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A molecular approach to studying Hymenoptera diets using polistine wasps

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The study of animal diets has benefited from the rise of high-throughput DNA sequencing applied to stomach content or faecal samples. The latter can be fresh samples used to describe recent meals, or older samples, which can inform about past feeding activities. For most invertebrates, however, it is difficult to access 'historical' samples, due to the small size of the animals and the absence of permanent defecation sites. Therefore, sampling must be repeated to account for seasonal variation and to capture the overall diet of a species. This study develops a method to describe the overall diet of social Hymenoptera based on a single sampling event, by analysing prey DNA from faeces accumulated in brood cells. We collected 48 nests from two species of introduced paper wasps (*Polistes chinensis*, and *P. humilis*) in the urban and peri-urban areas of Auckland, New Zealand, and selected two samples per nest. One from brood cells in the outer layer of the nest to represent the most recent diet, and one from brood cells in an inner layer to represent older diet. Diet differed between species, although both fed mainly on Thysanoptera, Lepidoptera, Diptera, Collembolla and Acariformes. Prey taxa identified to species level included both agricultural pests and native species. Prey communities consumed were significantly different between inner and outer nest samples suggesting seasonal variation in prey availability and/or a diversification of the wasps' diet as the colony grows. We also show for the first time potential predation of marine organisms by *Polistes* wasps.

KEYWORDS
eDNA, frass, metabarcoding, paper wasps, social insects, trophic interactions

1 | INTRODUCTION

In recent years, molecular detection of trophic interactions has become the standard to study the diet of invertebrates (Sheppard et al. 2005; Boyer et al. 2013; González-Chang et al. 2016). These methods rely on the sequencing of prey DNA present in regurgitates (Waldner & Traugott 2012), gut content (Olmos-Pérez et al. 2017) or faeces (Boyer et al. 2011) of predators (i.e. environmental DNA or eDNA). Often these studies are based on the analysis of fresh material upon the capture of live individuals. As such, these samples provide a snapshot of recently consumed food items whose DNA is still present in the gut of predators at the time of capture. Knowledge about temporal variation is essential to fully understand an animal's diet, but such data is difficult to collect and therefore under-represented in food web studies (McMeans et al. 2019). Nevertheless, significant seasonal variation in diet is observed in many taxa (Waterhouse et al. 2014; Lambert & Rothman 2015; Coulter et al. 2019; Amponsah-Mensah et al. 2019). The recent development of molecular techniques that can efficiently analyse low-quantity and low-quality DNA samples has been applied to the analysis of faecal samples for assessing diet (Monterroso et al. 2019, Waterhouse et al. 2014). This can be a particularly useful strategy for assessing seasonal variation in diet if faeces are deposited in permanent sites such as latrines, or other territory marking sites (Fretueg et al. 2015). In social nest-building wasps (Hymenoptera), faecal samples of larvae are deposited in permanent sites within the nest.

Social wasps live in colonies and build a nest organised around a caste system (Oster & Wilson 1978). During their development, the larvae of social wasps are fed a range of food, including other invertebrates, by adult wasps (Harris & Oliver 1993, Kasper et al. 2004, Todd et al. 2015). The larvae remain in individual cells in the nest until metamorphosis, while their faeces (or frass) accumulate at the bottom of their brood cell. This frass constitutes a historical record of the animal's diet throughout its development and, in multi-layered nests, we hypothesize that faeces in successive layers of brooding cells could be used to identify which prey species adult wasps have brought to their young over several weeks or months. Social wasps are considered pests in many regions of the world (Beggs et al. 2011; Lester & Beggs 2019), particularly species of Vespula and *Polistes* introduced into Australasia, Hawaii and South America (Clapperton et al. 1996; Matthews et al. 2000; Masciocchi & Corley 2013; Hanna et al. 2014). For example, in New Zealand, introduced Vespula cause significant problems particularly in beech forest ecosystems (Moller et al. 1991; Beggs et al. 2005) by monopolising the honeydew produced by native scale insects (Hemiptera: Sternorrhyncha) (Beggs & Wardle 2006). Through their feeding on this abundant resource, introduced wasps attain the highest densities in the world and compete for food resources with many native species including birds, bats, lizards and other native insects that are known to feed on honeydew (e.g. Harris 1991; Toft & Rees 1998; Beggs 2001; Elliott et al. 2010). Each year, social wasps also cause millions of dollars of damage to the New Zealand economy, primarily due to the impact of wasps on honeybees (Apis mellifera Linnaeus) and the loss of flow-on benefits to pastoral farming (MacIntyre & Hellstrom 2015). Beyond their economic and ecological impacts, wasps clearly impact human wellbeing through their painful and occasionally life-threatening stings (Golden et al. 2006). While the risks posed by these species are clearly recognised, they may also provide some ecological benefits if they replace functions of species lost from ecosystems through human impact (Beggs & Wardle 2006). Additionally, social wasps may have potential as biocontrol agents in agricultural systems, particularly *Polistes* because they are generalist predators of the main insect orders of crop pests, Lepidoptera and Diptera (Todd et al. 2015; Southon et al. 2019). A more comprehensive analysis of social wasp diets would allow a much better understanding of both the positive and negative impacts these insects have on other species.

The main aim of the study was to develop a new molecular method to study the diet of social wasps by retrieving prey DNA from faeces left by wasp larvae inside the nest. Historically, wasp diets have been studied by collecting and identifying the prey items carried by adult foraging wasps when they return to the nest to feed the larvae (Kasper et

al. 2004) or by dissecting the gut of adult wasps (e.g. Ward & Ramón-Laca 2013). These traditional recovery methods only provide a snapshot of the diet of the insect at a given point of time, while the method proposed here provides an overview of the diet of the colony throughout the lifespan of the nest. By applying our method to the nests of two sympatric polistine wasps, our analysis outlines their resource partitioning in urban and peri-urban areas in New Zealand, and their potential ecological role in relation to New Zealand invertebrates.

2 | MATERIALS AND METHODS

2.1 | Nest monitoring and sampling

Field collection of polistine nests was carried out in the Auckland region, New Zealand, during the peak of the 2017 season (i.e. January). Potential nesting sites were located based on previous occurrences recorded on iNaturalistNZ (URL: https://inaturalist.nz/), an open web platform built to record species occurrences across New Zealand. A total of 53 active nests were collected between 1st of March and 15th of May 2017 (Figure 1). After removing resident adults from the nest with commercial fly spray, nests were placed in individual sealable plastic bags in a cold storage container for transportation, before being stored at -80°C for optimal DNA preservation.

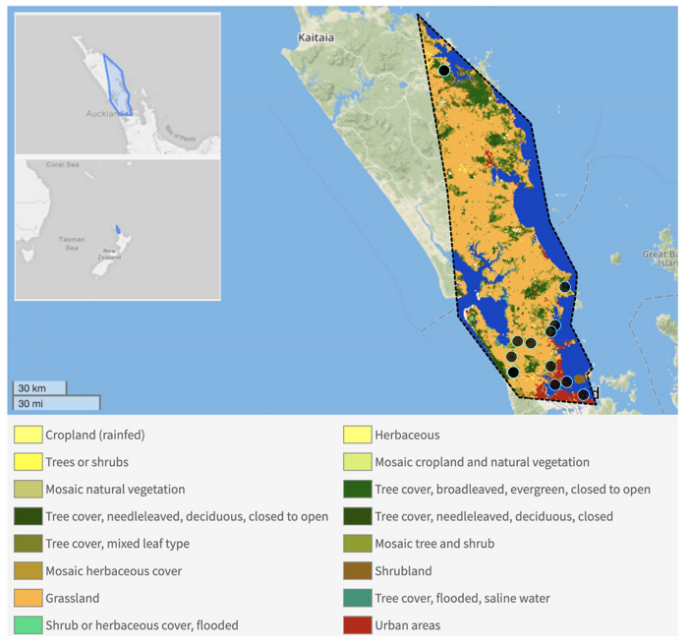

FIGURE 1 Location of samples collected in and around Auckland
Black dots correspond to areas where one or several nests were collected. See legend for colour code for land cover classification (from geofolio.org).

2.2 | DNA extractions

Among the collected nests, only the largest and better-preserved ones were used for the analysis (n=44 for *P. chinensis* and n=4 for *P. humilis*). Insect frass was sampled with sterile tweezers from one cell located on the outer (top) layer and from one cell located on the inner (bottom) layer of each nest. These two layers represent different batches of brood, hence they preserve a record of the diet of the colony at two different times. Faecal samples were ground directly in microcentrifuge tubes using a hand-held mortar and pestel and the DNA was extracted using a ZR Tissue & Insect DNA MicroPrep extraction kit (Zymo, Irvine CA) following an existing methodology for low quantity and degraded insect eDNA (Lefort et al. 2012). DNA concentration in the resulting eluates was measured by fluorometry using a Qubit® dsDNA HS Assay Kit (Life Technologies, Carlsbad, CA).

2.3 | DNA amplification and sequencing

Because prey DNA was likely to be highly degraded, primers targeting a short (313 bp) gene region of the mitochondrial gene COI were used for amplification. We selected the pair of primers mICOIintF (forward GGWACWGGWTGAACWGTWTAYCCYCC) and HCO2198 (reverse TAAACTTCAGGGTGACCAAAAAATCA), which are recommended for the amplification of invertebrate DNA (Leray et al. 2013; Lear et al. 2018). PCR reactions comprised 1 µL BSA, 10 µL Green GoTaq Mix, 0.6 µL of each primer [10 µM], 5.8 µL of ultrapure water, and 2 µL of template. A few recalcitrant samples were amplified using less template (0.5-1 µL) to limit the effect of potential PCR inhibitors. We used the touchdown PCR protocol recommended by Leray et al. (2013), with a 2 min initial denaturation (95°C) step. After the initial denaturation we conducted 16 cycles of 10 s denaturation (95°C), 30 s annealing (62°C -1°C per cycle) and 60 s elongation (72°C), followed by 25 cycles where the annealing temperature remained constant (46°C), a 7 min final elongation step was then carried out (72°C). According to standard protocol (Support Illumina 2016), all PCR products were purified using AMPure magnetic beads (Agencourt) and DNA concentration was standardised to 2 µM in all samples before pooling for high-throughput DNA sequencing. The resulting library was processed in high-throughput DNA sequencing analysis on one run of Illumina MiSeq using the 300 × 300 paired end protocol as recommended by the manufacturer. Ligation of individual barcodes, pooling of the libraries, and sequencing, were performed by New Zealand Genomics Limited, Auckland, New Zealand.

2.4 | Data analysis

The bioinformatic workflow was perfomed by NGBS (Nextgen Bioinformatic Services, New Zealand), based on the vsearch pipeline (Rognes et al. 2016) and included quality control, merging of paired end reads, removal of singletons, dereplication and chimera detection using uchime. The merged reads were then quality filtered and clustered at 97% identity to generate Molecular Operational Taxonomic Units (MOTUs). Following de novo and reference-based chimera detection, a final number of 1,436 MOTUs were detected, with an average read length of 364.7 bp (close to the expected amplicon size). Each MOTU was then compared to the NCBI NT database using BLASTn. To limit false positives and remove potential sequencing errors, only MOTUs for which read abundance within a sample was superior to 0.5% of the total were retained. To ensure reliable identification, only sequences for which the best hit had a query coverage over 70% were retained, which corresponded to an overlap of 250 bp or more between the query and the best hit. Retained MOTUs were identified at species level when the percentage identity was ≥ 98%, or assigned to the order of the corresponding best hit when the percentage identity was < 98%. MOTUs for which the best hit had more than 98% identity, but the species name was not available in Genbank (e.g. only genus or family name),

were searched against the Barcode Of Life Database (BOLD). Only MOTUs identified as terrestrial invertebrates were considered as potential prey. Reads corresponding to *Polistes* wasp DNA were used to confirm species identification of the wasp to which the nest belonged. MOTU confidently identified at the species level were categorised as native or introduced species based on Scott (1984) and the New Zealand Organisms Register (http://www.nzor.org.nz/).

2.5 | Statistical analysis

Because of an imbalance in the number of samples analysed for (n = 8) and *P. chinensis* (n = 88), we used the non-parametric Kruskall-Wallis test to compare the number of reads as well as prey richness between the two wasps species. To compare between inner and outer samples, we used the non-parametric Wilcoxon test to account for the non-normality and paired nature of the data. The cumulative number of MOTUs detected from the inner and outer layer of *P. chinensis*' nests was compared using a Koglomorov-Smirnov test. Because the analysis of only eight samples from four nests for *P. humilis* led to a low coverage of its diet, results for this species are only considered as indicative and a detailed analysis of prey community was only conducted for *P. chinensis*. Prey species assemblage differences between inner and outer samples of *P. chinensis* nests were investigated using an Analysis of Deviance on a multivariate generalised linear model. A negative binomial distribution was chosen for this analysis based on the dispersion of the residuals. In nests where more than 50 reads of marine origin were detected, the number of reads from commercial seafood and non-commercial sea organisms were analysed in relation to the distance to the sea using the non-parametric Kruskall-Wallis test. All analyses were performed with RStudio and the main packages used for statistical analyses were vegan (Oksanen et al. 2019) and mvabund (Wang et al. 2012).

3 | RESULTS

3.1 | Data quality and diet coverage

DNA was successfully amplified and sequenced from all faecal samples. A total of 7,767,737 high quality merged reads were obtained after clean-up of the raw illumina reads. Of these, 7,530,408 could be clustered at 97% identity in 1,436 MOTUs, which were then compared to the NCBI database and analysed thereafter (see supplementary Figure S1). An average of 79,967 DNA reads (± 6,412 s.e.) were obtained per sample. However, the vast majority of those reads (83%) corresponded to fungi, while 13.5% were from terrestrial invertebrate DNA, and 3% of the reads were other taxa, including marine organisms (Figure 2a). Lastly, 0.5% of the reads corresponded to DNA from the wasps themselves.

The number of reads per sample was not significantly different between the two species of wasps (KW, χ^2 = 0.32492, df = 1, p = 0.5687) (Figure 2b). Similarly, there was no significant difference for the number of invertebrate prey reads obtained per sample (χ^2 = 3.5433, df = 1, p = 0.05978), but a trend could be observed showing an average number of prey reads four times greater in *P. humilis* than in *P. chinensis*. No differences in the number of reads per sample were observed between the two wasp species with regard to contamination (χ^2 = 0.029289, df = 1, p = 0.8641) and marine organisms (χ^2 = 1.3172, df = 1, p = 0.2511). When comparing inner and outer samples, significantly more reads were obtained from outer samples (Wilcoxon, V = 376, p = 0.02916) (Figure 2b). However, this difference was mainly due to reads considered as contaminants (V = 244.5, p-value = 0.04297) and DNA of marine origin which was five times more abundant in outer samples than in inner samples (V = 299, p = 0.005126) (Figure 2c). In contrast, no differences were found in terms of reads from terrestrial invertebrates when comparing inner and outer samples (V = 524, p = 0.5182). Only 11% of the MOTUs corresponding to terrestrial invertebrates could be identified to the

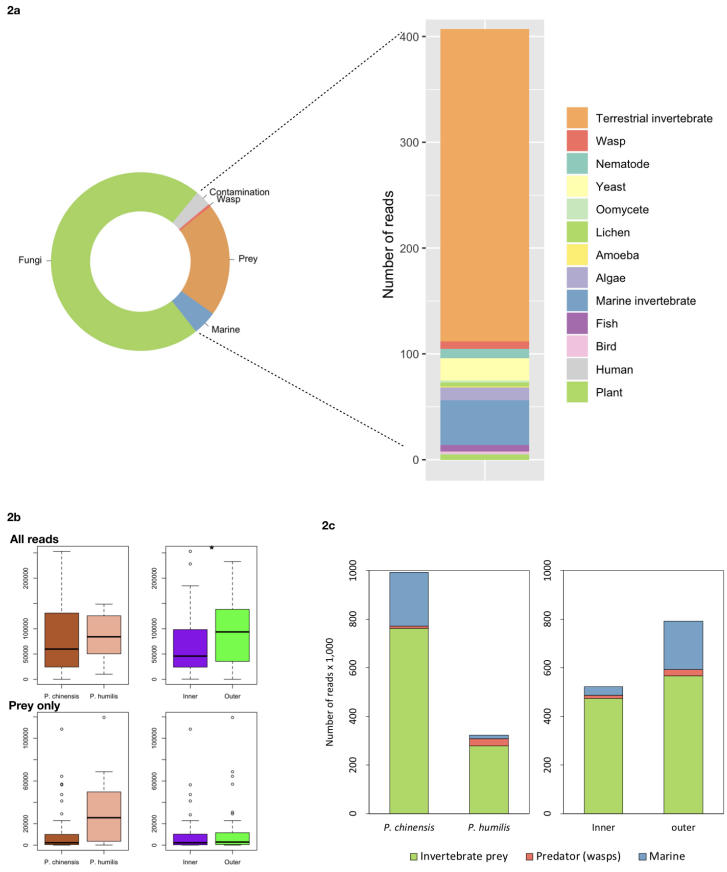

FIGURE 2 Detection of organisms in the faeces of wasp larvae
2a. Proportion of MOTUs corresponding to the main categories (left hand-side) and detailed categories excluding fungi (right hand-side) 2b. Number of reads per sample in relation to wasp species (left boxplots), and number of reads in inner versus outer samples (right boxplots) 2c. Total number of reads corresponding to marine, predator and terrestrial invertebrate prey DNA after removal of fungal DNA in both wasp species (left) and in outer versus inner samples (right).

species level, corresponding to 21 species of prey in the diet of *P. chinensis* and eight species in the diet of *P. humilis*. (Table S1, Supplementary figure S2). Therefore, the diet analysis was mainly performed at a higher taxonomic level (order) to provide a greater coverage of the wasps' diet (i.e. based on all invertebrate MOTUs). Using 88 samples from *P. chinensis* nests, 260 MOTUs were detected, corresponding to an estimated 91% of the species' overall terrestrial invertebrate diet (Figure 3). There was no significant difference in the cumulative number of MOTUs detected from inner and outer samples of *P. chinensis* nests (KS, D = 0.25, p = 0.1282) (Figure 3). For *P. humilis*, 141 MOTUs were detected in the eight samples analysed, which only provides a fraction of this species' diet (Figure 3).

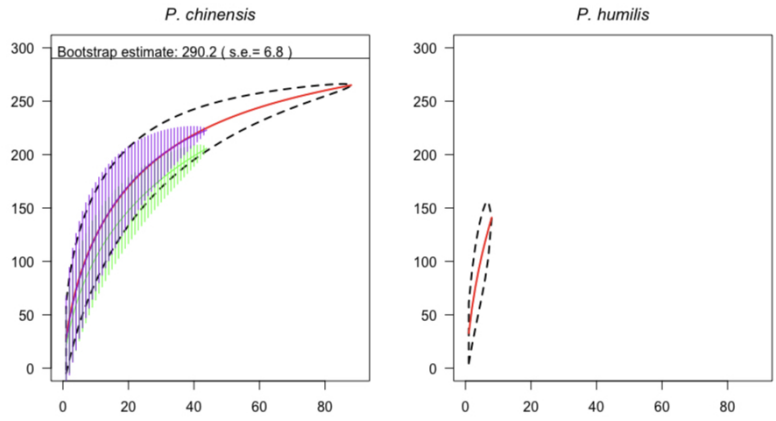

FIGURE 3 Detection of organisms in faeces
Cumulative curves of number of invertebrate prey MOTUs in relation to number of samples. Left: *P. chinensis*, right: *P. humilis*. Cumulative curves are in red, the area delimited by the dashed lines corresponds to the 95% confidence interval. On the left, the horizontal solid line represents the estimated total number of prey MOTUs in the diet of *P. chinensis*. The purple and green hatched areas correspond to cumulative curves obtained with inner and outer samples respectively.

3.2 | Prey identification

When considering only terrestrial invertebrates, the most common prey in the diet of *P. chinensis* belonged to the orders Thysanoptera, Lepidoptera, and Diptera, detected in 83%, 54%, and 35% of the respective samples (Figure 4a). The diet of *P. humilis* was also characterised by the strong presence of Thysanoptera, followed by Collembola and Lepidoptera, detected in 50%, 38% and 25% of the respective samples. At the MOTUs level, the prey community composition (based on RRA) varied with species (Analysis of Deviance: Dev1,94 = 708.2, p = 0.004) and to a lesser extent with the location of the samples in the nest (Dev1,93 = 574.7 p = 0.027). A significant interaction between species and sample location was also detected (Dev1,92=139.9, p = 0.030). Similar results were obtained at the order level, with regard to differences in prey community composition between wasp species (Dev1,94 = 114.19, p = 0.001) (Figure 4b), but not between inner and outer samples (Dev1,94 = 57.78, p = 0.050) (Figure 4b). Individual samples contained between 1 and 10 different orders of prey (Figure 4c), and prey richness was significantly greater in outer samples than in inner samples both at MOTU (V = 324.5, p = 0.01845) and at order level (V = 206, p = 0.00969). A total of 32 MOTUs could be identified at the species level corresponding to 23 species, most of which were lepidopteran species (18/23) (Table S1). Among these, eight were native species and 15 are considered introduced in New Zealand. The latter includes at least four serious agricultural pest species including the cosmopolitan Armyworm (*Mythimna separata* Walker), present in 20% of *P. chinensis* samples.

Over 236,000 reads clustered in 62 MOTUs were identified as marine organisms, mainly corresponding to polychaetes, fish, molluscs and cephalopods (Figure 5). Of these, 93% were recorded in the nests of *P. humilis* (Fig 2c). The vast majority of marine reads were from polychaete worms (Figure 5) and clustered in 31 MOTUs, all matching the genus *Dasybranchus* with 86 to 94% of identity. The only marine MOTU that could be identified to species with confidence (i.e. percentage identity ≥ 98%) was the squid *Nototodarus gouldi* (McCoy). This species was detected in

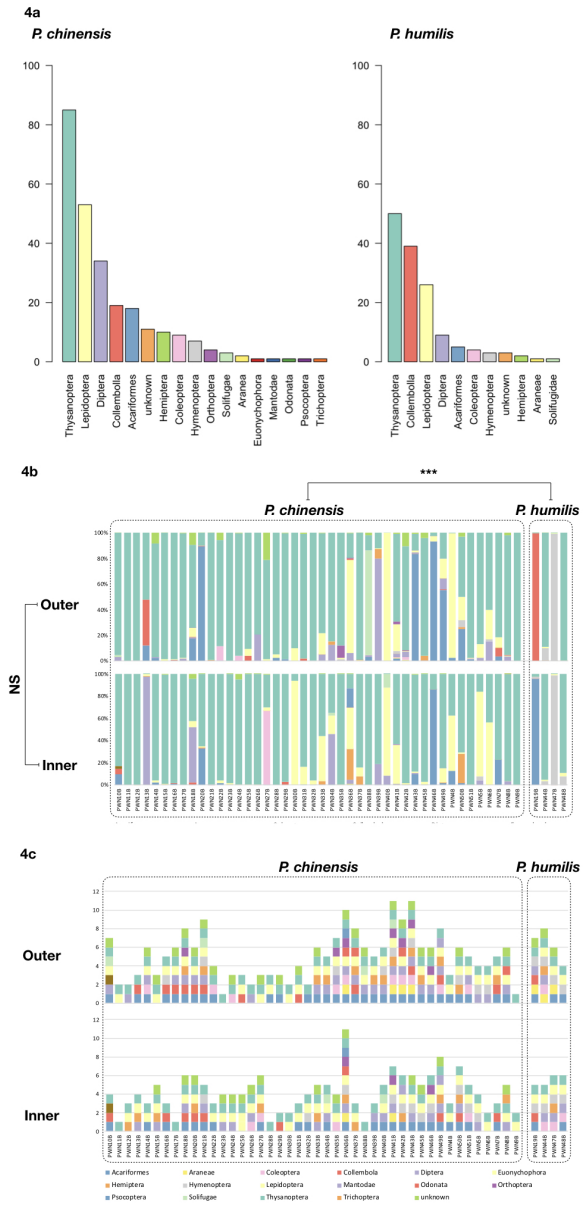

FIGURE 4 Diet composition of two Polistes wasp species from urban and sub-urban areas of Auckland (New Zealand), based on DNA analyses of larval faeces in nests (n=88 for *P. chinensis*; n=8 for *P. humilis*).
4a: Frequency of occurrence of different invertebrate orders in the diet of *P. chinensis* (left) and *P. humilis* (right). 4b: Relative read abundance of prey taxa as measured from each individual faecal sample (i.e. for each individual wasp larvae) in inner and outer cells of the nest. 4c: Occurrence of the different prey genera, in the diet of each individual wasp larvae.

the nests of both wasps and was present in inner and outer samples. There were no significant differences in the number of reads in nests located close to the sea (< 1 km) compared to those located further inland (> 1 km) both for seafood (KW, χ^2 = 1.0953, df = 1, p = 0.2953) and non-seafood marine organisms (KW, χ^2 = 0.15341, df = 1, p = 0.6953) (Figure 6).

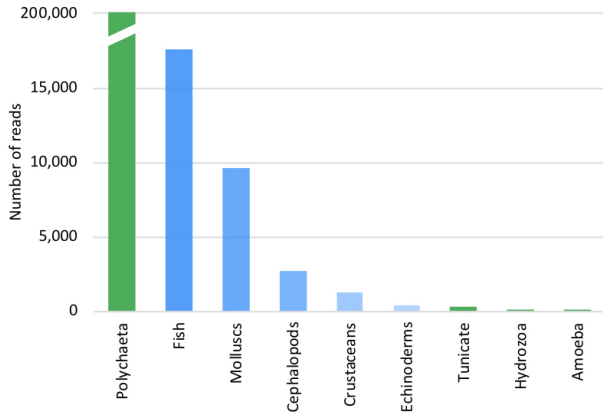

FIGURE 5 Marine organisms detected in the faeces of paper wasp larvae.
Blue bars correspond to potentially commercial sea products, green bars correspond to non-commercial taxa.

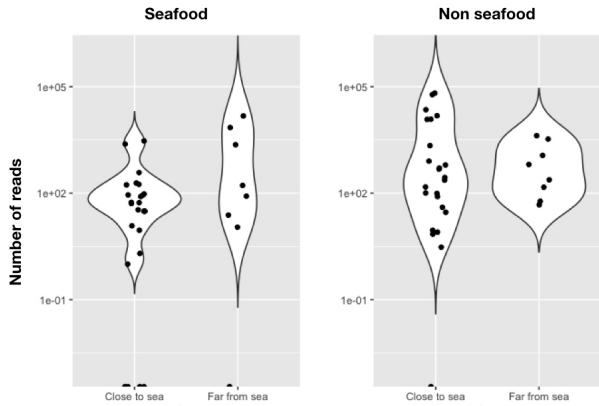

FIGURE 6 Number of reads for DNA of marine origin in relation to distance to the sea.
Samples collected within 1 km of the coast were considered close to sea while samples collected at more than 1 km inland were considered as distant from the sea. Only samples with more than 50 reads from marine origin are represented. Seafood include fish, molluscs, cephalopods, crustaceans and echinoderms, non-seafood include Polychaetes, Tunicates, Hydrozoa and Amoeba.

4 | DISCUSSION

4.1 | Method efficiency and improvements

The tested method led to the successful amplification of prey DNA from paper wasps' nests and the description of the diet of two polistine wasps in urban and peri-urban areas around Auckland, New Zealand. We used generalist degenerated PCR primers with the aim of amplifying a wide prey spectrum without a priori selection of particular taxa. However, wasp nests appeared to harbour a wide variety of fungi, which were strongly amplified by our primers (1,029 OUT detected). Although we still obtained good numbers of prey reads for most samples, the strong presence of fungal DNA (83% of all sequences generated) suggests a more targeted selection of primers is needed when working on wasp nests. Indeed, the sequencing power lost to fungal DNA would be better used to increase the sequencing depth of prey reads or multiplex more samples in the same sequencing run. Another crucial limitation lies in our inability to identify most prey taxa to species level (Wheeler 2018). The lack of taxonomic resolution encountered here is likely due to the limited availability of sequences for New Zealand invertebrates. Despite the high-resolution power of the chosen primers (Leray et al. 2013), the average percentage identity of prey MOTUs was only 88% (Supplementary Figure S2), which is far from the 98% threshold generally used for identification of invertebrates at the species level.

4.2 | The case of marine prey

A large number of reads corresponding to species of marine origin were obtained from the faeces of wasps, in particular that of *P. humilis*. To our knowledge, predation of marine organisms by Polistes wasps has never been reported before. The great majority of reads belonged to marine worms, probably from the genus *Dasybranchus* Grube, 1850, which includes species present in sand, rocky intertidal regions and shallow waters (e.g. Dean 2016; Mclachlan & Defeo 2018). Such species could be exposed at low tide and may have been targeted by the wasps. However, we found no significant relationship between distance to the sea and amount of reads from marine origin. Some of the other marine organisms detected in our samples are unlikely to have been predated by the wasps, such as squid. The presence of their DNA may correspond to true consumption, however, they probably reflects insects feeding on carrions washed up by the tide, or by-products of human activities (e.g. markets, fishing harbours, food waste). Many of the taxa detected include commercial sea products (fish, crustaceans, cephalopods, echinoderms), which would have been available near harbours or human habitations. Hence, these reads may not represent the natural diet of polistine wasps, however, they do suggest that human activity could strongly influence the diet of some colonies by providing an alternative food source. The greater presence of DNA from marine origin in outer samples (five times more abundant than in inner samples) could suggest a seasonal effect of human fishing activity or/and external contamination by sea spray. Such contamination is particularly likely for non-commercial life forms, mainly represented by polychaete worms, which release massive amount of DNA during swarming events.

4.3 | Diet composition and ecological role of Polistes wasps

The saturation of the MOTU accumulation curve (Figure 3) suggests that the diet of *P. chinensis* was well covered by our analysis. On the other hand, the low number of samples analysed for *P. humilis* did not allow an accurate characterisation of its diet. For the latter species, our results should therefore be taken as indicative only. Our results showed that the two species of wasps displayed a large overlap in their dietary niche (Figure 7) with 147 MOTUs

in common. For both species, the diet was dominated, both in terms of relative read abundance and percentage of occurrence, by Thysanoptera (Figure 3a, b), a group known to contain common pests such as thrips. None of the 85 MOTUs corresponding to Thysanoptera (totalling 670,850 reads) could be identified to species level. However, all Thysanoptera MOTUs pointed to the same best hit (with 82 to 92% identity), which was a species in the Phlaeotripidae family. This family is known to comprise a large number of fungus-feeding thrip species that are endemic to New Zealand (Mound & Walker, 1986). Another important group, especially in the diet of *P. chinensis*, was Lepidoptera. Identification to the level of species was successful for about a third of the lepidopteran MOTUs (Supplementary Figure S2). The improved identification power for this order is a direct consequence of the significant effort towards describing and barcoding the Lepidoptera fauna of New Zealand (Ball & Armstrong 2006). Lepidopteran prey included three notorious pest species: M. separata, Thysanoplusia orichalcea (Fabricius), and *Chrysodeixis eriosoma* (Doubleday), found in 41%, 16% and 14% of P. humilis samples, respectively. However, the diet of P. chinensis also comprised seven native lepidopteran species (mainly Noctuidae, Torticidae and Geometridae). Native and introduced Lepidoptera were also detected in the diet of P. humilis and identified to species level. The only pest species that could be identified to species level was the sawfly *Caliroa cerasi* Linnaeus. The ecological effetc of polistine wasps on pest species is therefore multifaceted. It is also interesting to note that the DNA of at least one parasitoid wasp, *Meteorus pulchricornis* (Wesmael), was detected in the nests of *P. humilis*. In New Zealand, this parasitoid is known to attack at least 20 lepidopteran species, both endemic and introduced (Berry & Walker 2004). Although these detections are likely to correspond to secondary predation (given the minute size of the adult parasitoid), by feeding on parasitized caterpillars, P. humilis could affect the parasitoid populations. These considerations highlight the complexity of the potential impact of Polistes wasp predation on New Zealand's invertebrate fauna.

4.3.1 | Seasonal variation in diet

The sizes of collected nests varied over the11 weeks of collection. Therefore, our samples were not suitable for a well-calibrated seasonal analysis of wasp diet. However, comparing samples taken from inner (older) and outer (younger) layers of the same nest does provide an estimate of how variable the colony diet was within the few weeks necessary to build one or more additional layers of brood cells. The composition of prey communities differed between inner and outer samples, which could reflect variation in prey availability at different times of the year. In addition, we detected greater prey richness in outer (younger) samples, which could indicate that the colony diet diversifies as the number of workers increases. However, this effect could also be explained by the higher degradation of specific prey DNA in inner (older) samples. We recorded greater amounts of contamination in samples located on the outer layers that were directly exposed to ambient air, than in samples located on inner layers that were somewhat sheltered inside the nest. The analysis of two faecal samples per nests (inner and outer samples) allowed for a greater coverage of each colony's diet. However, the number of samples per nest required to accurately estimate the diet of a whole colony may be greater and could significantly vary depending on the size of the nest and the species of interest. It is, for example, known that polistine nests usually contain between 20 to 400 cells, while some Vespula species can build significantly larger nests. For example, *Vespula germanica* Fabricius, can build nests comprising over 450,000 cells (Scott 1984) with a colony biomass of up to 600g (Malham et al. 1991). With large colonies in particular, it may be necessary to analyse many cells from the same layer of one nest to test whether a similar diet is provided to all larvae at a given time. Controlled feeding experiments may help to understand how food is divided among the brood. By providing known food items to a captive colony it would also be possible to measure the lifespan of prey DNA inside the nest as well as any potential DNA amplification biases between different types of prey. Sampling multiple cells from each nests provides more insight into how the colony diet varies over time, and it could display predatory

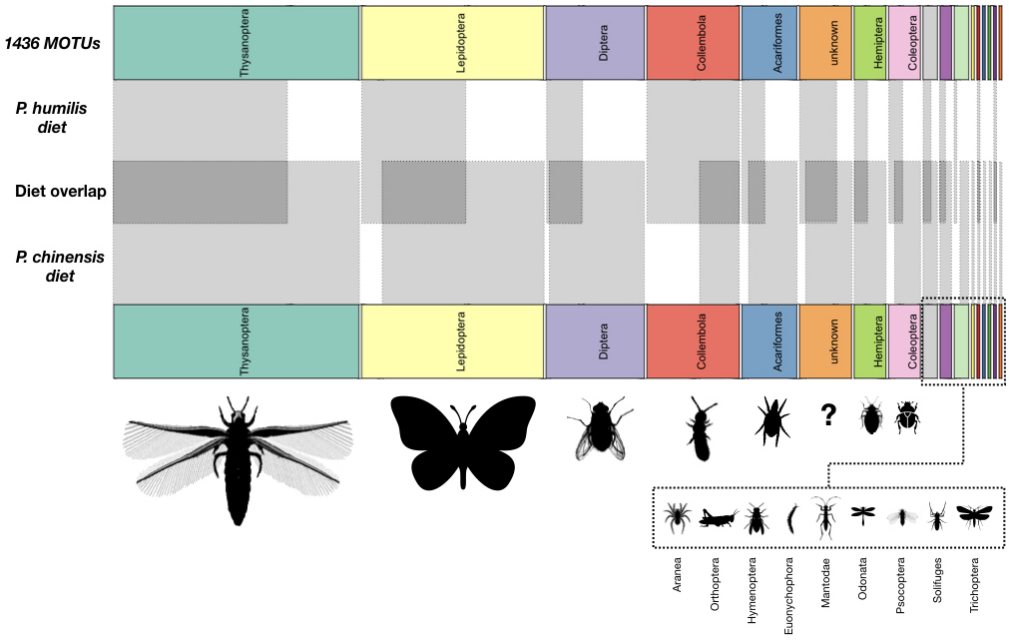

FIGURE 7 Prey diversity in the diet of *P. humilis* and *P. chinensis* and diet overlap.
The horizontal width of the rectangles correspond to the number of MOTU for each order. Light grey areas correspond to MOTUs detected in the diet of each wasp species. Darker grey areas correspond to MOTUs detected in the diet of both wasp species. Most silhouettes were obtained from Phylopic.org.

activity at a much higher resolution through time, potentially mirroring the phenology of the different prey species. However, in *Vespula* wasps, nests are often organised in multiple combs with little if any overlapping cell layers (Rome et al. 2015). In addition, cells remaining on the outer layer can be re-used multiple times by subsequent generations, which means a temporal analysis of the diet using requires precise knowledge of the nest construction.

4.4 | Concluding remarks

This study was geographically limited to the urban and peri-urban regions of Auckland. Given the relatively low number of nests analysed, our results should not be regarded as a comprehensive description of the diet of polistine wasps in New Zealand. Nevertheless, they provide a preliminary insight into the potentially controversial role that these species might play in New Zealand. Our results provide field evidence that polistine wasps feed on agricultural pests, supporting the hypothesis that some social wasp species do feed on and may suppress agricultural pests (Macintyre & Hellstrom 2015; Southon et al. 2019). Conversely, this study also clearly shows that native New Zealand Lepidoptera are consumed by polistine wasps, thereby illustrating the multifaceted ecological role of these generalist predators. The methodology developed in this project has the potential to greatly assist in the study of social wasp ecology. Compared to traditional DNA recovery methods, such as gut dissection (e.g. Ward & Ramón-Laca 2013), or the collection of prey pellets (e.g. Kasper et al. 2004), we have developed a method which can provide an overview of the diet of a colony through time based on a single sampling event. If nests are sampled after they are abandoned

at the end of the season, it might be possible to uncover a full record of the colony diet, assuming the degradation of prey DNA remains limited. We hope that this method will be applied to study the ecology of other nest building Hymenoptera, including native and invasive species such as Asian hornets (*Vespa velutina* Lepeletier). Regarding the latter, better knowledge of their diet is essential to measure the ecological impact of their invasion and to ensure the conservation of native invertebrate biodiversity and the ecosystem services they provide (Wycke et al. 2018; Cini et al. 2018).

5 | AUTHOR CONTRIBUTION

Designed the study: MCL, SB. Collected wasp nests: TS. Performed laboratory analyses: EJD, MCL, SB. Analysed the data, prepared the figures and wrote the first draft of the manuscript: MCL, SB. All authors contributed to the writing of the final manuscript.

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7 | SUPPLEMENTARY MATERIAL

Supplementary Table S1: Prey taxa identified at the species level in the diet of *Polistes humilis* and *P. chinensis*
Supplementary Figure S1: Data processing and number of reads and MOTUs retained or discarded at each step of the bioinformatics analysis
Supplementary Figure S2: Percentage identity for all prey MOTUs (left) and only Lepidoptera (right). Red dots are MOTUs identified at the species level (i.e. percentage identity $\geq$98%).

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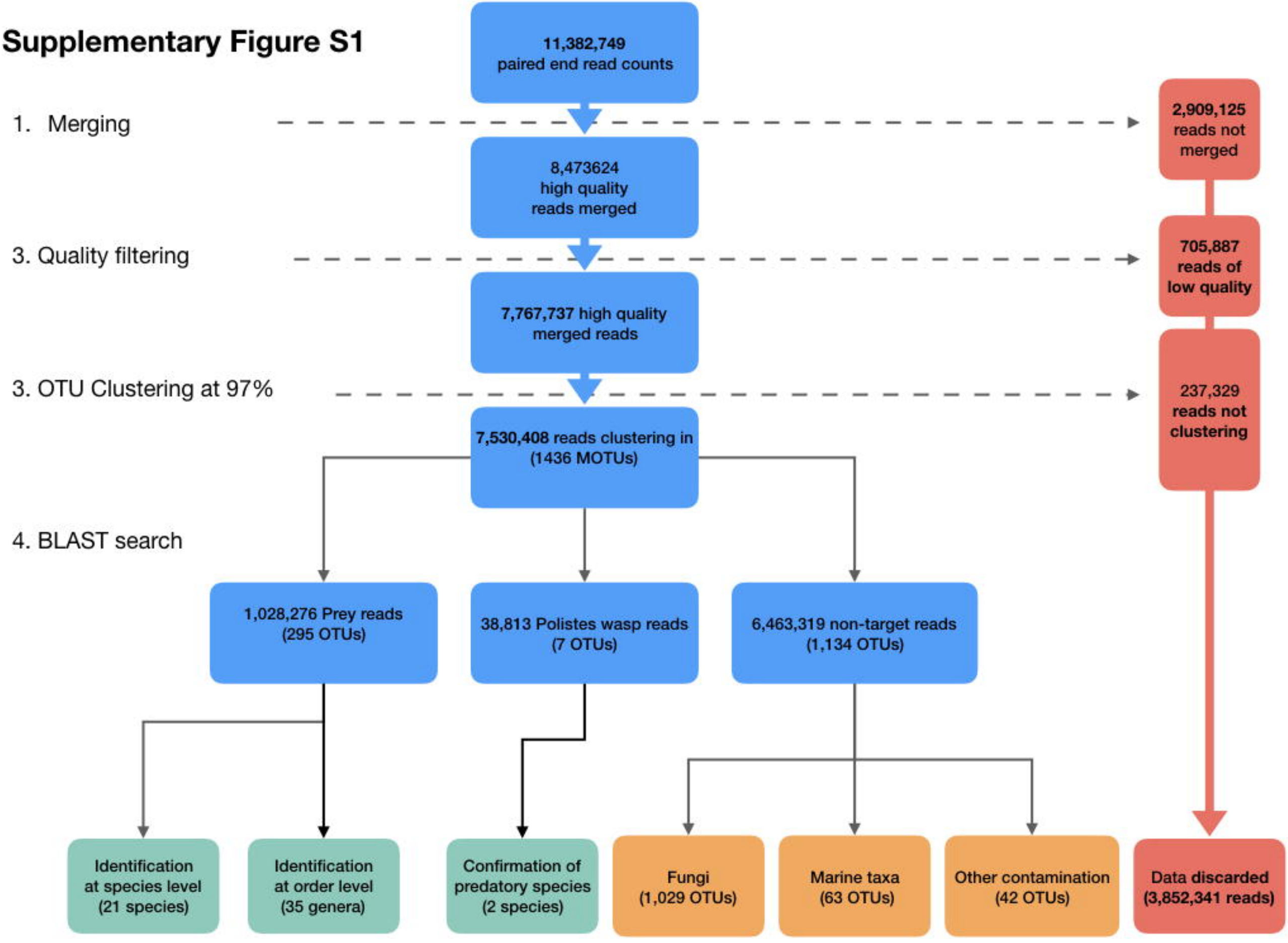
TABLE 1 Prey taxa identified at the species level in the diet of *Polistes humilis* and *P. chinensis*

Species	Family	Order	Native or Introduced	Agricultural pest	Number of samples tested positive	
					P. humilis (*n*=88)	*P. humilis* (*n*=8)
Anarsia dryinopa	Gelechiidae	Lepidoptera	Introduced		6	
Anatrachyntis badia	Cosmopterigidae	Lepidoptera	Introduced		2	6
Caliroa cerasi	Tenthredinidae	Hymenoptera	Introduced	Pest	1	
Chrysodeixis eriosoma	Noctuidae	Lepidoptera	Introduced	Pest		
Ctenoplusia limbirena	Noctuidae	Lepidoptera	Introduced			13
Ctenopseustis fraterna	Torticidae	Lepidoptera	Native		3	8
Ctenopseustis obliquana	Torticidae	Lepidoptera	Native		5	17
Declana floccosa	Geometridae	Lepidoptera	Native		3	3
Declana leptomera	Geometridae	Lepidoptera	Native			2
Ectopatria umbrosa	Noctuidae	Lepidoptera	Introduced			6
Ectopsocus meridionalis	Ectopsocidae	Psocoptera	Introduced			2
Eressa strepsimeris	Erebidae	Lepidoptera	Introduced			8
Graphania mutans	Noctuidae	Lepidoptera	Native			7
Isotenes miserana	Torticidae	Lepidoptera	Introduced			1
Leucania stenographa	Noctuidae	Lepidoptera	Introduced			11
Meteorus pulchricornis	Braconidae	Hymenoptera	Introduced			4
Mythimna separata	Noctuidae	Lepidoptera	Introduced	Pest		18
Oxysarcodexia varia	Sarcophagidae	Diptera	Introduced			1
Planotortrix notophaea	Torticidae	Lepidoptera	Native			2
Polistes humilis	Vespidae	Hymenoptera	Introduced			12
Scopula rubraria	Geometridae	Lepidoptera	Native			4
Tebenna micalis	Choreutidae	Lepidoptera	Native			2
Thysanoplusia orichalcea	Noctuidae	Lepidoptera	Introduced	Pest		7

Supplementary Figure S1



Supplementary Figure S2