ARTICLE

Natural selection and parallel clinal and seasonal changes in

Drosophila melanogaster

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Abstract

Spatial and seasonal variation in the environment are ubiquitous. Covariation between a trait

and space is often interpreted as the result of selection, but demography can complicate

inference. Finding seasonal changes driven by selection is challenging, given effect sizes are

small and the environment is subject to stochastic changes within seasons. *Drosophila*

melanogaster is known to harbor polymorphisms that change with latitude and seasons.

Identifying the role of selection in driving latitudinal and seasonal change is not trivial, but

one promising approach is to look for parallel clinal and seasonal variation. Here, we test

whether there is a genome-wide relationship between clinal and seasonal variation, and

whether the pattern is consistent with selection. We investigate the role of linked selection in

driving these patterns, and conservatively estimate the proportion of the variants that should

be under selection to explain our observations. Allele frequency estimates were obtained

from pooled samples from seven different locations along the east coast of the US, and 13

samples collected in the spring and fall in Pennsylvania. We show that there is a genome-

wide pattern of clinal variation mirroring seasonal variation, which cannot be explained by

linked selection alone. This pattern is stronger for coding than intergenic regions, consistent

with natural selection. We find that the genome-wide relationship between clinal and

seasonal variation could be explained by about 2.8% of the common variants being under

selection. Our results highlight the contribution of natural selection in driving allele

frequency changes with latitude and seasons in $D.\,melanogaster$.

Introduction

Many species can be found along environments that gradually change in time and

space (Ewing 1979; Cardini et al. 2007; Dionne et al. 2007; Hancock et al. 2008; Zuther et al.

2012; Campitelli and Stinchcombe 2013; Kooyers et al. 2015). Traits that covary with the

environment in space are called clinal. These clinal traits are usually thought to be a result of

spatially varying selection (Endler 1977), but non-adaptive processes such as isolation by

distance, range expansion and admixture can produce similar patterns of spatial change

(Wright 1943; Vasemägi 2006; Excoffier et al. 2009; Duchen et al. 2013; Bergland et al.

2016). Therefore, disentangling the relative contributions of natural selection and

demographic processes in producing clinal variation is challenging.

Drosophila melanogaster has been used as a model to study clinal variation because,

in addition to being a genetic model, it is a sub-Saharan fly species that has recently invaded

most of the world (David and Capy 1988). These flies migrated into Eurasia approximately

15,000 years ago (Li and Stephan 2006), but the colonization of the Americas and Australia

likely happened in a single event within the last several hundred years (Bock and Parsons

1981; Keller 2007). The establishment of populations in dramatically different environments

(*e.g.*, temperate regions) was likely followed by the emergence of adaptations to new

climates (David and Capy 1988; Yukilevich et al. 2010).

Clinal variation has been documented for *D. melanogaster* in many characters both at

the phenotypic and genetic level. For instance, at the phenotypic level, flies from higher

latitudes are darker (David et al. 1985), bigger (Arthur et al. 2008) and show higher incidence

of reproductive diapause than flies from lower latitudes (Schmidt et al. 2005). At the

genotypic level, latitudinal clines have been identified for allozymes, chromosome

inversions, copy-number variation and single nucleotide polymorphisms (Mettler et al. 1977;

Knibb 1982; Oakeshott et al. 1982; Schmidt et al. 2000; de Jong and Bochdanovits 2003;

Sezgin et al. 2004; Fabian et al. 2012; Kapun et al. 2016; Schrider et al. 2016). Surprisingly,

not much is known about the actual selective pressures in natural fly populations, but

temperature is directly or indirectly underlying much of clinal adaptation (Knibb 1982;

Krimbas and Powell 1992; Božičević et al. 2016).

A classical approach to discern between adaptive and neutral differentiation has been

to look at parallel latitudinal variation between continents (Hoffmann et al. 2002; Hoffmann

and Weeks 2007; Turner et al. 2008; Paaby et al. 2010; Reinhardt et al. 2014; Schrider et al.

2016). If a trait varies clinally in two or more continents, a simple, plausible scenario is

latitudinally varying selection acting on ancestral variation (Endler 1977; Barton 1983;

Barton 1999). However, some of the clinal patterns observed in North America may be

attributed to migration of African flies to low latitude regions and European flies to high

latitude regions (Caracristi and Schlotterer 2003; Yukilevich and True 2008; Duchen et al.

2013; Kao et al. 2015; Bergland et al. 2016). The main evidence in favor of the secondary

contact hypothesis is that there is a cline in the proportion of African ancestry in North

America (Kao et al.2015; Bergland et al. 2016), and a weak cline in African ancestry in

Australia (only marginally significant; see Bergland et al. 2016).

Separating the role of selection and demography in maintaining clinal polymorphisms

is not trivial, given that the axis of demography and environmental change are confounded in

this system (i.e., migration and the environment are structured along the south-north axis). A

simpler, more plausible model would be that admixture introduced "pre-adapted" alleles, that

then contributed to clinal adaptation (Flatt 2016). For instance, for proper modelling of

selection and demography in this system, one would need more information about rates of

migration, population sizes, changes in population size, historical founding of the

populations, history of admixture between populations, and sources and nature of selective

pressures that vary with latitude. Alternatively, one simple approach that can help disentangle

the role of adaptation is to explore signatures of parallel change with latitude and seasons

(Cogni et al. 2014; Cogni et al. 2015).

Drosophila has also been widely used in the study of seasonal adaptation. One of the

earliest examples of seasonal genetic variation was observed in chromosomal inversions in *D*.

pseudoobscura (Dobzhansky 1943). In *D. melanogaster*, it has been shown that flies

collected in the spring are more tolerant to stress (Behrman et al. 2015), show higher

diapause inducibility (Schmidt and Conde 2006), have increased immune function (Behrman

et al. 2018) and have different cuticular hydrocarbon profiles than those collected in the fall

(Rajpurohit et al. 2017). Furthermore, genome-wide analyses have identified polymorphisms

and inversions that oscillate in seasonal timescales in several localities in the United States

and Europe, but the mechanism behind it has not been fully resolved (Bergland et al. 2014;

Kapun et al. 2016; Machado et al. 2018; Wittman et al. 2017). Identifying polymorphisms

that fluctuate seasonally is challenging, because effect sizes are small and populations are

subject to stochastic environmental events that are not tied to seasonality (Machado et al.

2018). Indeed, a recent study put in question the interpretation that a set of previously

identified seasonal SNPs in *D. melanogaster* is due to seasonally varying selection (Buffalo

and Coop 2019).

One way to increase power would be to study the relationship between seasonal and

latitudinal change. Seasonal and latitudinal variation could be coupled because of two

different adaptive mechanisms. First, the environment changes similarly with latitude and

through seasons (at least with respect to temperature). Second, the onset of the spring

changes with latitude and so seasonal changes in allele frequencies alone could produce clinal

variation, a mechanism termed seasonal phase clines (Roff 1980; Rhomberg and Singh

1986). Demographic processes that can generate clinal patterns like isolation by distance and

secondary contact from diverged populations are independent to variation across seasons,

given the short time scale at which the seasonal processes happen (Bergland et al. 2014;

Cogni et al. 2015). Therefore, parallel latitudinal and seasonal variation in a trait is strong

evidence in favor of natural selection. For instance, it has been observed in *D. melanogaster*

that both the prevalence of reproductive diapause and the frequency of a variant in the couch

potato gene associated with diapause inducibility vary latitudinally and seasonally (Schmidt

et al. 2005; Cogni et al. 2014). The frequency of this diapause-inducing variant drops in the

summer and is positively correlated with latitude, as we would expect if it were being driven

by natural selection.

Besides this classic example, a few studies have looked for signatures of parallel

clinal and seasonal variation in *D. melanogaster* (Bergland et al. 2014; Cogni et al. 2015;

Kapun et al. 2016; Behrman et al. 2018; Machado et al. 2018). Using a candidate gene

approach, Cogni et al. (2015) found an association between clinal and seasonal change in

central metabolic genes, that are likely important drivers of climatic adaptation. Kapun et al.

(2016) dissected the role of selection in shaping clinal variation in *D. melanogaster*

cosmopolitan inversions. By integrating clinal and seasonal variation, information on the

stability of inversion clines, and analysis of the effect of climatic variables on inversion

frequencies, they concluded that several inversion clines are consistent with natural selection.

A recent study also investigated the association between clinal and seasonal variation, and

showed the direction of allele frequency change with latitude is concordant with the change

with seasons in many polymorphisms across the *D. melanogaster* genome (Machado et al.

2018). However, the informative value of parallel clinal and seasonal variation in

determining the role of selection was not fully explored.

Here, we aim to answer whether the clinal and seasonal patterns observed in *D.*

melanogaster are consistent with natural selection. If natural selection is a key process

underlying clinal and seasonal change, flies collected in the north should be more similar to

flies collected in early spring, whereas southern flies should be more like flies collected in the

fall (Cogni et al. 2014; Bergland et al. 2014; Cogni et al. 2015; Kapun et al. 2016; Behrman

et al. 2018; Machado et al. 2018). Thus, we quantified the relationship between clinal and

seasonal change along the *D. melanogaster* genome. Given the localized effects of selection,

and the expectation that constraint depends to some degree on genic annotations, we also

expect the parallelism to vary across genomic annotations. Finally, we estimated how much

of the common variants should be under selection to explain the degree of coupling between

clinal and seasonal change. By integrating these two independent sources of evidence, we

further disentangle the contributions of selection in maintaining the pervasive and long-

standing patterns of clinal variation in *D. melanogaster*.

Material and Methods

Population samples

We analyzed 20 samples from seven locations along the United States east coast, collected by

(Bergland et al. 2014) (10 samples), and (Machado et al. 2018) (10 samples) (see Table S1).

The samples were based on pools of wild-caught individuals. We decided to not include

previously collected samples from Maine because they were collected in the fall, whereas all

of our other samples were collected in the spring, and we also did not include the DGRP

sample from North Carolina, as it is hard to ascertain when they were obtained (Fabian et al.

2012; Mackay et al. 2012; Bergland et al. 2014). The Linvilla (Pennsylvania) population was

sampled extensively from 2009 to 2015 (six spring, seven fall samples), and was therefore

used in our analysis of seasonal variation. We also replicated our results using data from four

Australian samples (Anderson et al. 2005; Kolaczkowski et al. 2011). All the data used in this

project are available on the NCBI Short Read Archive (BioProject accession numbers

PRJNA256231, PRJNA308584 and NCBI Sequence Read Archive SRA012285.16).

Mapping and processing of sequencing data

Raw, paired-end reads were mapped against the FlyBase *D. melanogaster* (r6.15) and *D.*

simulans (r2.02) reference genomes (Gramates et al. 2017) using BBSplit from the BBMap

suite (https://sourceforge.net/projects/bbmap/; version from February 11, 2019). We removed

any reads that preferentially mapped to *D. simulans* to mitigate effects of contamination (the

proportion of reads preferentially mapping to *D. simulans* was minimal, never exceeding

3%). Then, reads were remapped to *D. melanogaster* reference genome using bwa (MEM

algorithm) version 0.7.15 (Li and Durbin 2010). Files were converted from SAM to BAM

format using Picard Tools (http://broadinstitute.github.io/picard). PCR duplicates were

marked and removed using Picard Tools and local realignment around indels was performed

using GATK version 3.7 (McKenna et al. 2010). Single nucleotide polymorphisms (SNPs)

and indels were called using CRISP with default parameters (Bansal et al. 2016). We applied

several filters to ensure that the identified SNPs were not artifacts. SNPs in repetitive regions,

identified using the RepeatMasker library for *D. melanogaster* (obtained from

http://www.repeatmasker.org), and SNPs within 5bp of polymorphic indels were removed

from our analyses, because these are more likely to be sequencing errors. SNPs with average

minor allele frequency in the clinal and seasonal samples less than 10%, with minimum per-

population coverage less than 10x (or 4x for the Australian samples) or maximum per-

population coverage greater than the 99th quantile were excluded from our analyses. Also,

SNPs with multi-allelic states were removed from the analysis. We did not include SNPs in

any of the sex chromosomes. SNPs were annotated with their genic classes using SNPeff

version 4.3o (Cingolani et al. 2012).

Clinal and seasonal changes in allele frequency

Pool-seq data contains an additional component of error due to sampling (Kofler et al. 2011;

Lynch et al. 2014). We decided to model this additional error by incorporating weights

proportional to the effective number of chromosomes (N_E):

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$$N_E = \left(\frac{1}{D} + \frac{1}{N_C}\right)^{-1}$$

where N_C is the number of chromosomes in the pool and D is the read depth at that site

(Kolaczkowski et al. 2011; Feder et al. 2012; Bergland et al. 2014).

To assess latitudinal variation, we used generalized linear models (GLM) to regress

allele frequency at each site against latitude according to the form,

$$y_i = \alpha + \beta^{clinal} Latitude + \epsilon_i$$

Where y_i is the allele frequency in the ith population, α is the intercept, β^{clinal} is the slope,

$Latitude$ is the latitude of the population, and ϵ_i is the binomial error given the N_E at that

SNP. This kind of regression is suitable for the analysis of clinal variation of allele

frequencies estimated from Pool-seq, because it accounts for differences in read depth and

number of chromosomes, and the non-normality of allele frequencies (Bergland et al. 2014;

Machado et al. 2016).

To assess seasonal variation, we followed a similar procedure. We regressed allele

frequency at each site against a binary variable corresponding to spring or fall according to

the form,

$$y_i = \alpha + \beta^{seasonal} Season + \gamma Year + \epsilon_i$$

where y_i is the allele frequency in the ith sample, α is the intercept, $\beta^{seasonal}$ is the slope,

$Season$ is the season in which the sample was obtained (June and July were encoded as

Spring, and September, October and November as Fall), $Year$ is the year, and ϵ_i is the

binomial error given the N_E at that SNP.

We defined clinal and seasonal SNPs using an outlier approach, because we do not

have an adequate genome-wide null distribution to compare our estimates. Thus, we

considered that SNPs were outliers if their regression P-value fell in the bottom 1% of the

distribution. All statistical analyses were performed in R 3.5.0 (R Core Team 2018).

Relationship between clinal and seasonal variation

We examined the relationship between clinal and seasonal variation by regressing the clinal

against seasonal slopes for all SNPs:

$$\beta_i^{clinal} = \alpha + \beta\beta_i^{seasonal} + \epsilon_i$$

where β_i^{clinal} is the estimated clinal slope for the ith SNP, α is the intercept, β is the slope,

$\beta_i^{seasonal}$ is the estimated seasonal slope for the i^{ith} SNP, and ϵ_i is the gaussian error at that

SNP. The clinal and seasonal slopes were z-normalized to facilitate interpretation. To

confirm our results are robust to potential model misspecifications, we implemented a

permutation test in which we rerun the regressions for each SNP using shuffled season and

latitude labels 10,000 times.

To assess differences in the relationship between clinal and seasonal variation across

different genic classes (exon, intron, 5' UTR, 3' UTR, upstream, downstream intergenic,

splice), we added the following interaction term:

$$\beta_i^{clinal} = \alpha + \beta\beta_i^{seasonal} + \gamma_i\beta_i^{seasonal} \times Class_j + \epsilon_i$$

where γ_i is the regression coefficient for the jth genic class, $Class_j$ is the genic class of the

i^{th} SNP. Furthermore, we performed a post-hoc Tukey test to find differences in the

relationship between clinal and seasonal variation among all genic classes. We also

confirmed our results with permuting season and latitude labels to build a null distribution of

the expected regression coefficients of each genic class.

There are some chromosomal inversions segregating in the populations we studied

and they are known to contribute to adaptation (Wright and Dobzhansky 1946; García-

Vázquez and Sánchez-Refusta 1988; Kapun et al. 2014). We tested whether the relationship

between clinal and seasonal variation is stronger inside within inversions. To do so, we

performed a regression similar to our previous one, that takes the form:

$$\beta_i^{clinal} = \alpha + \beta\beta_i^{seasonal} + \gamma_i\beta_i^{seasonal} \times Inversion_j + \epsilon_i$$

where $Inversion_j$ is the inversion status of the SNP, and γ_i is the regression coefficient for

the jth status. All statistical analyses were performed in R 3.5.0 (R Core Team 2018).

Composite statistic for clinal and seasonal variation

Our approach uses the relationship between clinal and seasonal variation in sets of SNPs to

infer selection, and so it is not possible to detect individual SNPs which are under spatial and

seasonal selection. We implemented a composite statistic based on Fisher's method (Fisher

1992) that considers clinal and seasonal regression P-values, and the sign of the clinal and

seasonal slopes:

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$$\chi_i = -2 \times sgn(\beta_i^{clinal} \times \beta_i^{seasonal})(\ln(P_i^{clinal}) + \ln(P_i^{seasonal}))$$

where P_i^{clinal} is the P-value of the clinal regression and $P_i^{seasonal}$ the P-value of the seasonal

regression. When both P-values are small, and the sign of the slopes are opposite, the statistic

χ will be negative in sign and have a large magnitude, suggesting that natural selection is

acting on that polymorphism. We defined as outliers those in the bottom 1% of the χ

distribution.

Enrichment tests

We tested for enrichment of genic classes using our sets of clinal, seasonal, and composite

SNPs. We tested for enrichment using Fisher's exact test for each genic region and statistic.

Because our choice of defining top 1% of outliers is arbitrary, we calculated our P-values

based on 10,000 permutations. In each iteration, we shuffled the season and latitude labels

and reran the generalized regressions. Using the P-values obtained from regressions in which

season and latitude labels were shuffled, we defined for each iteration the top clinal, seasonal

and composite SNPs. Then, we calculated the enrichment of each genic class using Fisher's

exact test for all iterations. To obtain a P-value for an enrichment of a given genic class, we

compared the observed odds ratio in the actual dataset to the distribution of odds ratios

observed for datasets in which season and latitude labels were shuffled.

We also performed Gene Ontology (GO) enrichment analysis to identify the

biological function underlying our sets of outliers (Ashburner et al. 2000). One common issue

with GO analysis is that they are biased, as longer genes are likely to harbor more SNPs. We

thus used Gowinda (Kofler and Schlötterer 2012), an unbiased (*i.e.*, that accounts for gene

length) permutation approach to GO analysis. We obtained annotations from Flybase r6.15

and used the following parameters in Gowinda: 10,000 simulations; a minimum of 1 gene per

GO category, and considered SNPs 1000bp up and downstream of a gene.

Some specific genes (Table 1) have been shown to be strongly involved with clinal or

seasonal variation in *D. melanogaster (*Vigue and Johnson 1973; Oakeshott et al. 1982; Berry

and Kreitman 1993; Schmidt et al. 2000; Verrelli and Eanes 2001; Duvernell et al. 2003;

Sezgin et al. 2004; Schmidt and Paaby 2008; Cogni et al. 2014; Paaby et al. 2014; Behrman

et al. 2018). Therefore, we sought to test whether they were enriched among the outliers of

our three statistics (clinal P-value, seasonal P-value and composite statistic). We performed

permutation tests comparing the proportion of outlier to non-outlier SNPs within a gene to

the number expected from a random sample of SNPs (excluding SNPs in intergenic regions).

The permutation allowed us to control for the fact that different genes harbor different

number of SNPs.

Mitigating the impact of linkage disequilibrium

Selection at one site affects genetic variation at nearby, linked neutral sites (Smith and Haigh

1974). Because we assume that sites are independent in our models, the indirect effects of

selection can inflate the magnitude of the patterns we investigated. To test the effect of

linkage disequilibrium (LD) on our outlier analyses, we computed for all SNPs how their

distance to a top SNP affects their P-value or composite statistic. We then smoothed the

scatterplot using cubic splines as implemented in ggplot2 (Wickham 2016). To test the effect

of linkage on the relationship between clinal and seasonal variation, we implemented a

thinning approach. Sampling one SNP per L base pairs one thousand times, we constructed

sets of SNPs with minimized dependency, where L ranged from 0 to 20kb. For each of these

sets for a given L, we regressed clinal against seasonal slopes, and compared the distribution

of the thinned regression coefficients to the coefficients we obtained using all SNPs.

Inferring proportion of selected SNPs

Although a negative, significant relationship between clinal and seasonal variation indicates

that a subset of the polymorphisms is under selection, it is difficult to measure how a given

slope is related to a biological parameter, such as the proportion of selected SNPs. Most of

the observed polymorphisms should be neutral with only a few under direct selection,

therefore we expect the relationship between clinal and seasonal change to be minor (i.e., for

most SNPs clinal and seasonal change should be unrelated). To get a sense of how the

proportion of SNPs with uncoupled clinal and seasonal change would impact our measure of

interest, the regression coefficient between clinal and seasonal variation, we performed a set

of simple simulations.

The polymorphisms most affected by the environment are going to be the ones that

change most consistently with seasons and latitude. For the subset of polymorphisms under

selection, we expect the relationship between the z-normalized regression coefficients for

latitude and seasons to be negative one. This is equivalent to assuming that the rank order of

clinal and seasonal change should be the same. Biologically, this would happen if the

environment changed in the same ways with latitude and seasons. Although this is probably

not the case, we stress that this assumption makes our test conservative, given variants under

latitudinal and seasonal selection might not fall along the 1:-1 line, even though we are

assuming they should.

Under this framework, we simulated datasets with different proportions of points

falling along a line ($y = -x$), representing selected SNPs and points without a linear

relationship, representing SNPs that are not under selection. For each simulation replicate, we

calculated the linear relationship between two variables with varying proportions of points

falling on the 1:-1 and points completely unrelated. Each value was drawn from the combined

distribution of observed z-normalized regression coefficients for season and latitude. By

calculating the observed slope between the regression coefficients of each of our simulated

datasets, we could establish a relationship between observed slope and the expected

proportion of SNPs that fall along 1:-1 line (or that are under selection). We then used

Approximate Bayesian Computation using a sequential sampling scheme (as implemented in

the R package EasyABC; Jabot et al. 2013) to estimate the posterior distribution of

proportion of selected SNPs that could explain our observed relationship between clinal and

seasonal change (using our thinned estimate to alleviate the effects of linkage

disequilibrium).

Results

We assembled 21 *D. melanogaster* population samples collected from seven localities across

multiple years in the east coast of the United States. All of these samples are the result of a

collaborative effort of many researchers from a consortium, the DrosRTEC (Bergland et al.

2014; Machado et al. 2018). Seven of our samples span from Florida to Massachusetts and

together comprise our clinal set. The seasonal samples were collected in Pennsylvania in the

spring (6 samples collected in June or July) and in the fall (7 samples collected in September,

October or November). For each sample, a median of 55 individuals (with a range of 33 to

116) was pooled and resequenced to an average 75x coverage (ranging from 17 to 215). We

also used four clinal samples from the Australia (Anderson et al. 2005; Kolaczkowski et al.

2011). More details about the samples can be found on Table S1 (also see Machado et al.

2018; Bergland et al. 2014). After all the filtering steps, we identified 572,914 common

autosomal SNPs, which were used in our downstream analyses.

Latitude explains more variation than seasons

Latitude explains much of allele frequency variation along the surveyed populations, as there

is an excess of low GLM P-value SNPs (Fig. S1A). The mean absolute difference in allele

frequency between the ends of the clines is 9.8%. Seasons, on the other hand, explain less of

the variation in allele frequency. There is a minor excess of low GLM P-value SNPs (Fig.

S1B) and the mean absolute difference in allele frequency between seasons is 2.75%.

344   Given we do not have enough information on the selective pressures and migration

patterns in nature to build an appropriate null distribution to calibrate our P-values, we sought

to demonstrate that top significant clinal and seasonal SNPs are enriched for true positive

SNPs that underlie adaptation. Latitudinal SNPs are more likely to be in exonic, UTR 3' and

UTR 5' regions (Fig. 1A), whereas seasonal SNPs are enriched for exonic and UTR 5'

regions (Fig. 1B). The enrichment of top latitudinal SNPs among functional genic classes has

been reported by previous studies which used different latitudinal samples (Fabian et al.

2012; Kolaczowski et al. 2011, Machado et al. 2016). Further, enrichment of top seasonal

SNPs among functional regions have been observed before (Bergland et al. 2014; Machado et

al. 2018). The distribution of the top SNPs along the genome is shown in Fig. S8 and see Fig.

S2 for the enrichment test using a 5% cut-off. Our P-values were obtained from a

permutation test in which we shuffled season and latitude labels to build our null distribution.

Clinal variation is related to seasonal variation

A clinal pattern can arise solely as a result of demographic processes, such as isolation by

distance and admixture (Duchen et al. 2013; Kao et al. 2015; Bergland et al. 2016). Although

seasonality is less affected by such processes, seasonal change is less pronounced and more

subject to stochastic changes in the environment, making it harder to detect seasonal change

with precision. Here, we integrate both clinal and seasonal change estimates across a large

number of SNPs in the genome (about 580,000 SNPs). We expect the overall pattern that

emerges to be informative of the relative role of natural selection, because selection is a

plausible process to produce a pattern of clinal variation mirroring seasonal variation (Cogni

et al. 2015). With higher latitudes, cold-favored alleles rise in frequency, whereas from spring

to fall, the frequency of cold-favored alleles decreases. Conversely, with higher latitudes,

warm-favored alleles decrease in frequency, whereas from spring to fall, the frequency of

warm-favored alleles increases.

To assess if clinal variation and seasonal variation are related, we performed a linear

regression of the slopes of clinal and seasonal allele frequency change (see Methods). We

find a significant negative linear relationship between clinal and seasonal regression

coefficients (Fig. 2A, S3, S4 and S5; Table S2). The relationship is strongest for SNPs within

exons, and the weakest for unclassified SNPs and those within intergenic regions (Fig. 2A;

Table S3 and S5). Nonetheless, the relationship is different than zero for all classes (except

for the unclassified), what would be consistent either with pervasive linked selection or

widespread distribution of variants that are important for adaptation, even within non-coding

regions. We stress that our results are robust to modelling assumptions, as they are significant

when compared to the datasets with permuted of latitude and season labels (Fig. 2A).

Given that previous studies have demonstrated the importance of cosmopolitan

inversions in climatic adaptation (Kapun et al. 2016), we looked at how clinal and seasonal

variation are related within common cosmopolitan inversions. Our results are in concordance

with previous findings, in that SNPs within the region In(3R)P spans have a strikingly strong

negative relationship between clinal and seasonal variation (Fig. 2B; Table S4). It is

noteworthy that the difference between the genome-wide estimate and the estimate using

only SNPs outside inversions is small, suggesting that inversions alone do not explain our

main result, and variants relevant for adaptation are widespread across the genome.

Another way of testing for parallelism between clinal and seasonal change is by

testing if clinal SNPs are more likely to be seasonal. We observed that clinal SNPs are

enriched for seasonal SNPs (Fig. 2C). The enrichment increases with more stringent lower P-

value quantile cut-offs, as we would expect if even strictly non-significant variants were

informative of the role of selection.

We also confirmed our main finding, that clinal and seasonal change are related using

samples from Australia. There is a genome-wide relationship between clinal variation in

Australia and seasonal variation in Pennsylvania that, although minor, is significant (Fig.

S6A). We also found a relationship between latitudinal regression significance and seasonal

significance using the Australian samples (Fig. S6B). To measure clinal change in Australia,

we only used four low coverage samples in Australia, two for each of low and high latitude

locations, thus our power was tremendously reduced.

Top SNPs for clinal and seasonal variation are enriched for metabolic functions

Our approach to detecting selection relies on a large set of SNPs, for which we fit a

regression of latitudinal against seasonal regression coefficients. It is not possible, however,

to single out individual polymorphisms and genes that are being driven by spatial and

seasonal selection. Thus, we implemented a composite statistic that incorporates clinal and

seasonal P-values, as well as the difference in sign (direction) of clinal and seasonal

regression coefficients.

The composite statistic outliers greatly overlap with clinal and seasonal variants (Fig.

S8). The overlap is greater among clinal and composite sites, as expected given clinal P-

values are generally much smaller than seasonal P-values. Nevertheless, the statistic

incorporates the difference in sign that can be helpful in distinguishing spurious from causal

variants involved with adaptation. Consistent with this reasoning, even though about one

third of the composite outliers are unique, outlier SNPs for the composite statistic are

enriched among exonic, UTR 3', and UTR 5' regions and depleted within intergenic regions

(Fig. 1C). The enrichment among genic regions seen for the composite statistic is comparable

to those for latitudinal P-values, suggesting that incorporating both clinal and seasonal

variation can inform us adaptive processes in natural populations.

416   We characterized the biological function of the outlier SNPs for each of these sets:

clinal SNPs, seasonal SNPs, and top composite SNPs. At an FDR corrected P-value < 0.05,

we found enrichment for 84 GO terms for the clinal SNPs, no enrichment for the seasonal

SNPs and five GO terms for the composite statistic SNPs (Tables S6, S7, S8). Although

caution is needed when interpreting GO analysis results (Pavlidis et al. 2012), some of these

enriched terms are involved in metabolism. These results agree with the previous observation

that metabolic genes are central to clinal adaptation and vary latitudinally and seasonally

(Lavington et al. 2014; Cogni et al. 2015).

424   There are many examples of genes that were empirically implicated with clinal and

seasonal adaptation in *D. melanogaster* (Vigue and Johnson 1973; Oakeshott et al. 1982;

Berry and Kreitman 1993; Schmidt et al. 2000; Verrelli and Eanes 2001; Duvernell et al.

2003; Sezgin et al. 2004; Schmidt and Paaby 2008; Cogni et al. 2014; Paaby et al. 2014;

Behrman et al. 2018). We thus tested whether those genes were enriched for top clinal,

seasonal and composite statistic SNPs (i.e., SNPs within introns and exons of a gene). Out of

10 genes with strong support from previous studies, four had fewer than ten SNPs in our

dataset, and thus we had no power to look for enrichment. Two out of the other six genes

were enriched for top clinal SNPs, one for top seasonal SNPs, and three for the composite

statistic (Table 1). We found an excess of top latitudinal, seasonal and composite SNPs in the

couch potato gene (*cpo*), which is associated with diapause and is known to harbor SNPs that

vary clinally and seasonally (Schmidt and Paaby 2008; Cogni et al. 2014). The insulin

receptor (*InR*) gene, which was shown to have seasonal and clinal variants (Paaby et al.

2014), is enriched with both latitudinal and composite SNPs (Table 1). Adh, a metabolic gene

implicated in climatic adaptation, shows an excess of top composite SNPs (Vigue and

Johnson 1973; Berry and Kreitman 1993; Sezgin et al. 2004). The genes *Tep2* and *Tep3*,

which have been associated with seasonal variation in immunity (Behrman et al. 2018), do

not show an excess of outlier SNPs for any of our statistics (Table 1). These results further

highlight the utility of the composite statistic, and could be used to guide future functional

studies.

The effects of linkage disequilibrium on clinal and seasonal variation

Variation at one site is linked to variation at other sites, and selection will increase this

dependency. To mitigate this effect, we first assessed if latitudinal and seasonal P-values, and

our composite statistic were dependent on how distance a SNP was from our top 1% SNPs.

We show that all statistics are dependent on distance from our outlier SNPs (Fig. 3A-C), but

the effect virtually disappears after 5kb.

450   Our regression analysis of clinal and seasonal variation assumes that SNPs are

independent. Thus, we assessed the impact of linkage on the relationship between clinal and

seasonal change by implementing a thinning approach. First, we tested how the genome-wide

regression estimate varied with changing window sizes. The dependency among sites affects

our estimates, but levels off after 5kb (Fig. 3D). Nevertheless, we chose a conservative

window length of 10kb for downstream analyses. The strength of the genome-wide

relationship between clinal and seasonal variation decreased from -0.041 to -0.028 when

randomly sampling one SNP per 10kb (P= 0.0449), but it still remained significantly

different from zero (P= 0.0029; Fig. 3E). The thinning did not significantly impact the signal

for exonic regions, but the strength of the signal within intronic regions significantly

decreased (P= 0.0209; Fig. 3E). For many of the thinned datasets, the regression coefficient

within intronic and intergenic regions reached values close to zero, as we would expect if

most of the signal within these regions were caused by linked selection. This suggests that the

approach is efficient in mitigating the impact of linked selection on our estimates.

Selection accounts for a substantial proportion of genome-wide clines

Although we identified a significant negative relationship between clinal and seasonal

variation, the magnitude of the regression coefficient appears small (10kb bootstrap estimate

$\beta = 0.0277$). It is hard to measure the biological relevance of such coefficient, and so we

designed a simple simulation approach that translates our statistic of interest into a biological

parameter, that is the proportion of selected sites. We assume that: (i) for the subset of

variants that are under selection, we would expect to see a perfectly linear relationship

between z-normalized clinal and seasonal regression coefficients; (ii) for the variants that are

not contributing to adaptation, then we would expect absolutely no relationship between

clinal and seasonal variation; (iii) variants are not linked. Assumption (i) renders our test

conservative, as the relationship would not necessarily be 1:-1 because some variants may be

strongly seasonal but not necessarily strongly latitudinal and vice-versa. Accounting for

linkage disequilibrium (assumption iii) is notoriously complicated, especially because we

cannot accurately measure LD from pooled sequencing (Feder et al. 2012). However, we

show that most of the effects of autocorrelation can be accounted for with our thinning

approach, thus we used the observed relationship between clinal and seasonal change with

10kb thinning.

We asked what the expected relationship between clinal and seasonal regression

coefficients would be if some of the variants were subject to spatially and seasonally varying

selection but most were neutral. To assess this, we built datasets with varying proportion of

adaptive variants (with linear relationship between clinal and seasonal variation) to neutral

variants (no relationship between clinal and seasonal variation) and calculated the expected

linear relationship between the two variables.

We found that the observed regression coefficient is consistent with approximately

2.8% of the common variants being under selection (Fig. 4), with 95% of the posterior

estimates ranging from 2.5% to 3%. Thus, our analysis suggests that a sizable fraction of

common, autosomal SNPs changes due to selection.

Discussion

Clinal patterns have been observed in both phenotypic and genotypic traits in many different

species (Hancock et al. 2008; Baxter et al. 2010; Adrion et al. 2015). Especially in systems in

which there is collinearity between the axis of gene flow and environmental heterogeneity,

disentangling the contribution of selection and demography to producing clines is not trivial.

Detecting seasonal cycling in allele frequencies is also challenging, mostly because the effect

size is likely to be small and the environment changes unpredictably within seasons. We

relied on integrating patterns of clinal and seasonal variation (Cogni et al. 2015), since

variation over short periods is independent of most confounding demographic processes

(Bergland et al. 2014). Parallel environmental change and the differences in the onset of

seasons with latitude can create parallelism between clinal and seasonal variation. Natural

selection would be the process driving the coupling between latitudinal and seasonal change

in both scenarios, thus investigating the relationship between clinal and seasonal change can

help discern the role of natural selection. Here, using *D. melanogaster* samples from the east

coast of the US, we show that (i) there is a genome-wide pattern of clinal variation mirroring

seasonal variation, (ii) this is consistent with the action of natural selection, and (iii) a

significant number of common variants are being driven by selection.

We show that clinal and seasonal change are related, as there is a significant negative

relationship between clinal and seasonal regression coefficients in the *D. melanogaster*

genome (Fig. 2A), and there is an enrichment for seasonal SNPs among clinal SNPs (Fig.

2C). The relationship between clinal and seasonal regression is stronger for genic when

compared to intergenic regions (Fig. 2A). Demographic processes would impact the genome

as a whole, but variants in genic regions are more likely to impact fitness (Andolfatto 2005).

We estimate that about 2.8% of common, autosomal SNPs should be under selection to

explain the genome-wide pattern of clinal and seasonal concordance uncovered (Fig. 4).

Further, we show that some genes previously known for their role in local adaptation are

enriched for variants that vary seasonally and latitudinally (Table 1).

Chromosomal inversions segregate along the cline and seasonally in Pennsylvania,

and thus could impact our estimates (Kapun et al. 2016). Although inversions do contribute

to adaptation (Fabian et al. 2012; Kapun et al. 2016), we demonstrate that the genome-wide

pattern of clinal reflecting seasonal variation is not being driven solely by inversions (Fig.

2B) and previous studies have also shown that not all clinal and seasonal variation can be

attributed to inversions (Fabian et al. 2012; Bergland et al. 2014; Machado et al. 2018). More

generally, because we expect selection to intensify linkage disequilibrium, the relationship

between clinal and seasonal variation could be mostly driven by a few large effect loci.

However, we show that our results cannot be fully explained by linked selection (Fig. 3).

When accounting for autocorrelation along the chromosomes, the relationship between clinal

and seasonal change disappears for variants within intergenic regions, but not for exons (Fig.

3E). The effects of linkage disequilibrium are important in this short time scale, but it decays

rapidly, and returns to background levels after 5kb (Fig. 3A-D). Therefore, our results

support a scenario where the genome-wide pattern of clinal variation mirroring seasonal

variation is a result of selection acting on many sites across the genome.

An important mechanism that can cause clinal patterns has been neglected from

recent discussions of clinal variation in *Drosophila*. The latitudinal variation on the onset of

seasons can produce clines, a phenomenon termed "seasonal phase clines" (Roff 1980;

Rhomberg and Singh 1986). Under this model, a correlation between clinal and seasonal

change is expected. Our latitudinal samples were all collected within one month of difference

(during the spring), and so our observations could be partially explained by differences in the

seasonal phase. Seasonality may be causing some of the patterns we uncovered, as we found

the relationship between clinal and seasonal change is strong when considering only the top

1% clinal SNPs (Fig. S5B), though the pattern is not replicated for the top 1% seasonal SNPs

(Fig. S5C). Our data does not allow for proper disentangling of seasonal phase clines from

parallel environmental change, but change on the onset of seasons alone cannot explain our

results. We found that latitude is usually a much better predictor of allele frequency

differences (Fig. S1), and the magnitude of change along the cline is much greater than what

we found within a population across seasons (9.8% vs. 2.75%). We show that including

Maine samples (which were sampled in the fall) in our analyses does not meaningfully

change our main results (Fig. S4). However, the magnitude of change in allele frequency

between Florida and Maine is smaller (mean change of 9.1% vs. 9.8% from Florida to

Massachusetts), which may be due to the difference in seasonal phase, as the samples from

Maine were collected in the Fall, but the sample from Massachusetts was collected in the

Spring.

Importantly, our results should be robust to differential admixture from Europe and

Africa to the ends of the clines (Duchen et al. 2013; Kao et al. 2015; Bergland et al. 2016).

Given that variation over seasonal time scales are less affected by broader scale migration

patterns, it is not plausible that secondary contact can create parallel clinal and seasonal

variation. Furthermore, the evidence for secondary contact in Australia is quite weak (see

Bergland et al. 2016), but we show that clinal variation in Australia is related to seasonal

variation in Pennsylvania (Fig. S6). Secondary contact may have contributed ancestral

variation, that have since been selectively sorted along the cline (Flatt 2016). Consistent with

this interpretation, the proportion of African ancestry is lower in low recombination regions,

suggesting selection is mediating admixture in *D. melanogaster* (Pool 2015).

Population substructure and migration could cause seasonal variation in allele

frequency in *D. melanogaster*. One possibility is that rural populations of *D. melanogaster* in

temperate regions collapse during the winter and recover from spring to autumn. However,

reproductive diapause cycles in orchards and reaches high frequencies early in the spring,

whereas its frequency in urban fruit markets in Philadelphia is much lower (Schmidt and

Conde 2006). Another possibility is that seasonal variation is produced by migration of flies

from the south in the summer, and from the north in the winter. There is little evidence that

long range migration produces clines in *D. melanogaster*, though this process seems

important in *D. simulans* (Bergland et al. 2014; Machado et al. 2016). Importantly, flies have

been shown to survive and reproduce during winter season in temperate regions, so rural flies

can withstand a harsh winter season and be subject to selection (Mitrovski and Hoffmann

2001; Hoffmann et al. 2003). More recent studies have also shown that fly populations adapt

to seasonal variation in outdoor cages (e.g., Rudman et al. 2019). These seasonal patterns

have been replicated in many populations across North America and Europe (Machado et al.

2018), bolstering the argument for seasonal selection. Regardless of the main process behind

seasonal change, comparing clinal and seasonal variation would still be worthwhile, as it

provides another way of independently assessing change in allele frequencies.

A recent study suggested the temporal change in allele frequencies reported in

Bergland et al. (2014) is not consistent with seasonal fluctuations driven by temporal

selection (Buffalo et al. 2019). Though we use a different set of samples, we show that the

distribution of seasonal P-values is close to uniform (Fig. S1), and there is only a slight

enrichment for coding SNPs. Finding truly seasonal SNPs is not trivial because the change in

allele frequency is generally much smaller and the populations are subject to stochastic

changes in the environment not related to seasons. For instance, Machado et al. 2018 found

that when they flipped the season labels of some samples the seasonal model fit was greatly

improved. On the other hand, finding SNPs driven by selection in space is hard because of

demography as a confounder. The main rationale behind our work is that integrating

signatures of clinal change with seasonal change can increase power to detect signatures of

selection. We show that the relationship between seasonal and latitudinal change disappears

when season labels are permuted (Fig. 2D). Thus, our results suggest that there is at least

some seasonal effect in our samples that is not pure noise.

A few previous studies have investigated the relationship between clinal and seasonal

variation (Bergland et al. 2014; Cogni et al. 2015; Kapun et al. 2016; Machado et al. 2018),

but here we further explore parallelism between clinal and seasonal variation to tease apart

the role of selection. Cogni et al. (2015) focused on a few SNPs in metabolic genes, and so

they could not draw robust conclusions about processes that impact most of the genome. Our

work is also fundamentally different from that of Machado et al. (2018), which revolves

around identifying seasonal variation in *D. melanogaster* by aggregating data from many

locations in North America and Europe. They also investigated the relationship between

clinal and seasonal variation, but do not interpret how parallel clinal and seasonal change can

help us identify the role of selection. In addition, because our interest is to ultimately

understand processes driving clinal variation in the east coast of the US, we only included

seasonal samples from Pennsylvania. This location had the greatest number of years surveyed

(7 years, as opposed to 2 years for most of the other locations; see Machado et al. 2018), and

it is the population for which we have the most information about how phenotypes vary

seasonally (Schmidt and Conde 2006; Behrman et al. 2015; Rajpurohit et al. 2017; Behrman

et al. 2018).

Many species occur along spatially structured environments and show clinal variation

in traits, so a question that remains open is: what is the role of selection in producing and

maintaining these patterns? Seasonal variation is also ubiquitous, especially in temperate

environments, so seasonal change could be an important feature of organisms that have

multiple generations each year (Behrman et al. 2015). Here, we demonstrate that by

integrating clinal and seasonal variation, we can discern the contributions of selection in

driving changes with the environment. This approach could potentially be applied to other

multivoltine species that occur along environmental gradients, including invasive species,

which are known to often have short generation times and to reproduce quickly (Sakai et al.

2001).

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Data accessibility

All the data used in this project are available on the NCBI Short Read Archive (BioProject

accession numbers PRJNA256231 and PRJNA308584, and NCBI Sequence Read Archive

SRA012285.16).

Author contributions

M.F.R. and R.C. designed the research. M.F.R. performed the research and analyzed the data.

M.F.R., M.D.V. and R.C. discussed the results and conclusions. M.F.R. wrote the manuscript

with input from M.D.V and R.C.

Table 1. Enrichment for outlier SNPs in genes involved with clinal and seasonal adaptation.

Gene	SNPs	Top Clinal		Top Seasonal		Top Composite	
		Top SNPs	P	Top SNPs	P	Top SNPs	P
Adh	11	1	0.1033	0	1.0000	2	**0.0059**
cpo	821	32	**0.0001**	14	**0.0439**	25	**0.0001**
Gpdh	10	0	1.0000	0	1.0000	0	1.0000
Hex-C	2	0	-	0	-	0	-
InR	167	25	**0.0001**	2	0.4928	40	**0.0001**
mth	7	0	-	0	-	0	-
Pgm	1	0	-	0	-	0	-
Tep2/3	190	0	1.0000	0	1.0000	0	1.0000
Treh	28	1	0.2409	0	1.0000	1	0.2518
UGP	1	0	-	0	-	0	-

Figure 1. Top SNPs are enriched for functionally relevant classes. Enrichment of top 1% SNPs in each genic class for A) latitudinal P-value, B) seasonal P-value, and C) composite statistic. The composite statistic takes into account the latitudinal and seasonal P-value, and the difference in sign between latitudinal and seasonal regression coefficients. Error bars are 95% CI. * P-value < 0.05, ** P-value < 0.005, *** P-value < 0.0005, **** P-value < 0.00005.

Figure 2. Clinal variation mirrors seasonal variation. A) Null distribution of the regression
coefficients between latitudinal and seasonal change for each genic class (after permuting the
latitude and season labels). Vertical bars represent the observed values. B) The relationship
between clinal and seasonal change depends on inversion statuses. C) Latitudinal and
seasonal significance are associated. Error bars are 95% CI. * P-value < 0.05, ** P-value <
0.005, *** P-value < 0.0005, **** P-value < 0.00005.

Figure 3. Effects of linkage disequilibrium. The A) latitudinal P-value, B) seasonal P-value
and C) composite statistics are dependent on the distance to the respective top 1% outliers.
The relationship between clinal and seasonal variation is affected by dependency among
SNPs. D) the strength relationship between clinal and seasonal variation decreases with the
size of the thinning window. E) comparison among original estimates (bar) and values
obtained after thinning using a window size of 10kb (point-range). Point-ranges show the
mean and standard deviation across simulations and gray dots in the background the values
for each simulation.
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Figure 4. Proportion of SNPs that should be under selection to explain the genome-wide
relationship between clinal and seasonal variation. A) background points represent each
simulation, and the line is the fitted loess curve. Dashed lines relate the observed scaled (or z-
normalized) regression coefficient after thinning (10kb) of our data to the expected
proportion of selected SNPs. B) posterior distribution of the proportion of selected SNPs,
given the observed relationship between clinal and seasonal variation (10kb thinned), with
horizontal line highlighting the mean.
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