

Bacteria maintain Slightly Beneficial Genes and Selfish Genetic Elements through the evolution of Horizontal Gene Transfer

B. van Dijk¹ | P. Hogeweg¹ | H.M. Doekes¹ | N. Takeuchi²

1= Utrecht University, Theoretical Biology, the Netherlands

2= University of Auckland, School of Biological Sciences, New Zealand

Correspondence: B. van Dijk (bramvandijk88@gmail.com), P. Hogeweg (p.hogeweg@uu.nl)

1 Abstract

2 Horizontal gene transfer (HGT) is a key component of bacterial evolution, which in concert with
3 gene loss can result in rapid changes in gene content. While HGT can evidently aid bacteria
4 to adapt to new environments, it also carries risks since bacteria may pick up selfish genetic
5 elements (SGEs). Here, we use modeling to study how bacterial growth rates are affected by
6 HGT of slightly beneficial genes, if bacteria can evolve HGT to improve their growth rates,
7 and when HGT is evolutionarily maintained in light of harmful SGEs. We find that we can
8 distinguish between four classes of slightly beneficial genes: indispensable, enrichable, rescuable,
9 and unrescuable genes. Rescuable genes – genes that confer small fitness benefits and are lost
10 in the absence of HGT — can be collectively retained by a bacterial community that engages
11 in HGT. Although this ‘gene-sharing’ cannot evolve in well-mixed cultures, it does evolve in a
12 spatially structured population such as a biofilm. Although HGT does indeed enable infection
13 by harmful SGEs, HGT is nevertheless evolutionarily maintained by the hosts, explaining the
14 stable coexistence and co-evolution of bacteria and SGEs.

15 Introduction

16 Horizontal Gene Transfer (HGT), the transmission of genetic material between unrelated in-
17 dividuals, is a major factor driving prokaryotic evolution (Ochman et al., 2000; Doolittle and
18 Zhaxybayeva, 2009; Vogán and Higgs, 2011). Recent estimates of the rate of HGT in closely
19 related bacteria are staggeringly high (Iranzo et al., 2019; Sakoparnig et al., 2019), with HGT
20 possibly even outpacing gradual sequence evolution (Hao and Golding, 2006; Puigbò et al., 2014;
21 Vos et al., 2015). Combining this with the fact that prokaryotes adapt mostly through rapid
22 gene loss (Kuo and Ochman, 2009; Morris et al., 2012), bacterial adaptation appears to be
23 mainly driven by changes in gene content (Snel et al., 2002; Treangen and Rocha, 2011; Nowell
24 et al., 2014). Rather than waiting for rare beneficial mutations to arise, taking up tried-and-true
25 genes from a shared ‘mobile gene pool’ allows bacteria to adapt quickly to different ecological
26 opportunities (Jain et al., 2003; Wiedenbeck and Cohan, 2011; Casacuberta and González, 2013;
27 Mell and Redfield, 2014; Niehus et al., 2015; Lopatkin et al., 2016). Indeed, many bacterial
28 species show patterns consistent with this rapid turn-over of genes, where strains from a single
29 niche contain a relatively small set of genes, while the set of genes found by sampling strains from
30 various niches (*i.e.* the pan-genome) is much richer (Welch et al., 2002; Lefébure and Stanhope,
31 2007; Touchon et al., 2009; Kim et al., 2015). Hence, genes appear to be rapidly lost from any
32 individual lineage, but are retained in a much larger gene pool through HGT.

33 When considering the effects of HGT on gene content, it is important to note that HGT does
34 not only *recombine* genes, but also has the ability to *copy* genes from one individual to another.
35 The latter process has been referred to as “additive HGT” (Thomas and Nielsen, 2005; Choi
36 et al., 2012; Soucy et al., 2015), and is quite distinct from processes like sex and recombination
37 because genes can now replicate independently from the cell cycle, and can thus spread at their
38 own pace (Hall et al., 2016; Nazarian et al., 2018; Takeuchi et al., 2015; Shapiro et al., 2012).
39 In additive HGT, a host cell picks up genes either from other cells or from the environment,
40 which may subsequently be expressed. Aside from the cost of expressing the machinery to do
41 so, this process also poses a risk in the form of Selfish Genetic Elements (SGEs), whose success
42 may depend on their ability to be transferred to new hosts (Bergstrom et al., 2000; Lili et al.,
43 2007; Slater et al., 2008). Hence, while picking up genes can be very beneficial for bacteria
44 when adapting to a new environment (Casacuberta and González, 2013; Mell and Redfield, 2014;
45 Lopatkin et al., 2016), taking up foreign DNA is also a costly and highly risky endeavour (Vogán
46 and Higgs, 2011; Baltrus, 2013). Given these disadvantages, is HGT ever adaptive for bacteria
47 when the environment does not change? Can HGT be considered an evolved trait of bacteria,
48 or is it only a side-effect of other unrelated processes like infection by SGEs or DNA repair
49 (Redfield, 2001)?

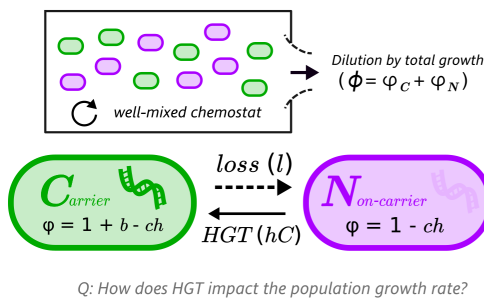
50 To address these questions, we here present and analyse a model of a bacterial population
51 undergoing additive HGT of a single gene, where we assume that HGT is a costly process
52 for the host cells. We show that HGT can have a positive impact on population growth rates
53 by recovering slightly beneficial genes, which are hard to maintain in the population through
54 selection alone. Based on whether or not the genes are lost from the population without HGT,
55 and whether HGT can improve the population growth rate, we find that genes fall into one
56 of five gene classes: (i) *indispensable genes*, that are never lost from the population, and for
57 which HGT is therefore unnecessary and deleterious, (ii) *enrichable genes*, that are not lost from
58 the population, but enriching the genes via HGT can nevertheless improve growth rates, (iii)
59 *rescuable genes*, which are lost from the population without HGT, but can be rescued by HGT
60 which improves population growth rates, and (iv) *unrescuable genes* which are also lost from
61 the population without HGT, but recovering them with HGT does not improve growth rates,
62 and (v) *selfish genetic elements*, which confer a fitness penalty but can persist through HGT.
63 For enrichable and rescuable genes, where HGT can increase population growth rates, we also

64 investigate if HGT can evolve *de novo*. While HGT can readily evolve for enrichable genes,
65 which have sufficient donor cells to interact with, evolving HGT to ‘rescue’ rescuable genes faces
66 a problem: HGT is needed for the gene to persist in the population, but sufficient donor cells
67 are required to make HGT adaptive. This paradox is however resolved in a spatially structured
68 population like a biofilm, as even a minority of donor cells can be locally abundant, giving rise to
69 a localised ‘gene-sharing’ community that eventually overgrows the whole population. Finally,
70 in this spatial eco-evolutionary context, HGT is evolutionarily maintained even when exploited
71 by harmful genetic parasites, resulting in stable coexistence of bacteria and SGEs. Our model
72 provides important insights and search images for how slightly beneficial genes may spread, or
73 fail to spread, in an evolving microbial population.

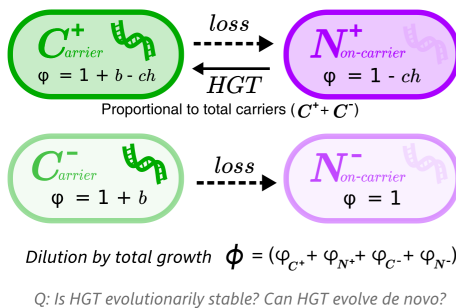
74 Results

75 Throughout this study, we analyse how the ability of HGT affects the growth rates of bacterial
 76 populations, and to what extent the ability of HGT is evolvable for the host cells. We do so by
 77 considering a ‘hard case’, where HGT is a continuously costly process for all cells, and is only
 78 beneficial under specific circumstances. Consider two cell types: cells that carry a beneficial gene
 79 (carriers, C), and cells that do not (non-carriers, N). The benefit of carrying the gene, b , makes
 80 carriers grow faster than non-carriers (or slower if $b < 0$, *i.e.* the gene is a selfish element), but
 81 carriers lose the beneficial gene at a fixed rate l . Non-carriers can recover genes by interacting
 82 with carriers through HGT. We have studied these dynamics with different models, first using
 83 simple ordinary differential equations (ODEs, **Figure 1A/B**), and later an individual-based
 84 model that takes spatial population structuring into account (IBM, **Figure 1C**). The equations
 85 and full description of the models can be found in the Methods section.

A) ODE model (carriers and non-carriers)



B) ODE model (carriers and non-carriers with / without HGT)



C) Individual-based, eco-evolutionary model

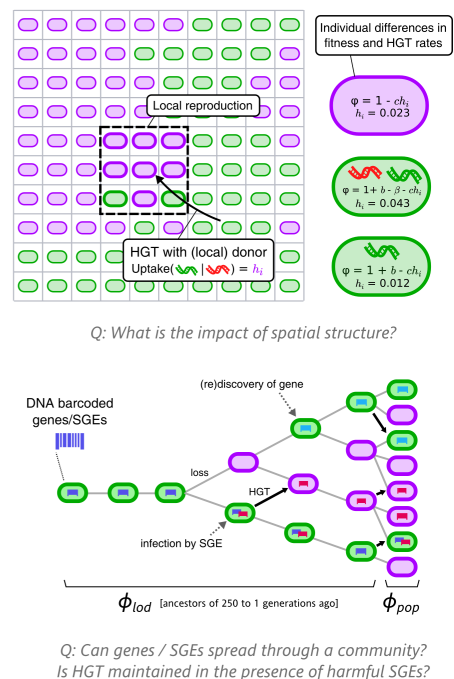


Figure 1: Graphical overviews of the different models: This study uses a series of models with gradually increasing complexity. The first two models are composed of Ordinary Differential Equations (ODEs), and the third model is an individual-based model (IBM). The models describe a population of bacterial cells which either carry a beneficial gene (carriers, C) or do not carry the genes (non-carriers, N). The cells are competing for a limited resource, where the intrinsic growth is 1, b is the growth rate advantage (or disadvantage) for carrying the gene, l is the rate at which the gene is lost, h is the rate of HGT, c is the cost of HGT, φ is the growth rate of sub-populations / individual cells, and ϕ represents the total growth rate. The IBM makes a distinction between the average growth rate of the population (ϕ_{pop}) and the average growth rate of the line of descent (ϕ_{lod} , previous 250 generations of cells). In the IBM, both beneficial genes (with benefit b , green) and harmful SGEs (with penalty β , red) are taken into account. Genes and SGEs are tagged with a unique barcode when they flux in, which are inherited upon reproduction or transfer. Parameters c , h and l are assumed to be positive. For b we focus on slightly beneficial genes ($b \simeq l$) and selfish genetic elements ($b < 0$).

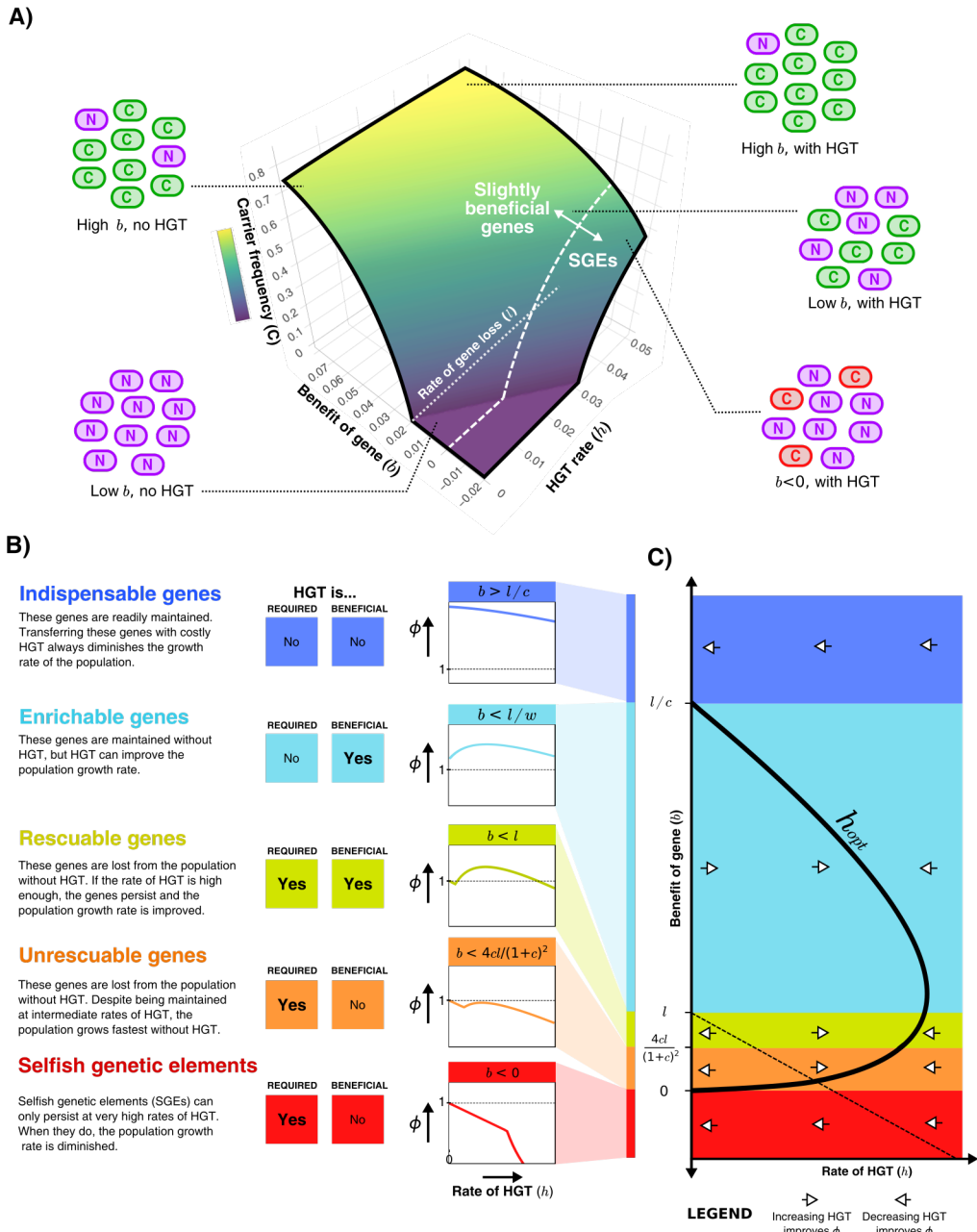


Figure 2: HGT can help genes persist in the population, resulting in distinct gene classes of slightly beneficial genes A) The frequency of carrier cells is shown in a 3D surface plot for different values of b and h . This function is derived in the Supplementary Material and given by Equation 10, and here drawn for $l = 0.02$ and $c = 0.2$. The white dashed line ($b = 0$) gives the boundary between slightly beneficial genes and SGEs. Cartoons illustrate how, for a very beneficial gene (high b), HGT leads only to a mild increase in carrier cells, how HGT has a large impact when the gene brings a smaller fitness effect (low b), and how SGEs can also persist with high HGT rates ($b < 0$). B) Different classes of slightly beneficial genes can be distinguished based on (i) if HGT is *required* for the gene to persist within the population and (ii) if HGT is *beneficial* for population growth rates. The graphs on the right-hand side show, for each of these classes, how an increasing rate of HGT (x -axis) influences the population growth ϕ (y -axis). C) A bifurcation diagram shows how the population growth rate is either improved or diminished by HGT for different values of the rate of HGT (h , x -axis) and the benefit parameter (b , y -axis). The HGT rate that optimises population growth rates ($h_{opt} = \sqrt{bl/c - b}$) is depicted by the thick black curve. The dashed line is given by $h = l - b$, above which the genes are able to persist in the population. Finally, white arrows depict whether $\delta\phi/\delta h$ is positive or negative, indicating how more/less HGT changes the population growth rate.

86 Starting with the simplest model depicted in **Figure 1A**, we first illustrate how the steady-state
87 frequency of carrier cells depends on the benefit of the gene (b) and the rate of HGT (h). **Figure**
88 **2A** shows that if the gene is sufficiently beneficial, most of the population will consist of carrier
89 cells with or without HGT. Despite being continuously lost, these genes are beneficial enough
90 to readily persist in the population through selection. An increased rate of HGT results in only
91 marginally more carrier cells. For genes with a much smaller benefit, HGT can have a large
92 impact on the frequency of carrier cells in the population. In fact, if the benefit is very small
93 ($b < l$, white dotted line), carriers do not survive in the absence of HGT at all, but can occur
94 in fairly high frequencies with sufficient HGT. Note however that the mere survival of carriers
95 with beneficial genes does not imply a positive impact on the population growth rate, as the
96 model assumes HGT comes at a cost. Actually, at sufficiently high rates of HGT, carrier cells
97 with costly genes ($b < 0$) can also persist in the population, which by definition is deleterious
98 for growth. These costly genes could either be genes that are expressed but not useful in the
99 current environment, or Selfish Genetic Elements (SGEs). Throughout this study, we consider
100 genes with $b < 0$ to be SGEs.

101 Slightly beneficial genes fall into distinct gene classes

102 To better understand the impact of HGT, we next study how HGT impacts the population
103 growth rate (ϕ). The population growth rate in steady state is given by **Equation 1** displayed
104 below (see full derivation in Supplementary Section 1). The function is comprised of two parts;
105 one where the population consists only of non-carriers (if $h \leq l - b$), and one where carriers
106 survive and the gene persists within the population (if $h > l - b$). When the gene persists, an
107 optimal growth rate is found at $h_{opt} = \sqrt{bl/c} - b$. (see Supplementary Material).

$$\phi^*(h) = \begin{cases} 1 - ch & \text{if } h \leq (l - b) \text{ (gene cannot persist)} \\ 1 - ch + b - \frac{bl}{b+h} & \text{if } h > (l - b) \text{ (gene persists).} \end{cases} \quad (1)$$

108 By analysing **Equation 1**, we find that we can distinguish distinct classes of genes depending
109 on (i) whether HGT is required for the gene to persist within the population, and (ii) whether
110 HGT is beneficial for the population growth rate (**Figure 2B**). When genes are highly beneficial
111 ($b > l/c$), HGT is not required for the gene to persist, and HGT does not improve the population
112 growth rate. In other words, although transferring these *indispensable genes* yields a small
113 increase in the number of carrier cells, this does not outweigh the costs of HGT. When considering
114 lower values of b , HGT is still not required for the gene to persist within the population, but
115 transferring these *enrichable genes* is nevertheless beneficial for population growth rates. For
116 even lower benefit ($b < l$), HGT is a necessity for the gene to persist within the population, but
117 the population growth rate can be improved by means of intermediate rates of HGT. We call these
118 genes *rescuable genes*. If we consider genes with even smaller fitness effects ($b < 4cl/(1 + c)^2$),
119 HGT is still required for the survival of these genes, but the population growth rates are highest
120 in the absence of HGT. Thus, despite being defined as a beneficial gene ($b > 0$), transferring
121 these *unrescuable genes* is not beneficial. Finally, we can consider SGEs, genes with a negative
122 effect on fitness ($b < 0$). These genetic parasites can only persist in the population at very high
123 rates of HGT, but are of course never beneficial for the population growth rate. **Figure 2C**
124 shows a bifurcation diagram that summarises how increasing or decreasing rates of HGT impact
125 the population growth rate for these different classes.

126

127 **HGT is an evolutionarily stable strategy, but cannot evolve to ‘rescue’ res-**
 128 **cuable genes**

129 By analysing the simple model of cells undergoing HGT, we have found 5 distinct gene classes.
 130 For two of these classes, namely enrichable and rescuable genes, moderate rates of HGT improve
 131 the population growth rates. We next study (i) whether HGT of enrichable and rescuable genes is
 132 an evolutionarily stable strategy, and (ii) if bacteria can evolve this strategy *de novo*. To answer
 133 these questions, we consider two competing species: one with that does engage in HGT, and does
 134 not (HGT^+ and HGT^- respectively, see **Figure 1B**). With this model, we have studied the
 135 evolution of HGT by means of adaptive dynamics (Metz et al., 1995). If HGT^- cannot invade
 136 HGT^+ , we call HGT an evolutionarily stable strategy, and if HGT^+ can invade HGT^- we call
 137 HGT evolvable.

138 We found that HGT is an evolutionarily stable strategy for both enrichable and rescuable
 139 genes, but that HGT is evolvable only for enrichable genes (see Supplementary Material for
 140 full analysis). Even when we assume that the invading HGT^+ -mutant has the optimal rate
 141 of HGT, it cannot invade into a population of HGT^- cells in steady state. These results were
 142 confirmed by numerical analysis, which indeed shows that HGT^+ only invades when the founding
 143 population size of HGT^+ (C^+/N^+) is relatively large (see **Figure 3A**). This failure to reach
 144 the alternative (fitter) evolutionary attractor is caused by positive frequency-dependent selection
 145 (known as the Allee effect). Invading mutants, *i.e.* a small population of HGT^+ cells, contain
 146 few carrier cells to act as donors for HGT. Moreover, since the resident population of HGT^- is
 147 also not able to retain the rescuable genes, the resident population can also not serve as a donor
 148 (see **Figure 3B**). As such, the costs of HGT for an invading HGT^+ -mutant do not outweigh
 149 the potential benefits. In summary, while HGT is an evolutionarily stable strategy, cells cannot
 150 evolve HGT to ‘rescue’ rescuable genes.

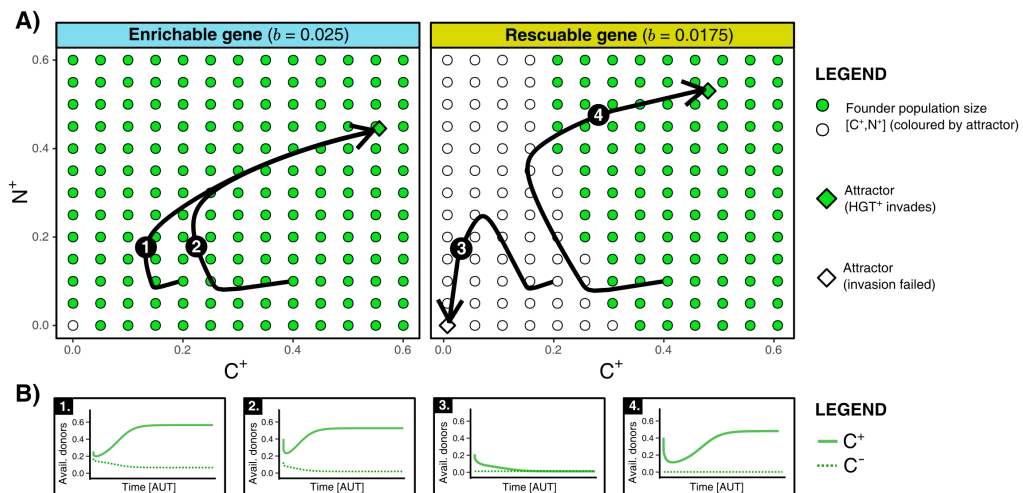


Figure 3: HGT is an evolutionarily stable strategy, but is evolutionarily inaccessible for rescuable genes due to a lack of gene-carrying donor cells. A) For an enrichable and a rescuable gene ($b = 0.025$ and $b = 0.0175$ respectively), a 2D projection of the 4D state space is shown. For various founder sizes (combinations of C^+ and N^+), the result of invasion of HGT^+ into HGT^- is shown. HGT^+ always successfully invades for enrichable genes. For a rescuable gene, low founder populations sizes of C^+ and N^+ (white dots) fail to invade, whereas they can invade at higher population sizes (green dots). Black arrows (1-4) show the trajectories starting from two founder population sizes. B) for the four trajectories from A, the graphs show the temporal dynamics of gene-carrying donor cells.

151 Spatial structure hinders the maintenance of genes, making HGT adaptive for 152 a wider range of genes

153 So far, we have studied a well-mixed population of cells that undergoes all-against-all competition,
154 and found that HGT is advantageous for slightly beneficial genes that (i) are not too beneficial,
155 as these genes readily persist within the population without HGT, and (ii) are beneficial enough
156 to compensate for the costly HGT. Next, we study the same dynamics of carrier and non-carrier
157 cells in a spatially explicit, eco-evolutionary context. We do this by implementing an individual-
158 based model (IBM), where bacterial cells reside on a grid, interactions are local, and events
159 like HGT and gene loss are implemented as stochastic processes (see Methods and **Figure 1C**).
160 When the cells on this grid are sufficiently mixed each time step, the IBM should approximate the
161 dynamics of the ODE model. However, when cellular mixing is minimal, the resulting spatially
162 structured population is more analogous to that of a biofilm. What is the effect of this spatial
163 structure?

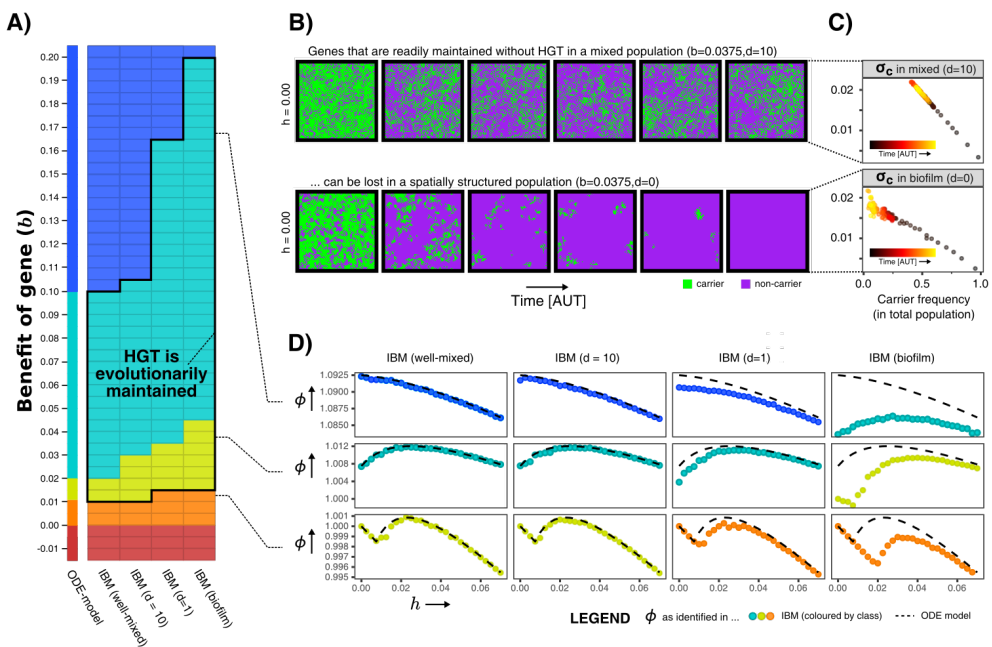


Figure 4: Spatial structure hinders the maintenance of slightly beneficial genes **A)** Each tile in this table represents a series of simulations in which we first test which gene class (background colour) is found when sweeping over different HGT-rates (h -values), and next test whether the observed optimal rate of HGT (h_{opt}) is evolutionarily maintained when starting with a population consisting of only carrier cells (shown with black outline). This was tested for the well-mixed IBM and the IBM with different levels of mixing (d). The continuum of gene classes from the ODE-model is presented for comparison. Colours are the same as in **Figure 2** (blue=indispensable, cyan=enrichable, yellow=rescuable, orange=unrescuable, red=SGE). **B)** Shown is the spatial grid of the IBM for two simulations with the same value of b , and no HGT. The gene readily persists in the mixed IBM (top panel, $d = 10$), while the gene does not persist in the spatially structured population (bottom panel, $d = 0$). **C)** For the simulations shown in B, the average competitive advantage of carrier cells with respect to their local competitors (σ_c , see Methods) is plotted against the frequency of carrier cells, showing how clumping hinders the effective benefit of carrying a gene. **D)** For 3 rows from the table of A, it is illustrated how the effect of spatial clumping illustrated in B and C modifies the gene class found for specific b -values. The dashed line indicates the growth rates predicted by the ODE model.

164 We first analysed the IBM for a wide variety of values for b and h , and measured the average

165 growth rates ϕ in the population. We can thus evaluate whether the aforementioned gene classes
166 (indispensable, enrichable, rescuable, unrescuable genes, and SGEs) are found under the same
167 conditions as in the ODE model. **Figure 4A** shows that, when the IBM is well-mixed, the gene
168 classes indeed occur at values of b identical to the ODE model. However, the gene classes shifts
169 to higher values of b when mixing is decreased, making the range of benefits which are classified
170 as enrichable and rescuable much broader. In these biofilm populations, HGT was indeed found
171 to be evolutionarily stable for this wider range of fitness-effects (black outline), illustrating that
172 it is not only the value of b , but also the ecological context in which a gene finds itself that
173 determines whether or not HGT is adaptive.

174 What causes these gene classes to shift depending on this spatial context? How does an enrichable
175 gene in the well-mixed system become rescuable in the spatially structured population, as though
176 it is less beneficial? **Figure 4B** shows how this can be intuitively understood by taking into
177 account how individuals in a spatial system mostly compete with their own kind (*i.e.* progeny and
178 conspecifics). Even when the majority of the population consists of non-carriers, carriers are still
179 competing mostly with other carrier cells. Thus, the effective benefit of carrying the gene is lower
180 in a biofilm, hence the gene becomes harder to maintain within the population. In **Figure 4C**
181 is shown that, while carrier cells in the well-mixed populations experience a competitive advantage
182 of $\sim 2\%$ when carriers make up approximately half the population, carriers in a biofilm only
183 reach a similar competitive advantage at very low carrier frequencies, *i.e.* when the carriers are
184 almost extinct. At this point, the gene will readily be lost stochastically. The hampered ability
185 of spatially structured population to retain slightly beneficial genes, indeed changes how the
186 population growth rate depends on the rate of HGT (**Figure 4D**).

187 HGT evolves for rescuable genes only in a spatially structured population

188 The results described in the previous section illustrate that HGT is an evolutionarily stable
189 strategy for a much broader range of b -values (fitness effects of genes) in a spatially structured
190 population than in a well-mixed culture. Many more genes are furthermore classified as rescuable
191 in these spatially structured populations, meaning that they can only persist through HGT. We
192 have concluded in the previous section that HGT cannot evolve to ‘rescue’ these rescuable genes
193 in populations that are well-mixed, fully deterministic, and by only considering a single HGT^+
194 mutant type at a time. In the IBM on the other hand, the population is not spatially structured,
195 events are stochastic, and each individual cell has its own rate of HGT. Can these different
196 assumptions help to alleviate the Allee effect mediated by a lack of donor cells, which prevents
197 the evolution of HGT?

198 To answer the question posed above, we allowed the HGT-rate (h) of all individuals in the IBM
199 to evolve (see Methods). When a non-carrier interacts with a (local) carrier, the h -value of this
200 non-carrier (*i.e.* the acceptor) determines the probability of accepting the gene. For simplicity,
201 we will call individuals with an h -parameter greater than 0.02 HGT^+ , and the others HGT^- .
202 We start with a non-carrier population of HGT^- cells (with $h = 0.00$), simulate this population
203 for some time (20,000 time steps), and then allow cells to sporadically discover rescuable genes.
204 Since rescuable genes cannot persist without HGT, the fate of this gene depends on the ability
205 of cells to engage in (local) HGT. Using this protocol, we investigate if the rescuable gene is
206 able to spread through the evolution of HGT. We found that HGT never evolved for rescuable
207 genes in well-mixed populations (**Figure 5A**), consistent with our prior results in the well-mixed
208 ODE model. Thus, we can conclude that the level of stochasticity in the IBM is insufficient to
209 overcome the aforementioned Allee effect caused by a lack of donor cells.

210 In the spatially structured population, HGT of rescuable genes *does* in fact evolve, therewith
211 ‘rescuing’ the rescuable genes (**Figure 5B**). Interestingly however, we found that HGT did
212 not always evolve immediately after the influx of rescuable genes started (yellow arrow), but
213 nevertheless spread steadily once attained. To further elucidate the spread of genes, we barcoded
214 each newly discovered gene with a unique ID, and visualised these on the spatial grid with
215 different colours (**Figure 5C**). Initially, rescuable genes fail to invade, even though different
216 barcodes may locally persist for a while (episode I). After some time however, one gene (green)
217 manages to persist within a local community of transferring cells (episode II). This sets in motion
218 a positive feedback mechanism, where the local abundance of the green gene alleviates the lack of
219 donor cells, transforming nearby HGT^+ -mutants into carriers, and so on (also see Supplementary
220 Movie). This emergent ‘gene-sharing’ community eventually overgrows the other cells, and the
221 rescuable gene ultimately persists in up to $\sim 70\%$ of the population. After the influx of rescuable
222 gene is stopped (episode III), the gene readily persists within the population, showing how this
223 transferring community does not depend on the continuous influx of genes. In summary, HGT
224 of rescuable genes can only evolve if transfer happens within spatially localised sub-populations,
225 and not under well-mixed conditions modelled by mass-action. Through a local ‘nucleation
226 event’, communities can reach the alternative stable state that can maintain the rescuable gene.
227 **Figure 5D** summarises the outcome of HGT evolution for a broad range of genes (b -values)
228 with different levels of mixing, revealing how HGT evolves for many more genes in a spatially
229 structured population. Moreover, while HGT of enrichable genes always evolved, HGT only
230 evolved for rescuable genes in spatially structured populations. Finally, as expected from prior
231 results, HGT never evolved for indispensable and unrescuable genes.

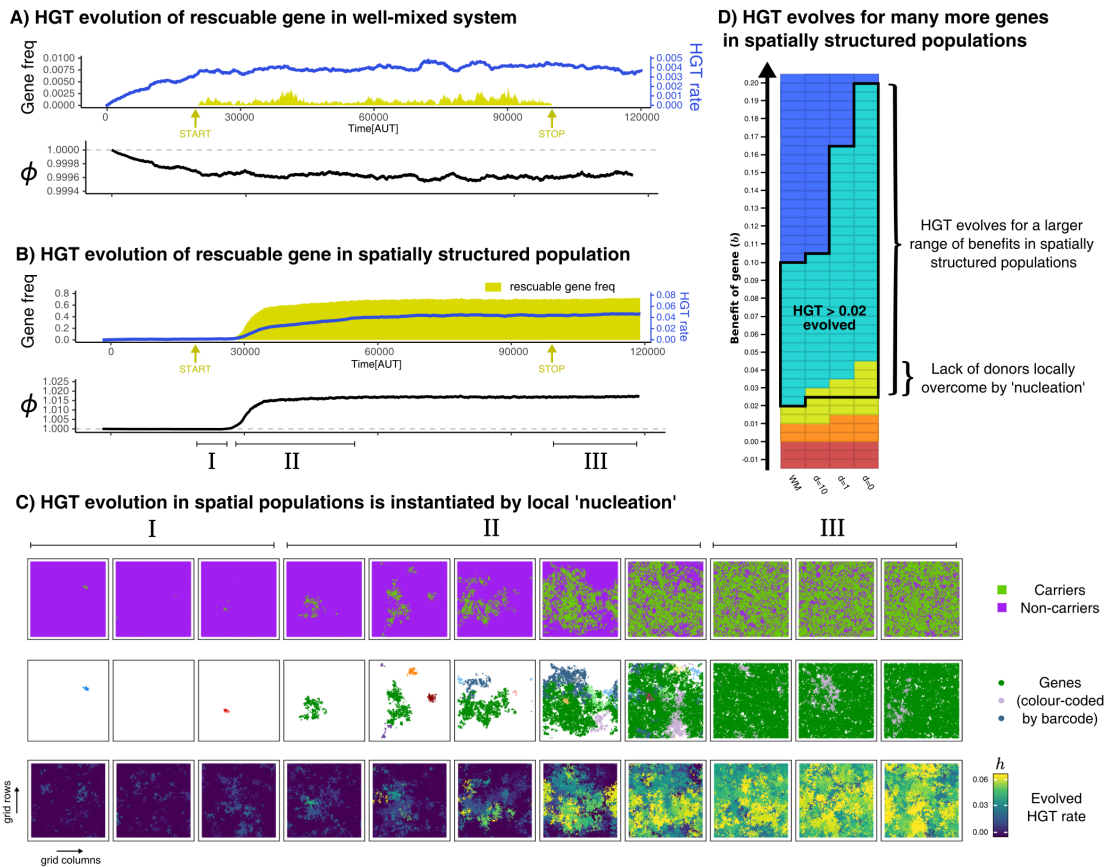


Figure 5: HGT of rescuable genes only evolves in spatially structured populations due to the emergence of ‘gene-sharing’ communities: Panel A and B both show the frequency of a rescuable gene (yellow area) that is discovered with a very low probability ($5 \cdot 10^{-6}$ per time step), the mean evolved HGT rate of the population (blue line), and the growth rate of the population (black). Note that A and B have a different range in the y-axis for clarity. C shows how in the spatially structured populations, carrier cells with a rescuable gene (colour coded by the unique barcodes) spread after a local ‘nucleation event’. A positive feedback loop follows, resulting in a ‘gene-sharing’ community which slowly overgrows the rest of the population. D shows the outcome of HGT evolution for the same combinations of fitness-effects and mixing as in Figure 4A. Parameters used: $h_{init} = 0.0$, $u = 5e - 3$, $m = 0.05$, $l = 0.02$, $c = 0.1$, $f = 5 \cdot 10^{-6}$, $f_{start} = 20.000$, $f_{stop} = 100.000$, $n = 400$ (*i.e.* $N = 400^2$). For the well-mixed population, we chose a rescuable gene with the highest benefit from **Figure 4A** ($b = 0.0175$), and for the spatially structured population we used $b = 0.030$ (the average of the much broader range of rescuable genes). Both these genes require HGT to persist, and are thus rescuable within their own spatial context.

232 HGT is evolutionarily maintained in the presence of harmful SGEs

233 We have shown that HGT can be adaptive and evolvable for bacteria in order to enrich or rescue
234 slightly beneficial genes. We next investigated if HGT can be maintained under the pressure of
235 harmful SGEs, genetic parasites that spread through horizontal transfer. For this, we consider a
236 population that evolved HGT of a rescuable gene ($b=0.03$), and expose this population to a low
237 influx of SGEs which confer a fitness penalty (β). We study if these SGEs, despite their fitness
238 penalty, can persist within this bacterial population, and if HGT is evolutionarily maintained
239 by the hosts. **Figure 6A** shows that, when the fitness penalty of the SGEs is small relative
240 to the benefit of the rescuable gene (hereafter called “weak SGEs”, $\beta = 0.01$), these genetic
241 parasites quickly rise to very high frequencies within the population. Although the host cells
242 gradually evolve lower HGT rates in response (from $h \pm 0.05$ it stabilises around $h \pm 0.04$, also
243 see **Supplementary Figure S2C**), HGT, the rescuable gene, and the SGEs are evolutionarily
244 maintained. When the influx of SGEs is stopped, the cells (and their beneficial gene) stably
245 coexists with these genetic parasites.

246 Strikingly, if we introduce SGEs whose fitness penalty is greater than the benefit of the gene
247 (“strong SGEs”, $\beta = 0.04$), we also observe the coexistence of cells, rescuable genes, and SGEs.
248 By looking at the initial invasion dynamics (**Figure 6B**), we can see that these strong SGEs
249 cannot rise to very high frequencies. As the hosts evolve lower rates of HGT, these genetic
250 parasites are pushed to very low frequencies. However, the reduced threat of genetic parasites
251 causes the host cells to once again increase their rates of HGT, leading to a secondary outbreak of
252 SGEs (**Figure 6B**, from $T=300,000$ onwards). It is interesting to note that, while the population
253 growth rates (ϕ_{pop}) clearly decrease due to this second infection, the growth rates along the line-
254 of-descent (ϕ_{lod} , see methods) remains largely unaffected. Thus, while a sub-set of the population
255 has been infected, individuals in this infected strain will not be amongst the long-term ancestors.
256 Counter-intuitively, strong SGEs only have a minor impact on bacterial growth rates, while
257 weaker SGEs impose a significant burden on the population by rising to much higher frequencies
258 (also see **Supplementary Figure S2C**). Finally, stopping the influx of SGEs does not impact
259 the long-term coexistence of cells, beneficial genes, and these strong SGEs (**Figure 6C** and
260 **Supplementary Figure S2C**).

261 To better understand the co-evolutionary process between SGEs and bacteria engaging in HGT
262 of rescuable genes, **Figure 6D** shows long-term dynamics of barcoded SGEs in this spatial
263 system. Although a diverse set of SGEs are initially discovered in parallel (coloured by their
264 unique barcode), eventually only a single barcode remains after the influx of SGEs is stopped.
265 Moreover, it can also be seen how SGEs are either locally abundant, or entirely absent. Thus,
266 spatially separated strains of bacteria experience opposing selection pressures for HGT. Lower
267 rates of HGT are favoured in the presence of these strong SGEs, but higher rates of HGT are
268 favoured when these genetic parasites have (locally) died out. Indeed, this heterogeneity of
269 SGEs is crucial for the strong SGEs to persist, as well-mixed populations can only retain weaker
270 SGEs (see **Supplementary Figure S2**). Interestingly, we also found that strong SGEs failed
271 to persist when HGT was too localised (*e.g.* only between neighbouring cells), as the SGEs then
272 could not escape to a new pool of hosts that have high rates of HGT (**Supplementary Figure**
273 **S3**). We conclude that, in a spatially structured population, strong SGEs can stably coexist in
274 a bacterial population which maintains HGT to ‘rescue’ rescuable genes.

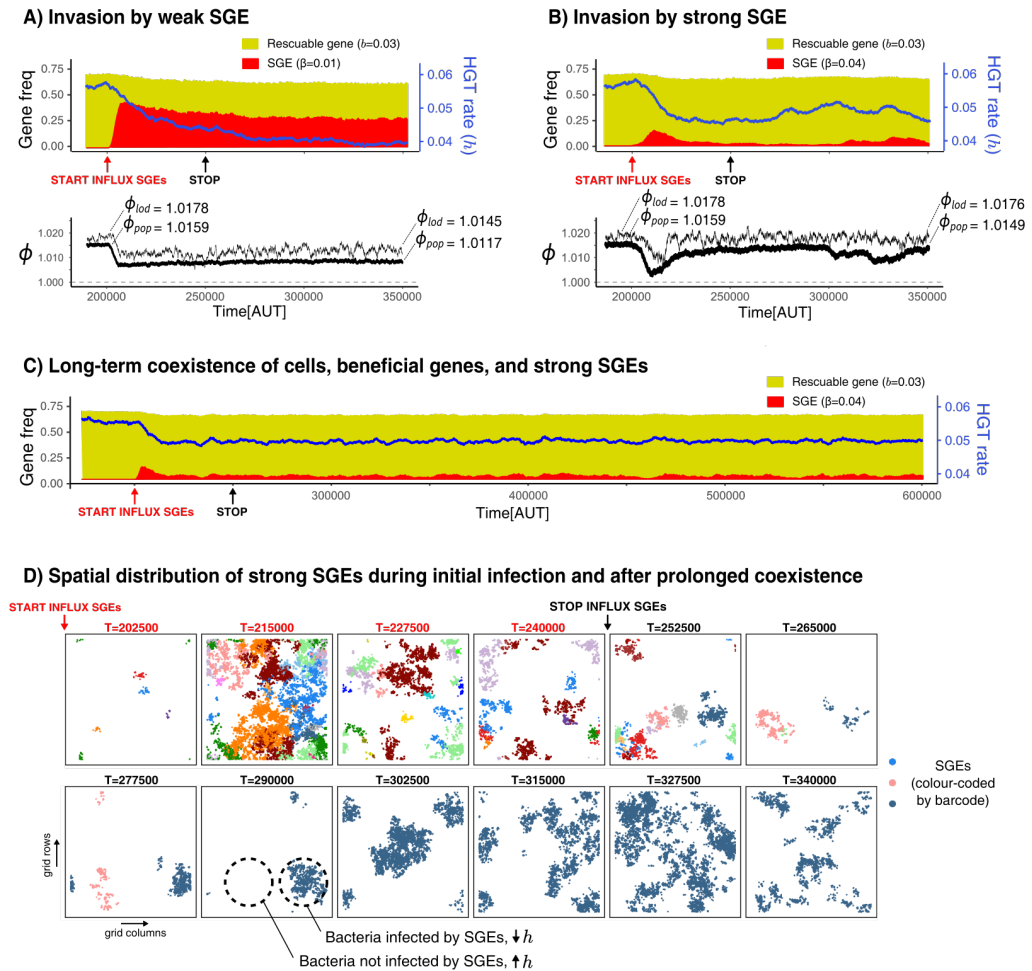


Figure 6: SGEs can invade and stably coexist with their host cell: A and B show the temporal dynamics for a population that has evolved to rescue a slightly beneficial gene ($b = 0.03$) invaded by a weak SGE (A, $\beta = 0.01$) and a strong SGE (B, $\beta = 0.04$) respectively. The blue line indicates the rate of HGT as evolved by the host cells. In the bottom graphs, the thick black line is the average growth rate of the population (ϕ_{pop}), and the thin black line is ϕ_{lod} , the average growth rate along the line of descent (250-1 generations ago). The ϕ_{pop} and ϕ_{lod} that are annotated with the dashed lines are the average of the first/final 200 generations. Panel C shows the long-term coexistence of cells, beneficial genes, and strong SGEs ($\beta = 0.04$). D shows the spatial distribution of SGEs (coloured by their unique barcodes). The top row shows this during the invasion (open ecosystem) and the bottom row shows this during prolonged coexistence (closed ecosystem). Note that the empty sites (white) only indicate the absence of SGEs, not of bacterial cells, which are instead present in every grid point. Parameters used: h -parameters and frequency of carriers as evolved from Figure 5, $u = 5e - 3$, $m = 0.05$, $l = 0.02$, $c = 0.1$, $i = 1e - 5$, $i_{start} = 200.000$, $i_{stop} = 250.000$, $n = 400$ (i.e. $N=400^2$).

275 Discussion

276 We have studied the balance between the advantages and disadvantages of HGT by modelling
277 transfer of a single gene within a simple bacterial population. Our analysis shows that we can
278 categorise slightly beneficial genes based on whether genes are lost from the population without
279 HGT, and whether HGT of these genes can improve the population growth rate. This results in
280 five distinct gene classes: (i) *indispensable genes*, that readily persist within the population and
281 for which HGT is therefore always deleterious, (ii) *enrichable genes* which are not lost from the
282 population without HGT, but moderate rates of HGT are adaptive, (iii) *rescuable genes* which are
283 lost from the population without HGT, but can be rescued by HGT which improves population
284 growth rates, and (iv) *unrescuable genes*, that are also lost from the population without HGT,
285 but recovering them with HGT does not improve population growth rates, and (v) *selfish genetic*
286 *elements*, genes that confer a fitness penalty, but can persist within the population with HGT. We
287 further investigated if HGT of these genes is an evolutionarily stable strategy, and if HGT of these
288 genes can evolve *de novo*. We found that horizontal transfer of enrichable and rescuable genes
289 is indeed a evolutionarily stable strategy, but can only evolve from scratch for enrichable genes.
290 The evolution of HGT to ‘rescue’ a rescuable gene faces a problem under well-mixed conditions:
291 HGT is required for the gene to persist, but sufficient carriers of the gene are necessary to evolve
292 HGT. By modelling this process in a spatially structured population, we show that HGT can
293 nevertheless evolve for these rescuable genes. As carriers of the gene can be locally abundant,
294 emergent communities form that locally retain the gene via HGT, therewith slowly outgrowing
295 other individuals. Finally we show that once stable transferring communities have evolved, selfish
296 genetic elements (SGEs) can stably coexist with the bacterial population and the beneficial genes.
297 In spite of these genetic parasites exploiting the host’s ability to transfer, HGT is evolutionarily
298 maintained, providing a doorway to the co-evolutionary process between bacteria and SGEs.

299 Our model reveals that HGT can be adaptive when considering genes with a fitness benefit
300 that does not sufficiently compensate for the rate of gene loss. While we studied this from the
301 perspective of genes that confer a constant fitness effect (*i.e.* a constant environment), bacteria
302 in natural microbial ecosystems frequently experience changing selection pressures. For example,
303 some genes confer a large fitness benefit under rare ecological circumstances, *e.g.* antimicrobial
304 resistance, toxin production, or cooperation (Riley and Wertz, 2002; Cordero et al., 2012; Vogwill
305 and MacLean, 2015; Gerardin et al., 2016; Hehemann et al., 2016; Dimitriu et al., 2019). However,
306 in between such rare opportunities, these traits are not beneficial or even costly. Although these
307 changing selection pressures have been used to explain how certain traits require HGT to persist
308 (Nogueira et al., 2009; Rankin et al., 2011; van Dijk and Hogeweg, 2015), our model shows that
309 a similar argument can be made under constant selection pressure, as long as the fitness effect of
310 the genes is small. In other words, our work shows how bacteria may benefit from preferentially
311 mobilising genes that are either constantly, or on average, slightly beneficial.

312 Besides investigating the impact of HGT for a range of different fitness effects, we have also shown
313 how spatial structure is a key component for the emergence of HGT of rescuable genes. Both
314 conjugation and transformation have indeed been observed to occur more frequently in biofilms
315 than in well-mixed cultures (Madsen et al., 2012), and plasmids have furthermore been shown
316 to be more persistent in biofilms (Stalder et al., 2020). On the other hand, spatial structure
317 can slow down adaptation in asexual populations because individuals are mostly competing with
318 their related conspecifics (Gordo and Campos, 2006; Habets et al., 2007; Chacón and Harcombe,
319 2019). Relatedness has indeed been shown to be an important factor in stabilising HGT, for
320 example of cooperative traits (Mc Ginty et al., 2011). Our model shows that, also without
321 explicitly taking cooperation into account, HGT can only evolve in ‘gene-sharing’ communities
322 which emerge in by local reproduction in spatially structured populations. Thus, not only are
323 relatedness and spatial structure necessarily intertwined, they are crucial for the rare ‘nucleation

324 events' that initiates evolution towards increased rates of HGT. Intriguingly, similar nucleation
325 events have been observed in origin of life studies (Wu and Higgs, 2012) and models of microbial
326 antagonistic interactions (Kotil and Vetsigian, 2018). These types of emergent evolutionary
327 transitions highlight how studying evolution under well-mixed conditions, and one mutant at
328 a time, can be highly misleading. Studying biological systems in a spatial context will help
329 us to better understand which eco-evolutionary outcomes are accessible, and maintainable, by
330 evolution.

331 **Horizontal Gene Transfer: rescue or catastrophe?**

332 In nature, HGT can happen through a variety of mechanisms that each have their own potential
333 advantages and disadvantages for the host cell (Vogan and Higgs, 2011; Baltrus, 2013). Bacteria
334 do not always have full control over the rates at which HGT happens, especially when considering
335 it as a side-effects of other processes (Redfield, 2001). However, it remains an intriguing question
336 under which specific circumstances bacteria benefit from HGT, whether it is a side-effect or not.
337 By abstracting away from the different mechanisms of HGT, and what it means for a gene to
338 be "beneficial", we have revealed the conditions under which HGT is an adaptive trait for the
339 host cells. In a similar spirit, earlier modelling by Vogan & Higgs has shown that HGT can
340 be adaptive with respect to genes that are frequently lost (Vogan and Higgs, 2011). However,
341 in their work, natural selection eventually favoured improved replication accuracy, therewith
342 decreasing the advantage of HGT. Other models have shown that HGT is beneficial to mitigate
343 the effects of Mullers Ratchet (Muller, 1964) by decreasing assortment load (Takeuchi et al., 2014;
344 Vig-Milkovics et al., 2019), analogous to the impact of sex and recombination on the balance
345 between drift and selection (Lynch et al., 1995; Schultz and Lynch, 1997; Lynch et al., 2016; Vos
346 et al., 2019). Our work complements these aforementioned studies by showing that, however
347 low the rate of gene loss may be, there may always be a class of slightly beneficial traits for
348 which HGT is adaptive and evolvable. Although genes with such small fitness effects are very
349 hard to detect experimentally (Bataillon, 2000; Wisner and Lenski, 2015), our model is a proof of
350 principle that HGT may play a key role in preventing the loss of these genes, which may explain
351 the differential rates of HGT as observed in the data (Nogueira et al., 2009; Rankin et al.,
352 2011; Madsen et al., 2012; Novick and Doolittle, 2020). With the upswing and improvement of
353 experimental techniques like Hi-C metagenomics (Beitel et al., 2014; Burton et al., 2014) and
354 DNA barcoding (Blundell and Levy, 2014; Ba et al., 2019), we will soon have more insights into
355 the eco-evolutionary dynamics of small-effect mutations (Li et al., 2018; Lerner et al., 2019) and
356 accessory genes (Quistad et al., 2019; Yaffe and Relman, 2020), and we may learn when HGT can
357 come to rescue a microbial population, and when it may be nothing more than a catastrophe.

358 Methods

359 General overview

360 In this work, we study the dynamics of bacteria undergoing HGT of slightly beneficial genes and
 361 Selfish Genetic Elements (SGEs). We do this by modelling the same processes with gradually
 362 increasing complexity, starting from simple Ordinary Differential Equations (ODEs), and then
 363 evaluating the same dynamics in an Individual-based Model (IBM). A graphical representation
 364 of these models is found in the main text (**Figure 1**). The models consider the competition
 365 between cells of two types: carrier cells (C) that carry a gene, and non-carrier cells (N). When
 366 carrier cells contain a beneficial gene (*i.e.* it is a beneficial trait), they grow faster than the
 367 non-carrier cells (N). However, carriers may lose this beneficial gene with a fixed rate l . Both
 368 cell types engage in HGT with rate h , which comes with a cost c . This cost is equal for both
 369 cell types, meaning that whatever the costs may entail, we assume they are continuously payed.
 370 Proportional to the density of available carrier cells, non-carriers can be transformed back into
 371 a carrier cell by means of “additive” HGT. Both models use a chemostat assumption, where cells
 372 wash out at a rate proportional to the rate of growth, ensuring a constant population size in
 373 steady state.

374 ODE model(s)

375 By modeling the dynamics described above by means of ODEs, we assume a well-mixed popula-
 376 tion of cells that compete according to all-against-all dynamics (*i.e.* mass-action). Our equations
 377 describing the density of carrier (C) and non-carrier (N) cells are given in **Equation 2**, where
 378 b is the benefit of the carried gene (or burden if $b < 0$), l is the rate of gene loss, h is the rate at
 379 which cells engage in HGT, c is the continuous cost for engaging in HGT, and HGT transforms
 380 a non-carrier into a carrier when they interact (hCN). This cost for HGT (c) is equal for both
 381 cell types, meaning that whatever the costs may entail, we assume they are continuously payed.
 382 Finally, the total amount of growth (ϕ) is subtracted from both populations, meaning that the
 383 population density in steady state is always 1.

$$\begin{aligned}
 \frac{dC}{dt} &= \underbrace{(1 - ch + b)C}_{\text{reproduction of C}} - \underbrace{lC}_{\text{gene loss}} + \underbrace{hCN}_{\text{HGT}} - \underbrace{\phi C}_{\text{chemostat}} \\
 \frac{dN}{dt} &= \underbrace{(1 - ch)N}_{\text{reproduction of N}} + \underbrace{lC}_{\text{gene loss}} - \underbrace{hCN}_{\text{HGT}} - \underbrace{\phi N}_{\text{chemostat}} \\
 \phi &= \underbrace{(1 - ch + b)C}_{\text{total growth of C}} + \underbrace{(1 - ch)N}_{\text{total growth of N}}
 \end{aligned}
 \tag{2}$$

$C + N = 1$ (constant population size, ensured by chemostat assumption.)

384 From the above model, we derived how the population growth rate (ϕ) depends on both b and
 385 h (see **Equation 1** in the main text), which shows the conditions under which HGT improves
 386 the total growth rate of the population. To analyse whether or not HGT could evolve, we
 387 extended the two-variable ODE model above (of cells with the same h) to a four-variable ODE
 388 model (of two species with a different h , see **Figure 1B** and **Equation 3** below). We use this
 389 extension to study whether or not a species with HGT (C^+ and N^+ , $h > 0$) could invade upon
 390 a species without HGT (C^- and N^- , $h = 0$), and *vice versa* (see Supplementary material for
 391 full analysis). Finally, we also extended the ODE model to study the impact on growth rates for
 392 cells that engage in HGT of both a beneficial gene and a Selfish Genetic Element (SGE), which
 393 can be found in the Supplementary Material.

$$\begin{aligned}\frac{dC^-}{dt} &= (1+b)C^- - lC^- - \phi C^- \\ \frac{dN^-}{dt} &= N^- + lC^- - \phi N^- \\ \frac{dC^+}{dt} &= (1+b-ch)C^+ - lC^+ + hN^+(C^- + C^+) - \phi C^+ \\ \frac{dN^+}{dt} &= (1-ch)N^+ + lC^+ - hN^+(C^- + C^+) - \phi N^+ \\ \phi &= (1+b)C^- + N^- + (1+b-ch)C^+ + (1-ch)N^+\end{aligned}\tag{3}$$

394 Individual-based model

395 The individual-based model (IBM) describes the same dynamics as the ODE models, but differs
396 in some important aspects. Firstly, individuals are discrete entities that live on a 2D grid, and
397 reproduce locally. This allows us to study the model with and without spatial pattern formation
398 by modifying the rate at which cells mix. When mixing is disabled or very limited, a spatially
399 structured population like that of a biofilm will form, while an increased amount of cellular
400 mixing will approximate a well-mixed culture. Under well-mixed conditions, individuals will
401 interact with random individuals in the population (approximating the all-against-all dynamics
402 of the ODEs), while individuals will interact mostly with their conspecifics in case of the biofilm.
403 We explicitly define a competition range (focal cell plus its 8 neighbouring grid points) and a
404 HGT range (all cells within distance t) which determine smaller samples of the total population
405 with which individuals can interact. Each individual (potentially) has its own h -parameter,
406 allowing us to study the evolution of HGT in an eco-evolutionary context (see implementation
407 of mutations below). As we primarily focus on the question if cells benefit from taking up
408 genes from their environment or other cells, we assume that the h -parameter of the acceptor
409 cell determines the probability of HGT. The IBM also includes a low rate (f) at which genes
410 with benefit b^* are (re)discovered, allowing us to study how and if newly discovered genes /
411 selfish elements spread through the population. Finally, note that processes such as gene loss,
412 HGT, and competition are no longer deterministic like in the ODEs, but implemented as events
413 that can stochastically happen at each simulated time step. To ensure the chance-events in
414 the IBM (reproduction, HGT, gene loss) accurately represent the rates as used in the ODE, all
415 probabilities were multiplied by a small constant $\Delta T=0.1$.

416 **Updating grid points:** All grid points i, j in the IBM contain a single cell which can be
417 a carrier or non-carrier ($b_{i,j} = b$ for carriers, $b_{i,j} = 0$ for non-carrier), which can carry a SGE
418 ($\beta_{i,j} = \beta$ for SGE infected cells, $\beta_{i,j} = 0$ for uninfected cells), and have an individual HGT-
419 parameter $h_{i,j}$. At each time step, local reproduction happens in each grid point i, j by drawing
420 a random individual from the Moore (9 cells) neighbourhood and letting it reproduce with a
421 probability proportional to its growth-rate $\varphi_{i,j}$:

$$\varphi_{i,j} = 1 + b_{i,j} - \beta_{i,j} - ch_{i,j}\tag{4}$$

422 When reproduction happens, the winner cell replicates and replaces the cell in grid point i, j .
423 This newborn cell is an exact copy of the mother cell. Next, all cells are also updated to include
424 the processes of stochastic gene loss with rate l , HGT with rate $h_{i,j}$, and gene/SGE discoveries

425 based on the influx-rate f . Finally, with a small probability u , the HGT rate of any individual
426 can mutate, where a cell uniformly samples a new parameter between $h_{i,j} - m$ and $h_{i,j} + m$.

427 **IBM growth rates:** With respect to growth rates, the simulated IBM model does not only
428 track the average growth rate of all cells in the population (ϕ_{pop}), but also tracks the growth rate
429 of the line of descent that gave rise to the current population (ϕ_{lod}). While ϕ_{pop} is comparable
430 to ϕ in the ODE-model, ϕ_{lod} gives us insights into how the long-term ancestors are impacted by
431 HGT. We also measure the competitive advantage that carrier cells have over non-carriers when
432 competing (locally) for reproduction (σ_c), which is defined as the average fitness advantage a
433 carrier has over its local competitors (8 neighbouring sites). When all competitors of a carrier
434 are carriers, σ_c approaches 0. When all competitors are non-carriers, σ_c approaches b .

435 **IBM barcoding:** We tag all influxed genes and SGEs with a unique identifier, allowing us
436 to visualise how genes / SGEs spread through the population (analogous to DNA barcoding
437 (Blundell and Levy, 2014; Levy et al., 2015; Ba et al., 2019; Lerner et al., 2019)). These barcodes
438 also allow us to investigate whether or not these genes are continuously rediscovered, or form
439 long lineages of genes that persist within the population.

440 **Parameters used:**

441 Throughout most of this study, the gene loss l was set to 0.02 and the cost for HGT was set to
442 $c = 0.2$. In general, our results do not depend on the precise value of these two parameters. For
443 example, when the rate of gene loss is set to much lower (arguably more realistic) values, the gene
444 classes discussed in **Figure 2** simply shift to lower values of b . Similarly, if the costs are higher,
445 the parameter-region where HGT is adaptive for the host cells (*i.e.* enrichable and rescuable
446 genes) is more narrow, but is still retained. Parameters such as the benefit (b), the HGT-rate
447 (h), the amount of mixing (d), and the HGT distance (t) have been extensively swept, as
448 discussed in the main text / Supplementary Material. In these cases, the used parameters are
449 given in the captions of the relevant figures. When comparing the IBM with the ODE models
450 (*e.g.* occurrence of gene classes), evolution of h was disabled ($u = 0.0$). For the *de novo* evolution
451 of HGT, the initial population consisted only of *non-carrier* cells, but genes fluxed in at a low
452 rate ($f = 5 \cdot 10e - 6$), while the initial level of HGT ($h = 0.0$) was allowed to evolve with
453 $u = 5 \cdot 10e - 5$ with a uniform step size of $m = 0.05$. Finally, when testing whether HGT could
454 be maintained, no influx of genes was present ($f = 0.0$), but the initial population consisted of
455 carrier-cells that, at least initially, all have the optimal rate of hgt ($h = h_{opt}$, see supplementary
456 material). All experiments in the IBM with Selfish Genetic Elements were done with slightly
457 lower costs ($c = 0.1$), to compensate for the extra costs imposed by these genetic parasites.
458 All the important parameters of our models are summarised in **Table 1**.

459 **Software used**

460 The analytical model was numerically analysed using grind.R by R.J. de Boer (<http://tbb.bio.uu.nl/rdb>), a R script that uses the deSolve R-package (Soetaert et al., 2010). The simulated
461 model was implemented in Cash (Cellular Automaton simulated hardware) version 2.1, an free
462 and easy-to-use library to make simple spatially explicit simulations (originally created by R.J. de
463 Boer & A.D. Staritsk, further developed by Nobuto Takeuchi and Bram van Dijk). Visualisation
464 of both models was done in R using ggplot (Wickham, 2016) and plotly (Inc., 2015). Simulations
465 were run in Linux Ubuntu 16.04 LTS using GNU parallel(Job).

467 Both the R-scripts for ODE analysis and the IBM code implemented in C, are available online
468 https://github.com/bramvandijk88/HGT_Genes_And_SGEs.

Table 1: Description of parameters used in the models

Parameter (general)	Description
Gene loss (l)	Rate at which carrier cells lose the beneficial gene
HGT rate (h)	Rate at which non-carriers are transformed into carriers (when interacting with carrier cells)
Benefit of gene (b)	Growth rate benefit for carrier cells (or penalty for negative b)
Costs of HGT (c)	Growth rate penalty for the rate of HGT
Parameter (IBM only)	Description
Grid size (n)	The simulation is done on a square grid of $n \times n$ cells
Mixing rate (d)	Every time step, the grid is mixed d times using the Margolus Diffusion algorithm (Toffoli and Margolus, 1987). Alternatively, the population was <i>well-mixed</i> by assigning new positions at random every time step.
Competition range (s)	Sub-population of $s \times s$ cells surrounding focal grid point that compete for reproduction
HGT distance (t)	Sub-population of $d \times d$ cells surrounding focal grid point from which a random potential donor is sampled for HGT
Influx genes (f)	A small probability for any cell to discover a gene <i>de novo</i>
Benefit of influxed gene (b^*)	Growth rate benefit for carrier cells (or penalty for negative b)
Influx SGEs (i)	A small probability for any cell to be infected by an SGE <i>de novo</i>
Fitness penalty of SGE (β)	The fitness penalty imposed by the SGE
Mutation rate (μ)	Chance of mutating the evolvable HGT-rate
Mutation step (m)	Uniform step size of mutations

469 Supplementary Material

470 This supplementary material includes the mathematical derivations of the results discussed in
 471 the main text and some extra insights and figures. The source code material to reproduce the
 472 numerical simulations we have done (both in the main text and in this supplementary material),
 473 is available online (https://github.com/bramvandiijk88/HGT_Genes_And_SGEs).

474 Part I: Mathematical analyses

475 Equilibria and population growth rate of a single population

As described in the main text, we consider a population of cells that either carry or do not carry a gene. The dynamics of the density of carriers (C) and non-carriers (N) are described by:

$$\frac{dC}{dt} = \underbrace{(1 - ch + b)C}_{\text{reproduction of } C} - \underbrace{lC}_{\text{gene loss}} + \underbrace{hCN}_{\text{HGT}} - \underbrace{\phi C}_{\text{chemostat}} \quad (5)$$

$$\frac{dN}{dt} = \underbrace{(1 - ch)N}_{\text{reproduction of } N} + \underbrace{lC}_{\text{gene loss}} - \underbrace{hCN}_{\text{HGT}} - \underbrace{\phi N}_{\text{chemostat}} \quad (6)$$

$$\phi = \underbrace{(1 - ch + b)C}_{\text{total growth of } C} + \underbrace{(1 - ch)N}_{\text{total growth of } N} \quad (7)$$

$$C + N = 1 \text{ (constant population size, ensured by chemostat assumption.)} \quad (8)$$

476 Equilibria and their stability

The equilibria of Eq 5–8 are found by solving $\frac{dC}{dt} = \frac{dN}{dt} = 0$.

$$\text{Let } \frac{dC}{dt} = (1 + b + ch)C - lC + hNC - \phi C = 0.$$

Then either $C = 0$,
 or $\phi = 1 + b - ch - l + hN$ and

$$\begin{aligned} 1 + b - ch - l + hN &= (1 + b - ch)C + (1 - ch)N \\ \Leftrightarrow 1 + b - ch - l + h(1 - C) &= (1 + b - ch)C + (1 - ch)(1 - C) \\ \Leftrightarrow 1 + b - ch - l + h &= (1 + b - ch + h - 1 + ch)C + 1 - ch \\ \Leftrightarrow b - l + h &= (b + h)C \\ \Leftrightarrow C &= \frac{b - l + h}{b + h} = 1 - \frac{l}{b + h} \end{aligned}$$

Using $C + N = 1$, we find that the system has two equilibria:

$$\text{equilibrium (i): } C^* = 0, \quad N^* = 1, \quad (9)$$

$$\text{equilibrium (ii): } C^* = 1 - \frac{l}{b + h}, \quad N^* = \frac{l}{b + h} \quad (10)$$

Next, we study under what conditions the gene can persist in the population described by Eq 5–8. Note that this is equivalent to asking when equilibrium (i) is unstable, *i.e.*, when the carrying cells (C) can invade on a resident population of non-carrying cells (N) at carrying capacity. When the system is in equilibrium (i), $C^* = 0$, $N^* = 1$, and $\phi^* = (1 - ch)$. The dynamics of the carrying cells can then be approximated by

$$\frac{dC}{dt} \approx (1 + b - ch - l + hN^* - \phi^*)C = (1 + b - ch - l + h - (1 - ch))C = (b + h - l)C,$$

477 and the carrying cells can invade iff $\frac{dC}{dt} > 0$, i.e., iff

$$b + h - l > 0. \quad (11)$$

478 From Eq 11 we can conclude that genes which yield a sufficient growth rate benefit to overcome
 479 the loss rate ($b > l$) do not need HGT in order to persist in a population. Slightly beneficial
 480 genes, however, only persist when $h > (l - b)$. HGT, serving as a plausible “back-mutation”,
 481 prevents the eventual loss of such a gene from the population.

482 **Population growth rate ϕ in steady state as a function of HGT rate h**

483 Even though we have shown above that some genes can only persist in a population at sufficiently
 484 high rates of HGT, the survival of these genes does not necessarily imply that HGT also improves
 485 the actual growth rate of the population under these conditions, as the model also assumes a
 486 cost for higher rates of HGT. To gain better insight into when HGT improves the steady state
 487 growth rate, we will next consider how the population growth rate ϕ depends on h .
 The population growth rate in steady state, ϕ^* , is given by:

$$\phi^*(h) = (1 + b - ch)C^* + (1 - ch)N^* \quad (12)$$

$$= \begin{cases} 1 - ch & \text{if } h \leq (l - b) \text{ (gene cannot persist);} \\ 1 - ch + b - \frac{bl}{b+h} & \text{if } h > (l - b) \text{ (gene persists).} \end{cases} \quad (13)$$

488 To determine the effect of the rate of HGT, h , on the steady state population growth rate ϕ^* ,
 489 we differentiate Eq 13 with respect to h :

$$\frac{\partial \phi^*}{\partial h} = \begin{cases} -c & \text{if } h \leq (l - b); \\ -c + \frac{bl}{(b+h)^2} & \text{if } h > (l - b). \end{cases} \quad (14)$$

As long as $h < (l - b)$, $\frac{\partial \phi^*}{\partial h} = -c < 0$ and an increase in HGT rate h will decrease the population
 growth rate at steady state $\phi^*(h)$. For, $h > (l - b)$, the population growth rate ϕ^* might however
 have a local optimum, which we can find by setting $\frac{\partial \phi^*}{\partial h}$ to 0:

$$\frac{bl}{(b+h)^2} - c = 0$$

$$\iff (b+h)^2 = \frac{bl}{c}$$

from which we can solve

$$h_{\text{opt}} = \sqrt{\frac{bl}{c}} - b \quad (15)$$

Note that this optimum is only obtained in the function $\phi^*(h)$ if $h_{\text{opt}} > (l - b)$:

$$\sqrt{\frac{bl}{c}} - b > l - b \quad (16)$$

$$\iff \frac{bl}{c} > l^2 \quad (17)$$

$$\iff b > lc. \quad (18)$$

490 *(This is the same condition found when solving $\frac{\partial \phi^*}{\partial h} > 0$ at $h = (l - b)$)*

Furthermore, since h is the rate of HGT, we are only interested in positive values of h . $h_{\text{opt}} > 0$ iff

$$\sqrt{\frac{bl}{c}} > b \quad (19)$$

$$b < \frac{l}{c}. \quad (20)$$

Under the conditions of Eq 18 and 20, the second derivative of ϕ^* to h is

$$\frac{\partial^2 \phi^*}{\partial h^2} = \frac{-2bl}{(b+h)^3},$$

which is negative if the parameters b and l are ≥ 0 . Hence, when $\phi^*(h)$ has an optimum for a positive HGT rate h_{opt} , this local optimum is a maximum. The growth rate in this local maximum is larger than the growth rate at $h = 0$, $\phi^*(0) = 1$, iff

$$\phi^*(h_{\text{opt}}) = 1 - ch_{\text{opt}} + b - \frac{bl}{b+h_{\text{opt}}} > 1 \quad (21)$$

$$\iff 1 + cb - \sqrt{bcl} + b - \frac{bl}{\sqrt{\frac{bl}{c}}} > 1 \quad (22)$$

$$\iff b(1+c) - 2\sqrt{bcl} > 0 \quad (23)$$

$$\iff b > \frac{4lc}{(1+c)^2}. \quad (24)$$

491 Summarising, the population growth rate at equilibrium, ϕ^* , decreases linearly with the risks
 492 ch when $h < (l - b)$ due to the costs of HGT (see Eq 13). Under these conditions, the growth
 493 rate does not depend on b because the gene cannot persist in the population. When $h > (l - b)$,
 494 the gene does persist within the population, resulting in an extra term $b - \frac{bl}{b+h}$ in the growth
 495 rate $\phi^*(h)$. This extra term approaches a maximal benefit of b for high values of h . The burden
 496 of HGT ch will however eventually outweigh this benefit for increasing rates of HGT. A (local)
 497 optimal rate of HGT can found at $h_{\text{opt}} = \sqrt{bl/c} - b$, as long as $b > lc$. This optimal HGT rate
 498 is greater than 1, meaning that HGT improves the population growth rate at steady state, if
 499 the genes have a minimal benefit (see Eq 24). However, when the benefit is too large ($b > l/c$),
 500 the optimal HGT rate becomes $h_{\text{opt}} < 0$. As negative values for HGT are biologically unsound,
 501 HGT never improves the population growth rate in steady state for genes with such a high fitness
 502 benefit. Following these derivations, genes can be divided in different classes based on the value
 503 of the fitness benefit b and the consequent effect of HGT on the population growth rate at steady
 504 state (see main text and **Figure 2**):

505 **Selfish Genetic Elements (SGEs) ($b < 0$)** Carrying the gene confers a fitness cost. Increasing
 506 HGT-rates only lower the equilibrium population growth rate ϕ^* .

507

508 **Unrescuable genes ($b < l$ and $b < \frac{4lc}{(1+c)^2}$)** Genes confer a small fitness benefit, but this ben-
 509 efit is too small to overcome gene loss. Furthermore, no positive HGT rate h improves
 510 the population growth rate $\phi^*(h)$ over the population growth rate in the absence of HGT
 511 ($\phi^*(0) = 1$).

512

513 **Rescuable genes ($\frac{4lc}{(1+c)^2} < b < l$)** Genes confer a small fitness benefit and cannot persist in
 514 a population in the absence of HGT, but can be rescued by a sufficiently high HGT rate
 515 ($h > (l - b)$). For some HGT rate $h_{\text{opt}} > 0$ the equilibrium growth rate $\phi^*(h) > 1$,
 516 indicating that HGT can improve the growth rate of the population.

517

- 518 **Enrichable genes** ($l < b < l/c$) Genes confer a sufficient fitness benefit to persist in a popula-
519 tion in the absence of HGT. HGT can however improve the equilibrium population growth
520 rate $\phi^*(h_{\text{opt}})$.
- 521 **Indispensable genes** ($b > l/c$) Genes confer a large fitness benefit and can persist in a popula-
522 tion in the absence of HGT. HGT furthermore does not improve the equilibrium population
523 growth rate.

524 Evolutionary stability of HGT^+ and HGT^- populations

525 To study whether HGT is an evolvable trait, we will consider 1) if HGT can evolve *de novo*, and
 526 2) if HGT can be evolutionarily maintained. For this, we extended the two-variable model of
 527 one species to a four-variable model of two species: a HGT^+ -species that engages in HGT, and a
 528 HGT^- -species that does not (**Supplementary Figure S1B, Equation 25-28**). We analysed
 529 under what conditions the HGT^+ -species can invade an equilibrium of the HGT^- -species, and
 530 vice versa. We found that HGT can only evolve for an enrichable gene, but is evolutionarily
 531 maintained for both enrichable and rescuable genes. The following paragraphs will elaborate on
 532 how these results are derived:

Consider a HGT^+ -species (C^+ , N^+) and a HGT^- -species (C^- , N^-) that differ in their HGT rate h , but are identical otherwise. The dynamics of the density of cells carrying and not carrying the gene of the two species can be described by the following equations:

$$\frac{dC^-}{dt} = (1 + b)C^- - lC^- - \phi C^- \quad (25)$$

$$\frac{dN^-}{dt} = N^- + lC^- - \phi N^- \quad (26)$$

$$\frac{dC^+}{dt} = (1 + b - ch)C^+ - lC^+ + hN^+(C^- + C^+) - \phi C^+ \quad (27)$$

$$\frac{dN^+}{dt} = (1 - ch)N^+ + lC^+ - hN^+(C^- + C^+) - \phi N^+ \quad (28)$$

$$\phi = (1 + b)C^- + N^- + (1 + b - ch)C^+ + (1 - ch)N^+ \quad (29)$$

$$C^- + N^- + C^+ + N^+ = 1. \quad (30)$$

533 Note that we include horizontal gene transfer from HGT^- -cells carrying the gene to HGT^+ -cells
 534 that do not yet carry the gene. In other words, we consider a situation in which the propensity
 535 of HGT is determined by the acceptor cell, and not by the donor. This is inspired by for instance
 536 the process of transformation, in which the acceptor cell “decides” whether or not it takes up
 537 extracellular DNA.

538 If HGT is evolvable *de novo*, the HGT^+ species should be able to invade a HGT^- population in
 539 steady state. In other words, the equilibrium state $(C^-, N^-, C^+, N^+) = (\hat{C}^-, \hat{N}^-, 0, 0)$ should
 540 be unstable.

541

Around the equilibrium $(\hat{C}^-, \hat{N}^-, 0, 0)$, the dynamics of the HGT^+ -species are linearly approximated by

$$\begin{pmatrix} \frac{dC^+}{dt} \\ \frac{dN^+}{dt} \end{pmatrix} \approx \mathbf{J} \begin{pmatrix} C^+ \\ N^+ \end{pmatrix},$$

where

$$\mathbf{J} = \begin{pmatrix} 1 + b - ch - l - \hat{\phi} & h\hat{C}^- \\ l & 1 - ch - h\hat{C}^- - \hat{\phi} \end{pmatrix}.$$

542 The HGT^+ -species can invade iff the dominant eigenvalue of \mathbf{J} is positive.

Note that the equilibrium densities of \hat{C}^- and \hat{N}^- depend on b and l . As derived in the previous section,

$$\text{if } b \leq l, \quad \hat{C}^- = 0 \quad \text{and} \quad \hat{N}^- = 1, \quad \text{while} \quad (31)$$

$$\text{if } b > l, \quad \hat{C}^- = 1 - \frac{l}{b} \quad \text{and} \quad \hat{N}^- = \frac{l}{b}. \quad (32)$$

543 We will consider both possibilities separately.

In the case of unrescuable and rescuable genes ($0 < b \leq l$), the equilibrium densities of \widehat{C}^- and \widehat{N}^- are given by Eq 31. Then, $\widehat{\phi} = 1$ and the Jacobian matrix

$$\mathbf{J} = \begin{pmatrix} b - ch - l & 0 \\ l & -ch \end{pmatrix}.$$

544 The eigenvalues of \mathbf{J} are $\lambda_1 = b - ch - l$ and $\lambda_2 = -ch$. The second eigenvalue $\lambda_2 < 0$ as long as
 545 HGT comes at some cost $c > 0$ (the HGT-rate h of a HGT⁺-species is always positive). At the
 546 same time, λ_1 is also negative because we consider genes with a small benefit, $0 < b \leq l$. Hence,
 547 we conclude that for unrescuable and more importantly for rescuable genes, an HGT⁺-species
 548 cannot invade on a HGT⁻-population at equilibrium, and HGT can hence never evolve *de novo*.
 549

In the case of enrichable and indispensable genes ($b > l$), the equilibrium densities of \widehat{C}^- and \widehat{N}^- are given by Eq 32. Now, $\widehat{\phi} = (1 + b)(1 - \frac{l}{b}) + \frac{l}{b} = 1 + b - l$, and the Jacobian matrix

$$\mathbf{J} = \begin{pmatrix} -ch & h(1 - \frac{l}{b}) \\ l & l - b - ch - h(1 - \frac{l}{b}) \end{pmatrix}.$$

The eigenvalues of \mathbf{J} should now be solved from

$$(-ch - \lambda)(l - b - ch - h(1 - \frac{l}{b}) - \lambda) - lh(1 - \frac{l}{b}) = 0 \quad (33)$$

$$\iff \lambda^2 - \lambda(l - b - 2ch - h(1 - \frac{l}{b})) + (bch - lch + c^2h^2 + ch^2(1 - \frac{l}{b}) - lh(1 - \frac{l}{b})) = 0. \quad (34)$$

Let

$$\beta = l - b - 2ch - h(1 - \frac{l}{b}), \quad \text{and} \quad (35)$$

$$\gamma = bch - lch + c^2h^2 + ch^2(1 - \frac{l}{b}) - lh(1 - \frac{l}{b}). \quad (36)$$

550 Then, the eigenvalues of \mathbf{J} are equal to $\lambda_{1,2} = \frac{1}{2}(\beta \pm \sqrt{\beta^2 - 4\gamma})$. Remember that we are interested
 551 in the sign of the dominant eigenvalue. If the eigenvalues are complex ($\beta^2 < 4\gamma$), the real part
 552 of the eigenvalues $\text{Re}(\lambda_{1,2}) > 0$ iff $\beta > 0$. If the eigenvalues are real, the dominant eigenvalue is
 553 $\lambda_1 = \frac{1}{2}(\beta + \sqrt{\beta^2 - 4\gamma})$, and $\lambda_1 > 0$ iff $\beta > 0$ or $\sqrt{\beta^2 - 4\gamma} > \beta \iff \gamma < 0$.
 First, consider the possibility $\beta > 0$. Then we should have

$$l - b - 2ch - h(1 - \frac{l}{b}) > 0 \quad (37)$$

$$\iff l - b > h(2c + (1 - \frac{l}{b})). \quad (38)$$

This is however a contradiction, since we here deal with genes for which $b > l$ and hence $l - b < 0$, but $\widehat{C}^- = 1 - \frac{l}{b} > 0$, $c > 0$ and $h > 0$. Hence, β is always negative and the dominant eigenvalue is positive only if $\gamma < 0$. From $\gamma < 0$, we find

$$bch - lch + c^2h^2 + ch^2(1 - \frac{l}{b}) - lh(1 - \frac{l}{b}) < 0 \quad (39)$$

$$\iff c(b - l + ch) + (ch - l)(1 - \frac{l}{b}) < 0 \quad (40)$$

554 Trying to solve Eq 40 for any value of h would yield a complicated condition on the value of
 555 b . However, we can further simplify Eq 40 by asking if a HGT⁺-species with a very small (but
 556 positive) HGT-rate could invade. For $h = \epsilon \approx 0$, Eq 40 reduces to

$$c(b - l) - l(1 - \frac{l}{b}) < 0, \quad (41)$$

from which we can solve

$$c(b-l) - l(1 - \frac{l}{b}) < 0 \quad (42)$$

$$\iff cb^2 - l(c+1)b + l^2 < 0 \quad (43)$$

$$\iff (cb-l)(b-l) < 0. \quad (44)$$

557 Since we consider enrichable and indispensable genes, with $b > l$, condition 44 can only be true if
 558 $cb < l \iff b < l/c$, which is exactly the condition that separates enrichable from indispensable
 559 genes. Hence, we conclude that for enrichable genes ($l < b < l/c$), a HGT⁺-species with a small
 560 but positive HGT-rate can always invade on a HGT⁻-population at equilibrium, and that HGT
 561 can hence evolve *de novo*.

562 So far, we have determined under what conditions a HGT⁻-population is evolutionarily stable.
 563 We can however ask the same for a HGT⁺-population. In other words, even though it may not be
 564 reached by gradual evolution, can HGT be *maintained*? To answer this question, we next consider
 565 the evolutionary stability of the HGT⁺-equilibrium: $(C^-, N^-, C^+, N^+) = (0, 0, \tilde{C}^+, \tilde{N}^+)$.
 Again, the densities of C^+ - and N^+ -cells at equilibrium depend on the values of b, l and h (see
 Eq 9–10 in the previous section):

$$\text{if } b \leq l - h, \quad \tilde{C}^+ = 0 \quad \text{and} \quad \tilde{N}^+ = 1, \quad \text{while} \quad (45)$$

$$\text{if } b > l - h, \quad \tilde{C}^+ = 1 - \frac{l}{b+h} \quad \text{and} \quad \tilde{N}^+ = \frac{l}{b+h}. \quad (46)$$

566 If $b \leq l - h$, the gene does not persist in the population and HGT hence does not confer any
 567 benefit, while still imposing a cost on the N^+ -cells. Under these conditions, the N^- -cells, that
 568 do not carry the cost of HGT, will always be able to invade.

For the more interesting case in which the gene does persist in a HGT⁺-population (Eq 46), we
 now linearise the dynamics of the HGT⁻-species around the equilibrium:

$$\begin{pmatrix} \frac{dC^-}{dt} \\ \frac{dN^-}{dt} \end{pmatrix} = \mathbf{J} \begin{pmatrix} C^- \\ N^- \end{pmatrix}$$

$$\text{with } \mathbf{J} = \begin{pmatrix} (1+b) - l - \tilde{\phi} & 0 \\ l & 1 - \tilde{\phi} \end{pmatrix}$$

$$\text{and } \tilde{\phi} = (1+b-ch)(1 - \frac{l}{b+h}) + (1-ch)\frac{l}{b+h} = (1-ch) + b(1 - \frac{l}{b+h}).$$

569 Again, the HGT⁻-species can invade if the dominant eigenvalue of \mathbf{J} is positive, and hence
 570 the HGT⁺-species of equilibrium is evolutionarily stable if both eigenvalues are negative. The
 571 eigenvalues of \mathbf{J} are $\lambda_1 = 1 + b - l - \tilde{\phi}$ and $\lambda_2 = 1 - \tilde{\phi}$.

For the first eigenvalue, we find

$$\lambda_1 < 0 \quad (47)$$

$$\iff 0 > 1 + b - l - \tilde{\phi} \quad (48)$$

$$\iff 0 > 1 + b - l - (1 - ch) - b(1 - \frac{l}{b+h}) \quad (49)$$

$$\iff 0 > bl + ch(b+h) - l(b+h) \quad (50)$$

$$\iff lh > ch(b+h) \quad (51)$$

$$\iff l > c(b+h) \quad (52)$$

$$\iff c < \frac{l}{b+h} \quad (53)$$

572 Hence, this first eigenvalue is negative as long as the costs of HGT are not too large.
573 For the second eigenvalue, we find:

$$\lambda_2 < 0 \quad (54)$$

$$\iff 0 > 1 - \tilde{\phi} \quad (55)$$

$$\iff 0 > 1 - (1 - ch) - b\left(1 - \frac{l}{b+h}\right) \quad (56)$$

$$\iff 0 > ch - b\left(1 - \frac{l}{b+h}\right) \quad (57)$$

$$\iff ch < b\left(1 - \frac{l}{b+h}\right) \quad (58)$$

$$\iff c < \frac{b\left(1 - \frac{l}{b+h}\right)}{h}. \quad (59)$$

573 Remember that we considered a HGT⁺-population in which the gene can persist, *i.e.*, $b+h > l$.
574 Hence $\frac{l}{b+h} < 1$ and the right hand side in Eq 59 is positive. Hence, we can again conclude that
575 there are some non-zero costs for which λ_2 is negative.

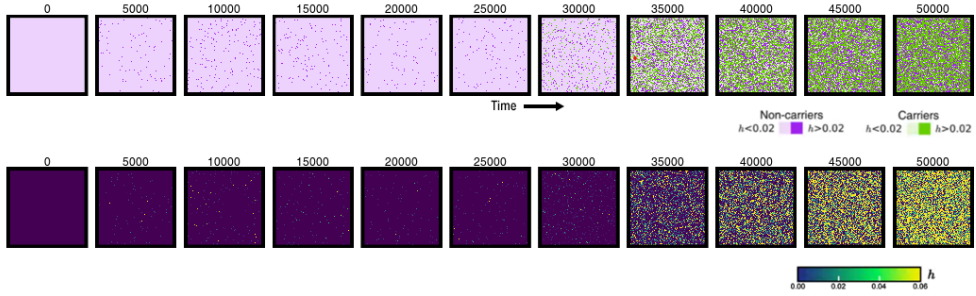
576 Combining the results in Eq 53 and 59, we see that for some costs, HGT can be maintained.
577 For rescuable genes with costs that satisfy conditions 53 and 59, there is an Allee effect with
578 respect to HGT: HGT can be evolutionarily maintained, but it cannot evolve *de novo*. This
579 result can be intuitively understood. Small (invading) HGT⁺-populations pay the continuous
580 costs for HGT, but hardly ever interact with their conspecifics, and hence the positive fitness
581 effects of maintaining the slightly beneficial gene are too small to overcome the costs for HGT.
582 Higher fitness can only be achieved when the population size is large enough, such that the
583 benefits conferred by HGT outweigh its costs. The presence of an Allee effect was confirmed by
584 numerically integrating Eq 25–28 for different initial conditions. We then indeed see that the
585 system converges to different equilibria depending on the initial frequency of HGT⁺-cells (see
586 **Figure 3**).

587 **Part II: Supplementary results and figures**

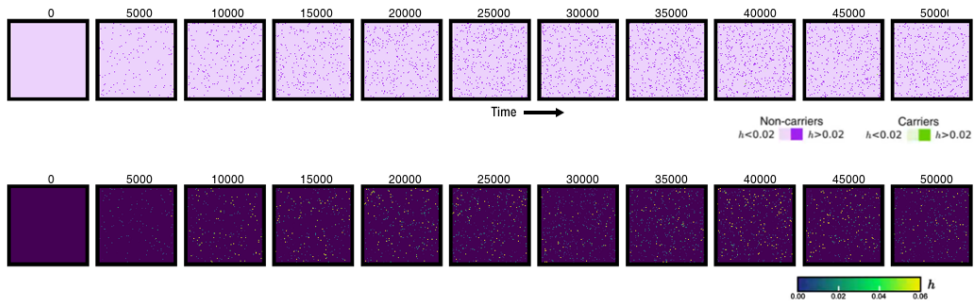
588 **In the well-mixed IBM, HGT only evolves for enrichable genes**

589 In the main text we discussed that HGT cannot evolve for genes that cannot persist without
590 HGT. For these genes, a lack of donor cells does not allow mutants that engage in HGT to get a
591 significant fitness benefit, even when they actually *do* carry the beneficial gene. To get over this
592 so-called Allee effect, a large number of gene-carrying individuals has to simultaneously start
593 engaging in HGT. We have also shown that, in the spatially structure populations, HGT *does*
594 evolve for genes that could not persist without HGT, as it is more likely that the lack of donor
595 cells is, at least locally, overcome. This supplementary figure summarises this result, by showing
596 that, even though HGT does evolve for enrichable genes under well-mixed conditions, it indeed
597 fails to evolve for rescuable genes.

A) HGT evolution for an enrichable gene ($b=0.03$, well-mixed)



B) HGT evolution for a rescuable gene ($b=0.0175$, well-mixed)



C) HGT evolution for a rescuable gene ($b=0.03$, no mixing)

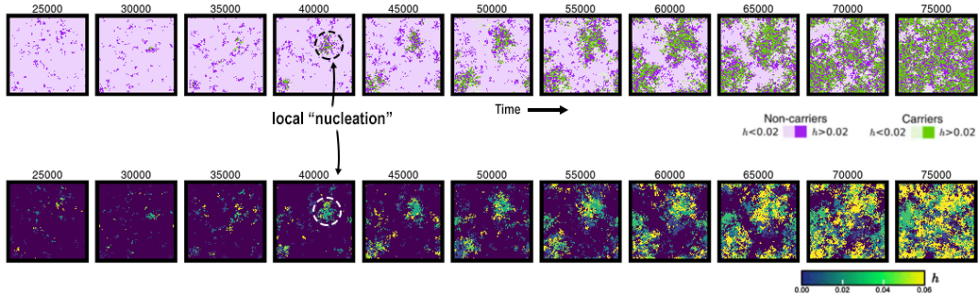
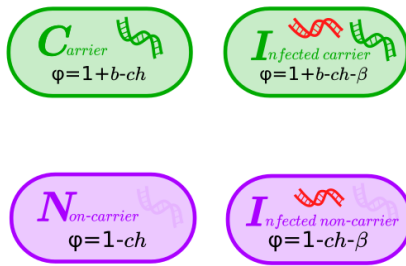


Figure S1: HGT evolution in IBM under various conditions

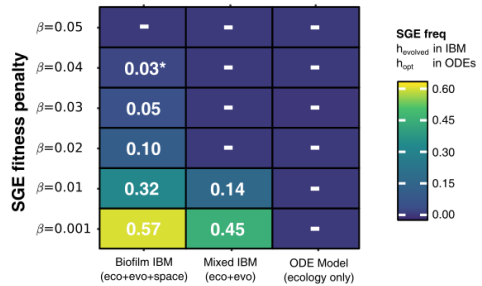
598 Maintaining weak and strong Selfish Genetic Elements

599 In the main text we have discussed how SGEs can coexist along-side their hosts and slightly
 600 beneficial genes, even when the their fitness-penalty is greater than the benefit of the gene.
 601 However, this was only observed in the spatially structured model, as illustrated in the figure
 602 below.

A) Cartoon of the 4 cell types



B) Persistence of SGEs



C) SGEs with intermediate fitness penalty impose the highest cost on the host

(simulated in the spatially structured populations of 200x200 cells)

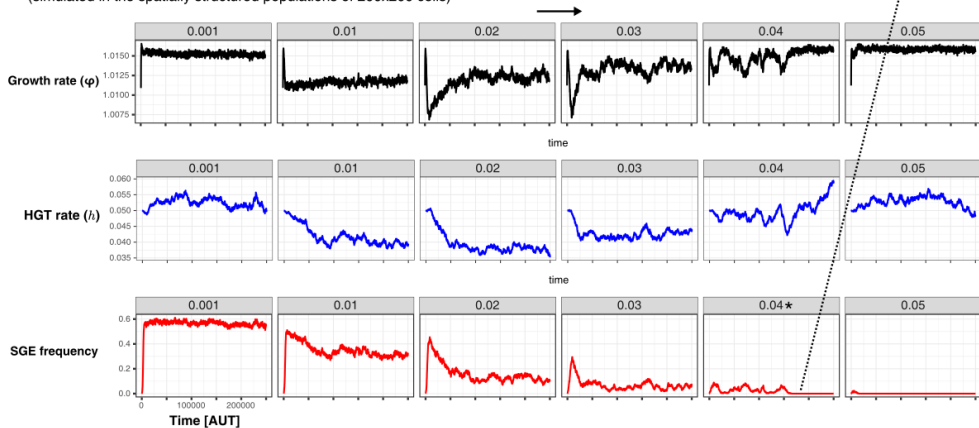


Figure S2: Persistence of SGEs in various implementations of our model. A shows a cartoon of the cell types, between which competition was modelled in a various ways. In B we show for these different implementations how many SGEs persist within the populations for SGEs with different penalties. For the IBM, we simulated for 250.000 time steps and calculated the average SGE-frequency in the final 100 generations. For the ODE model, we chose the optimal rate of HGT (h_{opt}), and numerically integrated the equilibrium concentrations of infected cells. Finally, C shows the temporal dynamics of the growth rate (ϕ), HGT-rate (h), and the SGE frequency, in the spatially structured simulations. As this parameter sweep had slightly smaller populations sizes as used in the main text, the strong SGE could eventually go extinct (this is annotated with an asterisk).

Equations for Supplementary Figure S2A

$$\begin{aligned}\frac{dC}{dt} &= \gamma(1 + b - ch)C - lC + lD + h(NC + 0.5ND - CP - CD) - \phi C \\ \frac{dN}{dt} &= \gamma(1 - ch)N + lC + lP - hN(C + P + D) - \phi N \\ \frac{dP}{dt} &= \gamma(1 - \beta - ch)P + lD - lP + h(NP + ND/2 - CP - PD) - \phi P \\ \frac{dD}{dt} &= \gamma(1 + b - \beta - ch)D - lD + h(CP + CD + PC + PD) - \phi D \\ \phi &= \gamma((1 + b - ch)C + (1 - ch)N + (1 - \beta - ch)P + (1 + b - \beta - ch)D)\end{aligned}\tag{60}$$

603 **Strong SGEs fail to spread / persist in the population at low HGT-distances**

604 In the main text we have discussed how we found that strong SGEs (genetic parasites with
605 a greater penalty than the beneficial gene) could nevertheless stably coexist with an evolving
606 population of cells. However, this persistence of SGEs relies on their ability to escape to new
607 susceptible hosts who have not experienced SGEs for some time (and therefore have evolved
608 elevated HGT rates). In this supplementary figure, it is indeed seen how the distance influences
609 the spread / persistence of SGEs. If the distance between donor and acceptor is very local ($d=1$),
610 SGEs cannot spread even while they are still fluxing in (top row). For an intermediate HGT-
611 distance ($1 < d < 10$), the SGEs persist for a bit as long as they flux in, but die out when influx
612 is stopped (middle row). For larger HGT distances ($d > 10$), we found that SGEs can persist
613 even after the influx was stopped.

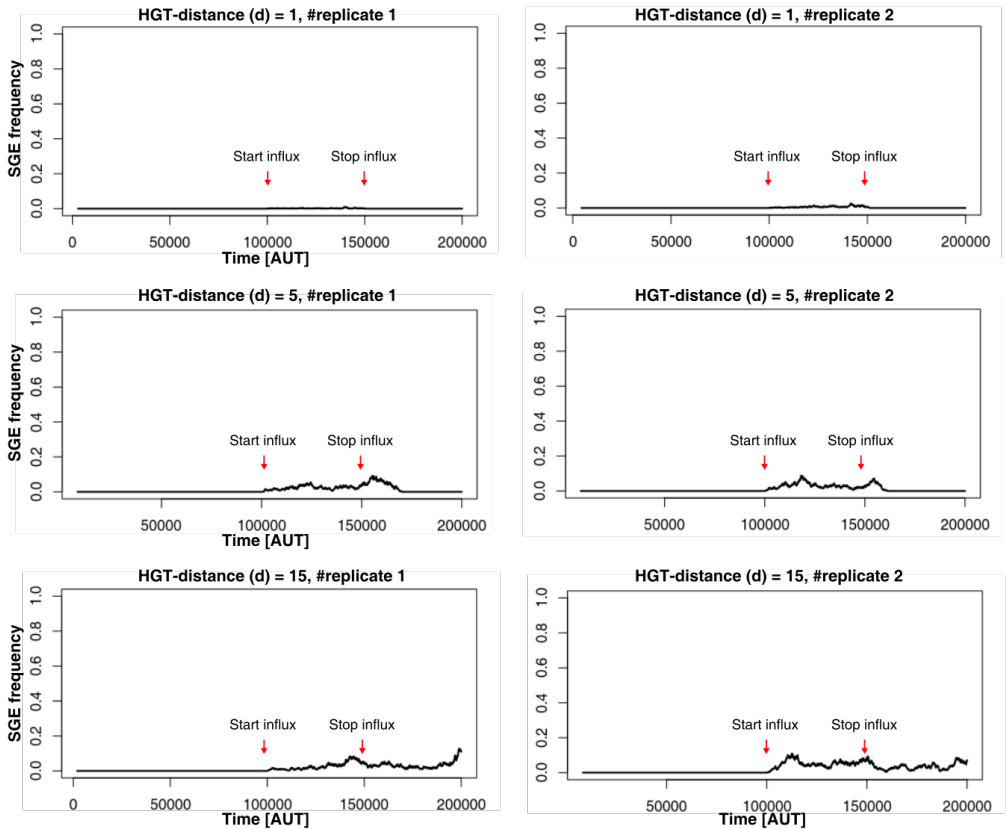


Figure S3: SGEs persistence in open and closed ecosystems. Parameters used:

614 **Supplementary Movie - Gene-sharing ‘nucleation events’ and coexistence with**
 615 **SGEs**

616 This annotated supplementary movie illustrates how gene-sharing of rescuable genes emerges
 617 through a ‘nucleation’-event, allowing local communities eventually overgrow all other cells.
 618 Where a local sub-community initially transitions to the alternative HGT^+ state, eventually
 619 the whole population will be taken over. Similar mechanisms have been observed in origin of life
 620 studies Wu and Higgs (2012) and microbial community transitions Kotil and Vetsigian (2018).
 621 We also show here how SGEs (here with $b = 0.01$) are able to infect, and stably coexist, with
 622 this gene-transferring community.

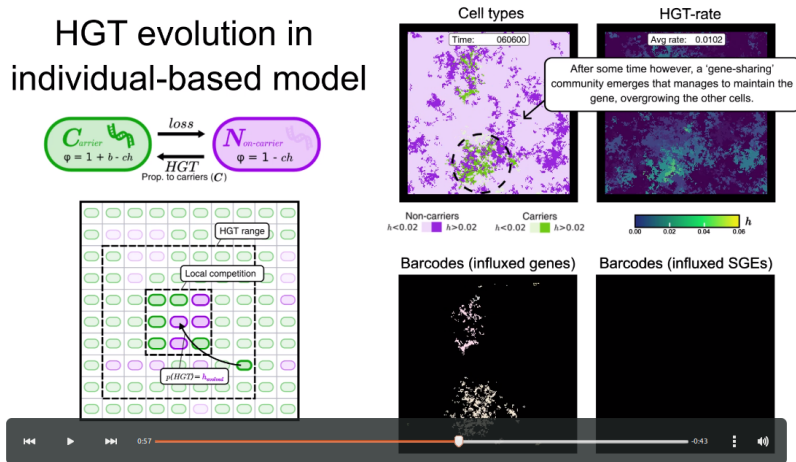


Figure S4: A snapshot from the supplementary movie.

623 <https://youtu.be/cpJh-CeFPm4>

624 Bibliography

- 625 A. N. N. Ba, I. Cvijović, J. I. R. Echenique, K. R. Lawrence, A. Rego-Costa, X. Liu, S. F. Levy,
626 and M. M. Desai. High-resolution lineage tracking reveals travelling wave of adaptation in
627 laboratory yeast. *Nature*, 575(7783):494–499, 2019.
- 628 D. A. Baltrus. Exploring the costs of horizontal gene transfer. *Trends in ecology & evolution*, 28
629 (8):489–495, 2013.
- 630 T. Bataillon. Estimation of spontaneous genome-wide mutation rate parameters: whither
631 beneficial mutations? *Heredity*, 84(5):497, 2000.
- 632 C. W. Beitel, L. Froenicke, J. M. Lang, I. F. Korf, R. W. Micheltore, J. A. Eisen, and A. E.
633 Darling. Strain-and plasmid-level deconvolution of a synthetic metagenome by sequencing
634 proximity ligation products. *PeerJ*, 2:e415, 2014.
- 635 C. T. Bergstrom, M. Lipsitch, and B. R. Levin. Natural selection, infectious transfer and the
636 existence conditions for bacterial plasmids. *Genetics*, 155(4):1505–1519, 2000.
- 637 J. R. Blundell and S. F. Levy. Beyond genome sequencing: lineage tracking with barcodes to
638 study the dynamics of evolution, infection, and cancer. *Genomics*, 104(6):417–430, 2014.
- 639 J. N. Burton, I. Liachko, M. J. Dunham, and J. Shendure. Species-level deconvolution of
640 metagenome assemblies with hi-c-based contact probability maps. *G3: Genes, Genomes,
641 Genetics*, 4(7):1339–1346, 2014.
- 642 E. Casacuberta and J. González. The impact of transposable elements in environmental
643 adaptation. *Molecular ecology*, 22(6):1503–1517, 2013.
- 644 J. M. Chacón and W. R. Harcombe. Increasing growth rate slows adaptation when genotypes
645 compete for diffusing resources. *bioRxiv*, page 616938, 2019.
- 646 S. C. Choi, M. D. Rasmussen, M. J. Hubisz, I. Gronau, M. J. Stanhope, and A. Siepel. Replacing
647 and additive horizontal gene transfer in streptococcus. *Molecular biology and evolution*, 29(11):
648 3309–3320, 2012.
- 649 O. X. Cordero, L.-A. Ventouras, E. F. DeLong, and M. F. Polz. Public good dynamics drive
650 evolution of iron acquisition strategies in natural bacterioplankton populations. *Proceedings
651 of the National Academy of Sciences*, 109(49):20059–20064, 2012.
- 652 T. Dimitriu, F. Medaney, E. Amanatidou, J. Forsyth, R. J. Ellis, and B. Raymond. Negative
653 frequency dependent selection on plasmid carriage and low fitness costs maintain extended
654 spectrum β -lactamases in escherichia coli. *Scientific Reports*, 9(1):1–7, 2019.
- 655 W. F. Doolittle and O. Zhaxybayeva. On the origin of prokaryotic species. *Genome research*, 19
656 (5):744–756, 2009.

- 657 Y. Gerardin, M. Springer, and R. Kishony. A competitive trade-off limits the selective advantage
658 of increased antibiotic production. *Nature microbiology*, 1(12):16175, 2016.
- 659 I. Gordo and P. R. Campos. Adaptive evolution in a spatially structured asexual population.
660 *Genetica*, 127(1-3):217, 2006.
- 661 M. G. Habets, T. Czaran, R. F. Hoekstra, and J. A. G. De Visser. Spatial structure inhibits
662 the rate of invasion of beneficial mutations in asexual populations. *Proceedings of the Royal
663 Society B: Biological Sciences*, 274(1622):2139–2143, 2007.
- 664 J. P. Hall, A. J. Wood, E. Harrison, and M. A. Brockhurst. Source–sink plasmid transfer dynamics
665 maintain gene mobility in soil bacterial communities. *Proceedings of the National Academy of
666 Sciences*, 113(29):8260–8265, 2016.
- 667 W. Hao and G. B. Golding. The fate of laterally transferred genes: life in the fast lane to
668 adaptation or death. *Genome Research*, 16(5):636–643, 2006.
- 669 J.-H. Hehemann, P. Arevalo, M. S. Datta, X. Yu, C. H. Corzett, A. Henschel, S. P. Preheim,
670 S. Timberlake, E. J. Alm, and M. F. Polz. Adaptive radiation by waves of gene transfer leads
671 to fine-scale resource partitioning in marine microbes. *Nature communications*, 7:12860, 2016.
- 672 P. T. Inc. Collaborative data science, 2015. URL <https://plot.ly>.
- 673 J. Iranzo, Y. I. Wolf, E. V. Koonin, and I. Sela. Gene gain and loss push prokaryotes beyond
674 the homologous recombination barrier and accelerate genome sequence divergence. *Nature
675 Communications*, 10(1):1–10, 2019.
- 676 R. Jain, M. C. Rivera, J. E. Moore, and J. A. Lake. Horizontal gene transfer accelerates genome
677 innovation and evolution. *Molecular biology and evolution*, 20(10):1598–1602, 2003.
- 678 Y. F. P. Job. Gnu parallel: The command-line power tool.
- 679 J.-N. Kim, Y. Kim, Y. Jeong, J.-H. Roe, B.-G. Kim, and B.-K. Cho. Comparative genomics
680 reveals the core and accessory genomes of streptomyces species. *J Microbiol Biotechnol*, 25
681 (10):1599–605, 2015.
- 682 S. E. Kotil and K. Vetsigian. Emergence of evolutionary stable communities through eco-
683 evolutionary tunneling. *bioRxiv*, page 271015, 2018.
- 684 C.-H. Kuo and H. Ochman. Deletional bias across the three domains of life. *Genome biology and
685 evolution*, 1:145–152, 2009.
- 686 T. Lefébure and M. J. Stanhope. Evolution of the core and pan-genome of streptococcus: positive
687 selection, recombination, and genome composition. *Genome biology*, 8(5):R71, 2007.
- 688 J. Lerner, M. Manhart, W. Jasinska, L. Gauthier, A. W. Serohijos, and S. Bershtein.
689 Chromosomal barcoding of e. coli populations reveals lineage diversity dynamics at high
690 resolution. *bioRxiv*, page 571505, 2019.
- 691 S. F. Levy, J. R. Blundell, S. Venkataram, D. A. Petrov, D. S. Fisher, and G. Sherlock.
692 Quantitative evolutionary dynamics using high-resolution lineage tracking. *Nature*, 519(7542):
693 181, 2015.
- 694 F. Li, M. L. Salit, and S. F. Levy. Unbiased fitness estimation of pooled barcode or amplicon
695 sequencing studies. *Cell systems*, 7(5):521–525, 2018.

- 696 L. N. Lili, N. F. Britton, and E. J. Feil. The persistence of parasitic plasmids. *Genetics*, 177(1):
697 399–405, 2007.
- 698 A. J. Lopatkin, S. Huang, R. P. Smith, J. K. Srimani, T. A. Sysoeva, S. Bewick, D. K. Karig,
699 and L. You. Antibiotics as a selective driver for conjugation dynamics. *Nature microbiology*, 1
700 (6):16044, 2016.
- 701 M. Lynch, J. Conery, and R. Bürger. Mutational meltdowns in sexual populations. *Evolution*,
702 49(6):1067–1080, 1995.
- 703 M. Lynch, M. S. Ackerman, J.-F. Gout, H. Long, W. Sung, W. K. Thomas, and P. L. Foster.
704 Genetic drift, selection and the evolution of the mutation rate. *Nature Reviews Genetics*, 17
705 (11):704, 2016.
- 706 J. S. Madsen, M. Burmølle, L. H. Hansen, and S. J. Sørensen. The interconnection between
707 biofilm formation and horizontal gene transfer. *FEMS Immunology & Medical Microbiology*,
708 65(2):183–195, 2012.
- 709 S. E. Mc Ginty, D. J. Rankin, and S. P. Brown. Horizontal gene transfer and the evolution
710 of bacterial cooperation. *Evolution: International Journal of Organic Evolution*, 65(1):21–32,
711 2011.
- 712 J. C. Mell and R. J. Redfield. Natural competence and the evolution of dna uptake specificity.
713 *Journal of bacteriology*, 196(8):1471–1483, 2014.
- 714 J. A. Metz, S. A. Geritz, G. Meszéna, F. J. Jacobs, and J. S. Van Heerwaarden. Adaptive
715 dynamics: a geometrical study of the consequences of nearly faithful reproduction. 1995.
- 716 J. J. Morris, R. E. Lenski, and E. R. Zinser. The black queen hypothesis: evolution of
717 dependencies through adaptive gene loss. *MBio*, 3(2):e00036–12, 2012.
- 718 H. J. Muller. The relation of recombination to mutational advance. *Mutation
719 Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 1(1):2–9, 1964.
- 720 P. Nazarian, F. Tran, and J. Q. Boedicker. Modeling multispecies gene flow dynamics reveals
721 the unique roles of different horizontal gene transfer mechanisms. *Frontiers in microbiology*,
722 9:2978, 2018.
- 723 R. Niehus, S. Mitri, A. G. Fletcher, and K. R. Foster. Migration and horizontal gene transfer
724 divide microbial genomes into multiple niches. *Nature communications*, 6:8924, 2015.
- 725 T. Nogueira, D. J. Rankin, M. Touchon, F. Taddei, S. P. Brown, and E. P. Rocha. Horizontal
726 gene transfer of the secretome drives the evolution of bacterial cooperation and virulence.
727 *Current Biology*, 19(20):1683–1691, 2009.
- 728 A. Novick and W. F. Doolittle. Horizontal persistence and the complexity hypothesis. *Biology
729 & Philosophy*, 35(1):2, 2020.
- 730 R. W. Nowell, S. Green, B. E. Laue, and P. M. Sharp. The extent of genome flux and its role in
731 the differentiation of bacterial lineages. *Genome biology and evolution*, 6(6):1514–1529, 2014.
- 732 H. Ochman, J. G. Lawrence, and E. A. Groisman. Lateral gene transfer and the nature of
733 bacterial innovation. *nature*, 405(6784):299, 2000.
- 734 P. Puigbò, A. E. Lobkovsky, D. M. Kristensen, Y. I. Wolf, and E. V. Koonin. Genomes in turmoil:
735 quantification of genome dynamics in prokaryote supergenomes. *BMC biology*, 12(1):66, 2014.

- 736 S. D. Quistad, G. Doucier, and P. B. Rainey. Experimental manipulation of selfish genetic
737 elements links genes to microbial community function. *BioRxiv*, page 608752, 2019.
- 738 D. J. Rankin, E. P. Rocha, and S. P. Brown. What traits are carried on mobile genetic elements,
739 and why? *Heredity*, 106(1):1, 2011.
- 740 R. J. Redfield. Do bacteria have sex? *Nature Reviews Genetics*, 2(8):634, 2001.
- 741 M. A. Riley and J. E. Wertz. Bacteriocins: evolution, ecology, and application. *Annual Reviews*
742 *in Microbiology*, 56(1):117–137, 2002.
- 743 T. Sakoparnig, C. Field, and E. van Nimwegen. Whole genome phylogenies reflect long-tailed
744 distributions of recombination rates in many bacterial species. *bioRxiv*, page 601914, 2019.
- 745 S. T. Schultz and M. Lynch. Mutation and extinction: the role of variable mutational effects,
746 synergistic epistasis, beneficial mutations, and degree of outcrossing. *Evolution*, 51(5):1363–
747 1371, 1997.
- 748 B. J. Shapiro, J. Friedman, O. X. Cordero, S. P. Preheim, S. C. Timberlake, G. Szabó, M. F.
749 Polz, and E. J. Alm. Population genomics of early events in the ecological differentiation of
750 bacteria. *science*, 336(6077):48–51, 2012.
- 751 F. R. Slater, M. J. Bailey, A. J. Tett, and S. L. Turner. Progress towards understanding the fate
752 of plasmids in bacterial communities. *FEMS Microbiology Ecology*, 66(1):3–13, 2008.
- 753 B. Snel, P. Bork, and M. A. Huynen. Genomes in flux: the evolution of archaeal and
754 proteobacterial gene content. *Genome research*, 12(1):17–25, 2002.
- 755 K. Soetaert, T. Petzoldt, and R. W. Setzer. Solving differential equations in R: Package deSolve.
756 *Journal of Statistical Software*, 33(9):1–25, 2010. ISSN 1548-7660. doi: 10.18637/jss.v033.i09.
757 URL <http://www.jstatsoft.org/v33/i09>.
- 758 S. M. Soucy, J. Huang, and J. P. Gogarten. Horizontal gene transfer: building the web of life.
759 *Nature Reviews Genetics*, 16(8):472–482, 2015.
- 760 T. Stalder, B. Cornwell, J. Lacroix, B. Kohler, S. Dixon, H. Yano, B. Kerr, L. J. Forney, and
761 E. M. Top. Evolving populations in biofilms contain more persistent plasmids. *Molecular*
762 *Biology and Evolution*, 2020.
- 763 N. Takeuchi, K. Kaneko, and E. V. Koonin. Horizontal gene transfer can rescue prokaryotes
764 from muller’s ratchet: benefit of dna from dead cells and population subdivision. *G3: Genes,*
765 *Genomes, Genetics*, 4(2):325–339, 2014.
- 766 N. Takeuchi, O. X. Cordero, E. V. Koonin, and K. Kaneko. Gene-specific selective sweeps in
767 bacteria and archaea caused by negative frequency-dependent selection. *BMC biology*, 13(1):
768 20, 2015.
- 769 C. M. Thomas and K. M. Nielsen. Mechanisms of, and barriers to, horizontal gene transfer
770 between bacteria. *Nature reviews microbiology*, 3(9):711, 2005.
- 771 T. Toffoli and N. Margolus. Cellular automata machines: a new environment for modeling
772 (scientific computation). In *Press Cambridge*, page 259. 1987.
- 773 M. Touchon, C. Hoede, O. Tenaillon, V. Barbe, S. Baeriswyl, P. Bidet, E. Bingen, S. Bonacorsi,
774 C. Bouchier, O. Bouvet, et al. Organised genome dynamics in the escherichia coli species
775 results in highly diverse adaptive paths. *PLoS genetics*, 5(1):e1000344, 2009.

- 776 T. J. Treangen and E. P. Rocha. Horizontal transfer, not duplication, drives the expansion of
777 protein families in prokaryotes. *PLoS genetics*, 7(1):e1001284, 2011.
- 778 B. van Dijk and P. Hogeweg. In silico gene-level evolution explains microbial population diversity
779 through differential gene mobility. *Genome biology and evolution*, 8(1):176–188, 2015.
- 780 Z. Vig-Milkovics, I. Zachar, Á. Kun, A. Szilágyi, and E. Szathmáry. Moderate sex between
781 protocells can balance between a decrease in assortment load and an increase in parasite
782 spread. *Journal of theoretical biology*, 462:304–310, 2019.
- 783 A. A. Vogan and P. G. Higgs. The advantages and disadvantages of horizontal gene transfer and
784 the emergence of the first species. *Biology direct*, 6(1):1, 2011.
- 785 T. Vogwill and R. C. MacLean. The genetic basis of the fitness costs of antimicrobial resistance:
786 a meta-analysis approach. *Evolutionary applications*, 8(3):284–295, 2015.
- 787 M. Vos, M. C. Hesselman, T. A. te Beek, M. W. van Passel, and A. Eyre-Walker. Rates of lateral
788 gene transfer in prokaryotes: high but why? *Trends in microbiology*, 23(10):598–605, 2015.
- 789 M. Vos, A. Buckling, and B. Kujiper. Sexual selection in bacteria? *Trends in microbiology*, 2019.
- 790 R. A. Welch, V. Burland, G. Plunkett, P. Redford, P. Roesch, D. Rasko, E. Buckles, S.-R. Liou,
791 A. Boutin, J. Hackett, et al. Extensive mosaic structure revealed by the complete genome
792 sequence of uropathogenic escherichia coli. *Proceedings of the National Academy of Sciences*,
793 99(26):17020–17024, 2002.
- 794 H. Wickham. *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York, 2016.
795 ISBN 978-3-319-24277-4. URL <https://ggplot2.tidyverse.org>.
- 796 J. Wiedenbeck and F. M. Cohan. Origins of bacterial diversity through horizontal genetic transfer
797 and adaptation to new ecological niches. *FEMS microbiology reviews*, 35(5):957–976, 2011.
- 798 M. J. Wisner and R. E. Lenski. A comparison of methods to measure fitness in escherichia coli.
799 *PLoS One*, 10(5):e0126210, 2015.
- 800 M. Wu and P. G. Higgs. The origin of life is a spatially localized stochastic transition. *Biology*
801 *direct*, 7(1):42, 2012.
- 802 E. Yaffe and D. A. Relman. Tracking microbial evolution in the human gut using hi-c reveals
803 extensive horizontal gene transfer, persistence and adaptation. *Nature Microbiology*, 5(2):
804 343–353, 2020.