The macroecology and evolution of avian competence for *Borrelia burgdorferi*

Daniel J. Becker[1,2] & Barbara A. Han[3]

[1]Department of Biology, Indiana University, Bloomington, IN

[2]Center for the Ecology of Infectious Disease, University of Georgia, Athens, GA

[3]Cary Institute of Ecosystem Studies, Millbrook, NY

Running head: Avian competence for *Bb*sl

Keywords: machine learning; boosted regression tree; birds; transmission; tick-borne disease;

Lyme borreliosis; phylogenetics

Abstract

Most efforts to predict novel reservoirs of emerging infectious diseases rely upon information about the infection status of host species rather than their competence, the ability to transmit pathogens to new susceptible hosts or vectors. Although competence can be difficult to quantify, tick-borne pathogens can provide a useful model system, as larval ticks can become infected only when feeding on a competent host during their first bloodmeal. For tick-borne diseases, competence has been best studied for *Borrelia burgdorferi* sensu lato (*Bb*sl), the pathogen that causes Lyme borreliosis, an increasing public health priority across the northern hemisphere. Major reservoirs include several small mammal species, but birds may play an underrecognized role in human risk of acquiring this tick-borne pathogen, given their ability to disperse infected ticks across large spatial scales. Here, we provide a global synthesis of the ecological and evolutionary factors that determine the ability of bird species to infect larval ticks with *Bb*sl. We find strong phylogenetic patterns in competence, which clusters in thrushes and has evolved multiple times across birds. With boosted regression trees, we distinguish competent birds with 76% accuracy and show competent species are more likely to fledge early, have low baseline corticosterone, breed at high latitudes, and have more diverse migratory movements into their breeding range. We use these trait profiles to predict over 100 likely but unsampled *Bb*sl-competent birds, including novel concentrations of avian reservoirs in the Neotropics and pelagic zones. Beyond expanding our understanding of avian reservoirs for tick-borne pathogens and generating new hypotheses, our findings also emphasize the need to consider competence in efforts to predict novel hosts of emerging diseases and in the study of multi-host pathogens.

Introduction

As most emerging infectious diseases originate in animals [1], a diverse array of efforts has

aimed to predict reservoir hosts and arthropod vectors of zoonotic pathogens [2–4]. Because such

predictions can guide surveillance and even interventions, they are necessary steps toward a

preemptive approach to minimizing pathogen spillover risks [5,6]. For example, PCR and

serological data on Nipah virus from wild bats were leveraged to prioritize field sampling targets

across India in response to a novel human outbreak [7], and similar approaches have been

successfully applied to identify hosts of filoviruses, rodent zoonoses, and Zika virus [8–10].

However, pathogen detection data likely better reflect host exposure rather than *competence*,

which is the ability of a host to transmit a pathogen to a new host or vector [11]. Greater

attention to competence could thus improve predictions about novel reservoirs of zoonoses.

Host competence is an inherently individual-level trait driven by immunological and

behavioral factors [12,13]. This individual heterogeneity mediates intra- and inter-specific

variation in the propensity to transmit pathogens, which is a key determinant of infection

dynamics in complex host communities [14]. Species-level variation in competence can produce

hosts with disproportionate contributions to pathogen transmission [15]. For example, American

robins (*Turdus migratorius*) infect up to 40% of mosquito vectors with West Nile virus despite

their lower relative abundance within the avian host community [16], and the transmission of

many helminths is often dominated by single host species in small mammal communities [17].

Identifying the broader ecological or evolutionary drivers of competence could in turn illuminate

how different host species contribute to the transmission of multi-host pathogens [18,19].

Competence can be difficult to quantify, as infection status of the donor and recipient

host must be known or directional transmission events must be inferred [12,20]. However, many

tick-borne pathogens provide an useful system to approximate competence, as tick larvae hatch

free of some pathogens and only become infectious with their first vertebrate bloodmeal [21].

Accordingly, competence can be inferred from the presence of pathogens in engorged larval ticks

on the donor host [22]. In this context, inter-specific variation in competence has been best

studied for *Borrelia burgdorferi* sensu lato (*Bb*sl), which causes Lyme borreliosis. *Bb*sl is

transmitted to humans by nymph *Ixodes* spp. ticks and has infection foci across the northern

hemisphere and parts of Latin America [23,24]. Lyme borreliosis is the most common vector-

borne disease in the United States (i.e., Lyme disease) [25], where fast-lived rodent species, in

particular white-footed mice (*Peromyscus leucopus*) and eastern chipmunks (*Tamias striatus*),

infect high proportions of larvae and are the most competent hosts among mammals [26–28].

Through infecting the majority of naïve vectors, these host species contribute disproportionally

to the production of infectious nymphs and therefore to human Lyme borreliosis risk [29,30].

68   In contrast to mammals, birds likely play an underrecognized role in the global ecology

of *Bb*sl, given that the capacity for flight, and especially long-distance migration, can allow avian

hosts to disperse infected ticks across entire continents [31–34]. Migratory birds likely transport

50–175 million ticks across Canada each spring [35], and the physiological stress of migration

itself may help drive reactivation of latent *Bb*sl infection [36]. Individual studies have suggested

some birds have important contributions to the enzootic maintenance of *Bb*sl [37,38]. For

example, birds play the dominant role in transmission of *B. garinii* and *B. valaisiana* to larvae

across Europe [39,40] and contribute to enzootic cycles of the primarily rodent genospecies, *B.*

afzelli [41]. In North America, *Bb* sensu stricto infects both rodent and avian species; however,

as with mammals, birds seemingly also display broad variation in competence. American robins

can infect up to 90% of naïve larvae [42], whereas gray catbirds (*Dumetella carolinensis*) and

veeries (*Catharus fuscescens*) display lower competence [43–45]. High tick burdens on some

avian species, such as ground foragers, may allow some birds to contribute more to community-

level *Bb*sl transmission than some rodent hosts [46,47]. Lastly, many bird species capable of

infecting larvae are also common in suburban and urban habitats [48,49], which could increase

human exposure to infectious nymphal ticks [50]. Yet despite these opportunities for birds to

play key roles in the global distribution of *Bb*sl and Lyme borreliosis risk, the inter-specific

drivers of reservoir competence across bird species have not yet been systematically identified.

Given the potential for birds to play important roles in the dynamics of this tick-borne

pathogen, we here compile a comprehensive, global dataset on avian competence for *Bb*sl and

assess its ecological and evolutionary drivers. We first describe spatial and temporal patterns in

competence, including testing for latitudinal gradients and seasonal variation among species.

Next, we characterize phylogenetic patterns in competence and reconstruct its evolution across

birds. We lastly use a flexible machine learning algorithm to evaluate the trait profiles associated

with competent avian species. For the latter, such data science approaches circumvent many

issues associated with traditional hypothesis testing (e.g., a large number of predictors, complex

interactions, non-randomly missing covariates) and can uncover new and surprising patterns in

data, thereby developing testable hypotheses [51,52]. Our work therefore aims to identify the

ecological and evolutionary drivers of avian competence while also generating predictions of

likely novel *Bb*sl reservoirs and directions for future studies of tick-borne disease more broadly.

Methods

Competence data

To collate data on avian competence for *Bb*sl, we searched Web of Science, PubMed, and CAB

Abstracts with the following string: ("reservoir competence" OR "host competence" OR

prevalence) AND (bird* OR Aves) AND (larva* OR tick* OR arthropod*) AND ("Lyme

disease" OR *Borrelia* OR "*B. burgdorferi*"). Using a systematic protocol (Fig. S1) [53], we

included only studies of experimental trials (e.g., infecting birds and using xenodiagnoses with

naïve larvae to test bird-to-tick transmission) or that approximated competence by testing

engorged larvae from wild birds for *Bb*sl. We excluded studies that only tested larvae for non-

*Bb*sl *Borrelia* (e.g., *B. lonestari*), only tested nymph or adult ticks, pooled ticks by life stage,

only tested birds themselves (e.g., blood or tissue), or pooled competence across bird species.

110   We identified 102 studies for inclusion, from which we recorded the sampling country

and coordinates (or used centroids of reported regions), sampling months and years, bird and

larval tick species, number of sampled and *Bb*sl-positive larvae, the *Bb*sl genospecies, and

competence assessment type (experimental trial or testing attached larvae). As most studies had

multiple lines of data (88/102), each record (*n*=1069) was a test of a bird–tick–*Bb*sl association

over space and time. Most records were from *Ixodes* ticks (90.11%, mostly *I. ricinus* and *I.*

scapularis), with the remainder unstated (1.24%) or from *Haemaphysalis* (4.85%), *Hyalomma*

(3.71%), or *Amblyomma* (0.10%). Whereas tick genera other than *Ixodes* are unlikely *Bb*sl

vectors [54], we retained these records as they still represent transmission from competent birds.

Meta-analysis of larval infection prevalence

We first used a phylogenetic meta-analysis to quantify heterogeneity in prevalence of *Bb*sl in

bird-fed larvae (*n*=964, Fig. S2; some studies only reported presence or absence of infected

larvae). We obtained a phylogeny of our 183 bird species from the Open Tree of Life with the

rotl package in R and used the *ape* package to calculate branch lengths [55,56]. We then used the

metafor package to estimate logit-transformed proportions and sampling variances [57]. We next

used restricted maximum likelihood to fit a random-effects model (REM), which included a

species-level random effect (the covariance structure used the phylogenetic correlation matrix),

observation nested within a study-level random effect, and weighting by sampling variances.

Resulting estimates of the variance components were used to derive I^2, the contribution of true

heterogeneity to the total variance in larval *Bb*sl prevalence, and to partition variance attributed

to each random effect. For avian species, we calculated phylogenetic heritability (H^2) [58].

132   To assess spatial and temporal variation in avian competence, we fit mixed-effects

models (MEMs) with the same random effects to the data subset describing *Bb*sl prevalence in

engorged larvae derived only from wild birds (n=922). Covariates included geographic region

(n=922), latitude (n=778), year (n=917), and season (n=654). As many studies pooled *Bb*sl

prevalence across time, we used either the given sampling year or the mid-point sampling year.

Similarly, we coded season as four binary covariates (winter, spring, summer, fall; only 10

prevalence estimates were from the southern hemisphere), as studies often reported data per

single season or pooled across multiple seasons. Given the differences in sample size between

predictors, we used Akaike information criterion corrected for small sample sizes (AICc) to

compare two sets of MEMs [59]: (*i*) space and year and (*ii*) space and season. Comparisons

included an intercept-only model, and we derived a pseudo-R^2 as the proportional reduction in

the summed variance components per MEM compared with that of an equivalent REM [60].

Phylogenetic analyses

We next aggregated tick–*Bb*sl data per avian species to assess phylogenetic patterns in binary

competence. Using the *caper* package, we calculated the *D* statistic, where a value of 1 indicates

a phylogenetically random trait distribution and a value of 0 indicates phylogenetic clustering

under a Brownian motion model of evolution [61]. Significant departure from either model was

quantified using a randomization test with 1000 permutations. However, because traits such as

competence may also arise under a punctuated equilibrium model of evolution, we next used a

graph-partitioning algorithm, phylogenetic factorization, to flexibly identify clades with

significantly different propensity to be competent at various taxonomic depths. We used the

taxize package to obtain a taxonomy from the NCBI database [62] and used the *phylofactor*

package to partition competence as a Bernoulli-distributed response in a generalized linear model

[63]. To account for variable study effort, we used the *rwos* package to quantify the number of

citations per species in Web of Science and used the square-root transformed values as weights

[7,8]. We determined the number of significant phylogenetic factors (clades) using a Holm's

sequentially rejective 5% cutoff for the family-wise error rate. Lastly, to assess if phylogenetic

patterns in competence could stem from study effort alone, we performed a secondary analysis to

partition Web of Science citation counts for each avian species as a quasi-Poisson response.

To investigate the evolution of competence along the avian phylogeny, we next used

the *ape* package and maximum likelihood to reconstruct the ancestral character state [56]. We

compared an equal-rate and an all-rates-different model of evolution using Akaike information

criterion (AIC) [64] and used the most competitive model to perform stochastic character

mapping with Markov chain Monte Carlo (*n*=1000) using the *phytools* package [65]. We

displayed the mean posterior probabilities of competence across our sampled avian phylogeny.

Trait-based analyses

We compiled avian trait data from EltonTraits [66], the Amniote Life History database [67],

International Union for the Conservation of Nature [68], and HormoneBase [69]. Traits included

diet composition and breadth, foraging strata (e.g., ground foraging), pace of life (e.g., maximum

lifespan, clutch size, fledging age, clutches per year), morphology (e.g., adult body mass,

hatching weight), maximum elevation, global population trend, and physiology (i.e., baseline

corticosterone; CORT). Using distribution maps from BirdLife International and the Handbook

of the Birds of the World [70], we derived total range size, latitude of the centroids of breeding

and non-breeding ranges, and mean migration distance (greater circle distance between these

centroids). We also quantified migratory dispersion, the extent to which species inhabit larger

(positive) or smaller (negative) areas in the non-breeding season relative to breeding range size,

and categorized migratory strategy (resident, full migrant, partial migrant) [71]. For resident

species, mean migration distance and dispersion were set to zero. To represent phylogeny, we

included binary covariates for each avian family and any clades identified by phylogenetic

factorization; we also had a binary covariate for Passeriformes given their overrepresentation in

our data (166/183 species). We also derived the degree of evolutionary isolation per species in

the avian phylogeny with the *picante* package [72]. We again used Web of Science citations per

species to approximate study effort. We transformed continuous predictors that spanned orders

of magnitude and excluded those with zero variance or missing values for over 75% of birds. We

compiled features for our 183 sampled birds and all remaining avian species to predict likely but

unsampled competent hosts. We limited out-of-sample species to the 39 families included in our

*Bb*sl dataset (approximately 4700 birds). Feature definitions and coverage are given in Table S1.

We used boosted regression trees (BRTs) to fit a predictive model relating binary

competence to a predictor matrix of all avian traits [73]. In our analysis, BRTs were trained to

maximize classification accuracy by learning patterns of features that best distinguish competent

and non-competent avian species. This generates recursive binary splits for randomly sampled

predictor variables, and successive trees are built using residuals of the prior best-performing tree

as the new response. Boosting generates an ensemble of linked trees, where each achieves

increasingly more accurate classification. Prior to analysis, we randomly split data into training

(80%) and test (20%) datasets while preserving the proportion of positive labels. Models were

trained with the *gbm* package [74], with the maximum number of trees set to 50000, a learning

rate of 0.0001, and an interaction depth of three [73]. BRTs used a Bernoulli error distribution

and 10-fold cross-validation, and we used the *ROCR* package to quantify classification accuracy

as area under the receiver operator curve (AUC) [75]. As BRT results can depend on random

splits between training and test data, we used five partitions to generate a model ensemble [76].

To diagnose if trait profiles of competent birds are driven by study effort, we ran a secondary

BRT ensemble that modeled Web of Science citation counts as a Poisson-distributed response.

After assessing accuracy of our BRTs against test data, we applied our model ensemble

to the full trait dataset to generate mean probabilities of *Bb*sl competence per avian species. To

assess potential phylogenetic signal in predictions, we estimated Pagel's λ in logit-transformed

mean probabilities with the *caper* package [77]. To guide surveillance for likely competent avian

reservoirs, we mapped the distributions of non-positive bird species with mean probabilities of

over 50%. We stratified distributions by avian phenology to emphasize when these unsampled

*Bb*sl reservoirs are most likely to encounter questing larvae (i.e., breeding and resident ranges).

Results

Avian competence on a global scale

Half of all sampled bird species were competent for Bbsl (91/183). Only nine species had

experimental evidence of competence, and all 91 competent species (with the exception of

Gallus gallus) had larvae from wild hosts test positive (Fig. S2). For records reporting Bbsl

genospecies from positive bird-fed larval ticks, our global-scale data were dominated by *B.*

garinii (31%), *B. valaisiana* (20%), *B. burgdorferi* sensu stricto (16%), and *B. afzelii* (15%),

representing the geographic skew of the data toward Europe (59%) and North America (37%);

3% of our data were from Eastern Asia, whereas 1% were from South America (Fig. 1A).

However, these 91 competent avian species were more broadly distributed across the Americas,

Africa, Asia, and Oceania, representing a mixture of resident, breeding, or wintering ranges.

We observed significant heterogeneity in bird-fed larval tick Bbsl prevalence (I^2=0.75,

Q_{963}=2970, p<0.0001). Avian species accounted for more of this variation ($I^2_{species}$=0.30) than

study (I^2_{study}=0.19) or individual record ($I^2_{observation}$=0.26), resulting in moderate phylogenetic

signal (H^2=0.40). Given the stronger effect of avian phylogeny, we found no effect of study-level

predictors such as space (Fig. 1B) or year (Fig. 1C) on the proportion of larval ticks infected by

wild birds (Table S2). Seasons were also generally uninformative (Table S3), but prevalence in

bird-infected larvae was lower for sampling conducted in summer (w_i=0.44, R^2=0.01; Fig. 1D).

Evolutionary patterns in donor transmissibility

We next considered competence as an intrinsic binary trait per avian species. We estimated

intermediate phylogenetic signal in the ability of birds to transmit Bbsl to larvae (D=0.78),

indicating significant phylogenetic clustering between randomness (p<0.001) and a Brownian

motion model of evolution ($p<0.001$). After controlling for study effort, phylogenetic

factorization identified one clade, the genus *Turdus*, as having a significantly greater likelihood

of including competent species when compared to other avian taxa (Fig. 2). In particular, all but

one member of this clade demonstrated the ability to infect larval ticks with *Bb*sl (92%).

Our secondary analysis identified seven taxa as having significantly different Web of

Science citation counts (Fig. S3), five of which were heavily studied single species: *Gallus*

gallus (4139), *Parus major* (3736), *Sturnus vulgaris* (2509), *Passer domesticus* (2162), and

Ficedula hypoleuca (1500). Disproportionately studied taxa also included the Phasianidae

($\overline{x}=609$) and a subclade of the Icteridae (genera *Quiscalus*, *Molothrus*, and *Agelaius*; $\overline{x}=545$).

These taxonomic patterns in Web of Science citation counts did not overlap with taxonomic

patterns in avian competence, suggesting the latter were not driven by study effort (Fig. S3).

The evolution of avian competence was best described by an equal-rate model ($w_i=0.73$).

Stochastic character mapping suggested equal transitions from non-competent to competent and

from competent to non-competent (Fig. 2), with the ancestral state being equivocal. Competence

was gained in the Turdidae as well as the Mimidae, Passerellidae, and Troglodytidae, with both

gains and losses in the Parulidae. We also observed a clear loss of competence within the

majority of the Carduelinae as well as within the Corvidae, Picidae, and Strigidae (Fig. 2).

Trait profiles of competent birds

Our BRT models distinguished competent from non-competent birds with a mean 76% accuracy

(AUC=0.76 $\pm$ 0.02 SE; Fig. 3A). The top features for describing *Bb*sl-competent avian species

included those relevant to life history (i.e., fledging age, maximum lifespan, birth and adult

weight, egg mass, incubation time, clutch size), physiology (i.e., baseline CORT), migration

(dispersion, distance), geography (breeding latitude, maximum elevation, geographic range size),

evolutionary isolation, and study effort. The family Turdidae was the only taxonomic feature

with non-trivial importance, and foraging was generally uninformative (Fig. 3B, Table S4).

*Bb*sl-competent bird species were partly described by a faster life history (Fig. 4).

Competent avian reservoirs have shorter incubation times and young that are smaller and fledge

earlier than their non-competent counterparts. However, competent birds also have longer

lifespans, lay smaller clutches, and have larger eggs and adults. Physiologically, competent birds

have lower baseline concentrations of CORT, the main avian glucocorticoid. Competent birds

also tend to breed at higher latitudes, have larger geographic ranges, occupy lower elevations,

and have stable or increasing population trends. The likelihood of competence was also greater

for species with negative migratory dispersion, indicating such hosts have larger breeding ranges

and greater diversity of migratory movements from their wintering to their breeding grounds.

Competence was less strongly related to mean migratory distance, with intermediate-distance

migrants being most likely to transmit *Bb*sl to larvae. Lastly, well-studied species are also more

likely to be competent. However, our secondary BRTs showed that Web of Science citations

were not predictable by these same traits (AUC=0.50 ± 0 SE), suggesting that the trait profile of

a competent bird is not confounded by the ecological traits associated with well-studied hosts.

Applying our BRT ensemble to trait and taxonomic data across the 39 sampled avian

families revealed at least 127 species that could be prioritized for *Bb*sl surveillance based on

feature similarity to known competent reservoirs, including six birds not currently known to be

competent and 121 unsampled birds (Fig. 5A). We observed strong phylogenetic signal in mean

predicted probabilities of competence (λ=0.95), indicating the influential traits revealed by our

BRTs are likely driven by lineages with high potential for being competent for *Bb*sl (Fig. 5B).

The geography of likely high-probability reservoirs revealed potential hotspots of

competent birds across their breeding (Fig. 5C), non-breeding (Fig. 5D), and resident (Fig. 5E)

ranges. Likely false negatives included the Indigo bunting (*Passerina cyanea*) [78–80], yellow-

breasted chat (*Icteria virens*) [81,82], common reed bunting (*Emberiza schoeniclus*) [39,83,84],

eyebrowed thrush (*Turdus obscurus*) [85], garden warbler (*Sylvia borin*) [86], and European pied

flycatcher (*Ficedula hypoleuca*) [86]. Likely but unsampled reservoirs included the American

goldfinch (*Spinus tristis*), Townsend's warbler (*Setophaga townsendi*), Abert's towhee (*Melozone*

aberti), Harris's sparrow (*Zonotrichia querula*), grasshopper sparrow (*Ammodramus*

savannarum), yellow-headed blackbird (*Xanthocephalus xanthocephalus*), northern mockingbird

(*Mimus polyglottos*), and eastern bluebird (*Sialia sialis*) in North America; Acadian flycatcher

(*Empidonax virescens*), red-eyed vireo (*Vireo olivaceus*), alder flycatcher (*Empidonax alnorum*),

Louisiana waterthrush (*Parkesia motacilla*), great crested flycatcher (*Myiarchus crinitus*), and

clay-colored thrush (*Turdus grayi*) across the Americas; corn bunting (*Emberiza calandra*) and

horned lark (*Eremophila alpestris*) in Eurasia; and thick-billed murre (*Uria lomvia*) and

rhinoceros auklet (*Cerorhinca monocerata*) across pelagic zones. The names and propensity

scores of bird species in the top 50th percentile of BRT predictions are provided in Table S2.

Discussion

Competence is increasingly recognized to be an important trait governing infectious disease

dynamics, especially in multi-host communities [11,12,27]. Efforts to predict new reservoirs of

zoonotic pathogens would be improved by identifying the ecological or evolutionary drivers of

the capacity for species to successfully transmit to recipient hosts or vectors [18,19]. To do this,

we examined Lyme borreliosis, a model system to study competence for a tick-borne disease.

Whereas inter-specific variation in this trait has been well-characterized for mammals [26–28],

competence has been comparatively understudied across avian hosts, despite their ability to

disperse infected ticks across large spatial scales and mounting evidence of high competence

across multiple species [42,43,87]. We show that taxonomic and life history traits of *Bb*sl

competence are likewise predictable across avian hosts. Our phylogenetic analyses demonstrate

that competence has evolved multiple times across birds and is pronounced in the genus *Turdus*.

Trait-based analyses distinguished competent avian hosts with 76% accuracy and emphasized

that such species fledge early, have low baseline CORT, breed at high latitudes, and have diverse

migratory movements into their breeding range. These trait profiles can be leveraged to prioritize

surveillance of likely but unsampled *Bb*sl-competent avian species and emphasize birds display

underrecognized variation in their contributions to the enzootic cycles of this zoonotic pathogen.

Although pathogen transmission inherently occurs from individual hosts, variation in

competence can arise across broader biological scales such as space or species [11,14,15]. Our

meta-analysis identified substantial heterogeneity in *Bb*sl prevalence from bird-fed larval ticks

(i.e., high I^2) that was better explained by bird phylogeny rather than space, and this intermediate

phylogenetic signal was also confirmed in our species-level analysis. Although intraspecific

variation in *Bb*sl competence may be high in some contexts (e.g., latitudinal gradients in Europe

[34]), the strong contribution of phylogeny likely obscured such patterns at a global scale across

bird assemblages. These analyses thus suggest that the ability to transmit *Bb*sl to larval ticks can

be considered an intrinsic trait of avian species. However, we note that most of our data only

inferred competence from presence of infected larvae on wild birds, and future experimental

infections and tests of bird-to-tick transmission are necessary to improve inference [43,87].

Our moderate estimates of phylogenetic signal in avian competence were reflected in

numerous gains and losses of this trait across the phylogeny, suggesting that competence has

evolved multiple times to form lineages of highly competent birds. Phylogenetic factorization

identified one of these clades, the genus *Turdus*, as being especially competent, even after

accounting for variable study effort across birds. Of the12 sampled *Turdus* species, all but one

(*T. obscurus*) had engorged larvae test positive for *Bb*sl [85], and two (*T. migratorius* and *T.*

merula) have experimentally infected naïve larvae [42,87]. Given the more extensive *Turdus*

clade (approximately 85 species), this result suggests the other remaining true thrushes could

also be competent reservoirs for *Bb*sl. *Turdus* thrushes are globally distributed owing to their

evolutionary origins in the eastern Palearctic before dispersal to Africa and radiation through

trans-Atlantic movements [88]. The genus displays wide variation in migratory strategy, with

resident, migrant, and partial migrants. However, most of the European *Turdus* are complete or

partial migrants [89], and photoperiod manipulations suggest annual migrations of at least one

species (*Turdus iliacus*) can reactivate latent *Bb*sl infections [36]. Future comparative studies

across *Turdus* thrushes that vary in both geography and migratory strategy could elucidate the

generality of this pattern and the broader inter-specific drivers of competence across this clade.

Prior work on competence for *Bb*sl in mammals suggests interspecific variation is shaped

by life history strategy. Fast-lived species, which display rapid development and high fecundity

at the expense of longevity, are more competent than slow-lived counterparts [26–28]. This

pattern is likely drive by tradeoffs with host immunity, such that fast-lived mammals invest more

in developmentally inexpensive innate defenses and in turn have weaker developmentally

expensive adaptive defenses [90]. Although host defense against *Bb*sl involves both innate and

adaptive mechanisms, robust *Bb*sl-specific antibody titers limit pathogen transmission to naïve

ticks [91]. This immunological basis to competence is likely to also occur in avian hosts, as fast-

lived bird species similarly invest less in adaptive defenses such as antibodies [92]. Our BRTs

provide some support for fast life histories shaping competence across avian hosts. Competent

birds had short incubation times and young that are small and fledge early, patterns consistent

with weaker antibody defenses that would facilitate *Bb*sl transmission to larvae. Yet competent

birds also had long lifespans, small clutches, and large eggs, which are consistent with a slow life

history that would support stronger adaptive immunity and thus weaker competence. Such

discrepancies could stem from competence occasionally occurring in families with slow life

histories, such as the Alcidae (i.e., *Fratercula arctica*) and Phasianidae (i.e., *Gallus gallus* and

Phasianus colchicus). While our sample size limited applying BRTs to only the passerines,

explicit comparisons of life history, antibody defense, and competence across the Passeriformes

could provide more robust and mechanistic tests of how pace of life shapes avian competence.

Our BRTs also identified CORT as an important predictor of avian competence for *Bb*sl.

Glucocorticoids such as CORT orchestrate physiological changes that allow animals to cope with

acute stressors; however, because elevated CORT can have deleterious effects over long

timescales, prolonged stressors can be immunosuppressive [93]. For example, prior work on

West Nile virus has shown that persistently elevated CORT can amplify competence by

increasing pathogen loads in birds and prolonging their infectious periods [94]. However, we

found that competent bird species were more likely to have lower baseline concentrations of this

hormone. At baseline, CORT has mostly metabolic functions that regulate energy storage and

mobilization, allowing animals to meet energetic demands and respond to adverse conditions

[93]. Low baseline CORT could also be linked to *Bb*sl competence by its association with

breeding latitude. Birds tend to display lower baseline CORT at high latitudes, which could

facilitate continued breeding in the face of suboptimal environments [95]. Such a mechanism

would be compatible with our finding that *Bb*sl competent species also generally breed at high

latitudes. Birds breeding further from the equator can have stronger tradeoffs between arms of

the immune system, such that high-latitude avian hosts mount weaker adaptive responses [96].

Birds breeding at high latitudes could thus display weaker antibody-mediated defenses that

increase *Bb*sl competence. Sampling *Bb*sl-competent birds across latitudinal gradients could

characterize such immunity tradeoffs and test if these restrict bird-to-tick transmission [97].

Alongside breeding latitude, our trait-based analyses identified several other predictors

related to bird geography and annual cycles, including low elevation, large distributions, and

negative migratory dispersion. For the latter, such values indicate avian species with a greater

diversity of migratory movements from their wintering to breeding grounds [71]. Associations

between more diverse spring migrations and competence could be driven by the physiological

changes incurred during this phase of the annual cycle, as long-distance movements demand

large energy expenditures that can in turn impair immune function [98]. These energetic costs

and shifts in immunity could cause latent *Bb*sl infections to reactivate, as previously illustrated

for redwings (*Turdus iliacus*) at the onset of migratory restlessness [36]. Such a mechanism

could facilitate competent birds arriving at their breeding grounds especially primed to infect

larval ticks. The generally positive association between migratory distance and competence

likewise supports the idea that longer biannual movements, in being more energetically costly,

could further promote these relapse events. Future work could explicitly test this hypothesis by

longitudinally sampling *Bb*sl-competent bird species across their annual cycles [99]. Similarly,

mathematical models of latent and relapsing infections could be expanded into a migratory

network to further understand the contexts when "migratory relapse" increases risk [100,101].

Results of our clade- and trait-based analyses also inform the surveillance of specific

species of wild birds for their contribution to the enzootic cycle of *Bb*sl and possibly other tick-

borne pathogens. In particular, phylogenetic factorization suggested unsampled *Turdus* thrushes

are especially likely to be competent, and our BRTs identified at least 14 unsampled species of

this genus as having a greater than 50% probability of being capable of infecting larvae (Table

S5). Similarly, this analysis suggested *T. obscurus* to be a likely false negative in our dataset

[85]. Some species such as *T. grayi* are known to be parasitized by ticks [102], whereas others

such as *T. torquatus* and *T. nigriceps* have had engorged nymphs test positive for *Bb*sl [83,103].

More broadly, the predicted probabilities from our BRTs displayed high phylogenetic

signal, suggesting general lineages of competent birds. Alongside the *Turdus*, the Parulidae

(New World warblers), Laridae (seabirds), Emberizidae (buntings), and Icteridae (icterids) had

relatively large numbers of likely but unsampled reservoirs. As with the *Turdus*, several species

in these families have had blood or nymphal ticks test positive for *Bb*sl, such as *Parkesia*

motacilla [81] and *Rissa tridactyla* [104]. Because some of these likely competent birds can be

common in suburban or urban habitats and could have unrecognized contributions to human

Lyme borreliosis risk [48,49], we suggest they be prioritized for spatiotemporal studies of

infection dynamics to identify when and where these species are most likely to infect ticks [105].

Experimental *Bb*sl infections and xenodiagnosis could also confirm competence of such species.

In addition to intrinsic factors, extrinsic environmental change could play an important

role in shaping how known and likely competent birds contribute to *Bb*sl dynamics and human

disease risk. Climate shifts and urbanization could alter bird geographic distributions and within-

host processes that influence intra-specific variation in competence. For the former, the breeding

distributions of many North American bird species are shifting north with climate change [106],

which could synchronize phenologies of competent birds and questing ticks if vectors also

expand northward [107]. Alternatively, warmer temperatures could facilitate shifts toward

residency, as observed for *Bb*sl-competent bird species such as *Turdus merula* [108]. If such

species become resident in parts of their range where ticks are abundant, sedentary behavior

could increase vector exposure and amplify bird–tick transmission cycles. Similarly, several

*Bb*sl-competent bird species (e.g., *Sylvia atricapilla*, *Turdus migratorius*, *Junco hyemalis*) and

unsampled but likely reservoirs (e.g., *Spinus tristis*) have shortened migration or become entirely

resident owing to abundant food in urbanized habitats [109–111]. However, urban residency

could also decrease host competence (e.g., if supplemental food increases antibody defense

[112]) or facilitate bird-to-tick transmission (e.g., if artificial night at light extends infectious

periods [113]). Combining sampling of known or likely competent birds across urban–rural

gradients or their historic and recent range with mathematical models could help forecast how

environmental change will alter bird distributions, competence, and contribution to *Bb*sl risk.

In conclusion, we demonstrate that host ability to transmit pathogens to new hosts or

vectors is predictable on the basis of ecological and evolutionary characteristics of bird species

within the Lyme borreliosis system. By combing flexible phylogenetic and trait-based analyses,

our work generates testable hypotheses for future comparative and theoretical studies of tick-

borne disease (e.g., how life history and migratory strategies shape competence) alongside

predictions that can inform bird surveillance efforts for *Bb*sl and similar pathogens (e.g.,

Anaplasma, *Ehrlichia*, *Babesia*) in the context of environmental change. Moving forward,

greater attention to the factors that shape competence within and between species could improve

our ability to predict and manage wild reservoirs for emerging infectious diseases more broadly.

Acknowledgments

We thank BirdLife International for providing avian distribution data, Tao Huang for assistance

with data processing, and JP Schmidt for technical advice on BRTs. We also thank Ellen

Ketterson, Alex Jahn. and members of the Ketterson lab for feedback on this manuscript.

Data availability

All spatiotemporal bird-fed larvae *Bb*sl infection prevalence data and binary competence per

sampled avian species are available in Dryad Digital Depository [114]:

https://datadryad.org/stash/share/4shvYd0_lCUkjEFhO5g1yc95sSLUfq3zbrd79_1J-0Q. All bird

species feature data used in the BRTs are available from their original sources [66–70].

Funding

DJB was supported by an appointment to the Intelligence Community Postdoctoral Research

Fellowship Program, administered by Oak Ridge Institute for Science and Education through an

interagency agreement between the U.S. Department of Energy and the Office of the Director of

National Intelligence. BAH was supported by the National Science Foundation Ecology and

Evolution of Infectious Diseases program (DEB-1717282 and DEB-1619072).

Competing interests

We declare no competing interests.

 Figures and legends

Figure 1. Global distribution of *Bb*sl infection prevalence in tick larvae sampled from wild birds.

(A) Sampled countries are shown with a black border alongside spatiotemporal bird–tick–*Bb*sl

prevalence estimates, which are sized by magnitude and colored by georegion. Shapefiles of the

91 competent species are overlaid in black and are from BirdLife International and the Handbook

of the Birds of the World [70]. Larval infection prevalence is plotted by absolute latitude (B),

year (C), and season (D), with each point representing a sampled bird–tick–*Bb*sl association;

points are colored by region and jittered to reduce overlap. Black points and lines (D) display

back-transformed predicted means and 95% confidence intervals from the MEM (Table S3).

Figure 2. Phylogenetic patterns in avian competence for *Bb*sl. The avian phylogeny displays observed species binary competence, and highlighted clades are those with significantly different competence from the paraphyletic remainder using phylogenetic factorization. Nodes show the mean posterior probabilities of being competent estimated from stochastic character mapping.

Figure 3. BRT performance in identifying traits predictive of avian competence for *Bb*sl and

their relative importance across five random partitions of training and test datasets. (A) Accuracy

is shown by the receiver operator curves obtained from 10-fold cross-validation on test data.

Grey lines show curves from each partition, whereas the black line displays the mean. (B)

Relative importance per feature is shown as the mean and standard error across partitions. Only

features with mean importance greater than 1% are shown (Table S4).

Figure 4. Trait profile of a *Bb*sl-competent avian species. Partial dependence plots of the top predictors across BRTs applied to five random partitions of training and test data are shown in order of mean relative importance. Grey lines or point show the marginal effect of a given variable for predicting *Bb*sl competence from each random data partition, whereas the black lines or squares display the average marginal effect. Histograms and rug plots display the distribution of continuous and categorical predictor variables, respectively, across the 183 sampled birds.

Figure 5. Distribution of the mean predicted probabilities of *Bb*sl competence across the 39

sampled avian families. (A) Density plots show mean predicted probabilities for currently

negative, positive, and unsampled species, and these propensity scores are also shown across the

avian phylogeny (B). The geographic distributions of species within a mean probability of over

50% are shown across by their breeding (C), non-breeding (D), and resident ranges (E).

Shapefiles are from BirdLife International and the Handbook of the Birds of the World [70].

References

1. Jones KE, Patel NG, Levy MA, Storeygard A, Balk D, Gittleman JL, Daszak P. 2008 Global trends in emerging infectious diseases. *Nature* **451**, 990–993. (doi:10.1038/nature06536)

2. Morse SS, Mazet JA, Woolhouse M, Parrish CR, Carroll D, Karesh WB, Zambrana-Torrelio C, Lipkin WI, Daszak P. 2012 Prediction and prevention of the next pandemic zoonosis. *The Lancet* **380**, 1956–1965. (doi:10.1016/S0140-6736(12)61684-5)

3. Babayan SA, Orton RJ, Streicker DG. 2018 Predicting reservoir hosts and arthropod vectors from evolutionary signatures in RNA virus genomes. *Science* **362**, 577–580. (doi:10.1126/science.aap9072)

4. Carroll D, Daszak P, Wolfe ND, Gao GF, Morel CM, Morzaria S, Pablos-Méndez A, Tomori O, Mazet JA. 2018 The global virome project. *Science* **359**, 872–874.

5. Becker DJ, Washburne AD, Faust CL, Mordecai EA, Plowright RK. 2019 The problem of scale in the prediction and management of pathogen spillover. *Philos. Trans. R. Soc. B Biol. Sci.* **374**, 20190224. (doi:10.1098/rstb.2019.0224)

6. Han BA, Drake JM. 2016 Future directions in analytics for infectious disease intelligence. *EMBO Rep.* , e201642534. (doi:10.15252/embr.201642534)

7. Plowright RK, Becker DJ, Crowley DE, Washburne AD, Huang T, Nameer PO, Gurley ES, Han BA. 2019 Prioritizing surveillance of Nipah virus in India. *PLoS Negl. Trop. Dis.* **13**, e0007393. (doi:10.1371/journal.pntd.0007393)

8. Han BA, Schmidt JP, Alexander LW, Bowden SE, Hayman DT, Drake JM. 2016 Undiscovered bat hosts of filoviruses. *PLOS Negl Trop Dis* **10**, e0004815.

9. Han BA, Schmidt JP, Bowden SE, Drake JM. 2015 Rodent reservoirs of future zoonotic diseases. *Proc. Natl. Acad. Sci.* **112**, 7039–7044.

10. Han BA *et al.* 2019 Confronting data sparsity to identify potential sources of Zika virus spillover infection among primates. *Epidemics* (doi:10.1016/j.epidem.2019.01.005)

11. Gervasi SS, Civitello DJ, Kilvitis HJ, Martin LB. 2015 The context of host competence: a role for plasticity in host–parasite dynamics. *Trends Parasitol.* **31**, 419–425.

12. Martin LB, Burgan SC, Adelman JS, Gervasi SS. 2016 Host Competence: An Organismal Trait to Integrate Immunology and Epidemiology. *Integr. Comp. Biol.* **56**, 1225–1237. (doi:10.1093/icb/icw064)

13. Barron D, Gervasi S, Pruitt J, Martin L. 2015 Behavioral competence: how host behaviors can interact to influence parasite transmission risk. *Curr. Opin. Behav. Sci.* **6**, 35–40. (doi:10.1016/j.cobeha.2015.08.002)

14. VanderWaal KL, Ezenwa VO. 2016 Heterogeneity in pathogen transmission: mechanisms
and methodology. *Funct. Ecol.* **30**, 1606–1622.

15. Paull SH, Song S, McClure KM, Sackett LC, Kilpatrick AM, Johnson PT. 2011 From
superspreaders to disease hotspots: linking transmission across hosts and space. *Front. Ecol.*
Environ. **10**, 75–82. (doi:10.1890/110111)

16. Kilpatrick AM, Daszak P, Jones MJ, Marra PP, Kramer LD. 2006 Host heterogeneity
dominates West Nile virus transmission. *Proc. R. Soc. Lond. B Biol. Sci.* **273**, 2327–2333.

17. Streicker DG, Fenton A, Pedersen AB. 2013 Differential sources of host species
heterogeneity influence the transmission and control of multihost parasites. *Ecol. Lett.* **16**,
975–984.

18. Downs CJ, Schoenle LA, Han BA, Harrison JF, Martin LB. 2019 Scaling of Host
Competence. *Trends Parasitol.* **0**. (doi:10.1016/j.pt.2018.12.002)

19. Han BA, Kramer AM, Drake JM. 2016 Global Patterns of Zoonotic Disease in Mammals.
Trends Parasitol. **32**, 565–577.

20. Archie EA, Luikart G, Ezenwa VO. 2009 Infecting epidemiology with genetics: a new
frontier in disease ecology. *Trends Ecol. Evol.* **24**, 21–30. (doi:10.1016/j.tree.2008.08.008)

21. Richter D, Debski A, Hubalek Z, Matuschka F-R. 2011 Absence of Lyme Disease
Spirochetes in Larval Ixodes ricinus Ticks. *Vector-Borne Zoonotic Dis.* **12**, 21–27.
(doi:10.1089/vbz.2011.0668)

22. Brunner JL, LoGiudice K, Ostfeld RS. 2008 Estimating Reservoir Competence of Borrelia
burgdorferi Hosts: Prevalence and Infectivity, Sensitivity, and Specificity. *J. Med. Entomol.*
45, 139–147. (doi:10.1093/jmedent/45.1.139)

23. Kurtenbach K, Hanincová K, Tsao JI, Margos G, Fish D, Ogden NH. 2006 Fundamental
processes in the evolutionary ecology of Lyme borreliosis. *Nat. Rev. Microbiol.* **4**, 660.

24. Ivanova LB *et al.* 2014 Borrelia chilensis, a new member of the Borrelia burgdorferi sensu
lato complex that extends the range of this genospecies in the Southern Hemisphere.
Environ. Microbiol. **16**, 1069–1080. (doi:10.1111/1462-2920.12310)

25. Schwartz AM. 2017 Surveillance for Lyme Disease — United States, 2008–2015. *MMWR*
Surveill. Summ. **66**. (doi:10.15585/mmwr.ss6622a1)

26. LoGiudice K, Ostfeld RS, Schmidt KA, Keesing F. 2003 The ecology of infectious disease:
Effects of host diversity and community composition on Lyme disease risk. *Proc. Natl.*
Acad. Sci. **100**, 567–571. (doi:10.1073/pnas.0233733100)

27. Ostfeld RS, Levi T, Jolles AE, Martin LB, Hosseini PR, Keesing F. 2014 Life History and
Demographic Drivers of Reservoir Competence for Three Tick-Borne Zoonotic Pathogens.
PLOS ONE **9**, e107387. (doi:10.1371/journal.pone.0107387)

28. Huang ZYX, de Boer WF, van Langevelde F, Olson V, Blackburn TM, Prins HHT. 2013
Species' Life-History Traits Explain Interspecific Variation in Reservoir Competence: A
Possible Mechanism Underlying the Dilution Effect. *PLoS ONE* **8**, e54341.
(doi:10.1371/journal.pone.0054341)

29. Mather TN, Nicholson MC, Donnelly EF, Matyas BT. 1996 Entomologic index for human
risk of Lyme disease. *Am. J. Epidemiol.* **144**, 1066–1069.

30. Ostfeld RS, Canham CD, Oggenfuss K, Winchcombe RJ, Keesing F. 2006 Climate, Deer,
Rodents, and Acorns as Determinants of Variation in Lyme-Disease Risk. *PLoS Biol.* **4**,
e145. (doi:10.1371/journal.pbio.0040145)

31. Smith Jr RP, Rand PW, Lacombe EH, Morris SR, Holmes DW, Caporale DA. 1996 Role of
bird migration in the long-distance dispersal of Ixodes dammini, the vector of Lyme disease.
J. Infect. Dis. **174**, 221–224.

32. Dubska L, Literak I, Kocianova E, Taragelova V, Sychra O. 2009 Differential role of
passerine birds in distribution of Borrelia spirochetes, based on data from ticks collected
from birds during the postbreeding migration period in Central Europe. *Appl Env. Microbiol*
75, 596–602.

33. Ishiguro F, Takada N, Masuzawa T. 2005 Molecular evidence of the dispersal of Lyme
disease Borrelia from the Asian continent to Japan via migratory birds. *Jpn. J. Infect. Dis.*
58, 184.

34. Norte AC *et al.* 2020 Host dispersal shapes the population structure of a tick-borne bacterial
pathogen. *Mol. Ecol.* **29**, 485–501. (doi:10.1111/mec.15336)

35. Ogden NH *et al.* 2008 Role of Migratory Birds in Introduction and Range Expansion of
Ixodes scapularis Ticks and of Borrelia burgdorferi and Anaplasma phagocytophilum in
Canada. *Appl. Environ. Microbiol.* **74**, 1780–1790. (doi:10.1128/AEM.01982-07)

36. Gylfe A, Bergström S, Lundström J, Olsen B. 2000 Reactivation of Borrelia infection in
birds. *Nature* **403**, 724–725. (doi:10.1038/35001663)

37. Brinkerhoff RJ, Folsom-O'Keefe CM, Tsao K, Diuk-Wasser MA. 2011 Do birds affect
Lyme disease risk? Range expansion of the vector-borne pathogen Borrelia burgdorferi.
Front. Ecol. Environ. **9**, 103–110. (doi:10.1890/090062)

38. Mysterud A, Heylen DJ, Matthysen E, Garcia AL, Jore S, Viljugrein H. 2019 Lyme
neuroborreliosis and bird populations in northern Europe. *Proc. R. Soc. B* **286**, 20190759.

39. Comstedt P, Bergström S, Olsen B, Garpmo U, Marjavaara L, Mejlon H, Barbour AG,
Bunikis J. 2006 Migratory Passerine Birds as Reservoirs of Lyme Borreliosis in Europe.
Emerg. Infect. Dis. **12**, 1087–1095. (doi:10.3201/eid1207.060127)

40. Hanincová K, Taragelová V, Koci J, Schäfer SM, Hails R, Ullmann AJ, Piesman J, Labuda
M, Kurtenbach K. 2003 Association of Borrelia garinii and B. valaisiana with songbirds in
Slovakia. *Appl. Environ. Microbiol.* **69**, 2825–2830.

41. Franke J, Moldenhauer A, Hildebrandt A, Dorn W. 2010 Are birds reservoir hosts for
Borrelia afzelii? *Ticks Tick-Borne Dis.* **1**, 109–112. (doi:10.1016/j.ttbdis.2010.03.001)

42. Richter D, Spielman A, Komar N, Matuschka FR. 2000 Competence of American robins as
reservoir hosts for Lyme disease spirochetes. *Emerg. Infect. Dis.* **6**, 133–138.

43. Ginsberg HS, Buckley PA, Balmforth MG, Zhioua E, Mitra S, Buckley FG. 2005 Reservoir
Competence of Native North American Birds for the Lyme Disease Spirochete, Borrelia
burgdorferi. *J. Med. Entomol.* **42**, 445–449. (doi:10.1603/0022-
2585(2005)042[0445:RCONNA]2.0.CO;2)

44. Mather TN, Telford SR, MacLachlan AB, Spielman A. 1989 Incompetence of Catbirds as
Reservoirs for the Lyme Disease Spirochete (Borrelia burgdorferi). *J. Parasitol.* **75**, 66–69.
(doi:10.2307/3282938)

45. Anderson JF, Johnson RC, Magnarelli LA, Hyde FW. 1986 Involvement of birds in the
epidemiology of the Lyme disease agent Borrelia burgdorferi. *Infect. Immun.* **51**, 394–396.

46. Wright SA, Thompson MA, Miller MJ, Knerl KM, Elms SL, Karpowicz JC, Young JF,
Kramer VL. 2000 Ecology of Borrelia burgdorferi in ticks (Acari: Ixodidae), rodents, and
birds in the Sierra Nevada foothills, Placer County, California. *J. Med. Entomol.* **37**, 909–
918.

47. Loss SR, Noden BH, Hamer GL, Hamer SA. 2016 A quantitative synthesis of the role of
birds in carrying ticks and tick-borne pathogens in North America. *Oecologia* **182**, 947–959.
(doi:10.1007/s00442-016-3731-1)

48. Battaly GR, Fish D. 1993 Relative importance of bird species as hosts for immature Ixodes
dammini (Acari: Ixodidae) in a suburban residential landscape of southern New York State.
J. Med. Entomol. **30**, 740–747.

49. Hamer SA, Goldberg TL, Kitron UD, Brawn JD, Anderson TK, Loss SR, Walker ED, Hamer
GL. 2012 Wild Birds and Urban Ecology of Ticks and Tick-borne Pathogens, Chicago,
Illinois, USA, 2005–2010. *Emerg. Infect. Dis.* **18**, 1589–1595. (doi:10.3201/eid1810.120511)

50. Mead P, Hook S, Niesobecki S, Ray J, Meek J, Delorey M, Prue C, Hinckley A. 2018 Risk
factors for tick exposure in suburban settings in the Northeastern United States. *Ticks Tick-
Borne Dis.* **9**, 319–324. (doi:10.1016/j.ttbdis.2017.11.006)

51. Hochachka WM, Caruana R, Fink D, Munson ART, Riedewald M, Sorokina D, Kelling S.
2007 Data-Mining Discovery of Pattern and Process in Ecological Systems. *J. Wildl. Manag.*
71, 2427–2437.

52. Yang LH, Han BA. 2018 Data-driven predictions and novel hypotheses about zoonotic tick
vectors from the genus Ixodes. *BMC Ecol.* **18**, 7. (doi:10.1186/s12898-018-0163-2)

53. Moher D, Liberati A, Tetzlaff J, Altman DG, Group P. 2009 Preferred reporting items for
systematic reviews and meta-analyses: the PRISMA statement. *PLoS Med.* **6**, e1000097.

54. Breuner NE *et al.* 2020 Failure of the Asian longhorned tick, Haemaphysalis longicornis, to
serve as an experimental vector of the Lyme disease spirochete, Borrelia burgdorferi sensu
stricto. *Ticks Tick-Borne Dis.* **11**, 101311. (doi:10.1016/j.ttbdis.2019.101311)

55. Michonneau F, Brown JW, Winter DJ. 2016 rotl : an R package to interact with the Open
Tree of Life data. *Methods Ecol. Evol.* (doi:10.1111/2041-210X.12593)

56. Paradis E, Claude J, Strimmer K. 2004 APE: analyses of phylogenetics and evolution in R
language. *Bioinformatics* **20**, 289–290.

57. Viechtbauer W. 2010 Conducting meta-analyses in R with the metafor package. *J. Stat.*
Softw. **36**, 1–48.

58. Nakagawa S, Santos ES. 2012 Methodological issues and advances in biological meta-
analysis. *Evol. Ecol.* **26**, 1253–1274.

59. Burnham KP, Anderson DR. 2002 *Model selection and multimodel inference: a practical*
information-theoretic approach. Springer Science & Business Media.

60. López-López JA, Marín-Martínez F, Sánchez-Meca J, Noortgate W, Viechtbauer W. 2014
Estimation of the predictive power of the model in mixed-effects meta-regression: A
simulation study. *Br. J. Math. Stat. Psychol.* **67**, 30–48.

61. Fritz SA, Purvis A. 2010 Selectivity in mammalian extinction risk and threat types: a new
measure of phylogenetic signal strength in binary traits. *Conserv. Biol.* **24**, 1042–1051.

62. Chamberlain SA, Szöcs E. 2013 taxize: taxonomic search and retrieval in R. *F1000Research*
2.

63. Washburne AD, Silverman JD, Morton JT, Becker DJ, Crowley D, Mukherjee S, David LA,
Plowright RK. 2019 Phylofactorization: a graph partitioning algorithm to identify
phylogenetic scales of ecological data. *Ecol. Monogr.* **0**, e01353. (doi:10.1002/ecm.1353)

64. Schluter D, Price T, Mooers AØ, Ludwig D. 1997 Likelihood of ancestor states in adaptive
radiation. *Evolution* **51**, 1699–1711.

65. Revell LJ. 2012 phytools: an R package for phylogenetic comparative biology (and other
things). *Methods Ecol. Evol.* **3**, 217–223.

66. Wilman H, Belmaker J, Simpson J, de la Rosa C, Rivadeneira MM, Jetz W. 2014 EltonTraits
1.0: Species-level foraging attributes of the world's birds and mammals. *Ecology* **95**, 2027–
2027. (doi:10.1890/13-1917.1)

67. Myhrvold NP, Baldridge E, Chan B, Sivam D, Freeman DL, Ernest SKM. 2015 An amniote
life-history database to perform comparative analyses with birds, mammals, and reptiles.
Ecology **96**, 3109–3109. (doi:10.1890/15-0846R.1)

68. Baillie J, Hilton-Taylor C, Stuart SN. 2004 *IUCN red list of threatened species: a global
species assessment*. IUCN.

69. Vitousek MN *et al.* 2018 HormoneBase, a population-level database of steroid hormone
levels across vertebrates. *Sci. Data* **5**, 180097.

70. BirdLife International, NatureServe. 2014 Bird species distribution maps of the world.
Camb. UK BirdLife Int.

71. Gilroy JJ, Gill JA, Butchart SH, Jones VR, Franco AM. 2016 Migratory diversity predicts
population declines in birds. *Ecol. Lett.* **19**, 308–317.

72. Kembel SW, Cowan PD, Helmus MR, Cornwell WK, Morlon H, Ackerly DD, Blomberg SP,
Webb CO. 2010 Picante: R tools for integrating phylogenies and ecology. *Bioinformatics* **26**,
1463–1464.

73. Elith J, Leathwick JR, Hastie T. 2008 A working guide to boosted regression trees. *J. Anim.
Ecol.* **77**, 802–813.

74. Ridgeway G. 2006 gbm: Generalized boosted regression models. *R Package Version* **1**.

75. Sing T, Sander O, Beerenwinkel N, Lengauer T. 2005 ROCR: visualizing classifier
performance in R. *Bioinformatics* **21**, 3940–3941. (doi:10.1093/bioinformatics/bti623)

76. Evans MV, Dallas TA, Han BA, Murdock CC, Drake JM. 2017 Data-driven identification of
potential Zika virus vectors. *elife* **6**, e22053.

77. Orme D. 2013 The caper package: comparative analysis of phylogenetics and evolution in R.
R Package Version **5**.

78. Sonenshine DE, Ratzlaff RE, Troyer J, Demmerle S, Demmerle ER, Austin WE, Tan S,
Annis BA, Jenkins S. 1995 Borrelia burgdorferi in Eastern Virginia: Comparison between a
Coastal and Inland Locality. *Am. J. Trop. Med. Hyg.* **53**, 123–133.
(doi:10.4269/ajtmh.1995.53.123)

79. Kinsey AA, Durden LA, Jr JHO. 2000 TICK INFESTATIONS OF BIRDS IN COASTAL
GEORGIA AND ALABAMA. *J. Parasitol.* **86**, 251–254. (doi:10.1645/0022-
3395(2000)086[0251:TIOBIC]2.0.CO;2)

80. Schneider SC, Parker CM, Miller JR, Page Fredericks L, Allan BF. 2015 Assessing the
Contribution of Songbirds to the Movement of Ticks and Borrelia burgdorferi in the
Midwestern United States During Fall Migration. *EcoHealth* **12**, 164–173.
(doi:10.1007/s10393-014-0982-3)

81. Anderson JF, Magnarelli LA. 1984 Avian and mammalian hosts for spirochete-infected ticks
and insects in a Lyme disease focus in Connecticut. *Yale J. Biol. Med.* **57**, 627–641.

82. Hamer SA, Tsao JI, Walker ED, Hickling GJ. 2010 Invasion of the Lyme Disease Vector
Ixodes scapularis: Implications for Borrelia burgdorferi Endemicity. *EcoHealth* **7**, 47–63.
(doi:10.1007/s10393-010-0287-0)

83. Hasle G, Bjune GA, Midthjell L, Røed KH, Leinaas HP. 2011 Transport of Ixodes ricinus
infected with Borrelia species to Norway by northward-migrating passerine birds. *Ticks Tick-*
Borne Dis. **2**, 37–43.

84. Kjelland V, Stuen S, Skarpaas T, Slettan A. 2010 Borrelia burgdorferi sensu lato in Ixodes
ricinus ticks collected from migratory birds in Southern Norway. *Acta Vet. Scand.* **52**, 59.
(doi:10.1186/1751-0147-52-59)

85. Ishiguro F, Takada N, Masuzawa T, Fukui T. 2000 Prevalence of Lyme Disease Borrelia
spp. in Ticks from Migratory Birds on the Japanese Mainland. *Appl. Environ. Microbiol.* **66**,
982–986. (doi:10.1128/AEM.66.3.982-986.2000)

86. Pascucci I *et al.* 2019 Assessing the role of migratory birds in the introduction of ticks and
tick-borne pathogens from African countries: An Italian experience. *Ticks Tick-Borne Dis.*
10, 101272. (doi:10.1016/j.ttbdis.2019.101272)

87. Norte AC, Carvalho IL de, Núncio MS, Ramos JA, Gern L. 2013 Blackbirds Turdus merula
as competent reservoirs for Borrelia turdi and Borrelia valaisiana in Portugal: evidence from
a xenodiagnostic experiment. *Environ. Microbiol. Rep.* **5**, 604–607. (doi:10.1111/1758-
2229.12058)

88. Nylander JAA, Olsson U, Alström P, Sanmartín I. 2008 Accounting for Phylogenetic
Uncertainty in Biogeography: A Bayesian Approach to Dispersal-Vicariance Analysis of the
Thrushes (Aves: Turdus). *Syst. Biol.* **57**, 257–268. (doi:10.1080/10635150802044003)

89. Ashmole MJ. 1962 The migration of European thrushes: a comparative study based on
ringing recoveries. *Ibis* **104**, 522–559.

90. Previtali MA, Ostfeld RS, Keesing F, Jolles AE, Hanselmann R, Martin LB. 2012
Relationship between pace of life and immune responses in wild rodents. *Oikos* **121**, 1483–
1492. (doi:10.1111/j.1600-0706.2012.020215.x)

91. Kurtenbach K, Dizij A, Seitz HM, Margos G, Moter SE, Kramer MD, Wallich R, Schaible
UE, Simon MM. 1994 Differential immune responses to Borrelia burgdorferi in European
wild rodent species influence spirochete transmission to Ixodes ricinus L.(Acari: Ixodidae).
Infect. Immun. **62**, 5344–5352.

92. Lee KA, Wikelski M, Robinson WD, Robinson TR, Klasing KC. 2008 Constitutive immune
defences correlate with life-history variables in tropical birds. *J. Anim. Ecol.* **77**, 356–363.

93. Sapolsky RM, Romero LM, Munck AU. 2000 How Do Glucocorticoids Influence Stress
Responses? Integrating Permissive, Suppressive, Stimulatory, and Preparative Actions.
Endocr. Rev. **21**, 55–89. (doi:10.1210/edrv.21.1.0389)

94. Gervasi SS, Burgan SC, Hofmeister E, Unnasch TR, Martin LB. 2017 Stress hormones
predict a host superspreader phenotype in the West Nile virus system. *Proc R Soc B* **284**,
20171090. (doi:10.1098/rspb.2017.1090)

95. Wingfield JC, Sapolsky RM. 2003 Reproduction and Resistance to Stress: When and How. *J.*
Neuroendocrinol. **15**, 711–724. (doi:10.1046/j.1365-2826.2003.01033.x)

96. Ardia DR. 2007 The ability to mount multiple immune responses simultaneously varies
across the range of the tree swallow. *Ecography* **30**, 23–30. (doi:10.1111/j.0906-
7590.2007.04939.x)

97. Becker DJ, Albery GF, Kessler MK, Lunn TJ, Falvo CA, Czirják GÁ, Martin LB, Plowright
RK. 2019 Macroimmunology: the drivers and consequences of spatial patterns in wildlife
immune defense. *J. Anim. Ecol.* (doi:10.1111/1365-2656.13166)

98. Owen JC, Moore FR. 2008 Swainson's thrushes in migratory disposition exhibit reduced
immune function. *J. Ethol.* **26**, 383–388. (doi:10.1007/s10164-008-0092-1)

99. Marra PP, Cohen EB, Loss SR, Rutter JE, Tonra CM. 2015 A call for full annual cycle
research in animal ecology. *Biol. Lett.* **11**, 20150552.

100.   Plowright RK, Peel AJ, Streicker DG, Gilbert AT, McCallum H, Wood J, Baker ML,
Restif O. 2016 Transmission or Within-Host Dynamics Driving Pulses of Zoonotic Viruses
in Reservoir–Host Populations. *PLoS Negl. Trop. Dis.* **10**, e0004796.
(doi:10.1371/journal.pntd.0004796)

101.   Hall RJ, Altizer S, Bartel RA. 2014 Greater migratory propensity in hosts lowers
pathogen transmission and impacts. *J. Anim. Ecol.* **83**, 1068–1077. (doi:10.1111/1365-
2656.12204)

102.   Miller MJ, Esser HJ, Loaiza JR, Herre EA, Aguilar C, Quintero D, Alvarez E,
Bermingham E. 2016 Molecular Ecological Insights into Neotropical Bird–Tick Interactions.
PLoS ONE **11**. (doi:10.1371/journal.pone.0155989)

103.   Saracho Bottero MN, Sebastian PS, Carvalho LA, Claps LG, Mastropaolo M, Mangold
AJ, Venzal JM, Nava S. 2017 Presence of Borrelia in different populations of Ixodes
pararicinus from northwestern Argentina. *Ticks Tick-Borne Dis.* **8**, 488–493.
(doi:10.1016/j.ttbdis.2017.02.008)

104.   Gasparini J, McCoy KD, Haussy C, Tveraa T, Boulinier T. 2001 Induced maternal
response to the Lyme disease spirochaete Borrelia burgdorferi sensu lato in a colonial
seabird, the kittiwake Rissa tridactyla. *Proc. R. Soc. Lond. B Biol. Sci.* **268**, 647–650.
(doi:10.1098/rspb.2000.1411)

105.   Plowright RK, Becker DJ, McCallum H, Manlove KR. 2019 Sampling to elucidate the
778   dynamics of infections in reservoir hosts. *Philos. Trans. R. Soc. B*
779   (doi:10.1098/rstb.2018.0336)

780   106.   Hitch AT, Leberg PL. 2007 Breeding Distributions of North American Bird Species
781   Moving North as a Result of Climate Change. *Conserv. Biol.* **21**, 534–539.
782   (doi:10.1111/j.1523-1739.2006.00609.x)

783   107.   Ostfeld RS, Brunner JL. 2015 Climate change and Ixodes tick-borne diseases of humans.
784   *Philos. Trans. R. Soc. B Biol. Sci.* **370**, 20140051.

785   108.   Vliet JV, Musters CJM, Keurs WJT. 2009 Changes in migration behaviour of Blackbirds
786   Turdus merula from the Netherlands. *Bird Study* **56**, 276–281.
787   (doi:10.1080/00063650902792148)

788   109.   Plummer KE, Siriwardena GM, Conway GJ, Risely K, Toms MP. 2015 Is supplementary
789   feeding in gardens a driver of evolutionary change in a migratory bird species? *Glob.*
790   *Change Biol.* **21**, 4353–4363. (doi:10.1111/gcb.13070)

791   110.   Bonnet-Lebrun A-S, Manica A, Rodrigues AS. 2020 Effects of urbanization on bird
792   migration. *Biol. Conserv.* , 108423.

793   111.   Yeh PJ, Price TD. 2004 Adaptive phenotypic plasticity and the successful colonization of
794   a novel environment. *Am. Nat.* **164**, 531–542.

795   112.   Becker DJ, Schultz EM, Atwell JW, Ketterson ED. 2019 Urban residency and leukocyte
796   profiles in a traditionally migratory songbird. *Anim. Migr.* **6**, 49–59. (doi:10.1515/ami-2019-
797   0002)

798   113.   Kernbach ME *et al.* 2019 Light pollution increases West Nile virus competence of a
799   ubiquitous passerine reservoir species. *Proc. R. Soc. B* **286**, 20191051.

800   114.   Becker DJ, Han BA. 2020 Data from: The macroecology and evolution of avian
801   competence for Borrelia burgdorferi. (doi:10.5061/dryad.r7sqv9s88)