Consideration of genetic and sex effects in mice enhances consilience with human addiction studies

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1 Abstract

Concerns about external validity of rodent models and translation of findings across species are

often based on narrow investigations of populations with limited diversity. Sources of individual

variation – including genetics and sex – are only infrequently encompassed in model organism

studies. As with most complex diseases, risk for cocaine use disorder is subject to considerable

inter-individual variation. Explicit inclusion of individual differences in rodent research may

reveal conserved phenotypes and molecular systems relevant to human addiction. We

surveyed cocaine-related traits in both males and females of eight inbred mouse strains whose

genomes collectively capture 90% of the genetic diversity of the mouse species. Across these

strains, individual differences explained a substantial proportion of variance in cocaine-

responsive or cocaine response-predictive behavioral and physiological phenotypes. Wild-

derived mouse strains often extended the phenotypic ranges of these behaviors beyond what is

observed in conventional laboratory strains. Striatum transcriptional responses to cocaine were

also highly dependent upon strain and sex differences; most cocaine-responsive genes were

differentially expressed in a manner moderated by strain, sex, or their combination. We

compared the strain- and sex-mediated transcriptional responses to cocaine in mice to

transcriptomic analysis of people with cocaine use disorder and found that mouse similarity to

humans was highly dependent upon mouse genetic background and sex. Specifically, male

WSB/EiJ mice and female NOD/ShiLtJ mice exhibited the greatest degree of neural

transcriptional consilience with humans with cocaine use disorder. Model organism diversity

thus represents a crucial source of biological information that can substantially improve

external validity of neuropsychiatric research.

Significance Statement

Laboratory mice are widely used in research on neurobiological mechanisms of addiction, but most studies use a single strain and often sex of mice. To assess how individual differences in mice modulate addiction-related traits and how this impacts comparative analysis with humans, we studied cocaine-relevant behaviors and brain molecular correlates in both males and females of genetically diverse mouse strains. In this population, individual differences related to sex and/or genetics explain large proportions of differences in cocaine-related traits. Importantly, brain gene expression data demonstrated that some strains mimic human genomic states more readily than others. Individual differences thus represent a crucial and underdeveloped source of biological information about addiction mechanisms that may influence the translational utility of such studies.

Introduction

Complex diseases such as neuropsychiatric disorders are typically characterized by the contributions of many genes (Hyman, 2018). Behavioral genetics in non-human animals aims to establish consilience with psychiatrically relevant human systems by studying conserved behaviors and their conserved underlying neural molecular substrates. However, despite mounting evidence of deep conservation in the complex gene systems driving behavior (Saul et al., 2019a; Sinha et al., 2020; Young et al., 2019), uncertainty about the psychiatric relevance of non-human animal systems abounds in the psychiatric genetics community (National Advisory Mental Health Council Workgroup on Genomics, 2018). These doubts likely arise from longstanding and well-characterized difficulties in replicating non-human animal behavioral findings (Crabbe et al., 1999) as well as the failure to translate psychiatric genetics results from any species into the clinic (Hyman, 2012). A critical reassessment of non-human animal behavioral genetics is needed for non-human animals to contribute to the human psychiatric literature.

To simplify experimental design, rodent behavioral and genomic studies of behavioral traits, though often quite detailed in their behavioral and biological scope, most often focus their efforts on identifying individual differences within a single inbred mouse strain for practical reasons (Pascoli et al., 2018; Walker et al., 2018) or within outbred rat populations that often ship from multiple vendors (Fitzpatrick et al., 2013), each with their own genetic bottlenecks. Sex differences are frequently ignored in both human and non-human animal research (Datta et al., 2020). Experimental methodologies agnostic to these known sources of individual differences can produce valuable biological insights in some circumstances, but the omission of sex and genetic variation in these studies limits generalizability without necessarily reducing experimental noise (Prendergast et al., 2014; Tuttle et al., 2018). Further, using a single inbred genetic background can suppress behavioral effects observed in even highly penetrant knockout alleles (Sittig et al., 2016). On the other hand, systematic inclusion of tractable genetic diversity may allow rodents to not just better emulate clinically relevant characteristics, but to contribute new genetic paradigms of understanding neuropsychiatric disorders (Neuner et al., 2019). These important advances in model organism genetics are often ignored in discussions

of how to best model neuropsychiatric phenotypes in non-human animals (e.g. Nestler and

Hyman, 2010).

As highly prevalent behavioral disorders, substance use disorders drive a public health crisis

associated with substantial morbidity and mortality. Illicit substance use disorders afflict

approximately 1 in 14 young adults in the United States (SAMHSA, 2017) and drug overdoses

are now the leading cause of accidental death among American adults under 55 (Kochanek et

al., 2017). Genetic variation and sex differences are both known to influence addiction

vulnerability; cocaine use disorder is highly heritable ($H^2 \approx 0.71$) (Goldman et al., 2005) and

substance use behaviors show sex differences in both humans and other animals (Becker et al.,

2012; Becker and Chartoff, 2019). Consequently, the neurobiology underlying addiction cannot

be fully understood without consideration of genetic background and sex in both humans and

non-human animals.

Given the genetic diversity in the population of inbred mouse strains (Beck et al., 2000) and the

multitude of derived mouse recombinant inbred and heterogeneous stock populations useful

for genetic mapping and trait correlation work (Chesler et al., 2005; Logan et al., 2013; Philip et

al., 2011), it is possible to sample over diverse genotypes and identify strains and sexes that

best mimic the human disease state. Such a strategy can improve translational relevance while

surveying these populations for heritable drivers of unidentified genetic mechanisms of disease

susceptibility. Though rodent genetic variation does not capture precise human variants, it can

be exploited to determine underlying mechanisms in addiction-relevant processes (Bogenpohl

et al., 2017; Huggett et al., 2020; Palmer et al., 2019). As a further benefit, genotypic and

phenotypic precision, along with high minor allele frequencies and in many cases, a well-

randomized population structure, allows genetics studies in rodents to be performed at orders

of magnitude lower cost than human GWAS.

To assess the influence of genetic variation, sex, and their potential interactions on cocaine-

related phenotypes in mice, we undertook a large-scale evaluation of behavioral, physiological,

and brain transcriptomic measures in both male and female mice from the eight inbred founder

strains of the Diversity Outbred (DO) mouse heterogenous stock (Saul et al., 2019b). Because

these strains include wild-derived inbred strains from the three dominant subspecies of mice,

their genomes together capture approximately 90% of the genetic diversity in the species *Mus*

musculus (Roberts et al., 2007). We further assessed how strain and sex differences drive

consilience with human addiction, identifying which strains and sexes capture significant

overlapping brain molecular correlates with human cocaine use disorder.

In the eight founder strains for the DO, we surveyed behavioral and physiological correlates of

vulnerability to cocaine use – including multiple novelty response behaviors, circadian

molecular rhythm phenotypes, and reversal learning as a measure of reward learning and

impulsivity. We directly measured cocaine-related behaviors such as initial locomotor

sensitivity to cocaine and intravenous self-administration (IVSA) of cocaine. Finally, using RNA

sequencing (RNAseq), we measured the striatum transcriptome response to cocaine in all eight

founder strains and compared them to a meta-analysis of postmortem brain tissue from

cocaine use disorder patients. Our work represents the first time that many of these behavioral,

physiological, and molecular traits have been studied using methods powered to detect sex and

genotype effects and the extent to which these interact to moderate one another's effects.

Results and Discussion

We first examined heritability, sex differences, and genetic differences moderated by sex

among the founders of the DO. Significant effects are reported in the text; all tests are reported

in **Supplementary Table S1.**

Response to novelty predicts psychostimulant addiction-related phenotypes in both humans

(Ersche et al., 2010) and mice (Dickson et al., 2015). We first assessed differences in behavioral

traits related to exploration and response to novelty using the open field, light-dark box, hole

board and novel place preference tests. Though novel place preference heritability was

moderately weak ($H^2 = 0.16$), the other three phenotypes displayed strong to very strong

heritability ($H^2 = 0.42$-0.55, **Figure 1A-D**). Further, for the phenotype of total entries in the hole

board test, an exploratory behavior, there was a significant sex difference ($F_{1,597} = 10.69$, p =

0.0014), which was apparently driven by lower hole board exploration in males than females

for all strains but PWK/PhJ. We detected strain-by-sex interactions in the total entries in the

hole board ($F_{7,597} = 2.74$, p = 0.0083) and in the proportion of distance traveled in the center of

the open field ($F_{7,684}$ = 2.09, p = 0.043), indicating that sex differences in these novelty response

traits are moderated by genetic background. Total entries in the hole board (**Figure 1A**) and

novel place preference (**Figure 1B**) exhibited extended range due to the inclusion of wild-

derived strains, demonstrating that wild-derived genetic variability functions in defining an

expanded phenotypic range (Wahlsten et al., 2003). Transitions between the sides of the light-

dark box also showed phenotypes toward the extreme in wild-derived mouse strains along with

NOD/ShiLtJ mice (**Figure 1D**).

Circadian rhythm and reward-related behavioral phenotypes are co-inherited and rhythm

disruptions are linked to development and progression of substance use disorders (Logan et al.,

Figure 1: Heritable differences in cocaine-related behavioral and physiological traits. All plots show the mean ±
the standard error in both sexes with females on the left. **A)** Hole board total entries, **B)** Novel place preference
(% time in novel zone), **C)** Open field (% time in center of open field), **D)** Transitions between light and dark in
the light-dark box, **E)** Circadian rhythm fibroblast *Bmal1-dLuc* luminescence amplitude, **F)** Reversal learning
premature responses to the hole rewarded during acquisition, **G)** Initial locomotor sensitivity to cocaine (cm
moved on Day 3 – Day 2), **H)** Cocaine intravenous self-administration number of infusions at 1.0 mg/kg FR1,
and **I)** Sessions to acquisition of cocaine intravenous self-administration.

2014). Further, the molecular clock system directly influences the expression of dopamine

receptors in the striatum involved in the modulation of cocaine reward-related behaviors

(Ozburn et al., 2015). A cell-based assay on fibroblasts derived from each of the founder strains

in which a *Bmal1-dLuc* reporter was utilized for circadian measurement of luciferase

bioluminescence to assess differences in circadian rhythmicity (Kim et al., 2016; Ramanathan et

al., 2014). We found very high heritability of the amplitude of these rhythmic patterns (H^2 =

0.59, **Figure 1E**), but no significant sex differences or strain x sex interactions. These results

suggest that genetic differences in the molecular clock are one potential mechanism for

individual differences in addiction-related phenotypes.

Reversal learning tasks evaluate impulsive and compulsive behaviors that predict addiction

liability (Izquierdo and Jentsch, 2012). Within reversal learning paradigms, one measure of

impulsivity is the number of premature responses produced during the reversal phase of the

task. This phenotype is similar to an impulsivity-related measure implemented in the five choice

serial reaction time task (Dalley et al., 2007). Premature reversal responses in our study showed

moderate heritability across our eight strains of mice (H^2 = 0.29, **Figure 1F**), but no significant

sex differences or strain x sex interactions. We observed the lowest rate of premature

responding in the wild-derived PWK/PhJ strain.

We next sought to assess individual differences in initial locomotor response to cocaine, a

behavioral phenotype that predicts subsequent drug use in humans (de Wit and Phillips, 2012).

In these diverse mouse strains, initial cocaine sensitivity showed strong heritability (H^2 = 0.48

for initial sensitivity, **Figure 1G**). Wild-derived strains, particularly WSB/EiJ and PWK/PhJ,

exhibited the highest initial sensitivity to cocaine.

Operant drug self-administration procedures directly quantify reinforced responding for drug

(Dickson et al., 2015) and produce profound transcriptional responses in inbred strain (Walker

et al., 2018). Heritability of cocaine IVSA traits in the founders of the DO was strong (H^2 = 0.47

for sessions to acquisition of IVSA, **Figure 1H**) to very strong (H^2 = 0.60 for total infusions at FR-1

1.0 mg/kg, **Figure 1I**). The high heritability of infusions earned is similar in magnitude to the

observed heritability of human cocaine use disorder (Goldman et al., 2005). Of note,

129S1/SvImJ mice do not acquire IVSA and do not take any cocaine during the acquisition

phase. The largest phenotypic range manifests in wild-derived mice; PWK/PhJ mice acquired

IVSA behavior very quickly and self-administered the most infusions of cocaine at FR-1 1.0

mg/kg – about four times as much as C57BL/6J – while WSB/EiJ mice self-administered the

fewest infusions of cocaine relative to

the other strains that do acquire IVSA

behavior. The broad phenotypic range

of volitional cocaine-taking behaviors in

these genetically diverse mouse strains

represents a clear opportunity to study

mechanisms underlying variation in the

initiation of cocaine of addiction.

Because addiction-related phenotypes

are highly heritable and sometimes

exhibit sex differences moderated by

genetics, we next sought to assess

heritable differences in the molecular

response to cocaine. Male and female

mice of the eight DO founder strains

were given multiple injections of

cocaine or saline over 19 days. Samples

for bulk RNAseq were collected from

striatum 24-48 hours after the final

injection of repeated administration of

either cocaine or vehicle (sham). First,

we documented that in the absence of

cocaine, most transcripts exhibited

expression that was moderate to

strong at baseline in the founder

strains (median H^2 = 0.29) and that

Figure 2: Individual differences in striatum cocaine response transcriptomes. **A)** High heritability of baseline transcriptome partitions the subspecies in a multidimensional scaling plot. Colors in Panel A carry forward to other figure panels. **B)** In a three-way analysis of strain, sex, and cocaine, many genes showed significant expression changes in response to cocaine in interaction factors; the strain x sex x cocaine interaction factor identified 1,282 DEGs. In contrast, there were very few significant DEGs in the main effect of cocaine. **C)** Two-way models show that the magnitude of transcriptional responses to cocaine differs by strain. B6 has few differentially expressed genes while 129 and PWK have many. The y-axis is $\log_{10}$. **D)** The serotonin 1D receptor gene *Htr1d* is differentially expressed in males of 129S1/SvImJ, PWK/PhJ, and CAST/EiJ strains. CAST/EiJ males show the opposite directionality of differential expression to PWK/PhJ and 129S1/SvImJ males.

subspecies of origin explains the greatest amount of variance in expression (**Figure 2A**). Using a

linear modeling approach (Chen et al., 2014; Phipson et al., 2016), we found few expression

differences attributable solely to the effect of cocaine treatment. Instead, significant effects of

cocaine arose in interaction with individual differences such as genetic background, sex, and

their interaction. The greatest quantity of differentially expressed transcripts was observed in

the strain-by-sex-by-drug treatment three-way interaction (1,282 genes at q < 0.01, **Figure 2B,**

Supplementary Table S2). Some strains showed stronger cocaine effects as measured by

number of differentially expressed genes than others; 129S1/SvImJ and PWK/PhJ had many

genes whose expression is influenced by cocaine and sex-by-cocaine interactions (PWK/PhJ: 89

genes at q < 0.01, **Figure 2C**) while the commonly used C57BL/6J strain had very few genes

influenced by cocaine (four genes at q < 0.01, **Figure 2C**). Because statistical power was

approximately equal for all strains involved in this study, these differences likely reflect real

individual differences in the brain's sex-specific responses to cocaine.

This genetics- and sex-inclusive approach identified many more changes in gene expression

following repeated cocaine exposure than had we used only a single sex in a single inbred

strain. For instance, we identified differential expression of the serotonin receptor 1D gene

Htr1d in the strain-by-sex-by-cocaine interaction factor. This gene, while not altered in

C57BL/6J animals, is upregulated in males of 129S1/SvImJ and PWK/PhJ strains, but

downregulated in males of the CAST/EiJ strain (**Figure 2D**).

Some of the heritable transcriptome differences we observed corroborate previous work

showing heritable differences in pharmacokinetics of cocaine (Wiltshire et al., 2015) – for

example, PWD/PhJ mice – closely related to the high cocaine taking PWK/PhJ strain – are

known to have some of the highest brain concentrations of cocaine shortly after injection.

These pharmacokinetic differences do not account for the strong differences between strains

such as 129S1/SvImJ and A/J, whose brain cocaine pharmacokinetic profiles are very similar

(Wiltshire et al., 2015). Further research on highly diverse mice may resolve the mechanisms

driving individual differences between strains with closely matched pharmacokinetic profiles.

We assessed to what degree strain and sex combinations mimic human genomic findings to

understand how individual differences' influence on consilience with human research. We used

Figure 3: Consilience differs by strain and sex. **A)** According to MSET p-values, a measure of higher-than-expected overlap, NOD/ShiLtJ female (left) and WSB/EiJ male (right) transcriptome differential expression shows statistically significant overlap with the union of multiple human cocaine use disorder brain transcriptome differential expression sets (multiple testing correction significance: ▪ = q < 0.20, * = q < 0.05). **B)** Overlapping gene lists show concurrent directionality. NOD/ShiLtJ female mice display increased expression of immediate early genes while WSB/EiJ males display decreased expression of oxidative phosphorylation genes.

the MSET method (Eisinger et al., 2013) to identify statistically significant overlaps of

orthologous genes between a human cocaine use disorder brain transcriptome dataset and

analogously analyzed cocaine response transcriptomes within each individual strain and sex

combination (Huggett et al., 2020). The magnitude of overlap was highly variable; the strongest

overlaps were in NOD/ShiLtJ female and WSB/EiJ male mice, though the NOD/ShiLtJ female

finding is not robust to different significance thresholds and does not survive stringent multiple

testing correction (**Figure 3A, Supplementary Table S3**. Other strains, including the most widely

used C57BL/6J, do not strongly mimic cocaine use disorder brain transcriptomes, a finding

consistent with previous observations of C57BL/6J males (Huggett et al., 2020). When we

examined the directionality of differential expression overlaps in NOD/ShiLtJ females and in

WSB/EiJ males, we found concordance in the upregulated and downregulated genes between

human and mouse data; in this experiment, WSB/EiJ males mostly captured downregulated

aspects of expression while NOD/ShiLtJ females mostly captured upregulated aspects (**Figure**

3B). Genes within the NOD/ShiLtJ female set include directional matches in immediate early

genes such as *Arc*, and *Junb* while genes downregulated in WSB/EiJ males include multiple

oxidative phosphorylation-related genes such as *Ndufa5*, *Cox6c2*, and *Ndufaf2*. The presence of

two mitochondrial Complex I subunit genes is particularly interesting; cocaine treatment

disrupts Complex I in the rat brain (Cunha-Oliveira et al., 2013) and a Complex I subunit gene is

associated with cocaine use disorder in humans (Huggett and Stallings, 2020a).

To describe the molecular systems dysregulated after repeated cocaine exposure in these

strains, we performed separate Gene Ontology Biological Process (GO BP) analysis of the genes

shared between human cocaine use disorder and WSB/EiJ male or the NOD/ShiLtJ female

differentially expressed gene sets. WSB/EiJ male differentially expressed genes were enriched

mostly for RNA binding and mRNA processing while NOD/ShiLtJ female differentially expressed

genes were enriched for nucleobase-related metabolic processes (**Supplementary Table S5**.

Together, these results imply that similarities mostly capture alterations in transcriptional

regulatory components.

Because the transcriptome results derive from the same group of mice as cocaine locomotor

sensitivity, it is of particular note that WSB/EiJ mice show the strongest behavioral response in

cocaine-induced locomotor sensitivity of all the strains tested (**Figure 1G**). Variants that alter

initial sensitivity to drugs are among the best supported and described addiction candidates in

humans, for example, nicotinic acetylcholine receptor polymorphisms associated with smoking

alter the receptor's sensitivity to nicotinic agonists (Bierut et al., 2008). This result implies that

WSB/EiJ male mice experience human-relevant alterations in their neurobiological states due to

differential cocaine sensitivity after repeated cocaine injection. However, WSB/EiJ mice take

fewer infusions of cocaine at FR-1 1.0 mg/kg than any other strain that reliably acquires IVSA

(**Figure 1H**). Though the finding of higher consilience associated with lower cocaine-taking

behavior appears to contradict the conventional interpretation of cocaine self-administration,

where higher cocaine intake indicates higher addiction vulnerability (Piazza et al., 2000), we

note that experimenter-administered cocaine affects neural signaling differently than self-

administered cocaine (McCutcheon et al., 2011). Consequently, this finding should demonstrate

that individual differences are highly important to consider for translational research using non-

human animals, but should not be taken as a specific recommendation of WSB/EiJ males as a

model for human cocaine use disorder phenotypes.

The translational relevance of non-human animals – often represented by mice – is the subject

of substantial discussion (Seok et al., 2013; Takao and Miyakawa, 2015). Within addiction

biology, a particularly active source of ferment is the question of how rodents may best

recapitulate aspects of addictive behaviors such as compulsive use and reinstatement (Ahmed,

2012). Here, we further this discussion by demonstrating that similarity of rodent molecular

systems to those involved in human neuropsychiatric-traits are heterogeneous and contingent

upon the specific rodents chosen. Within conventional inbred mouse strains, NOD/ShiLtJ

females and WSB/EiJ males showed the highest consilience with human cocaine phenotypes in

the brain as measured by similarity in genomic response. This finding has broad implications for

translational work. In many cases, the study of a single strain and/or only one sex of mice

within an experiment may limit the generalizability of the data and diminish potential relevance

for human addiction. Rather, it may be advantageous to explicitly take individual differences of

the type reported here into account. Strategies for querying diversity include identification of

specific strains that exhibit characteristics of the disease-susceptible population at a behavioral

or molecular level or surveys of a diverse population to ensure that natural variation is

represented, which allows queries of known biological sources of variation in addiction

vulnerability.

An important aspect of experimental rigor is the external validity of model organism research

paradigms. Here we show that this validity is in part driven by the organism under investigation

in addition to other characteristics of the research paradigm. Genetic variation is a valuable

resource for the discovery of biological mechanisms of addiction (Kumar et al., 2013; Ruan et

al., 2020). Similar to humans, individual differences among mice greatly influence behavioral,

physiological, and transcriptomic cocaine-related traits. For many of these traits, individual

differences explain a substantial proportion of the variation. This variation can be exploited to

enhance consilience of biological systems under study, or to discover the underlying biological

mechanisms of vulnerability to disease. Individual variation in addiction-related traits is a

largely untapped resource that can be exploited to improve and accelerate discovery of

neurobiological and genetic mechanisms related to risk for addiction as well as other complex

diseases.

Methods

Standard Operating Procedures

291 All methodologies used in this work are documented in depth in the Center for Systems

292 Neurogenetics of Addiction's Standard Operating Procedures (SOPs)

293 (https://www.jax.org/research-and-faculty/research-centers/systems-neurogenetics/data-

294 resources). The brief methods appearing below summarize these SOPs and reference specific

295 SOPs.

Animals

Mice from the following strains were surveyed in these experiments: A/J (JAX stock #000646),

C57BL/6J (JAX stock #000664), 129S1/SvImJ (JAX stock #002448), NOD/ShiLtJ (JAX stock

#001976), NZO/HlLtJ (JAX stock #002105), CAST/EiJ (JAX stock #000928), PWK/PhJ (JAX stock

#003715), and WSB/EiJ (JAX stock #001145). These strains are the founders of the DO

heterogeneous stock and CC recombinant inbred strains. Surveys of these eight strains can

demonstrate statistical heritability patterns that justify further dissection using the derived

resources (Saul et al., 2019b). The mice used in the Research Animal Facility at The Jackson

Laboratory came from breeding colonies maintained in the Research Animal Facility. These

colonies were derived from production colonies at The Jackson Laboratory and breeders were

replaced with animals from The Jackson Laboratory's production colony at least every five

generations. The mice used in the Jentsch Lab at Binghamton University were shipped to the

Jentsch Lab from either the Research Animal Facility colonies or from production colonies in

The Jackson Laboratory. The studies described utilized a total of 1,085 mice (**Supplementary**

Table S4). All procedures were approved by the Jackson Laboratory of Mammalian Genetics or

Binghamton University institutional animal care and use committees.

Mouse Husbandry and Housing

Mice were housed according to the CSNA animal housing SOP (https://www.jax.org/-/media/jaxweb/files/research-and-faculty/tools-and-resources/system-neurogenetics/csna-animal-housing.pdf).

Cocaine

Cocaine hydrochloride was provided by the National Institute on Drug Abuse Drug Supply Program Division of Therapeutics and Medical Consequences (catalog number: 9041-001). Cocaine was stored in powder form at room temperature until it was formulated into 0.9% Saline (100 mg/mL clear solution) in various concentrations specific to each experiment according to their individual SOPs.

Novelty Response Behavioral Phenotypes

Open field, light-dark box, hole board, and novel place preference behavioral paradigms were conducted in this order on consecutive days over the course of a week during the light phase of the light:dark cycle. Open field data were collected for 60 minutes according to the SOP (https://www.jax.org/-/media/jaxweb/files/research-and-faculty/tools-and-resources/system-neurogenetics/open-field-assay.pdf?la=en&hash=32DDAFF2B17B2D4961C136C5616C4982AC23EC3B). Light-dark data were collected for 20 minutes with the mouse starting in the light side of the chamber facing the dark side according to the SOP (https://www.jax.org/-/media/jaxweb/files/research-and-faculty/tools-and-resources/system-neurogenetics/light-dark-assay.pdf?la=en&hash=A63CF8D22EB7936CF6C69A3178373981F4016675). Hole board data were collected for 20 minutes according to the SOP (https://www.jax.org/-/media/jaxweb/files/research-and-faculty/tools-and-resources/system-neurogenetics/hole-board-assay.pdf?la=en&hash=EC343A797D37209CF64D34E6031608A511D8E15D). Novel place preference included a five minute acclimation period to a center chamber, a 10 minute exposure period to a randomized exposure side, a five minute acclimation period, and a final test period consisting of a five minute habituation period again to the center and a 20 minutes preference assessment for which both the novel side and the initial familiar exposed side were

accessible according to the SOP (https://www.jax.org/-/media/jaxweb/files/research-and-

faculty/tools-and-resources/system-neurogenetics/novelty-place-preference-

assay.pdf?la=en&hash=B5D2D0FC9028B408E84729C0C8832C580AB8E039). All mice from the

Center for Systems Neurogenetics of Addiction were tested through this novelty pipeline prior

to any other test and were then randomized and assigned into either reversal learning, cocaine

locomotor sensitization, or cocaine intravenous self-administration. The novelty study

produced observations from a total of 783 mice.

Bmal1-dLuc Circadian Rhythm Data

Data for circadian rhythm were measured in primary fibroblast cultures generated from skin

biopsies in the founders. To isolate fibroblasts, ear biopsies (one mm in diameter) were

digested in Dulbecco's Modified Eagle's Medium (DMEM, HyClone) containing 2.5 mg/ml

collagenase D (Gibco) and 1.25 mg/ml pronase (Millipore) for 90 mins and then plated in DMEM

growth media containing 10% Fetal Bovine Serum (FBS, HyClone), 292 μg/ml L-glutamine

(HyClone), 100 units/ml penicillin (Hyclone) and 100 μg/ml streptomycine (HyClone). *Bmal1-*

dLuc reporter was delivered to fibroblasts by lentiviral-mediated gene delivery (VectorBuilder).

Following synchronization of rhythms by 15 μM forskolin (Sigma) for two hours, the temporal

patterns of *Bmal1-dLuc* bioluminescence was recorded for ~70 seconds at intervals of 10

minutes over six to seven days from fibroblast cultures in DMEM recording media containing 15

μM forskolin, 25 mM HEPES (Gibco), 292 μg/ml L-glutamine, 100 units/ml penicillin, 100 μg/ml

streptomycine, and 10 μM luciferin (Promega) by an automated 32-channel luminometer

(Lumicycle, ActiMetrics) in a standard tissue culture incubator at 32°C. The amplitude of

bioluminescence rhythms was determined from baseline-subtracted data using the damped

sine fit and Levenberg-Marquardt algorithm (Izumo et al., 2003). The circadian study produced

observations from a total of 56 mice.

Reversal Learning

Data for reversal learning were collected at both JAX and Binghamton University using the SOP

(https://www.jax.org/-/media/jaxweb/files/research-and-faculty/tools-and-resources/system-

neurogenetics/reversal-learning-

assay.pdf?la=en&hash=8484E47B170462960E11C1FAEEE6FF3CE6FDFC08). The reversal

learning data produced observations from a total of 202 mice.

Initial Locomotor Sensitivity to Cocaine

Data for initial locomotor sensitivity were collected as described previously (Schoenrock et al.,

2020) using data from days 1-3 in the SOP for locomotor behavioral sensitization to cocaine

(https://www.jax.org/-/media/jaxweb/files/research-and-faculty/tools-and-resources/system-

neurogenetics/cocaine-locomotor-sensitization-

assay.pdf?la=en&hash=9E5D4C248C3BCCAD947C164AE81663C13A77EB0D). Briefly, mice were

placed into the open field arena for 30 minutes, removed, and injected i.p. with either saline

(days 1-2) or 10 mg/kg cocaine (day 3) and returned to the open field arena for 60 minutes.

Distance moved after injection on day 3 minus day 2 was uses as a measure of initial locomotor

sensitivity to cocaine. The sensitization study produced observations from a total of 230 mice.

Cocaine Intravenous Self-Administration

Prior to cocaine intravenous self-administration, mice were implanted with a jugular catheter

and allowed a minimum of 10 days for post-operative recovery. In an operant conditioning

paradigm, mice were allowed to acquire cocaine self-administration at 1.0 mg/kg, then

evaluated for dose-response effects at eight different doses. After a stabilizing dose at 1.8

mg/kg, extinction-related responses during seven days of withdrawal were recorded. Finally,

cued reinstatement was recorded for two days. Self-administration in these eight mouse strains

was performed according to v1.0 of the CSNA's SOP (https://www.jax.org/-

/media/jaxweb/files/research-and-faculty/tools-and-resources/system-

neurogenetics/intravenous-self-administration-ivsa-

paradigm.pdf?la=en&hash=FA64135F219C7DF65937A1CF9270301B0E771836). The

intravenous self-administration study produced observations from a total of 217 mice.

Data Deposit

393   Data for each phenotype will be deposited in the Mouse Phenome Database (MPD) (Bogue et

394   al., 2019) upon publication.

395 Heritability Calculations

396 For each trait, heritability was calculated from linear models using the isogenic strain as the

397 independent categorical variable using the following equation:

398
$$h^2 = \frac{MS_{strain}}{MS_{strain} + (n_{mean} - 1) * MS_{resid}}$$

where MS_{strain} is the mean square of the strain effect, n_{mean} is the mean number of samples

within each strain, and MS_{resid} is the mean square of the residuals. For the reversal learning

data, an additional additive covariate of site was included in the model to account for inter-lab

variation. This term was not utilized in the heritability calculation.

For some traits such as number of infusions at FR-1 1.0 mg/kg cocaine self-administration, a

single strain such as 129S1/SvImJ showed little to no variation, which may upwardly bias

heritability calculations. For these traits, heritability was calculated both with and without the

low variance strain. The data reported in the paper rely upon the inclusive calculation, but

results of both methods of calculation are reported for completeness (see **Supplementary**

Table S1).

RNAseq

Striatum tissue was collected during the light stage of the light:dark cycle between 24 and 48

hours after the final injection in the cocaine behavioral sensitization protocol according to the

SOP (https://www.jax.org/-/media/jaxweb/files/research-and-faculty/tools-and-

resources/system-neurogenetics/post-sensitization-tissue-

collection.pdf?la=en&hash=9E6CD8DEB39606B791A5D25F6CD0611EF14D96A7). Tissue was

collected for both sexes of each founder strain exposed to either sham (saline) or 10 mg/kg IP

cocaine.

RNA was isolated from striatum tissue using the MagMAX mirVana Total RNA Isolation Kit

(ThermoFisher) and the KingFisher Flex purification system (ThermoFisher). Tissues were lysed

and homogenized in TRIzol Reagent (ThermoFisher). After the addition of chloroform, the RNA-

containing aqueous layer was removed for RNA isolation according to the manufacturer's

protocol, beginning with the RNA bead binding step.

RNA concentration and quality were assessed using the Nanodrop 2000 spectrophotometer

(Thermo Scientific) and the RNA Total RNA Nano assay (Agilent Technologies). 2µl of diluted

1:1000 diluted ERCC Spike-in Control Mix 1 (Ambion by Life Technologies) was added to 100ng

of each RNA sample prior to library construction. Libraries were prepared by the Genome

Technologies core service at The Jackson Laboratory using the KAPA RNA Hyper Prep Kit with

RiboErase (HMR) (KAPA Biosystems), according to the manufacturer's instructions. Briefly, the

protocol entails depletion of ribosomal RNA (rRNA), RNA fragmentation, first and second strand

cDNA synthesis, ligation of Illumina-specific adapters containing a unique barcode sequence for

each library, magnetic bead size selection, and PCR amplification. Libraries were checked for

quality and concentration using the D5000 ScreenTape assay (Agilent Technologies) and

quantitative PCR (KAPA Biosystems), according to the manufacturers' instructions.

RNAseq libraries were pooled and sequenced by Novogene in 150 bp paired-end format on an

Illumina NovaSeq 6000 sequencer targeting 90 million read pairs per sample. Sequencing

achieved a median read depth of 132 million reads. The resultant reads were determined to be

of consistently high quality using fastqc v0.11.3 and MultiQC v1.2.

Reads were generated from raw data and demultiplexed using BCL2Fastq v2.18.0.12,

concatenated by sample, and aligned with the STAR aligner v2.6.1 (Dobin et al., 2013) to the

GRCm38 mouse reference genome with v94 of the Ensembl transcriptome. Transcript-level

quantification was estimated using RSEM v1.3.0 (Li and Dewey, 2011) on a transcriptome BAM

file produced as an output of this alignment. The data were imported into R v3.5.1 and

summarized to the gene level using tximport v1.10.1 (Soneson et al., 2016), TMM-normalized

using edgeR v3.24.3 (Chen et al., 2014), and imported into limma v3.38.3 (Ritchie et al., 2015)

using the $\log_2$-transformation function voom. We compared multivariate approaches modeling

with interaction factors between edgeR and voom+limma approaches and found that

voom+limma performs better than edgeR for controlling false negatives. Upon initial

examination of the findings, we identified intermittent contamination with choroid plexus,

which potentially derives from the ventricular aspect of the dorsal striatum. Correcting for this

contamination necessited an additive covariate for choroid plexus consisting of log-mean CPM

values of *Kl* and *Ttr* expression, unambiguous markers for choroid plexus (Sathyanesan et al.,

2012). These values were log$_2$ transformed for work in limma. For An overall model for all

strains included this choroid plexus factor as a nuisance variable plus the main effects of strain,

sex, and cocaine injection and all of their interactions. Individual models included the choroid

plexus nuisance variable plus sex, cocaine injection, and sex:cocaine injection interaction.

Correction for local false discovery rates utilized the qvalue package in R v2.14.1 (Storey and

Tibshirani, 2003). Because brain transcriptional changes are subtle (Hitzemann et al., 2014), all

results reported are at q < 0.01 with no fold-change cutoff (**Supplementary Table S2**).

Raw data and transcript-level expression estimates will be deposited in the Gene Expression

Omnibus (Barrett et al., 2012) upon publication (accession number: GSEXXXXX).

Cross-Species Gene List Comparison

To assess the molecular correspondence of mouse cocaine self-administration with human

cocaine use, we compared the results of the current study to differentially expressed genes

(BH-FDR < 0.05) associated with cocaine use disorder (CUD) in the midbrain (n = 20, 50% CUD,

M_{AGE} = 49.2, s.d. = 3.9; (Bannon et al., 2014), microarray), hippocampus (n = 15, 46.7% CUD,

M_{AGE} = 39.4, s.d.$_{AGE}$ = 39.4; Huggett and Stallings, 2020a, 2020b, RNA-sequencing) and dlPFC

neurons (n = 36, 52.7% CUD, M_{AGE} =35.0, s.d.$_{AGE}$ = 11.0; (Huggett and Stallings, 2020a); neuron-

specific RNA-sequencing). The aforementioned studies utilized methods that maximized power

for identifying differentially expressed genes and used case/control analyses that compared

individuals with CUD to matched cocaine free controls. A list of all the differentially expressed

genes can be found on GeneWeaver (https://www.geneweaver.org/; GS398242).

A key of 1:1 orthologs between humans and mice was generated from the MGI Vertebrate

Homology indices (accessed 2020-05-18). This key was used to compare the human gene set to

contrast tables within strain and sex combinations. To perform these comparisons, we used the

MSET algorithm (Eisinger et al., 2013) as contained in v1.16.6 of the msaul/msaul R package

(https://github.com/msaul/msaul). MSET p-values were corrected for multiple comparisons

using the qvalue package in R v2.14.1 (Storey and Tibshirani, 2003). Gene Ontology Biological

Process (GO BP) analysis on overlapping genes was performed on mouse Ensembl identifiers

using the AmiGO v2 web tool (accessed 2020-05-28) on GO Ontology Database

doi:10.5281/zenodo.3727280 with Fisher's exact tests and Bonferroni correction settings

(Carbon et al., 2009).

Supplemental Material

Supplementary Table S1: Statistical test results for heritability and genotype-by-sex

interactions.

Supplementary Table S2: Differential expression results for all tests discussed.

Supplementary Table S3: Quantification of overlap significance between human cocaine use

disorder transcriptome and individual strain and sex cocaine transcriptomes.

Supplementary Table S4: Sample sizes for all strain and sex combinations for studies reported.

Supplementary Table S5: Gene Ontology Biological Process results for WSB/EiJ male and

NOD/ShiLtJ female overlapping gene sets.

Raw behavioral data will be deposited in the Mouse Phenome Database upon publication.

Gene expression data will be deposited in the Gene Expression Omnibus upon publication.

All scripts, code, and metadata used for analysis are deposited in GitHub (repository:

github.com/msaul/csna_founders_survey_2020)

Author Contributions

EJC, PED, LMT, SAS, JDJ, RWL, CAM, LGR, VMP, and SJSR conceived the studies. SJSR, PED, JRB,

LSB, SAS, RD, ML, AO, TR, TW, and LHG designed and implemented the behavioral experiments.

SMK designed and implemented the circadian transcriptional experiment. JRB, LSB, UD, PED,

ML, SMK, AO, TR, SAS, TW, LHG, VMP, and MCS analyzed the behavioral data. SMK, MCS, and

VMP analyzed the circadian transcriptional data. MCS, VMK, and VMP analyzed the RNAseq

data. MCS, UD, PED, SBH, JRB, LSB, SAS, TW, LHG, VMP, and EJC interpreted the behavioral

results. SMK, MCS, and VMP interpreted the results of the circadian transcriptional experiment.

MCS, VMK, VMP, and EJC interpreted the RNAseq results. MCS, SBH, RHCP, and EJC conceived,

designed, analyzed, and interpreted the comparison between mouse and human gene sets.

MCS and EJC wrote the manuscript.

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