- Multivariate Analyses of Codon Usage of SARS-CoV-2 and other
- 2 betacoronaviruses
- 4 Haogao Gu¹, Daniel Chu, Malik Peiris, Leo L.M. Poon¹*
- 6 ¹School of Public Health, LKS Faculty of Medicine, The University of Hong Kong, Hong
- 7 Kong SAR

5

8

10

9 *Corresponding author: E-mail: llmpoon@hku.hk

Abstract

Coronavirus disease 2019 (COVID-19) is a global health concern as it continues to spread within China and beyond. The causative agent of this disease, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), belongs to the genus *Betacoronavirus* which also includes severe acute respiratory syndrome related coronavirus (SARSr-CoV) and Middle East respiratory syndrome related coronavirus (MERSr-CoV). Codon usage of viral genes are believed to be subjected to different selection pressures in different host environments. Previous studies on codon usage of influenza A viruses can help identify viral host origins and evolution trends, however, similar studies on coronaviruses are lacking. In this study, global correspondence analysis (CA), within-group correspondence analysis (WCA) and between-group correspondence analysis (BCA) were performed among different genes in coronavirus viral sequences. The amino acid usage pattern of SARS-CoV-2 was generally found similar to bat and human SARSr-CoVs. However, we found greater synonymous codon usage differences between SARS-CoV-2 and its phylogenetic relatives on spike and membrane genes, suggesting these two genes of SARS-CoV-2 are subjected to different evolutionary pressures.

Keywords: SARS-CoV-2; coronavirus; codon usage analysis; WCA.

30 Introduction 31 A novel coronavirus outbreak took place in Wuhan, Hubei province, China in December 2019¹. This novel coronavirus (SARS-CoV-2) causes pneumonia in patients² and it has 32 rapidly spread to other provinces in China and other countries³. This novel coronavirus 33 34 outbreak had raised global concern but current knowledge on the origin and transmission 35 route of the pathogen is still limited. The SARS-CoV-2 belongs to the genus *Betacoronavirus*, 36 which also includes two highly virulent human coronaviruses, SARS-CoV and MERS-CoV. 37 Apart from human, many animal species, such as bat, rat, camel, swine and hedgehog, can be 38 infected by different types of coronaviruses. Further sequence analyses of this novel and 39 other betacoronaviruses might provide additional information to better understand the 40 evolution of SARS-CoV-2. 41 Preferential codon usage is commonly seen in different organisms, and it has been evident 42 that the uneven codon usage is not neutral but related to gene expression or other selection pressures ^{4–6}. There are two levels of codon usage biases, one is at amino acid level and the 43 44 other is at synonymous codon level. The amino acid composition of proteins can be an 45 important factor that explaining certain sequence traits. For example integral membrane 46 proteins that are enriched in hydrophobic amino acids can create significant codon usage 47 bias'. Amino acid composition sometime can also introduce confounding effects when one 48 only focuses on studying the variations of synonymous codon usage. The use of global 49 correspondence analysis (CA) and its derivatives within-group correspondence analysis 50 (WCA) and between-group correspondence analysis (BCA) to analyze codon usages can 51 overcome the above problem. In fact, WCA becomes "model of choice" for analyzing 52 synonymous codon usage in recent years, as it is more robust than other traditional methods (e.g. CA with relative codon frequency or CA with RSCU values)^{7,8}. This analytic approach, 53 54 however, has not been used in studying viral sequences. As the natural history of the SARS-55 CoV-2 remains largely unknown, an in-depth codon usage analysis of this newly emerging 56 virus might provide some novel insights. 57 In this study, we used both CA and WCA to analyses codon usage patterns of a vast number 58 of betacoronavirus sequences. We found SARS-CoV-2 and bat SARSr-CoV have similar 59 amino acid usage. However, our analyses suggested that the spike and member genes of 60 SARS-CoV-2 have rather distinct synonymous codon usage patterns.

Methods

- 62 **Sequence data**
- To construct a reference sequence dataset, available full-length complete genome sequences
- of coronavirus were collected through Virus Pathogen Resource database
- 65 (https://www.viprbrc.org/brc/home.spg?decorator=corona, accessed 13 Jul 2019, ticket
- 66 958868915368). The sequences were filtered by the following steps: (1) Remove sequences
- 67 without protein annotation, (2) Keep only sequences with complete set of desired replicase
- and structural proteins (sequences coding for orflab, spike, membrane and nucleocapsid), (3)
- 69 Filter out sequences that are unusually long and short (>130% or <70% of the median length
- for each group of gene sequences), (4) Limit our analysis to genus *Betacoronavirus* and (5)
- 71 Concatenate orf1a and orf1b sequences to form orf1ab if necessary.
- 72 The final dataset comprised 769 individual strains (3076 individual gene sequences) that
- contain complete sets of coding regions for orflab, spike, membrane and nucleocapsid genes
- (see Supplementary Figure 1). The sequences for envelope gene were not included in the
- analysis because of the short length and potential bias in codon usage. Corresponding
- 76 metadata for the sequences were extracted by the sequence name field. 24 complete genome
- sequences of the newly identified SARS-CoV-2 and its phylogenetically close relatives were
- 78 retrieved from Genbank and GISAID (accessed 22 Jan 2020). Six genomes in this study were
- used as special references (BetaCoV/bat/Yunnan/RaTG13/2013|EPI_ISL_402131;
- 80 BetaCoV/pangolin/Guangxi/P1E/2017|EPI ISL 410539; MG772934.1 Bat SARS-
- 81 like_coronavirus_isolate_bat-SL-CoVZXC21; MG772933.1_Bat_SARS-
- 82 like_coronavirus_isolate_bat-SL-CoVZC45;
- 83 KY352407.1_Severe_acute_respiratory_syndrome-related_coronavirus_strain_BtKY72 and
- 84 GU190215.1_Bat_coronavirus_BM48-31/BGR/2008), as they have previously been reported
- to have close phylogenetic relationship with SARS-CoV- 2^{9-11} . Detailed accession ID for the
- above data are provided in the Supplementary Table S1.
- 87 The codon count for every gene sequence input for the correspondence analysis was
- 88 calculated by the SynMut¹² package. The implementation of the different correspondence
- analyses in this study was performed by functions in the package ade4¹³. Three stop codons
- 90 (TAA, TAG and TGA) were excluded in the correspondence analysis.

91 Global correspondence analysis (CA) on codon usage 92 Correspondence analysis (CA) is a dimension reduction method which is well suited for 93 amino acid and codon usage analysis. The concept in correspondence analysis is similar to Pearson's χ^2 test (i.e., the expected counts are calculated under the hypothesis of 94 95 independence, based on the observed contingency table). With the deduced expected count 96 table, the Euclidean distance or the χ^2 distance can be used to evaluate the difference between 97 two observations. The χ^2 distance that we are using in the global correspondence analysis is applied for the row profile (adjusted for the size effect among difference genes) and the 98 99 column profile (adjusted for the size effect among difference codons) and therefore the raw 100 codon count rather than the Relative Synonymous Codon Usage (RSCU) values are more 101 informative and suitable input for our model. The calculation of the χ^2 distance is included in 102 the Supplementary Method. 103 All the correspondence analyses in this study were performed individually for each gene, to 104 achieve better resolution on gene specific codon usage pattern. 105 Within-group correspondence analysis and between-group correspondence analysis 106 In contrast to the ordinary correspondence analysis, the within-block correspondence analysis ¹⁴ (WCA) can segregate the effects of different codon compositions in different 107 amino acids. WCA has been recognized as the most accurate and effective CA method for 108 studying the synonymous codon usage in various genomic profile⁸. WCA focuses on the 109 110 within-amino acid variability, and it technically excludes the variation of amino acid usage 111 differences. WCA was implemented based on the existing global CA, with additional 112 information for factoring. 113 Between-group correspondence analysis (BCA) is complementary to WCA; BCA focuses on 114 the between-group variability. BCA can be interpreted as the CA on amino acid usage. We 115 used BCA in this study to investigate the amino acid usage pattern in different coronaviruses. 116 Grand Average of Hydropathy (GRAVY) score Gravy score provides an easy way to estimate the hydropathy character of a protein¹⁵. It was 117 118 used in this study as a proxy to identify proteins that are likely to be membrane-bound 119 proteins. The GRAVY score was calculated in a linear form on codon frequencies as:

$$s = \sum_{i=1}^{64} \alpha_i f_i$$

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

148

149

150

Where α_i is the coefficient for a particular amino acid (provided by data EXP in Seqinr package¹⁶) encoded by codon i, f_i correspond to the relative frequency of codon i. **Results** General sequence features in *Betacoronavirus* A total of 3,076 individual gene sequences passed the filtering criteria and were included in this study. Viral sequences from 3 different species (Middle East respiratory syndrome related coronavirus (MERSr-CoV), Betacoronavirus 1, SARS related coronavirus (SARSr-Cov)) were the three most dominant species (see Supplementary Figure S1) in the filtered dataset. Four conserved protein sequence encoding regions of *Betacoronavirus* were analysed separately. The median lengths of the studied sequence regions were 21237 nt for orf1ab gene, 4062 nt for spike gene, 660 nt for membrane gene and 1242 nt for nucleocapsid gene. Spike gene has the lowest average and median G + C contents among these four genes (median: 37.45%, 37.31%, 42.60% and 47.22% for orf1ab, spike, membrane and nucleocapsid respectively). The G+C contents of the orflab and spike genes were found distributed in bi-modal patterns, and the G + C contents of SARS-CoV-2 were found located at the lesser half of the data of these two genes. The G + C contents for membrane and nucleocapsid genes of studied viral sequences were distributed in unimodal pattern (see Supplementary Figure S2). The overall amino acid and codon usage of the dataset are plotted in an ascending order (Figure 1). We observed that leucine and valine were the two most frequently used amino acids in the four studied genes, while tryptophan, histidine and methionine were the three least used ones. We also found that codons ending with cytosine or guanine were generally less frequent than the codons ending with adenine or thymine. This pattern of uneven usage in synonymous codons is in accordance with the G + C content distribution results (codons ending with guanine or cytosine were less frequently observed). We found a substantial bias in amino acid usage among these four genes, and this bias is well explained by the hydropathy of the encoded proteins (results from global correspondence analysis on all the four genes, collectively, data not shown). The GRAVY scores for every sequence were calculated to represent the degree of hydropathy. We discovered that the

151 nucleocapsid protein sequences had significantly lower GRAVY scores as compared to those 152 from other genes, while the membrane protein sequences had highest GRAVY scores (see 153 Supplementary Figure S3). 154 **Correspondence analysis** 155 We first conducted a multivariate analysis of codon usage on the dataset by using global 156 correspondence analysis. We also conducted WCA and BCA to study these sequences at 157 synonymous codon usage and amino acid usage levels, respectively. Given that there were 158 different amino acid usage biases among different genes (Supplementary Figure S3), we 159 performed correspondence analyses of these genes separately. 160 Of all the four correspondence analyses for the four genes, the extracted first factors 161 explained more than 50% of the total variance (see Supplementary Figure S4). The first two 162 factors in orf1ab global CA represented 67.7% and 16.8% of total inertia. Similarly, the first 163 two factors of the spike, membrane and nucleocapsid global CA represented 51.0% and 164 18.5%, 52.6% and 20.2%, and 54.8% and 14.2%, respectively, of total inertia. With only 165 these two factors, we could extract ~70% of the variability of the overall codon usage for each studied gene. These levels of representations were higher than or similar to those 166 deduced from other codon usage analyses^{8,17,18}. 167 168 The overall codon usage of SARS-CoV-2 in orf1ab, spike and membrane genes are 169 similar to those of bat and pangolin CoVs 170 Based on the above CA analysis, the data points are shown in different colours that represent 171 different features of the sequences (e.g. viral host or viral species). There were no 172 neighbouring human viruses around SARS-CoV-2 in CA results of orflab, spike and 173 membrane (Figure 2), suggesting that the overall codon usage of SARS-CoV-2 in the orf1ab, 174 spike or membrane gene was significantly different from those of human betacoronaviruses. 175 By contrast, the nucleocapsid genes of SARS coronavirus and SARS-CoV-2 are found to be 176 relatively similar (Supplementary Figure S5A). Except for the nucleocapsid gene, virus 177 sequences adjacent to the SARS-CoV-2 were all from bat coronaviruses (coloured in purple 178 in Figure 2). 179 There are five groups of viral sequences of human origin in the dataset (SARS-CoV-2, 180 Betacoronavirus 1, human coronavirus HKU 1, MERS-CoV and SARS-CoV). These five 181 groups of viral sequences were well separated from each other in terms of codon usage, 182 except the nucleocapsid gene sequences of SARS-CoV-2 and SARS-CoV as mentioned

187

188

189

191

195

201

209

214

above. There was no overlap between SARS-CoV-2 and human SARS-CoV in orf1ab, spike 184 and membrane, yet SARS-CoV codon usage processed more similar to SARS-CoV-2 185 compared to the other three types of human coronaviruses (i.e. yellow point always closest to 186 SARS-CoV-2 in Supplementary Figure S5A). Compared to human coronavirus sequences, the bat coronavirus sequences have more scattered codon usage, even within the same viral species (Supplementary S5B). Some viral species in bats formed their own clusters in all four genes (e.g. SARSr-CoV). SARSr-CoV is 190 a group of coronavirus that can be found in both humans and bats. We observed that the data points of human SARSr-CoV are clustered with those of bat SARSr-CoV in all the four genes 192 (by comparing the yellow points in Supplementary Figure S5A and S5B). The codon usage of 193 SARS-CoV-2 in orf1ab, spike and membrane were slightly different from the SARS-CoV 194 clusters and these data points are located in between SARSr-CoV and other coronavirus species (e.g. MERSr-CoV and bat coronavirus HKU9 etc.) 196 The global codon usages of bat RatG13 virus were found most similar to SARS-CoV-2 in 197 orf1ab, spike and nucleocapsid genes, but not in membrane gene (Figure. 2). In the analysis 198 of membrane protein, pangolin P1E virus had a more similar codon usage to SARS-CoV-2 199 than all the other viruses. We found the similarity in codon usage between pangolin P1E and 200 SARS-CoV-2 were also high in orf1ab, where P1E was the second closest data point to SARS-CoV-2. But this is not the case for spike and nucleocapsid genes. 202 We also observed that the codon usage pattern in spike gene was more complex than in other 203 genes. For example, data points adjacent to the spike gene of SARS-CoV-2 were 204 coronaviruses from bat, human and rodent hosts (Figure 2). The codon usage of rodent 205 coronaviruses was generally distinct from human or bat coronaviruses in orflab, membrane 206 and nucleocapsid gene sequences. By contrast, the spike gene sequences of murine 207 coronaviruses were found located between SARSr-CoV and other coronaviruses, just like 208 SARS-CoV-2 (Figure 2 and Supplementary Figure S6B). The codon usage from camel, swine and other coronaviruses were found to be well clustered and relatively distant to SARS-CoV-210 2 (see Supplementary Figure S6A, S5C, S5D). 211 The codon usage at synonymous level suggested novel patterns of SARS-CoV-2 in spike 212 and membrane genes 213 WCA and BCA were used to further differentiate codon usage of these betacoronaviruses at

synonymous codon usage and amino acid usage levels, respectively. After applying the row-

215 block structure to the original global CA model, we found that most of the variability in 216 codon usage can be explained at synonymous codon usage level (90.36% for orf1ab gene, 217 85.29% for spike gene, 83.71% for member gene and 84.07% for nucleocapsid gene) (Table 218 1). 219 Results from the BCA suggested that the amino acid usage of SARS-CoV-2 is closely related 220 to bat and human SARSr-CoVs in all four genes (Figure 3B and Figure 4B). Specifically, we 221 discovered that the SARS-CoV-2 had amino acid usage pattern most similar to bat RaTG13 222 virus, followed by pangolin P1E, bat CovVZC45 and bat CoVZXC21. The sequences of 223 BtKY72 and BM48-31 were from a more phylogenetically distant clade, and, accordingly, 224 they had relatively distinct amino acid usage to SARS-CoV-2 as expected in all four studied 225 genes. This result agrees with the result in the full-genome phylogenetic analysis 226 (Supplementary Figure S7). 227 The difference between SARS-CoV-2 and RaTG13 at synonymous codon usage level was 228 marginal in orflab and nucleocapsid sequences. Interestingly, there were noticeable 229 differences in the spike and membrane gene analyses. Our results suggest the synonymous 230 codon usage patterns in the spike and membrane gene of SARS-CoV-2 are different from 231 those of its genetically related viruses (i.e. RaTG13 and other reference relatives). For 232 example, the synonymous codon usage pattern of SARS-CoV-2 was found to be closer to a 233 cluster of rodent murine coronaviruses at the first two factorial levels (Figure 3A and Figure 234 4A). 235 Further analysis on spike gene, however, suggested that the codon usage of SARS-CoV-2 and 236 rodent murine coronaviruses were distinct at the third factorial level (Supplementary Figure 237 S8A). The results show that although RaTG13 was not the point most adjacent to SARS-238 CoV-2 at the first and second dimension, it surpassed murine coronaviruses at the third 239 dimension. Our results suggest a complex genomic background in the spike gene of SARS-240 CoV-2, which made its synonymous codon usage harder to differentiate from other genomic 241 sequences in our WCA analysis. Despite the proximity between RaTG13 and SARS-CoV-2 242 at three-dimensional level, they were still formed into two separated clusters (Supplementary 243 Figure S8A). It is evident that the synonymous codon usage pattern of SARS-CoV-2 is 244 distinct from other bat origin coronaviruses. The difference in synonymous codon usage is 245 largely explained by the first factor (more than 50%), and our analysis on codon usages 246 suggest that the first factor maybe highly related to the preferential usage of codons ending

248

249

250

251

252

253

254

255

256

257

258

259260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

278

with cytosine (Supplementary Figure S9). We also had similar observation for the membrane gene. Our three-dimensional analysis revealed that the synonymous codon usage of SARS-CoV-2 in membrane was most similar to P1E and CoVZXC21 (Supplementary Figure S8B). It is worth noting that comparing to RaTG13, P1E and CoVZXC21 had lower synonymous codon usage similarity to SARS-CoV-2 in the other three genes. Overall, our WCA results support a more complex synonymous codon usage background on spike and membrane genes, though we identified unique codon usage patterns of SARS-CoV-2 on these two genes. Discussion Codon usage can be affected by many sequence features, including nucleotide composition, dinucleotide composition, amino acid preference, host adaption, etc^{8,19,20}. The codon usages of viral sequences can vary by genes and host origins^{21–23}. The bias in codon usage is a unique and distinctive characteristic that can reflect the "signature" of a genomic sequence. Codon usage analyses are often complementary to ordinary sequence alignment-based analyses which focus on the genetic distance at nucleotide level, whereas codon usage analyses enable capturing signals at different sequence parameters. Therefore, codon usage bias can be another good proxy for identifying unique traits (e.g. virus origin, host origin, or some functions of proteins) of a genome. The goal of this study was to investigate the codon usage bias of betacoronaviruses. By studying the codon usags of these viruses in a systematic manner, we identified viral sequences carrying traits similar to those of SARS-CoV-2, which provided useful information for studying the host origin and evolutionary history of SARS-CoV-2. The codon usage of different genes in betacoronaviruses are very different. The G+C content, especially the GC3 content is known to be influential to the codon usage of some bacteria and viruses ^{7,24,25}. The GC3 content has pronounced effects on our WCA analysis of the orf1ab and spike genes. The GC3 content was found correlated with high WCA values on the first factor of orf1ab (Supplementary Figure S9). By contrast, codons ending with cytosine had lower factorial values in the spike gene analysis (Supplementary Figure S9). The G + C contents in membrane and nucleocapsid genes were less suppressed (Supplementary Figure S2). This can be partly explained by the fact that membrane and nucleocapsid are two genes with shorter lengths which may limit the flexibilities for mutation or codon usage adaptation.

280

281

282

283

284

285

286

287288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303304

305

306

307

308

309

310

In addition to global CA analysis, the application of WCA and BCA can eliminate the effects caused by amino acid compositions and synonymous codon usage, respectively. These alternative analytical tools were important to our study. It is because the amino acid sequences are expected to be more conserved such that they can preserve biological functions of the translated genes. By contrast, mutations at synonymous level tend to be more frequent, as most of these codon alternatives do not affect the biological function of a protein. Of all the existing genomes in the dataset, RaTG13 best matched the overall codon usage pattern of the SARS-CoV-2. Although the SARS-CoV-2 had amino acid usage similar to bat and human SARSr-CoVs, the synonymous codon usages between them were relatively different, which indicates similar protein characteristics but maybe different evolutionary histories. The codon usage of bat coronaviruses are more scattered than coronaviruses of other hosts. This result agrees with the fact that bat is a major host reservoir of coronavirus²⁶, thus it harbours coronaviruses with more complex genomic backgrounds. SARS-CoV-2 was first identified in human, but its codon usage pattern is very different from those of other human betacoroanviruses (Supplementary Figure S5A). In fact, the codon usage at both the amino acid level and synonymous level denote that the orflab gene in SARS-CoV-2 had closest relationship to SARSr-CoV, especially RaTG13. The CoVZX45 and CoVZXC21 had similar amino acid usage but relatively different synonymous codon usage to SARS-CoV-2 (Figure 3). Besides bat-origin SARSr-CoV, the pangolin P1E also had similar codon usage to SARS-CoV-2 both at amino acid and synonymous codon levels. The result in orflab is in accordance with the full-genome phylogenetic analysis (Supplementary Figure S7), showing a close relationship between SARS-CoV-2 and RaTG13 by the overall backbone of the genome. The S protein is responsible for receptor binding which is important for viral entry. The genetic variability is extreme in spike gene²⁷, and this highly mutable gene may possess valuable information about recent evolution history. In our results, the synonymous codon usage of SARS-CoV-2 in spike gene was distinct from those of RaTG13 and other phylogenetic relatives (Figure 3A), which was not observed in orflab or nucleocapsid gene. Although the codon usage in spike of SARS-CoV-2, RaTG13 and P1E were similar at amino acid level, the difference at synonymous codon usage level indicates that they are unlikely to share a very recent common ancestor. It is more likely that SARS-CoV-2, RaTG13 and P1E might have undergone different evolution pathways for a certain period of time. The amino

311 acid usage of SARS-CoV-2 in membrane was clustered with bat SARSr-CoV, however the 312 synonymous codon usage of SARS-CoV-2 was still distinct to these bat coronaviruses. 313 Notably, in membrane gene, pangolin P1E had a more similar synonymous codon usage to 314 SARS-CoV-2 than RaTG13. These findings suggest that there may be different selection 315 forces between genes. Our result supports different evolutionary background or currently 316 unknown host adaption history in SARS-CoV-2. The codon usage of SARS-CoV-2 in 317 nucleocapsid gene was similar to bat SARSr-CoV both at amino acid level and synonymous 318 level, suggesting that no highly significant mutation happened in this gene. 319 Codon usage can be shaped by many different selection forces, including the influence from 320 host factors. Some researchers have hypothesised that the codon usage in SARS-CoV-2 maybe directly correlated to the codon usage of its host²⁸. However our recent study on 321 322 influenza A viruses implied that these may not be the most influential factors shaping the 323 codon usage of a viral genome¹⁹. Our analysis took advantage of the existing genomes of 324 Betacoronavirus to study the complex host effect on codon usage, which warrants more 325 accurate but relatively conserved estimation. 326 Acknowledgements 327 We thank researchers for making viral sequences available for public access. We gratefully 328 acknowledge the Originating and Submitting Laboratories for sharing genetic sequences and 329 other associated data through the GISAID Initiative, on which this research is based. A list of 330 the authors can be found in Supplementary Table S2. 331 **Funding** 332 This work was supported by Health and Medical Research Fund (Hong Kong) and National 333 Institutes of Allergy and Infectious Diseases, National Institutes of Health (USA) (contract 334 HHSN272201400006C). 335 References 336 Wang, C., Horby, P. W., Hayden, F. G. & Gao, G. F. A novel coronavirus outbreak of global 337 health concern. Lancet (2020). doi:10.1016/S0140-6736(20)30185-9 338 2. Zhu, N. et al. A Novel Coronavirus from Patients with Pneumonia in China, 2019. N. Engl. J. 339 Med. NEJMoa2001017 (2020). doi:10.1056/NEJMoa2001017 340 3. WHO. Novel coronavirus – Republic of Korea (ex-China). Geneva: World Health

341

Organization. (2020).

- 4. Percudani, R. & Ottonello, S. Selection at the wobble position of codons read by the same
- tRNA in Saccharomyces cerevisiae. Mol. Biol. Evol. (1999).
- doi:10.1093/oxfordjournals.molbev.a026087
- 345 5. Pepin, K. M., Domsic, J. & McKenna, R. Genomic evolution in a virus under specific
- 346 selection for host recognition. *Infect. Genet. Evol.* (2008). doi:10.1016/j.meegid.2008.08.008
- 347 6. Akashi, H. & Eyre-Walker, A. Translational selection and molecular evolution. Curr. Opin.
- 348 Genet. Dev. (1998). doi:10.1016/S0959-437X(98)80038-5
- 7. Perriere, G. Use and misuse of correspondence analysis in codon usage studies. *Nucleic Acids*
- 350 *Res.* **30**, 4548–4555 (2002).
- 351 8. Suzuki, H., Brown, C. J., Forney, L. J. & Top, E. M. Comparison of correspondence analysis
- methods for synonymous codon usage in bacteria. DNA Res. 15, 357–365 (2008).
- 353 9. Lu, R. et al. Genomic characterisation and epidemiology of 2019 novel coronavirus:
- implications for virus origins and receptor binding. *Lancet (London, England)* (2020).
- 355 doi:10.1016/S0140-6736(20)30251-8
- 356 10. Zhou, P. et al. Discovery of a novel coronavirus associated with the recent pneumonia
- outbreak in humans and its potential bat origin. bioRxiv 2020.01.22.914952 (2020).
- 358 doi:10.1101/2020.01.22.914952
- 359 11. Lam, T. T.-Y. et al. Identification of 2019-nCoV related coronaviruses in Malayan pangolins
- 360 in southern China. *bioRxiv* 2020.02.13.945485 (2020). doi:10.1101/2020.02.13.945485
- 361 12. Gu, H. & Poon, L. L. Bioconductor SynMut. (2019). Available at:
- 362 https://doi.org/doi:10.18129/B9.bioc.SynMut. (Accessed: 24th January 2020)
- 363 13. Dray, S. & Dufour, A. B. The ade4 package: Implementing the duality diagram for ecologists.
- 364 *J. Stat. Softw.* **22**, 1–20 (2007).
- 365 14. Benzécri, J. P. Analyse de l'inertie intraclasse par l'analyse d'un tableau de correspondance.
- 366 *Cah. l'Analyse des données* **8**, 351–358 (1983).
- 367 15. Kyte, J. & Doolittle, R. F. A simple method for displaying the hydropathic character of a
- 368 protein. J. Mol. Biol. (1982). doi:10.1016/0022-2836(82)90515-0
- 369 16. Charif, D. & Lobry, J. R. SeqinR 1.0-2: A Contributed Package to the R Project for Statistical
- 370 Computing Devoted to Biological Sequences Retrieval and Analysis. in *Structural approaches*
- 371 to sequence evolution: Molecules, networks, populations (eds. Bastolla, U., Porto, M., Roman,
- 372 H. E. & Vendruscolo, M.) 207–232 (Springer Verlag, 2007). doi:10.1007/978-3-540-35306-
- 373 5_10

- 374 17. Lobry, J. R. Multivariate Analyses of Codon Usage Biases. Multivariate Analyses of Codon
- 375 *Usage Biases* (2018). doi:10.1016/c2018-0-02165-9
- 376 18. Zhou, T., Gu, W., Ma, J., Sun, X. & Lu, Z. Analysis of synonymous codon usage in H5N1
- virus and other influenza A viruses. *BioSystems* **81**, 77–86 (2005).
- 378 19. Gu, H., Fan, R. L. Y., Wang, D. & Poon, L. L. M. Dinucleotide evolutionary dynamics in
- influenza A virus. Virus Evol. 5, (2019).
- 380 20. Hershberg, R. & Petrov, D. A. Selection on Codon Bias. Annu. Rev. Genet. (2008).
- 381 doi:10.1146/annurev.genet.42.110807.091442
- 382 21. Wong, E. H., Smith, D. K., Rabadan, R., Peiris, M. & Poon, L. L. Codon usage bias and the
- 383 evolution of influenza A viruses. Codon Usage Biases of Influenza Virus. BMC Evol. Biol. 10,
- 384 253 (2010).
- 22. Cristina, J., Moreno, P., Moratorio, G. & Musto, H. Genome-wide analysis of codon usage
- 386 bias in Ebolavirus. *Virus Res.* (2015). doi:10.1016/j.virusres.2014.11.005
- 387 23. Jenkins, G. M. & Holmes, E. C. The extent of codon usage bias in human RNA viruses and its
- 388 evolutionary origin. Virus Res. (2003). doi:10.1016/S0168-1702(02)00309-X
- 389 24. Gu, W., Zhou, T., Ma, J., Sun, X. & Lu, Z. Analysis of synonymous codon usage in SARS
- 390 Coronavirus and other viruses in the Nidovirales. *Virus Res.* (2004).
- 391 doi:10.1016/j.virusres.2004.01.006
- 392 25. Woo, P. C. Y., Huang, Y., Lau, S. K. P. & Yuen, K. Y. Coronavirus genomics and
- 393 bioinformatics analysis. Viruses (2010). doi:10.3390/v2081803
- 394 26. Calisher, C. H., Childs, J. E., Field, H. E., Holmes, K. V. & Schountz, T. Bats: Important
- reservoir hosts of emerging viruses. *Clinical Microbiology Reviews* (2006).
- 396 doi:10.1128/CMR.00017-06

- 397 27. Gallagher, T. M. & Buchmeier, M. J. Coronavirus spike proteins in viral entry and
- 398 pathogenesis. *Virology* (2001). doi:10.1006/viro.2000.0757
- 399 28. Ji, W., Wang, W., Zhao, X., Zai, J. & Li, X. Homologous recombination within the spike
- 400 glycoprotein of the newly identified coronavirus may boost cross ☐ species transmission from
- 401 snake to human. *J. Med. Virol.* jmv.25682 (2020). doi:10.1002/jmv.25682

Table 1. Variability explained by the synonymous codon usage level and the amino acid level.

404

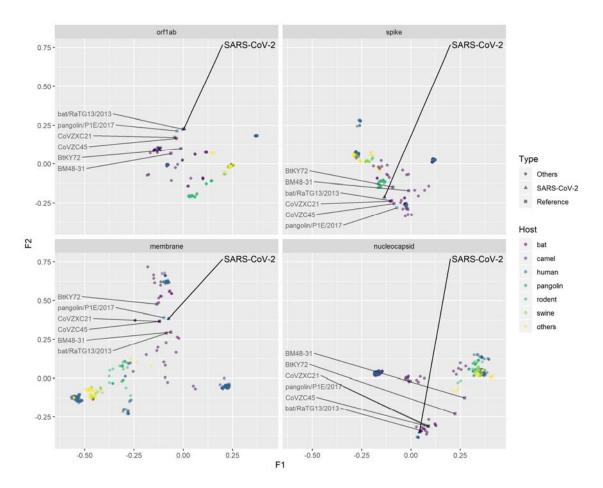
405

	Orf1ab	Spike	Membrane	Nucleocapsid
WCA (synonymous codon level)	90.36%	85.29%	83.71%	84.07%
BCA (amino acid level)	9.64%	14.71%	16.29%	15.93%

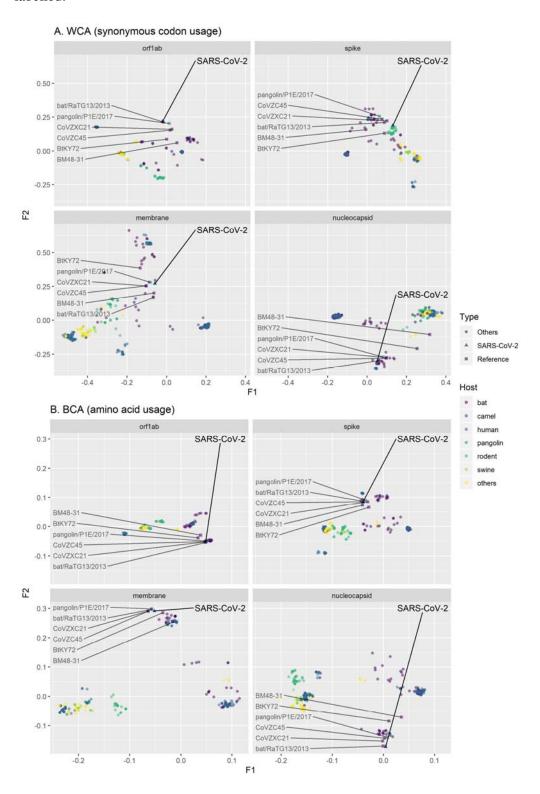
Figure 1. Codon usage in *Betacoronavirus* (Cleveland's dot plot). Points in green showed the count of codons in a sample SARS-CoV-2 genome (MN908947).



- 410 Figure 2. Factorial map of the first and second factors for global CA by different genes,
- 411 coloured by different viral host. The SARS-CoV-2 and related reference data points were
- 412 labelled.



- Figure 3. Factorial map of the first and second factors for WCA and BCA by different genes,
- 416 coloured by different viral host. The SARS-CoV-2 and related reference data points were
- 417 labelled.



- Figure 4. Factorial map of the first and second factors for WCA and BCA by different genes,
 - coloured by different viral species. The SARS-CoV-2 and related reference data points were
- 421 labelled.

