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Long-read error correction: a survey and qualitative comparison

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Abstract

Third generation sequencing technologies Pacific Biosciences and Oxford Nanopore Technologies were respectively made available in 2011 and 2014. In contrast with second generation sequencing technologies such as Illumina, these new technologies allow the sequencing of long reads of tens to hundreds of kbps. These so called long reads are particularly promising, and are especially expected to solve various problems such as contig and haplotype assembly or scaffolding, for instance. However, these reads are also much more error prone than second generation reads, and display error rates reaching 10 to 30%, according to the sequencing technology and to the version of the chemistry. Moreover, these errors are mainly composed of insertions and deletions, whereas most errors are substitutions in Illumina reads. As a result, long reads require efficient error correction, and a plethora of error correction tools, directly targeted at these reads, were developed in the past nine years. These methods can adopt a hybrid approach, using complementary short reads to perform correction, or a self-correction approach, only making use of the information contained in the long reads sequences. Both these approaches make use of various strategies such as multiple sequence alignment, de Bruijn graphs, hidden Markov models, or even combine different strategies. In this paper, we describe a complete survey of long-read error correction, reviewing all the different methodologies and tools existing up to date, for both hybrid and self-correction. Moreover, the long reads characteristics, such as sequencing depth, length, error rate, or even sequencing technology, can have an impact on how well a given tool or strategy performs, and can thus drastically reduce the correction quality. We thus also present an in-depth benchmark of available long-read error correction tools, on a wide variety of datasets, composed of both simulated and real data, with various error rates, coverages, and read lengths, ranging from small bacterial to large mammal genomes.

1 Introduction

Since their inception in 2011 and 2014, third generation sequencing technologies Pacific Biosciences (PacBio) and Oxford Nanopore Technologies (ONT) became widely used and allowed the sequencing of massive amount of data. These technologies distinguish themselves from second generation sequencing technologies, such as Illumina, by the fact that they allow to produce much longer reads, reaching length of tens of kbps on average, and up to 1 million bps [29]. Thanks to their length, these so called long reads are expected to solve various problems, such as contig and haplotype assembly of large and complex organisms, scaffolding, or even structural variant calling, for instance. These reads are however extremely noisy, and display error rates of 10 to 30%, while second generation short reads usually reach error rates of around 1%. Moreover, long reads error are mainly composed of insertions and deletions, whereas short reads mainly contain substitutions. As a result, in addition to a higher error rate, the error profiles of the long reads are also much more complex than the error profiles of the short reads. In addition, ONT reads also suffer from bias in homopolymer regions, and thus tend to contain systematic errors in such regions, when they reach more than 6 bps. As a consequence, error correction is often used as a first step in projects dealing with long reads. Since the error profiles and error rates of the long reads are much different than those of the short reads, this necessity led to new algorithmic developments, specifically targeted at these long reads.

Two major ways of approaching long-read correction were thus developed. The first one, hybrid correction, makes use of additional short reads data to perform correction. The second one, self-correction, on the contrary, attempts to correct long reads solely based on the information contained in their sequences. Both these approaches rely on various strategies, such as multiple sequence alignment, de Bruijn graphs, or hidden Markov models, for instance. As a result, a plethora of long-read correction methods were developed since 2012, and 29 different tools are available as of today.

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1.1 Contribution

In this paper, we propose a complete survey of long-read error correction tools. In particular, we dress a summary of every single approach described in the literature, both for hybrid correction and for self-correction. In addition, we also dress a list of all the available methods, and briefly describe the strategy they rely on. We thus propose the most complete survey on long-read correction up to date.

Additionally, long reads characteristics, such as the sequencing depth, the length, the error rate, and the sequencing technology, can impact how well a given tool or strategy performs. As a result, a given tool performing the best on a given dataset does not mean that this same tool will perform the best on other datasets, especially if their characteristics fluctuate from one another. As a result, we also present an in-depth benchmark of available long-read correction tools, on a total of 20 different datasets, displaying diverse characteristics. In particular, we assess both simulated and real data, and rely on datasets having varying read lengths, error rates, and sequencing depths, ranging from smaller bacterial to large mammal genomes.

2 State-of-the-art

As mentioned in Section 1, the literature describes two main approaches to tackle long-read error correction. On the one hand, hybrid correction makes use of complementary, high quality, short reads to perform correction. On the other hand, self-correction attempts to correct the long reads solely using the information contained in their sequences.

One of the major interests of hybrid correction is that error correction is mainly guided by the short reads data. As a result, the sequencing depth of the long reads has no impact on this strategy whatsoever. As a result, datasets composed of a very low coverage of long reads can still be efficiently corrected using a hybrid approach, as long as the sequencing depth of the short reads remains sufficient, *i.e.* around 50x.

Contrariwise, self-correction is purely based on the information contained in the long reads. As a result, deeper long reads coverages are usually required, and self-correction can thus prove to be inefficient when dealing with datasets displaying low coverages. The required sequencing depth to allow for an efficient self-correction is however reasonable, as it has been shown that from a coverage of 30x, self-correction methods are able to provide satisfying results [62].

In this section, we present the state-of-the-art of available long-read error correction methods. More particularly, we describe the various methodologies adopted by the different tools, and list the tools relying on each methodology, both for hybrid and self-correction. Details about performances, both in terms of resource consumption and quality of the results, are however not discussed here. Experimental results of a subset of available correction tools, on various datasets displaying diverse characteristics, are presented in Section 3. A summary of available hybrid correction tools is given Table 1. A summary of available self-correction tools is given Table 2.

2.1 Hybrid correction

Hybrid correction was the fist approach to be described in the literature. This strategy is based on a set of long reads and a set of short reads, both sequenced for the same individual. It aims to use the high quality information contained in the long reads to enhance the quality of the long reads. As first long read sequencing experiments displayed high error rates ($> 15\%$ on average), most methods relied on this additional use of short reads data. Four different hybrid correction approaches thus exist:

1. Alignment of short reads to the long reads. Once the short reads are aligned, the long reads can indeed be corrected by computing a consensus sequence from the subset of short reads associated to each long read. PBcR / PacBioToCA [35], LSC [4], Proovread [25], Nanocorr [23], LSCplus [27], CoLoRMap [26], and HECIL [15] are all based on this approach.

2. Alignment of long reads and contigs obtained from short reads assembly. In the same fashion, long reads can also be corrected with the help of the contig they align to, by computing consensus sequences from these contigs. ECTools [43], HALC [6], and MiRCA [30] adopt this methodology.

3. Use of de Bruijn graphs, built from the short reads' k-mers. Once built, the long reads can indeed be anchored to the graph. It can then be traversed, in order to find paths allowing to link together anchored regions of the long reads, and thus correct unanchored regions. LoRDEC [59], Jabba [52], FMLRC [67], and ParLECH [16] rely on this strategy.

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4. Use of Hidden Markov Models. These can indeed be used in order to represent the long reads. The models can then be trained with the help of short reads, in order to extract consensus sequences, representing the corrected long reads. Hercules [21] is based on this approach.

Other methods, such as NaS [49] and HG-CoLoR [53], combine different of the aforementioned approaches in order to counterbalance their advantages and their drawbacks. We describe each approach more into details, and summarize the core algorithm of each tool, in the following subsections.

2.1.1 Short reads alignment

This approach was the first long-read error correction approach described in the literature. It consists of two distinct steps. First, the short reads are aligned to the long reads. This step allows to cover each long reads with a subset of related short reads. This subset can then be used to compute a high quality consensus sequence, which can, in turn, be used as the correction of the original long read. The different methods adopting this approach mainly vary by the alignment methods they use, and also by algorithmic choices made during the consensus sequences computation.

PBcR / PacBioToCA (2012)

PBcR can compute alignment with the help of the aligner developed and included in the tool, with BLASR [11], or with Bowtie [40]. In order to reduce computational times, and avoid unsignificant alignments, alignments are only computed between short reads and long reads that share a sufficient number of k-mers. Moreover, only the short reads that align throughout their whole length are considered for the correction of the long reads they're associated to. Each short read can thus be aligned to multiple long reads. However, on a same, given long read, a short read is only considered once, for the correction of the region to which it aligns with the highest identity.

Moreover, to avoid covering repeated regions of different long reads with the same subset of short reads, the short reads are associated to the repeats they are more likely to belong to, according to the identity of the alignments. Each short read is thus only authorized to participate to the correction of the N long reads to which it aligns with the highest identity, where N roughly represents the sequencing depth of the long reads. Thus, the different repeats are only covered by the short reads representing them the best.

Finally, a consensus sequence is generated from the subset of short reads covering each long read, with the help of the consensus module included in the assembler AMOS. [58]. This module computes the consensus sequence in multiple steps, in a iterative way. At each step, all the short reads are aligned to the current consensus (being the long read itself during the first step), in order to generate a multiple sequence alignment (MSA). This MSA is then used to compute a new consensus sequence, with a majority vote at each position. Iterations stop when the new consensus sequence is identical to the previous step's consensus sequence.

PBcR thus produces split corrected long reads. Indeed, extremities of the long reads that are not covered by alignments are not included in the consensus sequence computation, and are thus not reported in the corrected versions of the long reads. Moreover, if gaps of coverage are spotted along a long read sequence, the corrected version of this read is split, and each covered region is reported independently, in order to avoid introducing incorrect bases in the corrected long reads.

LSC (2012)

LSC starts by compressing homopolymers regions, both in the short reads and in the long reads. To this aim, homopolymers are replaced by a single occurrence of the repeated nucleotide. For instance, the sequence `CTTAGGA` is compressed as `CTAGA`. This compression step is used to facilitate the alignments computation. Moreover, in order to be able to perform the symmetric operation, and decompress the reads, an index is also computed. This index is composed of two arrays for each compressed read, associating the position of the homopolymers to the length of these homopolymers in the original read. Once the reads are compressed, the long reads are concatenated, in order to obtain larger sequences. During this concatenation step, consecutive reads are separated by a block of n nucleotides N, where n represents the average length of the sort reads.

Short reads are then aligned to these new sequences, obtained after concatenation. By default, alignments are computed with Novoalign[1], although LSC is also compatible with other, less resource-consuming aligners,

[1] http://www.novocraft.com/products/novoalign

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such as BWA [47] or SeqAlto [55]. After this alignment step, a subset of short reads if associated to each long read.

Each long is then corrected by computing local consensus sequences from the short reads, on regions where alignments define substitutions, deletions or insertions between the long read and the short reads. Moreover, compressed homopolymers regions, whether they are located on the long read or on the short reads, are also locally corrected in the same fashion. For each aforementioned region, the sequences of all the short reads covering the region are temporarily decompressed. A local consensus sequenced is thus computed with a majority vote, in order to correct this region in the long read. The bases of the long read that do not correspond to any of these regions are thus left unmodified by the correction step. Once all the regions requiring correction have been processed, compressed homopolymers regions of the long reads, unmodified during the correction step, are decompressed and left unmodified in the sequences of the corrected long reads.

LSC thus produces trimmed corrected long reads, defined, for each read, as the sequence of the uncompressed long read, from the leftmost base to the rightmost base, covered by at least one short read. However, bases of the long reads that could not be corrected are not reported in a different case from the corrected bases. As a result, the corrected long reads cannot further be split after correction.

Proovread (2014)

Proovread computes alignments with the help of SHRiMP2 [17]. Although this tool/ is used as the default aligner, all the aligners reporting their results in SAM format, like BWA, Bowtie or Bowtie2 [39] are compatible with Proovread and can easily be used. A validation step is then applied, in order to filter out low quality alignments. To this aim, Proovread uses scores that are normalized according to the length of the alignments, in a local context. For this purpose, the long reads are represented as a series of consecutive, small windows of fixed length L ($L = 20$ by default). Each alignment is thus assigned to one of these windows, according to the position of its center.

Once the alignments have been allocated to the different windows, only the alignments having the highest scores in each window are considered for the consensus sequence computation, during the next step. To compute this consensus sequence, a matrix composed of a column per nucleotide used used for each long reads. This matrix is filled with the bases from the short reads, according to the information contained in the alignments selected during the previous step. The consensus sequence is then computed with a majority vote on each column of the matrix. If no base coming from a short read is available for a given column, the base of the original long read is conserved. Moreover, a score mimicking the Phred quality score is assigned to each base of the corrected long read. This score is defined from the support of the base that was chosen as the consensus, in the corresponding column of the matrix.

In order to reduce time and memory consumption of the alignment step, which can be extremely costly when the required level of sensitivity is high, Proovread proposes an iterative procedure for alignment and correction, where the alignment sensitivity is increased at each step. This strategy is composed of three cycles of pre-correction and of a finishing step. Thus, during the pre-correction cycles, only a subset of 20, 30, and 50% of the original set of reads is used, respectively for the first, second, and third cycle. During the first cycle, the short reads are quickly aligned, with a moderate level of sensitivity. Long reads are then pre-corrected with the help of the aligned short reads, as described in the previous paragraph. Moreover, regions of the long reads that are sufficiently covered by the short reads are masked, and are thus not further considered in the following alignment and correction cycles. Two additional pre-correction cycles are then applied, using larger subsets of the short reads, and increasing levels of sensitivity. On average, the first two cycles are enough to mask more than 80% of the long reads sequencing. Finally, for the finishing step, the mask are removed from the pre-corrected long reads, and the complet set of short reads is aligned with a high sensitivity, in order to perform a last correction step, and thus polish previous pre-corrections.

Proovread thus produces corrected long reads both in their trimmed version and in their split version. The splitting step is indeed not dependent on the correction step, and relies on the pseudo-Phred quality scores assigned to each bases of the corrected long reads. Low quality regions of the corrected long reads can thus easily be identified and removes, in order to only retain high quality regions.

Nanocorr (2015)

Nanocorr computes alignments with the help of BLAST [2]. A set of alignments, composed of both correct alignments, spanning almost through the whole length of the short reads, and incorrect of partial alignments of the short reads is thus obtained. A first filtering step is then applied to these alignments, in order to remove alignment that are too short, and fully included in another, larger alignment. A dynamic programming algorithm, based on the longest increasing subsequences problem [61] is then applied, in order to select the

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optimal subset of short reads alignments covering each long read.

From this subset, a consensus sequence is computed, with the help of a DAG, using the consensus module PBDAGCon, described Section 2.2.1.

Nanocorr thus produces untrimmed corrected long reads. Moreover, bases of the long reads that could not be corrected are not reported in a different case from the corrected bases, for the reasons mentioned in the description of PBDAGCon, Section 2.2.1. As a result, the corrected long reads cannot further be trimmed or split after correction.

LSCplus (2016)

LSCplus is based on the same principle as LSC, which we previously presented. It however brings a few modifications in terms of implementation and optimization during the alignment and correction steps. First, as in LSC, the homopolymers regions, both in the short reads and in the long reads, are compressed and replaced by a single occurrence of the repeated nucleotide. The index allowing to retrieve the original size of the homopolymers is however altered compared to that of LSC. Indeed, in LSCplus, this index is composed of a single array for compressed each read, registering, for each position (whether it compressed or not), the number of bases in the original read. Moreover, unlike LSC, the long reads are not concatenated in larger sequences before performing the alignment step. The short reads are thus directly aligned to the long reads, with the help of Bowtie2.

Each long read can then be corrected, using the subset of short reads associated to it. A consensus sequence is thus computed, with a majority vote, at each position of the compressed long read. Unlike LSC, which decompresses regions requiring correction before computing the consensus, the most frequent nucleotide at each position is determined before decompression, in LSCplus. One the most frequent nucleotide has been chosen, LSCplus performs decompression, and the size of the homopolymer is determined from the information contained in the indexes of the short reads that align at this position. The sequence thus obtained is concatenated at the end of the global consensus sequence, which is iteratively built, as the compressed long read bases are processed. Bases corresponding to uncovered positions of the long reads are decompressed without further modifications, and concatenated at the end of the global consensus sequence as well.

Unlike LSC, LSCplus thus produces corrected long reads both in their trimmed and in their untrimmed versions. As for LSC, the sequence of a trimmed long read is defined as the sequence of the uncompressed long read, from the leftmost base to the rightmost base, covered by at least one short read. Once again, as for LSC, bases of the long reads that could not be corrected are not reported in a different case from the corrected bases. As a result, the corrected long reads cannot further be split after correction.

CoLoRMap (2016)

CoLoRMap computes alignments with the help of BWA-MEM [44]. An alignment graph is then built for each long read, from the subset of short reads that align to it. This graph is directed and weighted, and each vertex is defined from the alignment of a short read to the long read. An edge is defined between two vertices u and v if the short reads associated to these two vertices share a prefix / suffix overlap, over a sufficient length, and with at most one error. At least, the weight of an edge between two vertices u and v is defined as the edit distance between the suffix of the read associated to v that is not included in the overlap, and the original long read.

Each connected component of this graph is then processed independently. For each of these connected components, a *source* vertex, corresponding to the leftmost aligned short read of the component, and a *target* vertex, corresponding to the rightmost aligned short read, are defined. Dijkstra's algorithm [18] is then used, in order to find the shortest path of the graph allowing to link the source and the target. This path, through the short reads associated to the traversed vertices, dictates a correction for the region of the long read which is covered by the alignments contained in this connected component.

To correct long reads regions that were not covered by any short read during the alignment step, CoLoRMap makes use of the information carried by the paired reads than can be sequenced from second generation sequencing technologies. *One-End Anchors (OEA)* are thus defined, in order to correct these uncovered regions. These OEA are short reads than could not be aligned, but whose mate could be aligned to a region flanking the uncovered region to correct. The set of OEA associated to a given uncovered region of a long read can thus be defined be observing the alignments located in the flanking, corrected regions.

For this second correction step, short reads are aligned to the corrected long reads, obtained at the end of the previous step, once again with the help of BWA. The short reads can indeed easily be aligned to the corrected regions of the long reads, due to the low divergence between their sequences and the corrected regions of the long reads. For each region of a given long read that could not be corrected during the previous step, CoLoRMap

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defines a set of OEA, by observing the alignments located in the flanking regions. The reads composing this set of OEA are then assembled, with the help of Minia [12], in order to obtain a set of contigs, which is then, in turn, used to correct the uncovered region.

CoLoRMap thus produces untrimmed corrected long reads. However, the bases of the long reads that could not be corrected are not reported in a different case from the corrected bases. Thus, the corrected long reads can easily be trimmed or split after correction, in order to get rid of their uncorrected regions.

HECIL (2018)

HECIL computes alignments with the help of BWA-MEM. From these alignments, erroneous positions, denoting disagreements (insertions, deletions or substitutions) between a given long read and the short reads that align to it are marked. For each of this positions, the set of aligned short reads is processed, and and two correction steps are applied.

First, in case of strong consensus (*i.e.* at least 90%) of the short reads at a given position, the nucleotide of the long read is replaced by the consensus nucleotide from the short reads. This first step, inspired by other methods relying on majority votes, allows for a quick first correction step. Moreover, it also allows to avoid spurious correction at certain positions, performed from high-frequency, but low-quality short reads. The corrections that are realized during this first step also allow to greatly reduce the number of positions to process during the next step.

In a second time, for the remaining erroneous positions, a score is associated to each aligned short read, by combining the quality of the short read, obtained through its Phred score, to the similarity score of the alignment between the short read and the long read. The main motivation behind this combined score is that the incorporation of short reads quality into the problem of long-read correction had not been tackled by any state-of-the-art method at that time. For each erroneous position, the base from the short read which minimizes the combined score is chosen as then correction of the long read's base. In case of a tie between multiple short reads, the base from the short read having the highest quality is chosen as the correction.

Moreover, HECIL also defines an iterative learning methodology. More precisely, the two previously described correction steps are applied iteratively. At each iteration, the previously defined combined scores are used in order to define a subset of high-confidence positions to correct. Only the positions contained in this subset are thus corrected during this given iteration. These positions are then fixed and marked, and are thus no longer modified during subsequent iterations. By only correcting the best subset of position for each iteration, HECIL aims to perform better correction during the following iterations, since these iterations will benefit from the higher quality of the long reads, increased by the previous iterations. Thus, at each iteration, the short reads are realigned to the updated long reads, and the two correction steps are performed on a new subset of high-confidence positions. The iterations are stopped when the number of unique k-mers of the long reads varies less than a given threshold between two consecutive iterations, or once a user-defined maximum number of iterations is reached.

HECIL thus produces untrimmed corrected long reads. Indeed, local corrections are performed on the long reads, on positions denoting disagreements with the aligned short reads. Thus, the uncovered extremities of the long reads cannot be modified or corrected. Moreover, the covered bases of the long read that could not be corrected are not reported in a different case from the corrected bases. As a result, the corrected long reads cannot further be split after correction.

2.1.2 Contigs alignement

Given their length, short reads can be difficult to align to repeated regions, or to extremely noisy regions of the long reads. This approach aims to address this issue by first assembling the short reads. Indeed, the contigs obtained after assembling the short reads are much longer than the original short reads. As a result, they can cover the repeated of highly erroneous regions of the long reads much more efficiently, by using the context of the adjacent regions during alignment. In the same fashion as the short reads alignment strategy described in Section 2.1.1, the contigs aligned with the long reads can then be used to compute high quality consensus sequences, and thus correct the long reads they're associated to. Once again, the different methods adopting this strategy vary by the alignment methods they use, and by algorithmic choices made during consensus computation.

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ECTools (2014)

ECTools requires a set of short reads assembled into unitigs as a basis for its error correction process. This means that the short reads assembly has to be performed before launching ECTools, and that it is thus not automatically generated during the error correction process. The assembly can thus be produced by any tool, although the authors recommend using Celera [56], as it guarantees that each short reads is included in a unitig, by creating unitigs composed of a single short read, if necessary. This allows ECTools to ensure all the information contained in the short reads will be transmitted to the set of unitigs, and that no information will be lost during the assembly process.

The unitings are aligned to the long reads with the help of Nucmer, which is available in the MUMmer [38] suite of tools. The subset of unitigs that covers the best each long read is then chosen and used for its correction. This optimal subset is computed with the help of an algorithm based on the longest increasing subsequences problem, which is also available in the MUMmer suite of tools. Once computed, this subset of unitigs is used to correct the long read. The bases of the long read which are different from those of the selected unitigs are identified, and the long read is corrected according to the bases of the unitigs.

ECTools thus produces split corrected long reads. Uncovered extremities of the long reads are first removed from the sequences of the corrected long reads. Then, internal, uncovered regions, are considered as uncorrected, and are labeled with an error rate of 15%. In the same fashion, covered regions are considered as corrected, and are labeled with an error rate of 1%. The average error rate of each corrected long read is then computed on its whole length, according to these labels, and is then compared to a minimal quality threshold. If the average quality of a given long read is below that threshold, the read is then split, by removing the longest uncorrected region from its sequence. This process is then recursively applied to each obtained fragment, until they are all above the minimum quality threshold.

HALC (2017)

In the same way as ECTools, HALC also requires a set of assembled short reads as a basis for its error correction process. Once again, assembly of the short reads thus has to be performed before launching HALC, and is not automatically generated during the error correction process. Unlike ECTools, HALC however makes use of the contigs generated by the assembly, instead of only using the unitigs. The long reads are thus aligned to the contigs, with the help of BLASR [11], to initiate the error correction process.

An undirected graph is then built from the alignments between the long reads and the short reads. This allows to represent the alternative alignments, in particular in repeated regions, by different paths. Vertices of the graph are thus defined from the regions of the contigs to which a long read aligned. An edge is present between two vertices if at least a pair of adjacent regions of a given long read aligned to the two regions of the contigs corresponding to theses vertices.

This graph is also weighted, and the weight of each edge is computed according to two rules. First, the weight of an edge (u, v) is set to 0 if the regions corresponding to vertices u and v are adjacent in one of the contigs. Contrariwise, for a given edge (u, v) for which the regions corresponding to vertices u and v are not adjacent in any of the contigs, the weight is set to $max\{C_0 - C, 0\}$. C_0 represents the long read coverage on the contigs, whereas C represents the number of adjacent regions of the long reads that align to the regions corresponding to vertices u and v.

Once the graph is built, each long read is processed and corrected independently. To this aim, paths representing alternative alignments are search through the graph. The path displaying the lowest cumulative weight is then computed via a dynamic programming algorithm, and the regions of the contigs corresponding to the traversed vertices are used to correct the long read.

However, with this approach, adjacent regions of the long reads that are corrected with the help of non-adjacent contigs regions have a high chance of being repeated regions. Thus, they also have a high chance of having been corrected with similar repeats whether than with the true genome regions they actually belong to. Such regions are recorded, and their correction is then polished with the help of LoRDEC, another hybrid correction tool presented in Section 2.1.3.

HALC thus produces untrimmed corrected long reads. However, the last polishing step, being based on LoRDEC, allows to report the corrected bases in a different case from the uncorrected bases. As a result, the corrected long reads can easily be trimmed or split after correction, in order to get rid of their uncorrected regions. In particular, untrimmed, trimmed, and split corrected long reads are all reported by HALC.

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MiRCA (2018)

Unlike previously presented methods, MiRCA starts by applying a filtering step to the short reads. Indeed, although they display a much higher quality than the long reads, short reads also contain sequencing errors that can lead to imprecisions during the assembly step and during the alignment step of the long reads to the obtained contigs. Thus, in MiRCA, short reads containing too many errors are removed, and are not considered during the following steps. This filtering step relies on a frequency analysis of the short reads' k-mers. Short reads containing too many weak k-mers (*i.e.* k-mers appearing less frequently than a given threshold) are thus filtered out.

High-quality short reads are then assembled with the help of SPAdes [5], an assembler based on a de Brujn graph, that allows to ensure that all the information contained in the short reads is preserved in the obtained set of contigs. These contigs are then aligned to the long reads with the help of BLAST+ [48].

The long reads can then be corrected from the obtained alignments. In the case a unique contig aligns to a given region of a long read, this region is simply replaced by the sequence of the aligned contig. In the case where multiple contigs are aligned to a same given region of a long reads, these alignments define a multiple sequence alignment, and a majority vote is used to correct the region of the long read. If the majority vote does not allow to define a consensus nucleotide at a given position, the long read nucleotide at this position is kept as the correction.

MiRCA thus produces trimmed corrected long reads. Indeed, extremities of the long reads to each no contig aligned cannot be processed by the algorithm, and are thus removed from the sequences of the corrected long reads. However, the internal regions of the long reads that were not covered by a single contig, and that could not be corrected, are not reported in a different case from the corrected bases. As a result, corrected long reads cannot further be split after correction.

2.1.3 De Bruijn graphs

Another alternative to the alignment of short reads to the long reads is the direct use of a de Bruijn graph, built from the short reads k-mers. This approach aims to avoid the explicit step of short reads assembly altogether, contrary to methods mentioned in Section 2.1.2, and instead directly use the graph to correct the long reads. The graph is first built from the k-mers of the short reads. The long reads can then be anchored to the graph according to their k-mers. Finally, the graph can be traversed, in order to find paths, and link anchored regions of the long reads together, and thus correct erroneous, unanchored, regions. Methods adopting this approach vary by the way they represent the graph, but also by the way they anchor the long reads to the graph, and by the way they correct unanchored regions.

LoRDEC (2014)

LoRDEC starts by building a de Bruijn graph from the solid k-mers (*i.e.* k-mers that appear more frequently than a given threshold) of the short reads. This filtering step of the weak k-mers allow the avoid the introduction of errors during the correction of the long reads, as it limits the presence of tips and bubbles in the graph. The solid k-mers of the long reads are then used as anchor points on the graph. The graph is then traversed, from and towards the vertices corresponding to these anchors, in order to find paths allowing to corrected regions of the long reads that are only composed of weak k-mers. Thus, if a given long reads does not contained any solid k-mer, it cannot be corrected.

Given the high error rates of the long reads, a small k-mer size has to be chosen for the graph construction, in order to allow the correction of a as much long reads as possible. Thus, it is recommended to build the graph for values of k between 19 and 23. Two different correction approaches are then applied, according to the localization of the region to correct. Internal regions and extremities of the long reads are indeed corrected differently.

In the first case, solid k-mers located in the flanking regions of the region to correct are used as anchor points on the graph. The solid k-mers located on the left of the region to corrected define *sources*, whereas the solid k-mers located on the left define *targets*. The graph is thus traversed, in order to find a path between a source and a target k-mer. Once found, such a path dictates a correction for the erroneous region of the long read. In order to find an optimal path, at each branching path, the edge minimizing the edit distance to the erroneous region of the long read is chosen. However, a limit on the maximal number of branching paths that can be explored is set, in order to avoid extensive and costly traversals of the graph. Several pairs of source and target

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k-mers are thus considered, in order to limit the cases where no path can be found to correct a weak k-mers region. As a result, the graph is traversed multiple times for the correction of a given region. Several of these traversals can thus find a path between two anchor k-mers, and dictate a correction for the erroneous region of the long read. In this case, the path dictating the sequence minimizing the edit distance to the erroneous region of the long read is chosen as the correction.

In the second case, one of the source of target k-mer is missing, and the search thus cannot be stopped when a path between two k-mers is found. In this case, the graph is only traversed from a solid k-mer close to the erroneous region. The traversal of the graph stops when the length of the erroneous extremity of the long read is reached, when the edit distance between the current path and the erroneous region is too high, or when to edge can be followed out of the current vertex. Once the graph traversal is over, the obtained sequence is realigned to the extremity of the long read. This allow to to determine the prefix (resp. the suffix) of the graph sequence that aligns the best to the extremity of the long read. The aligned prefix (resp. suffix) of the long read extremity is thus corrected with the prefix (resp. suffix) of the graph sequence. This alignment step allows to ensure divergent sequences, caused by inappropriate traversals of the graph, are no used as correction.

Two correction passes are thus applied for each long read, once from the left to the right, and once from the right to the left. Indeed, the long reads are corrected on the fly by the graph traversals. These corrections thus allow to generate new solid k-mers that can be used as anchor points by the second correction pass. Moreover, repeats and sequencing errors that are present in the short reads can lead to the traversal of different subgraphs, according to the extremity that is chosen as the starting point of the path search.

LoRDEC thus produces untrimmed corrected long reads. However, bases corresponding to solid k-mers are reported in a different case from bases corresponding to weak k-mers. As a result, the corrected long reads can easily be trimmed or split after correction, in order to get rid of their uncorrected regions. Two additional tools allowing to perform these operations are provided along with LoRDEC.

Jabba (2016)

Unlike previously presented methods, Jabba starts by correcting the short reads, in order to limit the impact of their sequencing errors on the quality of the de Bruijn graph, and thus on the quality of the correction. To this aim, the authors recommend using Karect [1], although any correction tool can be used. The de Bruijn graph is then built from the k-mers of the corrected short reads. To further reduce the impact of the errors contained in the graph on the correction of the long reads, another correction procedure is applied to the graph itself. More precisely, the tips are eliminated, and the bubbles are removed from the graph, by selecting, for each of them, the highest supported path. The construction and correction of the graph is performed with the help of Brownie[2].

Moreover, unlike LoRDEC, Jabba does not rely on the study of k-mers that appear both in the graph and in the long reads to find anchor points, but rather uses Maximal Exact Matches (MEM) between the long reads and the vertices of the graph. Thereby, since the graph also displays a high quality, due to the two correction processes, large values of k can be used for its construction. Usually, the graph is built with $k = 75$. Using such values of k allows to reduce the complexity of the graph, by resolving short repeats, of size smaller than k, directly inside the graph, which, in turns, allows to facilitate the alignment of the long reads to the graph, during the following step.

A seed-and-extend strategy is applied to align the long reads to the graph. First, the MEMs, which represent the seeds, are computed with the help of essaMEM [66]. This method is based on an enhanced suffix array, built for the sequence obtained by concatenating all the vertices of the graph, as well as their reverse-complements. Once the seeds of a given long read are computed, they are placed of the graph. To this aim, several passes are performed on the long read. For each iteration, all the regions of the long reads that have not yet been aligned are considered. For each of these regions, the longest seeds are used, in order to determine the vertices to which this region can be aligned. The quality of the alignments between each vertex and the long read region are then verified. Alignments that are fully included in longer alignments (called *containments*), and alignments covering a fraction of the vertex that is below a given threshold are then filtered, and are thus not considered during the following step.

Once the alignments between the regions of the long read and the vertices of the graph are computed for the current iteration, the alignments between the vertices are chained, by following the paths of the graph. Each alignment on a vertex is thus extended, by considering all the possible paths of the graph starting from this vertex. If the vertex only has one outgoing edge, the alignment is extended along that edge. If multiple outgoing edges are available from this vertex, the length of the target vertex of each edge is studied. The extension taking place between two regions of the long read, a maximal distance between the two edges can be defined, and edges that are too long are thus not considered. In the case where no outgoing edge can be followed from a given

[2]https://github.com/biointec/brownie

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vertex, the corresponding region of the long read reprocessed and realigned during the next iteration.

Once all the iterations have been performed, several alignments usually remain possible for each long reads. The alignment covering the largest proportion of the long read sequence is thus selected, and used for the correction. For the correction of the extremities of the long read, the alignment is extended along unique paths of the graph. If the long read extends outside of these unique paths, its extremities are trimmed. Indeed, correcting these extremities is extremely time consuming, since they need to be aligned to all the possible paths, as no seed can be used to guide the alignment. This problem is thus not tackled by Jabba, as the gain in quality if to weak compared to the computational cost.

Jabba thus produces split corrected long reads. Indeed, as mentioned in the previous paragraph, extremities of the reads are not corrected outside of the longest unique path of the graph to which they align. Moreover, in cases where, even after multiple iterations, no edge can be followed to extend the alignment of a given vertex, a direct path between the first and the last seed of the long read does not exist. In this case, Jabba splits the long read, and independently corrects and outputs the different regions of this read that could be aligned and chained along the graph.

FMLRC (2018)

FMLRC relies on a principle that is similar to that of LoRDEC. Indeed, it also builds a de Bruijn graph from the k-mers of the shorts reads, and uses the solid k-mers of the long reads as anchor points on this graph, in order to find paths allowing to correct the regions of the long reads only composed of weak k-mers. One of the main differences between FMLRC and LoRDEC is that the de Bruijn graph is implicitly represented in FMLRC, with the help of an FM-index built from the sequence obtained by concatenating the set of short reads. As a result, the edges between the different vertices are found by querying the FM-index. The size ok k thus does not have to be known at construction time, but is rather directly chosen at execution time. This way, the FM-index allows to represent all the de Bruijn graphs, and not simply a single, fixed order, graph. Such a graph is called a *variable-order* de Bruijn graph. However, unlike LoRDEC, the weak k-mers of the long reads are not filtered, and all the k-mers of the short reads are thus present in the graph.

The correction of the long reads is then realized in the same way as LoRDEC. For regions composed of weak k-mers bordered by two regions composed of solid k-mers, source and target k-mers are defined on each bordered regions, and used as anchor points on the graph. A path is then searched through the graph, from the source toward the target, and from the target toward the source. Indeed, the search can traverse different subgraphs, according to its starting point. As in LoRDEC, if multiple paths are found, the one minimizing the edit distance to the erroneous region of the long read is chosen as the correction. For erroneous regions located at the extremities of the long reads, the solid k-mers that is the closest to the erroneous region is chosen as the anchor point. The graph is thus traversed, and the traversal stops when the length of the path reaches the length of the erroneous region to correct. Several paths can thus be obtained, and the one minimizing the edit distance to the erroneous region of the long read is, once again, chosen as the correction.

However, unlike LoRDEC, FMLRC performs two passes of correction for each long read. A first pass is performed with a small value of k ($k = 20$ by default), and a second pass in then performed, with a larger value of k ($k = 59$ by default). The first pass usually manages to correct most errors, and the second pass allows to polish the correction, especially in repeated regions, which are easier to resolve with the help of a higher order de Bruijn graph. Moreover, unlike LoRDEC, the solidity threshold of the long reads' k-mers is dynamically adjusted for each long read, according to the chosen k-mer size and to the k-mers frequencies of this long read. The limit on the maximal number of branching paths that can be explored is also set according to the k-mer size, in order to reduce the number of branches exploration during the first pass. Indeed, the traversal of the first graph can be extremely costly, due to the important number of branching paths in repeated regions, when using a small k-mer size. These regions are rather resolved during the second correction pass, where a greater number of branches exploration is allowed, and where the k-mer size is higher.

FMLR thus produces untrimmed corrected long read. However, unlike LoRDEC, bases corresponding to solid k-mers are not reported in a different case from bases corresponding to weak k-mers. As a result, the corrected long reads cannot further be trimmed or split after correction.

ParLECH (2019)

Unlike other DBG based methods, ParLECH relies on two distinct correction steps, depending on the type of errors. A first step is thus dedicated to the correction of indels, and another step focuses on the correction of substitutions, which can be introduced by the short reads during the correction of indels.

For the correction of indels, ParLECH first builds the de Bruijn graph from the k-mers of the short reads.

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The graph is stored in a key-value hashmap, with the help of HazelCast [3], where the k-mers define the keys, and where predecessors and successors of these k-mers define the values. Moreover, a number of occurrences is associated to each key. In the same fashion as in previously presented methods, regions solely composed of weak k-mers are corrected by traversing the graph, using flanking, solid k-mers as anchor points. If multiple paths allow to link together a given pair of anchors, the path maximizing the minimum k-mer coverage between two vertices is chosen as the correction.

For the correction of substitutions, ParLECH first divides the long reads into shorter fragments, approximatively of the size of the short reads. Indeed, k-mers in smaller subregions usually have similar abundances. This allows ParLECH to divide the long reads into sequences of high and low coverage fragments. If a fragment belongs to a low coverage region of the genome, most of its k-mers are expected to have a low coverage. On the contrary, most of its k-mers are expected to have a high coverage. This methodology allows ParLECH to better distinguish low coverage genomic k-mers, and high coverage erroneous k-mers. Once the long reads have been fragmented, the Person's skew coefficient of the k-mer coverage of each fragment is computed, and used as a threshold to classify fragments as correct or erroneous. If the coefficient of a fragment falls within a given interval, the fragment is considered as correct. Otherwise, it is considered as erroneous. Moreover, fragments with mostly low-coverage k-mers are ignored.

After classification, the erroneous fragments are divided into subsets of high and low coverage. To this aim, if the median k-mer coverage of a fragment is greater than the median coverage of the whole set of k-mers, the fragment is considered as high coverage. Otherwise, it is considered as low coverage. To correct substitutions, ParLECH makes use of a majority vote algorithm similar to that of Quake [31]. However, unlike Quake, ParLECH makes use of different thresholds for high a low coverage regions, in order to improve accuracy. For each erroneous base detected during the previous step, ParLECH attempts to replace it with all the other possible nucleotides, and computes the coverage of all the k-mers with this new base. The erroneous base is thus replaced by the base such that all k-mers containing this base exceed or equal the threshold for this region.

2.1.4 Hidden Markov models

Hidden Markov models, used for short read error correction, were also adopted for the error correction of long reads. To this aim, models are first initialized in order to represent the original long reads. A subset of short reads is then assigned to each long read, by alignment. Each subset of short reads can then be used to train the model it is associated to. Finally, the trained models can be used to compute consensus sequences, and thus correct the long reads they represent.

Hercules (2018)

Like LSC and LSCplus, Hercules starts by compressing homopolymers regions of the long reads and of the short reads. The compressed short reads are then filtered, and those that are shorter than a given threshold are removed. Such short reads could indeed ambiguously align to the long reads. The remaining short reads are then aligned to the long reads. Bowtie2 is used by default, although any aligner can be used, as long as it outputs alignments in SAM format.

After computing the alignment, Hercules decompresses both the long reads and the short reads, and recomputes the beginning positions of the alignments, according to the uncompressed sequences. However, unlike other methods based on the alignment of short reads to the long reads, Hercules then only uses the beginning positions of the alignments, and does not make use of any other information reported by the aligner. Thus, the choice of the bases used to correct the long reads is not performed by the aligner.

Each long read is then represented with the help of a profile Hidden Markov Model (pHMM). These pHMMs have three different type of states, allowing to represent deletions, insertions, or matches / mismatches. The aim of Hercules is to make use of the pHMM to produce the consensus sequence of the long read, which is defined as the most probable sequence, among all the sequences that can be produced by the model. However, the consensus that can be produced by a classical pHMM is only based on match states. As a result, in order to take into account insertion and deletion states during consensus computation, the pHMM are modified in Hercules. Loops on insertion states are removed, and replaced by multiple, distinct, insertion states. Deletion states are also removed, and replaced by deletion transitions. Once the model of a given long read is built, emission and transmission probabilities of each state are initialized according to the error profile of the sequencing

[3] https://hazelcast.com/

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technology.

The model is then updated according to the information contained in the short reads. To this aim, each alignment of a short read to a long read is considered independently. First, the subgraph corresponding to the region of the long read which is covered by this short read is extracted. Then, this subgraph is trained, with the help of the information contained in the selected short read, with the help of the Forward-Backward algorithm [8]. Emission and transmission probabilities of each vertex are thus updated by the algorithm, independently and exclusively in each subgraph, using the probabilities of the initial model. In the case a vertex is considered and updated in multiple subgraphs, its final probabilities, in the trained model, are defined by computing the average probability of each subgraph it appears in. Uncovered regions of the long reads cannot yield subgraphs. Emission and transmission probabilities of the initial models are thus kept, for the corresponding vertices.

Once all the alignments have been considered, and once all the subgraphs have been trained, the model is updated according to these subgraphs. Viterbi algorithm [64] is then used, in order to decode the consensus sequences of the updated model. This algorithm considers the pHMM associated to each long read, and searches for a path, from the initial state to the final state, which produces the highest emission and transmission probabilities, via a dynamic programming approach.

Hercules thus produces untrimmed corrected long reads. Indeed, the consensus sequence of a long read is extracted from the pHMM by searching for a path from the initial to the final state, representing respectively the first and the last base of this read. The entire length of the long reads is thus preserved after correction. Moreover, bases associated to subgraphs that could be trained are not reported in a different case than the bases to which no short read aligned. As a result, the corrected long reads cannot further be trimmed or split after correction.

2.1.5 Combination of strategies

Other methods combine different of the aforementioned strategies, in order to balance their advantages and drawbacks. For instance, NaS combines a first step of short reads alignment to a second step of short reads recruitment and assembly, in order to correct the long reads. HG-CoLoR also relies on a first step relying on short reads alignment, but then makes use of a variable order de Bruijn graph, in order to correct regions of the long reads that were not covered by the original alignments. We describe these two tools below.

NaS (2015)

Unlike previously presented methods, which either perform local correction on specific long reads regions, or compute consensus sequences from short reads alignments, NaS generates corrected long reads by assembling subsets of related short reads. These sequences, called synthetic long reads, are computed as follows.

First, the alignments between the short reads and the long reads are computed either with the help of BLAT [32] or LAST [33], according to the chosen correction mode (fast or sensitive, respectively). The short reads thus aligned to the long reads define seeds. For each long read, a seed-and-extend strategy is then applied. The seeds are compared to the set of short reads, in order to recruit new similar short reads, that allow to cover the regions of the long reads that were not covered by the initial alignments. This recruitment step is performed with the help of Commet [50], which considers two short reads are similar if they share a sufficient number of non-overlapping k-mers.

After recruiting new short reads, the subset of short reads formed by these new reads and by the seeds is assembler with the help of Newbler [65]. A unique contig is usually built, although in repeated regions, a few short reads originating from the wrong copy of the repeat can be erroneously recruited, and thus yield additional contigs that should not be associated to the original long read. To address this issue, and produce a single contig for each long read, the contig graph is explicitly built. This graph is a directed and weighted graph, for which the contigs defined the vertices. An edge is present between two vertices if the associated contigs overlap. The weights are however associated to the vertices, and not to the edges. Thus, the weight of each vertex is defined as the coverage of the contig by the seed short reads. Once graph is built, Floyd-Warshall algorithm [22, 68] is used in order to extract the shortest path. The final contig is thus obtained by combining the contigs associated to the vertices traversed by this path.

Finally, the short reads are realigned to the obtained contig, in order to verify is consistency. The contig is then defined as the correction of the original long read, if it is sufficiently covered by short reads.

Unlike previous methods, that usually either trim or split the corrected long reads, NaS rather tends to extend the reads, and thus produce corrected long reads that are longer than the original reads they are associated to. This is caused by the fact that the recruitment process often includes short reads that are located outside the

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extremities of the original long read to the subset of short reads used for the assembly. Such a behavior can thus assemble corrected sequences that span outside of the original long reads.

HG-CoLoR (2018)

HG-CoLoR combines together a short read alignment and a de Bruijn graph approach into a seed-and-extend strategy. Unlike other methods, it starts by correcting the short reads, in order to get rid of as much sequencing errors as possible. Indeed, since these short reads will be aligned to the long reads, and will also be used to build the graph, they need to reach a high level of quality to avoid the introduction of errors into the corrected long reads.

HG-CoLoR then aligns the corrected short reads to the long reads with the help of BLASR. A set of seeds is thus defined for each long read, and covered regions of the long reads can directly be corrected with help of these seeds. After the alignment step, a variable order de Bruijn graph is built from the solid k-mers of the corrected short reads. Unlike FMLRC, this graph is built with the help of PgSA [37]. Moreover, HG-CoLoR allows to explore every order of the graph, between a minimum order k and a maximum order K, instead of limiting the graph explorations to two different orders. To correct uncovered regions of the long reads, the prefixes and suffixes of the seeds that aligned to covered regions are used as sources and targets on the graph, which is traversed to perform correction. The graph is always explored starting from its highest order. The order of the graph is thus only locally decreased if no edge can be followed out of the current node, or if all its edges, for the current order, have already been explored and did not allow to link the source and the target. At branching paths, for a given order k, HG-CoLoR performs a greedy selection and explores the edge leading to the k-mer having the highest number of occurrences. Thus, when a path between two anchors is found, it is considered as optimal, due to that greedy selection strategy, and due to the fact that the order of the graph is only locally decreased. As a result, it is thus directly chosen as the correction of the uncovered region of the long read. As in other methods, a maximal number of branches explorations is also set, in order to avoid resource consuming traversals of the graphs. Moreover, if a given pair of seeds cannot be link after reaching that limit, HG-CoLoR can attempt to skip a certain number of seeds, and thus try link together the same source to different targets. A maximal number of skips is also set, and if no source-target pair could be linked, the uncovered region of the long read remains uncorrected.

Finally, as seeds do not always cover the extremities of the long reads, HG-CoLoR keeps on traversing the graph after linking together all the seeds, in order to correct the extremities of the long reads. The graph is thus traversed, until the whole length of the processed long read has been corrected, or until a branching path is reached. If the extremities of the original long read could not be reached, they are reported unmodified in the corrected long read.

HG-CoLoR thus produces untrimmed corrected long reads. However, the corrected bases, whether they come from seeds or from graph traversals, are reported in a different case from the uncorrected bases. As a result, the corrected long reads can easily be trimmed or split after correction, in order to get rid of their uncorrected regions. In particular, untrimmed, trimmed, and split corrected long reads are all reported by HG-CoLoR.

2.2 Self-correction

Self-correction aims to avoid the use of short reads data altogether, and to correct long reads solely based on the information contained in their sequences. Third generation sequencing technologies indeed evolve fast, and now allow the sequencing of long reads reaching error rates of 10-12%. As a result, correction is still required to properly deal with errors, but self-correction has recently undergone important developments. Two different self-correction approaches thus exist:

1. Multiple sequence alignment. This approach is similar to the hybrid approaches relying on short reads or contigs alignments, described in Section 2.1.1 and in Section 2.1.2. Indeed, once aligned against each other, the long reads can be corrected by computing a consensus sequence for each of them, in the same fashion as in the hybrid approaches. PBDAGCon (the correction module used in the HGAP assembler) [13], PBcR-BLASR [34], Sprai[4] [24], PBcR-MHAP [9], FalconSense (the correction module used in the assembler Falcon) [14], Sparc [70], the correction module used in the assembler Canu [36], MECAT [69] and FLAS [7] rely on this approach.

2. Use of de Bruijn graphs. In a similar manner as the hybrid correction approach using de Bruijn graphs, described in Section 2.1.3, once the graph is built, it can be used to anchor the reads. The graph can then

[4]http://zombie.cb.k.u-tokyo.ac.jp/sprai/index.html

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be traversed, in order to find paths allowing to link together anchored regions of the long reads, and thus corrected unanchored, erroneous regions. LoRMA [60] and Daccord [63] are based on this approach.

As for hybrid correction, other methods, such as CONSENT [54], combine these two approaches, in order to counterbalance their advantages and their drawbacks. We describe each approach more into details, and list the related tools, in the following subsections.

2.2.1 Multiple sequence alignment

This approach is highly similar to the short reads alignment approach for hybrid correction, described in Section 2.1.1, and to the contigs alignment approach described in Section 2.1.2. It is thus composed of a first step of overlaps computation between the long reads, and of a second step of consensus computation from the overlaps. The overlaps computation can be performed either via a mapping strategy, which only provides the positions of the similar regions of the long reads, of via alignment, which provides the positions of the similar regions, as well as their actual base to base correspondence in terms of matches, mismatches, insertions and deletions. For the consensus computation step, a directed acyclic graph (DAG) is usually built in order to summarize the alignments, and extract a consensus sequence. Methods adopting this strategy thus vary by the overlapping strategy they rely on, but also by algorithmic choices made during the consensus computation step.

PBDAGCon (2013)

PBDAGCon is a consensus module, include in the assembler HGAP. First, the alignments between the reads are computed with the help of BLASR. For each read, a DAG is then built, from the set of similar reads determined during the previous step. The vertices of this graph are labeled according to the bases of the reads. An edge is present between two vertices if there exists a read containing the two bases corresponding to these vertices consecutively (*i.e.* there is an edge between u and v if uv is a factor of any read). This way, every read can be represented by a unique path in this graph. Each vertex is thus supported by a given set of reads, and each edge is also supported by a given set of reads. The graph can thus be weighted, by defining the weight of an edge as the number of reads that support it, *i.e.* the number of reads that contain the factor dictated by the two vertices.

The graph is built iteratively. First, it is initialized with the sequence of the read to be corrected. All the edges are thus initialized with weight 1. The graph is then updated, by adding the other reads sequentially, with the help of the information contained in the alignments. This way, if an alignment corresponds to vertices and edges that are already present in the graph, the weight of the corresponding edges are simply incremented. Otherwise, new vertices and edges are created if necessary, and the weight of the edges are set to 1.

One all the reads have been added to the graph, the consensus sequence of the original read can be computed. To this aim, a score is assigned to each vertex, according to the weight of its outgoing edges. The path that maximizes the cumulative score of the traversed vertices is then searched through the graph, with the help of a dynamic programming algorithm. The starting point of this path is defined as the vertex corresponding the first base of the original read, and the ending point is defined as the vertex corresponding to its last base.

PBDAGCon thus produces untrimmed corrected long reads. Indeed, the consensus sequence of a read is computed by searching for a path in the graph between the vertices corresponding to the first and last base of this read. The entire length of the original reads is thus preserved by the correction process. Moreover, the bases of the original reads that could not be corrected, due to a lack of coverage, are not reported in a different case from the corrected bases. As a result, the corrected long reads cannot further be trimmed or split after correction.

PBcR-BLASR (2013)

PBcR-BLASR is a modified version of the PBcR hybrid correction tool, described in Section 2.1.1, that adapts to the problem of self-correction. Thus, the alignment between the reads are first computed with the help of BLASR. The obtained alignments are then used to correct the reads, with the help of the PBcR correction algorithm.

Since it relies on the PBcR algorithm, PBcR-BLASR thus produces split corrected long reads, for the reasons mentioned in the description of the PBcR algorithm, in Section 2.1.1.

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Sprai (2014)

Sprai starts by computing the alignments between the reads with the help of BLAST+. The set of similar reads that align to a given read can thus be used to define a multiple sequence alignment. This multiple sequence alignment is then polished, with the help of ReAligner [3], in order to to maximize it score by modifying its layout, while conserving it global structure. The main aim is to gather similar bases onto identical columns of the matrix representing the multiple sequence alignment. Indeed, local choices are performed when aligning two reads together, in particular in regions denoting insertions and deletions. Such errors being frequent in long reads, the combination of several alignments into a single multiple sequence alignment does not necessarily ensures the coherence of the choices made in each independent alignment. The polishing thus aims to restore this coherence, by locally modifying the multiple sequence alignment, in order to gather enough support at each position.

To this aim, all the sequences includes in the multiple sequence alignment are considered independently. Each of these sequences is then realigned to the multiple sequence alignment, from which the sequence being processed has been excluded, via a dynamic programming algorithm. The multiple sequence alignment is thus updated, according to the result of the alignment computation. Other sequences are then considered iteratively, and the process is thus repeated, until the score of the multiple sequence alignment no longer increases. The final multiple sequence alignment obtained is then used to compute the consensus sequence of the original read, with a majority vote at each position.

Sprai thus produces untrimmed corrected long reads. Indeed, the multiple sequence alignments are always built at the scale of the whole reads. Therefore, the consensus computed via majority vote also contain the uncovered bases of the reads. Moreover, the bases coming from the original reads, and that could not be corrected due to a lack of coverage, are not reported in a different case from the corrected bases. As a result, the corrected long reads cannot further be trimmed or split after correction.

PBcR-MHAP (2015)

PBcR-MHAP is a upgraded version of PBcR-BLASR. In this version, the computation of the alignments between the reads, previously performed with BLASR, is replaced by a mapping strategy called MHAP (Min-Hash Alignment Process). In this approach, the k-mers of the reads are extracted and converted into integer fingerprints with the help of hash functions. A sketch, whose size is defined by the number of hash functions, is thus built for each read. For each of these hash functions, the k-mer of the read that generates the smallest value is chosen as a part of the sketch. Such a k-mer is called a *min-mer*. The sketch of a read is thus defined a the ordered set of the integer fingerprints of its min-mers. The fraction of entries shared by two reads sketches can thus be used to estimate their similarity. Thus, if two given sketches share a sufficient similarity, the min-mers they both share are localized on the corresponding reads, and the median difference between their positions is computed, in order to determine the positions of the overlap between the two reads. A second iteration of this process is then applied, only on overlapping regions, in order to obtain a better estimate of the similarity between these sequences.

Once the overlaps between all the reads have been computed, the overlaps can be used to correct the reads. However, the algorithm used both in PBcR and PBcR-BLASR is replaced by two new consensus modules : FalconSense, which is faster but less sensitive, and PBDAGCon, which is slower but more accurate. These two algorithms are not detailed here, but are described in their respective presentations. Moreover, these two algorithms are not applied one after the other, and the choice as to which one to use is left to the user.

PBcR-MHAP thus produces split corrected long reads if FalconSense is used, and untrimmed corrected long reads, for which uncorrected bases are not reported in a different case from the corrected bases, if PBDAGCon is used. We do not further detail the specificities of these corrected reads, since they are explained in the respective description of FalconSense and PBDAGCon.

FalconSense (2016)

FalconSense is a correction module, which is included in the assembler FALCON. First, the alignments between the reads are computed with the help of DALIGNER [57]. The set of similar reads which align to a given read is then independently realigned to this read, without allowing mismatches, and thus encoding every edit operation with insertions and deletions. For each alignment position between the original read A and one of its similar reads B, a tag is generated. Each of these tags is composed of the four following fields:

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1. The position of read A;
2. The offset from this position on the read A. This value differs from 0 when an insertion is present on read B at this alignment position, and thus indicates the size of the offset between the position on read A and the position on the alignment;
3. The base of read A at this position;
4. The base of read B at this position.

Once all the tags have been defined, for every position of each alignment between the original read and its similar reads, an alignment graph can be built. This graph is a weighted DAG, whose vertices are defined by the tags. An edge is created between two vertices if their corresponding tags are consecutive in one of the alignments. The weight of such an edge is thus set to 1 if it was not already present, and is incremented otherwise. Once the final graph is built, the tag associated to an edge between two vertices represents the number of reads supporting the connexion between these two vertices.

The consensus sequence of the original read can then be computed, by applying a dynamic programming algorithm to the graph, in order to find the highest weighted path between the vertices corresponding to the first and to the last tags associated to the original read. The bases associated to the fourth field of the tags along that path thus dictate the consensus sequence.

FalconSense thus produces split corrected long reads. Indeed, the consensus sequence of a read is computed by searching for a path in the graph, between the first and the last tag associated to this read. Since these tags are computed according to the alignments associated to this read, its uncovered extremities result in a lack of tags, and thus in a lack of vertices and edges in the graph. As a result, these uncovered extremities cannot be included in the consensus sequence computation, and are thus not reported. Moreover, internal regions of a given read which are not covered by alignments also imply a lack of tags, vertices, and edges in the graph. In this case, several traversals are thus performed in the different subgraphs corresponding to covered regions of this read. Several consensus sequences are thus computed independently for each of these regions.

Sparc (2016)

Sparc starts by computing the alignments between the reads with the help of BLASR. For each read, a modified de Bruijn graph is then built, from its set of similar reads, determined during the alignment step. Unlike a classical de Bruijn graph, the vertices of this graph are not only defined from the k-mers of the reads, but also from the positions of these k-mers in the read. Two independent vertices are thus built for two identical k-mers, if they are located at different positions, unlike a classical de Bruijn graph, which only defines a unique vertex in such a case. This necessity to create independent vertices for a same k-mer located at different positions can be explained by the small k values used by Sparc ($l \leq 3$ in practice). Moreover, representing identical k-mers at different positions with distinct vertices also allows to avoid cycles in the graph, and thus compel it to be a DAG. Although the values of k are small, this graph is different from the DAG used in PBDAGCon, as the latter only stores bases of the reads, and not small k-mers.

Moreover, Sparc builds a sparse graph, and only stores one k-mer in every n bases. Edges between the vertices represent consecutive k-mers of the reads, and are labeled with the sequences that allow to link the corresponding vertices. The graph is also weighted, each edge being weighted with the number of reads supporting it. The graph is the initialized from the original read being processed, setting the weight of every edge to 1, and is then iteratively updated according to the alignments associated to this read, in the same way as in PBDAGCon. The weight of the edges are thus are thus incremented if a given alignment corresponds to an existing region of the graph, and new vertices and edges are created otherwise. On the final graph is built, a consensus sequence can be computed, by searching for the highest weighted path, between the first and the last vertex associated to the original read, with the help of a dynamic programming algorithm, in the fashion as PBDDAGCon.

Sparc thus produces untrimmed corrected long reads. Indeed, the consensus sequence of a read is obtained by searching for a path between the first and the last vertex associated to this read. Since these vertices represent respectively the first and the last k-mer of the read, the entire length of the reads is preserved after correction. As for PBDAGCon, bases from the original reads than could not be corrected, due to a lack of coverage, are not reported in a different case from the corrected bases. As a result, the corrected long reads cannot further be trimmed or split after correction.

Moreover, Sparc also proposes a hybrid mode, allowing to use complementary short reads during the alignment step. In this case, the correction process globally remains unchanged. The major difference is that higher weights are associated to edges which are supported by short reads, so they have higher chances of being chosen during the consensus sequence computation.

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Canu (2017)

The correction module of the assembler Canu is an updated version of PBcR-MHAP. During the mapping step, the creation of the sketches is altered, in order to adjust the probability whether to include or not certain k-mers. Indeed, a k-mer that appears frequently in the set of reads should have a lower weight, as it not representative of the true origin of a read within the genome. In contrast, a k-mer which is present in few reads but that appears several times in a single read should have a higher weight, as it represents a greater proportion of the length of this read. A combination of these weights, called tf-idf weight (term frequency inverse document frequency) is thus applied to each k-mer of the reads.

To this aim, for each entry of a read sketch, instead of applying a single hash function to each k-mer, N hash functions are applied, where N corresponds to the tf-idf weight of the k-mer. This way, k-mers having a high tf-idf weight are hashed several times, which increases their chances of being of the sketch. This application of the tf-idf weight to the sketches allows to increase the sensitivity of the overlaps, and to decrease the number of repetitive overlaps. The second iteration of the process, allowing to better estimate the similarity between overlapping regions, is also altered compared to PBcR-MHAP. In particular, a single hash function is used to build the sketches of the overlapping regions.

As in PBcR-MHAP, the overlaps are then used to correct the reads. This step is also improved, and allows to select an optimal subset of overlaps to use for the correction of each read, via a strategy similar to that of the hybrid version of PBcR. However, since the base to base alignments are unavailable, the score associated to each overlap is computed according to its length and to the estimate similarity between the two sequences. Each read is thus only authorized to participate to the correction of its N most similar reads, where N represents the sequencing depth of the reads. This approach allows to reduce the bias induced by repeated regions, by avoiding to always cover these regions with the same sets of reads.

The retained overlaps of each read are then used for correction. The correction is performed with the help of FalconSense, which has been adopted as the default algorithm, while PBDAGCon support has been discarded. Moreover, the FalconSense algorithm is also slightly altered. In particular, the edges of the graph which have a low weight are removed, to ensure that only well supported edges are traversed during the consensus sequence computation.

The Canu correction module thus produces split corrected long reads, for the reasons mentioned in the description of FalconSense.

MECAT (2017)

MECAT also relies on a mapping strategy to compute the overlaps between the reads. This strategy is based on the determination of k-mers shared between the reads, in order to define the overlaps. To this aim, the reads are first divided in several blocks, of 1,000 to 2,000 bps. The reads are then indexed in a hash table, using their k-mers as keys, and the positions of these k-mers in the blocks as the values. To find overlaps between the reads, the k-mers from the blocks associated to the reads are processed and queried in the hash table. A block from a given read thus overlap a block from another read if these two blocks share a sufficient number of k-mers. By extension, a given read overlaps another read if at least a pair of blacks from these two reads do overlap.

A filtering step is the applied, via a scoring system, in order to filter out excessive and uninformative overlaps. To this aim, k-mers pairs are considered within two overlapping blocks. A distance, called DDF (Distance Difference Factor), is then computed between the pairs of k-mers that are shared by the two blocks. This distance is computed according to the occurrence positions of the k-mers within their respective block. A distance shorter than a given threshold indicates that the k-mers are supporting each other. In such a case, the scores of these k-mers are incremented. The k-mer having the highest score within a bloc is then defined as a seed for the following step, if this score is high enough. If several k-mer share the same highest score, one of these k-mers is randomly chosen.

The scoring system is then extended from the current block to its neighbor blocks, after obtaining the seed. The DDF are computed between this seed and the k-mers from the neighbor blocks. As before, if the DDF of a k-mers pair is below a given threshold, the score of the seed is incremented. If more than 80% of the DDFs of a neighbor block are below that threshold, the block is marked, and the scores of its k-mers are not computed. If some blocks remain unmarked after an iteration, the whole process is repeated on these blocks, and the score of their k-mers are computed, as described in the previous paragraph.

After computing all the scores of the k-mers that are shared by two overlapping reads, a base to base alignment of these two reads can be computed. To this aim, the k-mers of these reads are ordered according to their scores. The k-mers displaying the highest scores are then used as seeds, in order to ease the alignment computation. Is the length of the final alignment is at least as large as the size of a block, and if the divergence

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between the two sequences is low enough, the alignment is considered as valid, and is reported.

To correct a given read, the reads overlapping it are sorted, and considered in the decreasing order of the cumulative scores of their k-mers, which are computed during the previous step, via their DDF. Base to base local alignments are thus computed between the read to correct, and the reads overlapping it, in decreasing order of their scores. Moreover, in order to avoid the bias related to chimeric reads and to repeated sequences, an alignment between two reads is filtered out if its length is shorter than 90% of the length of the shortest read. The local alignments computation stops once 100 alignments have been produced, or when all the reads have been processed. Since the DDF score allows to estimate the length of the overlaps, computing local alignments only between the read to correct and its overlapping reads displaying the highest scores allow to gather sufficient information to perform a quick correction, all the while avoiding the computation of repetitive alignments, that would only bring little additional information.

A new correction strategy, combining the principles of both PBDAGCon and FalconSense is then applied. To this aim, the alignments related to the read to correct are summarized in a consensus table that contains the counts of matches, insertions, and deletions, for each position of the read. This table is then browsed, in order to determine three different type of regions, that require different correction approaches.

1. Trivial regions, which are composed of consistent matches and of a small number of insertions (usually < 6);
2. Regions which are composed of consistent deletions, and of a small number of insertions (< 6);
3. More complex regions, composed of a greater number of insertions (≥ 6).

For the two first types of regions, correction is performed by determining the consensus base according to the counts stored in the table. For the third type of regions, a DAG is built, and the consensus is computed by searching for the highest weighted path, via a dynamic programming approach. Since such regions are usually short (< 10 bps), consensus can be computed quickly from the DAG.

MECAT thus produces split corrected long reads. Indeed, the blocks of the reads for which no overlap could be found cannot be corrected, whether they are located at the extremities or in internal regions of the reads. These blocks are thus removed from the corrected reads sequences, and consecutive blocks for which overlaps could be found are reported independently.

FLAS (2019)

FLAS is a wrapper of the MECAT algorithm, which we presented previously. It allows to polish the overlaps before correction, but also to use corrected regions of the reads in order to correct their uncorrected regions. First, the overlaps between the reads are computed with the MECAT mapping strategy. A directed graph is then built from these overlaps. Each vertex of this graph represents a read, and an edge is present between two vertices u and v if the reads associated to these vertices share at least an overlap, over a sufficient length. Once the graph is built, its maximal cliques are computed, with the help of the Bron-Kerbosch algorithm [10, 19, 20].

Pairs of maximal cliques that share common vertices are then processed, in order to find additional overlaps between the corresponding reads, or to remove erroneous overlaps. Three different case can indeed be at the origin of common vertices in a given pair of maximal cliques. These common vertices thus require specific handling.

1. The reads from the two cliques originate from the same genome region, and the overlaps between the reads associated to the common vertices are thus correct;
2. The reads from the two cliques originate from different genome regions, but the reads associated to the common vertices span these two regions, and their overlaps are thus correct;
3. The reads from the two cliques originate from different genome regions, and the reads associated to the common vertices originate from a single of these two regions, and share erroneous overlaps with the reads originating from the other region.

These differences can be determined by comparing the number of common vertices, s, to the number of vertices of the smallest maximal clique of the pair, c. Indeed, among all the overlaps, the number of correct overlaps is usually larger than the number of erroneous overlaps. As a result, if s represents less than 50% of the vertices of c, the common vertices are considered as belonging to the third case.

Otherwise, common vertices are considered as belonging to the first or second case. To distinguish between

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these two cases, the number of reads associated to the common vertices that have a region overlapping the reads from one clique, and a different region overlapping the reads from the other clique, is computed. If this number represents at least 50% of the set of the common vertices of the two cliques, the common vertices are considered as belonging to the second case. In the other case, the common vertices are considered as belonging to the first case.

For the maximal cliques belonging to the first case, the overlaps between the reads associated to the vertices of the cliques are recomputed, in order to produce additional overlaps, and thus be able to correct larger regions of these reads.

For the maximal cliques belonging to the second case, the reads associated to the common vertices span the two genome regions corresponding to the cliques. No supplementary alignment can thus be computed, and no specific action is necessary.

Finally, for the maximal cliques belonging to the third case, the common vertices have to be assigned to a single of the two cliques. To this aim, each common vertex is considered independently. The average similarity of the overlaps between the read associated to the vertex and the other reads of each clique is then computed. The vertex is thus removed from the clique with which it displays the lowest average similarity.

Once the overlaps have been polished, according to these three different cases, the MECAT algorithm is used to perform error correction of the reads.

A second correction step is then applied, where the corrected regions of the reads are used in order to correct the regions that could not be corrected during the first step. To this aim, the overlaps between the corrected regions of the reads are computed with the MECAT mapping strategy. A graph representing the overlaps is then built, in the same fashion as in the previous step. The reads are then aligned to this graph, in order to perform correction. For reads composed of both corrected and uncorrected regions, the corrected regions can be uniquely aligned to the graph, and thus be used as anchor points. The uncorrected regions can then be aligned along paths that allow to link together the corrected regions, and can thus be corrected with the help of the information contained in the followed paths. Moreover, reads solely composed of uncorrected regions can also be aligned to paths of the graph, as it is generated from corrected data, and thus contains a small number of errors. In cases where an uncorrected region of a reads aligns with several paths, the path to which the region aligns with the highest identity is chosen as the correction. If no alignment stands out, the path which is supported by the greatest number of reads is chosen.

FLAS thus produces split corrected long reads. Indeed, FLAS relies on MECAT's mapping and correction strategies. The regions (or blocks, as defined in MECAT) of the reads for which no overlap is found thus cannot be corrected. These regions are thus removed from the corrected reads sequences whether their are located at the reads extremities or not. Consecutive regions for which overlaps could be found are thus reported independently. However, unlike MECAT, FLAS also produces a trimmed version of the corrected reads, which retains the internal regions that could not be corrected due to a lack of overlaps.

2.2.2 De Bruijn graphs

This approach is similar to the hybrid correction approach using de Bruijn graphs, mentioned in Section 2.1.3. In a first step, the graph is built from the long reads k-mers, and in a second step, the graph is traversed in order to find paths allowing to correct unanchored regions of the long reads. The main difference with the hybrid approach comes from the fact that the graph is, here, only constructed from the solid k-mers from the long reads. The methods adopting this approach mainly differ by the scale at which the graph is build. On the one hand, it can either be built globally, by studying the frequency of all the k-mers appearing in the reads. On the other hand, it can be built locally, by first computing overlaps between the long reads, in order to define small similar regions of the long reads, and then building small, local graphs at the scale of these regions.

LoRMA (2016)

LoRMA adapts the principle of the hybrid correction tool LoRDEC to the problem of self-correction. The graph is thus built from the solid k-mers of the reads. The reads can then be anchored to the graph, with the help of their solid k-mers. In the same fashion as LoRDEC, regions composed of weak k-mers, and bordered by regions composed of solid k-mers, are corrected by searching for paths of the graph that allow to link an anchor from the left flanking region to an anchor of the right flanking region. In a similar way, regions composed of weak k-mers, and located at the extremities of the reads, are corrected by using a neighbor solid k-mer as anchor point on the graph, and by traversing the graph, using the same stopping conditions as LoRDEC.

The principle of FMLRC, to perform several passes of correction, with increasing values of k, is also adopted

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in LoRMA. Given the initial error rates of the reads, the first correction pass is performed with a small value of k. Although such small values of k do not allow to correct large weak k-mers regions, due to the high number of branching paths in the graph, small regions can nevertheless be corrected. Hence, after each correction pass, solid k-mers regions grow longer, and a larger value of k can thus be used during the following passes. Moreover, the number of branching paths of the graph also decreases, and larger weak k-mers can thus be corrected. By iteratively reaching high values of k, the whole reads can thus eventually be corrected. Unlike FMLRC, LoRMA however does not use a variable order de Bruijn graph, but instead builds a new graph at each step, according to the chosen value of k.

The corrected reads are then split: regions only composed of weak k-mers are removed, in order to retain only regions solely composed of solid k-mers. The correction of these split reads is then polished via a second correction phase, relying on multiple sequence alignments. To this aim, a de Bruijn graph is built from the corrected reads k-mers, without setting any solidity threshold. The simple paths (*i.e.* with no branching paths) of the graph are then enumerated, in order to define the path of the graph which dictates the sequence of each read. Each path is thus composed of a set of simple paths, called segments. For each of these segments, the set of reads that traverse it is saved. Reads that are similar to a given read can thus be retrieved with the help of the graph, via their common k-mers, by following the path associated to this read, and selecting the reads that traverse common segments. This subset of similar reads is then filtered, in order to only retain the reads that share a sufficient number of k-mers with the initial read.

A consensus sequence can then be computed, from the original read and its set of similar reads. To this aim, the consensus sequence is first initialized as the initial read itself. This consensus is then iteratively updated, by aligning the similar reads. Each of these similar reads is thus considered independently, and aligned to the current consensus. The current multiple sequence alignment is thus updated, and the consensus is then, in turn, updated according to the multiple sequence alignment, via a majority vote at each position. This process is thus repeated until all the similar reads have been considered and aligned. The final multiple sequence alignment is then browsed, and the consensus positions supported by at least two reads are used to polish the correction of the initial read.

Due the the splitting step performed after the correction pass with the increasing-size de Bruijn graphs, LoRMA produces split corrected long reads. Moreover, during the multiple sequence alignment step, bases covered by less that two other reads are reported in a different case from bases covered by a greater number of reads. As a result, the corrected reads can easily further be trimmed or split after correction.

Daccord (2017)

Although it also relies on de Bruijn graphs, Daccord differentiates itself from LoRMA, and does not built a unique graph from the solid k-mers of the whole set of reads. Indeed, it rather builds a set of local graphs, on small regions of subsets of similar reads, defined after a preliminary alignment step. Daccord thus starts by computing the alignments between the reads, with the help of DALIGNER. An alignment between two reads A and B is thus represented by an alignment tuple $(A_b, A_e, B_b, B_e, S, E)$, where:

- A_b and A_e represent, respectively, the beginning and ending positions of the alignment, on A;
- B_b and B_e represent, respectively, the beginning and ending positions of the alignment, on B;
- S represents the orientation of B relatively to A (forward or reverse-complement);
- E represent the edit script (*i.e.* the sequence of edit operations allowing to transform $A[A_b..A_e]$ into $B[B_b..B_e]$, if $S = 0$, or into $\overline{B}[B_b..B_e]$ if $S = 1$ (where $\overline{B}$ represents the reverse-complement of B).

A set of alignment tuples between a given read A and other reads thus define an alignment pile for the read A. The alignment pile of a given read A thus allows to represent the set of reads that align to A, and that are required for the construction of the local de Bruijn graph allowing to correct this read. These graphs are defined on small windows of less than 100 bps of the alignment pile. With the help of the information contained in this alignment pile, the sequences of the read that need to be included in the different windows can easily be extracted. Moreover, only the sequences from the reads that fully span a given window are used to built its associated de Bruijn graph. In particular, reads whose alignments begin or end inside of this window are thus not considered. Furthermore, the windows can be overlapping, and do not necessarily partition the alignment pile into a set of non-overlapping intervals.

Each window of the alignment pile is then processed independently. First, the de Bruijn graph of the window is built from the k-mers of factors of the reads that are included in this window, and that respect the aforementioned conditions. As for the previously described methods that rely on de Bruijn graphs, a source vertex and a target vertex are defined in order to guide the traversal of the graph, which yields a consensus

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sequence for the window. The source vertex is defined, in the left part of the window, as the most frequent first k-mer found in the factors of the reads included in the window. Symmetrically, the target vertex is defined, in the right part of the window, as the most frequent last k-mer found in the factors of the reads included in the window. Several pairs of such k-mers are however considered, in order to reduce the number of cases in which no path could be be found.

Unlike other methods based on the traversal of de Bruijn graphs that were previously presented, the graph is not traversed from the source towards the target nor for the target towards the source. Indeed, Daccord rather traverses the graph in both directions simultaneously, and searches for common vertices that allow to join the two paths. The traversals can thus yield a set of different paths. In this case, the consensus of the window is chosen by aligning the sequences of the obtained paths to the factor of the read A which is included in the window, and by choosing the sequence having the smallest edit distance with this factor. The procedure is thus repeated with all the other windows of the alignment pile of the read to correct.

Once the consensus sequences of all the windows have been computed, the read can eventually be corrected. To this aim, the consensus associated to each window of the alignment pile is aligned to the corresponding factor of the read. The edit scripts that allow to transform the factors of the read A associated to each window into their respective consensus sequences are thus computed. The windows are then once again considered independently, in order to define position pairs, from these edit scripts. For a window beginning at position b on the read A, the position pair $(b+i, 0)$ is assigned to the i-th edit operation which is not an insertion, and the position pair $(b+i, -d)$ is associated to the d-th edit operation which is an insertion, before the i-th edit operation which is not an insertion, considering operation from right to left. Each position pair is annotated with the corresponding base of the window consensus. Once all the windows have been processed, position pairs are sorted, and the consensus sequence of the read is obtained with a majority vote on each position pair, by observing their associated bases.

Daccord thus produces split corrected long reads. Indeed, it is likely that the path searching in the graph does not yield a consensus for a given window. In this case, this windows cannot be considered during the realignment step of the consensus sequences to the initial read. As a result, its edit script, as well as the position pairs associated to it, cannot be determined. These missing consensus sequences thus result in missing position pairs in the sequence of ordered position pairs. The consensus sequence of the read thus cannot be computed on these missing position pairs, and multiple consensus sequences, corresponding to sequences of consecutive position pairs, are produced independently.

2.2.3 Combination of strategies

As for hybrid correction, some methods also rely on combinations of the two previously described strategies. For instance, CONSENT [54] relies on both multiple sequence alignment and de Bruijn graphs, which it combines into a two-step error correction process.

CONSENT (2019)

CONSENT first computes overlaps between the reads, using a mapping approach, with the help of Minimap2 [46]. From these alignments, as in Daccord, alignment piles are then defined for each read. Small windows, of a few hundreds of bp, are then defined on these alignment piles. Each window is processed independently, in two different steps. First, a multiple sequence alignment strategy is used, in order to compute a consensus sequence. Then, the consensus further goes through a second correction step, in which a local de Bruijn graph is built and traversed, in order to further polish remaining errors.

For the multiple alignment step, unlike other methods that would compute independent, pairwise alignments between the window to correct and other windows of the pile, and then summarize them alignments into a DAG, CONSENT computes an actual MSA between all the sequences. To this aim, it makes use of poaV2 [42, 41], a algorithm that computes MSA with the help of partial order graphs. These graphs are DAGs, and are used as data structures containing all the MSA information. Fore a given MSA, the graph is thus enriched at each step, by aligning the new sequences to it, and updating it with additional vertices and edges, if necessary. Unlike other methods, this strategy thus allows CONSENT to both compute MSA and built the DAG used for consensus computation at the same time.

However, POAv2 is resource consuming, even of windows of a few hundreds bp. To address this issue, CONSENT relies on an efficient segmentation strategy, that allows to compute MSA and consensus on much shorter sequences, thus greatly reducing computational costs. First, common k-mers between the sequences included in the window being processed are computed. Using a dynamic programming algorithm, the longest chain of

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consecutive k-mers, common to the sequences, is then computed. This way, MSA and consensus sequences only need to be computed on subsequences bordered by the k-mers of the chain. CONSENT then reconstructs the global consensus, by concatenating the local consensus and the k-mers that were used for segmentation.

After consensus computation, a few erroneous bases might remain. To further enhance the quality of the corrected window, CONSENT thus performs a second correction phase, where the consensus sequence is polished with the help of a de Bruijn graph. First, the graph is thus built from the solid k-mers of the sequences included in the window. Due to the small size of the windows, a small k-mers size (usually, $k = 9$) can also be used. As in other de Bruijn graph based methods presented previously, solid k-ùers of the consensus are used as anchor points on the graph, and weak k-mer regions are corrected, by searching for paths of the graph that allow to link together two anchors. In the same fashion, extremities of the consensus are polished by following the highest weighted edges of the graph, until the length of the path reaches the length of the extremity to correct, or until a branching path is reached.

Finally, after computing and polishing the consensus of a given window, the obtained sequence is locally aligned to the original read, around the positions the window originates from, to actually perform correction. This process is thus applied to each window of the alignment pile, until the read is completely corrected.

CONSENT thus produces trimmed corrected long reads. Indeed, since windows cannot be defined on the uncovered extremities of the long reads, CONSENT cannot process and correct these extremities. They are thus removed from the sequences of the corrected long reads. Moreover, during the consensus polishing step, CONSENT reports bases corresponding to solid k-mers and bases corresponding to weak k-mers in a different case. The corrected long reads can thus easily further be split after correction.

2.3 Summary

In the section, we described the state-of-the-art of available methods for the error correction of long reads, whether they adopt a hybrid or a self-correction approach. Four main approaches were described for self-correction: the alignment of short reads to the long reads, the alignment of contigs obtained from short reads assembly and long reads, the use of de Bruijn graphs, and the use of hidden Markov models. Other methods also combine different strategies to benefit from their different advantages. Two approaches were described for self-correction: multiple sequence alignment, and use of de Bruijn graphs. As for hybrid correction, other methods also combine these two strategies to counterbalance their advantages and drawbacks. The algorithmic principles of each error correction method have also been described. Available hybrid correction tools are summarized in Table 1. Available self-correction tools are summarized in Table 2. These tables also recall the approaches these different methods rely on, the way they output the reads (*i.e.* trimmed / split or not), the year they were released, as well as the technology (PacBio or ONT) they were validated on, in their respective publications.

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Method	Approach	Output reads			Case	Release	Validated on
		Complete	*Trimmed*	*Split*			
PBcR	Short reads alignment	✖	✖	✔	✖	2012	PacBio
LSC	Short reads alignment	✖	✔	✖	✖	2012	PacBio
ECTools	Contigs alignment	✖	✖	✔	✖	2014	PacBio
LoRDEC	De Bruijn graphs	✔	✖	✖	✔	2014	PacBio
Proovread	Short reads alignment	✔	✖	✔	✖	2014	PacBio
Nanocorr	Short reads alignment	✔	✖	✖	✖	2015	ONT
NaS	Short reads alignment	✔	✖	✖	✖	2015	ONT
CoLoRMap	Short reads alignment	✔	✖	✖	✔	2016	PacBio
Jabba	De Bruijn graphs	✖	✖	✔	✖	2016	PacBio
LSCplus	Short reads alignment	✔	✔	✖	✖	2016	PacBio
HALC	Contigs alignment	✔	✔	✔	✔	2017	PacBio
HECIL	Short reads alignment	✔	✖	✖	✖	2018	PacBio
Hercules	Hidden Markov models	✔	✖	✖	✖	2018	PacBio
FMLRC	De Bruijn graphs	✔	✖	✖	✖	2018	PacBio
HG-CoLoR	Short reads alignment + de Bruijn graphs	✔	✔	✔	✔	2018	PacBio + ONT
MiRCA	Contigs alignment	✖	✔	✖	✖	2018	ONT
ParLECH	De Bruijn graphs	-	-	-	-	2019	PacBio

Table 1: **List of available hybrid correction tools.** Complete reads correspond to reads which are neither trimmed nor split. The case column indicates whether the method reports corrected bases in a different case from the uncorrected bases.

Method	Approach	Output reads			Case	Release	Validated on
		Complete	*Trimmed*	*Split*			
PBcR-BLASR	Multiple sequence alignment	✖	✖	✔	✖	2013	PacBio
PBDAGCon	Multiple sequence alignment	✔	✖	✖	✖	2013	PacBio
Sprai	Multiple sequence alignment	✔	✖	✖	✖	2014	PacBio
PBcR-MHAP (PBDAGCon)	Multiple sequence alignment	✔	✖	✖	✖	2015	PacBio
PBcR-MHAP (FalconSense)	Multiple sequence alignment	✖	✖	✔	✖	2015	PacBio
FalconSense	Multiple sequence alignment	✖	✖	✔	✖	2016	PacBio
LoRMA	De Bruijn graphs	✖	✖	✔	✖	2016	PacBio
Sparc	Multiple sequence alignment	✔	✖	✖	✖	2016	PacBio + ONT
Canu	Multiple sequence alignment	✖	✖	✔	✖	2017	PacBio + ONT
Daccord	De Bruijn graphs	✖	✖	✔	✖	2017	PacBio + ONT
MECAT	Multiple sequence alignment	✖	✖	✔	✖	2017	PacBio + ONT
CONSENT	Multiple sequence alignment + de Bruijn graphs	✖	✔	✖	✔	2019	PacBio + ONT
FLAS	Multiple sequence alignment	✖	✔	✔	✖	2019	PacBio

Table 2: **List of available self-correction tools.** Complete reads correspond to reads which are neither trimmed nor split. The case column indicates whether the method reports corrected bases in a different case from the uncorrected bases.

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3 Qualitative comparison

In this section, we present an in-depth benchmark of available hybrid and self-correction tools described in Section 2, and summarized in Table 1 and in Table 2. The following methods are however excluded from the benchmark, for the reasons mentioned below:

- FalconSense: this tool could not be installed on any of the computers used in our experiments;
- HECIL: this tool did not manage to correct any read, and produced a set of corrected read identical to the set of uncorrected reads, for all the datasets;
- LSCplus: this tool is not available for download anymore;
- MiRCA: this tool stopped and reported an error during the correction of all the datasets;
- ParLECH: this tool could not be installed on any of the computers used in our experiments;
- PBcR: this tool stopped and reported an error during the correction of all the datasets;
- PBcR-BLASR and PBcR-MHAP: this two tools are preliminary versions of the correction module from the assembler Canu. Only the latter, which is the most recent, is thus evaluated in the benchmark;
- PBDAGCon: this tool did not manage to produce any corrected read for all the datasets;
- Sparc: this tool stopped and reported an error during the correction of all the datasets;
- Sprai: this tool stopped and reported an error during the correction of all the datasets.

3.1 Datasets

We evaluated these correction methods on a wide variety of datasets, composed of both simulated and real data, displaying various error rates, read length and sequences depths, and ranging from small bacterial to large mammal genomes. These datasets are summarized in Table 3.

Dataset	Number of reads	Number of bases (Mbp)	Average length (bp)	Coverage	Error rate (%)	Accession
Simulated PacBio data						
A. baylyi 20x	8,795	72	8,202	20x	18.57	-
E. coli 20x	11,306	93	8,226	20x	18.65	-
S. cerevisiae 20x	30,132	247	8,204	20x	18.61	-
C. elegans 20x	244,277	2,004	8,204	20x	18.62	-
E. coli 30x	16,959	140	8,235	30x	12.29	-
S. cerevisiae 30x	45,198	371	8,216	30x	12.28	-
C. elegans 30x	366,416	3,006	8,204	30x	12.28	-
E. coli 60x (low)	33,918	279	8,211	60x	12.28	-
S. cerevisiae 60x (low)	90,397	742	8,204	60x	12.29	-
C. elegans 60x (low)	732,832	6,011	8,220	60x	12.28	-
A. baylyi 60x (medium)	26,296	216	8,213	60x	18.59	-
E. coli 60x (medium)	33,918	279	8,217	60x	18.62	-
S. cerevisiae 60x (medium)	90,397	743	8,221	60x	18.61	-
C. elegans 60x (medium)	732,832	6,022	8,218	60x	18.60	-
Real ONT data						
A. baylyi real	89,011	381	4,284	106x	29.91	Genoscope[1]
E. coli real	22,270	134	5,999	29x	20.54	Genoscope[2]
S. cerevisiae real	205,923	1,173	5,698	95x	44.51	Genoscope[3]
C. elegans real	363,500	2,008	5,524	20x	28.93	ERR1802061
D. melanogaster	1,327,569	9,064	6,828	63x	14.55	SRX3676783
H. sapiens[4]	1,075,867	7,256	6,744	29x	17.60	PRJEB23027
Illumina data						
A. baylyi	900,000	224	250	50x	1	ERR788913[5]
E. coli	775,500	232	300	50x	1	Genoscope[6]
S. cerevisiae	2,500,000	625	250	50x	1	Genoscope[7]
C. elegans	20,057,100	5,000	250	50x	1	ART [28]

Table 3: Characteristics of the datasets used during the experiments.
[1]http://www.genoscope.cns.fr/externe/nas/datasets/MinION/acineto/
[2]http://www.genoscope.cns.fr/externe/nas/datasets/MinION/ecoli/
[3]http://www.genoscope.cns.fr/externe/nas/datasets/MinION/yeast/
[4] Only reads from chromosome 1 were used. This dataset contains ONT ultra-long reads, reaching lengths up to 340 kbps.
[5]http://www.genoscope.cns.fr/externe/nas/datasets/Illumina/acineto/
[6]http://www.genoscope.cns.fr/externe/nas/datasets/Illumina/ecoli/
[7]http://www.genoscope.cns.fr/externe/nas/datasets/Illumina/yeast/

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3.2 Benchmark summary

To evaluate the quality of the correction provided by each tool, we used ELECTOR [51], a software specially developed for large scale error correction tools benchmark, that allows to assess both the correction of simulated and real data. In particular, it reports the error rates of the reads before and after correction, as well as the recall and the precision of the assessed tools, among other metrics, when ran on simulated data. With real data, it is able to perform remapping of the reads to the reference genome with the help of Minimap2 [46], and reports the average identity of the alignments as well as the genome coverage, among other metrics. It is also able to perform assembly of the corrected reads, with the help of Miniasm [45], and reports the number of aligned contigs, the NGA50, the NGA75, and the genome coverage, among other metrics.
We present results in various subsections, according to the characteristics of the assessed datasets. In particular, we distinguish seven different categories of datasets.

1. Datasets with high error rate and high coverage. This corresponds to the *A. baylyi* real and *S. cerevisiae* real datasets of Table 3. We present these results in Section 3.3, Table 4.
2. Datasets with high error rate a low coverage. This corresponds to the *C. elegans* real dataset of Table 3. We present these results in Section 3.4, Table 5.
3. Datasets with medium error rate and low coverage. This corresponds to the *A. baylyi* 20x, *E. coli* 20x, *S. cerevisiae* 20x, *C. elegans* 20x and *E. coli* real datasets of Table 3. We present these results in Section 3.5, Tables 8 and 7.
4. Datasets with medium error rate and medium coverage. This corresponds to the *A. baylyi* 60x (medium), *E. coli* 60x (medium), *S. cerevisiae* 60x (medium) and *C. elegans* 60x (medium) datasets of Table 3. We present these results in Section 3.6, Table **??**.
5. Datasets with low error rate and low coverage. This corresponds to the *E. coli* 30x, *S. cerevisiae* 30x and *C. elegans* 30x datasets of Table 3. We present these results in Section 3.7, Table 9.
6. Datasets with low error rate and medium coverage. This corresponds to the *E. coli* 60x (low), *S. cerevisiae* 60x (low) and *C. elegans* 60x (low) datasets of Table 3. We present these results in Section 3.8, Table 10.
7. Datasets containing ONT ultra-long reads. This corresponds to the *H. sapiens* dataset of Table 3. We present these results in Section 3.9, Table 11.

3.3 High error rate, high coverage

Tool		Remapping			Assembly	
		A. baylyi	*S. cerevisiae*		*A. baylyi*	*S. cerevisiae*
CoLoRMap	Number of reads	36,422	72,017	Number of contigs	1	71
	Number of bases	141,415,030	165,218,405	Number of aligned contigs	1	68
	Average length (bp)	3,882	2,294	Number of breakpoints	54	47
	Aligned reads (%)	99.9973	99.7403	NGA50 (bp)	3,588,032	204,052
	Average identity (%)	99.5079	99.6958	NGA75 (bp)	3,588,032	88,468
	Genome coverage (%)	100.0000	99.1528	Genome coverage (%)	96.4454	87.1140
	Runtime	3 h 41 min	10 h 44 min			
	Memory (MB)	13,028	18,241			
ECTools	Number of reads	6,882	21,780	Number of contigs	101	313
	Number of bases	36,892,324	116,347,153	Number of aligned contigs	101	313
	Average length (bp)	5,360	5,341	Number of breakpoints	0	0
	Aligned reads (%)	100.0000	100.0000	NGA50 (bp)	12,654	7,294
	Average identity (%)	98.6844	98.6497	NGA75 (bp)	8,579	7,294
	Genome coverage (%)	95.3957	87.2324	Genome coverage (%)	55.6888	48.0243
	Runtime	26 min	3 h 13 min			
	Memory (MB)	862	2,256			
FMLRC	Number of reads	89,011	205,923	Number of contigs	1	54
	Number of bases	390,735,419	1,185,455,434	Number of aligned contigs	1	54
	Average length (bp)	4,389	5,756	Number of breakpoints	21	69
	Aligned reads (%)	27.7741	31.7497	NGA50 (bp)	3,673,304	378,469
	Average identity (%)	99.6779	96.7164	NGA75 (bp)	3,673,304	186,053
	Genome coverage (%)	100.0000	99.8120	Genome coverage (%)	96.8804	92.2118
	Runtime	2 h 01 min	6 h 15 min			
	Memory (MB)	449	876			

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Tool	Remapping			Assembly		
		A. baylyi	*S. cerevisiae*		*A. baylyi*	*S. cerevisiae*
HALC	Number of reads	42,188	135,050	Number of contigs	2	82
	Number of bases	189,757,075	255,641,564	Number of aligned contigs	2	82
	Average length (bp)	4,497	1,892	Number of breakpoints	36	75
	Aligned reads (%)	99.6444	99.5091	NGA50 (bp)	2,420,463	313,785
	Average identity (%)	99.8345	99.2933	NGA75 (bp)	1,207,743	146,613
	Genome coverage (%)	100.0000	99.1874	Genome coverage (%)	97.3183	92.0039
	Runtime	47 h 41 min	2 h 56 min			
	Memory (MB)	10,577	2,329			
Hercules	Number of reads	89,011	205,923	Number of contigs	1	174
	Number of bases	383,188,837	1,173,722,512	Number of aligned contigs	1	171
	Average length (bp)	4,304	5,699	Number of breakpoints	49	10
	Aligned reads (%)	21.5468	22.7682	NGA50 (bp)	3,537,383	3,384
	Average identity (%)	81.3432	71.4998	NGA75 (bp)	3,537,383	3,384
	Genome coverage (%)	100.0000	99.7669	Genome coverage (%)	97.4830	29.1838
	Runtime	16 h 53 min	12 h 13 min			
	Memory (MB)	4,438	19,885			
HG-CoLoR	Number of reads	25,536	76,193	Number of contigs	2	53
	Number of bases	284,883,716	512,438,767	Number of aligned contigs	2	53
	Average length (bp)	11,156	6,725	Number of breakpoints	5	72
	Aligned reads (%)	99.9883	99.7126	NGA50 (bp)	3,595,353	470,355
	Average identity (%)	99.9760	99.7176	NGA75 (bp)	3,595,353	212,761
	Genome coverage (%)	100.0000	99.5341	Genome coverage (%)	99.9182	95.3867
	Runtime	1 h 34 min	8 h 51 min			
	Memory (MB)	3,750	11,575			
Jabba	Number of reads	17,483	37,703	Number of contigs	17	176
	Number of bases	179,366,684	243,374,292	Number of aligned contigs	17	176
	Average length (bp)	10,259	6,455	Number of breakpoints	1	2
	Aligned reads (%)	99.9714	98.5386	NGA50 (bp)	216,570	45,205
	Average identity (%)	99.9226	99.8889	NGA75 (bp)	184,851	6,507
	Genome coverage (%)	99.8170	93.3275	Genome coverage (%)	93.7381	72.0151
	Runtime	2 min	7 min			
	Memory (MB)	1,217	1,217			
LoRDEC	Number of reads	50,776	196,091	Number of contigs	2	66
	Number of bases	175,140,999	220,706,773	Number of aligned contigs	2	66
	Average length (bp)	3,449	1,125	Number of breakpoints	73	126
	Aligned reads (%)	99.8779	98.5094	NGA50 (bp)	3,010,005	361,726
	Average identity (%)	99.9448	98.8168	NGA75 (bp)	3,010,005	178,604
	Genome coverage (%)	100.0000	98.8934	Genome coverage (%)	94.6755	90.0706
	Runtime	16 min	1 h 09 min			
	Memory (MB)	436	797			
LSC[1]	Number of reads	-	10,054	Number of contigs	-	2
	Number of bases	-	36,116,073	Number of aligned contigs	-	2
	Average length (bp)	-	3,592	Number of breakpoints	-	0
	Aligned reads (%)	-	93.2664	NGA50 (bp)	-	5,393
	Average identity (%)	-	88.5159	NGA75 (bp)	-	5,393
	Genome coverage (%)	-	88.2405	Genome coverage (%)	-	0.1190
	Runtime	-	8 h 58 min			
	Memory (MB)	-	1,431			
Nanocorr	Number of reads	24,105	66,953	Number of contigs	1	170
	Number of bases	173,666,898	231,317,559	Number of aligned contigs	1	170
	Average length (bp)	7,204	3,454	Number of breakpoints	41	26
	Aligned reads (%)	99.7884	98.8873	NGA50 (bp)	3,647,559	84,048
	Average identity (%)	98.3639	97.1970	NGA75 (bp)	3,647,559	44,028
	Genome coverage (%)	100.0000	99.5137	Genome coverage (%)	98.2877	89.5144
	Runtime	22 h 28 min	158 h 53 min			
	Memory (MB)	616	3,128			
NaS[2]	Number of reads	24,063	71,793	Number of contigs	1	159
	Number of bases	212,707,189	426,326,355	Number of aligned contigs	1	159
	Average length (bp)	8,839	5,938	Number of breakpoints	2	17
	Aligned reads (%)	100.0000	99.8231	NGA50 (bp)	3,600,775	85,205
	Average identity (%)	99.9900	99.8985	NGA75 (bp)	3,600,775	43,313
	Genome coverage (%)	100.0000	98.7695	Genome coverage (%)	99.9986	89.7248
	Runtime	94 h 19 min	> 16 days			
	Memory (MB)	2,099	-			

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Tool	Remapping			Assembly		
		A. baylyi	*S. cerevisiae*		*A. baylyi*	*S. cerevisiae*
Proovread	Number of reads	38,432	91,055	Number of contigs	3	77
	Number of bases	155,655,071	160,204,364	Number of aligned contigs	3	74
	Average length (bp)	4,050	1,759	Number of breakpoints	60	48
	Aligned reads (%)	100.0000	99.7156	NGA50 (bp)	1,304,077	176,285
	Average identity (%)	99.9686	99.8979	NGA75 (bp)	1,304,077	80,275
	Genome coverage (%)	100.0000	99.1089	Genome coverage (%)	92.7046	86.0619
	Runtime	3 h 25 min	13 h 42 min			
	Memory (MB)	10,618	8,709			
Canu[3]	Number of reads	8,632	-	Number of contigs	19	-
	Number of bases	80,666,491	-	Number of aligned contigs	19	-
	Average length (bp)	9,345	-	Number of breakpoints	3	-
	Aligned reads (%)	99.9421	-	NGA50 (bp)	2,146,043	-
	Average identity (%)	94.5919	-	NGA75 (bp)	99,247	-
	Genome coverage (%)	99.7861	-	Genome coverage (%)	93.6289	-
	Runtime	31 min	-			
	Memory (MB)	3,015	-			
CONSENT	Number of reads	16,928	24,862	Number of contigs	1	186
	Number of bases	183,088,372	178,658,682	Number of aligned contigs	1	185
	Average length (bp)	10,815	7,186	Number of breakpoints	23	18
	Aligned reads (%)	99.4506	96.7702	NGA50 (bp)	3,494,031	3,326
	Average identity (%)	91.9470	76.7265	NGA75 (bp)	3,494,031	3,326
	Genome coverage (%)	100.0000	98.1075	Genome coverage (%)	99.1676	36.9326
	Runtime	48 min	40 min			
	Memory (MB)	5,150	14,663			
Daccord[4]	Number of reads	53,926	-	Number of contigs	2	-
	Number of bases	174,962,080	-	Number of aligned contigs	2	-
	Average length (bp)	3,244	-	Number of breakpoints	4	-
	Aligned reads (%)	97.1906	-	NGA50 (bp)	3,472,403	-
	Average identity (%)	93.2546	-	NGA75 (bp)	3,472,403	-
	Genome coverage (%)	100.0000	-	Genome coverage (%)	100.0000	-
	Runtime	43 min	-			
	Memory (MB)	25,801	-			
FLAS	Number of reads	18,658	34,483	Number of contigs	2	114
	Number of bases	165,473,591	220,777,235	Number of aligned contigs	2	114
	Average length (bp)	8,868	6,402	Number of breakpoints	32	8
	Aligned reads (%)	99.8553	84.1197	NGA50 (bp)	1,610,377	2,974
	Average identity (%)	91.6074	77.1713	NGA75 (bp)	1,610,377	2,974
	Genome coverage (%)	100.0000	98.6900	Genome coverage (%)	96.9851	21.8854
	Runtime	32 min	39 min			
	Memory (MB)	3,015	7,398			
LoRMA[5]	Number of reads	333,041	48,824	Number of contigs	1	-
	Number of bases	76,354,713	11,246,944	Number of aligned contigs	1	-
	Average length (bp)	229	230	Number of breakpoints	0	-
	Aligned reads (%)	99.9168	57.6397	NGA50 (bp)	4,949	-
	Average identity (%)	98.0710	95.2610	NGA75 (bp)	4,949	-
	Genome coverage (%)	66.4788	3.6315	Genome coverage (%)	0.1406	-
	Runtime	29 min	1 h 35 min			
	Memory (MB)	31,575	1,505			
MECAT	Number of reads	16,811	14,740	Number of contigs	1	102
	Number of bases	154,436,057	83,553,890	Number of aligned contigs	1	102
	Average length (bp)	9,186	5,668	Number of breakpoints	3	2
	Aligned reads (%)	100.0000	99.4573	NGA50 (bp)	3,335,596	3,163
	Average identity (%)	91.4676	80.0763	NGA75 (bp)	3,335,596	3,163
	Genome coverage (%)	100.0000	92.6533	Genome coverage (%)	99.9985	15.1658
	Runtime	23 min	14 min			
	Memory (MB)	2,978	7,374			

Table 4: Comparison of the different error correction tools, on high error rate and high coverage datasets. This corresponds to the *A. baylyi* real and *S. cerevisiae* real datasets of Table 3. [1] LSC stopped and reported and error during the correction of the *A. baylyi* dataset. [2] NaS was stopped after 16 days on the *S. cerevisiae* datasets. Corrected long reads were obtained from the Genoscope website. [3] Canu stopped and reported an error during the correction of the *S. cerevisiae* dataset. [4] Daccord could not perform correction on the *S. cerevisiae* dataset, since the alignment step with DALIGNER required more than 128 GB of memory. [5] LoRMA corrected reads could not be assembled for the *S. cerevisiae* dataset.

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3.4 High error rate, low coverage

For these experiments, we exclude the following tools from the comparison, for the reasons mentioned below:

- HALC, because it stopped and reported an error during correction;
- LSC, because of its large runtimes on previous experiments;
- Nanocorr, because of its large runtimes on previous experiments;
- NaS, because of its large runtimes on previous experiments;
- Daccord, because it could not perform correction, since the alignment step with DALIGNER required more than 128 GB of memory;
- FLAS, because it stopped and reported an error during correction;
- MECAT, because it stopped and reported an error during correction.

Tool	Remapping		Assembly	
CoLoRMap	Number of reads	184,092	Number of contigs	1,246
	Number of bases	418,509,369	Number of aligned contigs	1,237
	Average length (bp)	2,273	Number of breakpoints	617
	Aligned reads (%)	99.9473	NGA50 (bp)	58,411
	Average identity (%)	98.6614	NGA75 (bp)	11,122
	Genome coverage (%)	96.4079	Genome coverage (%)	76.6546
	Runtime	91 h 18 min		
	Memory (MB)	31,349		
ECTools[1]	Number of reads	18	Number of contigs	-
	Number of bases	54,323	Number of aligned contigs	-
	Average length (bp)	3,017	Number of breakpoints	-
	Aligned reads (%)	100.0000	NGA50 (bp)	-
	Average identity (%)	98.4596	NGA75 (bp)	-
	Genome coverage (%)	0.016	Genome coverage (%)	-
	Runtime	81 h 25 min		
	Memory (MB)	5,023		
FMLRC	Number of reads	363,500	Number of contigs	1,047
	Number of bases	2,063,059,647	Number of aligned contigs	1,042
	Average length (bp)	5,675	Number of breakpoints	713
	Aligned reads (%)	78.1986	NGA50 (bp)	91,545
	Average identity (%)	96.5611	NGA75 (bp)	40,293
	Genome coverage (%)	99.9932	Genome coverage (%)	82.9120
	Runtime	7 h 54 min		
	Memory (MB)	7,141		
Hercules	Number of reads	363,500	Number of contigs	1,356
	Number of bases	2,012,600,582	Number of aligned contigs	1,348
	Average length (bp)	5,536	Number of breakpoints	425
	Aligned reads (%)	72.8468	NGA50 (bp)	36,090
	Average identity (%)	82.3142	NGA75 (bp)	690
	Genome coverage (%)	99.9776	Genome coverage (%)	66.7194
	Runtime	20 h 36 min		
	Memory (MB)	32,000		
HG-CoLoR	Number of reads	305,777	Number of contigs	989
	Number of bases	1,567,516,029	Number of aligned contigs	988
	Average length (bp)	5,126	Number of breakpoints	461
	Aligned reads (%)	99.8653	NGA50 (bp)	96,058
	Average identity (%)	99.5430	NGA75 (bp)	41,829
	Genome coverage (%)	99.9655	Genome coverage (%)	82.6239
	Runtime	83 h 10 min		
	Memory (MB)	19,836		
Jabba	Number of reads	270,929	Number of contigs	880
	Number of bases	433,372,687	Number of aligned contigs	880
	Average length (bp)	1,599	Number of breakpoints	15
	Aligned reads (%)	97.2901	NGA50 (bp)	2,046
	Average identity (%)	99.9430	NGA75 (bp)	2,046
	Genome coverage (%)	95.2852	Genome coverage (%)	30.0434
	Runtime	59 min		
	Memory (MB)	13,362		

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Tool	Remapping		Assembly	
LoRDEC	Number of reads	167,699	Number of contigs	21
	Number of bases	221,676,779	Number of aligned contigs	16
	Average length (bp)	1,321	Number of breakpoints	9
	Aligned reads (%)	96.0137	NGA50 (bp)	589
	Average identity (%)	98.2718	NGA75 (bp)	589
	Genome coverage (%)	85.0782	Genome coverage (%)	0.0854
	Runtime	1 h 15 min		
	Memory (MB)	2,373		
Proovread	Number of reads	364,244	Number of contigs	1,095
	Number of bases	1,329,812,416	Number of aligned contigs	1,086
	Average length (bp)	3,650	Number of breakpoints	509
	Aligned reads (%)	99.7194	NGA50 (bp)	80,171
	Average identity (%)	99.6065	NGA75 (bp)	31,114
	Genome coverage (%)	99.9358	Genome coverage (%)	79.6994
	Runtime	119 h 48 min		
	Memory (MB)	20,254		
Canu	Number of reads	340,826	Number of contigs	1,272
	Number of bases	1,843,278,326	Number of aligned contigs	1,259
	Average length (bp)	5,408	Number of breakpoints	512
	Aligned reads (%)	76.7204	NGA50 (bp)	54,194
	Average identity (%)	90.6784	NGA75 (bp)	416
	Genome coverage (%)	99.9783	Genome coverage (%)	76.9836
	Runtime	14 h 19 min		
	Memory (MB)	9,178		
CONSENT	Number of reads	256,568	Number of contigs	1,279
	Number of bases	1,475,025,831	Number of aligned contigs	1,268
	Average length (bp)	5,749	Number of breakpoints	624
	Aligned reads (%)	96.7615	NGA50 (bp)	49,620
	Average identity (%)	91.6888	NGA75 (bp)	1,606
	Genome coverage (%)	99.7682	Genome coverage (%)	73.1300
	Runtime	9 h 50 min		
	Memory (MB)	20,382		
LoRMA[2]	Number of reads	38,121	Number of contigs	-
	Number of bases	10,277,321	Number of aligned contigs	-
	Average length (bp)	269	Number of breakpoints	-
	Aligned reads (%)	91.5506	NGA50 (bp)	-
	Average identity (%)	97.6599	NGA75 (bp)	-
	Genome coverage (%)	1.6269	Genome coverage (%)	-
	Runtime	1 h 13 min		
	Memory (MB)	32,275		

Table 5: Comparison of the different error correction tools, on high error rate and low coverage datasets. This corresponds to the *C. elegans* real dataset of Table 3. [1] ECTools corrected long reads could not be assembled. [2] LoRMA corrected long reads could not be assembled.

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3.5 Medium error rate, low coverage

3.5.1 Simulated data

Tool	Metric	Dataset			
		A. baylyi	*E. coli*	*S. cerevisiae*	*C. elegans*
CoLoRMap	Number of bases	64,353,820	81,060,058	210,966,804	571,054,288
	Error rate (%)	0.1336	0.1946	0.2655	2.6255
	Deletions	71,639	165,412	434,983	14,745,290
	Insertions	7,062	9,193	51,627	374,284
	Substitutions	51,954	62,188	279,378	3,722,575
	Recall (%)	99.9923	99.9890	99.9805	99.8445
	Precision (%)	99.8707	99.8118	99.7413	97.4526
	Trimmed / split reads	1,666	2,067	6,970	64,051
	Mean missing size (bp)	939.2	1,497.5	1,396.2	3,234.8
	Extended reads	8,463	10,730	28,117	96,953
	Mean extension size (bp)	260.3	308.8	251.6	233
	Low quality reads	12	20	121	332
	Short reads	689	1,036	2,166	181,741
	Homopolymer ratio	1.0000	1.0000	0.9826	0.9962
	Runtime	57 min	1 h 25 min	4 h 42 min	125 h 44 min
	Memory (MB)	6,198	9,659	13,544	32,188
ECTools	Number of bases	36,892,324	6,261,214	116,347,153	54,323
	Error rate (%)	1.2086	1.3991	1.2428	1.4652
	Deletions	77,464	13,883	246,662	123
	Insertions	395,759	76,702	1,275,965	702
	Substitutions	40,670	7,026	140,703	45
	Recall (%)	98.8241	98.6228	98.8048	98.5410
	Precision (%)	98.7923	98.6012	98.7581	98.5348
	Trimmed / split reads	5,628	1,595	18,270	18
	Mean missing size (bp)	2,730.3	4,103.1	2,747.6	4,770.3
	Extended reads	0	0	0	0
	Mean extension size (bp)	0	0	0	0
	Low quality reads	0	0	0	0
	Short reads	0	0	0	0
	Homopolymer ratio	1.0064	1.0000	0.9991	1.0045
	Runtime	17 min	8 min	57 h 12 min	77 h 59 min
	Memory (MB)	862	917	2,256	5,028
FMLRC	Number of bases	64,715,552	83,732,334	223,529,739	1,832,773,801
	Error rate (%)	0.1194	0.1930	1.0016	3.3582
	Deletions	26,253	54,707	643,464	20,717,465
	Insertions	89,640	185,196	2,289,467	52,872,229
	Substitutions	22,637	45,795	546,972	16,628,231
	Recall (%)	99.9142	99.8498	99.1797	97.8709
	Precision (%)	99.8815	99.8081	99.0020	96.6643
	Trimmed / split reads	20	28	114	3,090
	Mean missing size (bp)	26.2	29.4	17.2	32.8
	Extended reads	0	0	9	295
	Mean extension size (bp)	0	0	290.8	31
	Low quality reads	13	29	120	744
	Short reads	0	0	0	0
	Homopolymer ratio	1.0000	1.0000	0.9741	1.0000
	Runtime	23 min	29 min	1 h 19 min	8 h 02 min
	Memory (MB)	387	408	906	7,939
HALC	Number of bases	63,708,698	81,199,351	212,266,193	1,588,220,052
	Error rate (%)	0.1113	0.1537	0.4212	1.5288
	Deletions	59,157	115,973	1,035,978	35,970,722
	Insertions	8,273	15,034	100,874	2,520,363
	Substitutions	5,206	13,764	198,853	2,919,936
	Recall (%)	99.9937	99.9918	99.9734	99.8945
	Precision (%)	99.8909	99.8497	99.5872	98.4961
	Trimmed / split reads	2,358	3,816	12,043	153,855
	Mean missing size (bp)	341.7	547.3	577.5	777.2
	Extended reads	24	24	71	772
	Mean extension size (bp)	25.2	31.1	53.2	40.9
	Low quality reads	2	2	160	2,582
	Short reads	101	254	3,436	170,934
	Homopolymer ratio	1.0000	1.0000	1.0580	0.9978
	Runtime	22 min	24 min	1 h 19 min	5 h 59 min
	Memory (MB)	597	965	2,115	2,323

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Tool	Metric	Dataset			
		A. baylyi	*E. coli*	*S. cerevisiae*	*C. elegans*
Hercules	Number of bases	70,375,149	91,064,066	242,189,693	1,976,944,911
	Error rate (%)	10.5079	10.6411	10.9420	11.9249
	Deletions	1,319,811	1,753,865	4,655,235	40,229,132
	Insertions	7,040,655	9,211,444	25,099,150	216,151,231
	Substitutions	1,113,866	1,447,112	4,003,822	35,147,508
	Recall (%)	89.6376	89.4974	89.3519	88.3131
	Precision (%)	89.4946	89.3613	89.0615	88.0792
	Trimmed / split reads	8	14	42	706
	Mean missing size (bp)	22.2	20.5	20.1	30.4
	Extended reads	5	14	65	133
	Mean extension size (bp)	22.6	24.6	194.1	23
	Low quality reads	13	29	143	678
	Short reads	0	0	0	0
	Homopolymer ratio	0.9982	1.0000	1.0000	1.0014
	Runtime	6 h 23 min	7 h 47 min	32 h 38 min	199 h 42 min
	Memory (MB)	1,724	1,633	18,771	31,039
HG-CoLoR	Number of bases	65,065,102	84,089,814	219,744,436	1,726,223,265
	Error rate (%)	0.0430	0.0691	0.2959	0.6524
	Deletions	11,561	28,955	339,174	9,986,160
	Insertions	15,803	33,409	548,419	5,156,404
	Substitutions	1,869	3,147	79,127	775,253
	Recall (%)	99.9991	99.9982	99.9900	99.9682
	Precision (%)	99.9574	99.9315	99.7071	99.3554
	Trimmed / split reads	133	498	4,562	71,079
	Mean missing size (bp)	264.7	274.5	677.4	866.3
	Extended reads	5,969	8,180	19,237	140,791
	Mean extension size (bp)	67	71.1	72.5	72.2
	Low quality reads	2	0	99	621
	Short reads	0	4	496	8,849
	Homopolymer ratio	1.0000	1.0000	0.9741	1.0000
	Runtime	51 min	51 min	4 h 55 min	88 h 10 min
	Memory (MB)	1,432	1,517	3,237	19,730
Jabba	Number of bases	68,653,634	87,851,697	216,794,630	1,480,673,578
	Error rate (%)	0.0862	0.0636	0.1058	0.1909
	Deletions	53,665	42,166	112,063	1,865,824
	Insertions	9,913	7,041	95,757	649,951
	Substitutions	1,919	466	21,954	102,639
	Recall (%)	99.9977	99.9975	99.9973	99.9929
	Precision (%)	99.9150	99.9370	99.8950	99.8106
	Trimmed / split reads	639	1,204	6,387	87,670
	Mean missing size (bp)	3,323.5	2,735.3	2,166.6	2,853.8
	Extended reads	8,267	10,402	25,214	168,261
	Mean extension size (bp)	850.4	922.7	1,080.5	1,043.1
	Low quality reads	10	18	115	661
	Short reads	179	386	2,954	34,239
	Homopolymer ratio	1.0000	1.0000	0.9563	1.0000
	Runtime	2 min	2 min	5 min	45 min
	Memory (MB)	1,217	1,218	1,217	13,360
LoRDEC	Number of bases	60,892,408	77,969,503	188,228,237	1,154,508,245
	Error rate (%)	0.0712	0.1474	0.5400	1.2643
	Deletions	48,836	128,344	1,385,989	23,642,857
	Insertions	8,473	21,205	122,252	1,584,355
	Substitutions	6,144	17,896	229,925	1,845,645
	Recall (%)	99.9921	99.9890	99.9483	99.8871
	Precision (%)	99.9313	99.8570	99.4730	98.7542
	Trimmed / split reads	2,951	4,786	22,470	210,051
	Mean missing size (bp)	1,155.2	1,033.9	857.6	1,350.9
	Extended reads	0	0	3	30
	Mean extension size (bp)	0	0	1,285.3	156.9
	Low quality reads	2	0	252	1,966
	Short reads	260	737	22,811	474,725
	Homopolymer ratio	1.0000	1.0000	0.9741	0.9995
	Runtime	6 min	8 min	28 min	6 h 01 min
	Memory (MB)	438	455	799	2,238

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Tool	Metric	Dataset			
		A. baylyi	*E. coli*	*S. cerevisiae*	*C. elegans*
LSC	Number of bases	37,111,831	44,310,080	122,224,269	771,778,062
	Error rate (%)	5.8260	5.2598	6.1549	6.7716
	Deletions	372,086	438,428	1,630,573	14,922,136
	Insertions	1,835,695	1,991,268	6,295,946	40,404,903
	Substitutions	253,915	286,969	895,587	5,874,295
	Recall (%)	94.6923	95.2419	94.6340	94.7734
	Precision (%)	94.2025	94.7706	93.8772	93.2623
	Trimmed / split reads	5,990	7,307	20,257	147,403
	Mean missing size (bp)	3,008.4	3,031.8	2,962.6	2,934.9
	Extended reads	24	85	145	289
	Mean extension size (bp)	22.7	24	24.3	26.1
	Low quality reads	0	0	47	271
	Short reads	599	944	2,453	29,243
	Homopolymer ratio	1.0026	1.0000	0.9833	0.9938
	Runtime	47 min	47 min	9 h 42 min	117 h 34 min
	Memory (MB)	337	328	1,756	1,852
Nanocorr[1]	Number of bases	64,372,749	83,338,435	220,133,609	-
	Error rate (%)	0.4099	0.3043	0.4723	-
	Deletions	72,260	77,573	276,444	-
	Insertions	301,605	261,194	1,023,357	-
	Substitutions	54,832	57,152	258,241	-
	Recall (%)	99.6491	99.7715	99.6679	-
	Precision (%)	99.5928	99.6989	99.5322	-
	Trimmed / split reads	1,574	1,602	5,076	-
	Mean missing size (bp)	322.8	279.4	349.6	-
	Extended reads	0	0	1	-
	Mean extension size (bp)	0	0	20	-
	Low quality reads	1	0	46	-
	Short reads	0	0	0	-
	Homopolymer ratio	1.0000	1.0000	1.0158	-
	Runtime	2 h 52 min	3 h 17 min	39 h 52 min	-
	Memory (MB)	173	166	2,345	-
NaS[2]	Number of bases	58,963,977	78,034,042	207,085,378	-
	Error rate (%)	2.1095	1.3035	2.7483	-
	Deletions	714,139	530,425	2,102,677	-
	Insertions	1,100,737	948,398	5,112,655	-
	Substitutions	94,241	86,401	504,562	-
	Recall (%)	99.9158	99.9468	99.9077	-
	Precision (%)	97.8969	98.7009	97.2615	-
	Trimmed / split reads	339	274	2,187	-
	Mean missing size (bp)	2,361.4	2,370.8	2,437.7	-
	Extended reads	6,688	8,070	22,658	-
	Mean extension size (bp)	1,371.8	1,781.2	1,488.4	-
	Low quality reads	1,651	1,751	5,040	-
	Short reads	0	0	0	-
	Homopolymer ratio	1.0000	1.0000	1.0000	-
	Runtime	24 h 24 min	28 h 48 min	217 h 20 min	-
	Memory (MB)	2,099	2,099	2,099	-
Proovread	Number of bases	62,131,071	80,950,481	210,578,217	1,651,572,671
	Error rate (%)	0.1122	0.1220	0.2075	0.5320
	Deletions	73,995	127,118	670,850	21,888,508
	Insertions	1,364	2,255	26,710	239,748
	Substitutions	1,066	3,344	30,808	464,923
	Recall (%)	99.9944	99.9938	99.9880	99.9599
	Precision (%)	99.8897	99.8808	99.7964	99.4820
	Trimmed / split reads	3,221	4,485	14,211	98,127
	Mean missing size (bp)	299.5	268.1	332.5	599.5
	Extended reads	11	18	45	242
	Mean extension size (bp)	27.3	23.8	257	79.3
	Low quality reads	1	0	124	1,357
	Short reads	861	752	2,834	24,984
	Homopolymer ratio	1.0000	1.0000	0.9669	0.9935
	Runtime	52 min	1 h 18 min	4 h 24 min	79 h 33 min
	Memory (MB)	5,784	6,029	10,435	20,213

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Tool	Metric	Dataset			
		A. baylyi	*E. coli*	*S. cerevisiae*	*C. elegans*
Canu	Number of bases	66,632,351	86,443,218	229,555,492	1,873,188,109
	Error rate (%)	4.9496	5.2422	5.0619	4.9561
	Deletions	738,704	1,011,985	2,574,320	20,233,048
	Insertions	3,517,266	4,804,729	12,252,413	97,517,920
	Substitutions	622,602	841,679	2,197,172	17,594,481
	Recall (%)	95.2430	94.9490	95.1477	95.2663
	Precision (%)	95.0573	94.7647	94.9459	95.0524
	Trimmed / split reads	547	764	2,216	14,407
	Mean missing size (bp)	27.1	27.2	35.1	29.5
	Extended reads	53	66	178	1,462
	Mean extension size (bp)	32.3	27.4	30.7	32.5
	Low quality reads	0	0	43	305
	Short reads	0	0	0	0
	Homopolymer ratio	1.0055	1.0000	0.9937	1.0120
	Runtime	12 min	14 min	32 min	4 h 37 min
	Memory (MB)	2,974	2,821	3,274	6,950
CONSENT	Number of bases	48,390,086	61,487,428	166,101,116	1,359,131,353
	Error rate (%)	8.2534	8.5423	8.2652	9.5548
	Deletions	4,778,369	6,371,753	16,549,340	155,680,250
	Insertions	1,613,389	2,076,177	5,404,368	50,056,903
	Substitutions	114,628	142,283	455,785	6,084,369
	Recall (%)	97.9538	97.9155	98.0349	97.9553
	Precision (%)	91.8579	91.5687	91.8483	90.5794
	Trimmed / split reads	8,332	10,835	28,621	230,249
	Mean missing size (bp)	1,507.1	1,581.7	1,498	1,401
	Extended reads	100	112	267	2,373
	Mean extension size (bp)	133.8	110.3	106.9	92.2
	Low quality reads	0	0	57	471
	Short reads	0	0	0	0
	Homopolymer ratio	1.0070	1.0000	0.9788	0.9765
	Runtime	6 min	8 min	22 min	3 h 49 min
	Memory (MB)	1,020	1,552	4,514	14,522
Daccord[3]	Number of bases	64,669,977	83,773,362	222,050,951	-
	Error rate (%)	0.4694	0.3965	0.5447	-
	Deletions	121,756	75,627	470,101	-
	Insertions	275,390	321,532	977,499	-
	Substitutions	25,146	33,860	200,541	-
	Recall (%)	99.8647	99.8817	99.8591	-
	Precision (%)	99.5371	99.6077	99.4630	-
	Trimmed / split reads	328	119	991	-
	Mean missing size (bp)	445.5	369.9	410.4	-
	Extended reads	0	0	0	-
	Mean extension size (bp)	0	0	0	-
	Low quality reads	0	0	49	-
	Short reads	17	4	54	-
	Homopolymer ratio	1.0061	1.0000	0.9810	-
	Runtime	16 min	24 min	1 h 10 min	-
	Memory (MB)	3,509	4,538	14,111	-
FLAS	Number of bases	35,567,295	44,048,436	125,103,108	662,199,783
	Error rate (%)	2.1958	2.2848	2.4742	9.1804
	Deletions	777,449	879,707	2,963,186	71,270,551
	Insertions	590,885	801,154	2,362,512	40,983,626
	Substitutions	75,331	89,736	322,562	9,060,762
	Recall (%)	99.6847	99.6914	99.6456	99.2995
	Precision (%)	97.8309	97.7424	97.5575	90.8881
	Trimmed / split reads	4,992	6,372	17,066	110,818
	Mean missing size (bp)	2,040.3	2,097.1	2,051.1	1,806.9
	Extended reads	44	78	215	5,956
	Mean extension size (bp)	196.4	343.2	280	198.2
	Low quality reads	1,868	2,591	6,081	85,161
	Short reads	97	149	339	2,725
	Homopolymer ratio	0.9806	1.0000	0.9783	0.9964
	Runtime	2 min	3 min	9 min	1 h 13 min
	Memory (MB)	1,231	1,354	2,259	10,371

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Tool	Metric	Dataset			
		A. baylyi	*E. coli*	*S. cerevisiae*	*C. elegans*
LoRMA	Number of bases	676,120	884,592	6,279,242	11,394,637
	Error rate (%)	0.6305	0.9261	1.6624	5.0838
	Deletions	3,032	5,139	195,481	996,348
	Insertions	1,112	744	21,776	83,683
	Substitutions	1,156	2,136	14,201	110,710
	Recall (%)	99.9766	99.9733	99.8771	99.7216
	Precision (%)	99.3790	99.0871	98.3563	94.9446
	Trimmed / split reads	340	667	2,990	36,351
	Mean missing size (bp)	3,183.5	3,115.5	1,927.7	1,165.9
	Extended reads	0	0	6	24
	Mean extension size (bp)	0	0	1,383.2	195.4
	Low quality reads	14	0	201	2,476
	Short reads	446	1,248	8,683	125,774
	Homopolymer ratio	1.0000	1.0000	1.0000	1.0000
	Runtime	5 min	5 min	20 min	3 h 42 min
	Memory (MB)	32,118	32,183	32,183	32,041
MECAT	Number of bases	46,854,371	58,979,203	162,057,920	870,965,775
	Error rate (%)	0.5340	0.5243	0.6555	0.6540
	Deletions	223,296	267,547	975,507	5,379,870
	Insertions	66,725	82,121	263,724	1,566,201
	Substitutions	5,883	7,575	52,975	139,668
	Recall (%)	99.8289	99.8317	99.8015	99.8196
	Precision (%)	99.4817	99.4915	99.3636	99.3597
	Trimmed / split reads	4,802	6,304	16,034	120,325
	Mean missing size (bp)	2,036.7	2,114.5	2,069.5	2,320.8
	Extended reads	0	0	1	41
	Mean extension size (bp)	0	0	23	354.7
	Low quality reads	0	0	50	583
	Short reads	0	0	0	1
	Homopolymer ratio	1.0008	1.0057	0.9810	0.9938
	Runtime	22 sec	26 sec	1 min 20 sec	18 min 20 sec
	Memory (MB)	1,202	1,322	2,207	10,340

Table 6: Comparison of the different error correction tools, on medium error rate and low coverage simulated datasets. This corresponds to the *A. baylyi* 20x, *E. coli* 20x, *S. cerevisiae* 20x and *C. elegans* 20x datasets of Table 3. [1] Nanocorr was not launched on the *C. elegans* dataset due to its large runtimes. [2] NaS was not launched on the *C. elegans* dataset due to its large runtimes. [3] Daccord could not perform correction on the *C. elegans* dataset, since the alignment step with DALIGNER required more than 128 GB of memory.

3.5.2 Real data

Tool	Remapping		Assembly	
CoLoRMap	Number of reads	25,635	Number of contigs	1
	Number of bases	114,722,711	Number of aligned contigs	1
	Average length (bp)	4,475	Number of breakpoints	40
	Aligned reads (%)	100.0000	NGA50 (bp)	4,657,185
	Average identity (%)	99.5751	NGA75 (bp)	4,657,185
	Genome coverage (%)	100.0000	Genome coverage (%)	98.5239
	Runtime	2 h 01 min		
	Memory (MB)	12,121		
ECTools[1]	Number of reads	1,639	Number of contigs	-
	Number of bases	6,261,214	Number of aligned contigs	-
	Average length (bp)	3,820	Number of breakpoints	-
	Aligned reads (%)	100.0000	NGA50 (bp)	-
	Average identity (%)	98.4789	NGA75 (bp)	-
	Genome coverage (%)	26.9638	Genome coverage (%)	-
	Runtime	12 min		
	Memory (MB)	922		

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Tool	Remapping		Assembly	
FMLRC	Number of reads	22,270	Number of contigs	1
	Number of bases	134,402,291	Number of aligned contigs	1
	Average length (bp)	6,035	Number of breakpoints	3
	Aligned reads (%)	98.2937	NGA50 (bp)	4,642,134
	Average identity (%)	99.9252	NGA75 (bp)	4,642,134
	Genome coverage (%)	100.0000	Genome coverage (%)	100.0000
	Runtime	43 min		
	Memory (MB)	384		
HALC	Number of reads	24,159	Number of contigs	1
	Number of bases	130,674,260	Number of aligned contigs	1
	Average length (bp)	5,408	Number of breakpoints	17
	Aligned reads (%)	100.0000	NGA50 (bp)	4,644,236
	Average identity (%)	99.9273	NGA75 (bp)	4,644,236
	Genome coverage (%)	100.0000	Genome coverage (%)	99.3693
	Runtime	2 h 14 min		
	Memory (MB)	2,237		
Hercules	Number of reads	22,270	Number of contigs	1
	Number of bases	134,641,551	Number of aligned contigs	1
	Average length (bp)	6,045	Number of breakpoints	63
	Aligned reads (%)	95.815	NGA50 (bp)	4,655,108
	Average identity (%)	85.7397	NGA75 (bp)	4,655,108
	Genome coverage (%)	100.0000	Genome coverage (%)	97.9754
	Runtime	10 h 15 min		
	Memory (MB)	2,104		
HG-CoLoR	Number of reads	21,986	Number of contigs	1
	Number of bases	133,956,298	Number of aligned contigs	1
	Average length (bp)	6,092	Number of breakpoints	5
	Aligned reads (%)	100.0000	NGA50 (bp)	4,643,849
	Average identity (%)	99.9589	NGA75 (bp)	4,643,849
	Genome coverage (%)	100.0000	Genome coverage (%)	99.9739
	Runtime	59 min		
	Memory (MB)	1,508		
Jabba	Number of reads	22,086	Number of contigs	47
	Number of bases	128,040,222	Number of aligned contigs	47
	Average length (bp)	5,797	Number of breakpoints	2
	Aligned reads (%)	99.9457	NGA50 (bp)	133,278
	Average identity (%)	99.9687	NGA75 (bp)	61,014
	Genome coverage (%)	99.4309	Genome coverage (%)	95.0742
	Runtime	2 min		
	Memory (MB)	1,220		
LoRDEC	Number of reads	31,728	Number of contigs	1
	Number of bases	125,542,707	Number of aligned contigs	1
	Average length (bp)	3,956	Number of breakpoints	71
	Aligned reads (%)	96.6118	NGA50 (bp)	4,659,557
	Average identity (%)	99.9300	NGA75 (bp)	4,659,557
	Genome coverage (%)	100.0000	Genome coverage (%)	97.7848
	Runtime	13 min		
	Memory (MB)	458		
LSC	Number of reads	16,744	Number of contigs	7
	Number of bases	93,660,098	Number of aligned contigs	7
	Average length (bp)	5,593	Number of breakpoints	42
	Aligned reads (%)	100.0000	NGA50 (bp)	1,214,279
	Average identity (%)	91.1945	NGA75 (bp)	548,948
	Genome coverage (%)	100.0000	Genome coverage (%)	98.6104
	Runtime	1 h 10 min		
	Memory (MB)	1,832		
Nanocorr	Number of reads	21,764	Number of contigs	1
	Number of bases	128,375,708	Number of aligned contigs	1
	Average length (bp)	5,898	Number of breakpoints	33
	Aligned reads (%)	99.9954	NGA50 (bp)	4,662,932
	Average identity (%)	99.1417	NGA75 (bp)	4,662,932
	Genome coverage (%)	100.0000	Genome coverage (%)	99.1316
	Runtime	5 h 48 min		
	Memory (MB)	354		
NaS	Number of reads	21,818	Number of contigs	1
	Number of bases	172,918,739	Number of aligned contigs	1
	Average length (bp)	7,925	Number of breakpoints	6
	Aligned reads (%)	100.0000	NGA50 (bp)	4,641,680
	Average identity (%)	99.9793	NGA75 (bp)	4,641,680
	Genome coverage (%)	100.0000	Genome coverage (%)	99.9674
	Runtime	72 h 02 min		
	Memory (MB)	2,099		

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Tool	Remapping		Assembly	
Proovread	Number of reads	25,338	Number of contigs	1
	Number of bases	125,795,081	Number of aligned contigs	1
	Average length (bp)	4,964	Number of breakpoints	48
	Aligned reads (%)	100.0000	NGA50 (bp)	4,658,520
	Average identity (%)	99.9633	NGA75 (bp)	4,658,520
	Genome coverage (%)	100.0000	Genome coverage (%)	98.4384
	Runtime	2 h 14 min		
	Memory (MB)	7,166		
Canu	Number of reads	17,223	Number of contigs	1
	Number of bases	121,934,133	Number of aligned contigs	1
	Average length (bp)	7,079	Number of breakpoints	4
	Aligned reads (%)	99.9942	NGA50 (bp)	4,457,202
	Average identity (%)	93.8627	NGA75 (bp)	4,457,202
	Genome coverage (%)	100.0000	Genome coverage (%)	99.9802
	Runtime	35 min		
	Memory (MB)	3,264		
CONSENT	Number of reads	18,143	Number of contigs	1
	Number of bases	124,479,635	Number of aligned contigs	1
	Average length (bp)	6,861	Number of breakpoints	14
	Aligned reads (%)	100.0000	NGA50 (bp)	4,519,816
	Average identity (%)	94.2336	NGA75 (bp)	4,519,816
	Genome coverage (%)	100.0000	Genome coverage (%)	99.7372
	Runtime	22 min		
	Memory (MB)	2,239		
Daccord	Number of reads	26,502	Number of contigs	6
	Number of bases	119,130,415	Number of aligned contigs	6
	Average length (bp)	4,495	Number of breakpoints	1
	Aligned reads (%)	99.9170	NGA50 (bp)	1,140,945
	Average identity (%)	96.7690	NGA75 (bp)	995,717
	Genome coverage (%)	100.0000	Genome coverage (%)	99.9572
	Runtime	20 min		
	Memory (MB)	6,515		
FLAS	Number of reads	16,775	Number of contigs	19
	Number of bases	100,467,965	Number of aligned contigs	19
	Average length (bp)	5,989	Number of breakpoints	36
	Aligned reads (%)	99.8808	NGA50 (bp)	307,489
	Average identity (%)	91.4421	NGA75 (bp)	210,170
	Genome coverage (%)	100.0000	Genome coverage (%)	99.3639
	Runtime	8 min		
	Memory (MB)	1,611		
LoRMA[2]	Number of reads	108,980	Number of contigs	-
	Number of bases	17,947,836	Number of aligned contigs	-
	Average length (bp)	164	Number of breakpoints	-
	Aligned reads (%)	99.9128	NGA50 (bp)	-
	Average identity (%)	98.6822	NGA75 (bp)	-
	Genome coverage (%)	25.0530	Genome coverage (%)	-
	Runtime	12 min		
	Memory (MB)	32,117		
MECAT	Number of reads	16,089	Number of contigs	7
	Number of bases	106,757,446	Number of aligned contigs	7
	Average length (bp)	6,635	Number of breakpoints	1
	Aligned reads (%)	100.0000	NGA50 (bp)	843,326
	Average identity (%)	92.2218	NGA75 (bp)	610,826
	Genome coverage (%)	100.0000	Genome coverage (%)	99.9204
	Runtime	4 min		
	Memory (MB)	1,681		

Table 7: Comparison of the different error correction tools, on a medium error rate and low coverage real dataset. This corresponds to the *E. coli* real dataset of Table 3. [1] ECTools corrected reads could not be assembled. [2] LoRMA corrected reads could not be assembled.

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3.6 Medium error rate, medium coverage

Tool	Metric	Dataset			
		A. baylyi	*E. coli*	*S. cerevisiae*	*C. elegans*
CoLoRMap[1]	Number of bases	163,469,531	195,090,745	496,462,088	-
	Error rate	0.2847	0.4841	0.5088	-
	Deletions	381,859	993,426	1,827,202	-
	Insertions	39,322	49,908	202,518	-
	Substitutions	227,605	251,725	1,023,892	-
	Recall	99.9831	99.9712	99.9679	-
	Precision	99.7247	99.5320	99.5053	-
	Trimmed / split reads	10,325	12,378	37,316	-
	Taille manquante moyenne	1,897.5	2,280.1	2,385.6	-
	Extended reads	22,578	27,910	69,906	-
	Taille moyenne d'extension	212.9	251.0	196.8	-
	Low quality reads	28	36	282	-
	Short reads	10,357	10,861	27,833	-
	Ratio taille homopolymers	0.9913	1.0000	1.0042	-
	Runtime	2 h 52 min	3 h 32 min	10 h 51 min	-
	Memory (MB)	13.132	15.990	18.972	-
FMLRC	Number of bases	195,182,619	252,027,858	674,591,322	5,526,058,984
	Error rate	0..005	0.2741	1.0792	3.4246
	Deletions	145,662	228,943	2,146,125	65,413,791
	Insertions	502,754	853,350	7,457,074	163,114,555
	Substitutions	130,122	210,729	1,885,718	53,017,848
	Recall	99.8400	99.7630	99.1029	97.8036
	Precision	99.8006	99.7269	98.9245	96.5988
	Trimmed / split reads	63	84	308	9,214
	Taille manquante moyenne	33.7	14.9	31.8	32.6
	Extended reads	4	6	18	984
	Taille moyenne d'extension	34.0	55.7	176.4	30.2
	Low quality reads	52	63	340	2,315
	Short reads	0	0	0	0
	Ratio taille homopolymers	1.0000	1.0000	1.0032	0.7904
	Runtime	1 h 07 min	1 h 25 min	3 h 52 min	23 h 36 min
	Memory (MB)	409	428	895	7,938
HALC	Number of bases	189,974,036	240,901,559	632,932,482	4,729,802,695
	Error rate	0.1175	0.1468	0.4350	1.5007
	Deletions	214,063	309,688	3,248,553	107,463,874
	Insertions	29,571	43,596	310,542	6,493,145
	Substitutions	17,315	42,546	594,421	8,320,720
	Recall	99.9918	99.9918	99.9714	99.8916
	Precision	99.8850	99.8564	99.5740	98.5250
	Trimmed / split reads	7,183	12,042	37,380	465,219
	Taille manquante moyenne	598.4	758.9	729.0	864.2
	Extended reads	61	55	218	1,956
	Taille moyenne d'extension	31.3	30.5	65.5	42.9
	Low quality reads	5	4	499	7,243
	Short reads	319	821	10,890	516,489
	Ratio taille homopolymers	1.0000	1.0000	0.9993	1.0000
	Runtime	5 h 16 min	3 h 05 min	4 h 14 min	18 h 12 min
	Memory (MB)	3,146	2,688	2,409	5,578
HG-CoLoR[2]	Number of bases	196,441,510	252,224,483	654,754,845	-
	Error rate	0.0447	0.0820	0.2949	-
	Deletions	43,219	99,355	1,048,431	-
	Insertions	45,639	123,916	1,485,531	-
	Substitutions	6,432	14,387	274,343	-
	Recall	99.9989	99.9984	99.9908	-
	Precision	99.9559	99.9188	99.7082	-
	Trimmed / split reads	734	2,587	15,953	-
	Taille manquante moyenne	575.7	734.0	1,108.5	-
	Extended reads	18,788	250,60	59,420	-
	Taille moyenne d'extension	103.5	106.8	110.0	-
	Low quality reads	0	1	271	-
	Short reads	12	14	1,230	-
	Ratio taille homopolymers	1.0000	1.0000	1.0000	-
	Runtime	1 h 33 min	1 h 39 min	10 h 40 min	-
	Memory (MB)	2,130	2,747	7,316	-

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Tool	Metric	Dataset			
		A. baylyi	*E. coli*	*S. cerevisiae*	*C. elegans*
Jabba	Number of bases	206,618,316	262,664,049	651,032,022	-
	Error rate	0.0874	0.0594	0.0922	-
	Deletions	152,141	106,718	266,144	-
	Insertions	31,500	28,159	274,263	-
	Substitutions	4,467	1,003	58,593	-
	Recall	99.9980	99.9988	99.9976	-
	Precision	99.9136	99.9411	99.9085	-
	Trimmed / split reads	1,917	3,748	19,668	-
	Taille manquante moyenne	3,116.2	2,776.5	2,224.5	-
	Extended reads	24,669	31,072	75,271	-
	Taille moyenne d'extension	836.8	896.4	1,079.2	-
	Low quality reads	37	52	354	-
	Short reads	578	1,142	9,443	-
	Ratio taille homopolymers	1.0000	1.0000	1.0000	-
	Runtime	2 min	2 min	5 min	-
	Memory (MB)	1,215	1,220	1,217	-
LoRDEC	Number of bases	181,008,925	231,286,209	562,268,349	3,486,299,514
	Error rate	0.0625	0.1423	0.5208	1.2020
	Deletions	114,292	392,797	4,294,506	71,409,761
	Insertions	22,578	57,835	360,338	4,710,829
	Substitutions	15,740	49,721	669,770	5,452,250
	Recall	99.9934	99.9887	99.9487	99.8970
	Precision	99.9398	99.8620	99.4919	98.8149
	Trimmed / split reads	9,286	14,599	68,952	687,330
	Taille manquante moyenne	1,265.7	1,124.1	839.9	1,159.5
	Extended reads	1	2	8	88
	Taille moyenne d'extension	60.0	25.0	879.9	162.7
	Low quality reads	52	62	879	7,235
	Short reads	3,695	6,479	175,136	2,415,613
	Ratio taille homopolymers	1.0000	1.0000	0.9988	0.8843
	Runtime	13 min	24 min	1 h 17 min	17 h 24 min
	Memory (MB)	430	458	797	2,320
Proovread[3]	Number of bases	185,254,462	238,343,371	582,618,950	-
	Error rate	0.2382	0.2626	0.3130	-
	Deletions	507,394	884,818	2,576,850	-
	Insertions	2,527	4,468	61,534	-
	Substitutions	5,967	19,282	113,279	-
	Recall	99.9928	99.9893	99.9871	-
	Precision	99.7656	99.7434	99.6923	-
	Trimmed / split reads	14,741	22,793	70,024	-
	Taille manquante moyenne	399.2	388.9	567.7	-
	Extended reads	12	21	26	-
	Taille moyenne d'extension	25n1	26.9	505.2	-
	Low quality reads	3	0	415	-
	Short reads	1,324	1,403	15,113	-
	Ratio taille homopolymers	0.9565	1.0000	1.0006	-
	Runtime	3 h 30 min	5 h 21 min	17 h 38 min	-
	Memory (MB)	14,475	14,306	20,100	-
Canu	Number of bases	113,074,830	84,731,584	233,891,897	2,018,823,460
	Error rate	0.6196	0.5674	0.6847	0.8828
	Deletions	315,662	181,791	738,971	11,392,385
	Insertions	395,443	275,110	896,245	9,151,300
	Substitutions	25,626	15,970	74,696	605,305
	Recall	99.6344	99.6564	99.5995	99.5294
	Precision	99.3873	99.4363	99.3205	99.1259
	Trimmed / split reads	14,624	15,448	41,826	349,404
	Taille manquante moyenne	2,656.7	2,409.7	2,381.8	2,429.7
	Extended reads	82	69	181	1,778
	Taille moyenne d'extension	26n1	27.7	53.7	32.7
	Low quality reads	1	0	104	1,043
	Short reads	362	737	1,817	15,036
	Ratio taille homopolymers	1.0182	1.0077	0.9773	0.9885
	Runtime	22 min	15 min	41 min	5 h 37 min
	Memory (MB)	4,628	3,540	3,650	7,052

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Tool	Metric	Dataset			
		A. baylyi	*E. coli*	*S. cerevisiae*	*C. elegans*
CONSENT	Number of bases	157,025,017	201,954,943	539,862,948	4,393,920,843
	Error rate	1.2774	1.2531	1.4309	2.1802
	Deletions	1,747,520	2,142,584	7,280,698	125,609,247
	Insertions	1,066,354	1,386,428	3,826,520	35,808,412
	Substitutions	86,763	110,746	392,819	4,833,713
	Recall	99.6019	99.5955	99.5868	99.5042
	Precision	98.7452	98.7682	98.5942	97.8579
	Trimmed / split reads	23,418	30,379	80,740	650,917
	Taille manquante moyenne	1,325.3	1,351.3	1,315.4	1,243.0
	Extended reads	1	1	2	149
	Taille moyenne d'extension	28.0	26.0	31.5	113.0
	Low quality reads	0	1	213	4,900
	Short reads	3,239	4,002	11,199	105,744
	Ratio taille homopolymers	1.0000	1.0000	1.0029	0.9837
	Runtime	23 min	30 min	1 h 24 min	14 h 22 min
	Memory (MB)	4,347	4,808	11,088	17,388
Daccord[4]	Number of bases	194,674,108	251,327,413	235,508,504	-
	Error rate	0.0484	0.0283	0.4190	-
	Deletions	89,549	15,689	1,288,109	-
	Insertions	59,010	50,132	550,838	-
	Substitutions	11,248	21,053	83,275	-
	Recall	99.9892	99.9930	99.9177	-
	Precision	99.9529	99.9723	99.5929	-
	Trimmed / split reads	341	117	3,081	-
	Taille manquante moyenne	302.5	130.0	1,377.0	-
	Extended reads	0	0	1	-
	Taille moyenne d'extension	0.0	0.0	22.0	-
	Low quality reads	0	0	48	-
	Short reads	7	2	315	-
	Ratio taille homopolymers	1.0000	1.0000	1.0011	-
	Runtime	36 min	49 min	47 min	-
	Memory (MB)	11,267	14,398	31,990	-
FLAS	Number of bases	164,962,117	209,893,136	555,375,307	3,601,229,111
	Error rate	0.8227	0.7795	0.9486	2.3761
	Deletions	1,358,586	1,561,943	5,346,704	86,057,509
	Insertions	725,577	915,376	2,921,977	78,458,765
	Substitutions	93,550	111,358	396,177	11,864,157
	Recall	99.7894	99.8018	99.7709	99.7365
	Precision	99.2001	99.2422	99.0760	97.6522
	Trimmed / split reads	10,326	13,544	36,178	357,506
	Taille manquante moyenne	1,854.2	1,887.9	1,868.7	1,983.0
	Extended reads	35	41	174	8,944
	Taille moyenne d'extension	10,047.3	228.0	10,051.8	199.2
	Low quality reads	917	1,315	3,795	95,284
	Short reads	64	94	263	4,899
	Ratio taille homopolymers	1.0000	1.0000	0.9951	1.0006
	Runtime	19 min	24 min	1 h 01 min	7 h 03 min
	Memory (MB)	2,081	2,443	5,056	11,429
LoRMA	Number of bases	30,907,458	38,839,669	108,376,053	302,853,378
	Error rate	0.2369	0.2552	0.4330	1.0741
	Deletions	69,492	82,841	533,434	4,198,185
	Insertions	6,059	8,965	65,060	314,039
	Substitutions	5,371	16,286	71,794	485,037
	Recall	99.9852	99.9847	99.9689	99.9391
	Precision	99.7666	99.7487	99.5720	98.9343
	Trimmed / split reads	29,426	37,783	103,451	740,225
	Taille manquante moyenne	1,841.2	1,781.0	1,718.8	1,030.7
	Extended reads	0	0	6	58
	Taille moyenne d'extension	0.0	0.0	821.0	259.6
	Low quality reads	36	10	608	5,170
	Short reads	166,606	213,126	593,186	4,223,980
	Ratio taille homopolymers	1.0000	1.0000	1.0000	1.0000
	Runtime	17 min	25 min	1 h 56 min	20 h 48 min
	Memory (MB)	31,976	31,818	31,687	31,884

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Tool	Metric	Dataset			
		A. baylyi	*E. coli*	*S. cerevisiae*	*C. elegans*
MECAT	Number of bases	97,093,166	121,976,082	322,237,718	1,283,874,393
	Error rate	0.4936	0.4693	0.5463	0.5107
	Deletions	427,238	515,933	1,585,591	5,839,768
	Insertions	131,467	153,655	480,274	1,918,807
	Substitutions	12,353	14,170	52,759	125,770
	Recall	99.8166	99.8276	99.8031	99.8437
	Precision	99.5224	99.5460	99.4707	99.5008
	Trimmed / split reads	9,968	12,617	32,642	151,297
	Taille manquante moyenne	1,617.3	1,617.1	1,649.4	1,670.8
	Extended reads	0	0	3	19
	Taille moyenne d'extension	0.0	0.0	2,855.7	116.2
	Low quality reads	0	0	79	285
	Short reads	0	0	0	0
	Ratio taille homopolymers	0.9942	1.0000	0.9972	0.9937
	Runtime	2 min	3 min	9 min	1 h 37 min
	Memory (MB)	2,031	2,391	4,991	10,942

Table 8: Comparison of the different error correction tools, on medium error rate and medium coverage datasets. This corresponds to the *A. baylyi* 60x (medium), *E. coli* 60x (medium), *S. cerevisiae* 60x (medium) and *C. elegans* 60x (medium) datasets of Table 3. [1] Colormap was not launched on the *C. elegans* dataset due to its large runtimes. [2] HG-CoLoR was not launched on the *C. elegans* dataset due to its large runtimes. [3] Proovread was not launched on the *C. elegans* dataset due to its large runtimes. [4] Daccord could not perform correction on the *C. elegans* dataset, since the alignment step with DALIGNER required more than 128 GB of memory.

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3.7 Low error rate, low coverage

For these experiments, we exclude the following tools from the comparison, for the reasons mentioned below:

- ECTools, because of its unsatisfying results on previous experiments;
- Hercules, because of its unsatisfying results and large runtimes on previous experiments;
- LSC, because of its unsatisfying results on previous experiments;
- Nanocorr, because of its large runtimes on previous experiments;
- NaS, because of its large runtimes on previous experiments;

Tool	Metric	Dataset		
		E. coli	*S. cerevisiae*	*C. elegans*
CoLoRMap	Number of bases	134,398,575	343,101,257	1,197,628,902
	Error rate (%)	0.1137	0.3183	0.8955
	Deletions	14,929	346,678	1,900,447
	Insertions	36,705	291,959	1,036,546
	Substitutions	92,329	449,592	7,663,104
	Recall (%)	99.9881	99.9135	99.9165
	Precision (%)	99.8880	99.6860	99.1230
	Trimmed / split reads	31	845	8,587
	Mean missing size (bp)	3,729.9	4,242.4	5,180.3
	Extended reads	16,644	42,853	168,676
	Mean extension size (bp)	305.5	305.5	252.3
	Low quality reads	13	65	208
	Short reads	269	1,606	194,369
	Homopolymer ratio	1.0000	1.0071	0.9967
	Runtime	1 h 33 min	4 h 36 min	150 h 21 min
	Memory (MB)	13,097	14,243	32,267
FMLRC	Number of bases	131,066,466	348,205,441	2,820,835,418
	Error rate (%)	0.0320	0.2447	1.4161
	Deletions	11,502	273,645	15,775,123
	Insertions	16,155	427,404	16,783,150
	Substitutions	8,752	196,210	10,932,609
	Recall (%)	99.9919	99.9032	99.6795
	Precision (%)	99.9686	99.7579	98.6018
	Trimmed / split reads	2	44	2,090
	Mean missing size (bp)	17	66	28.4
	Extended reads	0	3	106
	Mean extension size (bp)	0	0	31.3
	Low quality reads	41	151	1,121
	Short reads	0	0	0
	Homopolymer ratio	1.0000	1.0000	0.9915
	Runtime	45 min	1 h 59 min	11 h 55 min
	Memory (MB)	400	892	7,937
HALC	Number of bases	130,839,109	347,722,386	2,819,386,712
	Error rate (%)	0.1565	0.3611	1.0897
	Deletions	121,296	444,792	9,586,042
	Insertions	40,749	403,321	12,999,176
	Substitutions	23,408	405,608	9,627,633
	Recall (%)	99.9799	99.9387	99.7543
	Precision (%)	99.8466	99.6455	98.9252
	Trimmed / split reads	1,013	2,812	31,158
	Mean missing size (bp)	140.8	151.8	101.8
	Extended reads	6	47	599
	Mean extension size (bp)	28.8	28.8	40.2
	Low quality reads	41	136	1,195
	Short reads	0	1	2
	Homopolymer ratio	1.0000	1.0000	1.0000
	Runtime	1 h 17 min	1 h 53 min	9 h 30 min
	Memory (MB)	1,714	1,892	2,853

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Tool	Metric	Dataset		
		E. coli	*S. cerevisiae*	*C. elegans*
HG-CoLoR	Number of bases	131,346,207	346,708,590	2,794,826,522
	Error rate (%)	0.0726	0.5115	1.1664
	Deletions	39,908	1,342,278	26,965,036
	Insertions	49,798	550,820	9,324,074
	Substitutions	5,741	162,266	1,509,055
	Recall (%)	99.9986	99.9592	99.9104
	Precision (%)	99.9279	99.4937	98.8449
	Trimmed / split reads	626	4,141	59,301
	Mean missing size (bp)	178.9	331.7	214.8
	Extended reads	8,293	16,823	130,681
	Mean extension size (bp)	30.3	30.3	29.4
	Low quality reads	0	60	435
	Short reads	0	7	2
	Homopolymer ratio	1.0000	1.0012	1.0000
	Runtime	1 h 20 min	7 h 20 min	108 h 26 min
	Memory (MB)	1,538	3,656	27,212
Jabba	Number of bases	133,959,455	340,101,149	2,463,694,280
	Error rate (%)	0.0771	0.1067	0.2319
	Deletions	80,450	209,683	4,332,786
	Insertions	15,541	155,778	1,242,983
	Substitutions	573	33,825	236,640
	Recall (%)	99.9981	99.9975	99.9913
	Precision (%)	99.9237	99.8941	99.7702
	Trimmed / split reads	2,037	10,781	148,923
	Mean missing size (bp)	2,601.5	1,992.5	2,805.6
	Extended reads	15,416	37,196	247,049
	Mean extension size (bp)	638.6	638.6	947.3
	Low quality reads	2	77	367
	Short reads	874	5,946	69,905
	Homopolymer ratio	1.0000	1.0000	1.0000
	Runtime	2 min	5 min	43 min
	Memory (MB)	1,220	1,215	13,362
LoRDEC	Number of bases	131,052,792	348,420,743	2,823,537,319
	Error rate (%)	0.0695	0.3990	1.2710
	Deletions	32,861	297,482	8,411,355
	Insertions	35,620	694,323	19,478,144
	Substitutions	24,234	480,342	10,963,740
	Recall (%)	99.9831	99.8123	99.4191
	Precision (%)	99.9328	99.6093	98.7441
	Trimmed / split reads	181	2,354	56,726
	Mean missing size (bp)	63.7	79.7	126.2
	Extended reads	2	2	149
	Mean extension size (bp)	25.5	25.5	38.9
	Low quality reads	22	97	558
	Short reads	0	0	3
	Homopolymer ratio	1.0000	1.0000	0.9979
	Runtime	12 min	35 min	11 h 30 min
	Memory (MB)	460	799	2,320
Proovread	Number of bases	130,360,408	342,457,860	2,703,725,143
	Error rate (%)	0.1615	0.2365	0.4325
	Deletions	169,197	845,704	20,509,633
	Insertions	1,309	30,583	189,378
	Substitutions	6,636	43,845	654,789
	Recall (%)	99.9953	99.9912	99.9711
	Precision (%)	99.8409	99.7668	99.5738
	Trimmed / split reads	8,571	29,179	153,272
	Mean missing size (bp)	44.1	85.6	227.3
	Extended reads	0	3	45
	Mean extension size (bp)	0	0	87.9
	Low quality reads	0	135	1,899
	Short reads	29	1,202	15,249
	Homopolymer ratio	1.0000	1.0000	1.0000
	Runtime	1 h 59 min	5 h 37 min	85 h 23 min
	Memory (MB)	8,368	16,777	29,934

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Tool	Metric	Dataset		
		E. coli	*S. cerevisiae*	*C. elegans*
Canu	Number of bases	129,533,580	226,459,133	2,773,456,245
	Error rate (%)	0.4156	1.1052	0.5008
	Deletions	247,325	803,320	6,013,667
	Insertions	237,224	1,371,782	6,130,158
	Substitutions	19,506	145,574	517,864
	Recall (%)	99.7647	99.1766	99.7103
	Precision (%)	99.5887	98.9036	99.5040
	Trimmed / split reads	9,422	33,164	170,416
	Mean missing size (bp)	128.9	1,762.6	126.5
	Extended reads	55	51	1,100
	Mean extension size (bp)	26.7	28.5	28.1
	Low quality reads	0	41	393
	Short reads	3	509	147
	Homopolymer ratio	1.0116	1.0040	0.9990
	Runtime	19 min	29 min	9 h 09 min
	Memory (MB)	4,613	3,681	6,921
CONSENT	Number of bases	129,647,002	344,457,307	2,789,537,501
	Error rate (%)	0.3142	0.4091	0.7902
	Deletions	254,833	853,156	12,522,232
	Insertions	134,538	456,318	8,794,630
	Substitutions	18,772	112,883	2,296,195
	Recall (%)	99.9430	99.9321	99.8773
	Precision (%)	99.6906	99.5971	99.2204
	Trimmed / split reads	14,681	38,872	309,728
	Mean missing size (bp)	86.7	83.8	80.8
	Extended reads	0	5	107
	Mean extension size (bp)	0	0	83.1
	Low quality reads	0	41	424
	Short reads	0	0	0
	Homopolymer ratio	1.0000	0.9967	1.0077
	Runtime	17 min	46 min	9 h 36 min
	Memory (MB)	2,390	5,523	21,819
Daccord[1]	Number of bases	131,020,333	347,992,121	-
	Error rate (%)	0.0248	0.1259	-
	Deletions	4,326	73,057	-
	Insertions	13,333	123,760	-
	Substitutions	15,965	265,120	-
	Recall (%)	99.9965	99.9874	-
	Precision (%)	99.9757	99.8762	-
	Trimmed / split reads	5	102	-
	Mean missing size (bp)	478.8	188.5	-
	Extended reads	0	5	-
	Mean extension size (bp)	0	0	-
	Low quality reads	0	47	-
	Short reads	0	21	-
	Homopolymer ratio	1.0000	1.0000	-
	Runtime	14 min	1 h 19 min	-
	Memory (MB)	6,813	31,798	-
FLAS	Number of bases	129,692,582	344,252,752	2,728,691,582
	Error rate (%)	0.2720	0.3272	0.7613
	Deletions	308,007	967,396	22,672,322
	Insertions	92,042	273,904	7,047,712
	Substitutions	6,952	35,833	1,451,039
	Recall (%)	99.9291	99.9131	99.8613
	Precision (%)	99.7385	99.6843	99.2541
	Trimmed / split reads	1,900	5,082	72,527
	Mean missing size (bp)	434	438.3	581.6
	Extended reads	0	1	578
	Mean extension size (bp)	0	0	161.2
	Low quality reads	5	58	7,480
	Short reads	0	0	24
	Homopolymer ratio	1.0076	0.9972	1.0036
	Runtime	12 min	29 min	3 h 07 min
	Memory (MB)	1,639	2,935	10,565

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Tool	Metric	Dataset		
		E. coli	*S. cerevisiae*	*C. elegans*
LoRMA	Number of bases	2,378,146	14,071,752	32,644,556
	Error rate (%)	1.1962	2.1640	3.6960
	Deletions	27,701	474,954	2,058,432
	Insertions	1,631	40,421	123,001
	Substitutions	6,271	35,339	182,811
	Recall (%)	99.9209	99.8351	99.7269
	Precision (%)	98.8208	97.8564	96.3449
	Trimmed / split reads	11,737	35,483	227,760
	Mean missing size (bp)	225.6	179.1	158.1
	Extended reads	0	3	16
	Mean extension size (bp)	0	0	492.7
	Low quality reads	7	485	2,758
	Short reads	63,731	199,074	1,163,973
	Homopolymer ratio	1.0000	1.0000	1.0000
	Runtime	10 min	46 min	8 h 19 min
	Memory (MB)	32,155	31,899	31,827
MECAT	Number of bases	106,842,493	284,618,017	2,083,992,906
	Error rate (%)	0.2569	0.3040	0.3908
	Deletions	238,111	738,020	7,017,472
	Insertions	46,384	154,073	1,559,772
	Substitutions	2,525	16,281	93,949
	Recall (%)	99.9302	99.9160	99.8903
	Precision (%)	99.7533	99.7072	99.6212
	Trimmed / split reads	6,625	16,957	148,957
	Mean missing size (bp)	1,261	1,229.9	1,468.4
	Extended reads	0	0	9
	Mean extension size (bp)	0	0	598.1
	Low quality reads	0	44	188
	Short reads	0	0	0
	Homopolymer ratio	1.0104	1.0043	0.9966
	Runtime	2 min	5 min	48 min
	Memory (MB)	1,600	2,907	10,535

Table 9: Comparison of the different error correction tools, on low error rate and low coverage datasets. This corresponds to the *E. coli* 30x, *S. cerevisiae* 30x and *C. elegans* 30x datasets of Table 3. [1] Daccord could not perform correction, since the alignment step with DALIGNER required more than 128 GB of memory.

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3.8 Low error rate, medium coverage

- ECTools, because of its unsatisfying results on previous experiments;
- Hercules, because of its unsatisfying results and large runtimes on previous experiments;
- LSC, because of its unsatisfying results on previous experiments;
- Nanocorr, because of its large runtimes on previous experiments;
- NaS, because of its large runtimes on previous experiments;

Tool	Metric	Dataset		
		E. coli	*S. cerevisiae*	*C. elegans*
CoLoRMap[1]	Number of bases	266,210,819	663,617,071	-
	Error rate (%)	0.1621	0.6143	-
	Deletions	98,293	1,930,344	-
	Insertions	112,511	1,216,405	-
	Substitutions	199,934	1,036,618	-
	Recall (%)	99.9631	99.7755	-
	Precision (%)	99.8400	99.3917	-
	Trimmed / split reads	185	3,913	-
	Mean missing size (bp)	3,787.2	3,746.2	-
	Extended reads	33,047	82,438	-
	Mean extension size (bp)	297.7	297.7	-
	Low quality reads	33	145	-
	Short reads	653	4,679	-
	Homopolymer ratio	1.0089	0.9946	-
	Runtime	3 h 01 min	8 h 08 min	-
	Memory (MB)	19,898	24,375	-
FMLRC	Number of bases	261,387,632	695,239,831	5,652,356,620
	Error rate (%)	0.0292	0.2469	1.4213
	Deletions	24,753	551,321	31,671,968
	Insertions	31,069	872,824	33,701,759
	Substitutions	17,368	391,349	21,909,721
	Recall (%)	99.9944	99.9006	99.6764
	Precision (%)	99.9714	99.7557	98.5966
	Trimmed / split reads	14	106	4,147
	Mean missing size (bp)	17.7	48.5	28.9
	Extended reads	0	8	242
	Mean extension size (bp)	0	0	28.1
	Low quality reads	85	360	2,315
	Short reads	0	0	0
	Homopolymer ratio	1.0000	1.0000	0.9856
	Runtime	1 h 28 min	3 h 57 min	23 h 25 min
	Memory (MB)	403	893	7,937
HALC	Number of bases	260,933,848	694,333,491	5,649,091,260
	Error rate (%)	0.1522	0.3648	1.0880
	Deletions	233,633	883,877	19,179,388
	Insertions	84,745	822,236	25,917,654
	Substitutions	48,696	827,403	19,226,301
	Recall (%)	99.9785	99.9354	99.7550
	Precision (%)	99.8509	99.6420	98.9269
	Trimmed / split reads	2,033	5,707	61,894
	Mean missing size (bp)	147.3	131.6	102
	Extended reads	11	60	1,284
	Mean extension size (bp)	22.4	22.4	42.6
	Low quality reads	79	329	2,512
	Short reads	0	1	0
	Homopolymer ratio	1.0000	1.0000	0.9852
	Runtime	4 h 57 min	4 h 25 min	19 h 10 min
	Memory (MB)	2,747	2,487	5,716

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Tool	Metric	Dataset		
		E. coli	*S. cerevisiae*	*C. elegans*
HG-CoLoR[2]	Number of bases	261,879,085	690,178,663	-
	Error rate (%)	0.0771	0.5995	-
	Deletions	87,607	2,824,556	-
	Insertions	113,291	1,924,555	-
	Substitutions	14,055	374,714	-
	Recall (%)	99.9987	99.9433	-
	Precision (%)	99.9234	99.4059	-
	Trimmed / split reads	1,569	9,700	-
	Mean missing size (bp)	264.1	496.2	-
	Extended reads	19,498	40,638	-
	Mean extension size (bp)	33.2	33.2	-
	Low quality reads	1	235	-
	Short reads	0	45	-
	Homopolymer ratio	1.0000	1.0000	-
	Runtime	2 h 03 min	12 h 23 min	-
	Memory (MB)	2,744	7,297	-
Jabba	Number of bases	267,665,882	678,936,284	4,934,623,442
	Error rate (%)	0.0716	0.1040	0.2312
	Deletions	146,631	387,538	8,496,067
	Insertions	30,775	340,301	2,534,990
	Substitutions	1,171	63,194	419,150
	Recall (%)	99.9984	99.9975	99.9916
	Precision (%)	99.9292	99.8967	99.7710
	Trimmed / split reads	4,095	21,726	298,954
	Mean missing size (bp)	2,507.2	1,996	2,804.2
	Extended reads	30,786	74,343	493,302
	Mean extension size (bp)	633.2	633.2	949.5
	Low quality reads	1	183	731
	Short reads	1,649	11,922	140,889
	Homopolymer ratio	1.0000	1.0000	1.0000
	Runtime	2 min	5 min	49 min
	Memory (MB)	1,220	1,215	13,360
LoRDEC	Number of bases	261,360,235	695,578,111	5,657,161,182
	Error rate (%)	0.0684	0.3984	1.2731
	Deletions	65,549	613,150	16,882,129
	Insertions	70,097	1,389,483	38,979,226
	Substitutions	47,336	974,285	21,920,160
	Recall (%)	99.9832	99.8136	99.4201
	Precision (%)	99.9339	99.6100	98.7420
	Trimmed / split reads	393	4,657	113,843
	Mean missing size (bp)	65.3	78.9	126.3
	Extended reads	0	2	273
	Mean extension size (bp)	0	0	48
	Low quality reads	35	214	1,263
	Short reads	0	1	11
	Homopolymer ratio	1.0000	1.0000	1.0160
	Runtime	20 min	1 h 09 min	23 h 30 min
	Memory (MB)	457	794	2,332
Proovread[3]	Number of bases	259,647,167	671,307,477	-
	Error rate (%)	0.1689	0.2568	-
	Deletions	358,693	2,080,733	-
	Insertions	2,837	56,965	-
	Substitutions	15,499	102,130	-
	Recall (%)	99.9948	99.9902	-
	Precision (%)	99.8335	99.7467	-
	Trimmed / split reads	18,693	68,598	-
	Mean missing size (bp)	54.3	148.2	-
	Extended reads	4	10	-
	Mean extension size (bp)	20.2	20.2	-
	Low quality reads	0	501	-
	Short reads	172	7,658	-
	Homopolymer ratio	1.0000	1.0000	-
	Runtime	4 h 07 min	11 h 51 min	-
	Memory (MB)	15,245	23,591	-

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Tool	Metric	Dataset		
		E. coli	*S. cerevisiae*	*C. elegans*
Canu	Number of bases	218,652,792	599,325,043	5,112,430,638
	Error rate (%)	0.7404	0.7919	0.7934
	Deletions	563,365	1,678,564	14,729,573
	Insertions	827,596	2,412,967	20,996,517
	Substitutions	72,521	240,442	1,897,014
	Recall (%)	99.4781	99.4488	99.4573
	Precision (%)	99.2658	99.2148	99.2131
	Trimmed / split reads	27,690	73,001	569,082
	Mean missing size (bp)	985.8	858.1	651.3
	Extended reads	47	176	1,658
	Mean extension size (bp)	29.0	53.7	37.2
	Low quality reads	0	104	941
	Short reads	187	354	1,845
	Homopolymer ratio	1.0158	1.0108	1.0065
	Runtime	24 min	1 h 11 min	9 h 30 min
	Memory (MB)	3,674	3,710	7,050
CONSENT	Number of bases	259,078,481	689,502,352	5,603,760,432
	Error rate (%)	0.1722	0.2918	0.5730
	Deletions	341,549	1,280,545	18,391,216
	Insertions	55,787	582,869	12,737,445
	Substitutions	8,354	115,614	2,826,988
	Recall (%)	99.9843	99.9481	99.9119
	Precision (%)	99.8306	99.7125	99.4345
	Trimmed / split reads	27,380	71,027	571,107
	Mean missing size (bp)	71.8	68.1	67.1
	Extended reads	0	2	187
	Mean extension size (bp)	0	0	122.2
	Low quality reads	0	107	849
	Short reads	0	0	0
	Homopolymer ratio	1.0000	1.0000	0.9653
	Runtime	36 min	1 h 46 min	27 h 04 min
	Memory (MB)	4,849	11,325	32,284
Daccord[4]	Number of bases	261,274,775	694,845,174	-
	Error rate (%)	0.0214	0.0400	-
	Deletions	6,353	51,217	-
	Insertions	19,335	132,765	-
	Substitutions	28,842	75,540	-
	Recall (%)	99.9971	99.9928	-
	Precision (%)	99.9790	99.9606	-
	Trimmed / split reads	13	73	-
	Mean missing size (bp)	58.8	276.8	-
	Extended reads	0	4	-
	Mean extension size (bp)	0	0	-
	Low quality reads	0	113	-
	Short reads	0	0	-
	Homopolymer ratio	1.0000	1.0000	-
	Runtime	54 min	2 h 26 min	-
	Memory (MB)	18,450	32,190	-
FLAS	Number of bases	260,324,873	689,491,882	5,583,925,852
	Error rate (%)	0.1547	0.2034	0.3997
	Deletions	378,728	1,273,157	25,708,666
	Insertions	53,059	220,507	5,628,454
	Substitutions	3,141	30,665	1,249,658
	Recall (%)	99.9546	99.9418	99.9175
	Precision (%)	99.8526	99.8049	99.6104
	Trimmed / split reads	403	1,274	30,950
	Mean missing size (bp)	182.4	223.4	305.4
	Extended reads	1	5	588
	Mean extension size (bp)	102	102	156.7
	Low quality reads	4	241	7,506
	Short reads	0	0	14
	Homopolymer ratio	0.9967	1.0000	0.9890
	Runtime	38 min	1 h 30 min	10 h 45 min
	Memory (MB)	2,428	4,984	13,682

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Tool	Metric	Dataset		
		E. coli	*S. cerevisiae*	*C. elegans*
LoRMA	Number of bases	170,581,660	442,516,834	780,732,399
	Error rate (%)	0.1285	0.2225	0.6446
	Deletions	163,750	1,197,979	5,515,626
	Insertions	25,117	129,027	369,539
	Substitutions	21,604	106,535	581,292
	Recall (%)	99.9865	99.9785	99.9547
	Precision (%)	99.8743	99.7812	99.3649
	Trimmed / split reads	40,347	108,516	1,154,643
	Mean missing size (bp)	1,007	970.3	412.3
	Extended reads	0	4	13
	Mean extension size (bp)	0	0	685.7
	Low quality reads	4	797	3,575
	Short reads	187,376	522,390	9,256,368
	Homopolymer ratio	1.0000	1.0000	1.0000
	Runtime	1 h 39 min	5 h 25 min	31 h 04 min
	Memory (MB)	31,682	31,828	32,104
MECAT	Number of bases	232,519,767	616,247,287	4,937,676,257
	Error rate (%)	0.1714	0.2088	0.2675
	Deletions	392,085	1,228,519	12,514,560
	Insertions	43,487	168,401	1,985,795
	Substitutions	2,006	14,471	107,294
	Recall (%)	99.9547	99.9428	99.9258
	Precision (%)	99.8362	99.7996	99.7415
	Trimmed / split reads	3,622	9,606	112,978
	Mean missing size (bp)	365	380.7	566.8
	Extended reads	0	0	30
	Mean extension size (bp)	0	0	165
	Low quality reads	0	92	443
	Short reads	0	0	0
	Homopolymer ratio	0.9967	1.0000	1.0000
	Runtime	5 min	16 min	2 h 43 min
	Memory (MB)	2,387	4,954	10,563

Table 10: Comparison of the different error correction tools, on low error rate and medium coverage datasets. This corresponds to the *E. coli* 60x (low), *S. cerevisiae* 60x (low) and *C. elegans* 60x (low) datasets of Table 3. [1] Colormap was not launched on the *C. elegans* dataset due to its large runtimes. [2] HG-CoLoR was not launched on the *C. elegans* dataset due to its large runtimes. [3] Proovread was not launched on the *C. elegans* dataset due to its large runtimes. [4] Daccord could not perform correction on the *C. elegans* dataset, since the alignment step with DALIGNER required more than 128 GB of memory.

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3.9 Ultra-long reads

- ECTools, because of its unsatisfying results on previous experiments;
- Hercules, because of its unsatisfying results and large runtimes on previous experiments;
- LSC, because of its unsatisfying results on previous experiments;
- Nanocorr, because of its large runtimes on previous experiments;
- NaS, because of its large runtimes on previous experiments;
- Proovread, because it could not be installed on the computer we performed these experiments on;
- Daccord, because it could not perform correction, since the alignment step with DALIGNER required more than 128 GB of memory.

Tool	Remapping		Assembly	
CoLoRMap	Number of reads	419,332	Number of contigs	404
	Number of bases	1,510,947,079	Number of aligned contigs	404
	Average length (bp)	3,603	Number of breakpoints	52
	Aligned reads (%)	99.9764	NGA50 (bp)	1,126
	Average identity (%)	96.4502	NGA75 (bp)	1,126
	Genome coverage (%)	91.9475	Genome coverage (%)	4.5428
	Runtime	304 h 10 min		
	Memory (MB)	80,613		
FMLRC	Number of reads	1,075,867	Number of contigs	66
	Number of bases	7,647,637,034	Number of aligned contigs	63
	Average length (bp)	7,108	Number of breakpoints	275
	Aligned reads (%)	93.1494	NGA50 (bp)	2,010
	Average identity (%)	95.3985	NGA75 (bp)	2,010
	Genome coverage (%)	92.4947	Genome coverage (%)	47.8800
	Runtime	77 h 15 min		
	Memory (MB)	16,381		
HALC	Number of reads	1,065,270	Number of contigs	72
	Number of bases	7,256,690,308	Number of aligned contigs	69
	Average length (bp)	6,812	Number of breakpoints	256
	Aligned reads (%)	97.4287	NGA50 (bp)	1,239
	Average identity (%)	94.9728	NGA75 (bp)	1,239
	Genome coverage (%)	92.4495	Genome coverage (%)	48.0811
	Runtime	26 h 47 min		
	Memory (MB)	7,739		
HG-CoLoR	Number of reads	970,212	Number of contigs	196
	Number of bases	6,553,413,717	Number of aligned contigs	196
	Average length (bp)	6,754	Number of breakpoints	393
	Aligned reads (%)	99.8157	NGA50 (bp)	4,628
	Average identity (%)	98.8042	NGA75 (bp)	4,628
	Genome coverage (%)	92.4523	Genome coverage (%)	49.6926
	Runtime	167 h 47 min		
	Memory (MB)	50,898		
Jabba	Number of reads	1,059,671	Number of contigs	2,032
	Number of bases	3,460,536,974	Number of aligned contigs	2,032
	Average length (bp)	3,265	Number of breakpoints	83
	Aligned reads (%)	89.8926	NGA50 (bp)	3,418
	Average identity (%)	99.9888	NGA75 (bp)	3,418
	Genome coverage (%)	87.7037	Genome coverage (%)	28.8106
	Runtime	8 h 23 min		
	Memory (MB)	25,917		
LoRDEC	Number of reads	1,075,867	Number of contigs	111
	Number of bases	6,850,695,672	Number of aligned contigs	108
	Average length (bp)	6,368	Number of breakpoints	655
	Aligned reads (%)	89.9923	NGA50 (bp)	2,281
	Average identity (%)	91.7205	NGA75 (bp)	2,281
	Genome coverage (%)	92.4693	Genome coverage (%)	48.3585
	Runtime	12 h 52 min		
	Memory (MB)	7,902		
Canu	Number of reads	717,436	Number of contigs	361
	Number of bases	5,604,822,256	Number of aligned contigs	359
	Average length (bp)	7,812	Number of breakpoints	275
	Aligned reads (%)	97.6001	NGA50 (bp)	951,801
	Average identity (%)	90.4029	NGA75 (bp)	267,665
	Genome coverage (%)	92.3293	Genome coverage (%)	88.0860
	Runtime	14 h 04 min		
	Memory (MB)	10,295		

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Tool	Remapping		Assembly	
CONSENT	Number of reads	869,462	Number of contigs	248
	Number of bases	6,348,740,755	Number of aligned contigs	245
	Average length (bp)	7,301	Number of breakpoints	871
	Aligned reads (%)	99.5864	NGA50 (bp)	3,760,455
	Average identity (%)	93.0004	NGA75 (bp)	1,368,818
	Genome coverage (%)	92.3993	Genome coverage (%)	88.9587
	Runtime	8 h 30 min		
	Memory (MB)	45,869		
FLAS[1]	Number of reads	717,973	Number of contigs	259
	Number of bases	5,695,206,077	Number of aligned contigs	258
	Average length (bp)	7,932	Number of breakpoints	317
	Aligned reads (%)	99.0611	NGA50 (bp)	1,622,624
	Average identity (%)	90.9952	NGA75 (bp)	346,914
	Genome coverage (%)	92.3690	Genome coverage (%)	88.4068
	Runtime	4 h 57 min		
	Memory (MB)	14,957		
LoRMA	Number of reads	6,796,439	Number of contigs	4
	Number of bases	1,247,317,428	Number of aligned contigs	4
	Average length (bp)	183	Number of breakpoints	0
	Aligned reads (%)	96.4997	NGA50 (bp)	2,742
	Average identity (%)	97.8254	NGA75 (bp)	2,742
	Genome coverage (%)	28.6189	Genome coverage (%)	0.0060
	Runtime	13 h 07 min		
	Memory (MB)	50,435		
MECAT[1]	Number of reads	667,532	Number of contigs	249
	Number of bases	5,479,325,177	Number of aligned contigs	248
	Average length (bp)	8,208	Number of breakpoints	199
	Aligned reads (%)	99.9495	NGA50 (bp)	1,783,022
	Average identity (%)	91.6925	NGA75 (bp)	462,658
	Genome coverage (%)	91.4387	Genome coverage (%)	88.7002
	Runtime	1 h 53 min		
	Memory (MB)	11,075		

Table 11: Comparison of the different error correction tools, on a dasaset containing ONT ultra-long reads. This corresponds to the *H. sapiens* dataset of Table 3. [1] Reads longer than 50 kbp were filtered out, since they caused the programs to stop with an error.

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4 Conclusion

In this paper, we presented the state-of-the-art of long-read error correction, tackling both hybrid and self-correction. For each approach, we described the different existing methodologies, and listed all tools available at the moment. Four different approaches thus exist for hybrid correction: short reads alignment, contigs alignment, use of de Bruijn graphs and use of hidden Markov models. For self-correction, two main methodologies exist: multiple sequence alignment and use of de Bruijn graphs. As of today, a total of 29 different methods exist for performing long-read error correction.

We also showcased how the long reads datasets characteristics can impact the quality of the correction. In particular, our experiments show that self-correction performs better than hybrid correction as the sequencing depth grows. Oppositely, given high error rates, hybrid correction tends to perform the best, even when the sequencing depth is high. In addition, our experiments also underline the fact that self-correction tends to perform better when the complexity of the sequenced organism grows.

Further work shall focus on a more in-depth description of each available tool, to give the reader a better understanding of the algorithmic differences exiting between tools adopting the same approaches. In addition, more datasets could also be studied, in a order to provide better guidelines as to which tool to chose according to the datasets characteristics.

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