Alcohol-Induced Increase in the Content of CYP2E1 in Human Liver Microsomes Causes a Multifold Activation of CYP3A4 and Attenuates its Cooperativity

Bikash Dangi[1], Nadezhda Y. Davydova[1], Nikita E. Vavilov[2], Victor G. Zgoda[2,3] and Dmitri R. Davydov[1*]

[1]Department of Chemistry, Washington State University, Pullman, WA, 99164; [2]Institute of Biomedical Chemistry, Moscow, 119121, Russia; [3]Skolkovo Institute of Science and Technology, 143025 Skolkovo, Moscow region, Russia

Abstract

Here we investigate the effect of alcohol-induced increase in the content of CYP2E1 in human liver microsomes (HLM) on the function of CYP3A4. In these studies we used a model that implements enrichment of HLM samples with CYP2E1 through membrane incorporation of the purified protein. Enrichment of HLM with CYP2E1 considerably increases the rate of metabolism of 7-benzyloxyquinoline (BQ) and attenuates the homotropic cooperativity observed with this CYP3A4-specific substrate. Incorporation of CYP2E1 also eliminates the activating effect of α-Naphthoflavone (ANF) on BQ metabolism seen in some untreated HLM samples. To probe the physiological relevance of these effects we compared three pulled preparations of HLM from normal donors (HLM-N) with a preparation obtained from heavy alcohol consumers (HLM-A). The composition of the P450 pool in all four samples was characterized with mass-spectrometric determination of 11 cytochrome P450 species. The molar content of CYP2E1 in HLM-A was from 2.5 to 3.3 times higher than that found in HLM-N. In contrast, the content of CYP3A4 in HLM-A was the lowest among all four HLM samples. Despite of that, HLM-A exhibited much higher rate of metabolism and lower degree of homotropic cooperativity with BQ, similar to that observed in CYP2E1-enriched HLM-N. In order to substantiate the hypothesis on the involvement of physical interactions between CYP2E1 and CYP3A4 in the observed effects we probed hetero-association of these proteins in Supersomes™ containing recombinant CYP3A4 with a technique based on homo-FRET and employing CYP2E1 labeled with BODIPY-618 maleimide. These experiments demonstrated high affinity interactions between the two enzymes and revealed an inhibitory effect of ANF on their hetero-association. Our results demonstrate that the catalytic activity and allosteric properties of CYP3A4 are fundamentally dependent on the composition of the cytochrome P450 ensemble and imply a profound impact of chronic alcohol exposure on the pharmacokinetics of drugs metabolized by CYP3A4.

***Keywords*:** cytochrome P450; CYP2E1; CYP3A4; oligomerization; cooperativity; protein-protein interaction; alcohol-drug interactions, alcohol exposure

[*] ***Corresponding author**:* Dmitri R. Davydov, Department of Chemistry, Washington State University, MS 4630, Pullman, WA 99164-4630; Tel. +1 509 335-5983; E-mail: dmrdavyd@gmail.com

Introduction

The core of the drug-metabolizing ensemble in human liver is the ensemble of multiple P450 species co-localized in the membrane of endoplasmic reticulum (ER). There is an emerging recognition of the fact that multiple P450 enzymes interact with each other with the formation of heteromeric complexes where the functional properties of individual P450 enzymes are largely modified (Davydov, 2011; Reed and Backes, 2012; Davydov, 2016; Reed and Backes, 2016). Resent findings point out the association of dissimilar P450s as the major cause for poor correlation between the composition of the P450 ensemble and its drug metabolizing profile demonstrated with studies in a large set of normal human liver microsomes (Gao et al., 2016; Zhang et al., 2016).

Of particular importance are the changes in P450-P450 crosstalk induced by alcohol consumption. Although the multi-fold increase in the content of CYP2E1 in liver observed in alcohol consumers is well documented (Cederbaum, 1998; Dupont et al., 1998; Lieber, 1999; Cederbaum, 2006), the involvement of CYP2E1 in alcohol-drug interactions is commonly considered insignificant due to a minor role of this enzyme in drug metabolism (Jang and Harris, 2007). However, the effects of induction of CYP2E1 may stretch beyond the changes in CYP2E1-dependent drug metabolism and involve the effects of CYP2E1 on the functional properties of other drug-metabolizing enzymes.

In order to probe the crosstalk between CYP2E1 and other P450 species and elucidate its role in alcohol-drug interactions we established a model of alcohol-induced increase in CYP2E1 content that implements enrichment of HLM samples with CYP2E1 through membrane incorporation of the purified protein (Davydov et al., 2015; Davydov et al., 2017; Davydova et al., 2019). We demonstrated that the adopted CYP2E1 becomes a fully-functional member of the drug-metabolizing ensemble and interacts with other P450 enzymes with the formation of heteromeric complexes (Davydova et al., 2019). Studying the effect of enrichment of HLM with CYP2E1 on the function of other P450 enzymes we demonstrated a CYP2E1-induced activation of CYP1A2 and the associated re-routing of the metabolism of 7-ethoxy-4-cyanocoumarin (CEC), the substrate concurrently metabolized by CYP2C19 and CYP1A2, towards the latter enzyme (Davydova et al., 2019).

The present study continues our efforts on revealing the impact of alcohol-induced increase in CYP2E1 content on drug metabolism. Here we explore the effects of CYP2E1 on the functional properties of CYP3A4, the enzyme that metabolizes about 50% of drugs on

the market (Guengerich, 1999). As a probe substrate we used 7-benzyloxyquinoline (BQ), a fluorogenic substrate highly selective for CYP3A4 (Stresser et al., 2002). In addition to studying the effect of CYP2E1 on the parameters of BQ metabolism in a series of microsomal preparations with thoroughly characterized composition of the P450 pool, we also probed its impact on the effect of α-naphthoflavone, a prototypical allosteric effector of CYP3A4. In order to assess the relevance of the results obtained with CYP2E1-enriched microsomes to the changes caused by the effects of chronic alcohol exposure we also compared the parameters of BQ metabolism in a pulled HLM preparation obtained from 10 alcoholic donors with those exhibited by three pulled HLM preparations obtained from donors with no history of alcohol dependence. Our results demonstrate that the catalytic activity and allosteric properties of CYP3A4 are fundamentally dependent on the composition of the cytochrome P450 ensemble in human liver and imply a profound impact of chronic alcohol exposure on the pharmacokinetics of drugs metabolized by CYP3A4.

Furthermore, probing possible mechanisms of CYP2E1-dependent activation of CYP3A4 revealed in this study we investigated the interactions between the two proteins in the microsomal membrane using the technique based on homo-FRET in the oligomers of CYP2E1 labeled with BODYPY-618 maleimide (CYP2E1-BODIPY) (Davydova et al., 2019). High-affinity interactions between CYP2E1-BODIPY with CYP3A4 and their modulation by α-naphthoflavone revealed in these studies suggest a direct involvement of hetero-association between CYP2E1 and CYP3A4 in the observed effects.

Materials and methods.

Chemicals

7-benzyloxyquinoline (BQ) was a product of CypEx (Dundee, UK). 7-hydroxycoumarin was obtained from Acros Organics, a part of Thermo Fisher Scientific (New Jersey, NJ). 7,8-benzoflavone (α-naphthoflavone, ANF) was a product of Indofine Chemical Company (Hillsborough, NJ). All other reagents were of ACS grade and used without additional purification.

Protein expression and purification

N-terminally truncated (Δ3-20) and C-terminally His-tagged CYP2E1 (Spatzenegger et al., 2003) was expressed in *E. coli* TOPP3 cells and purified as described earlier (Davydova et al., 2019).

Pulled human liver microsomes and their characterization with mass-spectrometry

The preparation of Human Liver Microsomes (HLM) obtained from 10 donors (mixed gender) with a history of chronic alcohol exposure (lot FVT) was purchased from BioIVT corporation (Baltimore, MD). This preparation is referred hereinafter as HLM-A. We also studied three different preparations of pulled human liver microsomes from 50 donors (mixed gender) without reported history of alcohol exposure. The preparation obtained by differential centrifugation of pulled human liver S9 fraction (the product of BD Gentest, lot number 3212595) is referred to as HLM-N1. The HLM samples referred here as HLM-N2 and HLM-N3 are InVitroCYP™ M-class 50-donor mixed gender pooled HLM preparations, lots LBA and LFJ respectively obtained from BioIVT corporation (Baltimore, MD).

The composition of the cytochrom P450 ensemble in these preparations was characterized by mass-spectrometric analysis with a triple quadrupole mass spectrometer using the method of multiple reaction monitoring (MRM) as described previously (Davydova et al., 2019). Content of NADPH-cytochrome P450 reductase and cytochromes P450 1A2, 2A6, 2B6, 2D6, 2C8, 2C9, 2C18, 2C19, 2E1, 3A4 and 3A5 was determined based on the concentration of protein-specific tryptic peptides in trypsin-digested samples with the use of the stable isotope labeled internal standards (SIS) (Davydova et al., 2019). For each protein, one standard peptide with three transitions was used. The peptides, which sequences are provided in Table 1, were arranged into one Selected Reaction Monitoring (SRM) assay. Mass-spectrometry measurements of P450 content were performed using the equipment of "Human Proteome" Core Facilities of the Institute of Biomedical Chemistry (Moscow, Russia).

Characterization of the content of protein, phospholipids, NADPH-cytochrome P450 reductase and cytochromes P450 in HLM

Determinations of protein and phospholipid concentrations in microsomal suspensions were performed with the bicinchoninic acid procedure (Smith et al., 1985) and through the determination of total phosphorus in a chloroform/methanol extract according to Bartlett (Bartlett, 1959), respectively. The concentration of NADPH-cytochrome P450 reductase in microsomal membranes was determined based on the rate of NADPH-dependent reduction of cytochrome *c* at 25 °C and the effective molar concentration of CPR was estimated using the turnover number of 3750 min^{-1} (Davydova et al., 2019). The total concentration of cytochromes P450 in HLM was determined with a variant of "oxidized CO

versus reduced CO difference spectrum" method as described earlier (Davydova et al., 2019).

Preparation of CYP2E1-enriched HLM samples.

Incorporation of CYP2E1 into HLM was performed by incubation of undiluted suspensions of HLM (20-25 mg/ml protein, 10-13 mM phospholipid) in 125 mM K-Phosphate buffer containing 0.25M Sucrose with purified CYP2E1 for 16 - 20 hours at 4°C at continuous stirring. CYP2E1 was added in the amount ranging from 0.25 to 1 molar equivalent to the endogenous cytochrome P450 present in HLM. Following the incubation, the suspension was diluted 4-8 times with 125 mM K-Phosphate buffer, pH 7.4 containing 0.25 M sucrose and centrifuged at 53,000 rpm (150,000 g) in an Optima TLX ultracentrifuge (Beckman Coulter Inc., Brea, CA, USA) with a TLA100.3 rotor for 90 min at 4 °C. The pellet was resuspended in the same buffer to the protein concentration of 15-20 mg/ml. The amount of incorporated cytochrome P450 was calculated from the difference between the heme protein added to the incubation media and the enzyme found in the supernatant. According to the results of this assay, our procedure resulted in incorporation of 96 -98% of the added protein into the microsomal membrane.

Fluorimetric assays of BQ metabolism.

The rate of debenzylation of 7-benzyloxyquinoline was measured with a real-time continuous fluorometric assay using a Cary Eclipse fluorometer (Agilent Technologies, Santa Clara, CA, USA) or a custom-modified PTI QM-1 fluorometer (Photon Technology International, New Brunswick, NJ (Davydov et al., 2017). In the experiments with Cary Eclipse the excitation was performed with a monochromatic light centered at 405 nm with 5 nm bandwidth. In the case of PTI QM-1 the excitation light centered at 405 nm was emitted by a CPS405 collimated laser diode module (Thorlabs Inc, Newton, NJ). The emission wavelength was set at 516 nm with a 20 nm slit. The rate of formation of 7-hydroxyquinolyne was estimated by determining the slope of the linear part of the kinetic curve recorded over a period of 2 – 3 min.

All kinetic assays were performed in 0.1 M Na-HEPES buffer, pH 7.4, containing 60 mM KCl. In the case of the use of Cary Eclipse the total volume of the incubation mixture was equal to 300µl and a 5 x 5 mm quartz cell was used. In the experiments with PTI QM-1 fluorometer we used a 3 x 3 mm quartz cell and the volume of the sample was equal to 60µl. With both instruments the kinetic assays were carried out at continuous stirring and the temperature was maintained at 30 °C with a circulating water bath. An aliquot of 15-24 mM

stock solution of BQ in acetone was added to attain the desired substrate concentration in the range of 0.5 - 250 μM. The reaction was initiated by addition of 20 mM solution of NADPH to the concentration of 200 μM. Fitting of the dependencies of the reaction rate on the substrate concentration to Michaelis-Menten and Hill equations was performed with a combination of Marquardt and Nelder-Mead non-linear regression algorithms as implemented in our SpectraLab software (Davydov et al., 1995; Davydov et al., 2016).

Monitoring the interactions of CYP2E1-BODIPY with microsomal membranes.

Labeling of CYP2E1 with BODIPY-618 maleimide was performed at 2:1 label to protein ratio as described earlier (Davydova et al., 2019). The studies of interactions of CYP2E1-BODIPY with microsomal membranes were performed with the use of a Cary Eclipse spectrofluorometer equipped with a Peltier 4-position cell holder. The excitation of donor phosphorescence was performed with monochromatic light centered at 405 with 20 nm bandwidth. Alternatively the measurements were done with the use a custom-modified PTI QM-1 fluorometer (Photon Technology International, New Brunswick, NJ) equipped with a thermostated cell holder and a refrigerated circulating bath. In this case the excitation was performed at 405 nm with a CPS405 collimated laser diode module (Thorlabs Inc, Newton, NJ). The spectra in the 570 – 750 nm wavelength region were recorded repetitively with the time interval varying from 0.5 to 15 min during the course of monitoring (5 – 16 hours). The experiments were performed at continuous stirring at 4 °C in 100 mM Na-Hepes buffer (pH 7.4) containing 150 mM KCl and 250 mM sucrose.

Calculations of the surface density of cytochromes P450 in microsomal membranes (C_{CYP2E1}) were based on the molar ratio of membranous phospholipids to incorporated CYP2E1 ($R_{L/P}$). For our calculations we assumed the area of microsomal membrane corresponding to one molecule of phospholipid in a monolayer to be equal to 0.95 nm^2 (Wibo et al., 1971), similar to the approach used in our earlier reports (Davydov et al., 2015; Davydov et al., 2017; Davydova et al., 2019). The use of this value results in the following relationship between C_{CYP2E1} and the $R_{L/P}$:

$$C_{CYP2E1} = 349.6 / R_{L/P} \quad \text{pmol/cm}^2 \qquad \text{(Eq.1)}$$

Analysis of series of spectra obtained in fluorescence spectroscopy experiments was done by principal component analysis (PCA) (Davydov et al., 1995) as previously described (Davydova et al., 2019). The equation for the equilibrium of binary association

(dimerization) used in the fitting of oligomerization isotherms (dependencies of FRET efficiency on the concentration of P450 in membranes) had the following form:

$$[X]=\frac{1}{2}\cdot K_D+[E]_0-\sqrt{\frac{1}{4}\cdot K_D^2+[E]_0\cdot K_D} \qquad \text{(Eq. 2)}$$

where $[E]_0$, $[X]$, and K_D are the total concentration of the associating compound (enzyme), the concentration of its dimers, and the apparent dissociation constant, respectively. In order to use this equation in fitting of the dependencies of the relative increase in fluorescence observed at enzyme concentration $[E]_0$ (R_E) equation (1) was complemented with the parameter R_{max} . This parameter corresponds to the value of R_E observed upon a thorough dissociation of completely oligomerized enzyme:

$$R_E=R_{max}\cdot([X]/[E]_0) \qquad \text{(Eq. 3)}$$

The parameter R_{max} is determined by the efficiency of FRET (E_{FRET}) according to the following relationship:

$$E_{FRET}=R_{max}/(1+R_{max}) \qquad \text{(Eq. 4)}$$

Fitting of the titration isotherms to the above equations was performed with non-linear regression using a combination of Nelder-Mead and Marquardt algorithms as implemented in our SpectraLab software (Davydov et al., 1995).

Results.

Characterization of the composition of the cytochrome P450 pool in HLM samples.

The preparations of HLM used in this study were characterized by determining the content of NADPH-cytochrome P450 reductase and 11 major cytochrome P450 species, namely CYP1A2, CYP2A6, CYP2B6, CYP2C8, CYP2C9, CYP2C18, CYP2C19, CYP2E1, CYP3A4 and CYP3A5, with mass-spectrometry. The total content of cytochromes P450 and the concentration of NADPH-cytochrome reductase (CPR) were also quantified with absorbance spectroscopy and determination of the rate of NADPH-dependent reduction of cytochrome *c*, respectively.

Results of our analysis of the composition of the P450 pool in HLM are summarized in Table 1. The values shown in this table for HLM-N2, HLM-N3 and HLM-A are the mean values averaged over the results of 15, 20 and 11 individual measurements respectively. The values shown for HLM-N1 are the results of a single measurement. As expected, the fractional content of CYP2E1 in HLM-A was from 2.5 to 3.3 fold higher than that detected in the HLM-N samples.

Parameters of BQ metabolism exhibited by four HLM-preparations

In order to probe the effect of the composition of the P450 ensemble on the functional properties of CYP3A4 we determined the parameters of O-debenzylation of BQ by all four microsomal preparations under study. We also probed the effect of ANF, the prototypical allosteric effector of the enzyme. Results of these studies are summarized in Table 2 and illustrated in Fig. 1. It should be noted that the rate of metabolism of BQ in HLM in this study was normalized on the concentration of CPR in the microsomal membrane (see Materials and Methods). In view of a high excess of cytochromes P450 over CPR in HLM, this normalization was considered as the most appropriate approach for comparing the rates of metabolism in the HLM preparations with different composition of the cytochrome P450 ensemble.

In all three samples of HLM-N the metabolism of BQ revealed a prominent homotropic cooperativity characterized with the Hill coefficient of 1.9 – 2.2 (Fig. 1, curves shown in black open circles), in a good agreement with the previous reports (Stresser et al., 2002; Davydov et al., 2008). In HLM-N samples the maximal rate of reaction increases concomitant with increasing fractional content of CYP3A4 in microsomes (Table 2). Strikingly, despite the low content of CYP3A4 in HLM-A, this preparation is characterized with the highest maximal rate of BQ metabolism over all four HLM samples studied. V_{max} value exhibited by HLM-A is over 6 times higher than the value obtained with HLM-N1, where the fractional content of CYP3A4 (28%) is comparable to that observed in HLM-A (21%). The values of V_{max} obtained with HLM-N2 and HLM-N3, where the fractional content of CYP3A4 is around 40%, are respectively 1.4 and 2.2 times lower than that characteristic to HLM-A. This multifold activation of BQ metabolism in HLM-A is associated with the attenuation of homotropic cooperativity, which is revealed in a considerable decrease of the Hill coefficient (Table 2).

The contrasting difference between HLM-A and HLM-N preparation in the rate of BQ metabolism is illustrated in Fig. 3, where the V_{max} values are plotted against the molar ratio of CYP3A4 to CPR determined in these preparations with LC-MS/MS. As it might be expected, the V_{max} value observed with HLM-N preparations and calculated per molar content of CPR displays an evident proportionality to the content of CYP3A4A (Fig. 2a). However, the rate observed with HLM-A, where the content of CYP3A4 is the lowest out of all four preparations, deviates from this proportionality being the highest of all four values under comparison. The difference between HLM-A and HLM-N becomes even more

contrasting when the rates of turnover are calculated per molar content of CYP3A4. Obviously, this way of normalization attenuates the difference between the three HLM-N samples, whereas the turnover number calculated for HLM-A becomes almost an order of magnitude higher than the values calculated for HLM-N samples (Fig. 2b).

The effect of ANF.

Consistent with the previous observations (Domanski et al., 2001; Davydov et al., 2008; Davydov et al., 2013), the addition of 25 mM ANF completely eliminated homotropic cooperativity observed with BQ and considerably decreased the S_{50} values in all four HLM samples studied (Table 2). However, the effect of ANF on the maximal rate of reaction differs dramatically between the HLM samples. Whereas in the case of HLM-N1 the addition of ANF increased the maximal rate of reaction almost two times, it resulted in over two-fold inhibition of BQ metabolism in HLM-N2, while having no statistically significant effect on V_{max} in HLM-N3 and HLM-A (Table 2). The effect of ANF on substrate saturation curves in four HLM samples is illustrated in Fig. 1 (curves in black closed circles).

As shown in Fig. 3, where the V_{max} values observed in the presence of ANF are plotted against the content of CYP3A4 in HLM preparations, addition of this allosteric effector eliminates the proportionality between the rate of BQ turnover and the content of CYP3A4 in HLM-N samples. This ANF-dependent leveling of the turnover rates is achieved through the activating effect of ANF in HLM-N1, the sample with the lowest CYP3A4 content, and a substantial inhibition of BQ metabolism in CYP3A4-rich HLM-N2. At the same time, addition of ANF does not affect the contrasting difference between HLM-A and HLM-N samples in the rate of BQ turnover (Fig. 3).

Effect of incorporation of CYP2E1 into HLM on BQ metabolism

To probe if the loss of homotropic cooperativity with BQ and a multifold increase in the rate of its metabolism in HLM-A is caused by alcohol-induced increase in the fraction of CYP2E1 in the P450 pool we studied the effect of incorporation of exogenous CYP2E1 into HLM. The dependencies of the reaction rate on substrate concentration with and without addition of ANF obtained with all four studied HLM samples enriched with CYP2E1 are exemplified in Fig. 1 (curves shown in red). The parameters of BQ metabolism in these preparations may be found in Table 2. As seen from these results, incorporation of CYP2E1 essentially eliminates the heterotropic cooperatvity in all samples studied. Furthermore, in two of the three HLM-N preparations – HLM-N1 and HLM-N3 – this incorporation results in a significant increase in the maximal rate of BQ turnover. Therefore, in some sense, the

effect of incorporation of CYP2E1 into HLM on homotropic cooperativity of CYP3A4 is analogous to the effect of the addition of ANF.

Importantly, addition of ANF to CYP2E1-enriched HLM preparations largely eliminates their differences from the untreated HLM samples in the parameters of BQ metabolism. With all three CYP2E1-enriched HLM-N samples the addition of ANF considerably inhibits BQ turnover at substrate saturation, while decreasing S_{50} values and eliminating heterotropic cooperativity. In the case of CYP2E1-enriched HLM-A the effects of ANF on V_{max} and S_{50} are marginal, while the effect on the Hill coefficient is retained.

The effect of increasing amounts of incorporated CYP2E1 on the parameters of BQ metabolism in HLM-N and HLM-A preparations in the absence and in the presence of ANF is illustrated in Fig. 4 and Fig. 5 respectively. The dependencies of S_{50}, V_{max} and the Hill coefficient on CYP2E1 content obtained in the absence of ANF reveal a striking contrast between the "normal" HLM samples and the sample obtained from the alcohol-exposed donors. Enrichment of HLM-N in CYP2E1 resulted in a stepwise decrease in the Hill coefficient, while increasing the values of S_{50} and V_{max}. In contrast, incorporation of CYP2E1 into HLM-A, while also increasing $V_{max,}$, had no statistically significant effect on the values of S_{50} and the Hill coefficient (Fig 4).

At the same time, the dependencies of the parameters of BQ metabolism on CYP2E1 content obtained in the presence of ANF (Fig. 5) show that the addition of this allosteric effector eliminates most of the effects of CYP2E1, although the increased activity of CYP2E1 in HLM-A preparation remains to be seen (Fig. 5). This observation suggests that the grounds of heterotropic cooperativity in CYP3A4 and the origins of its activation by CYP2E1 may involve some common mechanistic elements.

Interactions of CYP2E1 with CYP3A4 in the membranes of Supersomes and their modulation by ANF.

In our recent study we introduced a homo-FRET-based technique to monitor the incorporation of CYP2E1 into the microsomal membrane and probe the effect of microsomal P450 proteins on the dissociation of CYP2E1 homo-oligomers (Davydova et al., 2019). This method employs the use of CYP2E1 labeled with BODIPY-618 maleimide in the molar ratio 1:2. Its principle is based on a deep quenching of the fluorescence of the label in the CYP2E1-BODIPY homo-oligomers due to homo-FRET between the labels attached to different loci of the interacting protein molecules (Davydova et al., 2019). Its setup employs registration of the process of dissociation of homo-oligomers after their incorporation into

the microsomal membrane through monitoring the consequent increase in the intensity of fluorescence (Fig. 5). The dependence of the amplitude of the observed increase on the amount of added microsomes (phosopholipid-to–CYP2E1-BODIPY ratio, $R_{L/P}$) allows determining the apparent dissociation constant (K_D) of the homo-oligomers, or the value of $R_{L/P}$ at which the degree of homo-oligomerization is equal to 50% ($R_{L/P,50\%}$). Effect of the presence of a given cytochrome P450 species in the microsomal membrane on $R_{L/P,50\%}$ is interpreted as reflecting its ability to form mixed oligomers with CYP2E1 (thus promoting the dissociatiation of the homo-oligomers) (Davydova et al., 2019).

In order to probe the relevance of the effects of CYP2E1 on the functional properties of CYP3A4 to direct physical interactions of these proteins we explored the ability of CYP2E1 to form mixed oligomers with CYP3A4 with the use of the homo-FRET based approach. In these experiments we compared SS(3A4) with the preparation of Supersomes containing recombinant CPR and cytochrome b_5, but lacking any P450 protein (SS(CPR)). Dependencies of the amplitude of increase in the intensity of fluorescence on the resulting surface density of CYP2E1-BODIPY in the membrane (or $R_{L/P}$ ratio) are shown in Fig. 6. The parameters deduced by fitting these dependencies to Eq. 2 are summarized in Table 3.

Comparison of the dependencies obtained with SS(3A4) and SS(CPR) reveal a dramatic effect of microsomal CYP3A4 on homo-oligomerization of the incorporated CYP2E1-BODIPY. If in the membranes of SS(CPR) 50% dissociation of the homo-oligomers requires the molar ratio of membrane phospholipids to CYP2E1 to be as high as 14,400:1, the presence of CYP3A4 decreases this ratio to 176:1 (Table 3). Another important finding is a two-fold decrease of the apparent maximal amplitude of the increase in fluorescence intensity in the presence of CYP3A4. This observation may reveal a conformational effect of the interactions of CYP3A4 with CYP2E1 resulting in a decreased intensity of BODIPY fluorescence in CYP3A4-CYP2E1 complexes.

We also probed the effect of ANF on hetero-association of CYP2E1 with CYP3A4 in a series of experiments with incorporation of CYP2E1-BODIPY into SS(3A4) in the presence of 25 μM ANF. As seen from Table 3, the addition of ANF increases the value of $R_{L/P,50\%}$ to 604:1 and eliminates the effect of CYP3A4 on the maximal intensity of fluorescence. These observations indicate that the addition of ANF attenuates the ability of CYP3A4 to form mixed oligomers with CYP2E1 and eliminates its apparent conformational effect revealed in decreased intensity of fluorescence of CYP2E1-BODIPY.

Discussion.

Changes in the composition of the cytochrome P450 ensemble under influence of chronic alcohol exposure.

Analysis of the content of 11 drug-metabolizing cytochrome P450 species in pooled HLM preparation obtained from alcoholic donors revealed a 2.5 – 3.3 fold increase in the fractional content of CYP2E1 as compared to the microsomal samples unaffected by alcohol exposure (Table 1). This observation is in a good agreement with the previous reports (Cederbaum, 1998; Dupont et al., 1998; Lieber, 1999; Cederbaum, 2006). Our data also indicate ~2 fold elevation of the fractional contents of CYP2D6 and CYP2C18 (Table 1). Although the results of our analysis of a single pooled preparation of HLM from alcohol-exposed donors cannot serve as a proof of alcohol-induced increase in the content of these P450 species, this observation may provide a clue to further identification of changes in P450 expression induced by alcohol exposure. Importantly, our results reveal no alcohol-induced increase in the contents of CYP3A4 and CYP2A6, notwithstanding with the existing reports on induction of expression of these enzymes by ethanol in model animals and in cell culture (Feierman et al., 2003; Jin et al., 2012).

Another notable difference of the HLM-A preparation from HLM-N samples is low degree of coverage of the total P450 concentration with the sum of the contents of 11 analyzed P450 species. The averaged degree of coverage of 84 ±19 % calculated for three HLM-N samples is in a good agreement with the estimate of 78%, calculated for the fractional content of the 11 analyzed P450 species based on the data of Wang and co-authors, who determined the content of 25 P450 species in a series of 102 individual HLM samples (Wang et al., 2020). In contrast, the degree of coverage calculated for HLM-A is as low as 18±8 %. A possible source of this difference may be an alcohol-induced increase in the content of some major P450 enzyme(s) not included in our assays, such as CYP4F2 and/ or CYP4A11.

Multifold activation of CYP3A4 by alcohol exposure is stipulated by the effect of CYP2E1 on the landscape of P450-P450 interactions

The most striking result of this study is a demonstration of a multifold activation of CYP3A4-dependent metabolism of BQ in HLM sample from alcohol-exposed donors. Although the fractional content of CYP3A enzymes in HLM-A was the lowest among all four studied preparations, these microsomes exhibited much higher rate of metabolism and

lower homotropic cooperativity with BQ as compared to HLM-N samples. Notably, in contrast to two of the three HLM-N preparations, HLM-A does not show any ANF-induced increase in V_{max} of debenzylation of BQ. Incorporation of additional CYP2E1 into HLM-A results in a further increase in the rate of BQ metabolism and makes this rate over an order of magnitude faster than that exhibited by the HLM-N1 sample, which is characterized by a similar CYP3A content.

The finding that the activation of BQ metabolism and attenuation of its cooperativity may be reproduced in HLM-N samples via incorporation of the exogenous CYP2E1 into their membranes suggests that these effects, at least in a part, represent direct results of the alcohol-induced increase in CYP2E1 content. It is extremely unlikely that the changes in the parameters of BQ metabolism upon CYP2E1 incorporation into HLM may result from the activity of CYP2E1 itself or be due to participation of any other P450 enzyme, as BQ metabolism is known to be almost entirely selective to CYP3A4. CYP2E1 itself lacks any potential to metabolize this substrate (Stresser et al., 2002; Davydov et al., 2015). According to the data of Stresser and co-authors (Stresser et al., 2002), the only other enzymes present in adult HLM and capable of BQ metabolism are CYP1A2 and CYP3A5. However, the turnover numbers exhibited by these enzymes with BQ are respectively 8.5 and 4.3 times lower than that characteristic to CYP3A4 (Stresser et al., 2002). Thus, due to relatively low abundance of CYP1A2 and CYP3A5 in HLM (Table 1) their sizable participation in BQ metabolism is extremely unlikely. We can conclude therefore that the effects of CYP2E1 incorporation demonstrate that the increased rate and decreased homotropic cooperativity in BQ metabolism seen in HLM-A are attributable, at least in a part, to direct effects of the increased CYP2E1 content in this sample.

The striking effects of one P450 protein on the functional properties of another P450 enzyme observed in this study and evidenced in earlier reports from our group and by others (Backes et al., 1998; Kelley et al., 2006; Reed et al., 2010; Davydov et al., 2015; Davydov et al., 2017) may be best explained with a concept of "positional heterogeneity" in P450 oligomers (Davydov, 2011; Davydov et al., 2015; Davydov, 2016; Reed and Backes, 2017; Davydova et al., 2019). This concept is based on considering heteromeric complexes of multiple P450 species as a predominant state of the P450 enzymes in the microsomal membrane. According to our hypothesis the P450 subunits forming these oligomers are not identical in their conformation and orientation, but are characterized by different abilities to be reduced, bind substrates and interact with redox partners (Davydov et al., 2015;

Davydova et al., 2019). In effect, oligomerization results in abstracting an important part of the P450 pool from the catalytic activity. A discussion of possible structural basis for positional heterogeneity in P450 oligomers may be found in a comprehensive review by Reed and Backes (Reed and Backes, 2017).

From the standpoint of the above concept, the activation of one P450 enzyme by its interaction with another one might reveal a difference between the two proteins in their propensities for occupying the positions of two different types. The results of our experiments with incorporation of CYP2E1-BODIPY into CYP3A4-containing Supersomes (Fig. 7, Table 3) provide a strong evidence of high affinity interactions between CYP3A4 and CYP2E1 resulting in the formation of their heteromeric complexes. This conclusion is supported by our earlier studies with LRET-based technique (Davydov et al., 2019). It might be therefore hypothesized that, when CYP3A4 interacts with CYP2E1, the former preferentially occupies the "active" positions in the oligomers, whereas CYP2E1 fills the "restrained" ones. Resulting re-distribution of CYP3A4 between the "active" and "restrained" positions increases the active fraction of this enzyme in HLM and results in increased catalytic turnover of CYP3A4-specific substrates.

Variability of heterotropic cooperativity of CYP3A4 in HLM and its modulation by P450-P450 interactions

Four preparations of HLM examined in this study revealed notable differences in their response to the addition of ANF, a prototypical allosteric effector of CYP3A enzymes. Although the attenuation of the homotropic cooperativity with BQ and increase in the affinity to this substrate were revealed with all four samples, the effect of ANF on the maximal rate of BQ turnover was contrastingly different. Whereas in the case of HLM-N1 the addition of ANF substantially increased the maximal rate of reaction, it resulted in over two-fold inhibition in HLM-N2 while having no statistically significant effect in on $V_{\max}$ in HLM-N2 and HLM-A (Table 2). Interestingly, these differences are correlated with the differences in the CYP3A4 content. While the activated HLM-N1 preparation has the lowest CYP3A4 content over all three HLM-N samples, the inhibited HLM-N2 preparation is characterized with the highest CYP3A4 concentration.

Importantly, increase in the fractional content of CYP2E1, either as a result of chronic alcohol exposure or through incorporation of the exogenous protein, resulted in a substantial decrease in the degree of heterotropic cooperativity seen with BQ (Table 2).

Therefore, the effect of enrichment of the microsomal membrane with CYP2E1 on BQ metabolism reveals some parallelism with the effect of ANF.

The mechanisms of heterotropic cooperativity in CYP3A4 became a subject of extensive studies and a vigorous discussion over the last two decades. The initial interpretation of all known examples of CYP3A4 cooperativity, either homo- or heterotropic, is that the large substrate-binding pocket of the enzyme sometimes requires simultaneous binding of several substrate molecules to assure a productive orientation of at least one of them (see (Davydov and Halpert, 2008; Denisov et al., 2009; Denisov and Sligar, 2012) for review). The presence of at least two molecules of some substrates in the binding pocket of CYP3A4 is well established (Roberts et al., 2005; Fernando et al., 2006; Denisov et al., 2007). Although this model provides a reasonable explanation for most cases of homotropic cooperativity, it fails to explain complex instances of heterotropic activation in CYP3A4 (Davydov et al., 2013).

More recent studies compellingly demonstrated the presence of a separate allosteric effector binding site and revealed its role in the instances of heterotropic activation of CYP3A4 by various effectors, including ANF (Davydov et al., 2012; Davydov et al., 2013; Polic and Auclair, 2017; Denisov et al., 2018; Marsch et al., 2018; Denisov et al., 2019; Ducharme et al., 2019). This allosteric site is formed at the vicinity of the *F*' and *G*' helices and located at the interface between the two CYP3A4 molecules in the crystallographic dimer of CYP3A4 complex with peripherally-bound progesterone (PDB 1W0F (Williams et al., 2004)). As we discussed earlier (Davydov et al., 2013), the high-affinity interactions at this site require the protein to be oliogomeric – ligands are unlikely to bind with high affinity at this site in the monomeric enzyme due to small size and low degree of "buriedness" of the pocket formed by *F – F'* and *G – G'* loops. Therefore, if this peripheral site is involved in the mechanism of heterotropic activation of CYP3A4, the allosteric properties of the enzyme are expected to be determined by the degree of its oligomerization. A close interconnection between the allosteric properties of CYP3A4 and the degree of its oligomerization we demonstrated in our earlier study of the activating effect of ANF in model microsomes with variable surface density of CYP3A4 in the membrane (Davydov et al., 2013).

Location of the allosteric site at the interface between two subunits of the P450 oligomer suggests a critical dependence of allosteric properties of the enzyme in HLM on the formation of mixed oligomers between multiple P450 species. Although in some pairs of

CYP3A4 with interacting partners this allosteric site may be retained, its affinity to the ligands and the effect of their binding on the functional properties of CYP3A4 must be considerably modified by hetero-oligomerization. Our observation that the addition of ANF attenuates the ability of CYP2E1-BODIPY to form hetero-oligomers with CYP3A4 may be interpreted as an indication of decreased ability of CYP2E1-CYP3A4 complexes to bind this allosteric effector, as compared to CYP3A4 homo-oligomers. This difference may explain the attenuation of the effect of ANF on CYP3A4 in CYP2E1-rich microsomes demonstrated in this study.

Certainly, unveiling the sources of the effect of composition of P450 ensemble on CYP3A4 allostery requires further rigorous investigation. At the current state of knowledge we are far away from understanding the mechanisms of P450 hetero-association and their relationship to the allosteric properties of the P450 ensemble. Nevertheless, the model based on the hypothesis of positional heterogeneity and the involvement of the allosteric binding site located at the interface between two subunits of P450 oligomer provides a reliable explanation for a remarkable connection between CYP3A4 allostery and the composition of the P450 pool revealed in our studies.

Concluding remarks

Our study with human liver microsomes obtained from alcoholic donors demonstrated that chronic alcohol exposure results in a multifold activation of CYP3A4, the enzyme responsible for metabolism of wide variety of drugs currently on the market. This activation and the associated attenuation of homo- and heterotropic cooperativity of CYP3A4 can be reproduced in model HLM preparations, where the content of CYP2E1 was elevated by incorporation of the purified enzyme. These results reveal a profound effect of alcohol-induced increase in CYP2E1 content on the functional properties of CYP3A4 and suggest an important role of CYP2E1-CYP3A4 interactions in the mechanisms of the effects of alcohol exposure on drug metabolism. Furthermore, our findings compellingly demonstrate that the profile of human drug metabolism cannot be unambiguously determined from a simple superposition of the properties of the P450 species constituting the drug-metabolizing ensemble. This profile rather reflects a complex, "non-linear" combination of the functionalities of the individual enzymes which is largely affected by their intermolecular interactions and the formation heteromeric complexes.

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Authorship Contributions.

Davydov and Zgoda designed the study; Davydov wrote the paper; Dangi, Davydova, Vavilov and Davydov conducted the experiments; Dangi, Davydov, Vavilov and Zgoda analyzed the results.

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Table 1. Composition of the cytochrome P450 pool in HLM preparations used in this study*

Cytochrome P450 Species	Peptide used in the assay	HLM-N1		HLM-N2		HLM-N3		HLM-A		A-to-N ratio[b]
		Content, pmol/mg	Specific fraction, %[a]	Content, pmol/mg	Specific fraction, %[a]	Content, pmol/mg	Specific fraction, %[a]	Content, pmol/mg	Specific fraction, %[a]	
CYP1A2	ASGNLIPQEK	38.1	15.6	15.4 ±3.5	5.0 ±0.6	14.6 ±3.0	5.4 ±0.7	4.1 ±0.8	8.16 ±1.47	0.9
CYP2A6	GTGGANIDPTFFLSR	43.1	17.6	66 ±21	21.3 ±2.4	47.5 ±10.8	16.9 ±1.8	8.7 ±3.5	15.2 ±2.8	0.8
CYP2B6	GYGVIFANGNR	1.1	0.5	3.4 ±0.9	1.13 ±0.16	2.8 ±0.8	1.0 ±0.2	0.29 ±0.06	0.58 ±0.11	0.6
CYP2C8	GLGIISSNGK	5.1	2.1	2.6 ±3.6	0.56 ±0.69	4.5	1.1	0.91 ±0.23	1.93 ±0.33	1.1
CYP2C9	GIFPLAER	34	14	46 ±14	15.6 ±1.6	39 ±11	14.5 ±2.2	5.6 ±0.9	10.7 ±2.3	0.8
CYP2C18	IAENFAYIK	2.6	1.1	0.60 ±0.11	0.24 ±0.05	0.96 ±0.30	0.33 ±0.05	0.53 ±0.22	0.93 ±0.21	2.2
CYP2C19	ICVGEGLAR	7.8	3.2	0.82 ±0.74	0.28 ±0.06	0.86 ±0.21	0.33 ±0.06	0.25 ±0.12	0.59 ±0.29	0.6
CYP2D6	LLDLAQEGLK	1.1	0.4	2.0 ±0.9	0.51 ±0.05	2.2 ±1.1	0.59 ±0.16	0.75 ±0.56	0.98 ±0.37	1.9
CYP2E1	GIIFNNGPTWK	38	15.4	36 ±11	11.3 ±1.3	41 ±11	13.9 ±1.3	22.6 ±8.4	38.9 ±2.3	2.7
CYP3A4	LSLGGLLQPEKPVVLK	67	27.5	185 ±91	43.7 ±6.6	164 ±64	43.5 ±5.8	16.3 ±12.3	21.4 ±6.2	0.6
CYP3A5	DVEINGVFIPK	6.9	2.8	1.6 ±1.3	0.41 ±0.26	7.8 ±4.8	2.4 ±1.1	0.38 ±0.11	0.71 ±0.12	0.5
Total of all species		208	85.2	355 ±135	86.1 ±35.8	323 ±94	81.9 ±23.8	60.0 ±25.5	17.6 ±7.5	
P450 content[c]		244		405 ±95		394 ±53		341 ±70		
CPR (LC-MS)	NPFLAAVTTNR	58.7	101	42.6 ±10.7	68 ±17.0	42.1 ±9.3	82 ±18.0	44.3 ±17.3	127 ±50	
CPR content[c]		57.8		63.0 ±12.2		51.4 ±11.6		34.9 ±4.3		

* The values given for HLM-N2, HLM-N3 and HLM-A are the averages of 15, 20 and 11 individual measurements respectively. The measurement with HLM-N1 was performed only once. In each individual assay we determined the concentration of all P450 species under analysis, except for CYP2C8, which content in HLM-N1 and HLM-N3 was determined only once. The results of CYP2C8 determination in HLM-N2 and HLM-A represent the averages of 2 and 5 measurements respectively. The"±" values are the confidence intervals calculated for p=0.05.

[a] The values of specific content of P450 species are the averages of the values calculated for each individual dataset. The fractions shown for the total P450 content and the content of CPR are the percents of the amounts detected with LS-MS/MS from the amounts determined with absorbance spectroscopy (P450) and activity (CPR) assays.

[b] "A-to-N ratio" is the ratio of the fractional content of a given P450 species in HLM-A to an average of the respective contents in HLM-N samples. This ratio characterizes the direction and the amplitude of apparent alcohol-induced changes in the content of P450 enzymes.

[c] The values determined with absorbance spectroscopy (P450) and activity (CPR) assays and expressed in pmol per mg of protein.

<u>Table 2.</u> Parameters of BQ metabolism obtained with four HLM preparations and their modulation by α-naphthoflavone*

HLM preparation[a]	S_{50}, μM		N		V_{max}, min^{-1}	
	No effector	+ANF	No effector	+ANF	No effector	+ANF
HLM-N1	40.2 ±12.2	**10.7 ±4.8 (0.012)**	2.24 ±0.37	**0.83 ±0.12 (0.002)**	16.4 ±1.8	**24.9 ±2.8 (0.002)**
HLM-N1+2E1	**89.2 ±20.4 (0.012)**	**12.4 ±4.3 (0.012)**	**1.26 ±0.12 (0.009)**	**0.87 ±0.16 (0.020)**	**31.9 ±5.0 ($<10^{-3}$)**	**19.9 ±2.7 (0.014)**
HLM-N2	36.7 ±10.3	**7.4 ±3.8 (0.006)**	2.09 ±0.44	**1.22 ±0.30 (0.033)**	71.4 ±11.4	**32.6 ±3.5 (0.003)**
HLM-N2+2E1	**89.1 ±34.3 (0.031)**	**12.0 ±4.7 (0.014)**	**1.28 ±0.09 (0.011)**	1.02 ±0.24 (0.068)	83.7 ±38.8 (0.573)	46.1 ±11.1 (0.131)
HLM-N3	63.3 ±3.2	**19.7 ±6.7 ($<10^{-3}$)**	**1.92 ±0.02**	**1.02 ±0.37 (0.010)**	45.7 ±5.1	49.3 ±7.9 (0.528)
HLM-N3+2E1	107 ±55 (0.248)	**16.3 ±6.9 (0.019)**	**1.13 ±0.03 ($<10^{-6}$)**	**0.86 ±0.08 (0.003)**	**77.3 ±4.9 (0.001)**	46.8 ±16.6 (0.082)
HLM-A	59.6 ±12.5	**25.0 ±10.7 (0.006)**	1.02 ±0.15	**0.99 ±0.20 (0.025)**	101 ±20	113 ±24 (0.934)
HLM-A+2E1	65.4 ±25.5 (0.665)	**23.0 ±13.1 (0.044)**	1.47 ±0.30 (0.295)	**0.85 ±0.26 (0.033)**	**147 ±30 (0.028)**	127 ±4 (0.318)

* The values given in the table are the averages of 3 - 7 individual measurements. The"±" values are the confidence intervals calculated for p=0.05. The values in parentheses represent the p-values of Student's t-test for the hypothesis of equality of the respective values with the value obtained with untreated microsomes (for the values in "No effector" columns) or at no ANF added (for the values in "+ANF" columns). The values characterized with p-values ≤0.05 are shown in bold to emphasize the effects with high statistical significance.

[a] The CYP2E1-enriched samples represented in this table are the samples where the content of added CYP2E1 was equal to the total P450 content in untreated HLM.

Table 3. Effect of membrane incorporation of CYP2E1-BODIPY on the degree of its homo-oligomerization explored with homo-FRET*

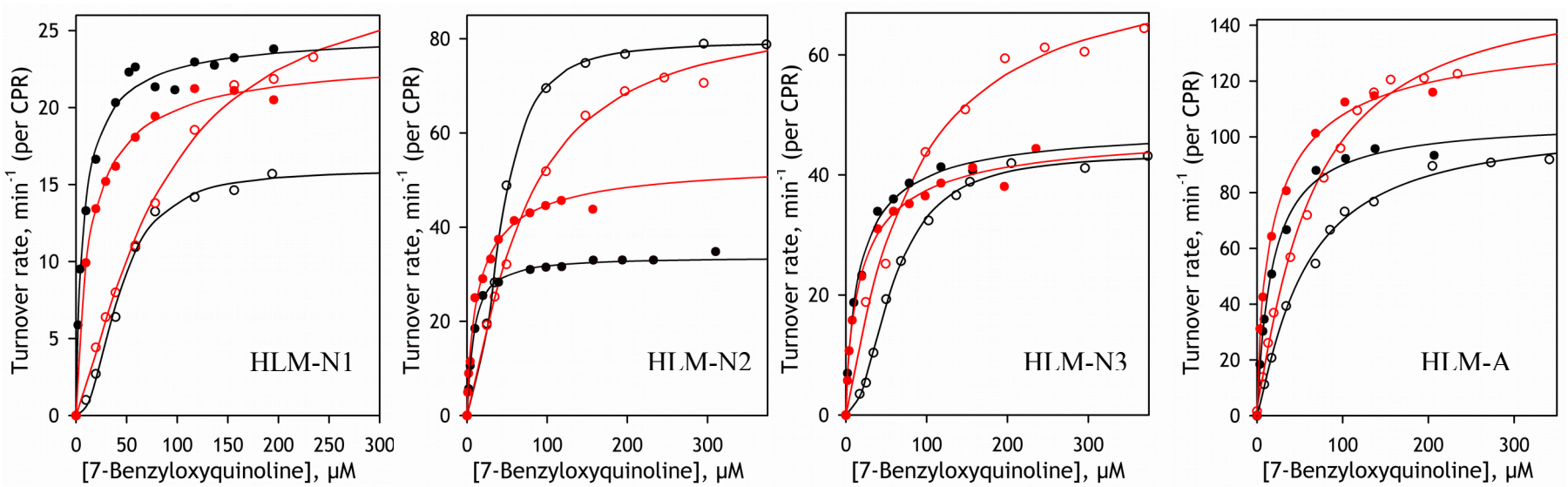
Membranous preparation	$[CYP2E1]_{50\%}$[a], pmol/cm^2	$(Lipid/CYP2E1)_{50\%}$[b]	FRET Efficiency, %
SS(CPR)	0.024 ± 0.005	14,400 ± 2,850	84 ± 10
SS(3A4)	2.0 ± 0.15	176 ± 14	74 ± 9
SS(3A4)+ANF	0.58 ± 0.06	604 ± 62	84 ± 20

[*] The values given in the Table were obtained by fitting of the titration curves with the binary association equation (Eq. 2) in combination with Eq. 3 that defines the relationship between the relative increase in the intensity of fluorescence and the steady-state concentration of the homo-oligomers. The "±" values show the confidence interval calculated for $p = 0.05$.

[a] Surface density of P450 in the membranes at which the amplitude of the titration curves reaches 50% of the maximal level. This value is equal to the value of K_D determined from the fitting of the titration curves with Eq. 2 combined with Eq. 3. The surface density was calculated from the RL/P ratio as described in Materials and Methods using Eq. 1.

[b] Lipid-to-P450 ratio at which the amplitude of the titration curves reaches 50% of the maximal level which was calculated from the respective $[CYP2E1]_{50\%}$ values.

[c] FRET efficiency was calculated from the values of R_{max} (Eq. 3) according to relationship given by Eq. 4.



<u>Fig. 1.</u> BQ metabolism in intact and CYP2E1-enriched HLM samples. The figure show the dependencies of the rate of BQ turnover on substrate concentration in intact (black) and CYP2E1-enriched (red) HLM preparations. The dependencies obtained in the absence of added effector are shown in open circles, while the data obtained in the presence of 25 μM ANF are designated by closed circles. The CYP2E1-enriched samples exemplified in this figure are the samples where the content of added CYP2E1 was equal to the total P450 content in HLM ("1:1 incorporation"). The solid line show the results of fitting of the datasets with the Hill equatrion.

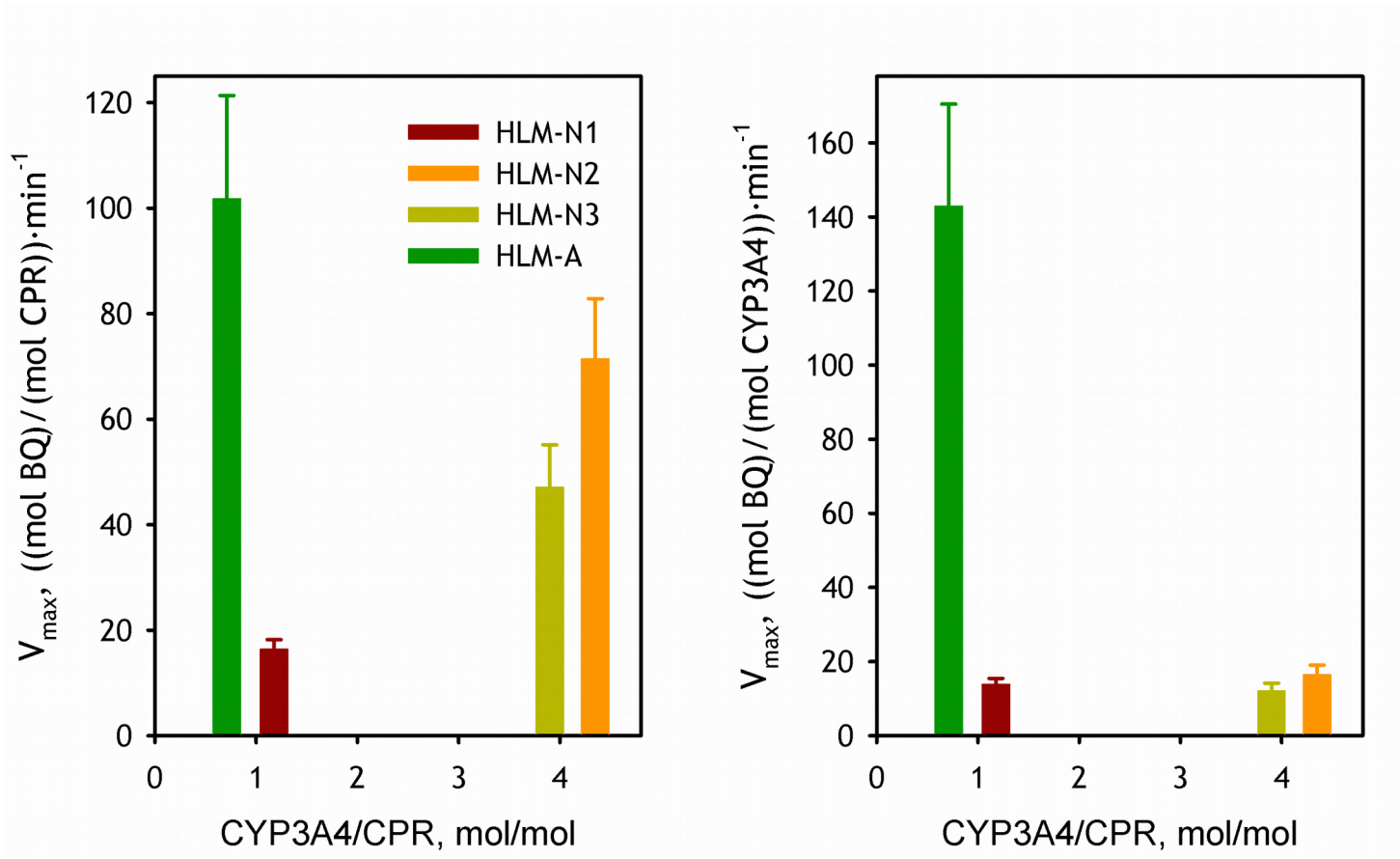


Fig. 2. The relationship between the CYP3A4 content in HLM preparations and the rate of BQ metabolism an no added effector. The maximal velocity of BQ debenzylation is plotted against the molar ratio of CYP3A4 to CPR in microsomal membranes calculated from the results of LC-MS/MS analysis (Table 1). Left panel shows the V_{max} values normalized on the concentration of CPR, while the right panel shows the same results normalized per CYP3A4 content.

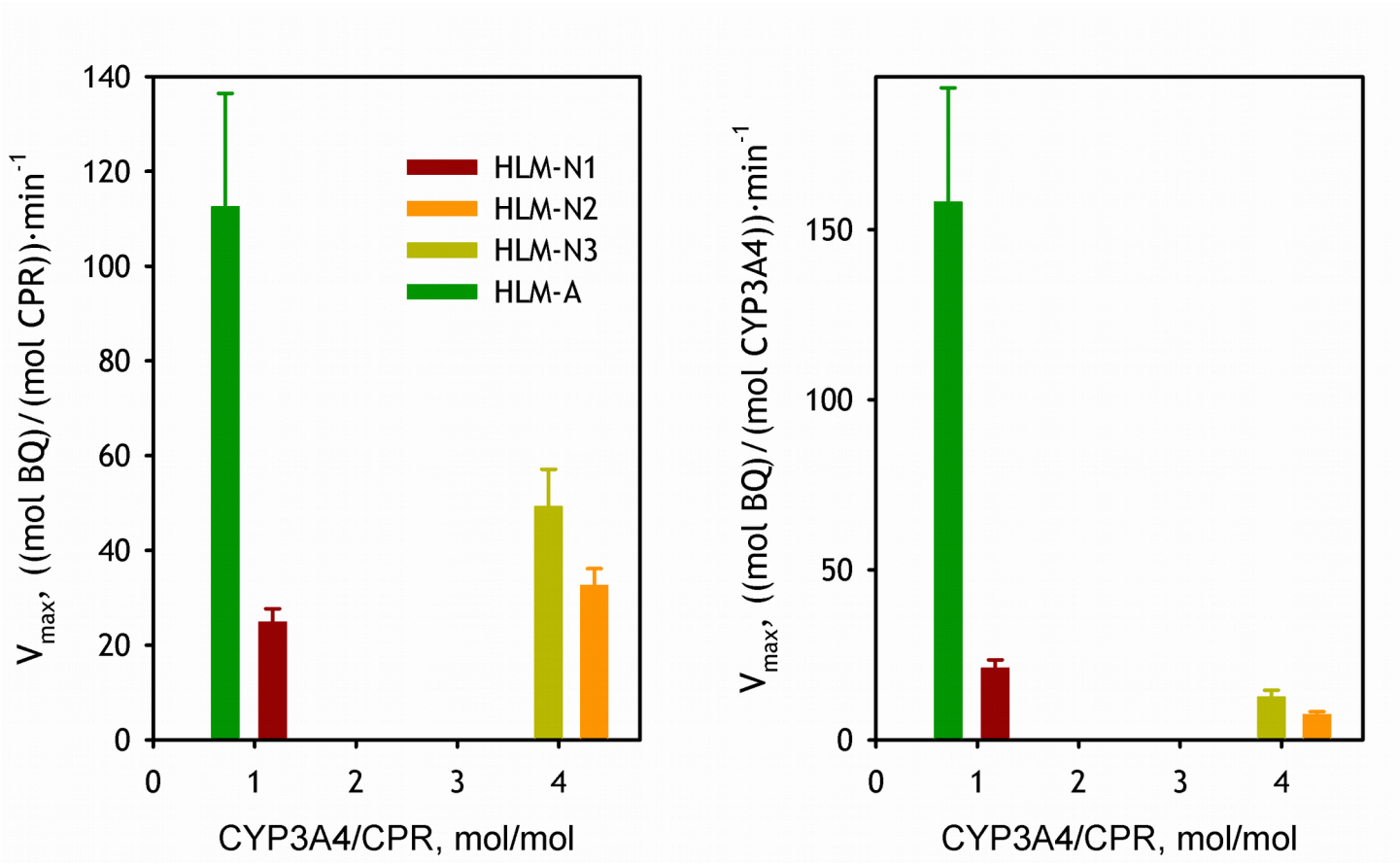
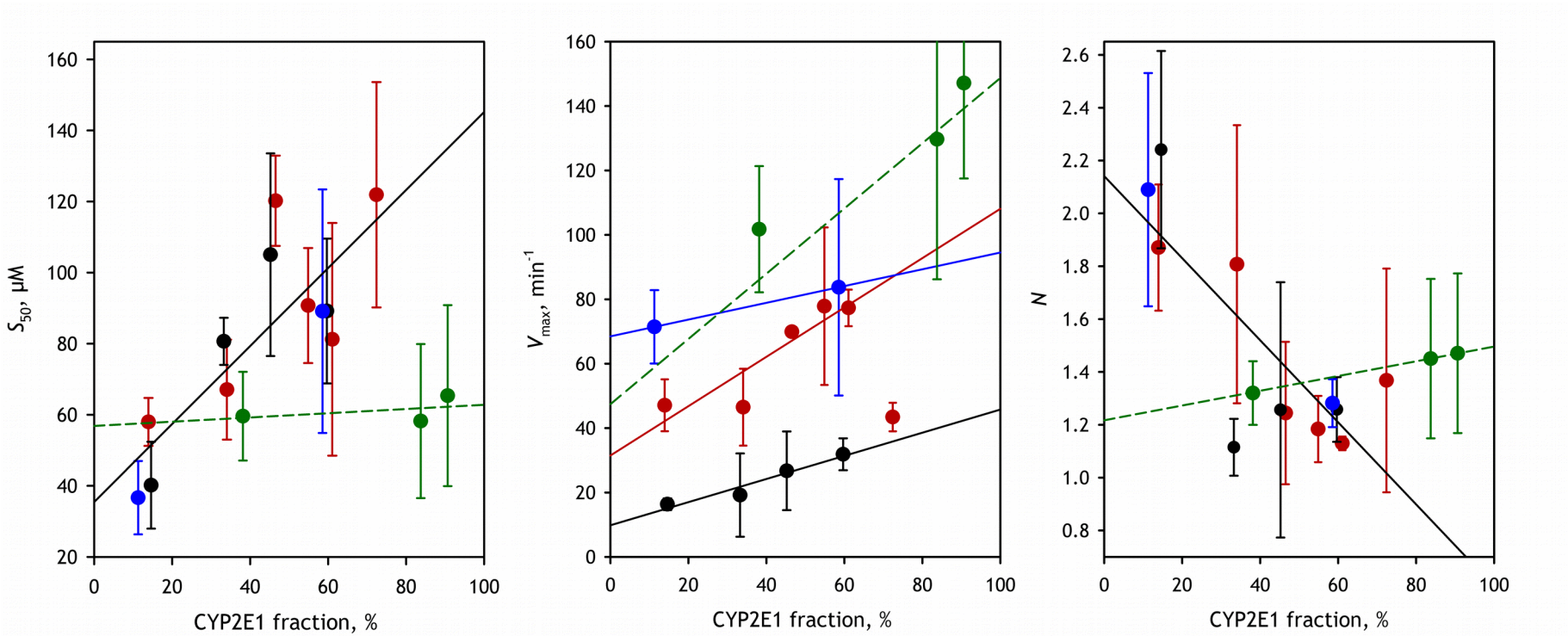
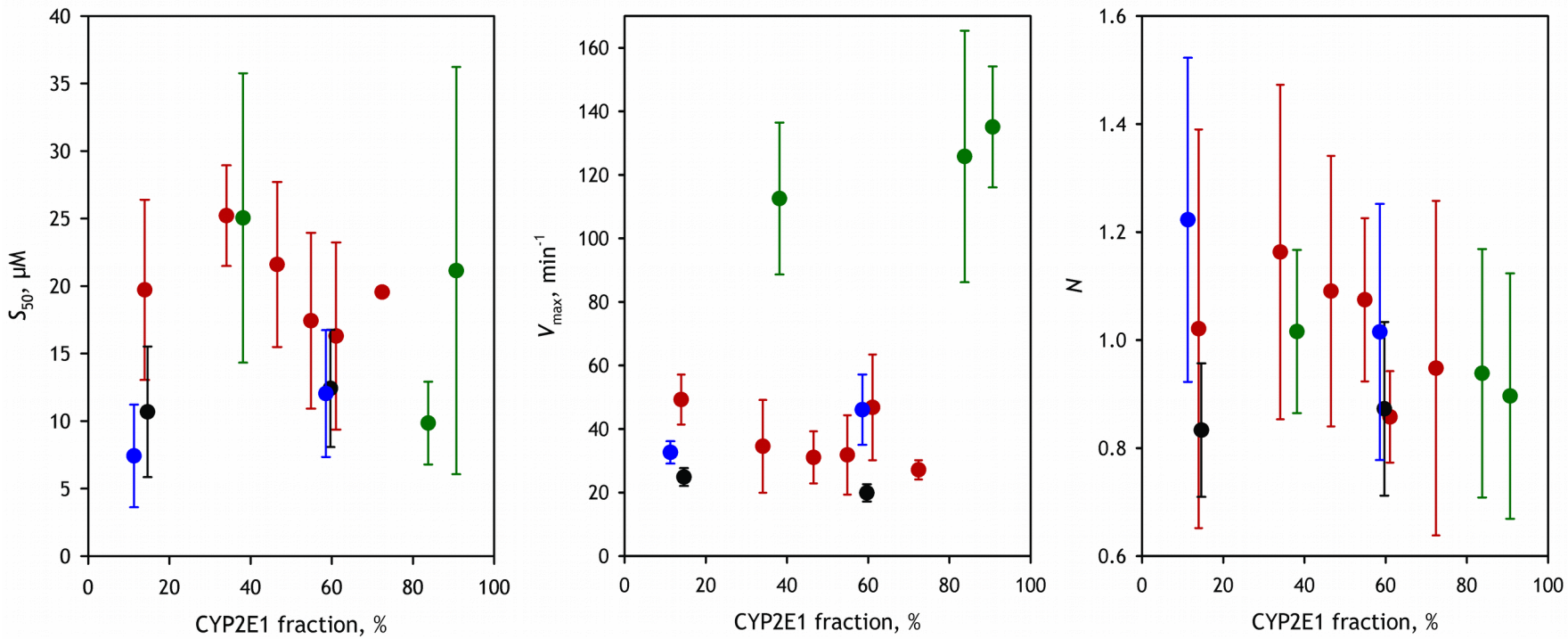


Fig.3. **The relationship between the CYP3A4 content in HLM preparations and the rate of BQ metabolism in the presence of 25 µM ANF.** The maximal velocity of BQ debenzylation is plotted against the molar ratio of CYP3A4 to CPR in microsomal membranes calculated from the results of LC-MS/MS analysis (Table 1). Left panel shows the V_{max} values normalized on the concentration of CPR, while the right panel shows the same results normalized per CYP3A4 content.



<u>Fig.4.</u> Effect of incorporation of CYP2E1 into HLM-N on the parameters of BQ metabolism at no added effector. Dependencies of S_{50}, V_{max} and N on the fractional content of CYP2E1 in the total of 11 analyzed P450 species. The datasets obtained with HLM-N1, HLM-N2 and HLM-N3 are shown in black , blue and red respectively. The error bars show the confidence intervals calculated for p=0.05. Solid lines show the linear approximations of the data.



<u>Fig.5.</u> Effect of incorporation of CYP2E1 into HLM-N on the parameters of BQ metabolism in the presence of ANF. Dependencies of S_{50}, V_{max} and N on the fractional content of CYP2E1 in the total of 11 analyzed P450 species. The datasets obtained with HLM-N1, HLM-N2 and HLM-N3 are shown in black , blue and red respectively. The error bars show the confidence intervals calculated for p=0.05.

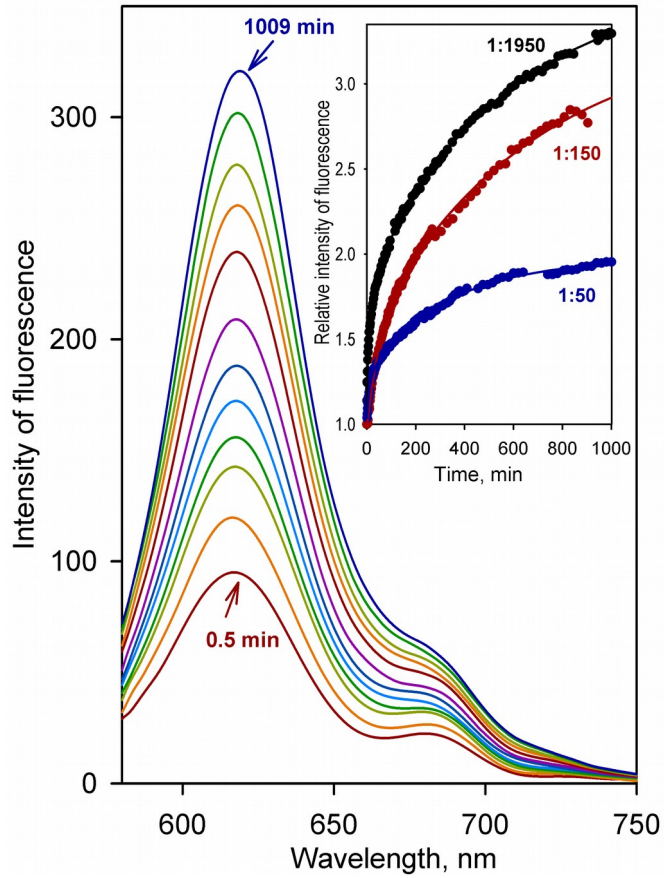


Fig. 6. Process of dissociation of homo-oligomers of CYP2E1-BODIPY after their incorporation into microsomal membrane monitored by the changes in BODIPY fluorescence. The spectra shown in the main panel were recorded in the process of incubation of 0.081 μM CYP2E1-BODIPY with SS(3A4) added to the phospholipid concentration of 158 μM ($R_{L/P}$=1950). The spectra were recorded at 0.5, 2, 8, 16, 34, 65, 126, 253, 382, 510, 766 and 1009 min after addition of Supersomes to CYP2E1-BODIPY. Panel ***c*** shows the changes in the relative intensity of fluorescence (Re) during incubations performed at $R_{L/P}$ ratios of 50 (blue), 150 (red) and 1950 (black). The solid lines represents the approximations of the datasets with a three-exponential equation.

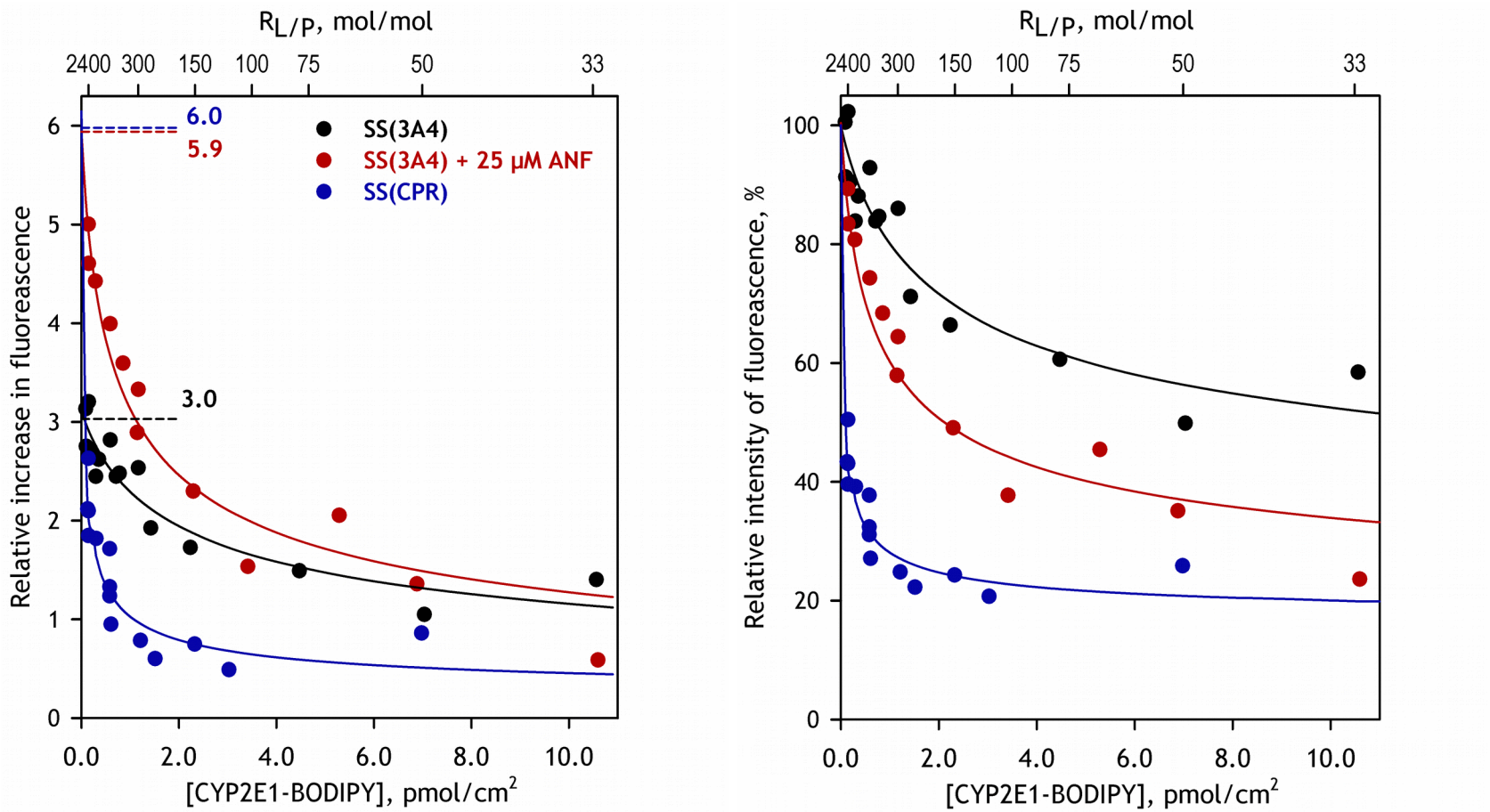


Fig. 7. The dependencies of the changes in fluorescence caused by incorporation of CYP2E1-BODIPY into Supersomes on the concentration (surface density) of CYP2E1-BODIPY in the membrane. The datasets obtained with SS(CPR), SS(3A4) at no effector added and SS(3A4) in the presence of 25 μM ANF are shown in blue, black and red respectively. Solid lines show the approximations of the data sets with a combination of Eq. 2 and Eq. 3. Left panel shows the changes in the relative increase in the intensity of fluorescence (R_E in Eq. 3). Dashed lines shown in this panel indicate the maximal level of increase (R_{max}) estimated from the fitting. Right panel shows the same datasets scaled to the percent of maximal fluorescence..