an aromatase inhibitor [36]. Interestingly, in spite of affecting the intensity of aggressive interactions, aromatase 361 inhibition did not affect electric signalling, which suggests that the electrogeneration system is 362 363 not sensitive to rapid estrogen effects per se. Our results, taken together with reports of 364 estrogenic modulation of male aggression [36] support estrogen as a key modulator of nonbreeding aggression, acting through rapid mechanisms in this species. Estrogen, most probably 365

366 brain derived, has been reported to have rapid effects underlying non-breeding aggression in 367 birds and mammals [18,19,23,24,63,64]. The magnitude of these rapid effects upon behavior 368 have been shown to depend on estrogen sensitivity i.e. higher estrogen receptor expression, in 369 key brain regions [21]. It is interesting to focus on how female aggression can bring novel and 370 sexually distinct mechanisms into consideration. Rapid effects of estrogens have been reported 371 to be more pronounced in female than male brains in zebra finch [13]. Non-breeding female 372 Siberian hamsters, which display robust aggression, have very low circulating estrogen levels 373 which are offset by a seasonal increase in estrogen sensitivity in brain areas associated with 374 aggressive behavior [63]. Teleost fish, which have exceptionally high aromatase activity that shows both seasonal plasticity and sexual differences (reviewed in [65]) emerge as an 375 376 advantageous model for this approach.

377

378 Concluding remarks

379 In this study we show for the first time in a female teleost that non-breeding aggression depends 380 on estrogen production. Females of *Gymnotus omarorum* rely on short term estrogen synthesis 381 to engage in territorial aggression, maintain high levels of aggression, and ultimately reach 382 conflict resolution from which dominant/subordinate status emerges. Our results highlight the 383 importance of fast acting estrogens in the control of non-breeding female aggression in G. 384 omarorum which taken together with our reports from males of this species, as well as contributions from bird and mammal models point to conserved strategies across species. 385 386 Further analysis of female non-breeding aggression may shed light on potential sexual differences in the fine tuning of social behaviors. 387

388

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- 587

588 Figure Legends

589

- 590 Figure 1
- Test of effectiveness of cyproterone acetate (CA) in *Gymnotus omarorum*. **A**. Animals implanted with testosterone (T) and subjected to a daily IP injection of mineral oil (n=12) for 15 days changed their EOD waveform as expected [41], increasing the amplitude of the V4 component in comparison to day 1, shown as a significant increase of the index V4 amplitude / V3 amplitude (AV4/AV3) (paired t-test, p = 0.006, n = 12). **B**. Animals implanted with T and subjected to a daily IP injection of CA for 15 days (n=10) showed no significant differences in V4 amplitude (paired t-test, p = 0.8, n = 10).

598

599 Figure 2

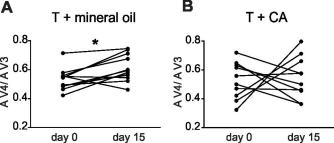
Female non-breeding aggression characterization. A. Female dyadic encounters displayed the
 three typical phases of agonistic behavior. Submission signals began during the contest phase

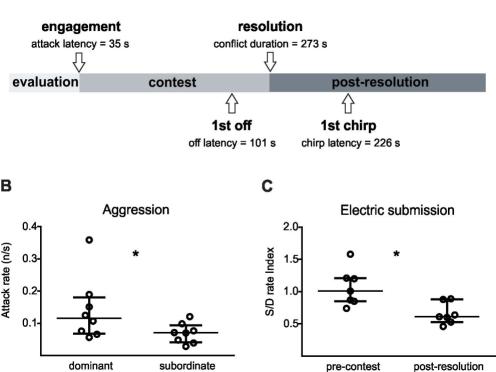
and continued into post-resolution (n=8 control dyads). **B.** Individuals which achieved dominance showed higher aggression levels than their counterparts during conflict (attack rate Wilcoxon Matched-Pairs test, p = 0.008, n = 8). **C.** Dominant and subordinate status was expressed by post contest EOD rate. Rates were compared by the subordinate / dominant EOD rate index (S/D rate index). Index values were near 1 before contests and significantly lower after conflict resolution (pre-contest vs post-resolution S/D rate index: Wilcoxon Matched-Pairs test, p = 0.016, n = 7).

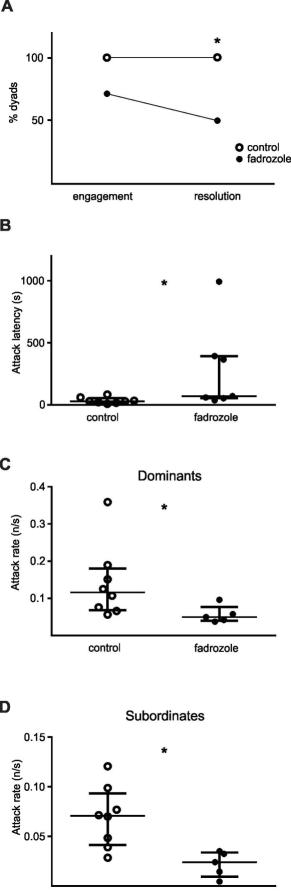
609 Figure 3

Effects of acute inhibition of aromatase on female non-breeding aggression. A. Fadrozole 610 611 treated dyads engaged less in conflict than control dyads and only 5 dyads reached conflict 612 resolution and established dominant/subordinate status (Chi square test 2x2, Fisher exact Test, p=0.035, $n_{FAD} = 10$, $n_{CTRL} = 8$). B. The latency to first attack in FAD treated dyads was significantly 613 lower than in control dyads (Mann-Whitney U test, p = 0.014, $n_{rad} = 7$, $n_{cre} = 8$). C. Individuals 614 615 which achieved dominance displayed lower attack rates during contests in FAD treated dyads compared to control dyads (Mann-Whitney U test, p = 0.019, $n_{rad} = 5$, $n_{cret} = 8$) **D**. Subordinates 616 617 showed lower attack rates in FAD treated dyads in comparison to controls (Mann-Whitney U 618 test, p = 0.006, $n_{FAD} = 5$, $n_{CTRL} = 8$).

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control

fadrozole