Cluster Analysis of Medicinal Plants and Targets Based on Multipartite Network

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Abstract

Although medicinal plants have played an important role in the development of therapeutics as
natural compound sources, the synergistic effects of the multiple chemicals in the plants on multiple
targets have not been studied widely. Recently, network-based methods for the analysis of drug-
target interactions have gained attention and relies on the paradigm that a single drug can act on
multiple targets rather than a single target. In this study, we have presented a novel approach to
analyze the interactions between the chemicals in the medicinal plants and multiple targets based on
the complex multipartite network of the medicinal plants, multi-chemicals, and multiple targets. The
multipartite network was constructed via the conjunction of two relationships: chemicals in plants
and the biological actions of those chemicals on the targets. By analyzing the structural
characteristics of the network using the network analysis techniques, we investigated the similarities
among the plants via their chemical profiles obtained from the hierarchical clustering analysis. In
doing so, we introduced an index of the efficacy of chemicals in a plant on a protein target of interest,
called target potency score (TPS). We showed that the analysis can identify specific chemical profiles
from each group of plants, which can then be employed for discovering new alternative therapeutic
agents. Furthermore, specific clusters of plants and chemicals acting on specific targets were
retrieved using TPS that suggested potential drug candidates with high probability of clinical success.

We expect that this approach may open a way to predict the biological functions of multi-chemicals
and multi-plants on the targets of interest and enable repositioning of the plants and chemicals.

1   Introduction

Plants have been used as therapeutic agents for a very long time in the human history (Petrovska,
2012). Single bioactive compounds in plants have been developed as well-known commercial drugs,
such as paclitaxel, vinblastine, morphine, and artemisinin (Koehn and Carter, 2005; Harvey et al.,
2015). Occasionally, single components from natural products have provided chemical scaffolds for
developing more potent semi-synthetic drugs, such as rosuvastatin, tramadol, and eribulin (Rodrigues
et al., 2016). Similar to these single compound drugs that act on a specific target, a mixture of
components from the whole plants or their sub-fractions have been widely used as botanical drugs,
such as Veregen® and Fulyzaq® (Galanakis, 2019). However, the synergistic effects of multi-
chemicals in plants on multiple targets and the underlying complex biological pathways involved are
less understood.

For the purpose of developing drugs composed of multi-chemicals, the identification of the
interactions between the multi-chemicals and multi-targets is important to understand the therapeutic
effects as well as the side effects, i.e., the on-target and off-target effects (Roth et al., 2004; Keiser et
al., 2009; Besnard et al., 2012; Lounkine et al., 2012; Anighoro et al., 2014; Zhang et al., 2015, 2017;
Ramsay et al., 2018). Identifying the synergistic drug combinations is also critical to develop
effective therapies against cancers and to improve the therapeutic efficacy (Jia et al., 2009; NCI-
DREAM Community et al., 2014; Sun et al., 2015; Han et al., 2017). However, the complexity of
screening all possible candidate combinations of multi-chemicals and multi-targets prevents the use
of the classical way of identifying the interactions via biological experiments, using *in vitro* and *in*
vivo assays. Since a medicinal plant produces a large number of chemical compounds, the target
identification of every single chemical is impossible for the development of botanical drugs,
especially to acquire approval in the global market requiring well-controlled clinical studies (Ahn,
2017).

In recent decades, computational approaches for the systematic analysis of drug-target interactions
have become available, improving efficiency and reducing costs of studying the interactions (Zheng
et al., 2013; Lavecchia and Cerchia, 2016; Luo et al., 2017; Zhang et al., 2017; Ezzat et al., 2019;
Forouzesh et al., 2019). Among the computational approaches, network-based approaches provide
alternative methods to deal with the complex molecular interactions (Hopkins, 2008; 2019; Monteiro
et al., 2020). Compared with other computational methods, network-based approaches do not require
three-dimensional structures of the targets or negative control samples (Cheng et al., 2019). These
approaches have also been proposed for botanical drug design in search of optimal combinations of
medicinal plant components and prediction of multiple target activities (Leung et al., 2013).
However, studies based on the integrated networks of medicinal plants, chemicals, and multi-targets
are still rare. Most studies have focused only on drug-target networks (Yıldırım et al., 2007) or the
molecular networks of traditional Chinese medicinal plants (Zhao et al., 2010).

In this study, we introduced an approach based on the integrated network of medicinal plants,
chemical components, and multi-targets to analyze the complex biological roles of medicinal plants
in human body. First, we constructed an integrated multipartite network of the biomedical entities
including Korean medicinal plants, chemicals, and protein targets from heterogeneous databases. The
databases have been curated for biomedical entities, such as genes, proteins, chemicals, and species
and are publicly accessible. In particular, we employed the ChEMBL database, which is one of the

largest public databases of the activities of small molecules on biological targets (Koutsoukas et al., 2011; Martínez-Jiménez et al., 2013; Davies et al., 2015; Gaulton et al., 2017; Bosc et al., 2019). It also provides the built-in, ligand-based target prediction methods that we used to generate the links between chemicals with targets in the interaction network. We also used the database on compound ingredients extracted from Korean medicinal plants, which was released in 2007 by the Korean Traditional Knowledge Portal (http://www.koreantk.com) (Shin et al., 2010).

We then preprocessed the multipartite network and adapted the spectral co-clustering method to identify the subnetworks of closely related biomedical entities within their own structural properties, such as the average bipartite clustering coefficient (Dhillon, 2001). The clustering results could be used for the single or combination use of Korean medicinal plants in common target domains. The hierarchical clustering analysis was followed for further identification of the plant clusters based on the similarities in their chemical composition. In the analysis, we introduced a network-based mathematical index of interactions between the medicinal plants and targets, called the target potency score (TPS). The suggested index is used for ranking the plant clusters or protein targets, and for retrieving the best subnetworks of the medicinal plants, multi-chemicals, and multi-targets customized to specific medicinal purposes. Finally, we have demonstrated the utility of the simplified multipartite network with the hierarchical clusters.

2   Materials and Methods

2.1   Raw Data Sets

We employed several heterogeneous data sets to construct a multipartite network of medicinal plants, chemicals, and targets. The data set of the medicinal plants was obtained from the Korean Traditional Knowledge Portal (KTKP, http://www.koreantk.com). The database was built by the Korean Intellectual Property Office, Ministry of Trade, Industry and Energy, Republic of Korea (Shin, Jin-Sup et al., 2010). The KTKP data sets contain a list of 5,500 Korean medicinal plants and 10,056 chemicals, and their relations. The data set of chemicals and targets was obtained from the ChEMBL database (ChEMBL25, March 2019, http://ftp.ebi.ac.uk/pub/databases/chembl/target_predictions). The ChEMBL25 database contains 1,879,206 compounds, 12,482 targets, and 15,504,603 bioactivity entries from 72,271 publications. All the chemicals in the KTKP data sets were identified in the ChEMBL25 database, and the links between the chemicals and single-protein targets were generated using the prediction model reported previously (Koutsoukas et al., 2011; Martínez-Jiménez et al., 2013), and is available on the ChEMBL25 database. Specifically, the probability of the activation of a compound against a target was estimated by the 10-uM activity cut-off model in the ChEMBL25 database, and we determined the interaction between a pair of a chemical and a target at the probability of >0.9.

2.2   Preprocessing

We excluded non-plants from the raw dataset. For instance, minerals (non-living things) and animals (including corals) were removed, but the fungi were not. After excluding the non-plants, the raw data sets containing 5,500 instances of plants and non-plants were reduced to a set containing 3,614 plants. Then, the duplicated instances were removed based on their identical "scientific name," "Latin name" and "use target", which led to a remaining list of 2,886 plants in the data set.

The multipartite network of the medicinal plants, chemicals, and targets was constructed by combining the two bipartite networks: the plants-chemicals network and chemicals-targets network (**Figure 1A**). The nodes in the network represented the plants, chemicals, and targets, and the edges

in the network represented the relationships between the plants and chemicals, and between the
chemicals and targets. From the multipartite network, we removed the nodes of degree zero, i.e., the
plants, chemicals, or targets that were not linked to any other entities. After all, 1,138 plants, 10,043
chemicals, and 441 targets remained for the combined multipartite network. The basic statistics of the
preprocessed network data are summarized in **Table 1**.

2.3   Methods

We analyzed the multipartite network using the spectral co-clustering algorithm (Dhillon, 2001). The
algorithm clusters the medicinal plants, chemicals, and targets by solving a network partitioning
problem. It partitions the set V of the connected nodes into k subsets, $V_1, V_2, ..., V_k$ in such a way that
the k subsets approximately minimize the normalized cut criterion:

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$$Q(V_1, V_2, ..., V_k) = \frac{1}{k} \sum_{i=1}^{k} \frac{cut(V_i, \bar{V_i})}{vol(V_i)}$$

where $\bar{V_i} = V \backslash V_i$ is the complement of V_i, the $cut(M, N) = \sum_{m \in M, n \in N} w_{mn}$ of two disjoint sets M and
N is defined by the sum of the edge weights, w_{mn}, between two nodes from each of M and N, and
$vol(V_i) = \sum_{m \in V_i} d_m = \sum_{m \in V_i} \sum_n w_{mn}$ is the sum of degrees of all nodes in V_i. The normalized cut
criterion represents the level of connectivity between the k subsets, where a smaller value indicates a
weaker connectivity between them.

To determine the number of clusters, k, we computed the modularity, defined by

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$$Modularity(k) = \frac{1}{2E} \sum_{i=1}^{k} \sum_{m \in V_i, n \in V_i} (w_{mn} - d_m d_n / 2E)$$

where E is the number of edges and dm is the degree of a node m (Newman, 2004, 2006; Newman
and Girvan, 2004). The modularity measures the strength of a partition with respect to the
distribution of edges because it compares the number of edges, $\sum_{m,n} w_{mn}$, with the expected number
of edges placed at random, $\sum_{m,n} d_m d_n / 2E$, within the partition. An optimal number of clusters is
determined at the maximum point of the modularity.

The hierarchical clustering analysis led to the identification of the clusters of medicinal plants based
on their chemical profiles. The goal of the analysis was to provide insight into the biologically or
chemically inter-related plants and search for their potentials medicinal effects in the integrated
network. We use the Bray-Curtis distance for the hierarchical clustering, which is defined by,
between two plants u and v,

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$$d(u, v) = \frac{\sum_j |u_j - v_j|}{\sum_j |u_j| + |v_j|}$$

where u_j (or v_j) is equal to one if the plant u (or v)is connected to the jth chemical in the network,
and zero otherwise. The Bray-Curtis distance takes a value between 0 and 1, and it has two favorable
properties for the multipartite network analysis: (1) It is a scale-free distance metric, that is, it
depends only on the fraction of the number of chemicals connected to the plants, rather than the total
number of chemicals in the network. The scale-freeness is a crucial property for analyzing sparsely

connected networks, such as the multipartite network in this study where most of the u_j (or v_j) values
are zero; see, for example, **Figure S1** in Supplemental Information for a visualization. (2) It is
suitable for the unweighted networks because it only considers the number of edges while ignoring
the edge weights.

We used the complete linkage method for hierarchical clustering, that is, the distance between two
clusters is defined by the distance between two elements that are farthest from each other. The
combined use of the Bray-Curtis distance and complete linkage yields a meaningful interpretation of
clustering results. That is, if we combine two clusters one whenever they are at a distance of less than
one, then every pair of elements will have a distance less than one. It implies that every pair of
medicinal plants in a cluster will have at least one common chemical because $d(u, v) < 1$ implies
that $u_j = v_j = 1$ for some chemical j.

Finally, we proposed an index called TPS, which is a similarity measure between a plant cluster and
a target. TPS of a plant cluster P to a target T is defined by the fraction

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$$TPS(P,T) = \frac{|C_P \cap C_T|}{|C_T|} = \frac{number\ of\ chemicals\ connected\ to\ both\ P\ and\ T}{number\ of\ chemicals\ connected\ to\ T}$$

where C_P is the set of all chemicals connected to at least one plant in the plant cluster P, C_T is the set
of all chemicals connected to the target T, and $|\cdot|$ denotes the number of elements in a set. TPS can
represent potential efficacy of a plant cluster for predicting a protein target. We used the TPS to
search for the multipartite network for plant clusters of the highest efficacy or most active targets.

3   Results

3.1   Multipartite Network Analysis

We constructed the multipartite network with the nodes of medicinal plants, chemicals, and targets
for the interpretation of the biological roles of these chemicals and plants containing them on multi-
targets. The preprocessed network is shown in **Figure 1B**, where 10% of the nodes are randomly
selected for visualization. Note that the plants and targets have connections only to the chemicals.
Figure 1C shows the distribution of the node degrees for the plants, chemicals, or targets, where the
vertical axes are on a log scale. The distribution of the power-law degree imply that the network is a
scale-free network (Goh et al., 2002; Newman, 2005; Lima-Mendez and van Helden, 2009).

The structural properties of the multipartite network were further analyzed by the network analysis
techniques. We divided the network into five subnetworks by applying the spectral co-clustering
algorithm. Then, the structural properties of the multipartite network and the five subnetworks were
computed and summarized in **Table S1** in Supplemental Information. The computed structural
properties are a standard set of theoretic graphical measures, such as the diameter, average bipartite
clustering coefficient, and degree of assortativity. **Figure S3** in Supplemental Information illustrates
the multipartite network, modularity, and five subnetworks, where only 10% of the whole nodes were
randomly selected for visualization.

3.2   Hierarchical Clustering Analysis of Plants

We performed the hierarchical clustering analysis of the medicinal plants in the multipartite network,
and generated dendrograms to visually inspect the clusters of the medicinal plants (**Figure 2** and
Figure S2 in Supplemental Information). In the two largest clusters of the plants (**Figure 2A** and **2C**,

Table S2 and **S3** in Supplemental Information), the same species with different usage parts have
been aggregated at the bottom level (i.e., at the height of zero) in the dendrogram, and different
species belonging to the same family are grouped together at low levels. However, some species
belonging to the same family are far apart or the ones belonging to a different family are closely
located. In the largest cluster, many different species sharing β-sitosterol, which is one of the most
common phytosterols and is distributed over a wide range in the plant kingdom, existed (Tao et al.,
2019); many plants belonging to different families share more chemicals than the ones belonging to
the same family (**Figure 2A**). The chemical diversity of the cluster shows that the specialized
distribution of the secondary metabolites in plants is not dependent on their family level. However, in
the second largest cluster, the chemical profiles of the plants depending on the family contributed to
the classification of the smaller clusters (**Figure 2C**). Additionally, the chemical diversity of the
plants in the second largest cluster (123 types of subclasses and 311 types of direct parent levels) was
bigger than the largest cluster (109 types of subclasses and 257 types of direct parent levels) (**Figures
2B** and **2D**). Among the smaller clusters, two clusters containing the specialized types of chemicals,
protoberberine alkaloids (**Figures 2E** and **2F**, and **Table S4** in Supplemental Information) and
vitamins (**Figure 2G** and **2H**, **Table S5** in Supplemental Information), were clearly categorized by
the family. We expect that the clustering information by the chemical profiles between the plants can
be utilized not only for the chemotaxonomic study of the medicinal plants, but also for alternative use
of the medicinal plants.

3.3   Interaction Between Plants and Targets Based on TPS

In the multipartite network, the interaction between the plants and targets is mediated by the
connectivity of the chemicals in the network. We used the TPS to quantitatively measure the
interaction that represents the potential efficacy of the plants for the targets. Because the plants in the
same cluster share chemicals in common, the TPS was calculated between a cluster of plants and
target.

The TPS was applied in two ways. First, the targets with the highest TPS values were selected for
each of the four clusters of plants that were mentioned in the previous section and were manually
investigated to validate our approaches. The subnetworks of the selected targets, chemicals connected
to the targets, and corresponding cluster of plants connected to the chemicals are illustrated in
Figures 3A-D, for the inspection of the effects of the mixture of chemicals or the plants in the
cluster. Specifically speaking, the targets with the TPS values greater than 0.05~0.5 were selected
while the number of chemicals connected to the targets was kept sufficiently large enough (at least
20) to produce reliable TPS values.

In each network, a single target had multiple edges with chemicals, but only a few chemicals
contributed to the connection between the plants and targets. In the largest cluster, six targets, signal
transducer and activator of transcription 3 (CHEMBL4026), phosphodiesterase 10A
(CHEMBL4409), anandamide amidohydrolase (CHEMBL3229), prostanoid EP2 receptor
(CHEMBL4909), sphingosine 1-phosphate receptor Edg-3 (CHEMBL3892), and pepsin A
(CHEMBL2714), are displayed (**Figure 3A**). For example, in case of phosphodiesterase 10A, which
has been studied as a therapeutic target for colon cancer and neurological symptoms (Li et al., 2015),
12 chemicals, mainly carboline-type alkaloids, are displayed with linkages to the target (**Table S6** in
Supplemental Information). The well-known targets, such as cannabinoid CB1 (CHEMBL218) and
CB2 (CHEMBL253) receptors, in the network from the second largest cluster (**Figures 2C** and **3B**)
shared the monoterpenoids and benzopyran-type chemicals including cannabinoids from the genus
Cannabis (Wu, 2019). In the third network, protoberberine-type alkaloids from three families,

Papaveraceae, Ranunculaceae, and Rutaceae, were linked with the targets, dopamine receptors
(CHEMBL217, CHEMBL265, CHEMBL1850, CHEMBL2056, and CHEMBL339), and alpha
adrenergic receptors (CHEMBL315 and CHEMBL1867), which are known as the main target of
these chemicals (Kawano et al., 2015; Fan et al., 2019). Tetraterpenoids, such as lycopene, lutein, and
β-carotene, which are the well-known chemicals targeting the retinoid signaling pathway, had
multiple linkages with retinoic acid receptor gamma (RAR-γ, CHEMBL2003) and retinoid X
receptor beta (RXR-β, CHEMBL4047) (Aydemir et al., 2012, 2013; Huang et al., 2016). These
results show that the target information predicted by the chemicals in the multipartite network is
reliable enough to investigate alternative targets of the chemicals, for which biological roles have not
been defined in previous studies or they existed as mixed forms with other chemicals in a plant.

Second, the proposed TPS was applied for searching the closest related plants, which are predictive
of a given target based on the highest TPS value. The results provide clues to understand the complex
biological roles of the combination of medicinal plants or the chemicals produced from the plants. In
this scenario, a cluster of plants is recommended from among the large-scale network as the
candidate medicinal plant for a single target of interest. For example, we used 'tyrosinase'
(CHEMBL3318) as the model target to validate the suggested method and to investigate the
medicinal effects of the retrieved cluster of plants (**Figure 4A**). In the multipartite network retrieved
for the target "tyrosinase," 165 chemicals from 31 different species were linked with the target node.
Among them, 50 chemicals from *Morus alba* were found in the middle of the network (**Table S7** in
Supplemental Information). Additionally, *Carthamus tinctorius*, *Cudrania tricuspidata*, *Glycyrrhiza*
inflata, and *Glycyrrhiza uralensis* had more than 10 chemicals, which were predicted to act on
tyrosinases (**Table S8** in Supplemental Information). These five plants have been known as the most
popular and successful sources for the whitening products used in the cosmetic industry (Smit et al.,
2009; Xin et al., 2017).

Next, we combined three subnetworks-networks from three targets, steroid 5α-reductase 1 (S5R1,
ChEMBL1787), steroid 5α-reductase 2 (S5R2, ChEMBL1856), and androgen receptor (AR,
ChEMBL1871), which are involved in the androgen metabolism, for the interaction of plants and
chemicals on multi-targets (**Figure 4B**). In total, 44 among 52 and 54 chemicals from S5R1 and
S5R2, respectively, shared with each other, but 10 of 52 from S5R1 and 7 of 54 from S5R2 only
shared those with 58 chemicals from AR (**Table S9-11** in Supplemental Information). The types of
chemicals between the pairs of targets showed significantly different patterns. The chemicals co-
targeting S5R1 and S5R2 were mainly the steroid derivatives, such as cholestanes, ergostanes, and
stigmastanes, which were synthesized via the triterpenoid pathways (**Figure 4C**); however,
sesquiterpenoids and diterpenoids were the main components sharing the pairs of S5R1 and AR, or
S5R2 and AR (**Figure 4D** and **4E**). We found that only 6 chemicals simultaneously acted on three
targets and plants containing them (**Figure 5** and **Table 2**).

4   Discussion

In this study, we tried to unravel the complex interaction between multi-chemicals and multiple
targets using the computational target prediction model and multipartite networks comprising the
relationship between the plants, chemicals, and targets. In hierarchical cluster analysis, plants
producing similar specialized metabolites are clustered only within a taxon. In particular, in
chemotaxonomy, secondary metabolites can be used as specialized markers to distinguish among
plants (Wink, 2003). However, not all secondary metabolites are produced in the same taxon. For
example, flavonoids are widely distributed in plants and even in fungi, fulfilling many functions,
such as ultraviolet filtration, symbiotic nitrogen fixation, and floral pigmentation (Kumar and

Pandey, 2013). The chemical profile of each plant has a minor effect on the classification of the
plant.

To unravel the clusters of multipartite networks from hierarchical clustering analysis, we introduced
an index of potential efficacy of plant clusters toward a target, called TPS. TPS indicated the
proximity of subnetworks of the metabolic relationships between plant clusters and targets. We
utilized it to search for and suggest candidate plants and chemicals acting on specific targets (**Figure
3**). It was validated that the plants with more than 10 chemicals had enough experimental evidence to
explain the biological roles on specific targets retrieved in the network. TPS was also used for the
extraction of a list of plants and chemicals acting on a specific target of interest (**Figure 4**).

We also obtained potent plant and chemical candidates for the specific target, "tyrosinase,"
confirming the validity of the approach. The node information of the plants in the network is
expected to guide toward new alternative sources developed by plant combinations.

The approach is based on the multipartite network of plants and chemicals for a single target using
the TPS and could be expanded for multi-targets that closely interact with each other. The plants and
chemicals, which are expected to act on multiple targets, including S5R1, S5R2, and AR, could be
suggested to control the levels of testosterone (T) and dihydrotestosterone (DHT), involved in the
androgen metabolism. It is known that the blockade of AR can decrease benign prostate hyperplasia
(BPH) symptoms and further relieve the lower urinary tract symptoms (Izumi et al., 2013). Since the
alterations in AR are associated with prostate cancer, the inhibition of AR by drugs is the prevention
target for prostate cancer growth (Tan et al., 2015). The 5α-reductase inhibitors, which suppress the
conversion of T into DHT, contribute to the reduction in BPH or preventing prostate cancer (Izumi et
al., 2013; Liss and Thompson, 2018). In the present study, the AR-targeted chemicals were
sesquiterpenoid and diterpenoid-type, while the steroid derivatives were suggested as the chemicals
targeting 5α-reductases (S5R1 and S5R2). We expected that the plants containing the chemicals
acting on all three targets, S5R1, S5R2, and AR, can be the new candidates for controlling the
symptoms or diseases involving androgen metabolism, such as BPH and prostate cancer. The well-
organized multipartite network is expected to explain the synergistic effects between the multi-
chemicals in single or multiple medicinal plants.

We have constructed a multipartite network by integrating the databases of the medicinal plants,
chemicals, and targets for the prediction of the biological roles of the plants and chemicals on
specific targets. We have presented a novel computational approach based on the multipartite
network with which the plant species can be distinguished based on their chemical profiles. In the
clustered bipartite network of plants and chemicals, the chemical profiles of the plants in identical or
similar species were clustered closely at the bottom level of the dendrogram. However, the plants in
the same family were found far apart in the dendrogram in many cases, which implies that the
chemical profiles of the plants do not significantly contribute to the classification of the species.

In conclusion, we have demonstrated that the network analysis helps to understand the biological
roles of multi-chemicals on multi-targets. With the target prediction models, it also provides a way to
discover alternative therapeutic agents from natural sources by extrapolating from the known
bioactivities of chemicals on targets. The predicted results about the biological roles of multi-
chemicals on multi-targets allow to explain the therapeutic activity of the medicinal plant extracts via
subnetworks of molecular mechanisms, and suggest the combinatory preparation of several medicinal
plants on multi-targets to be involved in the symptoms or diseases of interest. In our future study, we

will study the candidates, plants or chemicals, predicted from the multipartite network in *in vitro* and
in vivo models to validate our approaches and further develop novel therapeutic botanical drugs.

5   References

Ahn, K. (2017). The worldwide trend of using botanical drugs and strategies for developing global
drugs. BMB Rep. 50, 111-116.

Anighoro, A., Bajorath, J., and Rastelli, G. (2014). Polypharmacology: challenges and opportunities
in drug discovery: miniperspective. J. Med. Chem. 57, 7874-7887.

Aydemir, G., Carlsen, H., Blomhoff, R., and Rühl, R. (2012). Lycopene induces retinoic acid
receptor transcriptional activation in mice. Mol. Nutr. Food Res. 56, 702-712.

Aydemir, G., Kasiri, Y., Birta, E., Béke, G., Garcia, A. L., Bartók, E.-M., et al. (2013). Lycopene-
derived bioactive retinoic acid receptors/retinoid-X receptors-activating metabolites may be
relevant for lycopene's anti-cancer potential. Mol. Nutr. Food Res. 57, 739-747.

Besnard, J., Ruda, G. F., Setola, V., Abecassis, K., Rodriguiz, R. M., Huang, X. P., et al. (2012).
Automated design of ligands to polypharmacological profiles. Nature 492, 215-220.

Bosc, N., Atkinson, F., Felix, E., Gaulton, A., Hersey, A., and Leach, A. R. (2019). Large scale
comparison of QSAR and conformal prediction methods and their applications in drug
discovery. J. Cheminformatics 11.

Cheng, F., Kovács, I. A., and Barabási, A.-L. (2019). Network-based prediction of drug
combinations. Nat. Commun. 10, 1197. doi:10.1038/s41467-019-09186-x.

Davies, M., Nowotka, M., Papadatos, G., Dedman, N., Gaulton, A., Atkinson, F., et al. (2015).
ChEMBL web services: streamlining access to drug discovery data and utilities. Nucleic Acids
Res. 43, W612-W620.

Dhillon, I. S. (2001). Co-clustering documents and words using bipartite spectral graph partitioning.
in Proceedings of the seventh ACM SIGKDD international conference on Knowledge
discovery and data mining - KDD '01 (San Francisco, California: ACM Press), 269-274.

Ezzat, A., Wu, M., Li, X., and Kwoh, C.-K. (2019). "Computational Prediction of Drug-Target
Interactions via Ensemble Learning," in Computational Methods for Drug Repurposing, ed. Q.
Vanhaelen (New York, NY: Springer New York), 239-254.

Fan, J., Zhang, K., Jin, Y., Li, B., Gao, S., Zhu, J., et al. (2019). Pharmacological effects of berberine
on mood disorders. J. Cell. Mol. Med. 23, 21-28.

Forouzesh, A., Samadi Foroushani, S., Forouzesh, F., and Zand, E. (2019). Reliable target prediction
of bioactive molecules based on chemical similarity without employing statistical methods.
Front. Pharmacol. 10, 835.

Galanakis, C. M. (2019). Nutraceuticals and natural product pharmaceuticals.

Gaulton, A., Hersey, A., Nowotka, M., Bento, A. P., Chambers, J., Mendez, D., et al. (2017). The
ChEMBL database in 2017. Nucleic Acids Res. 45, D945-D954.

Goh, K.-I., Oh, E., Jeong, H., Kahng, B., and Kim, D. (2002). Classification of scale-free networks.
Proc. Natl. Acad. Sci. 99, 12583-12588.

Han, K., Jeng, E. E., Hess, G. T., Morgens, D. W., Li, A., and Bassik, M. C. (2017). Synergistic drug
combinations for cancer identified in a CRISPR screen for pairwise genetic interactions. Nat.
Biotechnol. 35, 463-474.

Harvey, A. L., Edrada-Ebel, R., and Quinn, R. J. (2015). The re-emergence of natural products for
drug discovery in the genomics era. Nat. Rev. Drug Discov. 14, 111-129.

Hopkins, A. L. (2008). Network pharmacology: the next paradigm in drug discovery. Nat. Chem.
Biol. 4, 682-690.

Huang, X., Gao, Y., Zhi, X., Ta, N., Jiang, H., and Zheng, J. (2016). Association between vitamin A,
retinol and carotenoid intake and pancreatic cancer risk: Evidence from epidemiologic studies.
Sci. Rep. 6, 38936.

Izumi, K., Mizokami, A., Lin, W.-J., Lai, K.-P., and Chang, C. (2013). Androgen receptor roles in
the development of benign prostate hyperplasia. Am. J. Pathol. 182, 1942-1949.

Jia, J., Zhu, F., Ma, X., Cao, Z. W., Li, Y. X., and Chen, Y. Z. (2009). Mechanisms of drug
combinations: interaction and network perspectives. Nat. Rev. Drug Discov. 8, 111-128.

Kawano, M., Takagi, R., Kaneko, A., and Matsushita, S. (2015). Berberine is a dopamine D1- and
D2-like receptor antagonist and ameliorates experimentally induced colitis by suppressing
innate and adaptive immune responses. J. Neuroimmunol. 289, 43-55.

Keiser, M. J., Setola, V., Irwin, J. J., Laggner, C., Abbas, A. I., Hufeisen, S. J., et al. (2009).
Predicting new molecular targets for known drugs. Nature 462, 175-181.

Koehn, F. E., and Carter, G. T. (2005). The evolving role of natural products in drug discovery. Nat.
Rev. Drug Discov. 4, 206-220.

Koutsoukas, A., Simms, B., Kirchmair, J., Bond, P. J., Whitmore, A. V., Zimmer, S., et al. (2011).
From in silico target prediction to multi-target drug design: Current databases, methods and
applications. J. Proteomics 74, 2554-2574.

Kumar, S., and Pandey, A. K. (2013). Chemistry and Biological Activities of Flavonoids: An
Overview. Sci. World J. 2013, 1-16.

Lavecchia, A., and Cerchia, C. (2016). In silico methods to address polypharmacology: current
status, applications and future perspectives. Drug Discov. Today 21, 288-298.

Leung, E. L., Cao, Z.-W., Jiang, Z.-H., Zhou, H., and Liu, L. (2013). Network-based drug discovery
by integrating systems biology and computational technologies. Brief. Bioinform. 14, 491-505.

Li, N., Lee, K., Xi, Y., Zhu, B., Gary, B. D., Ramírez-Alcántara, V., et al. (2015). Phosphodiesterase
10A: a novel target for selective inhibition of colon tumor cell growth and β-catenin-dependent
TCF transcriptional activity. Oncogene 34, 1499-1509.

Lima-Mendez, G., and van Helden, J. (2009). The powerful law of the power law and other myths in
network biology. Mol. Biosyst. 5, 1482.

Liss, M. A., and Thompson, I. M. (2018). Prostate cancer prevention with 5-alpha reductase
inhibitors: concepts and controversies. Curr. Opin. Urol. 28, 42-45.

Lounkine, E., Keiser, M. J., Whitebread, S., Mikhailov, D., Hamon, J., Jenkins, J. L., et al. (2012).
Large-scale prediction and testing of drug activity on side-effect targets. Nature 486, 361-367.

Luo, Y., Zhao, X., Zhou, J., Yang, J., Zhang, Y., Kuang, W., et al. (2017). A network integration
approach for drug-target interaction prediction and computational drug repositioning from
heterogeneous information. Nat. Commun. 8, 573.

Martínez-Jiménez, F., Papadatos, G., Yang, L., Wallace, I. M., Kumar, V., Pieper, U., et al. (2013).
Target Prediction for an Open Access Set of Compounds Active against Mycobacterium
tuberculosis. PLoS Comput. Biol. 9, e1003253.

Monteiro, N. R. C., Ribeiro, B., and Arrais, J. (2020). Drug-target interaction prediction: end-to-end
deep learning approach. IEEE/ACM Trans. Comput. Biol. Bioinform.

NCI-DREAM Community, Bansal, M., Yang, J., Karan, C., Menden, M. P., Costello, J. C., et al.
(2014). A community computational challenge to predict the activity of pairs of compounds.
Nat. Biotechnol. 32, 1213-1222.

Newman, M. (2005). Power laws, Pareto distributions and Zipf's law. Contemp. Phys. 46, 323-351.

Newman, M. E. J. (2004). Fast algorithm for detecting community structure in networks. Phys. Rev.
E 69, 066133.

Newman, M. E. J. (2006). Modularity and community structure in networks. Proc. Natl. Acad. Sci.
103, 8577-8582.

Newman, M. E. J., and Girvan, M. (2004). Finding and evaluating community structure in networks.
Phys. Rev. E 69, 026113.

Petrovska, B. (2012). Historical review of medicinal plants′ usage. Pharmacogn. Rev. 6, 1.

Ramsay, R. R., Popovic-Nikolic, M. R., Nikolic, K., Uliassi, E., and Bolognesi, M. L. (2018). A
perspective on multi-target drug discovery and design for complex diseases. Clin. Transl. Med.
7, 3.

Rodrigues, T., Reker, D., Schneider, P., and Schneider, G. (2016). Counting on natural products for
drug design. Nat. Chem. 8, 531-541.

Roth, B. L., Sheffler, D. J., and Kroeze, W. K. (2004). Magic shotguns versus magic bullets:
selectively non-selective drugs for mood disorders and schizophrenia. Nat. Rev. Drug Discov.
3, 353-359.

Shin, Jin-Sup, Lee, Yu-Seon, and Lee, Myeong-Seon (2010). Protection and utilization of
430   traditional knowledge resources through Korean traditional knowledge portal(KTKP). J. Korea
431   Contents Assoc. 10, 422-426.

432   Smit, N., Vicanova, J., and Pavel, S. (2009). The hunt for natural skin whitening agents. Int. J. Mol.
433   Sci. 10, 5326-5349.

434   Sun, Y., Sheng, Z., Ma, C., Tang, K., Zhu, R., Wu, Z., et al. (2015). Combining genomic and
435   network characteristics for extended capability in predicting synergistic drugs for cancer. Nat.
436   Commun. 6, 8481.

437   Tan, M. E., Li, J., Xu, H. E., Melcher, K., and Yong, E. (2015). Androgen receptor: structure, role in
438   prostate cancer and drug discovery. Acta Pharmacol. Sin. 36, 3-23.

439   Wink, M. (2003). Evolution of secondary metabolites from an ecological and molecular phylogenetic
440   perspective. Phytochemistry 64, 3-19.

441   Wu, J. (2019). Cannabis, cannabinoid receptors, and endocannabinoid system: yesterday, today, and
442   tomorrow. Acta Pharmacol. Sin. 40, 297-299.

443   Xin, L.-T., Yue, S.-J., Fan, Y.-C., Wu, J.-S., Yan, D., Guan, H.-S., et al. (2017). Cudrania
444   tricuspidata: an updated review on ethnomedicine, phytochemistry and pharmacology. RSC
445   Adv. 7, 31807-31832.

446   Yıldırım, M. A., Goh, K.-I., Cusick, M. E., Barabási, A.-L., and Vidal, M. (2007). Drug—target
447   network. Nat. Biotechnol. 25, 1119-1126.

448   Zhang, C., Hong, H., Mendrick, D. L., Tang, Y., and Cheng, F. (2015). Biomarker-based drug safety
449   assessment in the age of systems pharmacology: from foundational to regulatory science.
450   Biomark. Med. 9, 1241-1252.

451   Zhang, W., Pei, J., and Lai, L. (2017). Computational multitarget drug design. J. Chem. Inf. Model.
452   57, 403-412.

453   Zhao, J., Jiang, P., and Zhang, W. (2010). Molecular networks for the study of TCM Pharmacology.
454   Brief. Bioinform. 11, 417-430.

455   Zheng, M., Liu, X., Xu, Y., Li, H., Luo, C., and Jiang, H. (2013). Computational methods for drug
456   design and discovery: focus on China. Trends Pharmacol. Sci. 34, 549-559.

Figures and Tables

(A)

(B)

(C)

Figure 1. (A) The construction and visualization of the multipartite network database. **(B)** The

representative multipartite network generated from the nodes which were randomly selected from 10%

of the nodes 1,138 plants, 10,043 chemicals, and 441 targets in the whole network. **(C)** The distribution

of the node degrees for the plants, chemicals, or targets.

Figure 2. The four sub-clusters in the whole dendrogram (**Figure S2**). **(A)** The largest sub-cluster in

the whole network. **(B)** The classification of chemicals in **(A)**. **(C)** The second largest sub-cluster in

the whole network. **(D)** The classification of chemicals in **(C)**. **(E)** The sub-cluster of the nodes sharing

protoberberine-type chemicals in the whole network. **(F)** The classification of chemicals in **(E)**. **(G)**

The sub-cluster of the nodes sharing vitamins in the whole network. **(H)** The classification of chemicals

in **(G)**.

Figure 3. (A) The network diagram for the plants cluster in **Figure 2A** and the 6 targets retrieved based

on the TPS index greater than or equal to 0.1. The chemicals effective to the targets are depicted in red

circles. **(B)** The network diagram for the plants cluster in **Figure 2C** and the 2 targets retrieved based

on the TPS index greater than or equal to 0.5. The chemicals effective to the targets are depicted in red

circles. **(C)** The network diagram for the plants cluster in **Figure 2E** and the 7 targets retrieved based

on the TPS index greater than or equal to 0.05. The chemicals effective to the targets are depicted in

red circles. **(D)** The network diagram for the plants cluster in **Figure 2G** and the 5 targets retrieved

based on the TPS index greater than or equal to 0.05. The chemicals effective to the targets are depicted

in red circles.

(A)   (B)

(C)   (D)   (E)

Figure 4. (A) The network diagram for the plants cluster having the largest TPS index to the target

'Tyrosinase' (CHEMBL3318). **(B)** The combined network generated from three network diagrams for

the plants cluster having the largest TPS index to the three targets, 'steroid 5α-reductase 1' (S5R1,

ChEMBL1787), 'steroid 5α-reductase 2' (S5R2, ChEMBL1856) and 'androgen receptor' (AR,

ChEMBL1871). The chemical profiles sharing from chemicals from S5R1 and S5R2 **(C),** from S5R1

and AR **(D)** and from S5R2 and AR **(E)**.

Table 1. Basic statistics of the combined multipartite network data of plants, chemicals, and targets.

The density of a multipartite network is computed by E/(Np*Nc+Nc*Nt).

	Number of Nodes				Number of Edges			Density
	Plant (Np)	Chemical (Nc)	Target (Nt)	Total (N)	Plant-Chemical	Chemical-Target	Total (E)	
Raw Data	5,500	10,056	1,224	16,780	58,068	100,290	158,358	0.00234
without Non-plants	3,614	10,056	1,224	14,894	54,585	100,290	154,875	0.00318
without Non-plants or Duplicates	2,886	10,056	1,224	14,166	34,549	100,290	134,839	0.00326
Only targets with the probability of above 0.9	2,886	10,056	441	13,383	34,549	73,112	107,661	0.00322
Pre-processed Data	1,138	10,043	441	11,622	34,549	73,112	107,661	0.00679

498 **Table 2**. List of chemicals simultaneously acting on three targets, S5R1, S5R2 and AR.

Chemical ID	Chemical Name	inchikey	Chemicals classification				
			Kingdom	Superclass	Class	Subclass	Direct_parent
9612	Kusunol	MQWIFDHBNGIVPO-UHFFFAOYSA-N	Organic compounds	Lipids and lipid-like molecules	Prenol lipids	Sesquiterpenoids	Eremophilane, 8,9-secoeremophilane and furoeremophilane sesquiterpenoids
10638	Stigmasta-4,6-dien-3-one	KEAZWUZFBSXOMV-UHFFFAOYSA-N	Organic compounds	Lipids and lipid-like molecules	Steroids and steroid derivatives	Stigmastanes and derivatives	Stigmastanes and derivatives
2001	Stigmastan-3-one (5-alpha)	BVVFRHKBULZQCQ-UHFFFAOYSA-N	Organic compounds	Lipids and lipid-like molecules	Steroids and steroid derivatives	Stigmastanes and derivatives	Stigmastanes and derivatives
11158	Dehydroabietinal	YCLCHPWRGSDZKL-UHFFFAOYSA-N	Organic compounds	Lipids and lipid-like molecules	Prenol lipids	Diterpenoids	Diterpenoids

1847	(-)-alpha-Copaene	VLXDPFLIRFYIME-UHFFFAOYSA-N	Organic compounds	Lipids and lipid-like molecules	Prenol lipids	Sesquiterpenoids	Sesquiterpenoids
9654	(-)-Ylangene	VLXDPFLIRFYIME-UHFFFAOYSA-N	Organic compounds	Lipids and lipid-like molecules	Prenol lipids	Sesquiterpenoids	Sesquiterpenoids

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