bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

Pannexin 1 regulates spiny protrusion dynamics in cortical neurons

Juan C. Sanchez-Arias[1], Rebecca C. Candlish[1], and Leigh Anne Swayne[1]*

[1]Division of Medical Sciences, University of Victoria, Victoria, British Columbia V8P 5C2, Canada.

Author Contributions

J.C.S.A. and L.A.S. designed the research. J.C.S.A. and R.C.C. performed the research. J.C.S.A. and L.A.S. analyzed the research. J.C.S.A. and L.A.S. wrote the manuscript.

Correspondance

*Lead contact. Correspondence: lswayne@uvic.ca; twitter: @dr_swayne

Number of figures: 5

Number of tables: 2

Number of words for abstract: 199

Number of words for significance statement: 90

Number of words for introduction: 442

Number of words for discussion: 545

Acknowledgements

We are thankful for technical assistance from Sarah N. Ebert who was supported by a University of Victoria Summer Undergraduate Student Award.

Funding sources

This work was supported by operating grants from the Canadian Institutes of Health Research (CIHR Grant MOP142215), from the Natural Sciences and Engineering Research Council (NSERC) [RGPIN-2017-03889], The Scottish Rite Charitable Foundation of Canada (15118) and the University of Victoria-Division of Medical Sciences to L.A.S. L.A.S. was also supported by a Michael Smith Foundation for Health Research and British Columbia Schizophrenia Society Foundation Scholar Award (5900). J.C.S.A. was supported by a University of Victoria Fellowship Graduate Award. L.A.S. is also grateful for infrastructure support from the Canada Foundation for Innovation (29462) and the BC Knowledge Development Fund (804754) for the Leica SP8 confocal microscope system.

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

Abstract

The integration of neurons into networks relies on the formation of dendritic spines. These specialized structures arise from dynamic filopodia-like spiny protrusions. Recently, it was discovered that cortical neurons lacking the channel protein Pannexin 1 (Panx1) exhibited larger and more complicated neuronal networks, as well as, higher dendritic spine densities. Here, we expanded on those findings to investigate whether the increase in dendritic spine density associated with lack of Panx1 was due to differences in the rates of spine dynamics. Using a fluorescent membrane tag (mCherryCD9-10) to visualize spiny protrusions in developing neurons (at 10 days-*in-vitro*, DIV10) we confirmed that lack of Panx1 leads to higher spiny protrusion density while transient transfection of Panx1 leads to decreased spiny protrusion density. To quantify the impact of Panx1 expression on spiny protrusion formation, elimination, and motility, we used live cell imaging in DIV10 neurons (1 frame every 5 seconds for 10 minutes). We discovered, that at DIV10, lack of Panx1 KO stabilized spiny protrusions. Notably, re-expression of Panx1 in Panx1 knockout neurons resulted in a significant increase in spiny protrusion motility and turnover. In summary, these new data revealed that Panx1 regulates the development of dendritic spines by controlling protrusion dynamics.

Significance statement

Cells in the brain form intricate and specialized networks - *neuronal networks* - in charge of processing sensations, executing movement commands, and storing memories. To do this, brain cells extend microscopic protrusions - *spiny protrusions* - which are highly dynamic and survey the local environment to contact other cells. Those contact sites are known as synapses and undergo further stabilization and maturation establishing the function and efficiency of neuronal networks. Our work shows that removal of Panx1 increases the stability and decreases the turnover of spiny protrusion on young neurons.

Introduction

Pannexin 1 (Panx1) is a four transmembrane domain protein that forms channels permeable to ion and metabolites with various activation mechanisms and diverse (patho)physiological implications (for review Boyce et al., 2018; Chiu et al., 2018). Panx1 is broadly and highly expressed in the brain during postnatal early development (Ray et al., 2005; Vogt et al., 2005) and localized and enriched in synaptic compartments (Sanchez-Arias et al., 2019; Zoidl et al., 2007).

Recent reports have implicated Panx1 in neurite outgrowth, hippocampal synaptic plasticity, and the development of neuronal networks and dendritic spines in cortical neurons (Ardiles et al., 2014; Prochnow et al., 2012; Sanchez-Arias et al., 2019; Wicki-Stordeur & Swayne, 2013). While the behavioural features resulting from a loss of Panx1 have not been thoroughly characterized, a handful of studies have detected important phenotypes like anxiety, increased wakefulness, and spatial learning deficits (Ardiles et al., 2014; Gajardo et al., 2018; Kovalzon et al., 2017; Prochnow et al., 2012). Notably, dendritic spine development has been linked to each

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

of these behaviours. For example, dendritic spine density is increased in various neurodevelopmental disorders in which clinical manifestations include anxiety, intellectual disability, and stereotypical movements (Phillips & Pozzo-Miller, 2015). Moreover, sleep promotes dendritic spine and spiny protrusion turnover in the cortex and hippocampus (Spano et al., 2019; G. Yang & Gan, 2012), which facilitates network sparsity and memory consolidation (Frank et al., 2018; Li et al., 2017). Dendritic spine-based synapses result from spiny protrusions (including dendritic filopodia) actively extending to contact presynaptic boutons during developmental excitatory synaptogenesis; upon contact, spiny protrusions stabilize and evolve into mature dendritic spines along active presynaptic boutons (Fiala et al., 1998; Ziv & Smith, 1996). These steps are critical in establishing network ensembles and Hebbian plasticity (Hoshiba et al., 2017).

In light of this evidence, we investigated the role of Panx1 in spiny protrusion dynamics in cultured primary cortical neurons at 10 days-*in-vitro* (DIV10). We first established an approach to study spiny protrusions using a fluorescent membrane tag (mCherryCD9-10), allowing us to visualize these characteristically long and thin structures. Then, we transiently transfected wildtype (WT) and Panx1 knock-out (KO) neuronal cultures with EGFP or Panx1EGFP (as well as mCherry-CD9-10) and analyzed spiny protrusions in fixed and living neurons at DIV10. We confirmed that lack of Panx1 leads to higher spiny protrusion density while over-expression and rescue of Panx1 leads to decreased density. Using live cell imaging we observed increased stability and decreased turnover of spiny protrusions in Panx1 KO neurons, while re-expression of Panx1 resulted in a significant increase in spiny protrusion motility and turnover. In summary, these new data reveal an inverse relationship between Panx1 expression and dendritic spine stability.

Materials and Methods

Table 1. Key Resources Table

Reagent or Resource	Source	Identifier	RRID
Experimental Models: Organisms/Strains			
Cortical neuron cultures from P0 C57BL/6J	The Jackson Laboratory	Cat# JAX:000664	RRID:IMSR_JAX:000664
Cortical neuron cultures from P0 Panx1 KO on a C57BL/6J background	(Dvoriantchikova et al., 2012; Sanchez-Arias et al., 2019)	NA	NA
Recombinant DNA			
mCherry-CD9-10	Addgene	Plasmid #55013	RRID:Addgene_55013
pEGFP-N1	Clontech (Takara Bio) - discontinued	Cat# 6085-1	NA
Panx1EGFP	(Penuela et al., 2007)	NA	NA
Chemicals, Recombinant Proteins			
DMEM/F12	Thermo Fisher Scientific		NA
NeuroCult™	STEMCELL Tech.	Cat# 05713	NA
BrainPhys™	STEMCELL Tech.	Cat# 05790	NA

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

Neurocult™ SM1	STEMCELL Tech.	Cat# 05711	NA
GlutaMAX	Thermo Fisher Scientific	Cat# 35050061	NA
Penicillin/Streptomycin	Thermo Fisher Scientific	Cat# 15140122	NA
Gentamicin	MilliporeSigma	G1397	NA
Poly-D-lysine hydrobromide (PDL)	MilliporeSigma	P6407	NA
Dispase-1	MilliporeSigma	D4818-2MG	NA
Papain	MilliporeSigma	P4762-25MG	NA
DNAse-1	MilliporeSigma	11284932001	NA
Cytosine β-D-arabinofuranoside (ara-C)	MilliporeSigma	C1768	NA
Lipofectamine2000	Thermo Fisher Scientific	Cat# 11668027	NA
OptiMEM™ I	Thermo Fisher Scientific	Cat# 31985062	NA
Probenecid (water-soluble)	Thermo Fisher Scientific	Cat# P36400	NA
My-Taq Extract PCR Kit	Bioline	BIO-21126	NA
Vectashield	Vector Laboratories	H-1000	RRID:AB_2336789
Software and Algorithms			
FIJI (FIJI is just ImageJ)	NIH, (Schindelin et al., 2012)	NA	RRID:SCR_002285
MultiStackReg v1.45	Brad Busse (http://bradbusse.net/Multi-StackReg1.45_.jar)	NA	NA
R Project for Statistical Computing (version 3.6.2)	The R Foundation	NA	RRID:SCR_001905
Rstudio	Rstudio Inc.	NA	RRID:SCR_000432
tidyverse package for R	CRAN	NA	RRID:SCR_014601
DaBest package for R	CRAN, (Ho et al., 2019)	NA	NA
Adobe Photoshop CS6	Adobe Systems Inc.	NA	RRID:SCR_014199
Leica Application Suite Software version 3.1.3.16308	Leica Microsystems GmbH	NA	RRID:SCR_013673
Equipment			
Leica TCS SP8	Leica Microsystems GmbH	NA	NA
8-well Nunc™ Lab-Tek™ Chambered-coverglass	Thermo Fisher Scientific	155411PK	NA
PDL precoated coverslips	NeuVitro	GG-12-PDL	NA

Experimental animals

All animal procedures were approved by the University of Victoria Animal Care Committee and performed in accordance with the guidelines set by the Canadian Council on Animal Care. Male and female postnatal day (P)0-P1 were used in this study. C57BL/6J mice were obtained from The Jackson Laboratory. The global Panx1 KO strain was derived from a strain originally generated by Dr. Valery Shestopalov (Dvoriantchikova et al., 2012). These mice have been back-crossed in-house onto a C57BL/6J for at least 6 generations (Sanchez-Arias et al., 2019). Mice were housed under a 12 h light/dark cycle starting at 8:00 A.M., with food and water *ad libitum*; temperature was maintained between 20 and 25ºC and humidity at 40-65%.

Primary cortical neuron cultures and transfections

Primary cortical neuron cultures were prepared as previously described (Sanchez-Arias et al., 2019). Briefly, cortices from male and female P0 pups from timed-pregnant WT and Panx1 KO breeding pairs were microdissected and incubated with papain, dispase-1, and DNAse-1 for 40 minutes in HBSS followed by mechanical dissociation in DMEM/F12 medium supplemented with Neurocult™ SM1, GlutaMAX, and penicillin/streptomycin (P/S). Then, 125,000 cells were plated in Nun™ Lab-Tek™ 8-well chambered coverglasses coated with PDL. After 1-2

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

hours after plating, the medium was replaced with Neurocult™ supplemented with Neurocult™ SM1, GlutaMAX, P/S, and gentamicin. From 4 days-*in-vitro* (DIV) onwards, partial (half) medium changes were done with BrianPhys™ maturation medium (Bardy et al., 2015); to limit proliferation of glial cells, ara-C was added to the medium at DIV4. Transfections were performed at DIV6 using Lipofectamine©2000. DNA/lipid complexes were diluted in OptiMEM-I® at ratio of 2 µg DNA:1 µL lipofectamine ratio and incubated at room temperature for 30 minutes. Then, these DNA/lipid complexes were added to cells in BrainPhys™ medium without antibiotics and incubated for 1-1.5 hours. Neurons were transfected with either pEGFP-N1 (250 ng) or Panx1EGFP (250 ng, gift from Silvia Penuela and Dale Laird). All transfections contained mCherry-CD9-10 (250 ng, was a gift from Michael Davidson; Addgene plasmid #55013; http://n2t.net/addgene:55013; RRID:Addgene_55013) to visualize neurons and spiny protrusions (**Figure 1**). All neurons used for this study were used at DIV10. Neurons used for fixed quantifications were plated on PDL-coated coverslips

Genotyping

Primers for LoxTGF, LoxTGR, and Panx1 LoxR (CTTTGGCATTTTCCCAGTGT, CGCGGTTGTAGACTTTGTCA, and GTCCCTACAGGAGGCACTGA) were used to genotype mice as previously described (Sanchez-Arias et al., 2019). Genomic DNA was extracted from tail-clips using MyTaq™Extract PCR Kit. DNA from WT mice amplifies a single 585 bp band, whereas DNA from global Panx1 KO mice have a single 900 bp band.

Imaging and analysis of spiny protrusions in fixed cortical neurons

Spiny protrusions (including filopodia) were defined as any membranous protrusions between 0.4 µm and 10 µm. Neurons on coverslips were fixed with 4% PFA and 4% sucrose for 10 minutes and mounted on microscope slides with VectaShield antifade mounting medium. High resolution images (3320×3320, pixel size: 0.088 µm, z-step size: 0.4 µm) were acquired using a Leica TSC SP8 microscope using a 40× immersion oil objective (1.30 NA) and exported to FIJI for analysis (Schindelin et al., 2012). Individual spiny protrusions were traced along the longest neurite (primary neurite) and their density was calculated by dividing the total number of spiny protrusions by the segment length and multiplying by 10 (spiny protrusions per 10 µm). Representative images were processed uniformly with a Gaussian blur of 0.5 pixels, and uniform adjustments to levels and contrast were made using Photoshop CS6 Extended suite (Adobe Systems).

Imaging and analysis of spiny protrusions in live cortical neurons

Cortical neurons plated on chambered coverglasses in BrainPhys™ at 37°C and 5% CO_2 and primary and secondary dendrite segments of 67-76 µm were imaged (1024×256, pixel size: 0.06 µm) every 5 seconds for 10 minutes and 0.7 µm z-step using a Leica TSC SP8 microscope in resonant mode (8,000 Hz) with a 63× water immersion objective (1.20 NA).

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

Images were exported to FIJI for analysis. First, the four-dimensionality (x,y,z,t) was reduced by creating maximum z projections before additional image processing and x-y drift was corrected with MultiStackReg v1.45 (developed by Brad Busse http://bradbusse.net/MultiStackReg1.45_.jar) when required. Then, images were subjected to a low-pass filter using a Gaussian blur (kernel size 2) and thresholded using the triangle method (Zack et al., 1977). From these binary images, outlines for each time frame were created and temporal colour-coded (**Figure 3*A*,*B***). Spiny protrusions were manually counted, and four basic characteristics were recorded: formation, elimination, lability, and motility. We defined formation as any *de novo* appearance of a spiny protrusion within the time-lapse recording; elimination was defined as the complete disappearance of a spiny protrusion. Lability was defined as spiny protrusions that were formed and eliminated within the duration of the time-lapse, typically short-lived and lasting 1-3 minutes (**Figure 3*C***). To assess spiny protrusion motility, we annotated partial extensions and partial retractions of individual spiny protrusions (**Figure 3*C***). The survival fraction of spiny protrusions was calculated by dividing the number of spiny protrusions at the end of each time-lapse (10-minute mark) by the number of spiny protrusions at the start (0-minute mark). The overall turnover rate was calculated as the net per cent gain and loss (sum of formation, elimination, and lability) of spiny protrusions divided by the number of spiny protrusions at the start of the time-lapse. Lastly, the overall movement change of spiny protrusions (Δ movement) was calculated by adding the basic dynamic characteristics (formation, elimination, lability, and motility) divided by the number of spiny protrusion at 0 min. Representative images were processed uniformly with a Gaussian blur of 0.5 pixels, and uniform adjustments to levels and contrast were made using Photoshop CS6 Extended suite (Adobe Systems Inc.).

Experimental design and statistical analysis

For all experiments 3 independent cultures were used. All images were blindly acquired and analyzed. Relevant details are described in *Results*, figure legends, and where appropriate, illustrated on the figures themselves. Data are presented as mean ± standard deviation. Data analysis using bootstrap estimation (5000 bootstrap resamples), determination of effect size, bias-corrected confidence intervals, and Cumming estimation plots were generated using the dabestR package for R (Bernard, 2019; Calin-Jageman & Cumming, 2019; Ho et al., 2019). Null-hypothesis significance testing was performed using R (version 3.6.2) and a p value < 0.05 was used as the significant threshold for these tests. Normality was tested using the Shapiro-Wilk test (McDonald, 2014). Group analyses for normally distributed data were performed with a two-way ANOVA coupled to multiple comparisons with Bonferroni's correction. For non-normally distributed data Kruskal-Wallis pairwise comparisons with Bonferroni's correction were used.

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

Results

Table 2. Statistical Table

	Fig.	*Comparison*	*Type of test*					*Pair.Comp.*	*Effect size*
a1	2Bi	Spiny protrusion density (# / 10 µm)	Two-way ANOVA with Bonferroni's correction						
				Df	Mean^2	F	Pr(>F)		
		WT-EGFP vs Panx1 KO-EGFP	genotype	1	9.61	2.679	0.1095	0.03517 *	2.36 [95CI 1.28; 3.54]
		WT-EGFP vs WT-Panx1EGFP	plasmid	1	252.48	70.432	2.34e^{-10}	0.00268 **	-3.24 [95CI -4.54; -2.21]
		WT-EGFP vs Panx1 KO-Panx1EGFP	interaction	1	22.05	6.151	0.0174	0.00026 ***	-3.72 [95CI -5.17; -2.15]
		Panx1 KO-EGFP vs Panx1 KO-Panx1EGFP						7.10e^{-09} ****	-6.08 [95CI -7.84; -4.51]
a2	2Bii	Spiny protrusion length (µm)	Two-way ANOVA with Bonferroni's correction						
				Df	Mean^2	F	Pr(>F)		
		WT-EGFP vs Panx1 KO-EGFP	genotype	1	0.1133	0.618	0.436	>0.9999	-0.13 [95CI -0.397; 0.131]
		WT-EGFP vs WT-Panx1EGFP	plasmid	1	0.2018	1.101	0.300	>0.9999	0.105 [95CI -0.21; 0.417]
		WT-EGFP vs Panx1 KO-Panx1EGFP	interaction	1	0.0085	0.046	0.831	>0.9999	0.031 [95CI -0.31; 0.36]
		Panx1 KO-EGFP vs Panx1 KO-Panx1EGFP						>0.9999	0.161 [95CI -0.178; 0.53]
b1	4Bi	Spiny protrusion formation (%)	Kruskal-Wallis chi-squared = 14.593, df = 3, p-value = 0.0022						
		WT-EGFP vs Panx1 KO-EGFP						0.2267	-3.02 [95CI -5.39; -0.803]
		WT-EGFP vs WT-Panx1EGFP						>0.9999	3.75 [95CI -0.755; 10.3]
		WT-EGFP vs Panx1 KO-Panx1EGFP						0.2111	5.21 [95CI 1.21; 9.14]
		Panx1 KO-EGFP vs Panx1 KO-Panx1EGFP						0.0028 **	8.23 [95CI 4.54; 11.8]
b2	4Bii	Spiny protrusion elimination (%)	Kruskal-Wallis chi-squared = 25.245, df = 3, p-value = 1.372e-05						
		WT-EGFP vs Panx1 KO-EGFP						0.62307	-2.57 [95CI -5.85; -0.209]
		WT-EGFP vs WT-Panx1EGFP						0.15616	5.27 [95CI 0.455; 11.5]
		WT-EGFP vs Panx1 KO-Panx1EGFP						0.00959	9.06 [95CI 4.02; 13.8]
		Panx1 KO-EGFP vs Panx1 KO-Panx1EGFP						0.00024 ***	11.6 [95CI 7.5; 15.8]
b3	4Biii	Spiny protrusion lability (%)	Kruskal-Wallis chi-squared = 13.421, df = 3, p-value = 0.00381						
		WT-EGFP vs Panx1 KO-EGFP						>0.9999	-1.11 [95CI -3.17; 0.786]
		WT-EGFP vs WT-Panx1EGFP						>0.9999	2.34 [95CI -0.966; 6.76]
		WT-EGFP vs Panx1 KO-Panx1EGFP						0.0291 **	6.2 [95CI 2.5; 9.81]
		Panx1 KO-EGFP vs Panx1 KO-Panx1EGFP						0.0034 **	7.31 [95CI 3.92; 10.6]
b4	4Biv	Spiny protrusion motility (%)	Kruskal-Wallis chi-squared = 20.442, df = 3, p-value = 0.0001374						
		WT-EGFP vs Panx1 KO-EGFP						0.00016 ***	-13.3 [95CI -18.5; -8.42]
		WT-EGFP vs WT-Panx1EGFP						>0.9999	0.816 [95CI -7.12; 9.33]
		WT-EGFP vs Panx1 KO-Panx1EGFP						>0.9999	-3.1 [95CI -9.89; 3.97]
		Panx1 KO-EGFP vs Panx1 KO-Panx1EGFP						0.03582 *	10.2 [95CI 4.53; 16]
c1	4Bvi	Spiny protrusion survival fraction (%)	Kruskal-Wallis chi-squared = 24.351, df = 3, p-value = 2.11e^{-05}						
		WT-EGFP vs Panx1 KO-EGFP						0.81748	2.21 [95CI 0.0663; 5.03]
		WT-EGFP vs WT-Panx1EGFP						0.2034	-4.4 [95CI -9.04; -0.308]
		WT-EGFP vs Panx1 KO-Panx1EGFP						0.00909 **	-8.19 [95CI -12.3; -3.65]
		Panx1 KO-EGFP vs Panx1 KO-Panx1EGFP						0.00028 ***	-10.4 [95CI -14; -6.58]
c2	4Biv	Spiny protrusion turnover (%)	Kruskal-Wallis chi-squared = 19.895, df = 3, p-value = 0.0001784						
		WT-EGFP vs Panx1 KO-EGFP						0.0092 **	-4.48 [95CI -7.73; -2.01]
		WT-EGFP vs WT-Panx1EGFP						0.5205	6.68 [95CI 0.967; 14.7]
		WT-EGFP vs Panx1 KO-Panx1EGFP						>0.9999	8.07 [95CI 2.84; 13]
		Panx1 KO-EGFP vs Panx1 KO-Panx1EGFP						0.0027 **	12.5 [95CI 8.06; 17]
c3	5Biii	Spiny protrusion Δ movement (%)	Kruskal-Wallis chi-squared = 28.526, df = 3, p-value = 2.816e-06						
		WT-EGFP vs Panx1 KO-EGFP						6.2e-^{05} ****	-17.8 [95CI -23.6; -11.9]
		WT-EGFP vs WT-Panx1EGFP						>0.9999	7.49 [95CI -3.45; 20.1]
		WT-EGFP vs Panx1 KO-Panx1EGFP						>0.9999	4.98 [95CI -4.1; 13.8]
		Panx1 KO-EGFP vs Panx1 KO-Panx1EGFP						0.00033 ***	22.8 [95CI 15; 30.4]

Pair.Comp, Pairwise comparison. Significance. codes: <0.0001 '****', <0.001, '***', <0.01 '**', <0.05 '*'

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

A novel approach to visualize and quantify spiny protrusions in cortical neurons

Spiny protrusions (including filopodia) are characteristically highly dynamic, thin, and long. As dendritic arbors mature, these transient structures stabilize into mature dendritic spines. Most methods used to detect these structures rely on cytoplasmic volume markers such as GFP (and its variants) or membrane-bound lipophilic dyes (DiI, DiO, etc). The former approach allows for sparse labelling but fails to fully label thin processes such a spiny protrusions (**Figure 1**), while the latter achieves clear visualization of these structures by labelling the membrane at the expense of widespread labelling (Mancuso et al., 2013). We co-transfected cortical neurons with the tetraspanin CD9-10 fused to a monomeric red fluorescent protein mCherry (mCherry-CD9-10 was a gift from Michael Davidson, Addgene plasmid #55013), mCherry-CD9-10 facilitated detailed resolution of spiny protrusions in sparsely transfected cells (**Figure 1**).

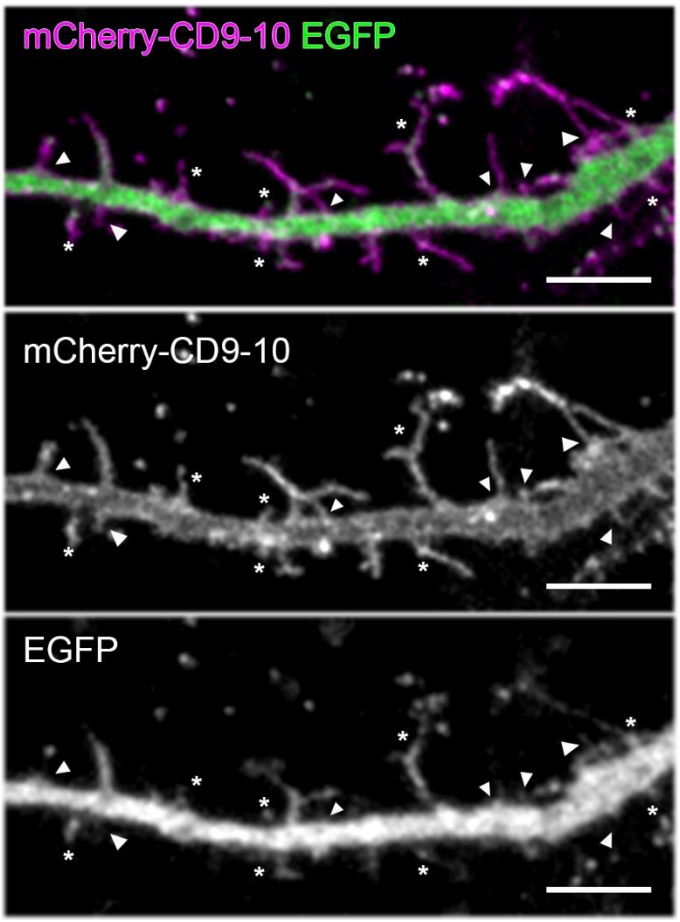


Figure 1. ***A novel approach to visualize and quantify spiny protrusions in cortical neurons***. Representative maximum projection of a dendritic segment from a neuron co-transfected with mCherryCD9-10 and EGFP at DIV6 and fixed at DIV10. Thin and long spiny protrusions are clearly visualized with mCherry-CD9-10 (mid), while the use of a volume marker such as EGFP (bottom) fails to accurately label these structures (*) or misses them entirely (arrowheads). Scale bar 5 µm.

Transfection of Panx1 decreases spiny protrusion density in WT and Panx1 KO DIV10 neurons

To investigate the impact of Panx1 expression, we co-transfected WT and Panx1 KO cortical neuronal cultures with mCherry-CD9-10 as well as EGFP (control) or Panx1EGFP (over-expression/rescue) at DIV6 and fixed the cells 4 days later at DIV10 (**Figure 2*A***). With EGFP control transfection we observed a 20% increase in spiny protrusion density in primary neurites of Panx1 KO neurons (effect size: 2.36 [95CI 1.28; 3.54], p = 0.03517, [a1]). In Panx1EGFP-expressing cultures we observed a 27% decrease in spiny protrusion density in WT neurons (**Figure 2*Bi***, effect size: -3.24 [95CI -4.54; -2.21], p = 0.00268, [a1]) and a 42.5% density reduction in Panx1 KO neurons (**Figure 2*Bi***, effect size: -6.08 [95CI -7.84; -4.51], p <0.0001, [a1]). Spiny protrusion length was not significantly different amongst the groups (**Figure 2B*ii***, [a2]). These results suggest spiny protrusion density is inversely proportional to Panx1 expression levels.

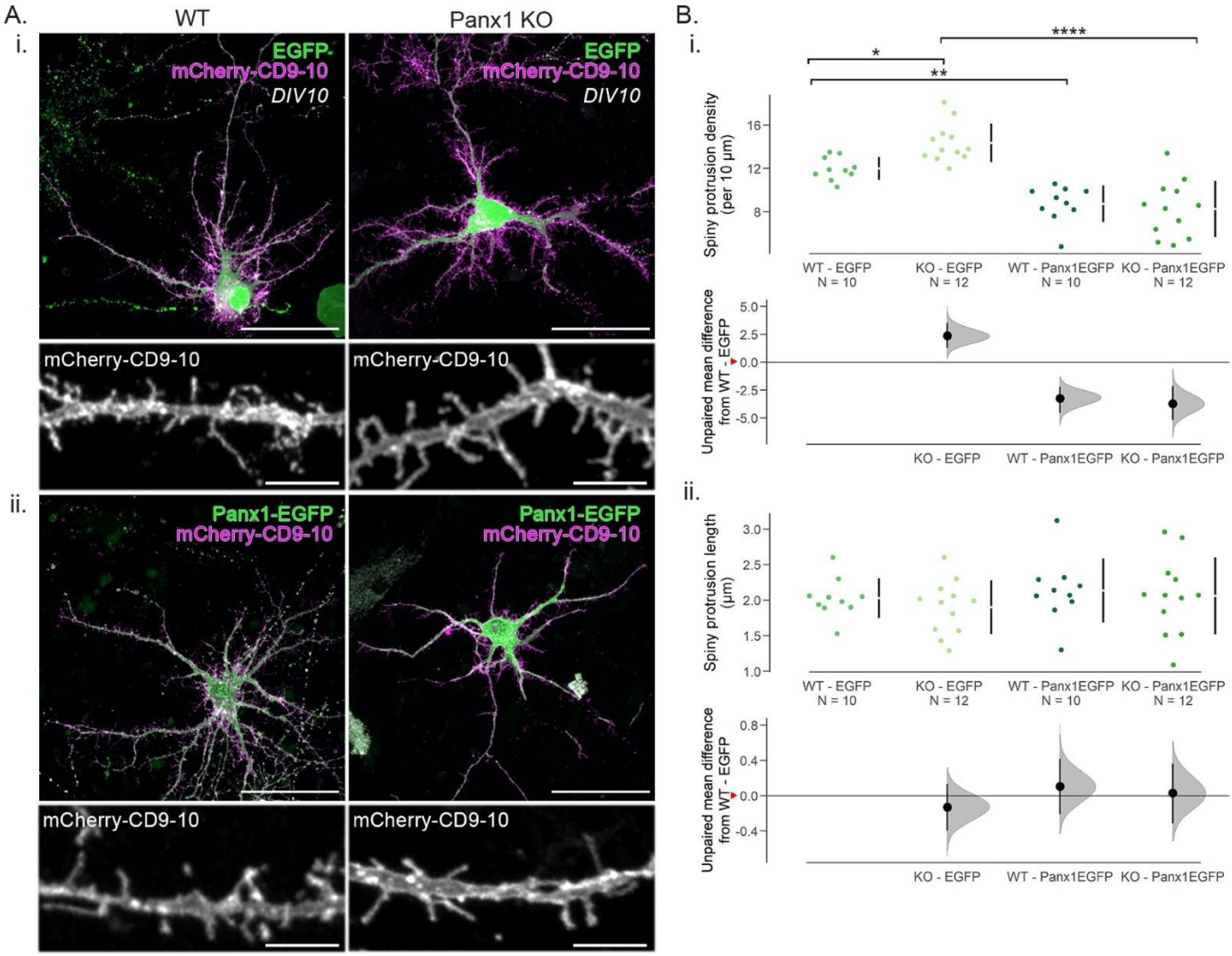
bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

Measuring spiny protrusion dynamics in living neurons using a membrane marker

To investigate the mechanisms contributing to differences in spiny protrusion densities between groups, we acquired 10-minute time-lapses (one frame every 5 seconds) of primary and secondary dendrites from cortical neurons at DIV10. These cultures were transfected with mCherry-CD9-10 and either EGFP or Panx1EGFP at DIV6. At DIV10, dendrites harbour highly dynamic, thin, and long spiny protrusion that are the precursors for dendritic spines (Fiala et al., 1998; Ziv & Smith, 1996). We reduced the dimensionality of the time-lapses by creating maximum z-projections, and then images were passed through a low-pass filter and thresholded to create outlines (**Figure 3*A***). The dendritic silhouettes (**Figure 3*B*)** were then temporally colour-coded to facilitate the detection of formation, elimination, liability, retraction, and growth of spiny protrusions (**Figure 3*C*)**.

Figure 2. ***Spiny protrusion density is inversely related to Panx1 expression levels in DIV10 neurons. A.*** Representative maximum projections of WT and Panx1 KO cultured cortical neurons co-transfected with mCherry-CD9-10 and either EGFP (***Ai***) or Panx1EGFP (***Aii***) and their respective dendritic segments from a primary neurite. Scale bar: 50, and 5 µm. ***B.*** Effect of Panx1 expression in spiny protrusion density and length in developing cortical neurons co-transfected with mCherry-CD-9-10 and either EGFP or Panx1EGFP using Cumming estimation plots. ***Bi.*** With EGFP expression, spiny protrusion density was higher with Panx1 KO neurons (WT-EGFP: 12.0 ± 0.3 spiny protrusions per 10 µm; Panx1 KO-EGFP: 14.4 ± 0.5 spiny protrusions per 10 µm, p = 0.03517, two-way ANOVA with Bonferroni's multiple-comparison test). With Panx1EGFP expression, spiny protrusion density was decreased in both WT and Panx1 KO neurons (WT-Panx1EGFP: 8.8 ± 0.5 spiny protrusions per 10 µm, p = 0.00268; Panx1 KO-Panx1EGFP: 8.3 ± 0.8 spiny protrusions per 10 µm, p < 0.0001, two-way ANOVA with Bonferroni's multiple-comparison test, [a1]). ***Bii.*** No significant differences in spiny protrusion length were found between groups (WT-EGFP: 2.0 ± 0.3 µm; Panx1 KO-EGFP: 1.9 ± 0.4 µm, p > 0. 9999, two-way ANOVA with Bonferroni's multiple-comparison test, [a2]; WT-Panx1EGFP: 2.1 ± 0.1 µm; Panx1 KO-Panx1EGFP: 2.1 ± 0.2 µm, p > 0. 9999, two-way ANOVA with Bonferroni's multiple-comparison test). Data are presented as mean ± standard deviation. Effect sizes are reported in the main text and Table 2. s.p., spiny protrusion. <0.0001, '****'; <0.001, '***'; <0.01 '**'; <0.05 '*'. *Figure on next page.*

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.



Basic characteristics of spiny protrusion dynamics in WT and Panx1 KO neurons at DIV10

Using the above approach, we observed that transfection of Panx1EGFP in Panx1 KO neurons significantly increased the percentage of formation and elimination of spiny protrusions compared to EGFP transfection of Panx1 KO neurons (**Figure 4*A* & 4*Bi-ii***, formation – effect size KO-EGFP *vs.* KO-Panx1EGFP: 8.23% [95CI 4.54%; 11.8%], $p = 0.0028$, [b1]; elimination - effect size KO-EGFP *vs.* KO-Panx1EGFP: 11.6% [95CI 7.5%; 15.8%], $p = 0.00024$ [b2]), while no significant differences were observed between Panx1EGFP and EGFP transfection in WT neurons ($p > 0.9999$, [b1]). Similarly, no significant differences were observed between genotypes with EGFP (control) transfection (**Figure 4*Bi-ii***, formation – effect size WT-EGFP *vs.* KO-EGFP: -3.02% [95CI -5.39%; -0.803%], $p = 0.2267$, [b1]; elimination – effect size WT-EGFP *vs.* KO-EGFP: -2.57% [95CI -5.85%; -0.209%], $p = 0.62307$, [b2]). We next quantified spiny protrusion lability within our experimental groups. Transient

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

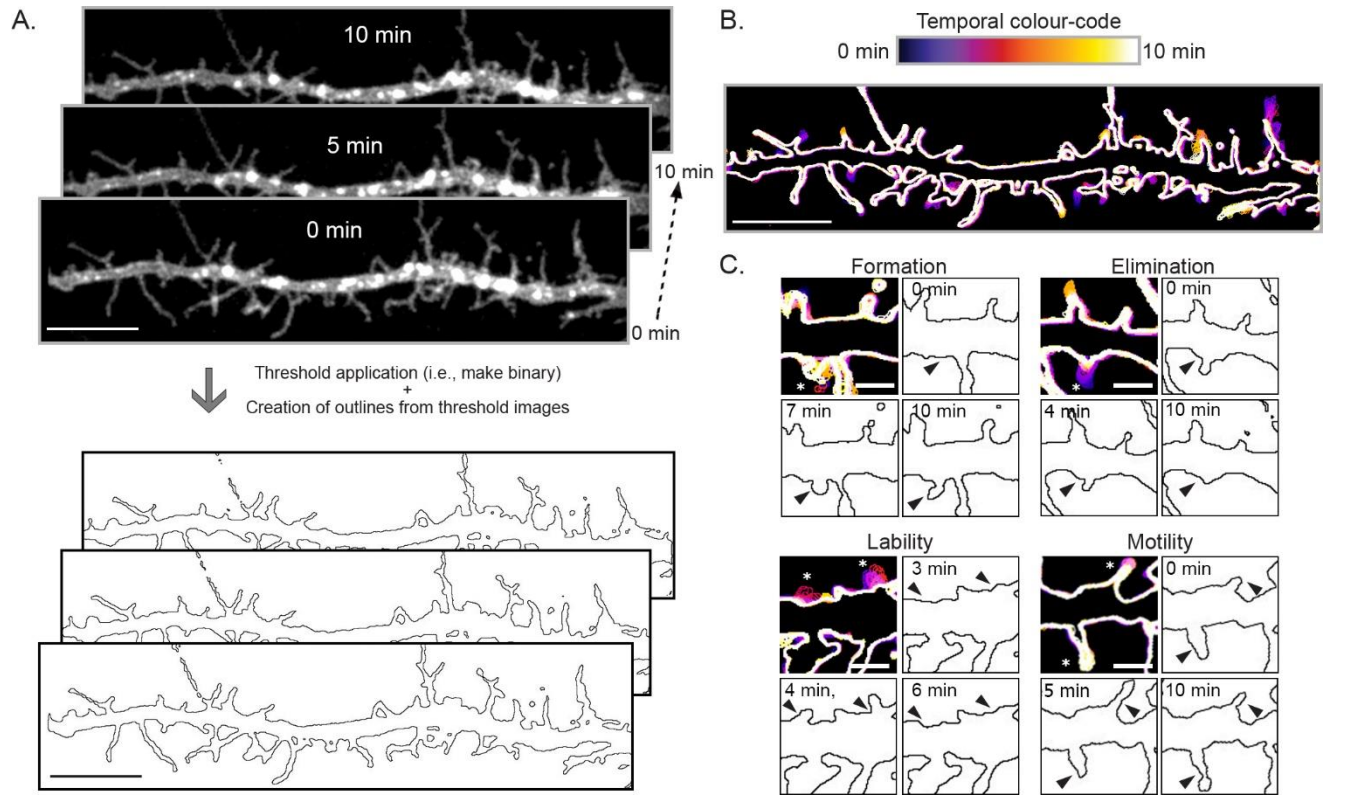


Figure 3. ***Image analysis strategy to quantify spiny protrusion dynamics in cortical cultures.*** Ten minutes time-lapses were acquired by imaging dendrite segments from cortical neurons every 5 seconds. The dimensionality of these recordings was reduced by creating maximum z projections. Images were thresholded to create outlines (A, scale bar 10 µm), which were temporally colour-coded (B, scale bar 10 µm), allowing the visualization of various events such as the percentage of spiny protrusion (relative to time 0) undergoing formation (*de novo* appearance), elimination (complete disappearance by the end of the time-lapse), lability (appearance and disappearance by the end of the time-lapse), and retraction/extension (incomplete shrinkage or growth to an existing protrusion) shown in (C, scale bar 2 µm). See Methods for further details.

expression of Panx1EGFP in Panx1 KO neurons significantly increased spiny protrusion lability; there were no significant effects of Panx1EGFP expression in WT neurons (**Figure 4*Biii***, effect size KO-EGFP *vs.* KO-Panx1EGFP: 7.31% [95CI 3.92%; 10.6%], $p = 0.0034$; effect size WT-EGFP *vs.* WT-Panx1EGFP: 2.34% [95CI -0.966%; 6.76%], $p > 0.9999$, [b3]). There was also no significant effect of EGFP expression between WT and Panx1 KO neurons (effect size WT-EGFP *vs.* KO-EGFP: -1.11%[95CI -3.17%; 0.786%], $p > 0.9999$, [b3]). Additionally, within groups transiently expressing EGFP, Panx1 KO neurons exhibited significantly reduced spiny protrusion motility (**Figure 4*Biv***, effect size WT-EGFP *vs.* KO-EGFP: -13.3% [95CI -18.5%; -8.42%], $p = 0.00016$, [b4]). Intriguingly, transient Panx1EGFP expression increased spiny protrusion motility in Panx1 KO neurons only (effect size KO-EGFP *vs.* KO-Panx1EGFP: 10.2% [95CI 4.53%; 16%], $p = 0.03582$, [b4]). Together these results suggest that spiny protrusion dynamics roughly correlate with Panx1 expression levels.

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

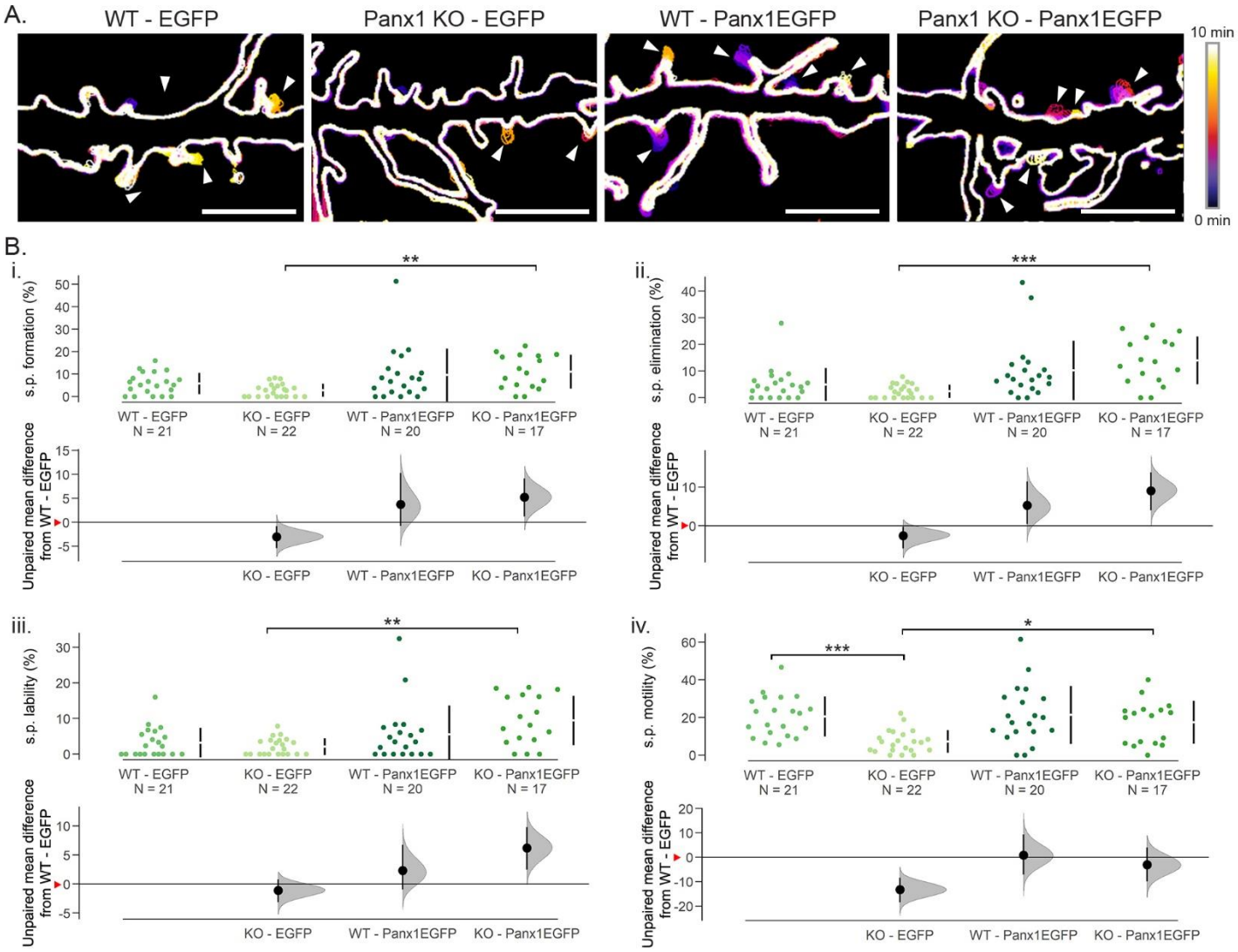


Figure 4. ***Basic characteristics of spiny protrusion dynamics in WT and Panx1 KO neurons at DIV10. A.*** Representative colour-coded traces of WT and Panx1 KO neurons co-transfected with mCherry-CD9-10 and either EGFP or Panx1EGFP showing examples of spiny protrusion formation, elimination, and lability events. ***B.*** Effect of Panx1 expression on spiny protrusion formation, elimination, lability, and motility in WT and Panx1 KO using Cumming estimation plots. ***Bi.*** Spiny protrusion formation was significantly higher in Panx1 KO neurons transiently expressing Panx1EGFP compared to those expressing EGFP (KO-EGFP: 0.2% ± 0.1%, KO-Panx1EGFP: 4.6% ± 1.3%, p = 0.0028, Kruskal-Wallis test, [b1]). No significant differences were observed between genotypes in EGFP-expressing neurons (WT-EGFP: 1.7% ± 0.7%; Panx1 KO-EGFP: 0.2% ± 0.1%, p = 0.2267, Kruskal-Wallis test, [b1]). ***Bii***. Similarly, only transient expression of Panx1EGFP in Panx1 KO neurons increased spiny protrusion elimination (KO-EGFP: 0.3% ± 0.15%; KO-Panx1EGFP: 4.6% ± 1.28%, p = 0.00024, Kruskal-Wallis test, [b2]). No significant differences were found between WT and Panx1 EGFP-expressing cells (p = 0.62307, [b2]). ***Biii.*** Spiny protrusion lability was higher in Panx1 KO neurons transfected with Panx1EGFP (KO-EGFP: 2.1% ± 0.5%; KO-Panx1EGFP: 9.4% ± 1.7%, p = 0.0034, Kruskal-Wallis test, [b3]), beyond that observed in WT expressing EGFP control (p = 0.0291, Kruskal-Wallis test, [b3]). Transient expression of Panx1EGFP in WT neurons had no significant effects (p >0.9999, [b3]). ***Biv.*** Spiny protrusion motility was significantly reduced in Panx1 KO neuron expressing EGFP control (WT-EGFP: 20.5% ± 2.3%; KO-EGFP: 7.2% ± 1.3%, p = 0.00016, Kruskal-Wallis test, [b4]). Transient Panx1EGFP expression increased spiny protrusion motility in Panx1 KO neurons only (KO-Panx1EGFP: 17.4% ± 2.8%, p = 0.03582, Kruskal-Wallis test, [b4]). Effect sizes are reported in the main text and Table 2. s.p., spiny protrusion. <0.0001, '****'; <0.001, '***'; <0.01 '**'; <0.05 '*'.

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

Panx1 KO neuron spiny protrusions are more stable

We used the basic characteristic measurements devised in Figure 3C to calculate spiny protrusion survival fraction, turnover, and overall change in movement (Δ movement). The number of spiny protrusions persisting at the end of the analysis period relative to time 0 min, referred to as survival, was significantly reduced in Panx1EGFP expressing Panx1 KO neurons (**Figure 5*A* & 5*Bi***, KO-EGFP *vs.* KO-Panx1EGFP: -10.4% [95CI -14%; -6.58%], $p = 0.00028$, [c1]). This reduction of survival surpassed that seen in WT, EGFP expressing cells (effect size WT-EGFP *vs.* KO-Panx1EGFP: -8.19% [95CI -12.3%; -3.65%], $p = 0.00909$, [c1]). We next calculated turnover by adding together formation, elimination, and lability, divided by the total number of spiny protrusions at time 0 min. With EGFP expression, turnover was significantly reduced in Panx1 KO neurons (**Figure 5*Bii***, effect size WT-EGFP *vs.* KO-EGFP: -4.48% [95CI -7.73%; -2.01%], $p = 0.0092$, [c2]). Within Panx1 KO cultures transient Panx1EGFP expression significantly increased turnover compared to EGFP control (effect size KO-EGFP *vs.* KO-Panx1EGFP: 5.24% [95CI 2.87%; 8.66%], $p = 0.0027$, [c2]). Finally, to measure the overall change in spiny protrusion movement (Δ movement), we calculated the sum of the four basic dynamic characteristics (formation, elimination, lability, and motility). Within EGFP expressing cells, Δ movement was significantly reduced in Panx1 KO neurons compared to WT controls (**Figure 5*Biii panel***, effect size WT-EGFP *vs.* KO-EGFP: -17.8 [95CI -23.6; -11.9], $p < 0.0001$). Transient Panx1EGFP expression resulted in increased Δ movement in Panx1 KO cultures only (effect size KO-EGFP *vs.* KO-Panx1EGFP: 22.8 [95CI 15; 30.4], $p = 0.00033$, [c3]). Altogether, these results suggest that Panx1 KO neuron spiny protrusions are more stable.

Discussion

Dendritic spine-based synapses account for the bulk of excitatory neurotransmission in the cerebral cortex and have been implicated in neurodevelopmental and neuropsychiatric disorders (Forrest et al., 2018; Kwon et al., 2019; Lima-Caldeira et al., 2019; Nishiyama, 2019). Although the mechanisms underlying plasticity of existing dendritic spines have been well characterized (Araya et al., 2014; Holtmaat et al., 2005; Sala & Segal, 2014; Schätzle et al., 2018), the processes involved in their formation are less well understood (Sando et al., 2017; Sigler et al., 2017; reviewed in Südhof, 2018). Here we identified a novel role for Panx1 in regulating the dynamics of developing dendritic spines, building on previous work showing that Panx1 KO cortical neurons exhibit higher dendritic spine density and more complex networks. Our results found a reciprocal relationship between Panx1 expression levels and spiny protrusion stability. Additionally, in order to make these discoveries, we optimized methods for the visualization and analysis of spiny protrusion dynamics, thereby providing a framework for others. While the current study is limited to a single timepoint (DIV10), our intriguing results suggest that additional longitudinal

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

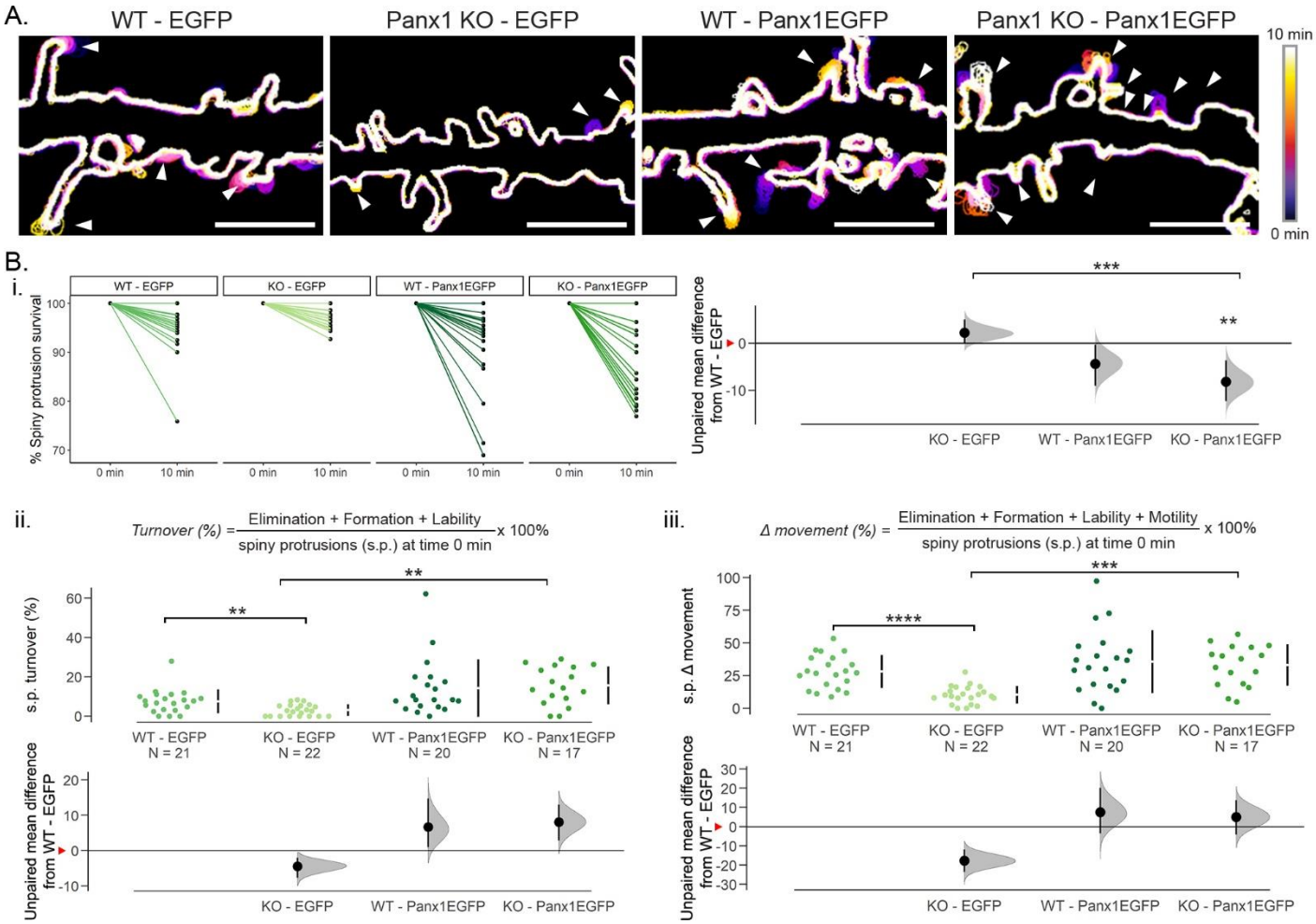


Figure 5. Panx1 KO neuron spiny protrusions are more stable. A. Representative examples of spiny protrusion motility in WT and Panx1 KO neurons co-transfected with mCherry-CD9-10 and either EGFP or Panx1EGFP. ***B.*** Cumming estimation plots of spiny protrusion second order metrics: survival fraction, turnover, and overall change in movement (Δ movement). ***Bi.*** Transient Panx1 expression in WT and Panx1 KO neurons decreased the survival fraction of spiny protrusions, albeit only significantly in Panx1 KO neurons (WT-EGFP: 94.5 ± 1.2%; WT-Panx1EGFP: 91.1% ± 1.9%, $p = 0.2034$, ^{c1}; Panx1-EGFP: 97.7% ± 0.5%; Panx1 KO-Panx1EGFP: 87.3% ± 1.9%, $p = 0.00028$, Kruskal-Wallis test, ^{c1}). ***Bii.*** In the EGFP-control-expressing group, spiny protrusion turnover was reduced in Panx1 KO neurons (WT-EGFP: 7.5% ± 1.3; Panx1-EGFP: 3.1% ± 0.6%, $p = 0.0092$, Kruskal-Wallis test, ^{c2}). Transient expression of Panx1 increased significantly spiny protrusion turnover in Panx1 KO neurons but not WT neurons (WT-Panx1EGFP: 14.2% ± 3.3%, $p > 0.9999$; Panx1 KO-Panx1EGFP: 15.6% ± 2.34%, $p = 0.0027$, Kruskal-Wallis test, ^{c2}). ***Biii.*** Spiny protrusion overall movement change (Δ movement) was reduced in Panx1 KO neurons (WT-EGFP: 28% ± 2.8%; KO-EGFP: 10.3% ± 1.5%, $p < 0.0001$, Kruskal-Wallis test, ^{c3}). Transfection of Panx1EGFP increased Δ movement in both WT (WT-Panx1EGFP: 35.5% ± 5.4%) and Panx1 KO neurons, however this effect was only significant in Panx1 KO neurons (KO-Panx1EGFP: 33% ± 3.8%, $p = 0.00033$, Kruskal-Wallis test, ^{c3}). Effect sizes are reported in the main text and Table 2. s.p., spiny protrusion. <0.0001, '****'; <0.001, '***'; <0.01 '**'; <0.05 '*'.

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

analysis is now warranted. Although *in vitro* spine plasticity characteristics correlate highly with those observed in more complex models (e.g. slice), it would be useful to confirm the current observations within these systems.

The relatively muted impact of Panx1EGFP over-expression in WT neurons, suggests that Panx1 effect on spiny protrusion dynamics is subject to saturation. Possible mechanisms of saturation could be limited machinery for trafficking supplementary Panx1 to spiny protrusions or self-regulation via ATP-dependent internalization. Alternatively, the effects of supplementary Panx1 could be constrained by limited amounts of endogenous interacting partners, such as Crmp2, Arp3c, and actin or saturation of downstream autocrine or paracrine purinergic signalling pathways related to its ATP release function (e.g. ATP stimulating glia) (Abbracchio et al., 2009; Bhalla-Gehi et al., 2010; Dahl, 2015; Wicki-Stordeur & Swayne, 2013, p.; Xu et al., 2018; D. Yang et al., 2015). In contrast to WT neurons, transient expression of Panx1EGFP exhibited significant effects on spiny protrusion dynamics in Panx1 KO cultures. Somewhat consistent with previous results, in EGFP-control-expressing cultures, loss of Panx1 precociously stabilized spiny protrusions, pointing to a fundamentally different underlying molecular organization.

Consistent with this idea, recent work has identified brain enriched and autism associated single nucleotide polymorphisms (SNPs) resulting in changes in Panx1 expression levels; although the direction of this change (i.e. decrease or increase Panx1 expression) was not identified (Davis et al., 2012). Further supporting a role for Panx1 in neuronal development, intellectual disability was observed in an individual with a germline single nucleotide polymorphism in PANX1 (Shao et al., 2016).

In addition to playing a direct role in neurodevelopment, Panx1 is also indirectly involved through its interaction with Crmp2 and purinergic receptor signalling (Boyce et al., 2015; Boyce & Swayne, 2017; reviewed in Swayne & Boyce, 2017). Crmp2 auto-antibodies have been implicated in ASD (Braunschweig et al., 2013), while suramin treatment corrected synaptic and behavioural phenotypes in the Fragile X mouse model (J. C. Naviaux et al., 2015; R. K. Naviaux et al., 2013, 2017).

In summary, this work significantly advances our understanding of the role on Panx1 in dendritic spine development and underscores the importance of additional molecular mechanistic studies investigating intrinsic (e.g. Crmp2) and extrinsic (e.g. glia) pathways.

References

Abbracchio, M. P., Burnstock, G., Verkhratsky, A., & Zimmermann, H. (2009). Purinergic signalling in the nervous system: An overview. *Trends in Neurosciences*, *32*(1), 19–29. https://doi.org/10.1016/j.tins.2008.10.001

Araya, R., Vogels, T. P., & Yuste, R. (2014). Activity-dependent dendritic spine neck changes are correlated with synaptic strength. *Proceedings of the National Academy of Sciences*, *111*(28), E2895–E2904. https://doi.org/10.1073/pnas.1321869111

Ardiles, A. O., Flores-Muñoz, C., Toro-Ayala, G., Cárdenas, A. M., Palacios, A. G., Muñoz, P., Fuenzalida, M., Sáez, J. C., & Martínez, A. D. (2014). Pannexin 1 regulates bidirectional

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

hippocampal synaptic plasticity in adult mice. *Frontiers in Cellular Neuroscience*, *8*. https://doi.org/10.3389/fncel.2014.00326

Bardy, C., Hurk, M. van den, Eames, T., Marchand, C., Hernandez, R. V., Kellogg, M., Gorris, M., Galet, B., Palomares, V., Brown, J., Bang, A. G., Mertens, J., Böhnke, L., Boyer, L., Simon, S., & Gage, F. H. (2015). Neuronal medium that supports basic synaptic functions and activity of human neurons in vitro. *Proceedings of the National Academy of Sciences*, *112*(20), E2725–E2734. https://doi.org/10.1073/pnas.1504393112

Bernard, C. (2019). Changing the Way We Report, Interpret, and Discuss Our Results to Rebuild Trust in Our Research. *ENeuro*, *6*(4). https://doi.org/10.1523/ENEURO.0259-19.2019

Bhalla-Gehi, R., Penuela, S., Churko, J. M., Shao, Q., & Laird, D. W. (2010). Pannexin1 and Pannexin3 Delivery, Cell Surface Dynamics, and Cytoskeletal Interactions. *The Journal of Biological Chemistry*, *285*(12), 9147–9160. https://doi.org/10.1074/jbc.M109.082008

Boyce, A. K. J., Epp, A. L., Nagarajan, A., & Swayne, L. A. (2018). Transcriptional and post-translational regulation of pannexins. *Biochimica et Biophysica Acta (BBA) - Biomembranes*, *1860*(1), 72–82. https://doi.org/10.1016/j.bbamem.2017.03.004

Boyce, A. K. J., Kim, M. S., Wicki-Stordeur, L. E., & Swayne, L. A. (2015). ATP stimulates pannexin 1 internalization to endosomal compartments. *Biochemical Journal*, *470*(3), 319–330. https://doi.org/10.1042/BJ20141551

Boyce, A. K. J., & Swayne, L. A. (2017). P2X7 receptor cross-talk regulates ATP-induced pannexin 1 internalization. *Biochemical Journal*, *474*(13), 2133–2144. https://doi.org/10.1042/BCJ20170257

Braunschweig, D., Krakowiak, P., Duncanson, P., Boyce, R., Hansen, R. L., Ashwood, P., Hertz-Picciotto, I., Pessah, I. N., & Van de Water, J. (2013). Autism-specific maternal autoantibodies recognize critical proteins in developing brain. *Translational Psychiatry*, *3*(7), e277–e277. https://doi.org/10.1038/tp.2013.50

Calin-Jageman, R. J., & Cumming, G. (2019). Estimation for Better Inference in Neuroscience. *ENeuro*, *6*(4). https://doi.org/10.1523/ENEURO.0205-19.2019

Chiu, Y.-H., Schappe, M. S., Desai, B. N., & Bayliss, D. A. (2018). Revisiting multimodal activation and channel properties of Pannexin 1. *Journal of General Physiology*, *150*(1), 19–39. https://doi.org/10.1085/jgp.201711888

Dahl, G. (2015). ATP release through pannexon channels. *Philosophical Transactions of the Royal Society B: Biological Sciences*, *370*(1672), 20140191. https://doi.org/10.1098/rstb.2014.0191

Davis, L. K., Gamazon, E. R., Kistner-Griffin, E., Badner, J. A., Liu, C., Cook, E. H., Sutcliffe, J. S., & Cox, N. J. (2012). Loci nominally associated with autism from genome-wide analysis show enrichment of brain expression quantitative trait loci but not lymphoblastoid cell line expression quantitative trait loci. *Molecular Autism*, *3*(1), 3. https://doi.org/10.1186/2040-2392-3-3

Dvoriantchikova, G., Ivanov, D., Barakat, D., Grinberg, A., Wen, R., Slepak, V. Z., & Shestopalov, V. I. (2012). Genetic Ablation of Pannexin1 Protects Retinal Neurons from Ischemic Injury. *PLOS ONE*, *7*(2), e31991. https://doi.org/10.1371/journal.pone.0031991

Fiala, J. C., Feinberg, M., Popov, V., & Harris, K. M. (1998). Synaptogenesis Via Dendritic Filopodia in Developing Hippocampal Area CA1. *Journal of Neuroscience*, *18*(21), 8900–8911. https://doi.org/10.1523/JNEUROSCI.18-21-08900.1998

Forrest, M. P., Parnell, E., & Penzes, P. (2018). Dendritic structural plasticity and neuropsychiatric disease. *Nature Reviews Neuroscience*, *19*(4), 215–234. https://doi.org/10.1038/nrn.2018.16

Frank, A. C., Huang, S., Zhou, M., Gdalyahu, A., Kastellakis, G., Silva, T. K., Lu, E., Wen, X., Poirazi, P., Trachtenberg, J. T., & Silva, A. J. (2018). Hotspots of dendritic spine turnover facilitate clustered spine addition and learning and memory. *Nature Communications*, *9*(1), 1–11. https://doi.org/10.1038/s41467-017-02751-2

Gajardo, I., Salazar, C. S., Lopez-Espíndola, D., Estay, C., Flores-Muñoz, C., Elgueta, C., Gonzalez-Jamett, A. M., Martínez, A. D., Muñoz, P., & Ardiles, Á. O. (2018). Lack of Pannexin 1 Alters Synaptic GluN2 Subunit Composition and Spatial Reversal Learning in Mice. *Frontiers in Molecular Neuroscience*, *11*. https://doi.org/10.3389/fnmol.2018.00114

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

Ho, J., Tumkaya, T., Aryal, S., Choi, H., & Claridge-Chang, A. (2019). Moving beyond P values: Data analysis with estimation graphics. *Nature Methods*, *16*(7), 565–566. https://doi.org/10.1038/s41592-019-0470-3

Holtmaat, A. J. G. D., Trachtenberg, J. T., Wilbrecht, L., Shepherd, G. M., Zhang, X., Knott, G. W., & Svoboda, K. (2005). Transient and Persistent Dendritic Spines in the Neocortex In Vivo. *Neuron*, *45*(2), 279–291. https://doi.org/10.1016/j.neuron.2005.01.003

Hoshiba, Y., Wada, T., & Hayashi-Takagi, A. (2017). Synaptic Ensemble Underlying the Selection and Consolidation of Neuronal Circuits during Learning. *Frontiers in Neural Circuits*, *11*. https://doi.org/10.3389/fncir.2017.00012

Kovalzon, V. M., Moiseenko, L. S., Ambaryan, A. V., Kurtenbach, S., Shestopalov, V. I., & Panchin, Y. V. (2017). Sleep-wakefulness cycle and behavior in pannexin1 knockout mice. *Behavioural Brain Research*, *318*, 24–27. https://doi.org/10.1016/j.bbr.2016.10.015

Kwon, T., Merchán-Pérez, A., Rial Verde, E. M., Rodríguez, J.-R., DeFelipe, J., & Yuste, R. (2019). Ultrastructural, Molecular and Functional Mapping of GABAergic Synapses on Dendritic Spines and Shafts of Neocortical Pyramidal Neurons. *Cerebral Cortex*, *29*(7), 2771–2781. https://doi.org/10.1093/cercor/bhy143

Li, W., Ma, L., Yang, G., & Gan, W.-B. (2017). REM sleep selectively prunes and maintains new synapses in development and learning. *Nature Neuroscience*, *20*(3), 427–437. https://doi.org/10.1038/nn.4479

Lima-Caldeira, G., Peça, J., & Carvalho, A. L. (2019). New insights on synaptic dysfunction in neuropsychiatric disorders. *Current Opinion in Neurobiology*, *57*, 62–70. https://doi.org/10.1016/j.conb.2019.01.004

Mancuso, J. J., Chen, Y., Li, X., Xue, Z., & Wong, S. T. C. (2013). Methods of dendritic spine detection: From Golgi to high-resolution optical imaging. *Neuroscience*, *251*, 129–140. https://doi.org/10.1016/j.neuroscience.2012.04.010

McDonald, J. (2014). *Handbook of Biological Statistics. [Online]. Baltimore: Sparky House*.

Naviaux, J. C., Wang, L., Li, K., Bright, A. T., Alaynick, W. A., Williams, K. R., Powell, S. B., & Naviaux, R. K. (2015). Antipurinergic therapy corrects the autism-like features in the Fragile X (Fmr1 knockout) mouse model. *Molecular Autism*, *6*(1), 1. https://doi.org/10.1186/2040-2392-6-1

Naviaux, R. K., Curtis, B., Li, K., Naviaux, J. C., Bright, A. T., Reiner, G. E., Westerfield, M., Goh, S., Alaynick, W. A., Wang, L., Capparelli, E. V., Adams, C., Sun, J., Jain, S., He, F., Arellano, D. A., Mash, L. E., Chukoskie, L., Lincoln, A., & Townsend, J. (2017). Low-dose suramin in autism spectrum disorder: A small, phase I/II, randomized clinical trial. *Annals of Clinical and Translational Neurology*, *4*(7), 491–505. https://doi.org/10.1002/acn3.424

Naviaux, R. K., Zolkipli, Z., Wang, L., Nakayama, T., Naviaux, J. C., Le, T. P., Schuchbauer, M. A., Rogac, M., Tang, Q., Dugan, L. L., & Powell, S. B. (2013). Antipurinergic Therapy Corrects the Autism-Like Features in the Poly(IC) Mouse Model. *PLOS ONE*, *8*(3), e57380. https://doi.org/10.1371/journal.pone.0057380

Nishiyama, J. (2019). Plasticity of dendritic spines: Molecular function and dysfunction in neurodevelopmental disorders. *Psychiatry and Clinical Neurosciences*, *73*(9), 541–550. https://doi.org/10.1111/pcn.12899

Penuela, S., Bhalla, R., Gong, X.-Q., Cowan, K. N., Celetti, S. J., Cowan, B. J., Bai, D., Shao, Q., & Laird, D. W. (2007). Pannexin 1 and pannexin 3 are glycoproteins that exhibit many distinct characteristics from the connexin family of gap junction proteins. *Journal of Cell Science*, *120*(21), 3772–3783. https://doi.org/10.1242/jcs.009514

Phillips, M., & Pozzo-Miller, L. (2015). Dendritic spine dysgenesis in autism related disorders. *Neuroscience Letters*, *601*, 30–40. https://doi.org/10.1016/j.neulet.2015.01.011

Prochnow, N., Abdulazim, A., Kurtenbach, S., Wildförster, V., Dvoriantchikova, G., Hanske, J., Petrasch-Parwez, E., Shestopalov, V. I., Dermietzel, R., Manahan-Vaughan, D., & Zoidl, G. (2012). Pannexin1 Stabilizes Synaptic Plasticity and Is Needed for Learning. *PLOS ONE*, *7*(12), e51767. https://doi.org/10.1371/journal.pone.0051767

Ray, A., Zoidl, G., Weickert, S., Wahle, P., & Dermietzel, R. (2005). Site-specific and developmental expression of pannexin1 in the mouse nervous system. *Eur J Neurosci*, *21*(12), 3277–3290.

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

Sala, C., & Segal, M. (2014). Dendritic Spines: The Locus of Structural and Functional Plasticity. *Physiological Reviews*, *94*(1), 141–188. https://doi.org/10.1152/physrev.00012.2013

Sanchez-Arias, J. C., Liu, M., Choi, C. S. W., Ebert, S. N., Brown, C. E., & Swayne, L. A. (2019). Pannexin 1 Regulates Network Ensembles and Dendritic Spine Development in Cortical Neurons. *ENeuro*, *6*(3), ENEURO.0503-18.2019. https://doi.org/10.1523/ENEURO.0503-18.2019

Sando, R., Bushong, E., Zhu, Y., Huang, M., Considine, C., Phan, S., Ju, S., Uytiepo, M., Ellisman, M., & Maximov, A. (2017). Assembly of Excitatory Synapses in the Absence of Glutamatergic Neurotransmission. *Neuron*, *94*(2), 312-321.e3. https://doi.org/10.1016/j.neuron.2017.03.047

Schätzle, P., Silva, M. E. da, Tas, R. P., Katrukha, E. A., Hu, H. Y., Wierenga, C. J., Kapitein, L. C., & Hoogenraad, C. C. (2018). Activity-Dependent Actin Remodeling at the Base of Dendritic Spines Promotes Microtubule Entry. *Current Biology*, *28*(13), 2081-2093.e6. https://doi.org/10.1016/j.cub.2018.05.004

Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., Preibisch, S., Rueden, C., Saalfeld, S., Schmid, B., Tinevez, J.-Y., White, D. J., Hartenstein, V., Eliceiri, K., Tomancak, P., & Cardona, A. (2012). Fiji: An open-source platform for biological-image analysis. *Nature Methods*, *9*(7), 676–682. https://doi.org/10.1038/nmeth.2019

Shao, Q., Lindstrom, K., Shi, R., Kelly, J., Schroeder, A., Juusola, J., Levine, K. L., Esseltine, J. L., Penuela, S., Jackson, M. F., & Laird, D. W. (2016). A germline variant in PANX1 has reduced channel function and is associated with multisystem dysfunction. *J Biol Chem*. https://doi.org/10.1074/jbc.M116.717934

Sigler, A., Oh, W. C., Imig, C., Altas, B., Kawabe, H., Cooper, B. H., Kwon, H.-B., Rhee, J.-S., & Brose, N. (2017). Formation and Maintenance of Functional Spines in the Absence of Presynaptic Glutamate Release. *Neuron*, *94*(2), 304-311.e4. https://doi.org/10.1016/j.neuron.2017.03.029

Spano, G. M., Banningh, S. W., Marshall, W., Vivo, L. de, Bellesi, M., Loschky, S. S., Tononi, G., & Cirelli, C. (2019). Sleep Deprivation by Exposure to Novel Objects Increases Synapse Density and Axon–Spine Interface in the Hippocampal CA1 Region of Adolescent Mice. *Journal of Neuroscience*, *39*(34), 6613–6625. https://doi.org/10.1523/JNEUROSCI.0380-19.2019

Südhof, T. C. (2018). Towards an Understanding of Synapse Formation. *Neuron*, *100*(2), 276–293. https://doi.org/10.1016/j.neuron.2018.09.040

Swayne, L. A., & Boyce, A. K. J. (2017). Regulation of Pannexin 1 Surface Expression by Extracellular ATP: Potential Implications for Nervous System Function in Health and Disease. *Frontiers in Cellular Neuroscience*, *11*. https://doi.org/10.3389/fncel.2017.00230

Vogt, A., Hormuzdi, S. G., & Monyer, H. (2005). Pannexin1 and Pannexin2 expression in the developing and mature rat brain. *Molecular Brain Research*, *141*(1), 113–120. https://doi.org/10.1016/j.molbrainres.2005.08.002

Wicki-Stordeur, L. E., & Swayne, L. A. (2013). Panx1 regulates neural stem and progenitor cell behaviours associated with cytoskeletal dynamics and interacts with multiple cytoskeletal elements. *Cell Communication and Signaling*, *11*(1), 62. https://doi.org/10.1186/1478-811X-11-62

Xu, X., Wicki-Stordeur, L. E., Sanchez-Arias, J. C., Liu, M., Weaver, M. S., Choi, C. S. W., & Swayne, L. A. (2018). Probenecid Disrupts a Novel Pannexin 1-Collapsin Response Mediator Protein 2 Interaction and Increases Microtubule Stability. *Frontiers in Cellular Neuroscience*, *12*. https://doi.org/10.3389/fncel.2018.00124

Yang, D., He, Y., Muñoz-Planillo, R., Liu, Q., & Núñez, G. (2015). Caspase-11 Requires the Pannexin-1 Channel and the Purinergic P2X7 Pore to Mediate Pyroptosis and Endotoxic Shock. *Immunity*, *43*(5), 923–932. https://doi.org/10.1016/j.immuni.2015.10.009

Yang, G., & Gan, W.-B. (2012). Sleep contributes to dendritic spine formation and elimination in the developing mouse somatosensory cortex. *Developmental Neurobiology*, *72*(11), 1391–1398. https://doi.org/10.1002/dneu.20996

Zack, G. W., Rogers, W. E., & Latt, S. A. (1977). Automatic measurement of sister chromatid exchange frequency. *Journal of Histochemistry & Cytochemistry*, *25*(7), 741–753. https://doi.org/10.1177/25.7.70454

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.02.973917; this version posted March 2, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.

Ziv, N. E., & Smith, S. J. (1996). Evidence for a Role of Dendritic Filopodia in Synaptogenesis and Spine Formation. *Neuron*, *17*(1), 91–102. https://doi.org/10.1016/S0896-6273(00)80283-4

Zoidl, G., Petrasch-Parwez, E., Ray, A., Meier, C., Bunse, S., Habbes, H.-W., Dahl, G., & Dermietzel, R. (2007). Localization of the pannexin1 protein at postsynaptic sites in the cerebral cortex and hippocampus. *Neuroscience*, *146*(1), 9–16. https://doi.org/10.1016/j.neuroscience.2007.01.061