Full Title: Boysenberry and apple juice concentrate reduces acute lung inflammation through increased alternatively activated macrophage activity in an acute mouse model of allergic airways disease

Odette M. Shaw[1*], Roger D Hurst[2], Janine Cooney[3], Gregory M. Sawyer[1], Hannah Dinnan[1], Sheridan Martell[1]

[1]Nutrition & Health Group, Food Innovation Portfolio, The New Zealand Institute for Plant and Food Research Limited, Palmerston North, New Zealand

[2] Food Innovation Portfolio, The New Zealand Institute for Plant and Food Research Limited, Palmerston North, New Zealand

[3]Biological Chemistry & Bioactives Group, Food Innovation Portfolio, The New Zealand Institute for Plant and Food Research Limited, Hamilton, New Zealand

*__Correspondence:__

Odette Shaw

odette.shaw@plantandfood.co.nz

Short title: M2 macrophage reduce acute allergic airways inflammation

Abstract:

Bioactive compounds such as anthocyanins, proanthocyanins and other polyphenols are found in a wide variety of fruits and vegetables, and consumption of these have been associated with reduced lung inflammation and improved lung function in asthma and other lung diseases. We investigated whether a combination of Boysenberry and apple juice, found in BerriQi® Boysenberry and apple juice concentrate, could reduce the allergic airways inflammation associated with asthma. We characterised the polyphenolic components in BerriQi® Boysenberry and apple juice concentrate and identified the main compounds as cyanidin glycosides, ellagitannins, and chlorogenic acid. We found that consumption of 2.5 mg/kg of total anthocyanins from the BerriQi® Boysenberry and apple juice concentrate significantly reduced eosinophil infiltration following acute ovalbumin (OVA) exposure in a mouse model of allergic airways inflammation. We found that BerriQi® Boysenberry and apple juice concentrate consumption increased M2 (CD206+) macrophages and the production of the M2-associated cytokines CXCL10 and CCL4 within the lung. These results suggest that consumption of BerriQi® Boysenberry and apple juice concentrate promotes a shift towards an anti-inflammatory environment within the lung leading to reduced immune cell infiltration and tissue damage.

Keywords: allergic airways inflammation, Boysenberry, apple, anthocyanins, ellagitannins, alternatively activated macrophages.

1. Introduction

Asthma is a heterogeneous, chronic, inflammatory lung disease characterized by reversible airways obstruction, bronchospasm and infiltration of immune cells (1-3). It is estimated that 150 million people are affected by asthma worldwide, with a 5–15% prevalence in children (4) and there is evidence that early life exposure to air pollution caused by vehicle exhaust, environmental dust and industrial processes, increases the severity of asthma in children (5-7). The respiratory symptoms such as cough, and wheeze are worsened by exposure to pollution (8). Proinflammatory cytokine production in response to allergens by immune cells are further increased with concomitant pollution exposure (9-13). Eosinophils, in particular, produce reactive oxygen species, and cytokines, leading to epithelial damage and contribute to mucosal inflammation and the recruitment of other proinflammatory immune cells (14-17). These repeated acute inflammatory responses lead to tissue damage and remodelling, contributing to airway hyperresponsiveness, mucus cell hyperplasia, fixed airway flow obstruction, and loss of lung function over time (18-21).

Large-scale epidemiological studies have found that increased fruit and vegetable consumption correlates with reduced asthma symptoms (22-25). These dietary-related improvements in lung function benefits are also seen in people living in polluted environments (26-28). Fruits and vegetables contain numerous bioactive compounds, including anthocyanins and procyanidins, which have been shown to attenuate lung inflammation in cell and animal models of allergy and asthma (29-34). Human population studies have identified that dietary intake of foods high in polyphenols (28) such as apples, pears (35), carrots, tomatoes (25) and citrus is inversely correlated with the frequency and severity of reported asthma symptoms, especially wheezing and coughing (22, 25, 35, 36). Previously, we have identified

that Boysenberry consumption led to decreased chronic lung inflammation, and improved lung tissue repair in an animal model of chronic allergic lung inflammation (33). Boysenberries contain high concentrations of anthocyanins and ellagitannins (37, 38). We have also found that procyanidin-rich extracts from apple suppressed IL-4 mediated cytokine production in cell culture models of lung epithelial allergic inflammation (32, 39).

There is increasing interest in understanding the mechanisms of action that specific plant bioactives have in the human body. This is partially to better understand the benefits of consuming specific fruits and vegetables, and partially to add value to specific foods through validated health claims. There is also interest in determining if combining specific plants containing different polyphenols can augment the health benefits above those seen with the individual plant. Use of animal models, where dietary intake can be tightly controlled are useful for both demonstrating/revealing the efficacy for identified compounds, and determining the biological mechanisms of action. The aim of this study was to determine whether the combination of Boysenberries and apple, as found in BerriQi® Boysenberry and apple juice concentrate at a dose of 2.5 mg/kg total anthocyanins (TAC) could reduce allergic airways inflammation in response to acute ovalbumin exposure in a mouse model system. We also sought to determine the mechanisms involved in any ameliorating effect.

2. Methods

2.1 Mice and Materials

C57BL/6J male mice were group housed on 12h light/dark cycle in a conventional animal facility at The New Zealand Institute for Plant and Food Research Limited

(Palmerston North, New Zealand). Mice were fed Prodiet RMH1800 standard chow for rodents (Lab Diet, St Louis, MO, USA) and filtered water ad libitum throughout the study, all attempts to minimise suffering were made. All experimental procedures were approved by the AgResearch Grasslands Animal Ethics Committee (AE approvals #14839, #14731 and #14016) and carried out in accordance with the Animal Welfare Act (1999). A proprietary Boysenberry and apple juice concentrate (BerriQi®) was supplied by Anagenix Ltd (Auckland, New Zealand). Legendplex™ 13-plex Th cytokine, proinflammatory cytokine and proinflammatory chemokine panels, Zombie NIR™ fixable viability dye, and anti-mouse CD3 (clone 17A2), CD4 (clone GK1.5), CD8a (clone 53-6.7), CD80 (clone 16-10A1), CD86 (clone GL-1), CD11c (clone N418), CD45 (clone 30-F11), CD206 (clone C068C2), CD14 (clone Sa14-2), Ly6C (clone HK1.4), Gr-1 (clone RB6-8C5), I-A/I-E (MHC class II; clone M5/114.15.2), and F4/80 (clone BM8) were purchased from Biolegend (San Diego, CA, USA). Anti-mouse SiglecF (clone E50-2440) and CD11b (clone M1/70) were from BD Biosciences (San Jose, CA, USA). Ovalbumin (OVA), and Alum were purchased from Sigma (Auckland, New Zealand). iScript Advanced cDNA kit was from Bio-Rad Laboratories (Hercules, CA, USA). Unless otherwise stated, all cell culture media, supplements, Taqman probes and buffers were purchased from Life Technologies NZ (Auckland, NZ).

2.2 BerriQi® Formulation and Chemical Composition Analysis

Boysenberries (*Rubus ursinus* var loganbaccus cv Boysenberry), harvested between January-March 2017 from the Nelson region of NZ, were processed to a juice concentrate by Boysenberries NZ (Nelson, NZ). Apples (*Malus domestica* –mixed varieties) were harvested between April-June 2017 from the Hawkes Bay region of New Zealand, and juice concentrates prepared by Profruit Ltd. (Hastings, NZ). The

freshly harvested Boysenberries and apples were graded, milled and pressed into juice, de-seeded by centrifugation, pasteurised, depectinised by enzymatic treatment, clarified by centrifugation, and concentrated by evaporation by their respective manufacturers to yield fruit juice concentrates, each with an 8-fold concentration factor.

Shott NZ Ltd. (Wellington, NZ) then blended the juice concentrates (proprietary proportions), and added potassium sorbate preservative to a final concentration of 0.12% (w/w), yielding a dark red, non-cloudy syrup. The final BerriQi® Boysenberry and apple juice concentrate product measured pH 3.39 (by pH meter) and was 68.3° Brix (by refractometry), with a specific gravity of 1.35 mg/mL comprising 68% solids measured by both gravimetric and drying methods.

The polyphenol content of the BerriQi® Boysenberry and apple juice concentrate was determined by liquid chromatography-mass spectrometry (LC-MS) using an LTQ linear ion trap mass spectrometer fitted with an ESI interface (ThermoFisher Scientific, San Jose, CA, USA) coupled to an Ultimate 3000 UHPLC and PDA detector (Dionex, Sunnyvale, CA, USA). Anthocyanins were separated on a Poroshell 120 SB-C18, 2.1x150 mm, 2.7 µm, analytical LC column (Agilent, Torrance, CA, USA), maintained at 70°C. The solvents were (A) 5:3:92 acetonitrile:formic acid:water v/v/v and (B) acetonitrile + 0.1% formic acid (flow rate, 200 µL/min). The initial mobile phase, 100% A, was held for 2 min before being ramped linearly to 88% A at 14 min, returning to 5% A at 15 min and held for 4 min before resetting to the original conditions. The sample injection volume was 10 µL. The MS data were acquired in the positive mode. Anthocyanin concentrations are reported as cyanidin-3-*O*-glucoside equivalents.

Other phenolic compound separation was achieved using a Hypersil GOLD aQ 1.9µ C18 175Å (Thermo Scientific, Waltham, MA, USA), 150 × 2.1 mm column maintained at 45°C. The solvents were (A) water + 0.1% formic acid and (B) acetonitrile + 0.1% formic acid (flow rate, 200 µL/min). The initial mobile phase, 95% A/5% B, was ramped linearly to 85% A at 10 min, held for 3.75 min, then ramped linearly to 75% A at 18 min, 67.2% A at 25 min, 50% A at 28 min, 3% A at 29 min and held for 4 min before resetting to the original conditions. The sample injection volume was 4 µL. The MS data were acquired in the negative mode.

2.3 OVA-Induced Airway Inflammation Model

Allergic airway disease was induced as previously described (34, 40). For the Boysenberry and apple interventions, mice were randomized into receiving either water (vehicle control) or 2.5 mg/kg TAC in the BerriQi® Boysenberry and apple juice concentrate as previously described (34). Briefly, mice were fasted for 4 h before being orally gavaged with water (control) or at a dose of 2.5 mg/kg body weight TAC in the BerriQi® Boysenberry and apple juice concentrate made up to a total volume of 200 µL in water 1h before OVA challenge and again 2 days post-challenge. Mice were sacrificed 4 days following intranasal ovalbumin challenge and immune parameters were analyzed.

2.4 Immune Parameter Analysis

Bronchoalveolar lavage fluid (BALF) and lung tissues were collected as previously described and immune cells were phenotyped by flow cytometry (40). Lung tissue supernatant for cytokine analysis was prepared as previously described (34). Briefly, the left lung lobe was minced into 500 µL phenol red-free complete RPMI media and incubated at 37˚C for 30 min before being filtered through a 40 µm mesh and

centrifuged to remove cellular material. The resulting supernatant was analysed for cytokine concentration using Legendplex bead-based multiplex immunoassays as per the manufacturer's instruction. Both cell phenotyping and the cytokine multiplex assays were analyzed using a BD FACSverse (BD Biosciences, San Jose, CA, USA). H&E and AB-PAS histological staining were performed by Massey IVABS histology unit.

2.5 Real-time qPCR analysis

Mouse lung tissue was collected 4 days following OVA challenge and snap frozen in liquid nitrogen. The lung samples were crushed into powder using a mortar and pestle with liquid nitrogen to keep the samples frozen. The RNA was extracted from the powder using a TRIZOL total RNA extraction protocol. RNA was quantified using an LVis plate in a POLARstar Omega plate reader (BMG) and the quality of the ribosomal RNA bands confirmed by agarose gel electrophoresis (data not shown). Five µg of RNA from each sample was used as the template for cDNA synthesis using the iScript™ cDNA Synthesis Kit. Taqman® Gene Expression Assays were purchased for each gene of interest. Two housekeeping genes, GAPDH and β-actin, were used as controls to determine the amount of relative gene expression. Taqman® Gene Expression Master Mix was used to PCR amplify the genes in a Bio-Rad™ CFX384™ Real-Time PCR Detection System. Three lung samples per treatment group were prepared and amplified in quadruplicate with the housekeeping genes amplified on the same 384-well plate.

2.6 Statistical Analysis

Data were analyzed using one-way analysis of variance (ANOVA) with a Tukey's post hoc test and graphed in SigmaPlot 12.5 (Systat Software Inc., San Jose, CA, USA).

3. Results

3.1 Chemical Composition of the Boysenberry and Apple Juice Concentrate

The results of the LC-MS analysis showed that cyanidin glycosides, ellagitannins, and chlorogenic acid were the major components in BerriQi® Boysenberry and apple juice concentrate (Table 1, supplementary figures 1, 2). Minor components included phloretin 2-O-glucoside, and a mix of phenolic acids, flavonol glycosides, flavanol monomers and procyanidins. The major classes of phenolic compounds were anthocyanins (1969 µg/mL) and hydrolysable tannins (946 µg/mL), accounting for 56% and 27%, respectively, of the total phenolics quantified. The most abundant tannins were ellagic acid (449 µg/mL) and sanguiin H6 (213 µg/mL).

3.2 Effect of Boysenberry and Apple juice concentrate Intervention on Ovalbumin-Induced Allergic Airways Inflammation

Acute intranasal OVA exposure resulted in an infiltration of immune cells into the lung (Figure 1A) and increased mucous production (Figure 1B). Consumption of 2.5 mg/kg TAC BerriQi® Boysenberry and apple juice concentrate reduced the infiltration of immune cells and decreased OVA-induced mucous production (Figure 1A-B). We quantified the type and number of immune cells infiltrating into the lung, and found that acute OVA exposure significantly increased ($P<0.001$) infiltrating eosinophils (CD45+/CD11b+/SiglecF+), neutrophils (CD45+Ly6C+Gr-1+) and T-cells (CD45+/CD3+/CD4+ or CD45+/CD3+/CD8a+), compared with the lung of naïve animals (Figure 1C-F). Compared with animals only exposed to OVA, those that also

consumed 2.5 mg/kg TAC BerriQi® Boysenberry and apple juice concentrate showed a significant decrease ($P<0.001$) in the number of infiltrating eosinophils, neutrophils and T cells in the lung (Figure 1C-F). We saw no change in the number of CD4+ or CD8+ T cells in the mediastinal (lung-draining) lymph node for any of the treatment groups (Figure 2A-B).

There was a trend towards an increased percentage of CD206+/CD14- macrophages in the lungs of mice that consumed 2.5 mg/kg TAC BerriQi® Boysenberry and apple juice concentrate (Figure 3A). We measured the gene expression of Arg1, Ym-1 and Fizz1 in lung tissue and found that 2.5 mg/kg TAC BerriQi® Boysenberry and apple juice concentrate consumption led to a significant fold- increase in Arg1 ($P<0.05$) and Fizz1 ($P<0.01$) gene expression compared to naïve mice (Table 2). We found no significant fold-change in Nos2 or Ym-1 gene expression between any of the treatment groups (Table 2).

Consumption of 2.5 mg/kg TAC BerriQi® Boysenberry and apple juice concentrate led to increased levels of the cytokines IL-17A, CXCL10, and CCL4 (Figure 3B-D) 4 days following OVA challenge, but did not affect the IL-17F concentration (Figure 3E). We saw no effect on the concentrations of the classical Th1/2 cytokines IFNγ, IL-5, IL-9 or IL-10 in either the BerriQi® Boysenberry and apple juice concentrate treated or the OVA alone mice compared to naïve controls (Figure 2C-F).

4. Discussion

We evaluated the effects of dietary supplementation with 2.5 mg/kg TAC BerriQi® Boysenberry and apple juice concentrate, on the immune responses in a mouse model of acute allergic airways inflammation. Our results show that consumption of 2.5 mg/kg TAC BerriQi® Boysenberry and apple juice concentrate reduced

granulocyte and local T cell infiltration into the lung after OVA challenge, but did not alter T cell activation within the lung draining lymph node or the levels of classical Th-2 and Th-1 cytokines in the lung at four days following OVA challenge. Our current results indicated that BerriQi® Boysenberry and apple juice concentrate had little impact on the Th-2/Th-1 mediated allergic response of mice, but rather targeted innate proinflammatory immune pathways. This is consistent with our previously reported finding in a mouse model of chronic allergic airways inflammation using 10 mg/kg TAC Boysenberry juice concentrate (33). Chemical composition analysis showed that the BerriQi® Boysenberry and apple juice concentrate formulation contained high concentrations of cyanidin glycosides, ellagitannins, and chlorogenic acid. These compounds have been previously shown to reduce inflammatory signalling in vitro (38, 41, 42), and in vivo animal models of inflammation (34, 43-46). Our current results suggest that consumption of 2.5 mg/kg TAC BerriQi® Boysenberry and apple juice concentrate, which also contains high levels of ellagitannins and chlorogenic acid, could have broader lung health benefits beyond allergic asthma disease by promoting the resolution of inflammation caused by innate immune cell overactivation. Current asthma therapies also suppress both the adaptive and innate immune responses without affecting the aberrant sensitivity to the allergen and this can lead to adverse events.

Consumption of BerriQi® Boysenberry and apple juice concentrate had less of an effect on monocyte/macrophage infiltration into the lung than on granulocyte infiltration, and there was increased percentage of CD206+ monocytes. This could represent a shift to an M2 anti-inflammatory phenotype. We then measured the changes in gene expression for Arg1, Ym-1 and Fizz1, the classic genes for identifying alternatively activated macrophages (47, 48). Arg1 and Fizz1 gene

255 expression was significantly increased with BerriQi® Boysenberry and apple juice
256 concentrate consumption. Consistent with a shift to a more anti-inflammatory
257 macrophage phenotype, we detected increased levels of CXCL10 and CCL4
258 cytokines, which are produced by M2 macrophages, in the lungs of mice that
259 consumed BerriQi® Boysenberry and apple juice concentrate. These results suggest
260 that the consumption of Boysenberry and apple juice concentrate led to a switch to
261 the M2 phenotype in OVA-challenged mice. This could be one of the mechanisms by
262 which BerriQi® Boysenberry and apple juice concentrate consumption contributed to
263 the resolution of inflammation. Previously, we reported that 10 mg/kg TAC
264 Boysenberry juice concentrate can increase the abundance of alternatively activated
265 (M2) macrophages, which promote tissue repair in a chronic model of airways
266 inflammation (33). Our current results suggest that BerriQi® Boysenberry and apple
267 juice concentrate, had a similar effect in this mouse model of acute inflammation, in
268 particular the increase in Arg1 gene expression is similar to our previously reported
269 study showing increased arginase protein expression by alternatively activated
270 macrophages (33). Further, research looking at an animal model Th2-mediated
271 inflammation has identified M2 macrophage derived Fizz1 as a key limiting factor for
272 Th2-mediated pulmonary inflammation (49).

273 The mice that consumed 2.5 mg/kg TAC BerriQi® Boysenberry and apple juice
274 concentrate showed increased levels of the cytokines IL-17A, CXCL10, and CCL4,
275 but the levels of IL-17F were not affected. High IL-17 and IL-17F levels have been
276 implicated in asthma pathogenesis, however, there is also evidence that IL-17A (50)
277 can increase the abundance of MMP-9, an important tissue remodelling protein in
278 asthma (33) as well as inducing apoptosis of neutrophils and eosinophils (50).
279 CXCL10 and CCL4 are chemokines that attract monocytes/macrophages, and

CXCL10 may also inhibit the infiltration of eosinophils in response to allergic airways inflammation (51). The combination of increased IL-17A-mediated granulocyte apoptosis and CXCL10-mediated inhibition of granulocyte infiltration could explain how the consumption of the BerriQi® Boysenberry and apple juice concentrate resulted in decreased allergic airways inflammation, in particular the reduced number of eosinophils and neutrophils.

This switch to a M2 macrophage phenotype may be through the Boysenberry and apple polyphenols identified in the BerriQi® Boysenberry and apple juice concentrate directly inhibiting proinflammatory pathways, or through an indirect shortening of the proinflammatory phase. Other studies have shown that increased dietary fibre shortened the duration of the proinflammatory phase, leading to reduced tissue damage in a chronic mouse model of allergic asthma (52). Based on the estimated fibre content in BerriQi® Boysenberry and apple juice concentrate (Supplementary table 1), it is unlikely that the dietary fibre component played a significant role in the immune modulation we observed. However, the combination of anthocyanins with other polyphenols identified in the BerriQi® Boysenberry and apple juice concentrate could have a similar effect on inflammation either by shortening the proinflammatory time course, or promoting the production of anti-inflammatory proteins. Ellagitannins have been shown in cell culture and animal models of chronic inflammatory diseases to reduce proinflammatory prostaglandins (53), cytokines (45, 54), and other proteins (42, 55, 56). Anthocyanins have also been shown to inhibit proinflammatory proteins (57, 58), and activate anti-inflammatory pathways in models of inflammation (59-63). It is possible that the combination of the different polyphenols in the BerriQi® Boysenberry and apple juice concentrate act on a number of different immune pathways to regulate the immune responses to OVA.

We found that mice that consumed BerriQi® Boysenberry and apple juice concentrate had reduced immune cell infiltration in response to acute OVA challenge and this could be as a result of a shift towards an anti-inflammatory environment within the lung. These results highlight the potential of anthocyanin-rich Boysenberry and apple dietary supplementation to modulate innate immune pathways during acute allergic lung inflammation. Further work is needed to determine if these pathways are also altered in other lung inflammatory conditions, such as air pollution exposure. Clinical studies are needed to show if these findings are translatable to human health.

Conflict of Interest

All authors were employed by The New Zealand Institute of Plant & Food Research Limited, a New Zealand Crown Research Institute wholly owned by the New Zealand Government for the purposes of research into sustainable production, elite breeding, food and health science of horticultural, arable, and seafood products.

OMS and RDH report that they are named on patents related to the formulation of BerriQi® Boysenberry and apple juice concentrate, but have not received any financial compensation, nor will receive any personal royalty payments as a result of this. None of the other authors declare any other conflicts of interest. Under the terms of the Innovation Cell™ Collaboration Agreement Plant & Food Research and Anagenix have a royalty sharing agreement for any royalties that result from the sale of BerriQi® Boysenberry and apple juice concentrate product.

Author Contributions

OMS designed, performed, analyzed and interpreted the in vivo studies, and wrote and edited the manuscript; JC performed, analyzed and interpreted the chemcial composition experiments and GMS performed, analyzed and interpreted in vivo studies, and both contributed to the writing and editing of the manuscript; HD and SM performed the in vivo studies and helped edit the manuscript; RDH designed and directed the overall research programme and helped edit the manuscript.

Funding

This work was funded by The New Zealand Institute of Plant & Food Research Limited as a contribution to the Innovation Cell™ Collaboration Agreement executed between Anagenix Limited and The New Zealand Institute for Plant and Food Research Limited on 5 September 2016 (PFR reference #33609).

The authors declare that this study received funding from The New Zealand Institute of Plant & Food Research Limited. The funder was not involved in the study design, collection, analysis, and interpretation of data, the writing of this article or the decision to submit it for publication.

Acknowledgements

The authors acknowledge the efforts of TC Chadderton, and Brendan Vercoe in securing internal funding for this work. The authors would like to thank Andrew Carroll for assisting with data analysis and Jenny Smith for her invaluable help preparing the manuscript.

5. References

1. Agache I, Akdis CA. Endotypes of allergic diseases and asthma: An important step in building blocks for the future of precision medicine. Allergol Int. 2016;65(3):243-52.

2. Agrawal DK, Bharadwaj A. Allergic airway inflammation. Curr Allergy Asthma Rep. 2005;5(2):142-8.

3. Barnes PJ. Pathophysiology of asthma. Br J Clin Pharmacol. 1996;42(1):3-10.

4. WHO. Prevention of allergy and allergic asthma: based on the WHO/WAO Meeting on the Prevention of Allergy and Allergic Asthma, Geneva, 8-9 January 2002. Management DoND, Arthritis CRDa, Organization WA, Organization. WH, editors. Geneva: World Health Organization; 2003.

5. Hsu HH, Chiu YH, Coull BA, Kloog I, Schwartz J, Lee A, et al. Prenatal Particulate Air Pollution and Asthma Onset in Urban Children. Identifying Sensitive Windows and Sex Differences. Am J Respir Crit Care Med. 2015;192(9):1052-9.

6. Miller RL, Peden DB. Environmental effects on immune responses in patients with atopy and asthma. J Allergy Clin Immunol. 2014;134(5):1001-8.

7. Jung DY, Leem JH, Kim HC, Kim JH, Hwang SS, Lee JY, et al. Effect of Traffic-Related Air Pollution on Allergic Disease: Results of the Children's Health and Environmental Research. Allergy Asthma Immunol Res. 2015;7(4):359-66.

8. Hoek G, Pattenden S, Willers S, Antova T, Fabianova E, Braun-Fahrlander C, et al. PM10, and children's respiratory symptoms and lung function in the PATY study. Eur Respir J. 2012;40(3):538-47.

9. Acciani TH, Brandt EB, Khurana Hershey GK, Le Cras TD. Diesel exhaust particle exposure increases severity of allergic asthma in young mice. Clin Exp Allergy. 2013;43(12):1406-18.

10. Brandt EB, Biagini Myers JM, Acciani TH, Ryan PH, Sivaprasad U, Ruff B, et al. Exposure to allergen and diesel exhaust particles potentiates secondary allergen-specific memory responses, promoting asthma susceptibility. J Allergy Clin Immunol. 2015;136(2):295-303 e7.

11. Carlsten C, Blomberg A, Pui M, Sandstrom T, Wong SW, Alexis N, et al. Diesel exhaust augments allergen-induced lower airway inflammation in allergic individuals: a controlled human exposure study. Thorax. 2016;71(1):35-44.

12. Kim J, Natarajan S, Vaickus LJ, Bouchard JC, Beal D, Cruikshank WW, et al. Diesel exhaust particulates exacerbate asthma-like inflammation by increasing CXC chemokines. Am J Pathol. 2011;179(6):2730-9.

13. van Voorhis M, Knopp S, Julliard W, Fechner JH, Zhang X, Schauer JJ, et al. Exposure to atmospheric particulate matter enhances Th17 polarization through the aryl hydrocarbon receptor. PLoS One. 2013;8(12):e82545.

385 14. Trivedi SG, Lloyd CM. Eosinophils in the pathogenesis of allergic airways
386 disease. Cell Mol Life Sci. 2007;64(10):1269-89.
387 15. Amin K, Janson C, Bystrom J. Role of Eosinophil Granulocytes in Allergic
388 Airway Inflammation Endotypes. Scand J Immunol. 2016;84(2):75-85.
389 16. Bossley CJ, Fleming L, Gupta A, Regamey N, Frith J, Oates T, et al. Pediatric
390 severe asthma is characterized by eosinophilia and remodeling without T(H)2
391 cytokines. J Allergy Clin Immunol. 2012;129(4):974-82 e13.
392 17. Brown JR, Kleimberg J, Marini M, Sun G, Bellini A, Mattoli S. Kinetics of
393 eotaxin expression and its relationship to eosinophil accumulation and activation in
394 bronchial biopsies and bronchoalveolar lavage (BAL) of asthmatic patients after
395 allergen inhalation. Clin Exp Immunol. 1998;114(2):137-46.
396 18. Bergeron C, Al-Ramli W, Hamid Q. Remodeling in asthma. Proc Am Thorac
397 Soc. 2009;6(3):301-5.
398 19. Ahdieh M, Vandenbos T, Youakim A. Lung epithelial barrier function and
399 wound healing are decreased by IL-4 and IL-13 and enhanced by IFN-gamma. Am J
400 Physiol Cell Physiol. 2001;281(6):C2029-38.
401 20. Al-Muhsen S, Johnson JR, Hamid Q. Remodeling in asthma. J Allergy Clin
402 Immunol. 2011;128(3):451-62.
403 21. Brightling CE, Gupta S, Gonem S, Siddiqui S. Lung damage and airway
404 remodelling in severe asthma. Clin Exp Allergy. 2012;42(5):638-49.
405 22. McKeever TM, Britton J. Diet and asthma. Am J Respir Crit Care Med.
406 2004;170(7):725-9.
407 23. Nurmatov U, Devereux G, Sheikh A. Nutrients and foods for the primary
408 prevention of asthma and allergy: systematic review and meta-analysis. J Allergy
409 Clin Immunol. 2011;127(3):724-33 e1-30.

410 24. Okoko BJ, Burney PG, Newson RB, Potts JF, Shaheen SO. Childhood
411 asthma and fruit consumption. Eur Respir J. 2007;29(6):1161-8.
412 25. Rosenlund H, Kull I, Pershagen G, Wolk A, Wickman M, Bergstrom A. Fruit
413 and vegetable consumption in relation to allergy: disease-related modification of
414 consumption? J Allergy Clin Immunol. 2011;127(5):1219-25.
415 26. Stevens EL, Rosser F, Forno E, Peden D, Celedon JC. Can the effects of
416 outdoor air pollution on asthma be mitigated? J Allergy Clin Immunol.
417 2019;143(6):2016-8 e1.
418 27. Burbank AJ, Duran CG, Pan Y, Burns P, Jones S, Jiang Q, et al. Gamma
419 tocopherol-enriched supplement reduces sputum eosinophilia and endotoxin-
420 induced sputum neutrophilia in volunteers with asthma. J Allergy Clin Immunol.
421 2018;141(4):1231-8 e1.
422 28. Pounis G, Arcari A, Costanzo S, Di Castelnuovo A, Bonaccio M, Persichillo M,
423 et al. Favorable association of polyphenol-rich diets with lung function: Cross-
424 sectional findings from the Moli-sani study. Respir Med. 2018;136:48-57.
425 29. Park SJ, Shin WH, Seo JW, Kim EJ. Anthocyanins inhibit airway inflammation
426 and hyperresponsiveness in a murine asthma model. Food Chem Toxicol.
427 2007;45(8):1459-67.
428 30. Coleman SL, Shaw OM. Progress in the understanding of the pathology of
429 allergic asthma and the potential of fruit proanthocyanidins as modulators of airway
430 inflammation. Food Funct. 2017;8(12):4315-24.
431 31. Coleman S, Kruger M, Sawyer G, Hurst R. Procyanidin A2 Modulates IL-4-
432 Induced CCL26 Production in Human Alveolar Epithelial Cells. Int J Mol Sci.
433 2016;17(11):1888.

434 32. Sawyer GM, Stevenson DE, McGhie TK, Hurst RD. Suppression of CCL26
435 and CCL11 generation in human alveolar epithelial cells by apple extracts containing
436 procyanidins. J Funct Foods. 2017;31:141-51.
437 33. Shaw OM, Hurst RD, Harper JL. Boysenberry ingestion supports fibrolytic
438 macrophages with the capacity to ameliorate chronic lung remodeling. Am J Physiol
439 Lung Cell Mol Physiol. 2016;311(3):L628-38.
440 34. Shaw OM, Nyanhanda T, McGhie TK, Harper JL, Hurst RD. Blackcurrant
441 anthocyanins modulate CCL11 secretion and suppress allergic airway inflammation.
442 Mol Nutr Food Res. 2017;61(9):1600868.
443 35. Garcia-Larsen V, Thawer N, Charles D, Cassidy A, van Zele T, Thilsing T, et
444 al. Dietary Intake of Flavonoids and Ventilatory Function in European Adults: A
445 GA(2)LEN Study. Nutrients. 2018;10(1).
446 36. Garcia V, Arts IC, Sterne JA, Thompson RL, Shaheen SO. Dietary intake of
447 flavonoids and asthma in adults. Eur Respir J. 2005;26(3):449-52.
448 37. Cooney JM, Jensen DJ, McGhie TK. LC-MS identification of anthocyanins in
449 boysenberry extract and anthocyanin metabolites in human urine following dosing. J
450 Sci Food Agr. 2004;84(3):237-45.
451 38. McGhie TK, Martin H, Lunken RC. The combination of analytical-scale HPLC
452 separation with a TR-FRET assay to investigate JAK2 inhibitory compounds in a
453 Boysenberry drink. Food Funct. 2012;3(11):1170-5.
454 39. Coleman SL, Hurst RD, Sawyer GM, Kruger MC. The in vitro evaluation of
455 isolated procyanidins as modulators of cytokine-induced eotaxin production in
456 human alveolar epithelial cells. J Berry Res. 2016;6(2):115-24.
457 40. Shaw OM, Harper JL. An efficient single prime protocol for the induction of
458 antigen-induced airways inflammation. J Immunol Methods. 2013;395(1-2):79-82.

459 41. Cassidy A, Rogers G, Peterson JJ, Dwyer JT, Lin H, Jacques PF. Higher
460 dietary anthocyanin and flavonol intakes are associated with anti-inflammatory
461 effects in a population of US adults. Am J Clin Nutr. 2015;102(1):172-81.
462 42. El-Shitany NA, El-Bastawissy EA, El-desoky K. Ellagic acid protects against
463 carrageenan-induced acute inflammation through inhibition of nuclear factor kappa
464 B, inducible cyclooxygenase and proinflammatory cytokines and enhancement of
465 interleukin-10 via an antioxidant mechanism. Int Immunopharmacol. 2014;19(2):290-
466 9.
467 43. Auclair S, Silberberg M, Gueux E, Morand C, Mazur A, Milenkovic D, et al.
468 Apple polyphenols and fibers attenuate atherosclerosis in apolipoprotein E-deficient
469 mice. J Agric Food Chem. 2008;56(14):5558-63.
470 44. Denis MC, Roy D, Yeganeh PR, Desjardins Y, Varin T, Haddad N, et al. Apple
471 peel polyphenols: a key player in the prevention and treatment of experimental
472 inflammatory bowel disease. Clin Sci (Lond). 2016;130(23):2217-37.
473 45. Guan S, Zheng YY, Yu X, Li WL, Han B, Lu J. Ellagic acid protects against
474 LPS-induced acute lung injury through inhibition of nuclear factor kappa B,
475 proinflammatory cytokines and enhancement of interleukin-10. Food Agr Immunol.
476 2017;28(6):1347-61.
477 46. Impellizzeri D, Talero E, Siracusa R, Alcaide A, Cordaro M, Maria Zubelia J,
478 et al. Protective effect of polyphenols in an inflammatory process associated with
479 experimental pulmonary fibrosis in mice. Br J Nutr. 2015;114(6):853-65.
480 47. Chung S, Lee TJ, Reader BF, Kim JY, Lee YG, Park GY, et al. FoxO1
481 regulates allergic asthmatic inflammation through regulating polarization of the
482 macrophage inflammatory phenotype. Oncotarget. 2016;7(14):17532-46.

483 48. Kurowska-Stolarska M, Stolarski B, Kewin P, Murphy G, Corrigan CJ, Ying S,
484 et al. IL-33 amplifies the polarization of alternatively activated macrophages that
485 contribute to airway inflammation. J Immunol. 2009;183(10):6469-77.
486 49. Nair MG, Du Y, Perrigoue JG, Zaph C, Taylor JJ, Goldschmidt M, et al.
487 Alternatively activated macrophage-derived RELM-{alpha} is a negative regulator of
488 type 2 inflammation in the lung. J Exp Med. 2009;206(4):937-52.
489 50. Linden A, Dahlen B. Interleukin-17 cytokine signalling in patients with asthma.
490 Eur Respir J. 2014;44(5):1319-31.
491 51. Su YC, Rolph MS, Hansbro NG, Mackay CR, Sewell WA. Granulocyte-
492 macrophage colony-stimulating factor is required for bronchial eosinophilia in a
493 murine model of allergic airway inflammation. J Immunol. 2008;180(4):2600-7.
494 52. Trompette A, Gollwitzer ES, Yadava K, Sichelstiel AK, Sprenger N, Ngom-Bru
495 C, et al. Gut microbiota metabolism of dietary fiber influences allergic airway disease
496 and hematopoiesis. Nat Med. 2014;20(2):159-66.
497 53. Karlsson S, Nanberg E, Fjaeraa C, Wijkander J. Ellagic acid inhibits
498 lipopolysaccharide-induced expression of enzymes involved in the synthesis of
499 prostaglandin E2 in human monocytes. Br J Nutr. 2010;103(8):1102-9.
500 54. Cornelio Favarin D, Martins Teixeira M, Lemos de Andrade E, de Freitas
501 Alves C, Lazo Chica JE, Arterio Sorgi C, et al. Anti-inflammatory effects of ellagic
502 acid on acute lung injury induced by acid in mice. Mediators Inflamm.
503 2013;2013:164202.
504 55. Marin M, Maria Giner R, Rios JL, Recio MC. Intestinal anti-inflammatory
505 activity of ellagic acid in the acute and chronic dextrane sulfate sodium models of
506 mice colitis. J Ethnopharmacol. 2013;150(3):925-34.

507 56. Saba, Khan S, Parvez S, Chaudhari B, Ahmad F, Anjum S, et al. Ellagic acid
508 attenuates bleomycin and cyclophosphamide-induced pulmonary toxicity in Wistar
509 rats. Food Chem Toxicol. 2013;58:210-9.
510 57. Esposito D, Chen A, Grace MH, Komarnytsky S, Lila MA. Inhibitory effects of
511 wild blueberry anthocyanins and other flavonoids on biomarkers of acute and chronic
512 inflammation in vitro. J Agric Food Chem. 2014;62(29):7022-8.
513 58. Fu Y, Zhou E, Wei Z, Wang W, Wang T, Yang Z, et al. Cyanidin-3-O-beta-
514 glucoside ameliorates lipopolysaccharide-induced acute lung injury by reducing
515 TLR4 recruitment into lipid rafts. Biochem Pharmacol. 2014;90(2):126-34.
516 59. Chen YF, Shibu MA, Fan MJ, Chen MC, Viswanadha VP, Lin YL, et al. Purple
517 rice anthocyanin extract protects cardiac function in STZ-induced diabetes rat hearts
518 by inhibiting cardiac hypertrophy and fibrosis. J Nutr Biochem. 2016;31:98-105.
519 60. Edirisinghe I, Banaszewski K, Cappozzo J, McCarthy D, Burton-Freeman BM.
520 Effect of black currant anthocyanins on the activation of endothelial nitric oxide
521 synthase (eNOS) in vitro in human endothelial cells. J Agric Food Chem.
522 2011;59(16):8616-24.
523 61. Khanna S, Roy S, Bagchi D, Bagchi M, Sen CK. Upregulation of oxidant-
524 induced VEGF expression in cultured keratinocytes by a grape seed
525 proanthocyanidin extract. Free Radic Biol Med. 2001;31(1):38-42.
526 62. Koh ES, Lim JH, Kim MY, Chung S, Shin SJ, Choi BS, et al. Anthocyanin-rich
527 Seoritae extract ameliorates renal lipotoxicity via activation of AMP-activated protein
528 kinase in diabetic mice. J Transl Med. 2015;13(1):203.
529 63. Liu Y, Tan D, Tong C, Zhang Y, Xu Y, Liu X, et al. Blueberry anthocyanins
530 ameliorate radiation-induced lung injury through the protein kinase RNA-activated
531 pathway. Chem Biol Interact. 2015;242:363-71.

Table 1. Phenolic compounds detected in BerriQi® (µg/mL). M^+ and $(M-H)^-$ ions are the pseudomolecular ions used for identification of compounds by liquid chromatography-mass spectrometry (LC-MS). All identifications confirmed by MS/MS^n experiments. Peak numbers refer to chromatograms shown in supplementary data. # detected as $[M+formate]^-$ adduct

Peak	M^+	$(M-H)^-$	Compound	µg/mL
			Anthocyanins	
1	611		Cyanidin 3-*O*-sophoroside	883
2	449		Cyanidin 3-*O*-glucoside	571
3	481		Cyanidin 3-*O*-sambubioside	24
4	757		Cyanidin 3-*O*-(2-glucosylrutinoside)	411
5	595		Cyanidin 3-*O*-rutinoside	62
6	727		Cyanidin 3-*O*-xylosylrutinoside	18
			Phenolic acids	
7		169	Gallic acid	140
8		153	Protocatechuric acid	35
10		353	Chlorogenic acid	69
11		179	Caffeic acid	7
15		337	4-*p*-Coumaroylquinic acid	26
17		337	5-*p*-Coumaroylquinic acid	3
			Flavan-3-ols and procyanidins	
9		335#	Catechin	3
12		577	Procyanidin B2	6
13		335#	*Epi*-catechin	21
25		579	unknown procyanidin isomer	3
			Hydrolysable tannins	
14		1567	Sanguiin H10 isomer 1	9
16		469	Sanguisorbic acid dilactone	120
19		2036.5	Galloyl-SH6	66
20		1567	Sanguiin H10 isomer 2	61
21		2501	Lambertian C (minus ellagic acid)	11
22		2803	Lambertian C	17
24		1869	Sanguiin H6	213
26		301	Ellagic acid	449
			Flavonols	
27		609	Quercetin 3-*O*-rutinoside	6
28		463	Quercetin 3-*O*-galactoside	17

29	477	Quercetin 3-*O*-glucuronide	36
30	463	Quercetin 3-*O*-glucoside	13
31	433	Quercetin 3-*O*-pentoside 1	9
32	433	Quercetin 3-*O*-pentoside 3	6
33	433	Quercetin 3-*O*-pentoside 2	13
35	447	Quercetin 3-*O*-rhamnoside	15
37	301	Quercetin	19
		Chalcones	
34	567	Phloretin 2-*O*-xylo-glucoside	9
36	481[#]	Phloretin 2-*O*-glucoside	62
		Unknowns	
18	563[#]	unknown	71
23	639	unknown	22
		Total	**1557**

Table 2: BerriQi® Boysenberry and apple juice concentrate increases alternatively activated macrophage gene expression in the lung. Mice were primed with ovalbumin (OVA)/Alum i.p. and then challenged 7 days later with OVA i.n. (Day 0). Mice were orally gavaged with 2.5 mg/kg total anthocyanins (TAC) in the BerriQi® Boysenberry and apple juice concentrate (BerriQi) 1h before OVA challenge and again 2 days post-challenge. Mean fold-change (SEM) in gene expression was measured by real-time qPCR in lung tissue 4 days post-OVA challenge. *P<0.05, **P<0.01 compared with naïve (one-way ANOVA with Tukey's Post Hoc test) for 4 experimental replicates with n=3 per treatment groups.

Target Gene	Naïve	OVA	OVA + BerriQi
Arg1	1 (0.5)	9.4 (5.3)	26.0 (7.9) *
Ym-1	1 (0.4)	4.4 (1.9)	5.5 (1.0)
Fizz1	1 (0.5)	15.4 (0.9) **	19.6 (3.3) **
Nos2	1 (0.3)	1.5 (0.8)	1.7 (0.5)

548 **Figure 1: BerriQi® Boysenberry and apple juice concentrate suppresses**
549 **ovalbumin-induced airway inflammation, and immune cell infiltration.** Mice
550 were primed with ovalbumin (OVA)/Alum i.p and then challenged 7 days later with
551 OVA i.n (Day 0). Mice were orally gavaged with 2.5 mg/kg total anthocyanins (TAC)
552 in the BerriQi® Boysenberry and apple juice concentrate (BerriQi) 1h before OVA
553 challenge and again 2 days post-challenge. (A) Haematoxylin and eosin stained lung
554 tissue from naïve, OVA-challenged and OVA-challenged mice treated with BerriQi®
555 Boysenberry and apple juice concentrate. Magnification 10x (top) and 20x (bottom)
556 Asterisk =cell infiltration. (B) Alcian-blue Periodic acid-Schiff stained lung tissue from
557 naïve, OVA-challenged and OVA-challenged mice treated with BerriQi®
558 Boysenberry and apple juice concentrate. Magnification 10x (top) and 20x (bottom).
559 Arrow=mucous producing goblet cells. (C) Total eosinophil, (D) Total neutrophil, (E)
560 CD4+ T cells and CD8+ T cells in bronchioalveolar lavage fluid (BALF) were
561 determined 4 days post-OVA challenge. Data presented as mean ± SEM $P<0.001$
562 compared with naïve and OVA challenge + BerriQi® Boysenberry and apple juice
563 concentrate (one-way ANOVA with Tukey's Post Hoc test) for two experimental
564 replicates with n=10 per treatment groups.

565 **Figure 2: BerriQi® Boysenberry and apple juice concentrate does not alter**
566 **classical Th-1/Th-2 cells and cytokines.** Mice were primed with ovalbumin
567 (OVA)/Alum i.p and then challenged 7 days later with OVA i.n (Day 0). Mice were
568 orally gavaged with 2.5 mg/kg total anthocyanins (TAC) in the BerriQi® Boysenberry
569 and apple juice concentrate (BerriQi) 1h before OVA challenge and again 2 days
570 post-challenge. Mediastinal lymph node (MSLN) (A) CD4+ and (B) CD8+ T cells
571 number; and lung tissue production of (C) IL-5, (D) IL-9, (E) IL-10 and (F) IFNγ were

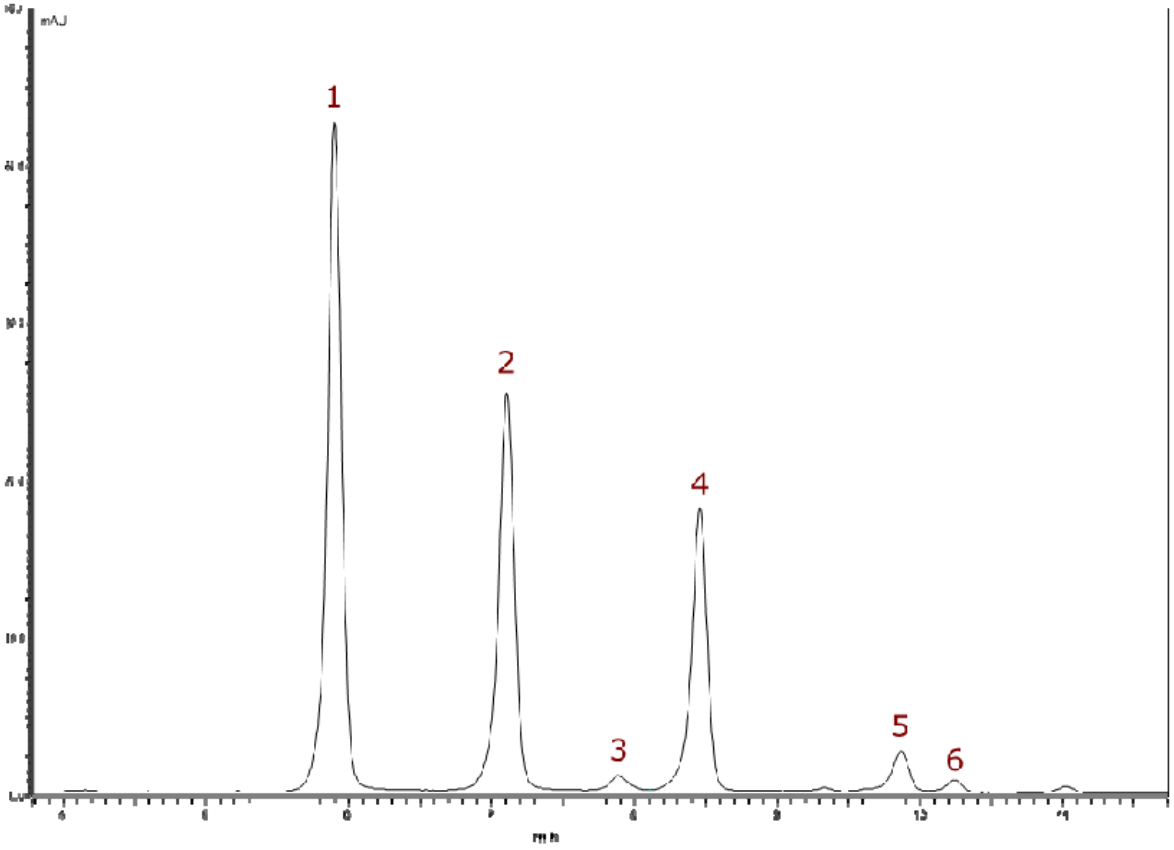
572 determined 4 days post-OVA challenge by Legendplex. Data presented as mean ±
573 SEM for two experimental replicates with n=10 per treatment groups.

574 **Figure 3: BerriQi® Boysenberry and apple juice concentrate suppresses**
575 **ovalbumin-induced airway inflammation through increased IL-17A, CXCL10**
576 **and CCL4 concentration.** Mice were primed with ovalbumin (OVA)/Alum i.p and
577 then challenged 7 days later with OVA i.n (Day 0). Mice were orally gavaged with 2.5
578 mg/kg total anthocyanins (TAC) in the BerriQi® Boysenberry and apple juice
579 concentrate (BerriQi) 1h before OVA challenge and again 2 days post-challenge. (A)
580 Percentage of CD206+ macrophages in bronchioalveolar lavage fluid (BALF) and
581 lung tissue production of (B) CXCL10, (C) IL-17A, (D) CCL4 and (E) IL-17F was
582 determined 4 days post-OVA challenge by Legendplex. Data presented as mean ±
583 SEM, $P<0.05$ compared with OVA challenge + BerriQi® Boysenberry and apple juice
584 concentrate, $P<0.01$ compared with naïve and OVA challenge (one-way ANOVA with
585 Tukey's Post Hoc test) for two experimental replicates with n=10 per treatment
586 groups.

587

588 ## SUPPLEMENTARY DATA

589



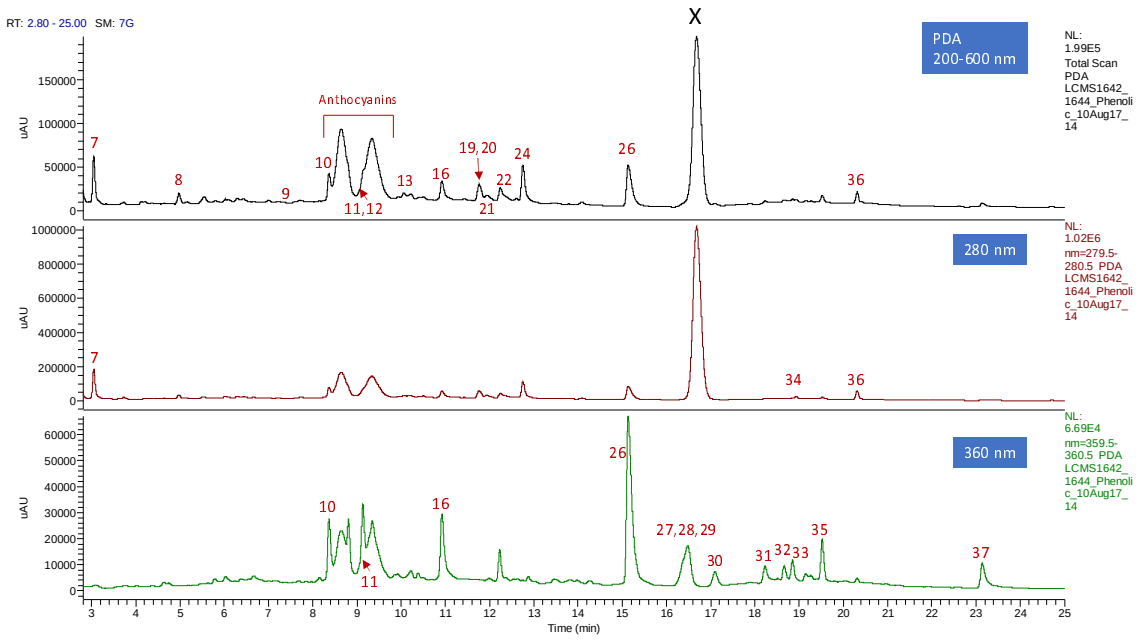
590

591 **Supplementary Figure 1.** Liquid Chromatography-Mass Spectrometry (LC-MS)
592 chromatogram for BerriQi® concentrate, showing the anthocyanin UV/VIS profile
593 measured at 520 nm. Peak numbers refer to compounds listed in Table 1.

594

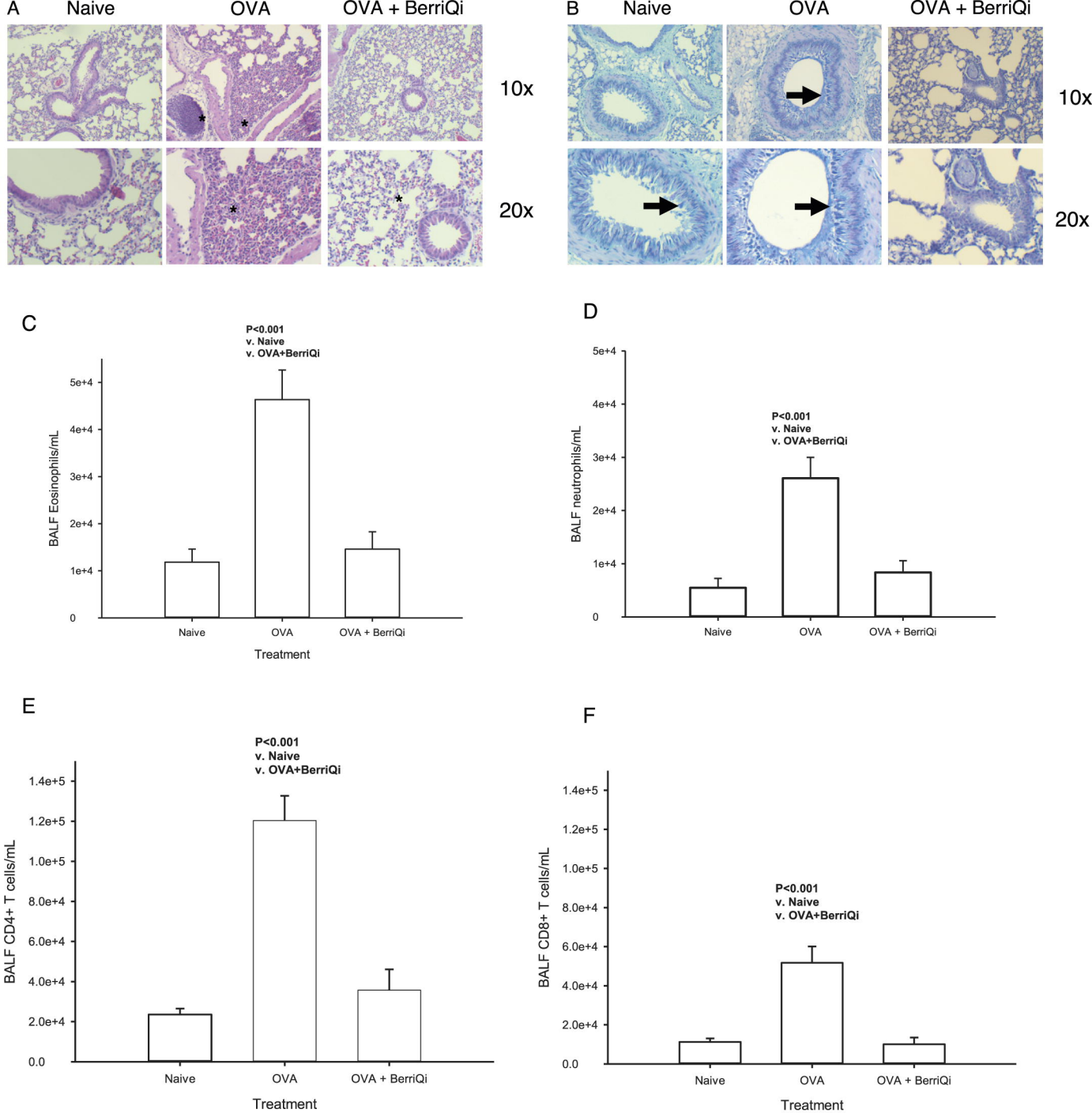
595

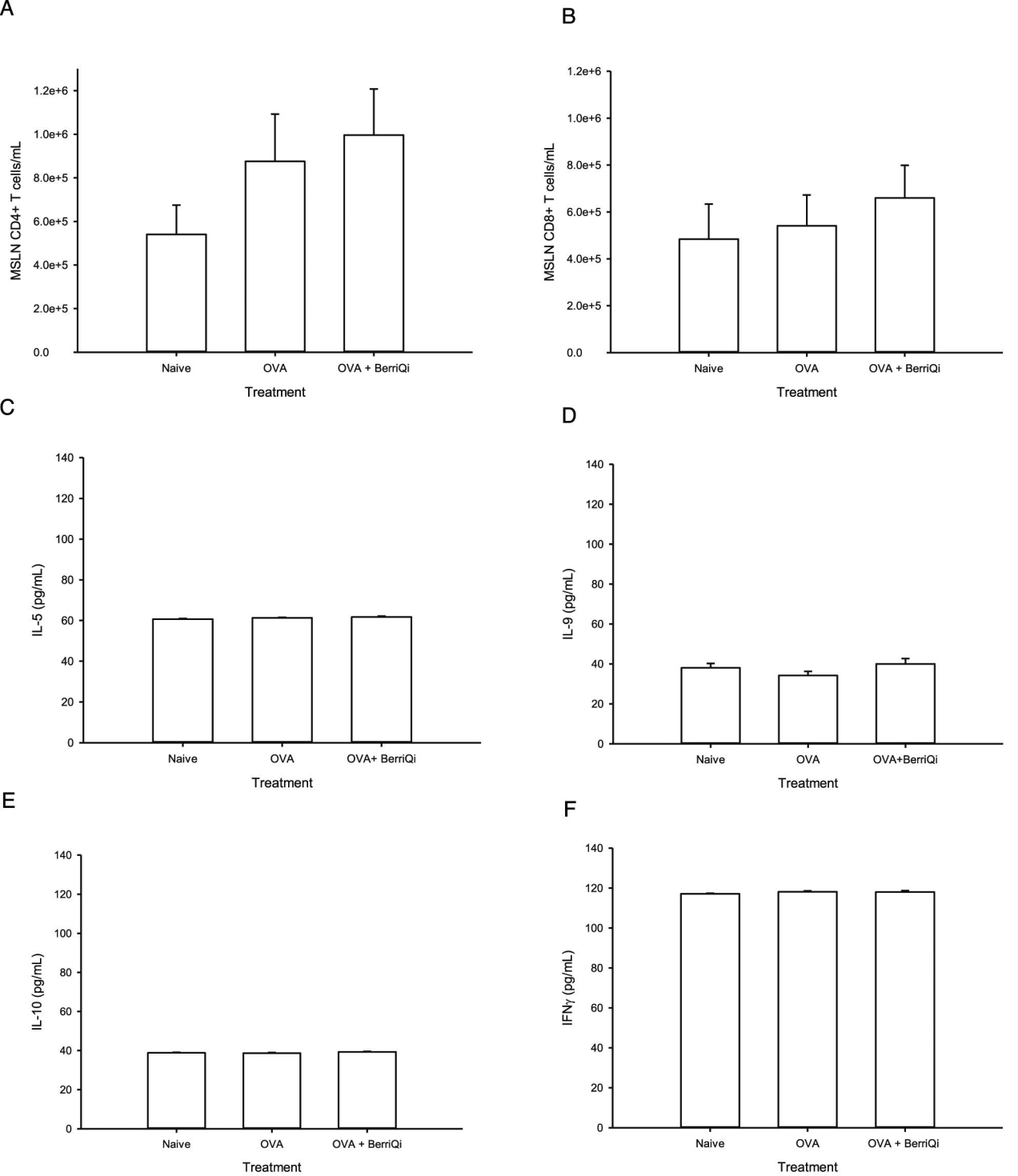
596

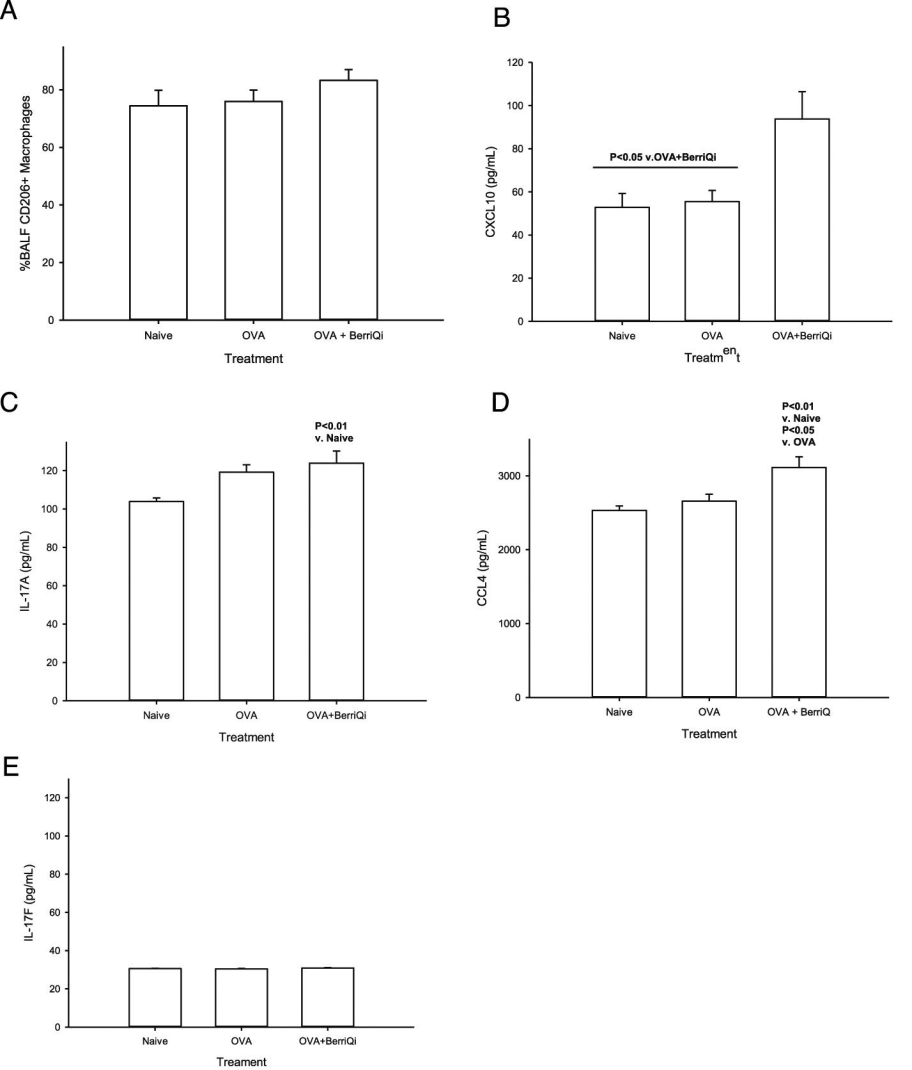


597

598 **Supplementary Figure 2.** Liquid Chromatography-Mass Spectrometry (LC-MS)
599 chromatograms for BerriQi® concentrate, showing PDA (photodiode array) phenolic
600 profiles measured at 200–600 nm, 280 nm and 360 nm. Peak numbers refer to
601 compounds listed in Table 1. X, denotes sorbic acid, a preservative added to the
602 BerriQi concentrate during formulation. This was present at a concentration of ~2000
603 µg/mL.


A
Naive
OVA
OVA + BerriQi
10x
20x
B
Naive
OVA
OVA + BerriQi
10x
20x
C
P<0.001
v. Naive
v. OVA+BerriQi
BALF Eosinophils/mL
5e+4
4e+4
3e+4
2e+4
1e+4
0
Naive
OVA
OVA + BerriQi
Treatment
D
P<0.001
v. Naive
v. OVA+BerriQi
BALF neutrophils/mL
5e+4
4e+4
3e+4
2e+4
1e+4
0
Naive
OVA
OVA + BerriQi
E
P<0.001
v. Naive
v. OVA+BerriQi
BALF CD4+ T cells/mL
1.4e+5
1.2e+5
1.0e+5
8.0e+4
6.0e+4
4.0e+4
2.0e+4
0.0
Naive
OVA
OVA + BerriQi
Treatment
F
P<0.001
v. Naive
v. OVA+BerriQi
BALF CD8+ T cells/mL
1.4e+5
1.2e+5
1.0e+5
8.0e+4
6.0e+4
4.0e+4
2.0e+4
0.0
Naive
OVA
OVA + BerriQi
Treatment


A
MSLN CD4+ T cells/mL
1.2e+6
1.0e+6
8.0e+5
6.0e+5
4.0e+5
2.0e+5
0.0
Naive
OVA
OVA + BerriQi
Treatment
B
MSLN CD8+ T cells/mL
1.2e+6
1.0e+6
8.0e+5
6.0e+5
4.0e+5
2.0e+5
0.0
Naive
OVA
OVA + BerriQi
Treatment
C
IL-5 (pg/mL)
140
120
100
80
60
40
20
0
Naive
OVA
OVA+ BerriQi
Treatment
D
IL-9 (pg/mL)
140
120
100
80
60
40
20
0
Naive
OVA
OVA+BerriQi
Treatment
E
IL-10 (pg/mL)
140
120
100
80
60
40
20
0
Naive
OVA
OVA + BerriQi
Treatment
F
IFNγ (pg/mL)
140
120
100
80
60
40
20
0
Naive
OVA
OVA + BerriQi
Treatment


A
%BALF CD206+ Macrophages
0
20
40
60
80
Naive
OVA
OVA + BerriQi
Treatment
B
CXCL10 (pg/mL)
0
20
40
60
80
100
120
P<0.05 v.OVA+BerriQi
Naive
OVA
OVA+BerriQi
Treatment
C
IL-17A (pg/mL)
0
20
40
60
80
100
120
P<0.01
v. Naive
Naive
OVA
OVA+BerriQi
Treatment
D
CCL4 (pg/mL)
0
1000
2000
3000
P<0.01
v. Naive
P<0.05
v. OVA
Naive
OVA
OVA + BerriQ
Treatment
E
IL-17F (pg/mL)
0
20
40
60
80
100
120
Naive
OVA
OVA+BerriQi
Treament