

Nairobi Sheep Disease Virus: a historical and epidemiological perspective

Running title: Nairobi Sheep Disease Virus distribution

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Abstract

Nairobi Sheep Disease virus (NSDv) is a zoonotic and tick-borne disease that can cause over 90% mortality in small ruminants. NSDv has historically circulated in East Africa and has recently emerged in the Asian continent. Despite efforts to control the disease, some regions, mostly in warmer climates, persistently report disease outbreaks. Consequently, it is necessary to understand how environmental tolerances and factors that influence transmission may shed light on its possible emergence in other regions. In this study, we quantified the available literature of NSDv from which occurrence data was extracted. In total, 308 locations from Uganda, Kenya, Tanzania, Somalia, India, Sri Lanka and China were coupled with landscape conditions to reconstruct the ecological conditions for NSDv circulation and identify areas of potential disease transmission risk. Our results identified areas suitable for NSDv in Ethiopia, Malawi, Zimbabwe, Southeastern China, Taiwan, and Vietnam. Unsuitable areas included Democratic Republic of Congo, Zambia, and Southern Somalia. In summary, soil moisture, livestock density, and precipitation predispose certain areas to NSDv circulation. It is critical to investigate the epidemiology of NSDv in order to promote better allocation of resources to control its spread in regions that are more at risk. This will help reduce disease impact worldwide as climate change will favor emergence of such vector-borne diseases in areas with dense small ruminant populations.

INTRODUCTION

Background

Small ruminant populations have become one of the pillars of socio-economic wellbeing for developing countries due to their direct contribution to food security, however newly emerging diseases pose a constant threat (1–3). The demand for small ruminant products is growing globally, especially in China, the biggest producer and importer of ovine meat (4), and now as an alternative to pork consumption due to the circulation of African Swine Fever (5). Other countries such as the United States, Islamic Republic of Iran, Japan, and Qatar have also increased import demand for these products (4). Some countries such as Somaliland have economies that depend entirely on their livestock industries and employ over 70% of the population (6,7). Others such as Saudi Arabia import 5 million live ruminants a year, most of which are sheep and goats (8). However, this economic value is directly dependent on animal health, which has been compromised in regions with high incidence of tick-borne diseases (9–11). It has been proposed that the effects of ticks and tick-borne diseases on livestock pose the greatest barrier to economic development (12–14), and thus, a better understanding of current emerging disease distributions is important for human socio-economic development and animal welfare.

Some of the most pathogenic diseases of small ruminants include viruses from the *Bunyaviridae* family, which are spreading to novel areas and currently threaten small ruminant populations worldwide (15–18). There are seven serogroups within the *Nairovirus* genus, with the most impactful ones being i) Crimean-Congo hemorrhagic fever group which includes the human pathogen Crimean-Congo hemorrhagic fever virus (CCHFv), and ii) Nairobi Sheep Disease group that contains Nairobi Sheep Disease virus (NSDv) and Dugbe virus (19). Due to the similarities

46 between NSDv and CCHFv, studies on NSDv will be useful in furthering our understanding of this important human pathogen (20), and will serve
47 as a good model system to study other nairoviruses (21).

48 NSDv is characterized by hemorrhagic gastroenteritis, fever, abortion, and high mortality in small ruminants (22) and febrile illness,
49 nausea, vomiting, and headache in humans (23). Mortality rates in susceptible animals exceed 90%, causing significant economic losses for
50 production systems (22). This disease is listed as notifiable to the World Organization for Animal Health (OIE) (24), and it has the potential to
51 impose trade barriers and consequently have a substantial impact on small ruminant producers worldwide (25).

52 Although the occurrence of NSDv has been reported in numerous countries (16,26–32), there has been limited understanding of the
53 biogeographic factors shaping its distribution and the potential areas at risk for future epidemics. The available literature on NSDv ranges from
54 1910 to 2019, with host serology and virus isolation found in 14 of these studies (Table S1 in Supplementary Material). It has been found in
55 environmentally varying areas, leaving an open question regarding the requirements of this virus and its vectors to efficiently spread infection. For
56 that reason, the aim was to develop a systematic review and distribution model of NSDv.

57

58 ***NSDv vectors and affected species***

59 Ticks are known to transmit a greater variety of pathogenic microorganisms than any other arthropod vectors, and are among the most important
60 vectors affecting livestock and humans (33–37). NSDv spreads via feeding of competent infected ticks, and its geographic distribution is therefore
61 limited to the areas comprising suitable environment conditions for them (20). The main tick species related to the spread of NSDv are
62 *Rhipicephalus appendiculatus* in East Africa and *Haemaphysalis intermedia* in Asia (20). Lewis (38) demonstrated that *R. appendiculatus* can
63 potentially retain NSDv for long periods (138 to 871 days) depending on the life stage of the tick. NSDv has also been isolated from *Amblyomma*
64 *variegatum* (39), *Rhipicephalus hemaphysaloides* (40), and *Haemaphysalis longicornis* (16).

65 NSDv is also known to have the potential to infect humans. The first human infection was recorded in a young boy (41) and some
66 laboratory acquired infections occurred in following years (42,43). Human sera has been shown to contain antibodies against NSDv in India
67 (41,44), Uganda (29), Kenya (45), and Sri Lanka (32). Other isolates have also been obtained from different vector species such as *Culex vishnui*
68 mosquitoes, where the virus was unable to replicate without a tick host (46), *Haemaphysalis wellingtoni* ticks feeding on red spurfowl
69 (*Galloperdix spadicea*) (47), and from a pool of *Culicoides* 23 midges (48) and *Culicoides tororensis*, although this could be due to blood meal
70 residues in the midges (49). Low titers were found in various wild ruminants, but this was likely due to an antibody cross-reaction with other
71 viruses (28).

72

73 ***The emerging pathway of NSDv***

74 The first reports of NSDv came from Nairobi livestock markets in Kenya in 1910 as the result of an investigation carried out by veterinary
75 pathologist Eustace Montgomery (27). NSDv was first identified as the causative agent in a classic study that showed virus transmission
76 transovarially and transstadially in the Ixodid tick *R. appendiculatus* (27), findings that were later confirmed by Daubney and Hudson (50). In the
77 following years, it was identified in various other areas of Kenya (28,39,51), Uganda (22,29), Somalia (attributed to *Rhipicephalus pulchellus*)
78 (30), and Tanzania (52). NSDv is also known as Ganjam virus in Asia. Similarities between NSDv and Ganjam virus were highly debated due to
79 their occurrence on different continents and association with different vectors, but genetic and serologic analyses demonstrated that Ganjam virus
80 is an Asian variant of NSDv, instead of a different virus as previously described (23,52–54). NSDv was first isolated on the Asian continent in
81 1954 from *H. intermedia* in India (26) and Sri Lanka in 1996 (32). More recently, Gong et al. (16) were the first to discover NSDv present in *H.*
82 *longicornis* ticks in China. Small ruminants bred in endemic areas do not appear to be affected by the virus (20), which could be due to the
83 presence of maternal antibodies that provide sufficient protection until the animal's own immunity can be established (20).

84 In August 2017, *H. longicornis* was found in the United States for the first time in all three of its life stages, infecting an Icelandic sheep in
85 Hunterdon County (New Jersey) that had not been transported or had any contact with foreign animals (55). The following year, this tick was
86 found in seven other states along the Eastern US and Arkansas (56), and its presence was verified by reexamination of archived historical samples,
87 confirming that *H. longicornis* was present in West Virginia in 2010 and New Jersey in 2013 (56). Over the past 30 years, the US Department of
88 Agriculture has identified this tick at least six times from imported horses in quarantine (57). *H. longicornis* is unique for its parthenogenesis,
89 which allows a single female to produce a clonal population without mating with a male (58). *H. longicornis* is also found in Australia, which may
90 facilitate the appearance of NSDv in the country (59). Due to the potential global distribution of this disease, the Australian Veterinary
91 Association has stated that changes in climate could lead to an increase in disease incidence with devastating consequences (59). *H. longicornis* is

92 the only exotic tick established in New Zealand that has an economic impact on livestock (60) from which NSDv was most recently isolated from
93 in China (16). The range of NSDv and its vectors is likely spreading, and it will become even more important as we continue to push for breed
94 improvement and maximizing land use to manage increasing global demands for small ruminant products (21).

95

96 ***Variables affecting NSDv spread***

97 Climate change is known to impact the distribution of a great number of diseases (61–65), especially in tropical and subtropical regions (66–68).
98 Our understanding of the environmental preferences facilitating the survival of NSDv vectors and hosts will allow at-risk regions to better prepare
99 for potential incursions of this disease. These tick species mentioned above are known to span large geographical ranges, such as *R.*
100 *appendiculatus*, which has a territory that extends from the tropical regions of East Africa to the temperate regions of South Africa (69), as well as
101 India and Pakistan (70), or *H. longicornis*, which occupies a wide range of climates from equatorial New Guinea and the Pacific Islands, to snow
102 and cool summer conditions in northeast Primorsky Krai, Russia (71,72). For that reason, a deeper understanding of the environmental conditions
103 facilitating the survival of these ticks is key to predicting novel areas at risk for NSDv spread, which could potentially lessen the impact of this
104 disease on naive populations.

105

106 ***Economic impact of NSDv***

107 As global trade continues to increase, infectious diseases are posing a greater threat to our economies, food supply, and health (73,74). NSDv has
108 the potential to have devastating effects on naive sheep and goat populations (22), and therefore, efforts to increase awareness and surveillance
109 should become a priority, especially for countries with economies that rely heavily on small ruminant products. NSDv vectors such as *H.*
110 *longicornis* have already adapted to New Zealand and Australia (59,60), threatening the NSD-free status of these countries. Crossbreeding and the
111 introduction of new breeds of small ruminants have been the target of recent efforts to increase the efficiency of meat production and commerce in
112 some countries. This increases the chances of NSDv outbreaks in populations lacking immunity against the virus (75–77) since the main route for
113 NSDv spread occurs upon movement of susceptible animals into enzootic areas (51). Since small ruminants are vital to people's livelihoods in
114 low-income communities (1), efforts to preserve their health and better understand tick-borne diseases are essential.

115 Given the critical importance of small ruminant production to global food security and the potential for NSDv spread in new regions, the
116 current information gaps in the epidemiology of this disease have to be explored. This allows us to better predict future outbreaks and potential
117 regions at risk for this disease. This is the first systematic review and predictive model of NSDv, allowing us to assess the potential distribution of
118 the disease and predict regions that are most at risk.

119

120 **METHODS**

121 We divided this study into two main sections: i) a systematic review to identify all susceptible hosts, potential vectors, and the ecological
122 dependencies for virus spread and ii) an assessment of the geographic and environmental distribution of hosts and vectors using an ecological
123 niche modeling approach.

124

125 **Figure 1 HERE**

126

127 **i) Systematic review**

128 ***Literature selection***

129 We followed the steps outlined by O'Connor and Sargeant and others (78–80) (Table S2 in Supplementary Material) to scope the global
130 distribution of NSDv. Our search terms were designed to recover all information and reports under the terms ‘‘Nairobi Sheep Disease’’ and
131 ‘‘Ganjam virus’’. The study protocol followed the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines
132 (81,82) (Figure 2).

133 These terms were used to search all databases using the EBSCOhost platform, which yielded 339 scientific publications in 11 databases
134 (Table S2 in Supplementary Material). Exclusion criteria included duplicates (132), non-English papers (22), and irrelevant studies (5) which
135 brought the total number of eligible papers for systematic review to 180 (Figure 2). This database search was completed in November 2018.

136

137 **Figure 2 HERE**

138

139 **ii) Ecological niche model**

140 ***Data collection***

141 From the literature review, we extracted a total of 308 locations from 14 studies. The information retrieved included occurrence data classified as
142 follows: i) host occurrence- if the virus was identified from sheep, goat, or human serology, ii) tick occurrence- if the virus was identified directly
143 from tick vectors. Data points were georeferenced to the centroid of the smallest administrative division of each country from where the data was
144 collected. One exception was the use of the locations reported by Gong et al. (16), which were also verified before being considered for analysis.
145 Diagnostics based on post-mortem examinations were not considered since lesions are not specific enough to reliably distinguish NSDv from
146 heartwater, *Pasteurella pneumonia*, or babesiosis (30). Additional occurrences from the following studies in South Africa (83), Uganda (29),
147 Kenya (39), and India (41) were not included in the study due to non-specific locations from which we were unable to recover the centroid of the
148 smallest administrative boundary.

149

150 ***Model calibration area***

151 To define the model calibration area for the circulation of NSDv, we followed the framework proposed by Soberón and Peterson (84), which
152 restricts the ecological niche model to ecological features for the organism in question, the resolution of the environmental variables employed,
153 and the extent of the region where the organisms are able to disperse due to their biogeographic barriers (see M in the BAM framework in
154 Soberón and Peterson (84)). This region is defined as the area accessible to the species that has been sampled, so presence records can exist within
155 a suitable area (85). We measured the maximum distance between all NSDv locations and used this distance as the accessible area for the virus. A
156 buffer around the occurrences was applied to establish the model calibration area.

157

158 ***Environmental variables selection***

159 The environmental variables used to estimate the distribution of NSDv were selected based on the described requirements of the hosts and ticks,
160 including their survival in landscapes with suitable temperature and humidity and their amplification by the presence of livestock reservoir
161 species. We used six environmental variables to calibrate all models, and candidate models were selected based on the described requirements of
162 the virus, including its capacity to survive in the landscape based on the presence of susceptible host species. We included the variables shown in
163 Table 1, which will serve as an appropriate approximation of the required biogeographic conditions.

164 Previous studies have highlighted the importance of temperature for the main host of NSDv, *R. appendiculatus* (69,86,87). Although it is
165 not a critical limiting factor for ticks, it is still important, especially when it approaches the developmental threshold for that species (71).
166 Likewise, variables related to humidity are also important because it has been observed that prolonged drought and dry pastures cause high
167 mortality, especially among engorged *H. longicornis* (71,86) and *R. appendiculata* larvae and eggs (87), and *H. intermedia* and *R.*
168 *haemaphysaloides* numbers which seem to increase after rains (88). For that reason, we used precipitation, runoff, evapotranspiration, and soil
169 moisture as predictors of tick prevalence. Finally, livestock population was represented by the density of sheep and goats, which are most
170 clinically affected by NSDv (53,89).

171

172 **Table 1 HERE**

173

174 ***Ecological niche model***

175 Niche modeling presents a framework based on ecological theory by which to interpret, understand, and anticipate geographic distributions
176 of species and other biological phenomena, such as disease transmission (90). For this study, we considered an ecological niche as the set of
177 environmental conditions in a region necessary for a species to persist (85). Information about the presence of susceptible hosts for NSDv were
178 also added to this study through a layer that contained sheep and goat densities. We explored three scenarios based on different combinations of
179 occurrences: i) the occurrences of ticks that tested positive for NSDv; ii) the occurrences of sheep and goats that tested positive for NSDv (hosts);
180 and iii) the combination of both positive tick and host occurrences. For these analyses, we considered that the factors explaining the distribution of
181 NSDv and its potential hosts follow the Biotic-Abiotic-Mobility (BAM) framework (84). The analysis was set individually for each occurrence
182 data set for all models in which a hypothesis for each accessible area M was constructed (further details of the importance of this step can be
183 found in Barve et al. (85)). The study area for positive ticks and host cases was designed based on the union of buffer areas created around each

184 occurrence location. The buffer was built based on the average distance between the external points and the most central point (centroid). Thus,
185 for positive ticks this buffer was defined as 1,334 km, while for positive hosts, it was 1,555 km. The polygons drawn around each study area were
186 then used to intercept the environmental predictor layers (Table 1).

187 The total occurrences for each set were randomly subdivided into 70% of the data set for model calibration and 30% for model evaluation.
188 This data division allowed for both model calibration and internal testing. For each combination of data, we created 493 candidate models by
189 combining 17 regularization multipliers (0.1-1.0 by the interval of 0.1, from 1-10 at the interval of 1), with all 29 possible combinations of the five
190 MaxEnt feature classes (linear=L, quadratic=Q, product=P, threshold=T, and hinge=H). We evaluated and selected the candidate model
191 performances based on significance rates (5%), and model complexity penalizations (AICc). The best models for each dataset were selected
192 according to the following criteria: significant models with omission rates $\leq 5\%$, and from those selected models, we used delta AICc values of
193 ≤ 2 to determine the final candidates. Both the full model calibration and selection step were performed using the “kuenm” package in the R
194 environment and used Maxent as the modeling algorithm (91).

195 The final ENM models were performed in Maxent version 3.3.3 k (92). The specific configuration of all models included 10 bootstrap
196 replicates and random seed with logistic outputs. Finally, to identify extrapolation risk in the model transfer steps, we used the mobility-oriented
197 parity (MOP) index for each data set, which is an improved metric proposed by Owens et al. (93). The interpretation of the output models
198 followed Merow et al. (94) as a suitable index to account for the probability of disease risk. To further evaluate model predictions, continuous
199 outputs were converted into binary maps based on a threshold, removing 10% of the calibration occurrences (error = 10%) to reduce the
200 uncertainty of estimations and facilitate model interpretation (95). Finally, the models were transferred to a global scale applying the same
201 predictor layers used for the calibration step (Table 1).

202

203 **RESULTS**

204

205 **Descriptive Analysis**

206 A descriptive analysis of both the disease and vector distribution is provided in Figure 3, even though the total number of recovered disease
207 occurrences were not used as offset information. A total of 308 data points were extracted from 14 studies, from which 12 were isolated from ticks
208 and the rest were obtained from host serology. Data coming from hosts was distributed throughout East Africa, India, and Sri Lanka, while tick
209 data was found only in India, Sri Lanka, and China.

210

211 **Figure 3 HERE**

212

213

214 **Ecological niche models**

215

216 *NSDv host potential distribution*

217 The positive host occurrences of NSDv were used to calibrate 493 models along with the full set of environmental predictors. In total, 490
218 candidate models were significant from which 157 were significant and met the omission rate criteria. For the global minimum AICc values, 1
219 model had delta AICc values ≤ 2 , and met the full criteria used in the selection step (Figure 4). We emphasize that this selected model met both
220 criteria but did not have lower AICc, which is often utilized as single decision criteria in model selection (96–98) (see the blue triangle which
221 highlights the selected final mode- Figure 4). In order to identify areas of higher risk for NSDv occurrences, the selected model identified the
222 geographic risk areas for small ruminants. This model showed countries presenting low suitability for NSDv spread including Democratic
223 Republic of Congo, Zambia, and southern Somalia. There were clusters of unsuitability throughout Saudi Arabia, eastern Yemen, Oman, and
224 western China of the study area. High-risk regions were found throughout Eritrea, Ethiopia, Kenya, Uganda, Rwanda, Burundi, Tanzania, Malawi,
225 Zimbabwe, and the southern tip of India (Figure 7). The livestock density variable was the most influential (48.7%) in describing the risk area,
226 followed by runoff (21.8%), evapotranspiration (11.6%), precipitation (10.2%), minimum temperature (4.7%), and soil moisture (3%).

227

228 **Figure 4 HERE**

229

230 *NSDv tick potential distribution*

231 The occurrences of NSDv in vectors was modeled applying the selected parameterizations found in Table 1. Using the locations for tick
232 occurrences, 367 candidate models out of 493 were statistically significant if compared with the assumed null expectations. All 367 of these
233 statistically significant models also met the omission rate criteria. For the global minimum AICc values for ticks, 2 models had delta AICc values
234 ≤ 2 and 1 model met the criteria (Figure 5). The model selected had lower AICc, was significant, and within the omission error cutoff (see the
235 blue triangle which highlights the selected final mode- Figure 5). This model was used to identify areas with a potential circulation of ticks within
236 the calibration study area. It showed unsuitable areas clustered in Sri Lanka, central Vietnam, eastern Taiwan, the western shores of Japan, and
237 southeast China. High-risk regions were found throughout the southern half of India, Sri Lanka, Bangladesh, Myanmar, Laos, Thailand,
238 Cambodia, Taiwan, China's Hainan Island and the southern tip of Guangdong province, and the northern and southern portions of Vietnam
239 (Figure 7). The minimum temperature variable was the most influential (53.5%) in describing the risk areas, followed by livestock (38.4%), soil
240 moisture (4.6%), and runoff (3.5%). Precipitation and evapotranspiration were not influential (0%).

241

242 **Figure 5 HERE**

243

244 *NSDv host and tick potential distribution*

245 The last model exploration we did included the combination of data associated with infected hosts and the occurrences of potential vector ticks.
246 From 493 candidate models, 209 were significant, from which 109 also met the omission rate criteria. For the global minimum AICc values, 1
247 model had delta AICc values ≤ 2 . Only 1 model met the full criteria used in the model selection step (Figure 6) (see the blue triangle which
248 highlights the selected final model-Figure 6). Suitable areas in the study area included Ethiopia, Uganda, Kenya, Tanzania, the southern tip of
249 India, the southeastern coast of China, and Taiwan (Figure 7). The livestock density variable was the most influential (38.1%) in describing the
250 risk area, followed by evapotranspiration (27.2%), runoff (14.5%), minimum temperature (9.2%), soil moisture (7.8%), and precipitation (3.3%).

251

252 **Figure 6 HERE**

253

254 **Figure 7 HERE**

255

256 *NSDv environmental preferences*

257 Overall, our ecological niche models coincided well with the reported geographical locations of the host and tick occurrences identified during the
258 systematic review. These regions are the most environmentally suited for NSDv, and they indicate areas where the disease would most easily
259 become established and continue spreading to naive small ruminant populations. For the host-based model, the most suitable areas occurred with
260 the highest values of evaporation (46.1 mm; IQR 83.0 mm), precipitation (40.3 mm; IQR 63.0 mm), and runoff (2.8 mm; IQR 3.0 mm). In the
261 case of the tick-based model, the most suitable areas presented the highest values of soil moisture (62.2 mm; IQR 588.0 mm) and livestock density
262 (11812.0 Ind/km²; IQR 8953.8 Ind/km²). Finally, the model combining hosts and ticks presented the highest values of minimum temperature
263 (12.86 °C; IQR 5.50 °C) (see Table 2 for detailed information).

264

265 **Table 2 HERE**

266

267 **DISCUSSION**

268

269 We identified novel countries at risk for infection with NSDv, including Ethiopia, Malawi, Zimbabwe, Southeastern China, Taiwan, and Vietnam.
270 These findings suggest that NSDv may have a wider range than was previously thought, and therefore, these predictions can help direct
271 surveillance efforts to those regions where NSDv could efficiently spread if introduced in the future.

272 Livestock diseases can result in devastating trade bans with wide-ranging consequences (7). Specifically, NSDv has the potential to easily
273 spread to naive populations and cause significant economic losses (22). Therefore, it is imperative for at-risk countries to be aware of these risks
274 and take necessary precautions to protect their animals, peoples' livelihoods, local economies, and global trade partners.

275 To the author's knowledge, this is the first systematic review of NSDv that has gathered all available geographic locations where this
276 disease has occurred. Systematic reviews are increasingly being used in animal agriculture and veterinary medicine since they offer a replicable
277 evidence-based method to identify, evaluate, and summarize primary research (78). Based on the reports obtained, we determined the countries
278 which are currently at risk for the introduction of this disease. We performed extensive model fitting from which we selected the best
279 performance-based model, focusing on significance rather than AIC values as our main criteria for model selection. Using significance rather than
280 AIC allows us to choose the single best model which outperforms the rest and will give us the most realistic prediction of occurrence. We found
281 varying results in our models, with notable differences in the environmental preferences needed for the optimal spread of the virus when host and
282 tick-based models were compared. Because our data points included serology from hosts and virus isolation from ticks, these models should be
283 interpreted together rather than separately. Multiple environmental variables play a role in the transmission of NSDv, and they should all be
284 considered holistically. We support that this combination of data would more accurately predict potential regions of outbreak since the main
285 transmission of NSDv will likely occur when both hosts and ticks coincide together in the same environment.

286 ENMs have been widely applied in epidemiology to assess species introduction into novel areas and disease emergence (96,99–101), or
287 the effect of climate on disease distribution (102–105). However, previous ENM methods rely solely on the lowest AIC to identify the best fit
288 model, which alone may not lead us to the best option for that set of occurrences (106). Significance should be used as the first criteria when
289 filtering through candidate models, followed by performance and simplicity (91). This approach reduces the time spent on transferring these
290 models, and uses the mobility-oriented parity (MOP) index to prevent outcome over-interpretation and identify extrapolation risks, making it a
291 critical tool when transferring ENMs to current and future scenarios, and resulting in a more accurate prediction of species' ecological niche as the
292 environment changes. These calibrations have become more popular and are being applied throughout epidemiology (91,107), but our model of
293 NSDv will be the first time that this calibrated approach has been used to model the potential distribution of a virus.

294 Ticks are described as the main route of NSDv transmission and therefore, they are the most likely route of infection for naive populations
295 given that the virus can survive for such extended periods of time inside its vector (38). International trade is a known pathway for introduction of
296 exotic ticks into new areas, which have the potential to harbor pathogens with significant consequences for human health and agriculture
297 (108,109), highlighting the importance of increased surveillance measures and regulatory framework over animal trade-related interactions. In the
298 tick-based model, NSDv preferred areas shown to have elevated soil moisture and a denser population of livestock. Although the density of sheep
299 and goats was expected to have a direct influence on the number of infected ticks found in some locations, soil moisture may have facilitated the
300 ability of ticks to survive in the field and continue transmitting the disease to other individuals. In contrast, the model based on host occurrences
301 had elevated mean evaporation, precipitation, and runoff. This may correlate with areas of intense feed and water resources, thus concentrating
302 more animals in one area, which will facilitate the spread of the disease between and within the population.

303 Even though these two models can independently explain and predict areas where NSDv is most likely to occur, the combination and
304 overlap of the most suitable environments for hosts and ticks will represent the areas where hosts and their vectors are more likely to proliferate
305 and coincide, greatly increasing the possibility of disease spread. Thus, we recommend the comparison and combination of these models for future
306 epidemiological assessments in order to get the most accurate prediction of at-risk areas.

307 We encountered some limitations during the development of this study. First, a number of non-English papers (n=22) were excluded in our
308 systematic review which may have included additional data for our models. We only included data points that had a specific location and excluded
309 those that were labeled as having been collected in general areas or districts, so it is possible that additional occurrences were not recorded.
310 Additionally, we used a centroid to approximate the location of occurrences, which entails the possibility of some data points being farther away
311 from the center. Multiple species of Ixodid ticks were modeled together due to their similar environmental requirements, which may
312 overgeneralize the distribution of these ticks.

313 Vector-borne diseases are expected to be the most climate-sensitive subset of infectious diseases (110,111), and using environmental
314 variables is critical to predicting their distribution. Recent outbreaks of devastating ruminant diseases such as Bluetongue and epizootic
315 hemorrhagic disease, serve as a reminder of the consequences that climate change has on vector-borne diseases (112), and illustrates the
316 importance of using the best modeling techniques we have at our disposal in order to more accurately predict the spread of these vector-borne
317 diseases.

318

319 **Conclusions**

320 The data compiled here will be useful for additional spatial and environmental modeling of NSDv and its vectors, informing governments
321 and policy makers about their prevalence and spread in order to direct surveillance efforts. Further studies would also help the development of
322 case studies for NSDv and other infectious diseases affecting small ruminants, as well as to understand and predict where to best implement
323 surveillance strategies, increasing response time and decreasing economic losses. Further study of the tick species shown to carry and transmit
324 NSDv will be imperative as the distribution of these species changes with environmental factors and increasing international trade. As the ticks'
325 distribution changes, so too should surveillance strategies. Additionally, many countries that could potentially be at risk for NSDv have never had
326 a cross-sectional study done on their livestock or tick species of interest, which would hinder their ability to quickly and effectively respond to a
327 disease outbreak. As an OIE reportable disease, NSDv can cause significant trade implications for countries and could be underreported as a
328 result. Rising global demand for these products and recognition of the socioeconomic importance of small ruminants merits further discussion of
329 the diseases that impact them and pose a threat to their populations in other parts of the world and the communities that depend on them.

330

331 **Author contributions**

332 SK, MJ, GM contributed to the design of the project. SK collected and curated the data used in both the systematic review and modeling exercise.
333 MJ and GM performed the modeling section. SK wrote the manuscript. All authors reviewed and edited the manuscript.

334

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338

339 **Conflict of interest**

340 The authors declare that there are no conflicts of interest.

341

342 **References**

- 343 1. Kosgey IS. Breeding objectives and breeding strategies for small ruminants in the tropics. (2004) Available at: <http://edepot.wur.nl/121527>
344 [Accessed March 15, 2019]
- 345 2. Umunna MO, Olafadehan OA, Arowona A. Small ruminant production and management systems in urban area of southern guinea Savanna
346 of Nigeria. *Asian J Agric Food Sci* (2014) **2**:107–114. Available at: <https://ajouronline.com/index.php/AJAFS/article/view/1041> [Accessed
347 March 15, 2019]
- 348 3. Adams F. Socio-Economic Analysis of Small Ruminant Livestock Production in Northern Ghana. *Socio-Economic Anal Small Rumin*
349 *Livest Prod North Ghana* (2015)1–200. Available at: <http://ir.knust.edu.gh/xmlui/bitstream/handle/123456789/9264/Faizal>
350 *Adams.pdf?sequence=1* [Accessed March 15, 2019]
- 351 4. FAO. Meat market review: 2019 Outlook. (2019). Available at: <http://www.fao.org/3/ca3880en/ca3880en.pdf>
- 352 5. Dixon LK, Stahl K, Jori F, Vial L, Pfeiffer DU. African Swine Fever Epidemiology and Control. *Annu Rev Anim Biosci* (2020) **8**:
353 doi:10.1146/annurev-animal-021419-083741
- 354 6. Godiah LM, Baker D, Elmi II, Costagli R, Gulaid I, Wanyoike F. Enhancing the provision of livestock marketing information in
355 Somaliland. Nairobi (2014). Available at:
356 https://cgspace.cgiar.org/bitstream/handle/10568/56737/ilriResearchReport_35.pdf?sequence=8&isAllowed=y [Accessed March 17, 2019]
- 357 7. Muhumed MM, Yonis AM. The Future of Somaliland Livestock Exports: Examining the Sustainability of Livestock Trade. *Int J Manag*
358 *Account Econ* (2018) **5**: Available at:
359 https://www.researchgate.net/publication/328319505_The_Future_of_Somaliland_Livestock_Exports_Examining_the_Sustainability_of_Livestock_Trade_International_Journal_of_Management_Accounting_and_Economics_Vol_5_No_8_2018_With_Abdiqadir_M_Yonis
360 [Accessed March 17, 2019]
- 361 8. Abd El-Rahim IHA, Asghar AH, Mohamed AM, Fat'hi SM. The impact of importation of live ruminants on the epizootiology of foot and
362 mouth disease in Saudi Arabia. *Rev Sci Tech* (2016) **35**:769–778. doi:10.20506/rst.35.3.2567
- 364 9. Kosgey IS, Rowlands GJ, van Arendonk JAM, Baker RL. Small ruminant production in smallholder and pastoral/extensive farming
365 systems in Kenya. *Small Rumin Res* (2008) **77**:11–24. doi:10.1016/J.SMALLRUMRES.2008.02.005

- 366 10. Derso S, Demessie Y. Tick Borne Hemoparasitic Diseases of Ruminants: A Review. *Adv Biol Res (Rennes)* (2015) **9**:210–224.
367 doi:10.5829/idosi.abr.2015.9.4.9516
- 368 11. Mureithi DK, Mukiria EW. An Assessment of Tick-Borne Diseases Constraints to Livestock Production in a Smallholder Livestock
369 Production System: a Case of Njiru District, Kenya. *Int J Res Agric For* (2015) **2**:43–49. Available at: <http://www.ijraf.org/pdf/v2-i10/5.pdf>
370 [Accessed March 15, 2019]
- 371 12. Young AS, Grocock CM, Kariuki DP. Integrated control of ticks and tick-borne diseases of cattle in Africa. *Parasitology* (1988) **96**:403–
372 432. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/3287285> [Accessed March 15, 2019]
- 373 13. Perry BD, Young AS. The past and future roles of epidemiology and economics in the control of tick-borne diseases of livestock in Africa:
374 the case of theileriosis. *Prev Vet Med* (1995) **25**:107–120. doi:10.1016/0167-5877(95)00546-3
- 375 14. Minjauw B, McLeod A. *Tick-borne diseases and poverty: the impact of ticks and tick-borne diseases on the livelihoods of small-scale and*
376 *marginal livestock owners in India and eastern and southern Africa*. Edinburgh: DFID Animal Health Programme, Centre for Tropical
377 Veterinary Medicine (2003). Available at: <https://www.cabdirect.org/cabdirect/abstract/20063155090> [Accessed March 17, 2019]
- 378 15. Linthicum KJ, Britch SC, Anyamba A. Rift Valley Fever: An Emerging Mosquito-Borne Disease. *Annu Rev Entomol* (2016) **61**:395–415.
379 doi:10.1146/annurev-ento-010715-023819
- 380 16. Gong S, He B, Wang Z, Shang L, Wei F, Liu Q, Tu C. Nairobi Sheep Disease Virus RNA in Ixodid Ticks, China, 2013. *Emerg Infect Dis*
381 (2015) **21**:718–720. doi:10.3201/eid2104.141602
- 382 17. Armstrong PM, Andreadis TG, Anderson JF. Emergence of a new lineage of Cache Valley virus (Bunyaviridae: Orthobunyavirus) in the
383 northeastern United States. *Am J Trop Med Hyg* (2015) **93**:11–17. doi:10.4269/ajtmh.15-0132
- 384 18. Elliott RM. Bunyaviruses and climate change. *Clin Microbiol Infect* (2009) **15**:510–517. doi:10.1111/j.1469-0691.2009.02849.x
- 385 19. Lasecka L, Bin-Tarif A, Bridgen A, Juleff N, Waters RA, Baron MD. Antibodies to the Core Proteins of Nairobi Sheep Disease
386 Virus/Ganjam Virus Reveal Details of the Distribution of the Proteins in Infected Cells and Tissues. *PLoS One* (2015) **10**:e0124966.
387 doi:10.1371/journal.pone.0124966
- 388 20. Baron MD, Holzer B. Nairobi sheep disease virus/Ganjam virus. *Rev Sci Tech* (2015) **34**:411–417. Available at:
389 <https://pdfs.semanticscholar.org/39a5/0d6747f8e2dcb3c85e593f91c3ca0d7036c.pdf> [Accessed March 15, 2019]
- 390 21. Bin Tarif A, Lasecka L, Holzer B, Baron MD. Ganjam virus/Nairobi sheep disease virus induces a pro-inflammatory response in infected
391 sheep. *Vet Res* (2012) **43**:71. doi:10.1186/1297-9716-43-71
- 392 22. Terpstra C. Nairobi sheep disease. Studies on virus properties, epizootiology and vaccination in Uganda. *Nairobi sheep Dis Stud virus Prop*
393 *Epizoot Vaccin Uganda* (1969) Available at: <https://www.cabdirect.org/cabdirect/abstract/19702201697> [Accessed March 17, 2019]
- 394 23. Yadav PD, Vincent MJ, Khristova M, Kale C, Nichol ST, Mishra AC, Mourya DT. Genomic analysis reveals Nairobi sheep disease virus to
395 be highly diverse and present in both Africa, and in India in the form of the Ganjam virus variant. *Infect Genet Evol* (2011) **11**:1111–1120.
396 doi:10.1016/j.meegid.2011.04.001
- 397 24. OIE. OIE-Listed diseases, infections and infestations in force in 2020. (2020) Available at: [https://www.oie.int/animal-health-in-the-](https://www.oie.int/animal-health-in-the-world/oie-listed-diseases-2020/)
398 [world/oie-listed-diseases-2020/](https://www.oie.int/animal-health-in-the-world/oie-listed-diseases-2020/) [Accessed December 4, 2020]
- 399 25. Ibrahim H. *Small ruminant production techniques*. Nairobi: ILRI (International Livestock Research Institute) (1998). Available at:
400 <https://cgspace.cgiar.org/handle/10568/509> [Accessed March 17, 2019]
- 401 26. Dandawate CN, Shah K V. Ganjam virus: a new arbovirus isolated from ticks *Haemaphysalis intermedia* Warburton and Nuttall, 1909 in
402 Orissa, India. *Indian J Med Res* (1969) **57**:799–804. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/5820427> [Accessed March 17,
403 2019]
- 404 27. MONTGOMERY E. On a Tick-borne Gastroenteritis of Sheep and Goats occurring in British East Africa. *J Comp Pathol Ther* (1917)
405 **30**:28–57. Available at: <https://www.cabdirect.org/cabdirect/abstract/19171000175> [Accessed March 17, 2019]
- 406 28. Davies FG. A survey of Nairobi sheep disease antibody in sheep and goats, wild ruminants and rodents within Kenya. *J Hyg (Lond)* (1978)
407 **81**:251–258. Available at: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2129766/pdf/jhyg00050-0085.pdf> [Accessed March 17, 2019]
- 408 29. Weinbren MP, Gourlay RN, Lumsden WHR, Weinbren BM. An Epizootic of Nairobi Sheep Disease in Uganda. *J Comp Pathol Ther*
409 (1958) **68**:174–187. doi:10.1016/S0368-1742(58)80018-1
- 410 30. Edelsten RM. The distribution and prevalence of Nairobi Sheep disease and other tick-borne infections of sheep and goats in northern
411 Somalia. *Trop Anim Health Prod* (1975) **7**:29–34. doi:10.1007/BF02383239

- 412 31. Jessett DM. Serological evidence of Nairobi sheep disease in Tanzania. *Trop Anim Health Prod* (1978) **10**:99–100. Available at:
413 <http://www.ncbi.nlm.nih.gov/pubmed/664022> [Accessed March 17, 2019]
- 414 32. Perera LP, Peiris JSM, Weilgama DJ, Calisher CH, Shope RE. Nairobi sheep disease virus isolated from *Haemaphysalis intermedia* ticks
415 collected in Sri Lanka. *Ann Trop Med Parasitol* (1996) **90**:91–93. doi:10.1080/00034983.1996.11813031
- 416 33. Jongejans F, Uilenberg G. The global importance of ticks. *Parasitology* (2004) **129 Suppl**:S3–14. Available at:
417 <http://www.ncbi.nlm.nih.gov/pubmed/15938502> [Accessed March 17, 2019]
- 418 34. de la Fuente J, Estrada-Pena A, Venzal JM, Kocan KM, Sonenshine DE. Overview: Ticks as vectors of pathogens that cause disease in
419 humans and animals. *Front Biosci* (2008) **13**:6938–6946. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/18508706> [Accessed March
420 17, 2019]
- 421 35. Parola P, Raoult D. Ticks and Tickborne Bacterial Diseases in Humans: An Emerging Infectious Threat. *Clin Infect Dis* (2001) **32**:897–
422 928. doi:10.1086/319347
- 423 36. Laurenson MK, Norman RA, Gilbert L, Reid HW, Hudson PJ. Identifying disease reservoirs in complex systems: mountain hares as
424 reservoirs of ticks and louping-ill virus, pathogens of red grouse. *J Anim Ecol* (2003) **72**:177–185. doi:10.1046/j.1365-2656.2003.00688.x
- 425 37. Keesing F, Belden LK, Daszak P, Dobson A, Harvell CD, Holt RD, Hudson P, Jolles A, Jones KE, Mitchell CE, et al. Impacts of
426 biodiversity on the emergence and transmission of infectious diseases. *Nature* (2010) **468**:647–652. doi:10.1038/nature09575
- 427 38. Lewis EA. Nairobi sheep disease: the survival of the virus in the tick *Rhipicephalus appendiculatus*. *Parasitology* (1946) **37**:55–59.
428 doi:10.1017/S0031182000013159
- 429 39. Johnson BK, Chanas AC, Squires EJ, Shockley P, Simpson DIH, Parsons J, Smith DH, Casals J. Arbovirus isolations from ixodid ticks
430 infesting livestock, Kano Plain, Kenya. *Trans R Soc Trop Med Hyg* (1980) **74**:732–737. doi:10.1016/0035-9203(80)90188-1
- 431 40. Joshi M V., Geevarghese G, Joshi GD, Ghodke YS, Mourya DT, Mishra AC. Isolation of Ganjam Virus from Ticks Collected off Domestic
432 Animals Around Pune, Maharashtra, India. *J Med Entomol* (2005) **42**:204–206. doi:10.1093/jmedent/42.2.204
- 433 41. Dandawate CN, Work TH, Webb JK, Shah K V. Isolation of Ganjam virus from a human case of febrile illness: a report of a laboratory
434 infection and serological survey of human sera from three different states of India. *Indian J Med Res* (1969) **57**:975–982. Available at:
435 <http://www.ncbi.nlm.nih.gov/pubmed/5823182> [Accessed March 17, 2019]
- 436 42. Rao C V, Dandawate CN, Rodrigues JJ, Rao GL, Mandke VB, Ghalsasi GR, Pinto BD. Laboratory infections with Ganjam virus. *Indian J*
437 *Med Res* (1981) **74**:319–324. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/6797936> [Accessed March 17, 2019]
- 438 43. Banerjee K, Gupta NP, Goverdhan MK. Viral infections in laboratory personnel. *Indian J Med Res* (1979) **69**:363–373. Available at:
439 <http://www.ncbi.nlm.nih.gov/pubmed/447375> [Accessed March 17, 2019]
- 440 44. Joshi M V, Elankumaran S, Joshi GD. A post-epizootic survey of Rift Valley Fever-like illness among sheep at Veerapuram, Chennai,
441 Tamil Nadu. *Indian J Virol* (1998) **14**:155–157. Available at: <http://krishikosh.egranth.ac.in/bitstream/1/2057191/1/TNV-979-51.pdf>
442 [Accessed March 17, 2019]
- 443 45. Morrill JC, Johnson BK, Hyams C, Okoth F, Tukei PM, Mugambi M, Woody J. Serological evidence of arboviral infections among
444 humans of coastal Kenya. *J Trop Med Hyg* (1991) **94**:166–168. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/2051522> [Accessed
445 March 17, 2019]
- 446 46. Dandawate CN, Singh KR, Dhanda V. Experimental infection of certain mosquitoes & ticks with Ganjam virus. *Indian J Exp Biol* (1981)
447 **19**:1185–1186. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/6120893> [Accessed March 17, 2019]
- 448 47. Rajagopalan PK, Sreenivasan MA, Paul SD. Isolation of Ganjam virus from the bird tick *Haemaphysalis wellingtoni* Nuttall and Warburton
449 1907. *Indian J Med Res* (1970) **58**:1195–1196. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/5534070> [Accessed March 17, 2019]
- 450 48. Walker AR, Davies FG. A preliminary survey of the epidemiology of bluetongue in Kenya. *J Hyg (Lond)* (1971) **69**:47–60.
451 doi:10.1017/S0022172400021239
- 452 49. Davies FG, Walker AR, Ochieng P, Shaw T. Arboviruses isolated from *Culicoides* midges in Kenya. *J Comp Pathol* (1979) **89**:587–595.
453 doi:10.1016/0021-9975(79)90049-5
- 454 50. Daubney R, Hudson JR. Nairobi Sheep Disease. *Parasitology* (1931) **23**:507–524. doi:10.1017/S0031182000013895
- 455 51. Davies FG. Nairobi sheep disease in Kenya. The isolation of virus from sheep and goats, ticks and possible maintenance hosts. *J Hyg*
456 *(Lond)* (1978) **81**:259–265. Available at: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2129769/pdf/jhyg00050-0093.pdf> [Accessed
457 March 17, 2019]

- 458 52. Davies FG, Casals J, Jesset DM, Ochieng P. The serological relationships of Nairobi sheep disease virus. *J Comp Pathol* (1978) **88**:519–
459 523. doi:10.1016/0021-9975(78)90005-1
- 460 53. Marczinke BI, Nichol ST. Nairobi Sheep Disease Virus, an Important Tick-Borne Pathogen of Sheep and Goats in Africa, Is Also Present
461 in Asia. *Virology* (2002) **303**:146–151. doi:10.1006/VIRO.2002.1514
- 462 54. Kuhn J, Wiley M, Rodriguez S, Bào Y, Prieto K, Travassos da Rosa A, Guzman H, Savji N, Ladner J, Tesh R, et al. Genomic
463 Characterization of the Genus Nairovirus (Family Bunyaviridae). *Viruses* (2016) **8**:164. doi:10.3390/v8060164
- 464 55. Rainey T, Occi JL, Robbins RG, Egizi A. Discovery of *Haemaphysalis longicornis* (Ixodida: Ixodidae) Parasitizing a Sheep in New Jersey,
465 United States. *J Med Entomol* (2018) **55**:757–759. doi:10.1093/jme/tjy006
- 466 56. Beard C Ben, Occi J, Bonilla DL, Egizi AM, Fonseca DM, Mertins JW, Backenson BP, Bajwa WI, Barbarin AM, Bertone MA, et al.
467 Multistate Infestation with the Exotic Disease–Vector Tick *Haemaphysalis longicornis* — United States, August 2017–September 2018.
468 (2018). doi:10.15585/mmwr.mm6747a3
- 469 57. Haddow AD. The Consequences of Medically Important Invasive Arthropods: The Longhorned Tick, *Haemaphysalis longicornis*. *Clin*
470 *Infect Dis* (2019) **68**:530–531. doi:10.1093/cid/ciy695
- 471 58. Mihara R, Umemiya-Shirafuji R, Abe Y, Matsuo T, Horiuchi N, Kawano S, Fujisaki K, Suzuki H. The development of oocytes in the ovary
472 of a parthenogenetic tick, *Haemaphysalis longicornis*. *Parasitol Int* (2018) **67**:465–471. doi:10.1016/j.parint.2018.04.006
- 473 59. Thomson S. Emergency Animal Diseases Bulletin No.119: Nairobi sheep disease. *Aust Vet J* (2018) **96**:N10. Available at:
474 <https://www.ava.com.au/17203> [Accessed March 17, 2019]
- 475 60. Heath A. Biology, ecology and distribution of the tick, *Haemaphysalis longicornis* Neumann (Acari: Ixodidae) in New Zealand. *N Z Vet J*
476 (2016) **64**:10–20. doi:10.1080/00480169.2015.1035769
- 477 61. Dantas-Torres F. Climate change, biodiversity, ticks and tick-borne diseases: The butterfly effect. *Int J Parasitol Parasites Wildl* (2015)
478 **4**:452–461. doi:10.1016/j.ijppaw.2015.07.001
- 479 62. Purse B V., Mellor PS, Rogers DJ, Samuel AR, Mertens PPC, Baylis M. Climate change and the recent emergence of bluetongue in
480 Europe. *Nat Rev Microbiol* (2005) **3**:171–181. doi:10.1038/nrmicro1090
- 481 63. Ready PD. Leishmaniasis emergence and climate change. *Rev Sci Tech* (2008) **27**:399–412. Available at:
482 <http://www.ncbi.nlm.nih.gov/pubmed/18819668> [Accessed March 17, 2019]
- 483 64. Burge CA, Mark Eakin C, Friedman CS, Froelich B, Hershberger PK, Hofmann EE, Petes LE, Prager KC, Weil E, Willis BL, et al. Climate
484 Change Influences on Marine Infectious Diseases: Implications for Management and Society. *Ann Rev Mar Sci* (2014) **6**:249–277.
485 doi:10.1146/annurev-marine-010213-135029
- 486 65. Wu X, Lu Y, Zhou S, Chen L, Xu B. Impact of climate change on human infectious diseases: Empirical evidence and human adaptation.
487 *Environ Int* (2016) **86**:14–23. doi:10.1016/J.ENVINT.2015.09.007
- 488 66. Lyon B, DeWitt DG. A recent and abrupt decline in the East African long rains. *Geophys Res Lett* (2012) **39**: doi:10.1029/2011GL050337
- 489 67. McSweeney C, New M, Lizcano G, Lu X, McSweeney C, New M, Lizcano G, Lu X. The UNDP Climate Change Country Profiles:
490 Improving the accessibility of observed and projected climate information for studies of climate change in developing countries. *Bull Am*
491 *Meteorol Soc* (2010) **91**:157–166. doi:10.1175/2009BAMS2826.1
- 492 68. Lott FC, Christidis N, Stott PA. Can the 2011 East African drought be attributed to human-induced climate change? *Geophys Res Lett*
493 (2013) **40**:1177–1181. doi:10.1002/grl.50235
- 494 69. Randolph SE. Abiotic and biotic determinants of the seasonal dynamics of the tick *Rhipicephalus appendiculatus* in South Africa. *Med Vet*
495 *Entomol* (1997) **11**:25–37. doi:10.1111/j.1365-2915.1997.tb00286.x
- 496 70. Ghosh S, Bansal GC, Gupta SC, Ray D, Khan MQ, Irshad H, Shahiduzzaman M, Seitzer U, Ahmed JS. Status of tick distribution in
497 Bangladesh, India and Pakistan. *Parasitol Res* (2007) **101**:207–216. doi:10.1007/s00436-007-0684-7
- 498 71. Heath A. Implications for New Zealand of potentially invasive ticks sympatric with *Haemaphysalis longicornis* Neumann, 1901 (Acari:
499 Ixodidae). *Syst Appl Acarol* (2013) **18**:1–26. doi:10.11158/saa.18.1.1
- 500 72. Hoogstraal H, Roberts FHS, Kohls GM, Tipton VJ. Review of *Haemaphysalis* (Kaiseriana) *longicornis* Neumann (resurrected) of Australia,
501 New Zealand, New Caledonia, Fiji, Japan, Korea, and northeastern China and USSR, and its parthenogenetic and bisexual populations
502 (Ixodoidea, Ixodidae). *J Parasitol* (1968) **54**:1197–1213. Available at: <http://hbs.bishopmuseum.org/fiji/pdf/hoogstraal-et-al1968.pdf>
503 [Accessed March 17, 2019]

- 504 73. Marano N, Arguin PM, Pappaioanou M. Impact of Globalization and Animal Trade on Infectious Disease Ecology. *Emerg Infect Dis*
505 (2007) **13**:1807–1809. doi:10.3201/eid1312.071276
- 506 74. Fèvre EM, Bronsvoort BM de C, Hamilton KA, Cleaveland S. Animal movements and the spread of infectious diseases. *Trends Microbiol*
507 (2006) **14**:125–131. doi:10.1016/j.tim.2006.01.004
- 508 75. Getachew T, Haile A, Wurzinger M, Rischkowsky B, Gizaw S, Abebe A, Sölkner J. Review of sheep crossbreeding based on exotic sires
509 and among indigenous breeds in the tropics: An Ethiopian perspective. *African J Agric Res* (2016) **11**:901–911.
510 doi:10.5897/AJAR2013.10626
- 511 76. Mbuku SM, Okeyo AM, Kosgey IS, Kahi AK. Optimum crossbreeding systems for goats in low-input livestock production system in
512 Kenya. *Small Rumin Res* (2015) **123**:55–61. doi:10.1016/J.SMALLRUMRES.2014.10.001
- 513 77. Kosgey IS, Baker RL, Udo HMJ, Van Arendonk JAM. Successes and failures of small ruminant breeding programmes in the tropics: a
514 review. *Small Rumin Res* (2006) **61**:13–28. doi:10.1016/J.SMALLRUMRES.2005.01.003
- 515 78. O'Connor AM, Sargeant JM. An introduction to systematic reviews in animal health, animal welfare, and food safety. *Anim Heal Res Rev*
516 (2014) **15**:3–13. doi:10.1017/S146625231400005X
- 517 79. Application of systematic review methodology to food and feed safety assessments to support decision making. *EFSA J* (2010) **8**:
518 doi:10.2903/j.efsa.2010.1637
- 519 80. Grindlay DJC, Brennan ML, Dean RS. Searching the veterinary literature: A comparison of the coverage of veterinary journals by nine
520 bibliographic databases. *J Vet Med Educ* (2012) **39**:404–412. doi:10.3138/jvme.1111.109R
- 521 81. Moher D, Liberati A, Tetzlaff J, Altman DG, PRISMA Group. Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The
522 PRISMA Statement. *PLoS Med* (2009) **6**:e1000097. doi:10.1371/journal.pmed.1000097
- 523 82. Liberati A, Altman DG, Tetzlaff J, Mulrow C, Gøtzsche PC, Ioannidis JPA, Clarke M, Devereaux PJ, Kleijnen J, Moher D. The PRISMA
524 Statement for Reporting Systematic Reviews and Meta-Analyses of Studies That Evaluate Health Care Interventions: Explanation and
525 Elaboration. *PLoS Med* (2009) **6**:e1000100. doi:10.1371/journal.pmed.1000100
- 526 83. Burt FJ, Spencer DC, Leman PA, Patterson B, Swanepoel R. Investigation of tick-borne viruses as pathogens of humans in South Africa
527 and evidence of Dugbe virus infection in a patient with prolonged thrombocytopenia. *Epidemiol Infect* (1996) **116**:353–361. Available at:
528 <http://www.ncbi.nlm.nih.gov/pubmed/8666081> [Accessed March 17, 2019]
- 529 84. Soberón J, Peterson AT. Interpretation of models of fundamental ecological niches and species' distributional areas. *Biodivers Informatics*
530 (2005)1–10. Available at:
531 https://kuscholarworks.ku.edu/bitstream/handle/1808/20560/soberon_interpretation.pdf?sequence=1&isAllowed=y [Accessed March 17,
532 2019]
- 533 85. Barve N, Barve V, Jiménez-Valverde A, Lira-Noriega A, Maher SP, Peterson AT, Soberón J, Villalobos F. The crucial role of the
534 accessible area in ecological niche modeling and species distribution modeling. *Ecol Modell* (2011) **222**:1810–1819.
535 doi:10.1016/J.ECOLMODEL.2011.02.011
- 536 86. Olwoch JM, Reyers B, Engelbrecht FA, Erasmus BFN. Climate change and the tick-borne disease, Theileriosis (East Coast fever) in sub-
537 Saharan Africa. *J Arid Environ* (2008) **72**:108–120. doi:10.1016/J.JARIDENV.2007.04.003
- 538 87. Tukahirwa EM. The effects of temperature and relative humidity on the development of *Rhipicephalus appendiculatus* Neumann (Acarina,
539 Ixodidae). *Bull Entomol Res* (1976) **66**:301–312. doi:10.1017/S0007485300006696
- 540 88. Latha BR, Aiyasami SS, Pattabiraman G, Sivaraman T, Rajavelu G. Seasonal activity of ticks on small ruminants in Tamil Nadu State,
541 India. *Trop Anim Health Prod* (2004) **36**:123–133. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/14998311> [Accessed March 17,
542 2019]
- 543 89. Uilenberg G. General review of tick-borne diseases of sheep and goats world-wide. *Parassitologia* (1997) **39**:161–165. Available at:
544 <http://www.ncbi.nlm.nih.gov/pubmed/9530703> [Accessed March 17, 2019]
- 545 90. Peterson AT. Ecologic Niche Modeling and Spatial Patterns of Disease Transmission. *Emerg Infect Dis* (2006) **12**:1822.
546 doi:10.3201/EID1212.060373
- 547 91. Cobos ME, Peterson AT, Barve N, Osorio-Olvera L. kuenm: an R package for detailed development of ecological niche models using
548 Maxent. *PeerJ* (2019) **7**:e6281. doi:10.7717/peerj.6281
- 549 92. Steven J. Phillips, Miroslav Dudík RES. Maxent software for modeling species niches and distributions (Version 3.3.3 k). Available at:

- 550 http://biodiversityinformatics.amnh.org/open_source/maxent/
- 551 93. Owens HL, Campbell LP, Dornak LL, Saupe EE, Barve N, Soberón J, Ingenloff K, Lira-Noriega A, Hensz CM, Myers CE, et al.
552 Constraints on interpretation of ecological niche models by limited environmental ranges on calibration areas. *Ecol Modell* (2013) **263**:10–
553 18. doi:10.1016/j.ecolmodel.2013.04.011
- 554 94. Merow C, Smith MJ, Silander JA. A practical guide to MaxEnt for modeling species' distributions: what it does, and why inputs and
555 settings matter. *Ecography (Cop)* (2013) **36**:1058–1069. doi:10.1111/j.1600-0587.2013.07872.x
- 556 95. Peterson AT, Soberón J, Pearson RG, Anderson RP, Martínez-Meyer E, Nakamura M, Araújo MB. *Ecological Niches and Geographic*
557 *Distributions (MPB-49)*. Princeton University Press (2011). doi:10.23943/princeton/9780691136868.001.0001
- 558 96. Machado G, Weiblen C, Escobar LE. Potential distribution of *Pythium insidiosum* in Rio Grande do Sul, Brazil, and projections to
559 neighbour countries. *Transbound Emerg Dis* (2018) **65**:1671–1679. doi:10.1111/tbed.12925
- 560 97. Cohen TM, McKinney M, Kark S, Dor R. Global invasion in progress: modeling the past, current and potential global distribution of the
561 common myna. *Biol Invasions* (2019)1–15. doi:10.1007/s10530-018-1900-3
- 562 98. Sutton GF. Searching for a needle in a haystack: Where to survey for climatically-matched biological control agents for two grasses
563 (*Sporobolus* spp.) invading Australia. *Biol Control* (2019) **129**:37–44. doi:10.1016/j.biocontrol.2018.11.012
- 564 99. Benedict MQ, Levine RS, Hawley WA, Lounibos LP. Spread of the tiger: Global risk of invasion by the mosquito *Aedes albopictus*.
565 *Vector-Borne Zoonotic Dis* (2007) **7**:76–85. doi:10.1089/vbz.2006.0562
- 566 100. Peterson AT, Bauer JT, Mills JN. Ecologic and Geographic Distribution of Filovirus Disease. *Emerg Infect Dis* (2004) **10**:40–47.
567 doi:10.3201/eid1001.030125
- 568 101. DeVaney SC, McNyset KM, Williams JB, Peterson AT, Wiley EO. A tale of four “carp”: Invasion potential and ecological niche modeling.
569 *PLoS One* (2009) **4**: doi:10.1371/journal.pone.0005451
- 570 102. González C, Wang O, Strutz SE, González-Salazar C, Sánchez-Cordero V, Sarkar S. Climate change and risk of leishmaniasis in North
571 America: Predictions from ecological niche models of vector and reservoir species. *PLoS Negl Trop Dis* (2010) **4**:
572 doi:10.1371/journal.pntd.0000585
- 573 103. Gálvez R, Descalzo MA, Guerrero I, Miró G, Molina R. Mapping the current distribution and predicted spread of the leishmaniasis sand fly
574 vector in the Madrid Region (Spain) based on environmental variables and expected climate change. *Vector-Borne Zoonotic Dis* (2011)
575 **11**:799–806. doi:10.1089/vbz.2010.0109
- 576 104. Daszak P, Zambrana-Torrel C, Bogich TL, Fernandez M, Epstein JH, Murray KA, Hamilton H. Interdisciplinary approaches to
577 understanding disease emergence: The past, present, and future drivers of Nipah virus emergence. *Proc Natl Acad Sci U S A* (2013)
578 **110**:3681–3688. doi:10.1073/pnas.1201243109
- 579 105. Baquero OS, Machado G. Spatiotemporal dynamics and risk factors for human Leptospirosis in Brazil. *Sci Rep* (2018) **8**:
580 doi:10.1038/s41598-018-33381-3
- 581 106. Velasco JA, Poe S, González-Salazar C, Flores-Villela O. Solitary ecology as a phenomenon extending beyond insular systems: Exaptive
582 evolution in *Anolis* lizards. *Biol Lett* (2019) **15**: doi:10.1098/rsbl.2019.0056
- 583 107. Raghavan RK, Peterson AT, Cobos ME, Ganta R, Foley D. Current and Future Distribution of the Lone Star Tick, *Amblyomma*
584 *americanum* (L.) (Acari: Ixodidae) in North America. *PLoS One* (2019) **14**:e0209082. doi:10.1371/journal.pone.0209082
- 585 108. Burrridge MJ. Ticks (Acari: Ixodidae) spread by the international trade in reptiles and their potential roles in dissemination of diseases. *Bull*
586 *Entomol Res* (2001) **91**:3–23. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/11354992> [Accessed March 17, 2019]
- 587 109. Keirans JE, Durden LA. Invasion: exotic ticks (Acari: Argasidae, Ixodidae) imported into the United States. A review and new records. *J*
588 *Med Entomol* (2001) **38**:850–861. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/11761384> [Accessed March 17, 2019]
- 589 110. Caminade C, McIntyre KM, Jones AE. Impact of recent and future climate change on vector-borne diseases. *Ann N Y Acad Sci* (2019)
590 **1436**:157–173. doi:10.1111/nyas.13950
- 591 111. Tabachnick WJ. Challenges in predicting climate and environmental effects on vector-borne disease epistystems in a changing world. *J Exp*
592 *Biol* (2010) **213**:946–954. doi:10.1242/JEB.037564
- 593 112. Maclachlan NJ, Zientara S, Wilson WC, Richt JA, Savini G. Bluetongue and epizootic hemorrhagic disease viruses: recent developments
594 with these globally re-emerging arboviral infections of ruminants. *Curr Opin Virol* (2019) **34**:56–62. doi:10.1016/J.COVIRO.2018.12.005
595

596 **Figure Legend**

597

598 **Figure 1.** Workflow of the modeling process. **A)** Data collection based on literature review, **B)** Study area definition, **C)** Environmental variables
599 selection, **D)** Model calibration, and **E)** Final ecological niche model.

600

601 **Figure 2.** PRISMA flow diagram detailing the literature selection process (81).

602

603 **Table 1.** Variables used for the NSDv ecological niche models.

Environmental Variable	Unit	Source
Minimum temperature	°C	MODISsp R package and https://neo.sci.gsfc.nasa.gov/
Annual precipitation	mm	https://climate.northwestknowledge.net/TERRACLIMATE/index_directDownloads.php
Runoff	mm	https://climate.northwestknowledge.net/TERRACLIMATE/index_directDownloads.php
Evapotranspiration	mm	https://climate.northwestknowledge.net/TERRACLIMATE/index_directDownloads.php
Soil moisture (Wetness index)	mm	http://data.isric.org
Livestock density	ind/km ²	https://dataverse.harvard.edu/dataverse/glw_3

604

605 **Figure 3.** Case distribution of NSDv used for model calibration. Points denote occurrence locations. Color represents positive host (red) and tick
606 (blue) occurrences and size denotes the year of publication.

607

608 **Figure 4.** Model calibration statistics for hosts.

609

610 **Figure 5.** Model calibration statistics for ticks.

611

612 **Figure 6.** Model calibration statistics for hosts and ticks.

613

614 **Figure 7.** Ecological niche models of **a)** hosts and ticks, **b)** hosts and **c)** tick circulation. Warmer colors represent areas with higher environmental
615 suitability.

616

617 **Table 2.** Environmental preference of hosts, ticks, and hosts & ticks based on their potential distributions.

618

	Statistic	Evapo. (mm)	Livestock (Ind/km ²)	Min. temp. (°C)	Precip. (mm)	Runoff (mm)	Soil moist. (mm)
Hosts							
	Mean	46.1	11471.6	12.8	40.3	2.8	39.6
	Minimum	0.0	472.6	-6.7	0.0	0.0	0.0
	Maximum	164.0	247138.0	22.9	343.0	223.0	371.0
	DesvEst	43.1	12683.9	4.61	48.8	8.2	50.8
	Range	164.0	246665.4	29.6	343.0	223.0	371.0
	IQR	83.0	9990.2	5.8	63.0	3.0	62.0

Ticks	Mean	24.0	11812.0	3.9	13.8	0.8	62.2
	Minimum	0.0	7.2	-32.0	0.0	0.0	0.0
	Maximum	145.0	182469.0	23.3	230.0	107.0	588.0
	DesvEst	23.1	9575.1	12.4	22.3	2.2	71.7
	Range	145.0	182461.8	55.3	230.0	107.0	588.0
	IQR	38.0	8953.8	15.2	13.0	1.0	74.0

Hosts and ticks	Mean	39.7	10267.0	12.8	35.4	2.2	31.8
	Minimum	0.0	472.6	-0.5	0.0	0.0	0.0
	Maximum	161.0	148964.0	22.2	343.0	223.0	440.0
	DesvEst	41.2	9545.4	4.1	42.8	5.4	43.8
	Range	161.0	148491.4	22.7	343.0	223.0	440.0
	IQR	65.0	9434.4	5.5	58.0	3.0	52.0

619
620
621

A

Literature selection

1998-2010-01-01 2010-12-31

Number of records about a pathogen of interest in each state and volume of trade are selected as input data for the model.

1998-2010-01-01 2010-12-31

Number of records about a pathogen of interest in each state and volume of trade are selected as input data for the model.

1998-2010-01-01 2010-12-31

1998-2010-01-01 2010-12-31



B

Study area definition



C

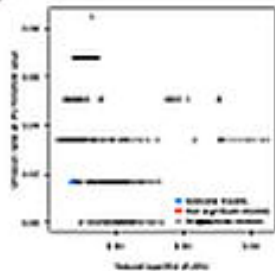
Environmental variables selection



D

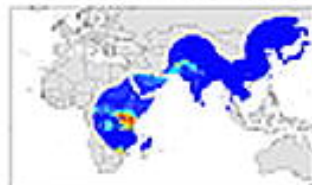
Model calibration

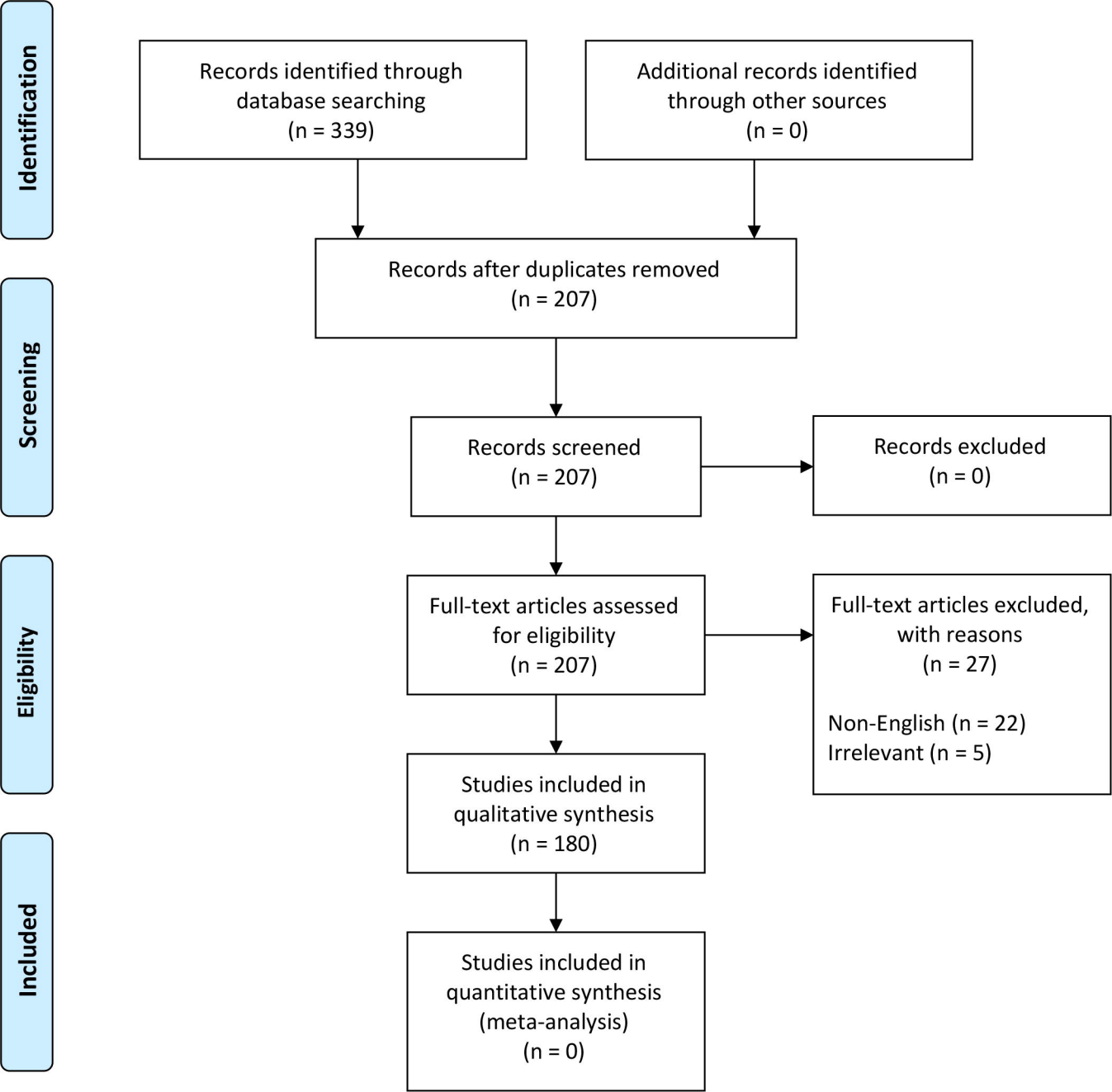
Year	Number of records	Volume of trade	Number of records	Volume of trade
2000	100	0	0	0
2001	100	0	0	0
2002	100	0	0	0
2003	100	0	0	0
2004	100	0	0	0
2005	100	0	0	0
2006	100	0	0	0
2007	100	0	0	0
2008	100	0	0	0
2009	100	0	0	0
2010	100	0	0	0

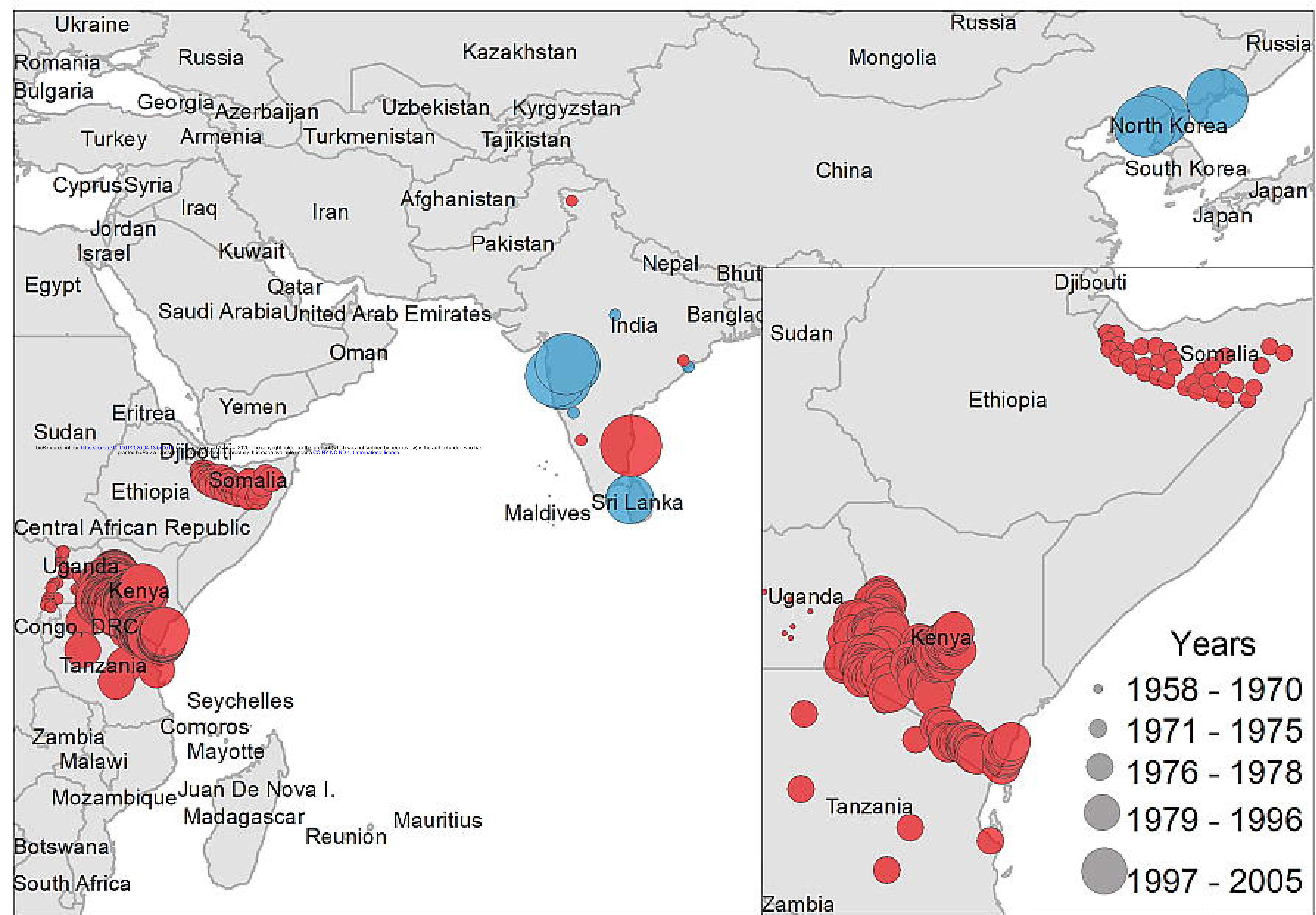


E

Ecological Niche Model

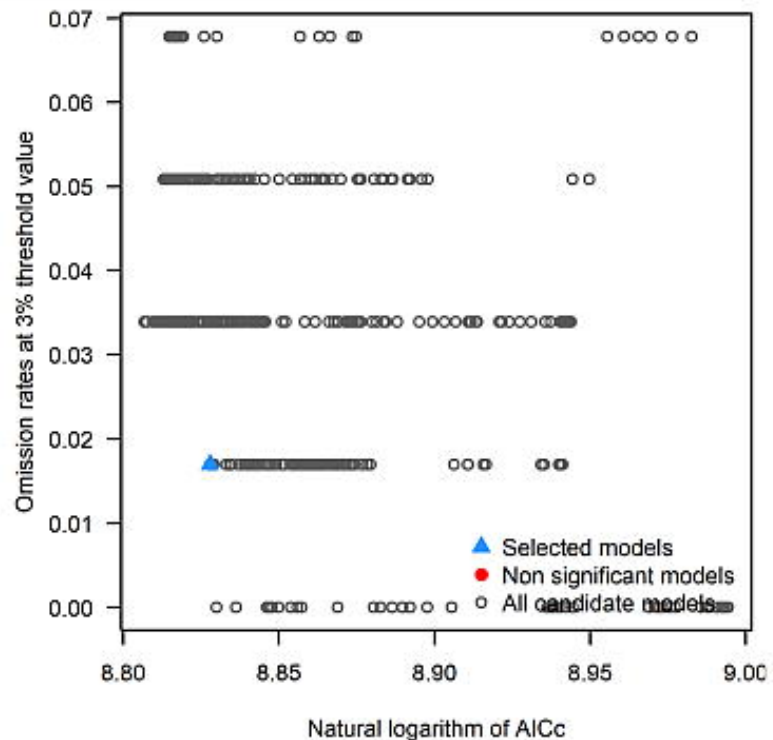






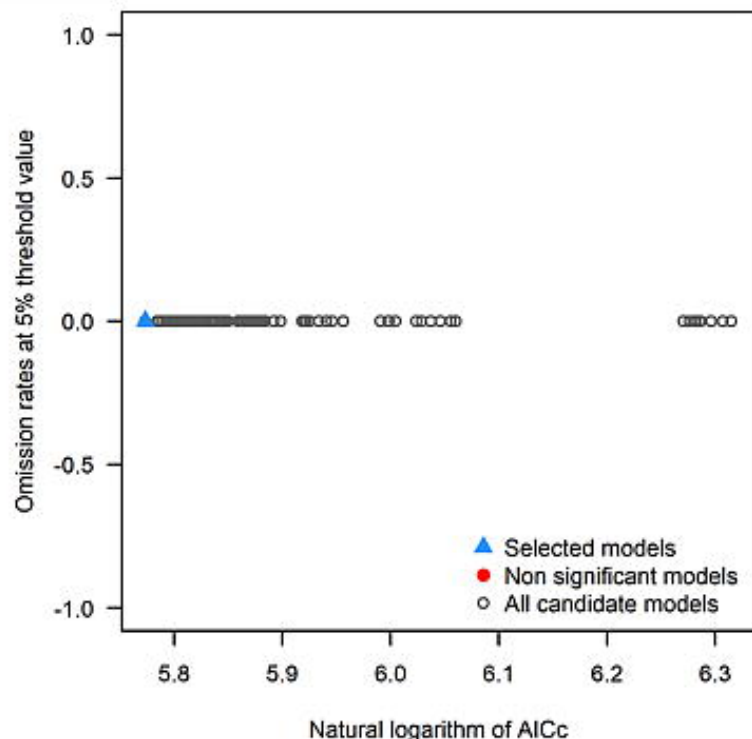
Model	Mean AUC ratio	Partial ROC	Omission rate 5%	AICc	Delta AICc	Weight AICc	Number of parameters
R=3.0; F=lph	1.53	0	0.017	6821.55	0	0.99	38

Criteria	Number of models
All candidate models	493
Statistically significant models	490
Models meeting omission rate criteria	160
Models meeting AICc criteria	1
Statistically significant models meeting omission rate criteria	157
Statistically significant models meeting AICc criteria	1
Statistically significant models meeting omission rate and AICc criteria	1



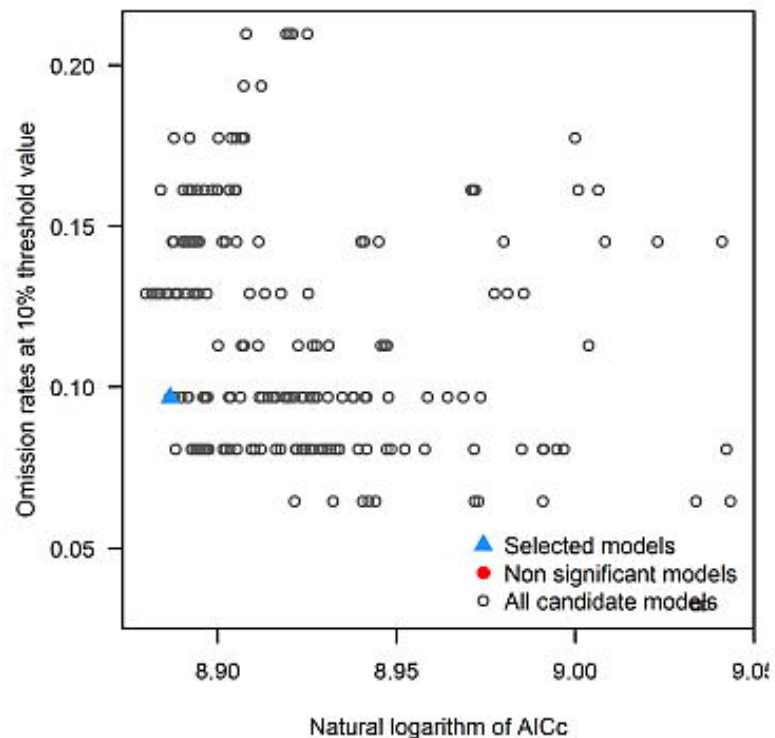
Model	Mean AUC ratio	Partial ROC	Omission rate 5%	AICc	Delta AICc	Weight AICc	Number of parameters
R=1.0; F=It	1.88	0	0	321.63	0	0.36	5

Criteria	Number of models
All candidate models	493
Statistically significant models	367
Models meeting omission rate criteria	493
Models meeting AICc criteria	2
Statistically significant models meeting omission rate criteria	367
Statistically significant models meeting AICc criteria	1
Statistically significant models meeting omission rate and AICc criteria	1

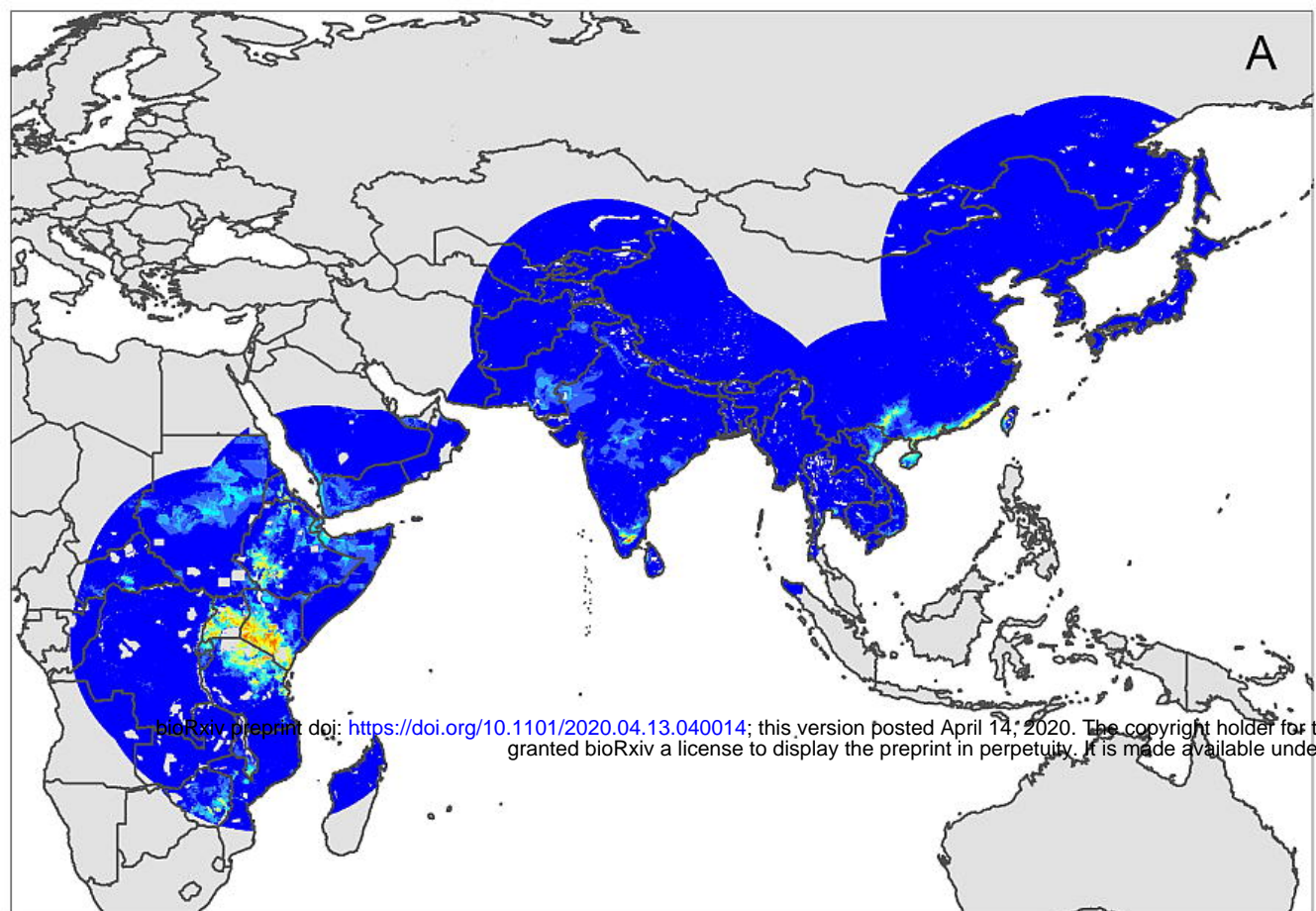


Model	Mean AUC ratio	Partial ROC	Omission rate 10%	AICc	Delta AICc	Weight AICc	Number of parameters
R=0.6; F=qph	1.79	0	0.097	7234.58	0	1	78

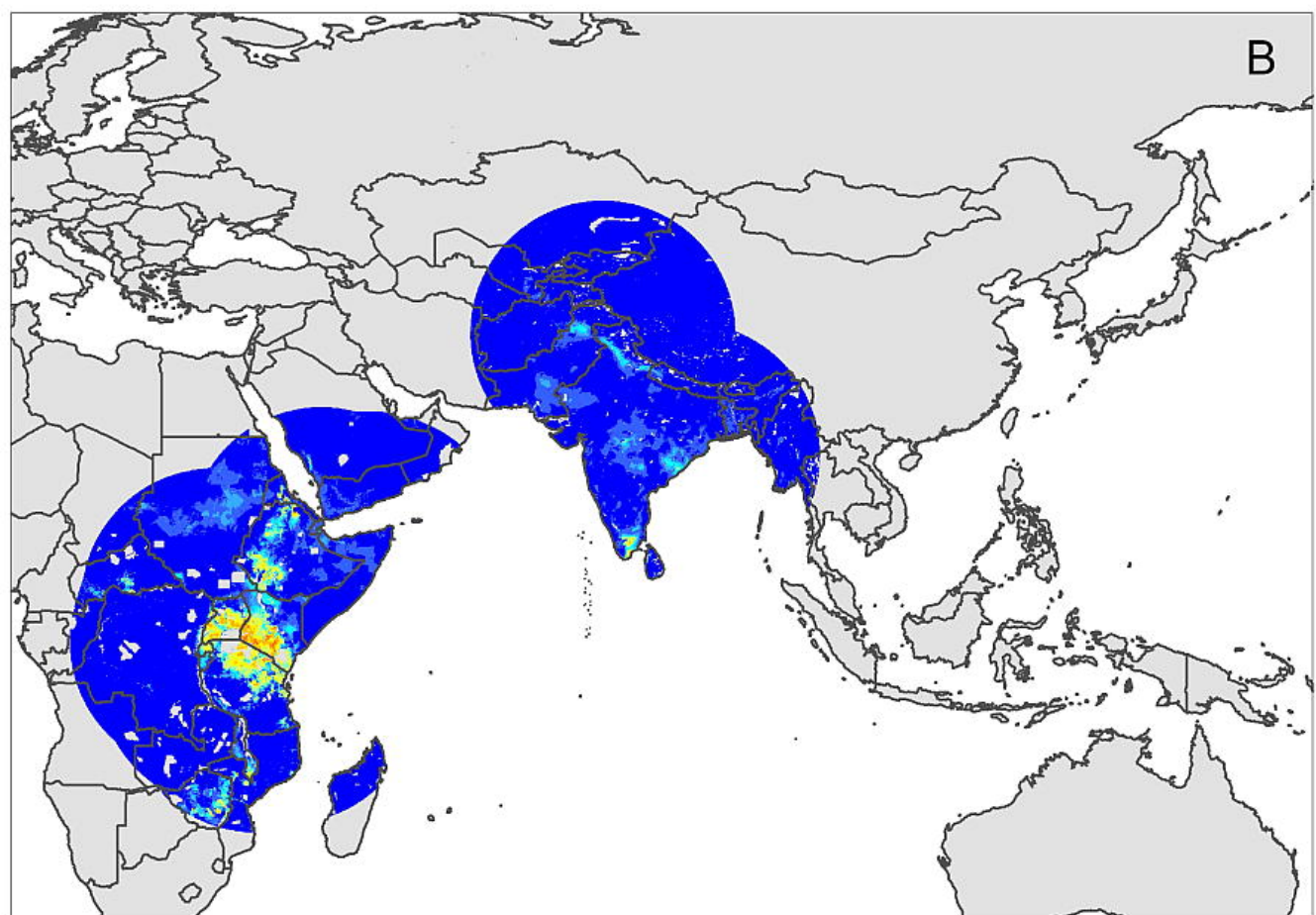
Criteria	Number of models
All candidate models	493
Statistically significant models	209
Models meeting omission rate criteria	267
Models meeting AICc criteria	1
Statistically significant models meeting omission rate criteria	109
Statistically significant models meeting AICc criteria	0
Statistically significant models meeting omission rate and AICc criteria	1



A



B



C

