

1 Assessment of Potential Health and Genetic Impacts in Releasing

2 Confiscated *Paroaria coronata* and *Saltator similis*

3 Short Title: Potential Health and Genetic Impacts in Releasing of Confiscated Songbirds.

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34

35 **Abstract**

36 Illegal capture and trade of wild birds has long been a threat to biodiversity.
37 Translocation—the release of individuals from one location into another—is a useful
38 conservation tool in the management of species. However, both health (such as different
39 pathogens) and adaptive (such as local adaptation), differences among populations must
40 be taken into account, as both can impact the recipient population negatively. Here, we
41 provide health and genetic information to support release planning for two of the most
42 trafficked Brazilian wild bird species (*Paroaria coronata* and *Saltator similis*). We
43 focused on two fundamental questions: Are there significant differences in pathogen load
44 between wild and captive populations? Is there significant genetic structure among
45 populations? In total, 223 free-living birds were captured, sampled, and released at the
46 same site. Devices and live decoys characteristics were top factors influencing captures.
47 We tested blood, feces, and oropharyngeal swabs from free-ranging (n=101) and
48 confiscated (n=92) birds for Newcastle disease virus, *Salmonella* spp., and *Mycoplasma*
49 *gallisepticum*. Genetic structure among populations was investigated using mtDNA in a
50 subsample of these birds. We found no evidence for Newcastle disease virus and
51 *Salmonella* spp. in seized and free-living birds from both species. However, seized *P.*
52 *coronata* and *S. similis* may be potential sources of *M. gallisepticum*. We found
53 significant but low genetic structure among populations occurring in different Biomes
54 ($\Phi_{CT}=0.26$ for *P. coronata*; $\Phi_{CT}=0.13$ for *S. similis*) and no significant structure among
55 populations occurring in the Pampa Biome. These results suggest that while it may be
56 important to screen seized birds for avian pathogens, genetic structure among populations
57 seems to be of lesser concern when planning the release of seized songbirds in the wild.

58

59 **Introduction**

60 Illegal trade, along with poaching, habitat loss, and pollution, are top causes of wild
61 bird decline in Brazil and in other developing countries that harbor a huge species
62 diversity [1–4]. The illegal wildlife trade has increased dramatically over the past decade,
63 along with enforcement efforts aimed at mitigating this threat [5]. For confiscated wild
64 birds, despite the shortage of studies on the subject, current translocation guidelines
65 include as an option the euthanasia of individuals belonging to species of low
66 conservation value [6,7]. In part, this policy can be justified on the grounds that a) seized
67 birds may harbor pathogens that will affect the wild population [8] and b) usually, seized
68 birds come from an unknown parental population, and their release in another population
69 may result in outbreeding depression [9,10], resulting, in both cases, in a negative impact
70 on the wild population. Health and genetic studies in wildlife species depend on direct
71 capture for sampling free-ranging individuals and collecting relevant data to evaluate the
72 sanitary and genetic risk of planned translocations. Ideally, such knowledge would allow
73 the translocation of seized and rehabilitated birds to their most probable area of origin,
74 with minimal health risks to the local populations and to the commercial flocks [5–8].
75 Unfortunately, both genetic and health information on the subject is scarce. Despite the
76 myriad of methods and tools available, mist nets are globally recognized as the top capture
77 method in biological studies [11–13] and are increasingly applied in association with bird
78 lure techniques to improve efficiency, mostly through playback audio [14]. In this sense,
79 the use of artificial [15,16] or live decoys [17], although routinely and widely used in
80 poaching, is only occasionally presented on scientific reports, albeit extensively
81 disseminated in the virtual media.

82 In this study, we describe the methods for capturing and sampling wild *Paroaria*
83 *coronata* (Red-crested Cardinal) and *Saltator similis* (Green-winged Saltator)—two of
84 the most heavily trafficked Brazilian wild bird species, which are classified as of least

85 concern on the IUCN red list [18]. Next, we generate sanitary and genetic data to answer
86 two fundamental questions: Are there significant differences in pathogen load between
87 wild and captive populations? Is there significant genetic structure among populations?
88 Finally, we discuss the potential impacts of a translocation program of confiscated
89 conspecifics for both species.

90

91 **Materials and Methods**

92 **Ethical statement**

93 This project (no. 23644) was approved by the Ethics Committee on the Use of
94 Animals—UFRGS—and licensed by the *Instituto Chico Mendes de Conservação da*
95 *Biodiversidade* (ICMBio), under the number 37567. Samples received from Argentina
96 were registered under the export permit code EX-2019-12969382-APN-DNBI#SGP.

97

98 **Sampled areas**

99 Sampling areas were selected to include representative areas of the species' usual
100 distribution, mostly in Rio Grande do Sul state but also in strategic areas from the total
101 species' distribution (Fig. 1, S1 Table). We captured *S. similis* in two different regions:
102 Pampa and Atlantic Forest. Even though the Pampa is associated with open grasslands
103 [19], we found *S. similis* always associated with forest paths as well as in the Atlantic
104 Forest region. On its turns, we captured *P. coronata* in three different regions: Pampa,
105 Atlantic Forest, and Chaco/Pantanal. In the Atlantic Forest, this species was associated
106 with anthropic deforested paths. The Chaco/Pantanal region is characterized by savannah
107 with a substantial forest component, especially in the Chaco, whose more humid portions
108 are connected to the seasonally flooded Pantanal plain in Brazil [20–22]. Sample size was
109 determined based on both pilot expeditions and on similar studies [23,24]. For the genetic

110 analyses we also included sequences from the GenBank (Bolívia n=1, Boracéia n=1,
111 Corrientes n=2, Mato Grosso n=1) and an additional five *P. coronata* samples were
112 received from the Buenos Aires area, Argentina.

113

114 **Fig. 1.** Geographical representation of sampled areas.

115

116 **Management of live decoys**

117 Dominant *P. coronata* and *S. similis* of both sexes, identified through bird song
118 defiance (playback) among groups of birds seized from illegal traffic at the *Centro de*
119 *Triagem de Animais Silvestres* (CETAS/IBAMA), Porto Alegre, RS were tentatively
120 selected, tested, and kept as live decoys for attracting free-ranging conspecific birds.
121 Upon joining the live decoy flock, birds had three consecutive fecal samples negative for
122 parasites. Birds with positive samples were treated (anthelmintic and anticoccidial drugs).
123 Only birds that systematically attracted conspecific birds in fieldwork were kept. Birds
124 that failed 2–4 times consecutively were returned to CETAS. Seed mixtures, commercial
125 rations, fruits, vegetables, arthropods, and minerals composed the birds' diets. During the
126 capturing time (breeding season—August to February), the birds were kept in cages, and
127 in the rest of the year, they were released in 12–24 m³ outdoor aviaries. Bath bowls,
128 sunbath, and cage tray cleaning (sand bedding exchange) were performed every other
129 day.

130

131 **Fieldwork**

132 Initially, we used mist nets and cages mounted with net-trapdoors (Fig. 2A) for
133 capturing both species. However, after successive unsuccessful attempts to capture *P.*
134 *coronata*, a bird trap-loop (Fig. 2B and 2C) was built according to an Indochinese model

135 demonstrated on the internet [25], and used next to the decoy's cage. All capturing
136 processes were timed.

137

138 **Fig. 2.** Schematic representation of capturing devices. A: cage with live decoy *Saltator*
139 *similis*, showing details of spring that closes the net-trapdoor (1), net-trapdoor (2), trigger-
140 perch (3), and metal hook holding the trigger-perch mounted and the net-trapdoor opened
141 (4). B: loop with its parts numbered as the spring that pulls the fishing-rod which pulls
142 the loop (5), fishing-rod (6), pin holding the trigger-perch mounted and fishing-rod (7),
143 trigger-perch (8), brackets (nylon line folds) holding the loop open (9 and 10), and spring
144 holding the trigger-perch up (11). C: *Paroaria coronata* captured in the loop.

145

146 Initial sampling was mostly conducted for studying genetic structure in both
147 species. Blood samples (0.1 ml) were obtained by right jugular vein puncture with an
148 insulin syringe [26] and conserved in FTA cards for subsequent genetic applications. A
149 second round of capture expeditions was conducted for obtaining most samples for the
150 sanitary study, in which 0.1 ml of blood (stored in Eppendorf tubes for serum separation),
151 oropharyngeal swab, and feces (mostly after keeping the bird in a holding bag for up to
152 30 min) were obtained. Such samples were tested for *M. gallisepticum* (MG) and
153 Newcastle disease virus (NDV) serology, MG-PCR, and *Salmonella* spp. isolation,
154 respectively. Samples for the sanitary studies were obtained from populations located up
155 to 200 km from Porto Alegre, to make it possible to deliver them in the laboratory in the
156 same day of collection. Samples were kept under refrigeration. The sampling period
157 included the reproductive seasons of 2017, 2018, and 2019. After sampling, birds were
158 banded and released in the same place of capturing. Additional data is shown in S1 Table.

159

160 **Sanitary tests**

161 The microbiological, serological, and molecular tests were performed at the Porto
162 Belo Laboratory, accredited by the Brazilian Ministry of Agriculture, Livestock, and
163 Supply (*Ministério da Agricultura, Pecuária e Abastecimento*—MAPA) to perform
164 official diagnostic tests within the National Plan of Avian Sanity (*Plano Nacional de*
165 *Sanidade Avícola*—PNSA).

166 Anti-NDV antibodies were searched for in serum samples by the Hemagglutination
167 Inhibition Test according to the SDA ordinance n° 182, 08 November 1994. *Salmonella*
168 spp. detection followed the methods presented in SDA ordinance n° 126, as of November
169 03, 1995, after replacing BHI broth by buffered pebble water 1%. Suspected colonies
170 were confirmed and characterized with specific antisera (serum agglutination on blade)
171 at the Fundação Oswaldo Cruz, according to World Health Association protocols [27].
172 For the detection of anti-MG antibodies, serum samples were tested by a rapid plate
173 agglutination (RPA) test according to SDA ordinance n° 44, as of November 08, 2001. A
174 real-time polymerase chain reaction assay using a Taqman-labeled probe for the detection
175 of *M. gallisepticum* (commercial kit MG—NewGene®) DNA [28] was applied, and the
176 positive results were sent for confirmation with a Multiplex real-time PCR [29] at Simbios
177 Biotechnology.

178 Serum samples from three *P. coronata* and three *S. similis* vaccinated against MG
179 and NDV served as controls for validation of the MG-qPCR and NDV-HI. Serum samples
180 from these birds were tested before and after vaccination to serve as negative and positive
181 controls, respectively. Vaccination protocols included one ocular drop from live vaccines
182 *M. gallisepticum* (MYCOVAX-TS-11, MERIAL, strain TS-11, P.400/17, V.02/18,
183 November 11, 2017) and NDV (MERIAL, strain La Sota 004/16, ND1873, P.004/16,
184 V.04/18, December 01, 2017), boosters with live vaccines for MG (BIOCAMP, Camp

185 VacMG-F, P.006/2016, V.11/19, February 02, 2018) and NDV (BIOVET, New-Vacin,
186 La Sota, P.007/17, V.05/19, February 23, 2018). An additional booster included an oil
187 inactivated NDV (BIOVET, New-BRONK-VET, P.003/17, V.08/19, virus B1 La Sota,
188 minimum titre before inactivation $10^{5.3}$ DIOE₅₀, March 23, 2018). Protocol was based on
189 a previous study on NDV vaccination of wild birds [30]. For the sanitary analyses, we
190 also included samples from birds confiscated from illegal handlers.
191 <http://dx.doi.org/10.17504/protocols.io.bac3iayn>. [PROTOCOL DOI].

192

193 **Genetic analysis**

194 In order to understand the genetic structure and diversity of *P. coronata* and *S.*
195 *similis*, we compared individuals from the Pampa region, in which both species occur in
196 relative abundance, with *P. coronata* individuals from the Chaco/Pantanal region and *S.*
197 *similis* from the Atlantic Forest region. There were few samples of *P. coronata* from the
198 Atlantic Forest and of *S. similis* from the Chaco/Pantanal because these are not major
199 occurrence regions for these species. This strategy also allowed us to compare two regions
200 of more similar vegetation patterns (Pampa vs. Chaco/Pantanal, in the case of *P.*
201 *coronata*) with two regions with more different vegetation patterns (Pampa vs. Atlantic
202 Forest, in the case of *S. similis*). Further information about the genetic sampling is given
203 in S2 Table.

204 DNA was extracted from the blood in FTA cards using the PureLink Genomic DNA
205 Mini Kit (Invitrogen), and the fragment of the mitochondrial gene NADH dehydrogenase
206 subunit 2 (*ND2*) was amplified using the same PCR protocol for both species. The
207 reaction was performed with concentrations of 20 ng/μl of DNA, 1x PCR Buffer
208 (Invitrogen), 3.5 mM MgCl₂, 0.2 mM dNTPs, 0.2 pmol/μl of each primer, and 0.04 U/μl
209 of Taq Platinum DNA Polymerase (Invitrogen). The primers used were MetL (described

210 by Hailer et al. [31], sequence 5'-AAGCTATCGGGCCCATACCCG-3') and RND2A
211 (this study, sequence 5'-CCTGAGTTGCATTYAGGGG-3'), and the PCR conditions
212 were as follows: 94°C for 2 min, 35 cycles of 94°C for 30 s, 59°C for 30 s, 72°C for 60
213 s, and a final extension of 72°C for 8 min. The amplification was confirmed through
214 electrophoresis in a 1% agarose gel. The amplified products purified enzymatically with
215 exonuclease I (GE Healthcare) and Sanger sequenced by ACTGene Inc., Brazil.

216

217 **Data analysis**

218 For field work data, descriptive variables for each capture were recorded in a
219 spreadsheet (S1 Table). A principal component analysis was performed based on Gower's
220 universal similarity index using Past v.3 (University of Oslo, Oslo, Norway.
221 <https://folk.uio.no/ohammer/past/>). We used the Spearman correlation coefficient to
222 relate the principal component with each of the studied variables. In addition, the
223 association among categorical variables was studied using the Pearson chi-square test for
224 association. These tests were conducted in Minitab v. 18 (State College, Pennsylvania,
225 USA. <http://www.minitab.com>) using 0.05 as the significance threshold. Responses were
226 also accessed using descriptive statistics.

227 For molecular data, DNA sequences were assembled and aligned in the software
228 Geneious v.10.2.3 (<https://www.geneious.com>) and checked by eye in MEGA X v.10.0.0
229 [32]. We used DnaSP v.6.12.03 [33] to define all different haplotypes whose evolutionary
230 relationships were represented using a Median-joining network [34], as estimated in the
231 software PopART (<http://www.popart.otago.ac.nz>). Standard genetic diversity indices,
232 including Tajima's D [35] and Fu's FS [36] neutrality tests were estimated using the
233 program Arlequin v.3.5.2.2 [37]. We also used this program to quantify the level of
234 genetic structure using both hierarchical and non-hierarchical Analysis of Molecular

235 Variance (AMOVA) [38]. The hierarchical AMOVA was performed for each species
236 using the three collection regions (Pampa, Atlantic Forest, and Chaco/Pantanal). Non-
237 hierarchical AMOVA was performed only for the Pampa region to explicitly compare the
238 level of structure in the same region using a comparable sample strategy for both species.

239 Finally, to understand the past demography of both species, Bayesian Skylines plots
240 (BSP) [39] were generated for the total population and the main occurrence regions of
241 each one of them, using BEAST v.2.6.1 [40]. We used 10,000,000 MCMC steps,
242 sampling every 1,000 steps and discarding the initial 10% of the sampling as burnin. We
243 used a partition scheme allowing each codon position to have a different substitution
244 model, which were estimated in MEGA X v.10 [32]. We checked sampling sufficiency
245 of the MCMC and built the BSP in Tracer v.1.7.1 [41]. The molecular substitution rate
246 for the ND2 gene was calculated as described previously [42], assuming the “calibration
247 set 2” for a 45g bird [43,44].

248

249 **Results**

250 **Capturing birds in the wild**

251 In total, we performed 223 captures: 122 Saltators and 101 Cardinals (S1 Table).
252 Net-trapdoors accounted for 36% of the captures, of which 97% were *S. similis* (Fig. 3).
253 Catches using the loop method were 33% of the total, with a high prevalence of *P.*
254 *coronata* (89%). The mist nets provided comparable catches in both species under study,
255 with 53% being *S. similis*.

256

257 **Fig. 3.** Frequency and duration of each capture according to bird species and capturing
258 method.

259

260 On average, it took 41.7 minutes (± 1.69 minutes) for each bird capture, with similar
261 values between species. Catches using the mist net were faster (32.8 ± 2.9 minutes)
262 compared to captures performed with the net-trapdoor (43.1 ± 1.9 minutes) or loop (48.5
263 ± 3.6 minutes) methods. The mist net also accounted for 62% of the captures performed
264 in less than 25 minutes, which correspond to the first quartile of the database. The
265 principal component analyses show that capture method was the most important factor
266 discriminating between *P. coronata* and *S. similis* captures (Fig. 4).

267

268 **Fig. 4.** Principal component analysis indicating captures (dots symbols) and the variables
269 having the highest explanatory power (lines).

270

271 Characteristics of the decoy were associated to all studied variables in the captured
272 birds (Table 1), showing an important effect of the decoy in the capturing process. Some
273 of these associations were very clear even before the statistical analysis. For instance, all
274 captures involved conspecific decoys. Male decoys accounted for 99% of the captures of
275 *S. similis*, while the captures of *P. coronata* were well distributed among decoy sexes
276 (couple: 42%, male: 33%, and female: 25%). Among the female birds sampled, only 26%
277 were captured using female decoys. A total of 60% of the female birds were captured
278 using loop (almost exclusively *P. coronata*), while 75% of the male birds were captured
279 using net-trapdoor or mist net, even though the low capture of female and young Saltators
280 may bias these results. Decoys with a high ranking for dueling disposition and vocal
281 performance (S3 Table) accounted for 96 and 98% of the fastest captures (performed in
282 less than 25 minutes), respectively. Seven birds (~3%) died due to capturing/sampling
283 procedures.

284

285 **Table 1.** Probability of association between characteristics of the captured bird (sex,
286 species, and age—or duration of the capture) with characteristics of the decoy or with the
287 method applied in the catch

288

Decoy / Method	Captured bird / Duration of capture			
	Sex	Species	Age	Duration ²
Sex	<0.001	<0.001	0.001	0.024
Species	<0.001	<0.001	0.009	<0.001
Dueling disposition ³	<0.001	0.047	0.015	0.037
Vocal. performance ³	0.027	<0.001	0.005	0.045
Method	<0.001	<0.001	0.095	<0.001

289

290 Probabilities obtained in the Pearson chi-square test for association. Significant values
291 are shown in bold. ²Duration was ranked in four classes according to the quartiles. ³Each
292 decoy was ranked in three classes according to its behavior and vocalizations during
293 captures (S3 Table).

294

295 **Sanitary tests**

296 Data on the PNSA-associated tests performed on samples from both free-ranging
297 and confiscated *P. coronata* and *S. similis* are presented in S4 Table. The serological
298 study revealed no presence of antibodies against Newcastle disease, irrespective of the
299 origin of the bird. *Salmonella* spp. isolation resulted in only one positive sample: *S.*
300 *enterica* serovar Cerro from a free-ranging *P. coronata*. Two tests were applied for the
301 detection of *M. gallisepticum* (MG). The RPA test indicated positive results for 25% of
302 the samples from free-ranging birds, but none was confirmed by the PCR assay. On the
303 other hand, seized birds showed positive results for MG in the RPA test in 53% of the
304 samples, which was reduced to 13% when samples were tested by PCR. Most
305 disagreement between the tests were 'false-positive' results. Despite this variation, results
306 obtained by the RPA method were highly correlated ($P < 0.001$) with the results obtained
307 by PCR when assessed by a Pearson chi-square test.

308

309 **Genetic structure and diversity**

310 DNA sequence analysis resulted in an alignment of 977 bp and 810 bp for *P.*
311 *coronata* and *S. similis*, respectively. All new sequences were deposited in GenBank (S2
312 Table). Genetic diversity indices for both species in all sampled regions are shown in
313 Table 2. Overall, there was higher genetic diversity in *S. similis* than in *P. coronata* for
314 both the whole sample as well as in the Pampa region. However, we found contrasting
315 patterns between species when the two major regions of occurrence were compared.
316 While for *S. similis*, the Pampa and Atlantic Forest have comparable levels of diversity,
317 for *P. coronata*, the Pampa has only a fraction of the diversity contained in the
318 Chaco/Pantanal region, which is also clear from the haplotype network (Fig. 5). For both
319 species, the sample size in regions of minor occurrence (Atlantic Forest in the case of *P.*

320 *coronata* or Chaco/Pantanal in the case of *S. similis*) is too low to allow for a through
321 characterization of the genetic diversity.

322 **Table 2.** Standard genetic diversity indices for *P. coronata* and *S. similis*.

	Source	Sample size	N Loc	Segregating sites	N Haplot	Haplotype diversity ¹	Nucleotide diversity ¹	Tajima's D ²	Fu's Fs ²
	Pampa	43	8	7	6	0.41 (0.09)	0.0006 (0.0005)	-1.82	-3.18
<i>Paroaria</i>	Chaco/Pantanal	13	3	25	9	0.92 (0.06)	0.0066 (0.0038)	-0.86	-1.02
<i>coronata</i>	Atlantic Forest	3	1	2	3	1.00 (0.27)	0.0014 (0.0014)	0.00	-1.22
	Total	59	12	30	14	0.60 (0.07)	0.0022 (0.0014)	-2.17	-4.76
	Pampa	45	9	26	14	0.86 (0.04)	0.0058 (0.0032)	-0.69	-1.24
<i>Saltator</i>	Chaco/Pantanal	3	2	2	2	0.67 (0.31)	0.0017 (0.0017)	0.00	1.06
<i>similis</i>	Atlantic Forest	27	7	26	18	0.93 (0.04)	0.0042 (0.0025)	-1.79	-11.37
	Total	75	18	43	27	0.90 (0.03)	0.0055 (0.0031)	-1.59	-10.65

323 N Loc: N umber of localities; N Haplot: Number of haplotypes; ¹Values in parenthesis are the standard deviation; ²Values in bold have a P-value

324 <0.05 for Tajima's D and <0.02 for Fu's Fs.

325

326 **Fig. 5.** Median-joining network for the haplotypes of *P. coronata* (A) and *S. similis* (B).
327 The circle's size is proportional to the number of individuals in the sample with that
328 haplotype. The transversal lines in the connectors represent the mutational steps.

329

330 The haplotype network for both species (Fig. 5) also suggests a relatively low
331 degree of genetic structure among regions for both species. Indeed, when all regions are
332 considered, 26.18% of the total genetic variance ($\Phi_{CT}=0.2618$; $P=0.040$) occurs among
333 regions for *P. coronata*, while for *S. similis*, this value is only 12.94% ($\Phi_{CT}=0.1294$;
334 $P<0.001$). On the other hand, most of the genetic variation occurs within populations:
335 65.17% ($\Phi_{ST}=0.3483$; $P<0.001$) and 82.72% ($\Phi_{ST}=0.1728$; $P=0.011$) for *P. coronata* and
336 *S. similis*, respectively. The genetic structure among populations within regions
337 corresponds to the remaining portion of genetic variance, but this value was not
338 significant for both species ($\Phi_{SC}=0.1171$; $P=0.069$ and $\Phi_{SC}=0.0499$; $P=0.090$ for *P.*
339 *coronata* and *S. similis*, respectively). When only the Pampa region is considered, there
340 is no significant genetic structure for either species ($\Phi_{ST}=-0.0510$; $P=0.923$ and
341 $\Phi_{ST}=0.0711$; $P=0.107$ for *P. coronata* and *S. similis*, respectively), corroborating the high
342 general genetic homogeneity within regions shown in the previous analysis.

343 We found evidence for population growth in both species judging by both Tajima's
344 D and Fu's FS statistics (Table 2). However, while in *P. coronata* this signal is only found
345 in the Pampa region, in *S. similis*, it is exclusive from the Atlantic Forest region. The
346 Bayesian Skyline analyses are congruent with these signals (Fig. 6). Population expansion
347 in *P. coronata* seems to be more recent (~50,000 years ago) and less dramatic than in *S.*
348 *similis*, whose expansion seems to have occurred ~350,000 years ago.

349

350 **Fig. 6.** Bayesian Skylines plots for *P. coronata* (A) and *S. similis* (B). The time scale is
351 in thousands of years. The effective population size is multiplied by the generation time
352 in the y-axis ($N_e \times G$). The colors indicate the region of the population considered: Total
353 population—grey; Pampa—purple; Chaco/Pantanal—orange; Atlantic Forest—green.

354

355 **Discussion**

356 Our planet faces many biodiversity conservation challenges to fulfill the needs of
357 an ever-growing human overpopulation [3,4]. Considering the limitation of financial
358 resources for conservation, it is understandable that non-threatened species do not retain
359 high conservation priority [3,5,6]. Indeed, neither the IUCN [5,6] nor the Brazilian
360 Ornithological Society [7] recommend translocation as a valuable conservation tool for
361 non-threatened confiscated wild birds, as the cost of returning birds to the wild in a
362 responsible manner can be prohibitive [6]. However, options other than humane killing
363 of several thousand wild passerines deserve consistent investigation, since euthanizing a
364 high numbers of wild birds annually [45,46] involves ethical issues and may be
365 incongruent with species conservation, especially in the long-term [1,3]. In fact, there has
366 been some evidence that common species also can be susceptible to population declines
367 [47]. The IUCN recognizes that returning confiscated animals of low conservation value
368 to the wild may be valid, given that there are available resources and that translocation is
369 undertaken in accordance with standard conservation guidelines, which should be based
370 on genetic and sanitary information in addition to a number of other studies [5,6]. *S.*
371 *similis* and *P. coronata* are among the most trafficked passerine species in Brazil
372 [45,46,48,49], which configures a legal and conservation challenge even if these species
373 are considered as of least concern on the IUCN red list [18]. Thus, genetic [50,51] and

374 health [52] data from these species may help to substantiate decisions on
375 rehabilitation/release programs.

376

377 **Capturing and sampling wild birds**

378 In this study, after consecutive unsuccessful attempts at capturing live birds, we
379 used live decoys in association with cages mounted with net-trapdoors, Indochinese
380 loops, and mist-nets. Live conspecific decoys have long been among the poachers'
381 practices for supplying illegal trade with wild birds [17]. This method simulates a
382 territorial intrusion, which provides adequate capture opportunities, especially during the
383 breeding season, since birds are prone to fight for defending their territories and are
384 willing to mate [53]. Previously, baiting an area for a couple weeks to entice birds could
385 facilitate the captures [12]; however, most of the capturing areas were visited only at the
386 capture time.

387 Of the 223 catches, 83 were trapped in the net-trapdoors, of which almost all were
388 *S. similis*, which readily engage in combat. On their turn, *P. coronata* are shyer,
389 evaluating their opponents for long periods, hardly perching on unknown devices, and
390 avoiding shaded areas that are suitable for mist netting. In this context, the use of loops
391 camouflaged under a perch allowed the capture of this elusive species, even with the
392 highest mean waiting-time recorded in the study. Loops were used to capture *S. similis*
393 only in specific occasions, such as when it systematically perched in the same place and
394 did not attack the decoy's cage. Mist nets provided comparable catches in both species
395 under study (Fig. 3, Fig. 4).

396 The average target-catches per day was ~ 2.5, much lower than the 10.2 reported
397 recently [14]. Unlike the aforementioned study, we spent considerable time on the road.
398 Furthermore, the increase in the number of daily captures over time (S1 Table) suggests

399 the importance of team learning by practice [54,55]. For example, mist-netting masked
400 by medium height vegetation or wood board fencing, under cloudy or shaded conditions,
401 created additional opportunities for capturing open area birds such as *P. coronata*.
402 Although with a greater effort for installing and linked to most of the non-target captures,
403 the mist net was the fastest (33 minutes) capturing method for both species (Fig. 3).

404 During the three-year period, only four out of 24 birds that were selected as live
405 decoys showed persistently high dueling disposition. The head down and retracted tuft of
406 mute Cardinals and the ruffled feathers, low chirping, and open fallen wings of Saltators
407 were submission behaviors observed under the free-ranging rival challenge. It has also
408 been shown that in some bird species, males utilize vocal performance to evaluate
409 competitors [53,56]. In our study, one specific *S. similis* decoy that whistled an obvious
410 longer and louder terminal trill was associated with many catches (n=58). This individual
411 likely elicited a wider territorial response to the song performance of this “intruder”,
412 similarly to what has been reported for *Zonotrichia leucophrys* males [56]. Thus, the
413 efficiency of the decoy and, consequently, of the capture was associated with the level of
414 the decoy’s song performance or underlying health and vigor [53]. In most occasions in
415 which decoys became mute and submissive, the free-living conspecific bird gave up the
416 fight and left. However, as long as we had a replacement decoy, methods were effective
417 and the target species were not captured, mostly when we couldn’t find them.

418 Data on mortality rates in surveys involving bird capture are scarce and usually rely
419 only on mist-netting [57,58]. While these studies report much lower mortality rates, they
420 do not involve sampling of biological tissues. It is likely that most of the birds that died
421 during collection, suffered from hemorrhages secondary to blood sampling. As an attempt
422 to minimize such losses (S1 Table), blood collection was limited to 0.1 ml per bird,
423 instead of 0.2 ml as we did at the beginning of the fieldwork.

424

425 **Sanitary and health issues**

426 While there is a long list of diseases affecting wild birds [59], we decided to
427 investigate only diseases addressed on the PNSA (i.e., Salmonellosis, mycoplasmosis,
428 and Newcastle disease) due to its associated economical relevance [60] and because
429 investigations about PNSA-associated pathogenic agents in wild passerines are rare.

430 All samples tested for the presence of anti-NDV antibodies were negative in this
431 study. Comparable results have been divulgated in NDV serological surveys in seized
432 Passeriformes, even though positive results have been observed in other avian orders [61].
433 Monitoring free-ranging wild birds have shown the same tendency [62].

434 *Salmonella Enteritidis* and *Salmonella Typhimurium* are, respectively, the
435 serotypes most prevalent and most commonly associated with disease in wild birds [63].
436 However, prevalence of *Salmonella* in wild birds seems to be low and expositions have
437 been attributed to man/domestic animals-contaminated environments [64]. Indeed,
438 previous studies estimated the prevalence of *Salmonella* in samples of wild birds,
439 including in passerines confiscated from illegal traffic, at between 1–7% [59,65–67]. In
440 our sample, only one fecal sample (0.7%) was positive for *Salmonella* cultivation and
441 isolation, which is consistent with a previous study on *Paroaria* sp. in which *Escherichia*
442 *coli* and *Klebsiella* spp., among others, were detected in much higher numbers than
443 *Salmonella* [67]. The positive sample was *Salmonella enterica* serovar Cerro (Group K).
444 This serovar has been linked to poultry feedstock and rations [68] and commercial laying
445 farms [69] and has high prevalence among cattle herds [70], which was the case here,
446 even though the meaning of this finding for the health of this free-ranging Red-crested
447 Cardinal population remains to be investigated.

448 We found higher prevalence for MG using the RPA test compared to the qPCR
449 assay for both free-living and seized birds, which was expected due to the limited
450 accuracy of RPA [29]. MG-prevalence in poultry and wild birds has usually been higher
451 under RPA than in other tests [71–73]. The qPCR-based estimate of 13% in MG
452 prevalence in the seized group is comparable to what has been found in other avian hosts
453 [74–76]. Similarly, the higher MG prevalence in seized birds probably reflects the poor
454 hygiene and stressful conditions to which these birds are subjected [59]. Even though MG
455 exposure has been reported for wild birds [71,74,77], the lack of qPCR-based positive
456 samples for MG is in agreement with another Brazilian study that sampled wild birds in
457 poultry farms surrounded by forest remnants [78]. Even though none of the positive
458 individuals in our sample showed any clinical evidence of infection, differential MG-
459 susceptibility across bird species has been observed after experimental infections that
460 resulted in both clinical and subclinical symptoms [79], which also occurred in wild birds
461 [74,77,80]. Disease outbreaks in free-ranging and captive wild bird have been recorded,
462 suggesting a potential reservoir for the pathogen in nature [81–83], reinforcing the need
463 of a reliable MG assay for elucidating the role of MG in populations of wild birds.

464

465 **Genetic diversity, structure, and implications of outbreeding depression**

466 The two species revealed different levels of genetic diversity and different
467 evolutionary demographic histories. *S. similis* had overall higher genetic diversity, a more
468 ancient population expansion restricted to the populations in the Atlantic Forest (while
469 the Pampa population remained constant), and similar levels of diversity between the
470 Pampa and the Atlantic Forest, its major regions of occurrence. On its turn, *P. coronata*
471 had lower diversity, a weaker and more recent population restricted to the populations in

472 the Pampa (while the Chaco/Pantanal population remained constant), and lower diversity
473 in the Pampa compared to the Chaco/Pantanal region (Table 2, Fig. 5, Fig. 6).

474 When compared to other passerines, mainly from open areas and/or dry forests in
475 Southern South America [84,85], *P. coronata* from the Chaco/Pantanal had a high
476 diversity at both the haplotype and nucleotide level. Even though some population of
477 *Gubernatrix cristata*, a highly endangered Thraupidae, had higher nucleotide diversity
478 values [84], this study used the mtDNA control region (mtDNA-CR) as a genetic marker,
479 which evolves faster than ND2 [86]. On the other hand, *P. coronata* from the Pampa had
480 a low diversity at both indices. This may indicate that the population inhabiting the Pampa
481 biome is a recent offshoot from the Chaco/Pantanal region, which may have acted as a
482 large source population for this species. This is compatible with the exclusive signal of
483 population expansion in the Pampa around 50,000 years ago (Fig. 6), as population
484 growth is usually expected in refuge areas [87].

485 Compared to other passerines more associated with the Brazilian Atlantic Forest
486 and other associated humid forests, *S. similis* revealed a high diversity both in the Pampa
487 as well as in the Atlantic Forest regions. Considering studies that used mtDNA coding
488 genes only [88–95], *S. similis* from the Atlantic Forest had the highest value for a forest-
489 associated species (only lower than *P. coronata* from the Chaco/Pantanal), while the
490 Pampa population had the fifth highest value. Haplotype diversity values for both
491 populations are also among the highest for passerines associated with the Atlantic Forest
492 [88–95]. The exclusive signal of an ancient population expansion for *S. similis* in the
493 Atlantic Forest region but not in the Pampa is curious. First, most studies that detected a
494 population expansion in specific populations or phylogroups suggest much more recent
495 timescales (similar to *P. coronata*) [90,92,94,96]. The two exceptions that report older
496 expansions are *Schiffornis virescens* [91] and *Basileuterus leucoblepharus* [92], around

497 150,000–300,000 years ago. The high genetic diversity and lack of a signal of population
498 expansion in the Pampa is intriguing. It is unlikely that the Pampa behaved as a refuge
499 area for this species, which is strongly associated with humid forests northern to the
500 Pampa region. Besides, the Atlantic Forest has higher genetic diversity than the Pampa,
501 which is not compatible with a scenario in which the population in a refuge – the Pampa
502 – occupies the Atlantic Forest and expands thereafter. Taken together, the lack of strong
503 genetic structure between these populations may indicate that the Pampa was occupied in
504 consequence of the population expansion in the Atlantic Forest but subsequently lost
505 genetic diversity, which eroded the signal of an ancient expansion.

506 We also found a very weak genetic structure for both *P. coronata* and *S. similis*
507 (Fig. 6). In the Pampa and associated dry forests, both *Gubernatrix cristata* [84] and
508 *Phytotoma rutila* [85] showed stronger structure among habitats ($\Phi_{ST} \sim 0.45$). For forest
509 species, most studies reported the presence of strongly differentiated phylogroups either
510 between different habitats or across the Atlantic Forest [88–90,93–99]. Again, the
511 exceptions are *Schiffornis virescens* [91] and *Basileuterus leucoblepharus* [92], which
512 showed no phylogeographic breaks across its distribution in the Atlantic Forest. The lack
513 of genetic structure in *S. similis* is corroborated by a study that, based on nine
514 microsatellite (SSR) loci, reported that only 0.1% of the total genetic variation occurred
515 among populations distributed in different Brazilian biomes (Atlantic Forest, Cerrado,
516 Caatinga and ecotones between these) [50]. Similarly, even though no study has
517 characterized *P. coronata* populations for SSR variation, the closely related species *P.*
518 *dominicana*, which occurs in the Brazilian northeast, showed that only 3% of the total
519 genetic variation occurred among populations. [50]. Thus, the high genetic connectivity
520 among populations does not seem to be an artifact from using only mtDNA.

521 The shallow population differentiation found in this study may have important
522 consequences for conservation policies. Even though genetic distance is not a good
523 predictor of outbreeding depression [100], outbreeding depression requires different
524 genetic adaptations to local environments that will be broken by crossing individuals from
525 exogenous populations [9,10,101]. The high historic genetic connectivity across
526 populations of both species suggests that the overall risk of outbreeding depression is low
527 [101].

528

529 **Concluding remarks**

530 The management of birds seized from illegal traffic is a complex and difficult
531 question; Even though the recommendation for euthanizing individuals belonging to
532 species of low conservation value may be justified [6,7], could new information for focal
533 species reduce the perceived risk of translocation to the point that this could become a
534 standard practice? In this study, we showed that it may be feasible to screen seized birds
535 for several pathogens and that while the overall prevalence rates are low, the risk of
536 disease transmission during bird management or after release in natural environments
537 justifies a systematic sanitary screening. In addition, applying an adequate quarantine
538 period for all seized birds seems to be both feasible and indispensable for maintaining
539 proper sanitary conditions of the wild bird populations in translocation/reintroduction
540 initiatives. Furthermore, systematic pathological investigations on confiscated birds
541 dying during the quarantine/rehabilitation processes may point to additional relevant
542 conditions to be included on the health screening.

543 The shallow genetic structure found for both species suggest that the overall risk
544 of outbreeding depression following translocation is low. Yet, additional criteria could be
545 used to choose an area for translocating individuals. For example, using different regional

546 vocal “dialects” as a proxy for the original parental population of an individual [102].
547 Vocal plasticity exhibited by several species may indicate that, for the undetermined
548 cases, reintroduced birds will be able to adapt to their release sites [102]. Another eventual
549 strategy could be releasing the birds close to the apprehension site. Conservationist
550 policymakers should consider when translocating seized individuals into wild populations
551 could represent a safe and useful strategy for maintaining large and healthy wild
552 populations for species highly targeted for illegal trade, irrespective of their conservation
553 status.

554

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885

886 **Supporting information captions**

887 **S1 Table** – Captures.

888 **S2 Table** – Genetic samples.

889 **S3 Table** – Live decoys.

890 **S4 Table** – Health tests.

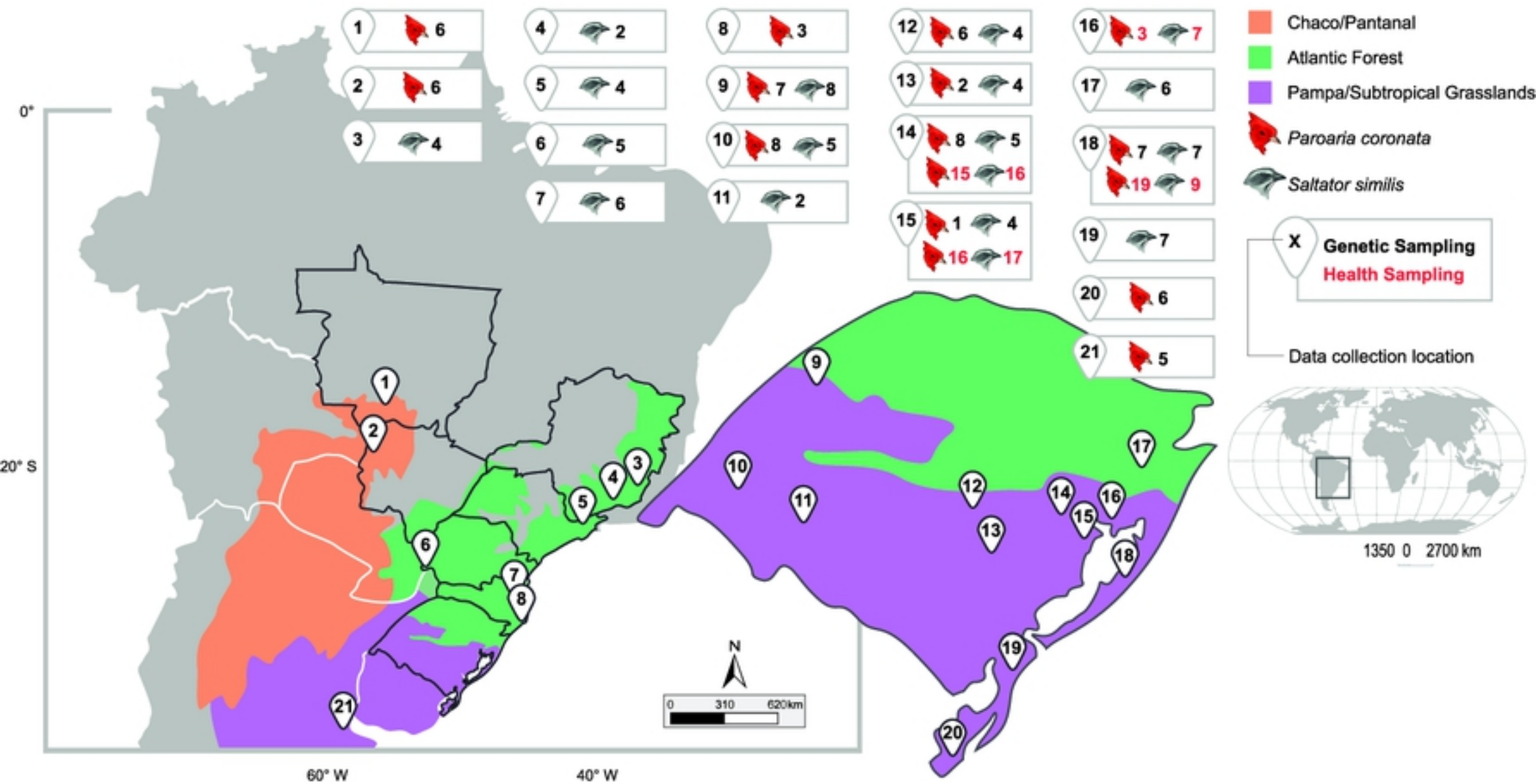
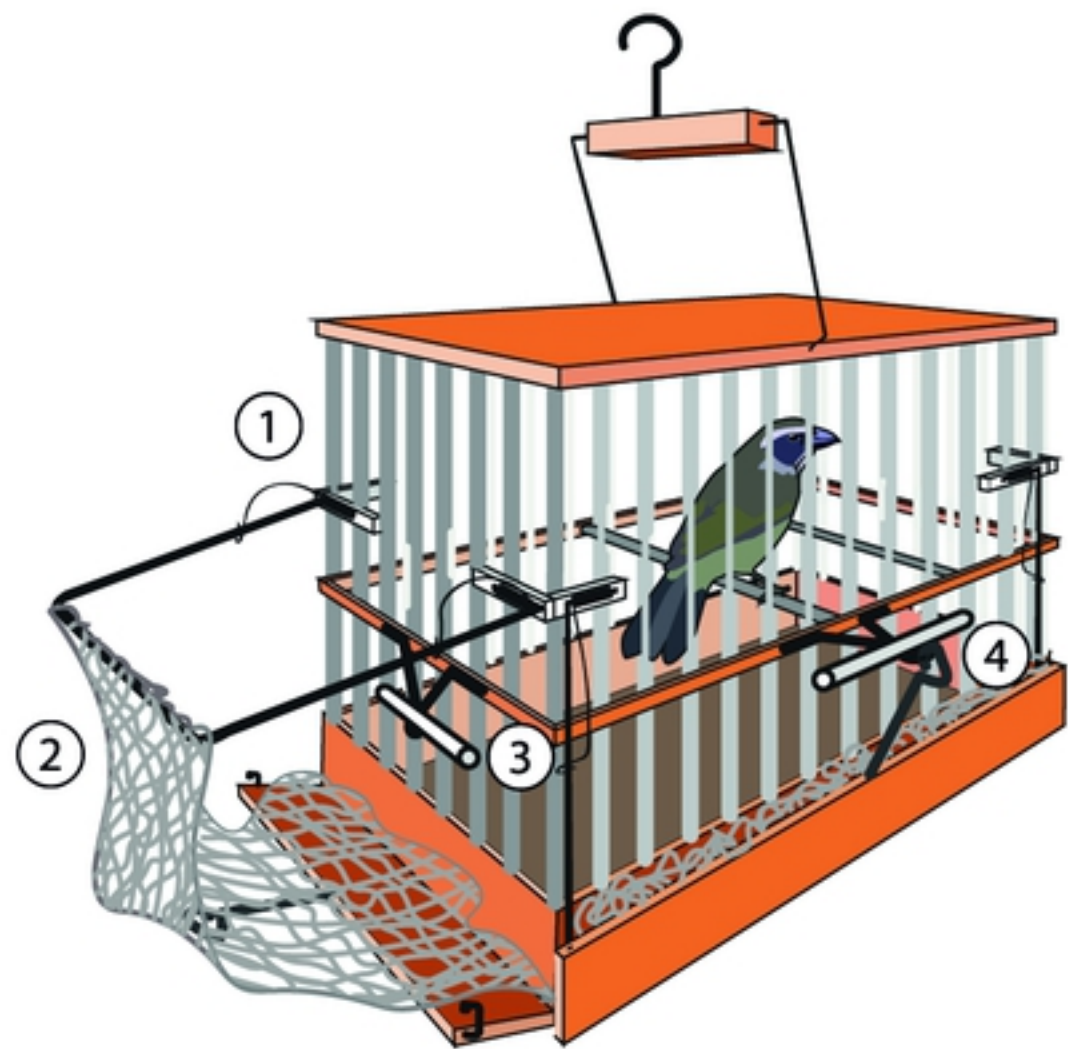
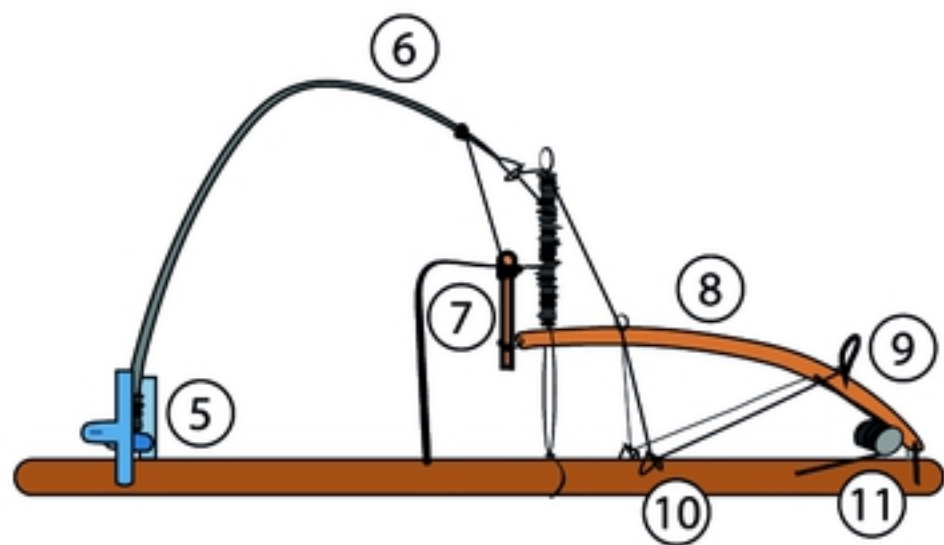


Figure 1

A.



B.



C.

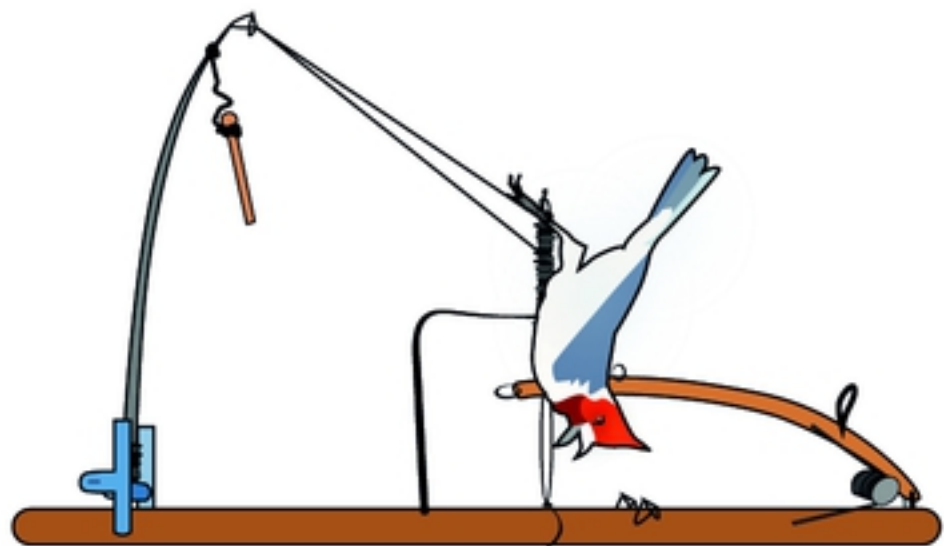


Figure 2

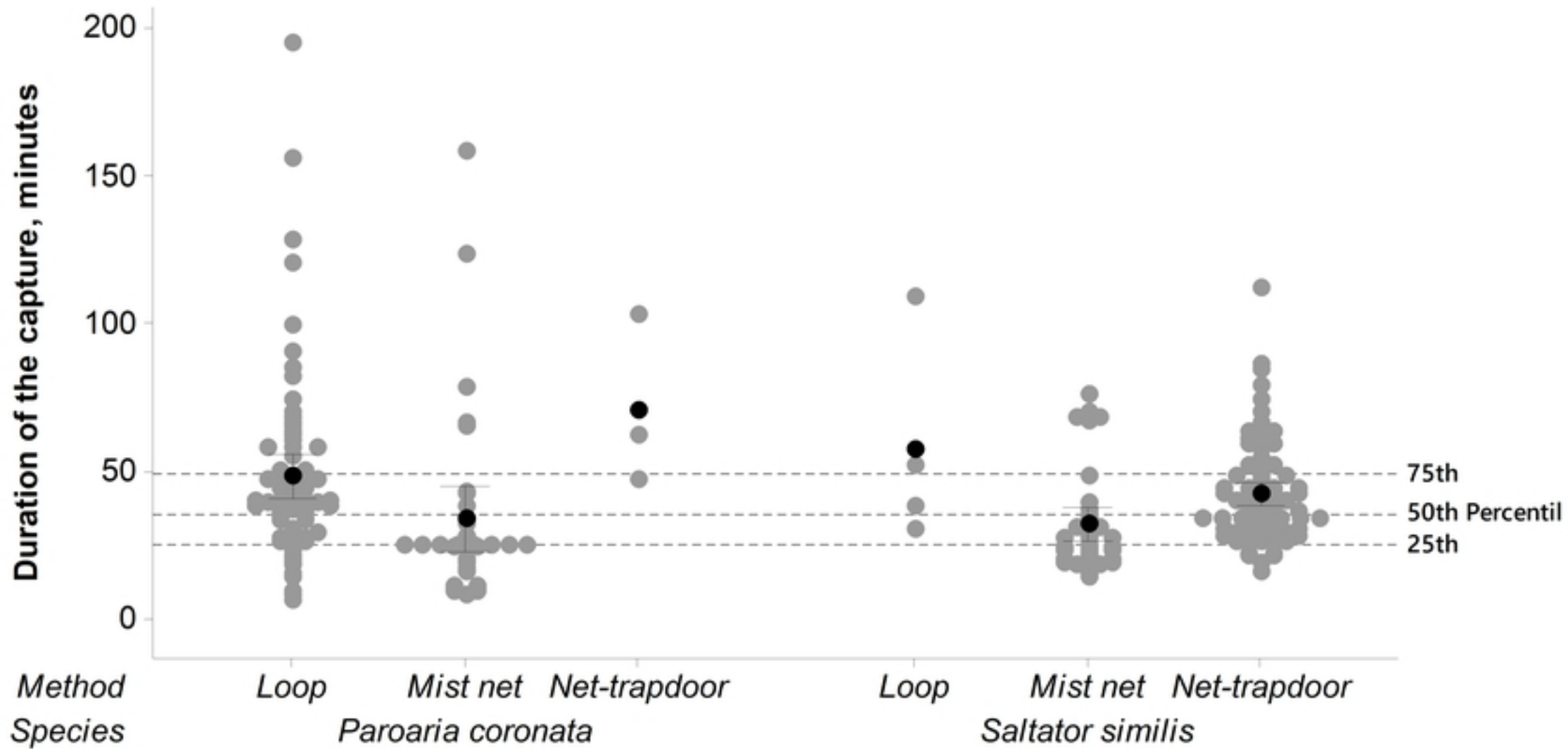


Figure 3

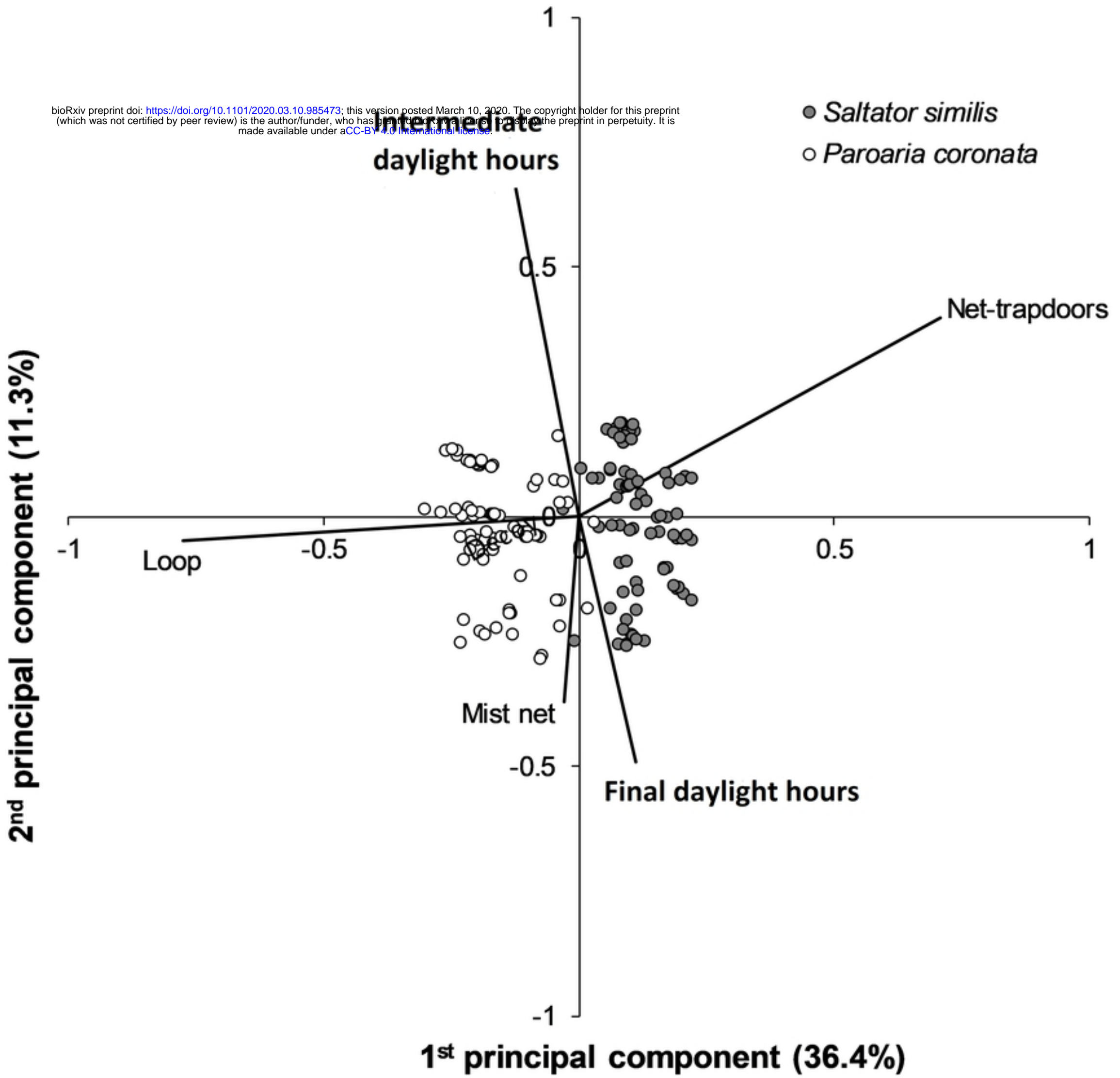


Figure 4

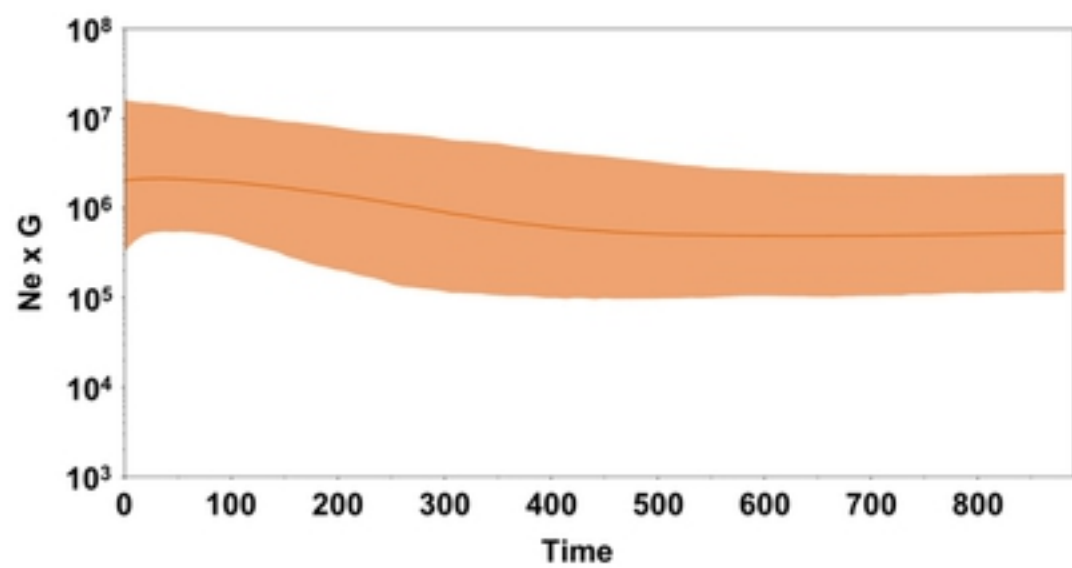
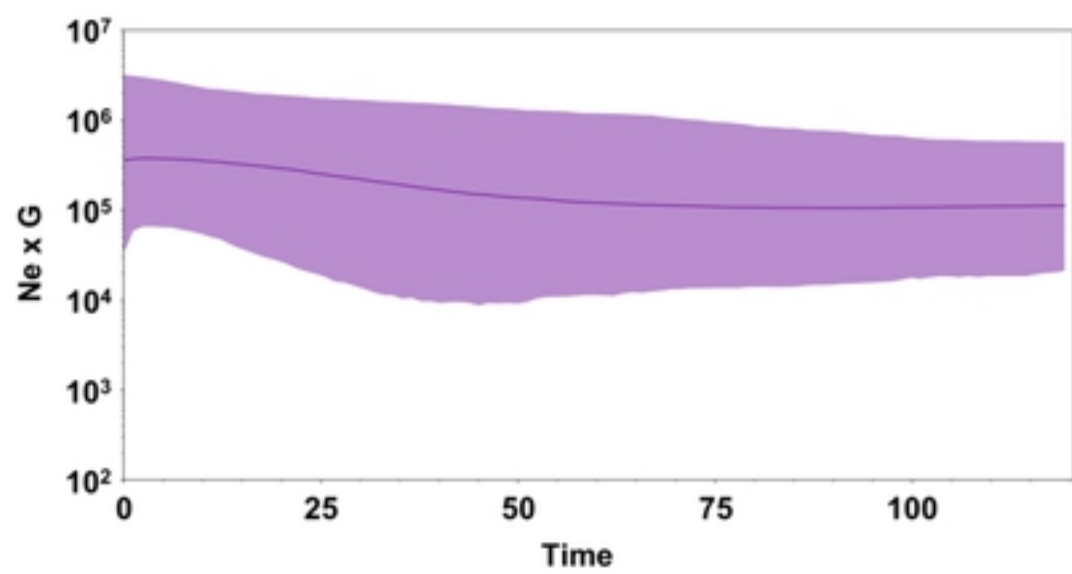
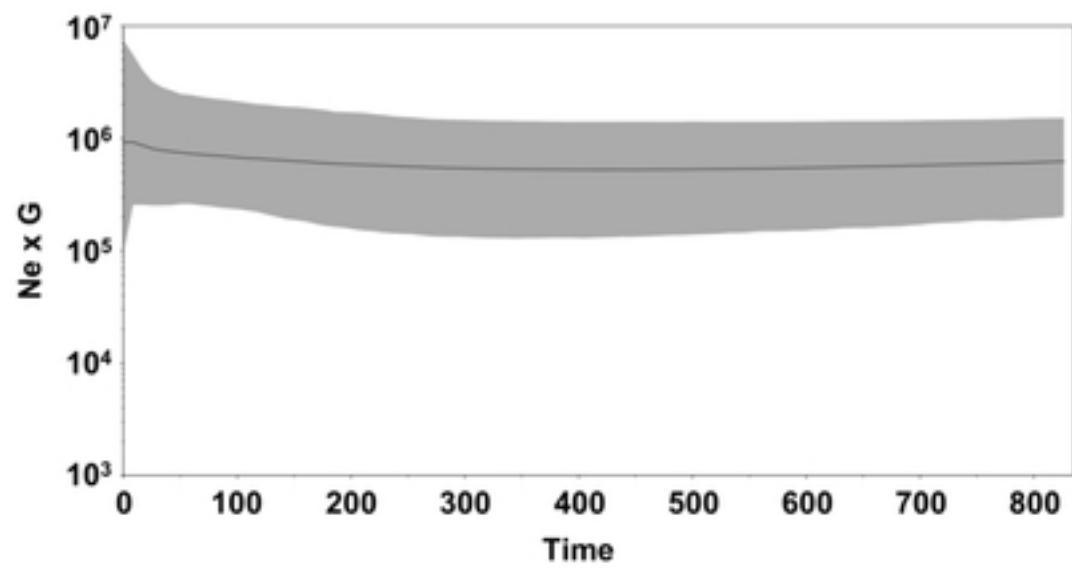
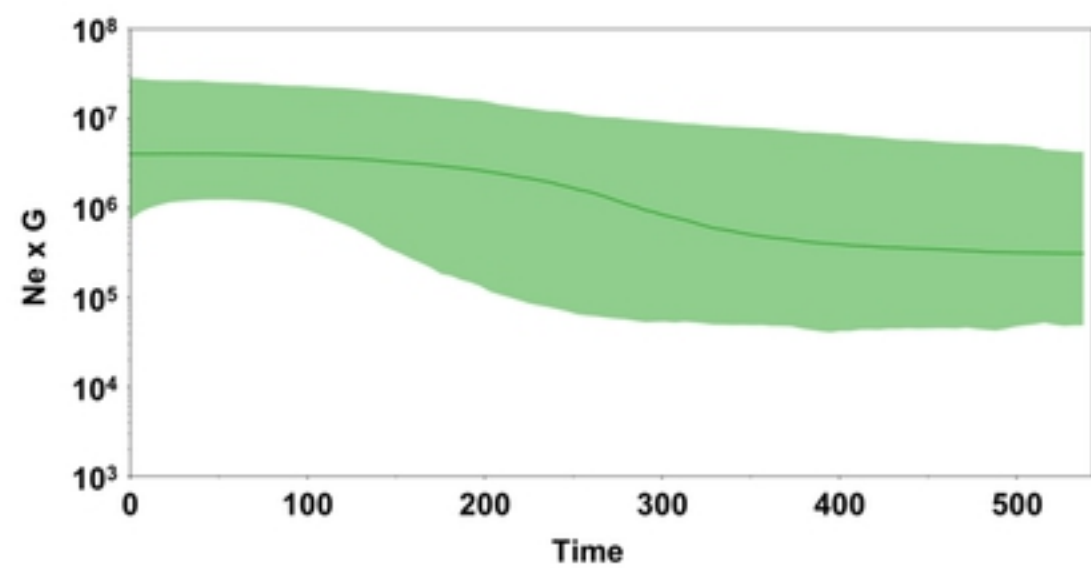
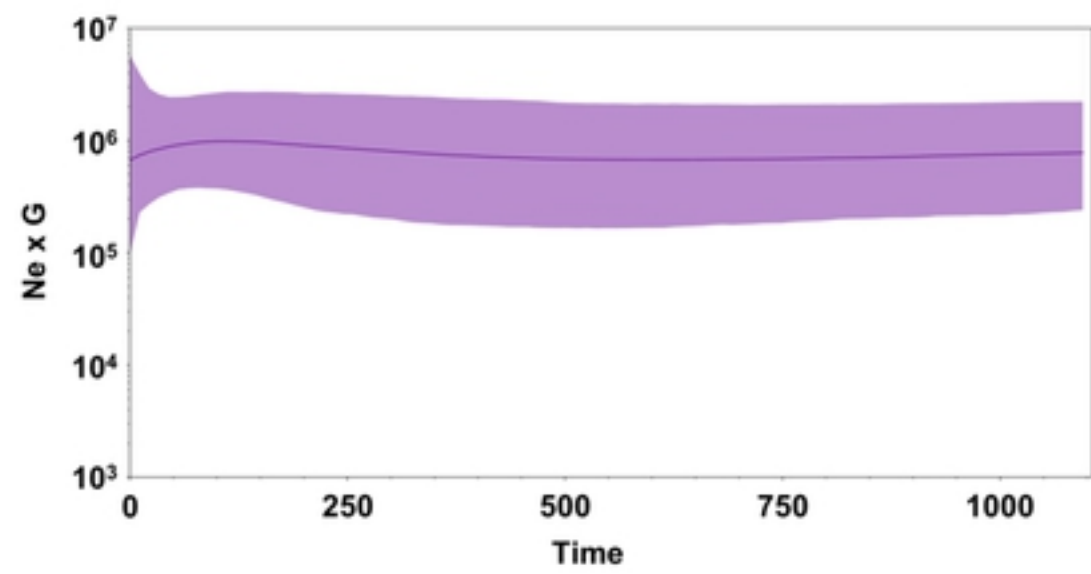
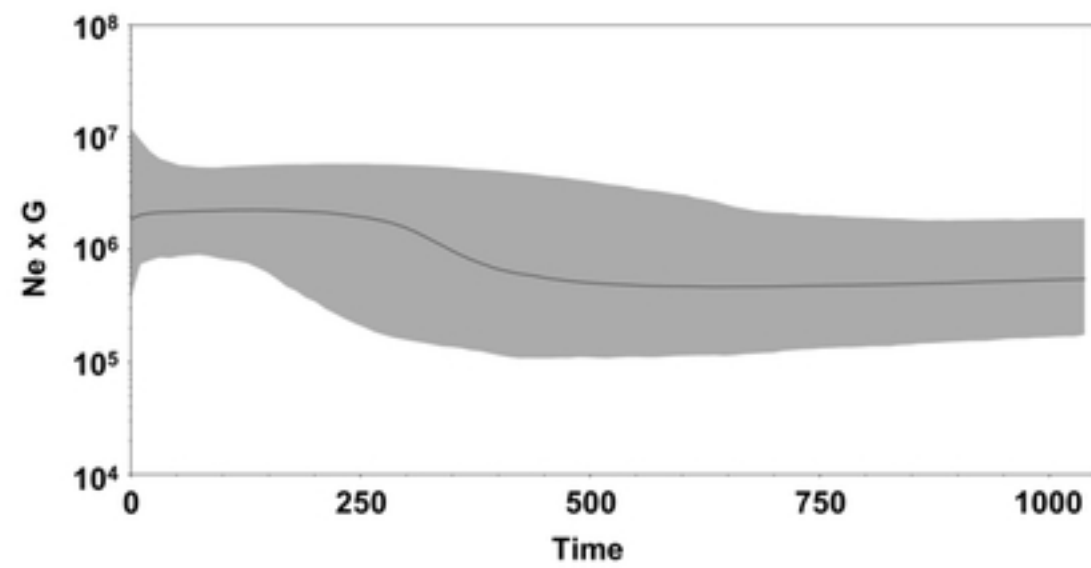
A**B**

Figure 6