Title: A data-driven geospatial workflow to improve mapping species distributions and assessing
extinction risk under the IUCN Red List.

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ABSTRACT

Species distribution maps are essential for assessing extinction risk and guiding conservation efforts. Here, we developed a data-driven, reproducible geospatial workflow to map species distributions and evaluate their conservation status consistent with the guidelines and criteria of the IUCN Red List. Our workflow follows five automated steps to refine the distribution of a species starting from its Extent of Occurrence (EOO) to Area of Habitat (AOH) within the species range. The ranges are produced with an Inverse Distance Weighted (IDW) interpolation procedure, using presence and absence points derived from primary biodiversity data. As a case-study, we mapped the distribution of 2,273 bird species in the Americas, 55% of all terrestrial birds found in the region. We then compared our produced species ranges to the expert-drawn IUCN/BirdLife range maps and conducted a preliminary IUCN extinction risk assessment based on criterion B (Geographic Range). We found that our workflow generated ranges with fewer errors of omission, commission, and a better overall accuracy within each species EOO. The spatial overlap between both datasets was low (28%) and the expert-drawn range maps were consistently larger due to errors of commission. Their estimated Area of Habitat (AOH) was also larger for a subset of 741 forest-dependent birds. We also found that incorporating geospatial data increased the number of threatened species by 52% in comparison to the 2019 IUCN Red List, and 103 species could be placed in threatened categories (VU, EN, CR) pending further assessment. The implementation of our geospatial workflow provides a valuable alternative to increase the transparency and reliability of species risk assessments and improve mapping species distributions for conservation planning and decision-making.

Keywords: *geospatial analysis, IUCN Red List, species range maps, Red List assessment.*

Introduction

An unprecedented rate of extinction is affecting millions of species worldwide (IPBES 2019). Hence, there is an urgent need to assess threats across species ranges to guide planning and priority-setting where conservation actions are most needed (Bachman et al. 2019). The International Union for Conservation of Nature (IUCN) Red List of Threatened Species employs two spatial metrics under criterion B (Geographic Range) to assess species extinction risk (IUCN 2012): Extent of Occurrence (EOO) and Area of Occupancy (AOO). These metrics represent the upper and lower bounds of a species distribution and have a different theoretical basis — the former measures the degree of risk spread and the latter is closely linked to population size (Gaston & Fuller 2009). However, the IUCN has recommended that for planning conservation actions, other metrics might be more appropriate (IUCN/SSC 2018; IUCN Standards and Petitions Committee 2019).

Recently, Brooks et al. (2019) suggested that IUCN Red List assessments should measure Area of Habitat (AOH), also known as Extent of Suitable Habitat. AOH is relevant to guide conservation by quantifying habitat loss and fragmentation within a species range, and is already part of the criteria for identifying Key Biodiversity Areas (KBA) (IUCN 2016a; KBA Standards and Appeals Committee 2019). To calculate AOH, researchers have relied mostly on refining expert-drawn range maps by clipping unsuitable areas based on published elevational limits and known habitat preferences (Harris & Pimm 2008; Ocampo-Peñuela et al. 2016). This approach has been used to inform the conservation status of different groups of terrestrial organisms on a global scale (Rondinini et al. 2011; Ficetola et al. 2015; Tracewski et al. 2016). However, expert-drawn range maps can have low accuracy due to errors of omission (known presences outside of the mapped area) and commission (known absences inside the mapped area) (Beresford et al. 2011; Peterson et al. 2018; Mainali et al. 2020). Generating range maps that minimize these errors is essential to avoid mischaracterization of species distributional patterns for local scale applications (Hurlbert & Jetz 2007) and to better inform conservation planning and decision-making (Rahbek 2005; Ficetola et al. 2014; Mainali et al. 2020)

There are different methods available to produce species ranges as an alternative to expert-drawn range maps, and it is crucial to understand the limitations and trade-offs when considering alternatives (Graham & Hijmans 2006; Cantú-Salazar & Gaston 2013; Maréchaux et al. 2017). One alternative is using species distribution models (SDM) that correlate environmental variables with known occurrences (Peterson et al. 2011). These methods have been successfully applied to estimate species ranges and inform Red List assessments (Pena et al. 2014; Syfert et al. 2014; Breiner et al. 2017). However, SDMs can be complex

and involve methodological choices that if not well understood or explicitly communicated, can reduce

transparency and reproducibility (Guisan et al. 2013; Sofaer et al. 2019; Feng et al. 2019). Furthermore,

these models usually require species-specific fine-tuning of parameters (Radosavljevic & Anderson 2014;

Muscarella et al. 2014) which makes them challenging to scale for hundreds (or more) species. Thus, there

is an active search for more straightforward approaches (García-Roselló et al. 2019).

Spatial interpolation methods are an alternative approach to support the mapping of species distributions

and have been widely employed for conservation applications (Li & Heap 2008). Of the different types of

interpolation procedures available, Inverse Distance Weighting (IDW) is a deterministic method that is

accessible, user-friendly, and could be readily employed to support producing species ranges. IDW uses

known presences and absences to produce a surface of probability values for the occurrence of a species

within a given area (Hijmans & Elith 2019). The main assumption is that species are more likely to be found

closer to presence points and farther from absences (*i.e.* spatial autocorrelation), and the local influence of

points (weighted average) diminishes with distance. IDW has been used for mapping invasive plants

(Roberts et al. 2004) and to understand the distribution of coral reef sessile organisms, with a higher

accuracy than more advanced geostatistical interpolation methods (Zarco-Perello & Simões 2017).

Recently, Gomes et al. (2018) found that Maxent SDMs models largely overlapped with abundance maps

derived from using IDW based inventory plots for 227 hyper-dominant Amazonian tree species.

Furthermore, IDW may outperform SDM's when environmental variables do not provide more explanatory

power than the spatial structure of the occurrence data (Bahn & McGill 2007; Hijmans 2012). Thus, IDW

is also useful to compare it to other approaches in species distribution modeling (Raes & ter Steege 2007;

Hijmans 2012; Hijmans & Elith 2019).

Here, we developed a geospatial workflow to map species distributions with presence and absence points

derived from primary biodiversity data. We aimed to provide a reproducible and transparent approach to

(1) improve the accuracy of species ranges and the estimation of Area of Habitat (AOH) for local-scale

applications and (2) increase the reliability of extinction risk assessments under the IUCN Red List. To

show the advantages of our approach we implemented our workflow on 2,273 bird species in the Americas,

55% of all terrestrial birds found in the region. Birds represent a standard in the evaluation of species

conservation status (BirdLife International 2018) and comprise more than half of the occurrence records

on biodiversity databases (Troudet et al. 2017). Overall, by highlighting differences in produced ranges and

derived spatial metrics for a well-known group of organisms in a species-rich region, we present the case

for an improved approach at mapping species distributions for conservation planning and decision-making.

Methods

<u>Geospatial workflow</u>

We developed a data-driven, reproducible geospatial workflow (Fig. 1) to refine the distribution of a species starting from its Extent of Occurrence (EOO) to producing a final Area of Habitat (AOH) map. First, the user must gather and process occurrence data, an essential pre-processing step of any range mapping procedure. We suggest users follow recent guidelines e.g. (Araújo et al. 2019; Feng et al. 2019) to that end. Our proposed method consisted of five main steps:

1. Draw the Extent of Occurrence (EOO) around presence points. The EOO is the upper bound of the distribution of a species (Brooks et al. 2019) and should be mapped as the Minimum Convex Polygon (MCP) due to its consistency and scale-free properties (Joppa et al. 2016; IUCN Red List Technical Working Group 2019; IUCN Standards and Petitions Committee 2019).

2. Clip the EOO to the elevational limits of a species by extracting the elevation of each occurrence point using a Digital Elevation Model (DEM). The choice of DEM resolution depends on the spatial extent of the data and the intended analysis for which 90 m to 250 m resolutions are commonly employed (Amatulli et al. 2018)

3. Overlay absence points to the clipped EOO. Absences can be derived from known surveys, monitoring schemes, or citizen science projects that have a measure of sampling effort such as eBird or biodiversity atlases (Johnston et al. 2019).

4. Interpolate presences and absence points using Inverse Distance Weighting (IDW). The output is a raster surface within the clipped EOO where each cell has a value between 0 and 1. The species range is obtained by using a threshold to convert continuous results to a binary score where the species is either present or absent (Liu et al. 2005). The ranges produced with IDW are consistent with the observation that distributions are naturally porous and have discontinuities (holes) at higher resolutions (Hurlbert & Jetz 2007).

5. Clip a habitat layer inside the species IDW range to remove unsuitable areas and derive Area of Habitat (AOH) (KBA Standards and Appeals Committee 2019). We recommend following the IUCN Habitat Classification Scheme for consistency. Users could use empirically developed products (e.g., Nature Map Explorer – www.explorer.naturemap.earth) or use expert criteria to select appropriate landcover classes that match the definition of species habitats coded by the IUCN.

<u>Case Study</u>

Here we show the implementation of our proposed geospatial workflow using data for terrestrial bird

species in the Americas. We followed the Handbook of the Birds of the World-Birdlife Taxonomic

Checklist v4, which is the basis for the IUCN Red List of Threatened Species and bird species distribution

maps of the world (HBW and BirdLife International 2019). We selected bird species with ranges smaller

than the global median [< 1M km^2 (Orme et al. 2005; Jenkins et al. 2013)] because they are typically more

at risk of extinction (Chichorro et al. 2018). We excluded species endemic to the Galapagos and Hawaiian

Islands and those classified as Extinct (EX), Extinct in the Wild (EW), and possibly extinct (CR- PE)

according to the 2019 IUCN Red List. Our final dataset comprised 2,273 bird species in the Americas, 55%

of all terrestrial birds found in the region (BirdLife International 2020). We generated ranges for all species

and derived Area of Habitat (AOH) for a subset of 741 species whose only habitat type is "forest" according

to the IUCN Habitat Classification Scheme (Donald et al. 2018).

1.   Primary biodiversity data

1.1 Gathering and processing occurrence data

Occurrence data was gathered from the Global Biodiversity Information Facility (GBIF;

https://www.gbif.org). The GBIF biodiversity database is a repository of multiple occurrence datasets

spanning from museum specimen records to the eBird and iNaturalist citizen science projects (GBIF 2020).

We obtained occurrence data with the R package *rgbif* (Chamberlain & Boettiger 2017) using the

'occ_search' function in April 2019. We removed duplicated coordinates and removed species with five

records or less (Pender et al. 2019). We used the R package *CoordinateCleaner* (Zizka et al. 2019) to

automatically filter common erroneous coordinates in public databases such as those assigned to the sea,

country capitals, or biodiversity institutions (Maldonado et al. 2015).

The GBIF occurrence data operates on a stable taxonomic backbone that although useful for database

management does not represent current knowledge on species limits (Burfield et al. 2017). Thus, we

consulted the Avibase database (https://avibase.bsc-eoc.org; Lepage et al. 2014) to check for species that

differed from the HBW-Birdlife taxonomy, and manually updated their occurrence data with the interactive

modular R platform *Wallace* (Kass et al. 2018). In this process we accounted for taxonomic splits and

removed obvious geographical outliers that we deemed as occurrence points in areas where a species is not

able to disperse given the presence of large geographical barriers (Barve et al. 2011; Hazzi et al. 2018). Our

final dataset of species occurrence data can be found in Appendix S1.

1.2 Deriving absences

We detected absences based on eBird hotspots – publicly accessible locations that are frequently birded and
have been previously approved by an administrator. Here, we deemed absences as eBird hotspots where a
given species *has never been recorded.* We used this approach as a stringent criterion to minimize the
uncertainties regarding absences (Lobo et al. 2010). We used the R package *auk* (Strimas-Mackey et al.
2016) to extract from the eBird Basic Dataset (2019) observational checklists for 57 countries and territories
in the Americas. We limited the data to complete checklists (birding events where observers report all
species detected) from January 1, 2000 to April 31, 2019. Using complete checklists is a critical step to
obtain better models of species distribution and is recommended as part of the eBird best practices (Johnston
et al. 2019; Strimas-Mackey et al. 2020). We obtained 142,703 points locations corresponding to eBird
hotspots which we used to identify absences (see mapping protocol).

2. Detailed Workflow

The implementation of our workflow followed methodological choices that were conceived to work for a
large number of species and computational efficiency, mostly using default parameters that users can adapt
or modify depending on the intended purpose. Our workflow was run on CyVerse's Atmosphere cloud-
computing platform to speed processing time (see acknowledgments). Following best practices for sharing
large datasets (Marwick et al. 2018), we provide the R code to reproduce it on a small sample of species
(see Appendix).

In our procedure we drew each species EOO and then buffered it to standard 10 km (Ocampo-Peñuela &
Pimm 2014). Then, we extracted elevational limits from occurrence points using the SRTM v4.1 DEM at
250 m resolution (Jarvis et al. 2008) which is a good compromise between computational ease and
resolution in complex topographical areas such as mountains (Hazzi et al. 2018). The interquartile range
(IQR) was used as a robust test for outlier detection, in which elevational data points below $Q_{25} - 1.5$ *IQR,
or above $Q_{75} + 1.5$ *IQR were removed (Wilcox 2017). We overlaid absence points to the clipped EOO and
removed those within a 5 km buffer of any presence points (Johnston et al. 2019; Strimas-Mackey et al.
2020).

We computed IDW in the R package *dismo* (Hijmans et al. 2017) using all presence and absence points
available for each species. To derive AOH for forest species we used a recently developed 50-m global
forest/non-forest map from the TanDEM-X satellite (Martone et al. 2018). This product is designed to

specifically estimate forest cover instead of other approaches that are more focused on evaluating forest

cover change [*e.g.* Hansen et al. (2013)]. We aggregated to 250 m with a nearest neighbor assignment to

match the resolution of our elevational data.

3. Comparison of species ranges

We compared the IDW and BirdLife ranges in terms of accuracy, range size, and range overlap. We also

evaluated differences in the estimation of Area of Habitat (AOH). For the BirdLife ranges, we examined

how accuracy influences their range size and the amount of spatial overlap with the IDW ranges.

3.1 BirdLife distribution maps

We obtained the BirdLife distribution maps v.2019-1 (created November 2019) that come assembled in an

ESRI File Geodatabase (http://datazone.birdlife.org/species/requestdis). Each species is represented by

polygons coded by different categories of presence, origin, and seasonality (BirdLife International and

HBW 2019). We combined all coded polygons to represent each species range into a single spatial feature

(Cantú-Salazar & Gaston 2013) using ArcGIS Pro 2.4.1. To estimate Area of Habitat (AOH) from the

BirdLife ranges, we refined the polygon of each species by elevation and forest cover (*sensu* Ocampo-

Peñuela et al. [2016]). We used the 50-m global forest/non-forest map that we employed in our geospatial

workflow. Species elevational limits were obtained from the IUCN Red List and HBW Alive.

3.2 Accuracy assessment

We evaluated the discriminant capacity of the IDW and BirdLife ranges to classify presences and absence

points within each species EOO. We used three basic metrics of classification performance: errors of

omission, errors of commission, and overall accuracy (Anderson et al. 2003). Omission errors were

quantified based on the amount of GBIF presences left outside of the mapped range (false negatives),

whereas commission errors were quantified based on the amount of eBird absences that remain inside the

mapped range (false positives). The overall accuracy was calculated using Jaccard's similarity index, which

summarizes the ability of the mapped range to (a) maximize true presences, (b) reduce both false negatives

and false positives, and (c) disregard true negatives that are easily classified due to issues of prevalence (Li

& Guo 2013; Leroy et al. 2018).

The accuracy metrics were calculated with a confusion matrix that cross-tabulates the match between
predictions and observations (Fig. 2 for reference). For the BirdLife ranges we computed the accuracy
metrics overlaying all the available presences and absences of each species. For the IDW ranges, we
conducted a *k*-fold cross-validation procedure using 80% of the data for training and 20% for testing the
model, resulting in five different folds for evaluation (Gareth et al. 2013). The resulting confusion matrices
of each fold were summed to represent the total performance of the model and compute the final accuracy
metrics.

3.3 Range size comparison

We evaluated differences in range size between the IDW and BirdLife ranges for our complete dataset of
2,273 species. For this, the IDW prediction surface was transformed into a binary map (presence/absence).
We used a probability threshold of > 0.5 to retain areas where the species is more likely to occur. This
threshold is particularly useful to compare the areas of different distribution maps that have been produced
for any given species (Zhang et al. 2005). We tested for a statistically significant difference between both
datasets with a Two-sample Kolmogorov–Smirnov test (K-S test). Then, we quantified the differences in
the distribution of range sizes across deciles with a shift function (Fig. 3) using the *rogme* package in R
(Rousselet et al. 2017).

We also computed a robust log-linear regression with an MM-type estimator in the R package *robustbase*
(Maechler et al. 2019) to examine how omission and commission errors influence difference in range size
between IDW and BirdLife datasets. We further calculated the 20% trimmed mean of range size for each
dataset to assess how the typical IDW and BirdLife ranges compare to each other. Trimmed means are a
robust measure of centrality recommend to estimate average values of skewed distributions (Wilcox 2017).
We estimated a 95% Confidence Interval (CI) of the range size difference using a percentile bootstrap
procedure. The same approach was used to estimate differences in Area of Habitat (AOH) for our subset of
741 forest species. We also measured the effect size with a Brunner–Munzel test, which estimated the
probability that a randomly sampled range from one of the datasets is larger than another randomly sampled
range from the other dataset. Statistical analysis was conducted with R functions provided by R.R. Wilcox
(https://dornsife.usc.edu/labs/rwilcox/software)

4. Comparison of IUCN extinction risk assessments

We used the R package *ConR* (Dauby et al. 2017) to compute with the gathered occurrence data, a

preliminary extinction risk assessment based on the IUCN criterion B [Geographic Range] parameters

(IUCN 2012): Extent of Occurrence (EOO) and Area of Occupancy (AOO). Species were assigned to

threatened categories when threshold levels are met for either EOO or AOO (e.g. AOO: Vulnerable <2,000

km^2). Note that AOO is calculated by tallying the area of 2 x 2 km grid cells with documented presences

(IUCN Standards and Petitions Committee 2019). Thus, it is a subset of Area of Habitat (AOH) (Brooks

et al. 2019). In addition to EOO or AOO thresholds, at least two out of three other conditions must be met

if a species is to be assigned to threatened categories (IUCN 2012). Two of these conditions are continued

decline in habitat and number of locations, defined as areas in which a single threatening event can affect

all individuals of a species (IUCN Standards and Petitions Committee 2019).

ConR assumes that habitat is declining for all species and quantifies locations based on 10 km^2 gridded

squares around presence data, for which 10 or fewer locations are the threshold to meet threatened

categories (IUCN 2012). These assumptions to automate the process help assessors to find and prioritize

species and should be interpreted as a first filter for a full IUCN extinction risk assessment. We used the R

package *rredlist* (Chamberlain 2019) to query from the IUCN Red List 2019 the following data: species

threat categories and criteria, upper and lower elevational limits, EOO and AOO metrics. We compared

differences in EOO and AOO measurements between our automated protocol and the Red List. Then we

evaluated the agreement between the threatened categories (Vulnerable - VU, Endangered - EN, Critically

Endangered - CR) in the preliminary automated assessment and the IUCN Red List 2019 using Kendall's

coefficient of concordance (Kendall's *W*) which ranges between 0 and 1 (no agreement to complete

agreement).

Results

1.   Accuracy

We found that for the complete dataset of 2,273 bird species, our ranges had reduced errors of omission

and commission with increased overall accuracy within the species EOO (Fig. 3). For the BirdLife ranges,

the 20% trimmed mean of omission errors was 0.23 (95% percentile bootstrap CI [0.22; 0.23], p < 0.001).

Commission errors were common (0.43; CI: 0.41-0.44, *p* < 0.001) and the overall accuracy of the ranges

was only moderate (0.52; CI: 0.51-0.53, *p* < 0.001). In comparison, For the IDW ranges omission errors

were lower (0.20; CI: 0.19-0.21, p < 0.001). Commission errors were very low (0.06; CI: 0.06-0.07, p <

0.001) and the overall accuracy was higher (0.76; CI: 0.75-0.76, p < 0.001). All of these differences were

statistically significant (Two-sided KS-test; $p < 0.001$). In percentages, 90% of our ranges had a higher

overall accuracy than the BirdLife ranges, 52% had lower omission errors and 85% had lower commission

errors.

2. Range size and spatial overlap

The IDW and BirdLife datasets had different distributions of range sizes for the 2,273 bird species analyzed

(Fig 4A; K-S test = 0.18, $p < 0.001$). A shift function showed the BirdLife ranges are consistently larger

than the IDW ranges across the entire distribution of values with positive differences between all deciles

(Fig 4B).

The 20% trimmed mean showed that the typical BirdLife range is almost twice as large as the typical IDW

range (BirdLife $_{tmean}$ = 163,809 km^2 ; IDW $_{tmean}$ = 82,843 km^2) with an estimated difference of 80,966 km^2

(CI:69,396 - 93,254 km^2], $p < 0.001$). The percentage of BirdLife ranges that are larger than ours is 79% (n

= 2,273). As a measure of effect size, the probability that a BirdLife range is larger than an IDW range for

a given species if selected at random is 0.61 (CI 0.60 - 0.63, $p < 0.001$). The estimated Area of Habitat

(AOH) derived from BirdLife ranges was also larger than those derived from the IDW ranges (BirdLife-

AOH $_{tmean}$ = 47,499 km^2 ; IDW-AOH$_{tmean}$ = 38,745 km^2) with an estimated difference of 8,753 km^2 (CI:

1,913 - 15,488 km^2, $p = 0.013$).

We found that commission errors increased differences in range size for the BirdLife ranges. A robust log-

linear regression showed that commission errors have a larger positive effect (*coef* = 1.24, *SE* = 1.14, t =

6.45, $p <0.001$) than the negative effect of omission errors (*coef* = -0.79, *SE* = 0.25, t= -3.13, $p = 0.02$) on

range size difference (*intercept* = 10.40, 0.14, t = 74.47, p < 0.001). Both independent variables were

significant predictors of difference in range size (X^2 =9.82, *df* =1, $p = 0.002$). The spatial overlap between

the BirdLife and IDW ranges was only 28% (95% percentile bootstrap CI [0.26; 0.29], p < 0.001). A robust

regression showed that for the BirdLife ranges, the overall accuracy had a significant effect on range overlap

(*coef* = 0.54, *SE* = 0.015, t = 35.01, $p <0.001$). This means that decreasing accuracy in the BirdLife ranges

decreases the spatial overlap with the IDW ranges and the expected estimates of AOH (Fig. 5)

3. IUCN extinction risk assessments

For the 2,273 bird species analyzed, we found that based on criterion B the 2019 IUCN Red List (hereafter
Red List) had 131 species in threatened categories whereas our preliminary assessment had 199, an increase
of 51.9%. Moreover, there is only a moderate agreement between both criterion B assessments in terms of
species assigned categories (Kendall's $W = 0.65$, $p < 0.001$). We also identified 103 species as threatened
that are currently classified as non-threatened categories (LC or NT) according to criteria B in IUCN (2019).

We analyzed the estimates of AOO and EOO derived from the occurrence data and those from the Red List.
There was not a statistically significant difference in EOO values between both datasets (K-S test = 0.03, p
$= 0.122$). Nevertheless, we identified 142 species that could be placed in threatened categories based on
EOO thresholds, 66 of which are currently classified in non-threatened categories. Critically, only 36
species (1.6%) had a value of AOO listed in the Red List (all species had EOO values), whereas we
identified 57 species that triggered AOO thresholds, 37 of which are currently considered as non-threatened.

We found 148 species that do not have any recorded information on their elevational limits (upper or lower)
based on the data available on the Red List. Additionally, 886 species do not have a recorded lower
elevation but have an upper limit, whereas 11 species do not have an upper limit but have a lower limit
recorded. In total, 1045 (46%) have missing information regarding elevational limits. For the remaining
1,228 species that have complete elevational limits, 892 species (73%) have a narrower elevational range
than the one found in our geospatial workflow, whereas 336 (27%) have a broader one. The difference in
elevational range for both datasets was significant (K-S test = 0.30, $p < 0.001$).

Discussion

We have developed an automated and reproducible geospatial workflow to map species distributions and
assess extinction risk starting from primary biodiversity data, in a way that is consistent with the guidelines
and criteria of the IUCN Red List. Using a dataset of 2,273 bird species in the Americas, 55% of all
terrestrial birds found in the region, we have shown that our approach produced ranges that have reduced
errors of omission, fewer errors of commission, and a higher accuracy when compared to expert-drawn
alternatives. Thus, it also provides a more reliable, transparent, and consistent way to derive estimates of
Area of Habitat (AOH), the newly recommended measure for the IUCN Red List (Brooks et al. 2019).

Our workflow emphasized confirmed records and excluded potential distributional areas outside of each
species EOO (Extent of Occurrence). Therefore, the generated ranges are especially useful for systematic
conservation planning (Margules & Pressey 2000) where resources are directed to areas in which a species
is known to be present. Unlike most expert-drawn range maps, we also excluded areas of known absence.
Although absences always carry a high degree of uncertainty (Lobo et al. 2010) they are crucial for
increasing the predictive accuracy of species ranges and therefore deriving better estimates of AOH (Elith
et al. 2006; Mainali et al. 2020). The maps of AOH derived from our ranges could be valuable for
conservationists in activities such as establishing protected areas or designing habitat corridors. On the other
hand, given the low accuracy that we found on the expert-drawn ranges, we concur with Peterson et al.
(2018) and do not recommend procedures that refine such maps to obtain estimates of AOH for guiding
conservation efforts.
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We have provided an objective and transparent way to produce species ranges, but we do not intend it for
every scenario — there are no 'silver bullets' to represent species distributions for every purpose (Guillera-
Arroita et al. 2015; Qiao et al. 2015). We do not recommend using our ranges for macroecological studies
(*e.g.* species richness or endemism patterns) given that our predictions stay close to the presence points
available and are limited by sampling bias. These biases might have influenced our findings that the
BirdLife ranges were consistently larger than our generated ranges, but our findings are in line with previous
research demonstrating substantial overestimation of range size from expert-drawn maps (Hurlbert & Jetz
2008; Jetz et al. 2008). Moreover, we also showed that range overestimation was driven by commission
errors (false presences inside the range) that are prevalent in the BirdLife ranges. In contrast, our ranges
markedly reduced commission errors and have a higher overall accuracy. Hence, our ranges are a reliable
alternative for representing current knowledge on species distributions for conservation applications, with
the advantage of being produced with an explicit and reproducible method.
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Our study also highlighted the importance of incorporating geospatial data within the IUCN red listing
process to improve species extinction risk estimates. We show with an automated assessment that when
using occurrence data (GBIF), the number of potentially threatened species increased by 52% and identified
up to 103 species likely threatened that are currently considered in non-threatened categories (LC or NT)
based on criterion B of the 2019 IUCN Red List. We stress that this analysis is preliminary and is not
intended to replace a full Red List assessment. Nevertheless, it does suggest that the IUCN Red List might
be underestimating species extinction risk given the missing spatial information in key assessment
parameters (EOO and AOO). For instance, while we derived AOO estimates for all the species in our
dataset, less than 2% of species had a reported value of AOO in the IUCN Red List and 46% had missing

elevational limits. Although the estimates of EOO between our methods and the Red List were not
statistically different, it was sufficient to produce differences in the threat categorization of many species,
with only moderate agreement between our preliminary conservation assessment and the 2019 IUCN Red
List.

We recommend that the IUCN Red List incorporates more transparent and reproducible protocols, such as
the one we have presented in this paper, to aid in mapping species distributions and increase transparency
in species risk assessments. Our workflow could be useful to narrow down species that meet threatened
criteria for further consultation with experts. Additional suggestions can be found in the literature - these
include adopting species range model metadata standards (Merow et al. 2019), checklists to reproduce
ecological niche models (Feng et al. 2019) and standards for biodiversity assessments (Araújo et al. 2019).
These could help update the IUCN Rules of Procedure (IUCN 2016b) and Mapping Standards and Data
Quality (IUCN Red List Technical Working Group 2019). Following suit, we suggest that all the spatial
data used for assessments under the IUCN Red List should be made publicly available so that it can be fully
understood, reviewed, and updated (Durant et al. 2017).

Our study demonstrated the utility and importance of a data-driven, reproducible approach to integrate
mapping species distributions and extinction risk assessments in a framework compatible with the IUCN
Red List. We have shown how our proposed workflow (1) assisted the red listing process by incorporating
geospatial data and analysis to detect more potentially threatened species; (2) improved the accuracy of
species ranges over expert-drawn range maps, and; (3) generated more reliable estimates of Area of Habitat
(AOH) for local-scale planning and decision-making. In the urgency to evaluate the conservation status of
many more species and implement measures to halt biodiversity loss (IUCN Species Survival Commission
2019), our workflow is a valuable addition to the toolkit of conservation practitioners and decision-makers
because it is consistent, easily understood and widely applicable. Notwithstanding, we acknowledge the
scarcity of spatial data is limiting in many groups and there are no easy shortcuts to address this shortfall
(Rodrigues 2011). Only with more field explorations and carefully designed biological surveys, we will
overcome the scarce knowledge on species occurrences and distributions around the globe for effective on-
the-ground conservation actions (Wilson 2017).

REFERENCES

Amatulli G, Domisch S, Tuanmu M-N, Parmentier B, Ranipeta A, Malczyk J, Jetz W. 2018. A suite of global, cross-scale topographic variables for environmental and biodiversity modeling. Scientific Data **5**:180040.

Anderson RP, Lew D, Peterson AT. 2003. Evaluating predictive models of species' distributions: criteria for selecting optimal models. Ecological Modelling **162**:211–232.

Araújo MB et al. 2019. Standards for distribution models in biodiversity assessments. Science Advances **5**:eaat4858.

Ayerbe-Quiñones F. 2019. An Illustrated Field Guide to the Birds of Colombia. Wildlife Conservation Society Colombia.

Bachman SP, Field R, Reader T, Raimondo D, Donaldson J, Schatz GE, Lughadha EN. 2019. Progress, challenges and opportunities for Red Listing. Biological Conservation **234**:45–55.

Bahn V, McGill BJ. 2007. Can niche-based distribution models outperform spatial interpolation? Global Ecology and Biogeography **16**:733–742.

Barve N, Barve V, Jiménez-Valverde A, Lira-Noriega A, Maher SP, Peterson AT, Soberón J, Villalobos F. 2011. The crucial role of the accessible area in ecological niche modeling and species distribution modeling. Ecological Modelling **222**:1810–1819.

Beresford AE, Buchanan GM, Donald PF, Butchart SHM, Fishpool LDC, Rondinini C. 2011. Poor overlap between the distribution of Protected Areas and globally threatened birds in Africa: Protected Areas and threatened African birds. Animal Conservation **14**:99–107.

BirdLife International. 2018. State of the world's birds: taking the pulse of the planet. Cambridge, UK: BirdLife International.

BirdLife International. 2020. Birdlife Data Zone. http://datazone.birdlife.org/.

BirdLife International and HBW. 2019. Bird species distribution maps of the world. Version 2019.1. Available at http://datazone.birdlife.org/species/requestdis.

Breiner FT, Guisan A, Nobis MP, Bergamini A. 2017. Including environmental niche information to improve IUCN Red List assessments. Diversity and Distributions **23**:484–495.

Brooks TM et al. 2019. Measuring Terrestrial Area of Habitat (AOH) and Its Utility for the IUCN Red List. Trends in Ecology & Evolution. Available from https://linkinghub.elsevier.com/retrieve/pii/S0169534719301892 (accessed July 17, 2019).

Burgman MA, Fox JC. 2003. Bias in species range estimates from minimum convex polygons: implications for conservation and options for improved planning. Animal Conservation **6**:19–28.

Cantú-Salazar L, Gaston KJ. 2013. Species richness and representation in protected areas of the Western

hemisphere: discrepancies between checklists and range maps. Diversity and Distributions

19:782–793.

Chamberlain S. 2019. rredlist: "IUCN" Red List Client. R package version 0.5.0. Available from

https://CRAN.R-project.org/package=rredlist.

Chamberlain SA, Boettiger C. 2017. R Python, and Ruby clients for GBIF species occurrence data.

preprint. PeerJ Preprints. Available from https://peerj.com/preprints/3304 (accessed September

11, 2019).

Chichorro F, Juslén A, Cardoso P. 2018. A systematic review of the relation between species traits and

extinction risk. Available from http://biorxiv.org/lookup/doi/10.1101/408096 (accessed

September 5, 2018).

Collar NJ, editor. 1992. Threatened birds of the Americas.3. ed. nternational Council for Bird

Preservation ; International Union for Conservation of Nature, Cambridge, U.K. : Gland,

Switzerland.

Dauby G et al. 2017. ConR: An R package to assist large-scale multispecies preliminary conservation

assessments using distribution data. Ecology and Evolution **7**:11292–11303.

Donald PF, Arendarczyk B, Spooner F, Buchanan GM. 2018. Loss of forest intactness elevates global

extinction risk in birds. Animal Conservation. Available from

http://doi.wiley.com/10.1111/acv.12469 (accessed December 22, 2018).

Durant SM et al. 2017. The global decline of cheetah *Acinonyx jubatus* and what it means for

conservation. Proceedings of the National Academy of Sciences **114**:528–533.

Elith J et al. 2006. Novel methods improve prediction of species' distributions from occurrence data.

Ecography **29**:129–151.

Feng X, Park DS, Walker C, Peterson AT, Merow C, Papeş M. 2019. A checklist for maximizing

reproducibility of ecological niche models. Nature Ecology & Evolution. Available from

http://www.nature.com/articles/s41559-019-0972-5 (accessed October 23, 2019).

Ficetola GF, Rondinini C, Bonardi A, Baisero D, Padoa-Schioppa E. 2015. Habitat availability for

amphibians and extinction threat: a global analysis. Diversity and Distributions **21**:302–311.

Ficetola GF, Rondinini C, Bonardi A, Katariya V, Padoa-Schioppa E, Angulo A. 2014. An evaluation of

the robustness of global amphibian range maps. Journal of Biogeography **41**:211–221.

García-Roselló E, Guisande C, González-Vilas L, González-Dacosta J, Heine J, Pérez-Costas E, Lobo

JM. 2019. A simple method to estimate the probable distribution of species. Ecography **42**:1613–

1622.

Gareth J, Witten D, Hastie T, Tibshirani R, editors. 2013. An introduction to statistical learning: with

applications in R. Springer, New York.

Gaston KJ, Fuller RA. 2009. The sizes of species' geographic ranges. Journal of Applied Ecology **46**:1–9.

GBIF. 2020. GBIF: The Global Biodiversity Information Facility (year) What is GBIF?. Available from

https://www.gbif.org/what-is-gbif [26 February 2020].

Gomes VHF et al. 2018. Species Distribution Modelling: Contrasting presence-only models with plot

abundance data. Scientific Reports **8**:1003.

Graham CH, Hijmans RJ. 2006. A comparison of methods for mapping species ranges and species

richness. Global Ecology and Biogeography **15**. Available from

http://doi.wiley.com/10.1111/j.1466-822X.2006.00257.x (accessed November 30, 2017).

Guillera-Arroita G, Lahoz-Monfort JJ, Elith J, Gordon A, Kujala H, Lentini PE, McCarthy MA, Tingley

R, Wintle BA. 2015. Is my species distribution model fit for purpose? Matching data and models

to applications: Matching distribution models to applications. Global Ecology and Biogeography

24:276–292.

Guisan A et al. 2013. Predicting species distributions for conservation decisions. Ecology Letters

16:1424–1435.

Hansen MC et al. 2013. High-Resolution Global Maps of 21st-Century Forest Cover Change. Science

342:850–853.

Harris G, Pimm SL. 2008. Range Size and Extinction Risk in Forest Birds: *Range Size and Extinction*

Risk. Conservation Biology **22**:163–171.

Hazzi NA, Moreno JS, Ortiz-Movliav C, Palacio RD. 2018. Biogeographic regions and events of isolation

and diversification of the endemic biota of the tropical Andes. Proceedings of the National

Academy of Sciences **115**:7985–7990.

HBW and BirdLife International. 2019. Handbook of the Birds of the World and BirdLife International

digital checklist of the birds of the world. Version 4. Available at:

http://datazone.birdlife.org/userfiles/file/Species/Taxonomy/HBW-

BirdLife_Checklist_v4_Dec19.zip [.xls zipped 1 MB].

Hijmans RJ. 2012. Cross-validation of species distribution models: removing spatial sorting bias and

calibration with a null model. Ecology **93**:679–688.

Hijmans RJ, Elith J. 2019. Spatial Distribution Models. Available from https://rspatial.org/.

Hijmans RJ, Phillips S, Leathwick J, Elith J. 2017. dismo: Species Distribution Modeling. Available from

https://CRAN.R-project.org/package=dismo.

Hurlbert AH, Jetz W. 2007. Species richness, hotspots, and the scale dependence of range maps in

ecology and conservation. Proceedings of the National Academy of Sciences **104**:13384–13389.

IPBES. 2019. Summary for policymakers of the global assessment report on biodiversity and ecosystem
services of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem
Service. IPBES secretariat, Bonn, Germany. 56 pages.
IUCN. 2012. IUCN Red List categories and criteria, version 3.1, second edition. Gland, Switzerland and
Cambridge, UK: IUCN.
IUCN. 2016a. A Global Standard for the Identification of Key Biodiversity Areas, Version 1.0. Gland,
Switzerland: IUCN.
IUCN. 2016b. Rules of Procedure for IUCN Red List Assessments 2017–2020.Version 3.0. Approved by
the IUCN SSC Steering Committee in September 2016. Downloadable from:
http://cmsdocs.s3.amazonaws.com/keydocuments/Rules_of_Procedure_for_Red_List_2017-
2020.pdf.
IUCN Red List Technical Working Group. 2019. Mapping Standards and Data Quality for IUCN Red
List Spatial Data. Version 1.18. Prepared by the Standards and Petitions Working Group of the
IUCN SSC Red List Committee. Downloadable from:
https://www.iucnredlist.org/resources/mappingstandards.
IUCN Species Survival Commission. 2019. The Abu Dhabi Call for Global Species Conservation Action.
Available from
https://www.iucn.org/sites/dev/files/content/documents/the_abu_dhabi_call_for_global_species_c
onservation_action_adopted_01112019.pdf.
IUCN Standards and Petitions Committee. 2019. Guidelines for Using the IUCN Red List Categories and
Criteria. Version 14. Prepared by the Standards and Petitions Committee. Downloadable from
http://www.iucnredlist.org/documents/RedListGuidelines.pdf. Available from
http://www.iucnredlist.org/documents/RedListGuidelines.pdf.
IUCN/SSC. 2018. Guidelines for Species Conservation Planning - version 1.0. Species Conservation
Planning Sub-Committee. Gland, Switzerland. IUCN, International Union for Conservation of
Nature. Available from https://portals.iucn.org/library/node/47142 (accessed July 3, 2019).
Jarvis A, Reuter HI, Nelson A, Guevara E. 2008. Hole-filled SRTM for the globe Version 4, available
from the CGIAR-CSI SRTM 90m Database (http://srtm.csi.cgiar.org).
Jenkins CN, Pimm SL, Joppa LN. 2013. Global patterns of terrestrial vertebrate diversity and
conservation. Proceedings of the National Academy of Sciences **110**:E2602–E2610.
Johnston A, Hochachka W, Strimas-Mackey M, Gutierrez VR, Robinson O, Miller E, Auer T, Kelling S,
Fink D. 2019. Best practices for making reliable inferences from citizen science data: case study
using eBird to estimate species distributions. preprint. Ecology. Available from
http://biorxiv.org/lookup/doi/10.1101/574392 (accessed February 24, 2020).

Joppa LN et al. 2016. Impact of alternative metrics on estimates of extent of occurrence for extinction risk
assessment: Extent of Occurrence and Extinction Risk. Conservation Biology **30**:362–370.

Kaschner, K, Kesner-Reyes K, Garilao C, Rius-Barile J, Rees T, Froese R. 2016. AquaMaps: Predicted
range maps for aquatic species. World wide web electronic publication, www.aquamaps.org,
Version 10/2019. Available from www.aquamaps.org.

Kass JM, Vilela B, Aiello-Lammens ME, Muscarella R, Merow C, Anderson RP. 2018. Wallace: A
flexible platform for reproducible modeling of species niches and distributions built for
community expansion. Methods in Ecology and Evolution **9**:1151–1156.

KBA Standards and Appeals Committee. 2019. Guidelines for using a Global Standard for the
Identification of Key Biodiversity Areas. Version 1.0. Prepared by the KBA Standards and
Appeals Committee of the IUCN Species Survival Commission and IUCN World Commission on
Protected Areas. Gland, Switzerland: IUCN. Available from
https://portals.iucn.org/library/sites/library/files/documents/2019-001.pdf.

Lepage D, Vaidya G, Guralnick R. 2014. Avibase – a database system for managing and organizing
taxonomic concepts. ZooKeys **420**:117–135.

Leroy B, Delsol R, Hugueny B, Meynard CN, Barhoumi C, Barbet-Massin M, Bellard C. 2018. Without
quality presence-absence data, discrimination metrics such as TSS can be misleading measures of
model performance. Journal of Biogeography **45**:1994–2002.

Li J, Heap AD. 2008. A review of spatial interpolation methods for environmental scientists. Geoscience
Australia, Record 2008/23.

Li W, Guo Q. 2013. How to assess the prediction accuracy of species presence-absence models without
absence data? Ecography **36**:788–799.

Liu C, Berry PM, Dawson TP, Pearson RG. 2005. Selecting thresholds of occurrence in the prediction of
species distributions. Ecography **28**:385–393.

Lobo JM, Jiménez-Valverde A, Hortal J. 2010. The uncertain nature of absences and their importance in
species distribution modelling. Ecography **33**:103–114.

Maechler M, Rousseeuw P, Croux C, Todorov V, Ruckstuhl A, Salibian-Barrera M, Verbeke T, Koller
M, Conceicao ELT, di Palma MA. 2019. robustbase: Basic Robust Statistics R package version
0.93-5. Available from http://CRAN.R-project.org/package=robustbase.

Mainali K, Hefley T, Ries L, Fagan W. 2020. Matching expert range maps with species distribution
model predictions. Conservation Biology:cobi.13492.

Maldonado C, Molina CI, Zizka A, Persson C, Taylor CM, Albán J, Chilquillo E, Rønsted N, Antonelli
A. 2015. Estimating species diversity and distribution in the era of Big Data: to what extent can
we trust public databases? Global Ecology and Biogeography **24**:973–984.

Maréchaux I, Rodrigues ASL, Charpentier A. 2017. The value of coarse species range maps to inform
local biodiversity conservation in a global context. Ecography **40**:1166–1176.

Margules CR, Pressey RL. 2000. Systematic conservation planning. Nature **405**:243–253.

Martone M, Rizzoli P, Wecklich C, González C, Bueso-Bello J-L, Valdo P, Schulze D, Zink M, Krieger
G, Moreira A. 2018. The global forest/non-forest map from TanDEM-X interferometric SAR
data. Remote Sensing of Environment **205**:352–373.

Marwick B, Boettiger C, Mullen L. 2018. Packaging Data Analytical Work Reproducibly Using R (and
Friends). The American Statistician **72**:80–88.

Merow C, Maitner BS, Owens HL, Kass JM, Enquist BJ, Jetz W, Guralnick R. 2019. Species' range
model metadata standards: RMMS. Global Ecology and Biogeography **28**:1912–1924.

Muscarella R, Galante PJ, Soley-Guardia M, Boria RA, Kass JM, Uriarte M, Anderson RP. 2014.
ENMeval: An R package for conducting spatially independent evaluations and estimating optimal
model complexity for MAXENT ecological niche models. Methods in Ecology and Evolution
5:1198–1205.

Ocampo-Peñuela N, Jenkins CN, Vijay V, Li BV, Pimm SL. 2016. Incorporating explicit geospatial data
shows more species at risk of extinction than the current Red List. Science Advances
2:e1601367–e1601367.

Ocampo-Peñuela N, Pimm SL. 2014. Setting practical conservation priorities for birds in the Western
Andes of Colombia. Conservation Biology **28**:1260–1270.

O'Hara CC, Afflerbach JC, Scarborough C, Kaschner K, Halpern BS. 2017. Aligning marine species
range data to better serve science and conservation. PLOS ONE **12**:e0175739.

Orme CDL et al. 2005. Global hotspots of species richness are not congruent with endemism or threat.
Nature **436**:1016–1019.

Pena JC de C, Kamino LHY, Rodrigues M, Mariano-Neto E, de Siqueira MF. 2014. Assessing the
conservation status of species with limited available data and disjunct distribution. Biological
Conservation **170**:130–136.

Pender JE, Hipp AL, Hahn M, Kartesz J, Nishino M, Starr JR. 2019. How sensitive are climatic niche
inferences to distribution data sampling? A comparison of Biota of North America Program
(BONAP) and Global Biodiversity Information Facility (GBIF) datasets. Ecological Informatics
54:100991.

Peterson AT, Navarro-Sigüenza AG, Gordillo A. 2018. Assumption-versus data-based approaches to
summarizing species' ranges. Conservation Biology **32**:568–575.

Peterson AT, Soberon J, Pearson RG, Anderson RP, Martinez-Meyer E, Nakamura M, Araújo MB,

editors. 2011. Ecological niches and geographic distributions. Princeton University Press,

Princeton, N.J.

Qiao H, Soberón J, Peterson AT. 2015. No silver bullets in correlative ecological niche modelling:

insights from testing among many potential algorithms for niche estimation. Methods in Ecology

and Evolution **6**:1126–1136.

Radosavljevic A, Anderson RP. 2014. Making better Maxent models of species distributions: complexity,

overfitting and evaluation. Journal of Biogeography **41**:629–643.

Raes N, ter Steege H. 2007. A null-model for significance testing of presence-only species distribution

models. Ecography **30**:727–736.

Rahbek C. 2005. The role of spatial scale and the perception of large-scale species-richness patterns:

Scale and species-richness patterns. Ecology Letters **8**:224–239.

Roberts EA, Sheley RL, Lawrence RL. 2004. Using sampling and inverse distance weighted modeling for

mapping invasive plants. Western North American Naturalist **64**. Available from

https://scholarsarchive.byu.edu/wnan/vol64/iss3/4.

Rodrigues ASL. 2011. Improving coarse species distribution data for conservation planning in

biodiversity-rich, data-poor, regions: no easy shortcuts. Animal Conservation **14**:108–110.

Rondinini C et al. 2011. Global habitat suitability models of terrestrial mammals. Philosophical

Transactions of the Royal Society B: Biological Sciences **366**:2633–2641.

Rousselet GA, Pernet CR, Wilcox RR. 2017. Beyond differences in means: robust graphical methods to

compare two groups in neuroscience. European Journal of Neuroscience **46**:1738–1748.

Sofaer HR et al. 2019. Development and Delivery of Species Distribution Models to Inform Decision-

Making. BioScience **69**:544–557.

Strimas-Mackey M, Hochachka WM, Ruiz-Gutierrez V, Robinson OJ, Miller ET, Auer T, Kelling S, Fink

D, Johnston A. 2020. Best Practices for Using eBird Data.Version 1.0.

https://cornelllabofornithology.github.io/ebird-best-practices/. Cornell Lab of Ornithology, Ithaca,

New York. Available from https://doi.org/10.5281/zenodo.3620739.

Strimas-Mackey M, Miller E, Hochachka W. 2016. auk: eBird Data Extraction and Processing with

AWK. R package version 0.3.0. Available from https://cornelllabofornithology.github.io/auk/.

Syfert MM, Joppa L, Smith MJ, Coomes DA, Bachman SP, Brummitt NA. 2014. Using species

distribution models to inform IUCN Red List assessments. Biological Conservation **177**:174–184.

Tracewski Ł, Butchart SHM, Di Marco M, Ficetola GF, Rondinini C, Symes A, Wheatley H, Beresford

AE, Buchanan GM. 2016. Toward quantification of the impact of 21st-century deforestation on

the extinction risk of terrestrial vertebrates: Effects of Deforestation on Vertebrates. Conservation Biology **30**:1070–1079.

Troudet J, Grandcolas P, Blin A, Vignes-Lebbe R, Legendre F. 2017. Taxonomic bias in biodiversity data and societal preferences. Scientific Reports **7**. Available from http://www.nature.com/articles/s41598-017-09084-6 (accessed December 4, 2017).

Velásquez-Tibatá J, Olaya-Rodríguez MH, López-Lozano D, Gutiérrez C, González I, Londoño-Murcia MC. 2019. BioModelos: A collaborative online system to map species distributions. PLOS ONE **14**:e0214522.

Wilcox RR. 2017. Introduction to robust estimation and hypothesis testing.4th edition. Elsevier, Waltham, MA.

Wilson EO. 2017. Biodiversity research requires more boots on the ground. Nature Ecology & Evolution **1**:1590–1591.

Zarco-Perello S, Simões N. 2017. Ordinary kriging vs inverse distance weighting: spatial interpolation of the sessile community of Madagascar reef, Gulf of Mexico. PeerJ **5**:e4078.

Zhang L, Liu S, Sun P, Wang T, Wang G, Zhang X, Wang L. 2015. Consensus Forecasting of Species Distributions: The Effects of Niche Model Performance and Niche Properties. PLOS ONE 10(3): e0120056.

Zizka A et al. 2019. CoordinateCleaner: standardized cleaning of occurrence records from biological collection databases. Methods in Ecology and Evolution. Available from http://doi.wiley.com/10.1111/2041-210X.13152 (accessed January 21, 2019).

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Author contributions

R.D.P. conceived and developed the geospatial workflow, performed the analysis and drafted the manuscript. P.J.N., J.V-T, and A.P.J. provided critical input to the analysis and contributed with extensive edits. All authors gave final approval for publication.

Data Availability

The bird species distribution maps of the world are available upon request to BirdLife International (http://datazone.birdlife.org/species/requestdis). Access to the IUCN Red List API is granted upon request (https://apiv3.iucnredlist.org/api/v3/token) and information is freely accessible at www.iucnredlist.org. Species occurrences can be gathered from its data providers through GBIF.org. The eBird observations and associated metadata can be obtained upon completion of a data request form (https://ebird.org/data/download). The SRTM v4.1 DEM is freely available for download from the CGIAR-CSI Consortium for Spatial Information (http://srtm.csi.cgiar.org/srtmdata/) The TanDEM-X Forest/Non-Forest Map - Global can be freely downloaded from (https://download.geoservice.dlr.de/FNF50/)

Code Availability

The R code for the geospatial workflow will be made available upon reasonable request, previous to its deposition in an open-access repository with the peer-reviewed version of this study. Requests should be sent to the corresponding author.

Competing Interests

The authors declare no competing interests.

772   **Figures**

Draw a minimum convex polygon around presence points to obtain EOO

Extract the elevation of presence points and clip EOO to species elevational limits

Overlay absence points within the clipped EOO

Use Inverse Distance Weighted (IDW) interpolation to obtain range

Clip habitat layer within the IDW range

Final map of Area of Habitat (AOH)

777   ***Figure* 1.** Mapping protocol to refine the distribution of a species from Extent of Occurrence (EOO) to

778   Area of Habitat (AOH). This is an illustrative example for the Glittering Starfrontlet (*Coeligena orina*), a

779   hummingbird species in the cloud forests of the western Andes of Colombia (please refer to figure 2 for a

780   picture of the species). The EOO was buffered by 10 km following the implementation in our workflow

781   (see methods).

Figure 2. Example of accuracy analysis for the BirdLife range of the Glittering Starfrontlet (*Coeligena orina*) in the western Andes of Colombia, using a confusion matrix to calculate accuracy metrics. The overall accuracy was high (0.78) but note the arbitrary division of the range into four areas, including a fifth one indicated with the arrow in which no data points are available on GBIF (up to April 2020). This area did not enter into the accuracy calculation because it is outside of the EOO (see methods). The IDW range is shown for visual comparison. TP = True Presences; FP = False Presences; TN= True Negatives; FN = False Negatives. Photo by Andres Cuervo.

Figure 3. Values for each of the three evaluated metrics of accuracy (commission error, omission error

and overall accuracy) between the IDW and the BirdLife ranges. The boxes show the Interquartile Range

(IQR) which visualize the spread of 50% of the data points between the 25th and 75th percentile. The notches

represent 95% confidence intervals of the median indicating differences because of no overlap. Note that

most distributions are skewed, and data points outside of the whiskers (1.5 x IQR) are not necessarily

outliers. There was a statistically significant difference between the values of the IDW and BirdLife ranges

for each evaluated metric (two-sample K-S test, $p < 0.001$).

Figure 4. **A.** Comparison of the distribution of range sizes between IDW and BirdLife for 2,273 birds in

the Americas. The thicker line corresponds to the median of the distribution and the thinner lines represent

deciles **B.** Pairwise differences showed that for each decile across the distribution the BirdLIfe ranges are

consistently larger in value than the IDW ranges. These difference are estimated with 95% confidence

intervals derived using a percentile bootstrap procedure (Rousselet et al. 2017)

Figure 5. **A.** The Colombian endemic Yellow-eared parrot (*Ognorhynchus icterotis*) showed particularly poor spatial overlap between BirdLife ranges and IDW ranges, and respective estimated Area of Habitat (AOH). Note that BirdLife range extends the distribution north of the Eastern Andes where there are no records. On the inset, the expert-derived range map from Ayerbe-quiñones (2019) represented the species distribution much closer to the IDW range. **B.** For the Tawny-tufted Toucanet (*Selenidera nattereri*) in the Amazonian lowlands, the AOH derived from BirdLife is nearly three times larger than the estimated AOH derived from the IDW range. **C.** The BirdLife range for the Brown-rumped Tapaculo (*Scytalopus latebricola*) is a polygon covering the whole area of the Santa Marta mountains. In contrast, the IDW range restricted the AOH to the western side where observations have been recorded.

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