1 **The impact of purifying and background selection on the inference of population history:**
2 **problems and prospects**
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4 Parul Johri[*,‡], Kellen Riall[*], Hannes Becher[†], Brian Charlesworth[†], and Jeffrey D. Jensen[*,‡]
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6 [*]School of Life Sciences, Arizona State University, Tempe, AZ 85287, USA
7 [†]Institute of Evolutionary Biology, School of Biological Sciences, University of Edinburgh, EH9
8 3FL, United Kingdom
9 [‡]Correspondence can be addressed to these authors
10
11 **Corresponding authors:**
12 Parul Johri
13 pjohri1@asu.edu
14
15 Jeffrey D. Jensen
16 Jeffrey.D.Jensen@asu.edu

ABSTRACT

Current procedures for inferring population history are generally performed under the

assumption of complete neutrality - that is, by neglecting both direct selection and the effects of

selection on linked sites. We here examine how the presence of direct purifying and background

selection may bias demographic inference by evaluating two commonly-used methods (MSMC

and *fastsimcoal2*), specifically studying how the underlying shape of the distribution of fitness

effects (DFE) and the fraction of directly selected sites interact with demographic parameter

estimation. The results show that, even after masking functional genomic regions, background

selection effects may result in the mis-inference of population growth under models of both

constant population size as well as decline. This effect is amplified as the strength of purifying

selection and the density of directly selected sites increases, as indicated by the distortion of the

site frequency spectrum and levels of nucleotide diversity at linked neutral sites. We also show

how simulated changes in background selection effects caused by population size changes can be

predicted analytically. We propose a potential method for correcting for the mis-inference of

population growth caused by selection. By treating the DFE as a nuisance parameter and

averaging across all potential realizations, we demonstrate that even directly selected sites may

be used to infer demographic histories with reasonable accuracy.

Keywords: demographic inference, background selection, distribution of fitness effects, MSMC,

fastsimcoal2, approximate Bayesian computation (ABC)

Running title: Demographic inference with selection

INTRODUCTION

The characterization of past population size change is a central goal of population genomic analysis - with applications ranging from anthropological to agricultural to clinical (see review by Beichman *et al.* 2018). Furthermore, use of an appropriate demographic model provides a necessary null model for assessing the impact of selection across the genome (*e.g.*, Teshima *et al.* 2006; Thornton and Jensen 2007; Jensen *et al.* 2019). Multiple strategies have been proposed for performing demographic inference, utilizing expectations related to levels of variation, the site frequency spectrum, linkage disequilibrium, and within- and between-population relatedness (*e.g.*, Gutenkunst *et al.* 2009; Li and Durbin 2011; Lukic and Hey 2012; Harris and Nielsen 2013; Excoffier *et al.* 2013; Bhaskar *et al.* 2015; Sheehan and Song 2016; Ragsdale and Gutenkunst 2017; Steinrücken *et al.* 2019; Kelleher *et al.* 2019; Speidel *et al.* 2019).

Although many methods perform well when evaluated under the standard assumption of neutrality, it is difficult in practice to assure that the nucleotide sites used in empirical analyses experience neither direct selection nor the effects of selection at linked sites. For example, inference is often performed using intergenic, 4-fold degenerate, or intronic sites. While there is evidence for weak direct selection in all of these categories in multiple organisms (*e.g.*, Haddrill *et al.* 2005; Chamary and Hurst 2005; Andolfatto 2005; Lynch 2007; Zeng and Charlesworth 2010; Choi and Aquadro 2016; Jackson *et al.* 2017), it is also clear that such sites near or in coding regions will also experience background selection (BGS; Charlesworth *et al.* 1993; Charlesworth 2013), and may periodically be affected by selective sweeps as well (Messer and Petrov 2013; Schrider et al. 2016). These effects are known to affect the local underlying effective population size, and alter both the levels and patterns of variation and linkage disequilibrium (Charlesworth et al. 1993; Kaiser and Charlesworth 2009; O'Fallon et al. 2010; Charlesworth 2013; Nicolaisen and Desai 2013; Ewing and Jensen 2016; Johri et al. 2020).

However, commonly-used approaches for performing demographic inference that assume complete neutrality, including *fastsimcoal2* (Excoffier et al. 2013) and MSMC/PSMC (Li and Durbin 2011; Schiffels and Durbin 2014), have yet to be thoroughly evaluated in the light of this assumption, which is likely to be violated in practice. There are, however, some exceptions. Rather than investigating existing software, Ewing and Jensen (2016) implemented an approximate Bayesian (ABC) approach to quantify the impact of BGS effects, demonstrating that weak purifying selection can generate a skew towards rare alleles that would be mis-

interpreted as population growth. Under certain scenarios, this resulted in a many-fold mis-

inference of population size change. However, the effects of the density of directly selected sites

and the shape of the distribution of fitness effects (DFE), which are probably of great

importance, have yet to be considered. Spanning the range of these potential parameter values is

important for understanding the implications for empirical application. For example, the

proportion of the genome experiencing direct purifying selection can vary greatly between

species, with estimates ranging from ~3-8% in humans, to ~12% in rice, to 37-53% in *D.*

melanogaster, to 47-68% in *S. cerevisiae* (Siepel et al. 2005; Liang et al. 2018). Further, many

organisms have highly compact genomes, with ~88% of the *E. coli* genome (Blattner et al. 1997)

and effectively all of many virus genomes, being functional (*e.g.*, >95% of the SARS-CoV-2

genome, Wu *et al.* 2020).

105   While such estimates allow us to approximate the effects of BGS in some model

organisms, in which recombination and mutation rates are well known, it is difficult to predict

these effects in the vast majority of study systems. Moreover, while the genome-wide mean of *B*,

a widely-used measure of BGS effects that measures the level of variability relative to neutral

expectation, can range from ~0.45 in *D. melanogaster* to ~0.94 in humans (Charlesworth 2013;

but see Pouyet *et al.* 2018), existing demographic inference approaches are usually applied

across organisms without considering this important source of differences in levels of bias. Here,

we examine the effects of the DFE shape and functional density on two common demographic

inference approaches - the multiple sequentially Markovian coalescent (MSMC) and

fastsimcoal2. Finally, we propose an extension within the approximate Bayesian computation

(ABC) framework to address this issue, treating the DFE as a nuisance parameter and

demonstrating greatly improved demographic inference even when using directly selected sites

alone.

RESULTS and DISCUSSION

Effects of SNP numbers and genome size on inference under neutral equilibrium

The accuracy and performance of demographic inference was evaluated using two popular

methods, MSMC (Schiffels and Durbin 2014) and *fastsimcoal2* (Excoffier et al. 2013). In order

to assess performance, it was first necessary to determine how much genomic information is

required to make accurate inference when the assumptions of neutrality are met. Chromosomal

segments of varying sizes (10 Mb, 50 Mb, 200 Mb, and 1 Gb) were simulated under neutrality

and demographic equilibrium (*i.e.,* a constant population size of 5000 diploid individuals) with

100 independent replicates each. For each replicate this amounted to the mean [SD] number of

segregating sites for each diploid individual being 1,944 [283], 9,996 [418], 40,046 [957] and

200,245 [1887]; for 50 diploid individuals, these values were 10,354 [225], 51,863 [567],

207,118 [1139] and 1,035,393 [2476] for 10 Mb, 50 Mb, 200 Mb and 1 Gb, respectively. Use of

MSMC resulted in incorrect inferences for all segments smaller than 1 Gb (Supp Figure 1).

Specifically, very strong recent growth was inferred instead of demographic equilibrium,

although ancestral population sizes were correctly estimated. In addition, when 2 or 4 diploid

genomes were used for inference, MSMC again inferred a recent many-fold growth for all

segment sizes even when the true model was equilibrium, but performed well when using 1

diploid genome for large segments (Supp Figure 1).

When using *fastsimcoal2* to perform demographic inference, parameters were accurately

estimated for all chromosomal segment sizes when the correct model (*i.e.*, equilibrium) was

specified (left panel of Supp Figure 2). However, when model selection was performed using a

choice of three models (equilibrium, instantaneous size change, and exponential size change),

64% of replicates of the 10 Mb chromosomal segment incorrectly inferred a size change (Supp

Figure 3a), although the current population sizes were estimated accurately (middle panel of

Supp Figure 2). When the instantaneous bottleneck model was added to the set of competing

models, the true model (equilibrium) was correctly inferred in only 11-33% of replicates (right

panel of Supp Figure 2; Supp Figure 3b). These results suggest caution when performing

inferences on smaller regions or genomes, specifically when the number of SNPs are less than

~200,000 per single diploid individual, or less than 1 million per 50 diploid individuals. Extra

caution should be used when interpreting population size changes inferred by MSMC when

using more than 1 diploid individual.

Given this performance, all further analyses were restricted to characterizing

demographic inference on data that roughly matched the structure and size of the human genome

- for every diploid individual, 22 chromosomes (autosomes) of size 150 Mb each were simulated,

which amounted to roughly 3 Gb of total sequence. Ten independent replicates of each parameter

combination were performed throughout, and inference utilized 1 and 50 diploid individuals for

MSMC and *fastsimcoal2,* respectively.

Effect of the strength of purifying selection on demographic inference

In order to test demographic inference in the presence of BGS, all 22 chromosomes were

simulated with exons of size 350 bp each, with varying sizes of introns and intergenic regions

(see Methods) being used in order to vary the fraction (5%, 10% and 20%) of the genome under

selection. Because the strength of selection acting on deleterious mutations affects how far the

effects of BGS extend, demographic inference was evaluated for various DFEs (Table 1). The

DFE was modelled as a discrete distribution with four fixed classes: $0 \le 2N_e s < 1$, $1 \le 2N_e s \le$

10, $10 < 2N_e s \le 100$ and $100 < 2N_e s \le 2N_e$, where N_e is the effective population size and s is the

reduction in fitness of the homozygous mutant relative to wildtype. The fitness effects of

mutations were uniformly distributed within each bin, assumed to be semi-dominant, and the

DFE shape was altered by varying the proportion of mutations belonging to each fixed class,

given by f_0, f_1, f_2, and f_3, respectively (see Methods). Three DFEs highly skewed towards a

particular class were initially used to assess the impact of the strength of selection on

demographic inference (with the remaining mutations equally distributed amongst the other three

classes):

DFE1: a DFE in which 70% of mutations have weakly deleterious fitness effects (*i.e.*, $f_1 = 0.7$),

DFE2: a DFE in which 70% of mutations have moderately deleterious fitness effects (*i.e.*, $f_2 =$

0.7), and DFE3: a DFE in which 70% of mutations have strongly deleterious fitness effects (*i.e.*,

$f_3 = 0.7$).

In order to understand the effects of BGS, exonic sites were masked, and only linked

neutral intergenic sites were used for demographic inference. The three demographic models

examined were (1) demographic equilibrium, (2) a 30-fold exponential growth, mimicking the

recent growth experienced by European human populations, and (3) ~6-fold instantaneous

decline, mimicking the out-of-Africa bottleneck in human populations (Figure 1a). Although

these models were parameterized using previous estimates of human demographic history (Supp

Table 1; Gutenkunst *et al.* 2009), these basic demographic scenarios are applicable to many

organisms, although the magnitudes of population size changes in this case may represent an

extreme. Although for the 30-fold exponential growth model the ancestral population size was

consistently over-estimated when using *fastsimcoal2*, even under neutrality, the site frequency

spectrum (SFS) obtained from the test simulations and from the best-fit model inferred by

fastsimcoal2 are virtually indistinguishable (Supp Table 2). In other words, the model fitting is

working correctly, but the models themselves result in nearly identical expected frequency

spectra (and see Rosen et al. 2018). In order to better characterize this mis-inference, we

simulated 2-fold, 5-fold, 10-fold, and 20-fold exponential growth for comparison. Generally

speaking, *fastsimcoal2* performs better under models of instantaneous size change compared to

those involving exponential change (Supp Table 3).

Under demographic equilibrium, when 20% of the genome experiences direct selection

(with masking of the directly selected sites), we found the true population size to be

underestimated, as well recent population growth mis-inferred (Figure 1), with a larger bias in

MSMC than *fastsimcoal2*. Stronger growth is inferred with larger fitness effects of mutations.

Conversely, when the true demographic model is characterized by recent 30-fold growth,

demographic inference is accurate and performs equally well for both MSMC and *fastsimcoal2*,

with the exception of a slight underestimation of the ancestral population size for all DFE types.

When the true model is population decline, weakly deleterious mutations alone did not affect

inference drastically with either method, and it was possible to recover the true model by

fastsimcoal2. However, moderately and strongly deleterious mutations resulted in an

underestimation of population size and an inference of strong growth, to the extent that

population decline would be misinterpreted as growth. We further tested the effect of BGS on

demographic inference when changes in population size were less severe, namely, when

population growth and decline was only 2-fold, with qualitatively similar results (Supp Figure 4).

Finally, given the strong evidence that most organisms have a bi-modal DFE with a

significant proportion of strongly deleterious or lethal mutations (Sanjuán 2010; Jacquier et al.

2013; Kousathanas and Keightley 2013; Bank et al. 2014; Charlesworth 2015), we investigated

the effect of this strongly deleterious class further. Thus, for comparison with the above, we

simulated a rather extreme case in which 30% or 50% of exonic mutations are strongly

deleterious with fitness effects uniformly sampled between $100 \le 2N_{anc}s \le 2N_{anc}$, with the

remaining mutations being neutral (*i.e.*, DFE5 and DFE6; see Table 1). As with the above

results, both equilibrium and decline models were falsely inferred as growth, with an order of

magnitude underestimation of the true population size (Figure 2).

In sum, neglecting BGS frequently results in the inference of population growth, almost

regardless of the true underlying demographic model.

Effects of density and inclusion/exclusion of directly selected sites on inference

Although we have shown that the presence of purifying selection biases demographic inference,

the extent of mis-inference necessarily depends on the fraction of the genome experiencing direct

selection. We therefore compared models in which 5%, 10% or 20% of the genome was

functional. For this comparison, equal proportions of mutations in each DFE bin were assumed

(*i.e.,* $f_0 = f_1 = f_2 = f_3 = 0.25$) corresponding to DFE4 (Table 1). As before, when the true model

was growth, inference was unbiased, with a slight underestimation of ancestral population size

when 20% of the genome experiences selection (Figure 3). Population decline was inferred

reasonably well if less than 10% of the genome experienced direct selection, but could be mis-

inferred as growth with greater functional density, as shown in Figure 3. Similarly, the extent to

which population size is under-estimated at demographic equilibrium increases with the fraction

of the genome under selection. Finally, it is noteworthy that many changes in population size that

were falsely inferred were greater than 2-fold in size, suggesting the need for great caution when

inferring such changes from real data.

Importantly, the results presented do not significantly differ between inference performed

while including directly selected sites (*i.e.*, no masking of functional regions; Supp Figure 5)

versus inference performed using linked neutral sites (*i.e.*, masking functional regions; Figures 1-

3). These results suggest that the exclusion of exonic sites, which is often assumed to provide a

sufficiently neutral dataset to enable accurate demographic inference, is not necessarily a

satisfactory solution unless gene density is low.

Effect of BGS on model selection and inferred time of size change using *fastsimcoal2*

In order to quantify the effects of BGS on model selection, four competing models were used for

inference: equilibrium, instantaneous size change (growth/decline), exponential size change

(growth/decline), and an instantaneous bottleneck. Although demographic equilibrium was

almost always inferred as instantaneous size change (70-100% of replicates), when there existed

a substantial proportion of highly deleterious mutations (DFE3 and DFE5) exponential growth

was generally inferred instead. However, when there was a true size change, *fastsimcoal2*

performed well in distinguishing between exponential *vs.* instantaneous change models even in
the presence of BGS (Supp Figure 6), provided that the magnitude of size change was large.
When size changes were on the order of 2-fold, exponential growth was consistently inferred to
be instantaneous.
With respect to model choice between growth and decline in the presence of BGS
(irrespective of instantaneous *vs.* exponential change), as the density of selected sites and
strength of purifying selection increased, both equilibrium and decline models were more likely
to be inferred as growth (Supp Figure 7), while true growth models were generally chosen
correctly. It should be added that with such large chromosome sizes (3 Gb of total sequence
data), model selection was not observed to vary between replicates using *fastsimcoal2* for any
given parameter combination. Thus, in the presence of BGS, high-confidence calls of an
incorrect underlying demographic model appear likely.
With regards to the time of inferred size change, when the true model was exponential
growth, the model was always correctly identified and inference of the time of change was only
slightly under-estimated in the presence of BGS (Supp Figure 8 and 9), consistent with the fact
that BGS will further skew the site frequency spectrum towards rare alleles. When the true model
was decline, and the model was correctly identified as such, the time of change was modestly
over-estimated (Supp Figure 10, 11) – up to 1.3-fold for 6× growth and 2.9-fold for 2× growth
(when 20% of the genome is exonic). However, when the decline model was incorrectly
identified as growth, the timing of size change was severely under-estimated (Supp Figure 10,
11).
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Effect of heterogeneity in recombination rates, mutation rates, and repeat masking
Variation in recombination and mutation rates, as well as the masking of repeat regions, may
also affect demographic inference procedures. We evaluated this issue by simulating
heterogeneity in both mutation and recombination rates (based on estimated human genome
maps, as described in the Methods section), and masking 10% of each simulated segment
drawing from the empirical distribution of repeat lengths in the human genome (Supp Figure 12).
In general, inferences under neutrality (Supp Figures 13-15) as well as under BGS (Supp Figures
16-18) were unaffected under all demographic models. These investigations suggest that such
demographic inference is robust to genome-wide variation in rates of recombination and

mutation, as well as the presence of repeat elements, at least for the parameters considered here.

Thus, serious mis-inference is more likely to be caused by selection. These observations also

suggest that simulations performed with mean rates of recombination and mutation, as in this

study, are sufficient to evaluate biases caused by BGS.

Effects of BGS on diversity and the SFS under various demographic models: theoretical

expectations versus simulation results

To better understand how BGS can lead to different biases in the inference of population history,

we investigated the extent of BGS effects under all three demographic models, with respect to

both the reduced diversity relative to neutrality, as well as the shape of the SFS at linked neutral

sites. First, we found that B, the diversity relative to the neutral expectation, differed among

demographic scenarios, disproportionately amplifying mis-inference under equilibrium and

decline (Figure 4). After a population decline, B was lower than that before the size change;

while after population expansion, B increased relative to that in the ancestral population,

sometimes approaching 1 (Figure 4). This may seem paradoxical, given that the magnitude of the

scaled selection coefficient ($2N_e s$) decreases with decreasing N_e (*i.e.*, the efficacy of purifying

selection decreases, and could thus be expected to result in larger values of B under population

decline). Conversely, with increasing N_e, B may be reduced.

However, these expectations apply only once a population has maintained a given N_e for

sufficient time such that equilibrium has been approached. During the initial stages of population

size change, and shortly afterwards, the dynamics of B tend to show a trend opposite to this long-

term expectation (see also Figure 5 of Torres *et al.* 2020). This is because differences in N_e

caused by different initial levels of BGS cause differences in the rates of response to changes in

population size – a small value of N_e (corresponding to low B) results in a faster response

compared with a high value (Fay and Wu 1999; Hey and Harris 1999; Pool and Nielsen 2007;

Pool and Nielsen 2009; Campos et al. 2014; Torres et al. 2020). The relative diversity values

observed with different initial equilibrium B values after a short period of population size change

may thus be very different from both the initial and final equilibrium values, so that the apparent

B values estimated by comparing diversities with and without BGS differ from the equilibrium

values. The overall effect is that there is an apparent increase in B immediately following a

population decline, and a decrease immediately following an expansion. Analytical models

describing these effects are presented in the Appendix. These models used the simulated values

of B at equilibrium before the population size changes to predict the apparent B values at the

ends of the periods of size change (see the Methods and Appendix). It can be seen from Figure 4

that there is good agreement between these predictions and the simulation results.

Because several demographic estimation methods are based on fitting a demographic

model to the SFS, it is also of interest to understand whether BGS can skew the SFS to different

extents under different demographic models. Although it is well known that BGS causes a skew

of the SFS towards rare variants under equilibrium models (Charlesworth et al. 1995; Nicolaisen

and Desai 2013), the effect of BGS on the SFS with population size change has not been much

explored (but see Johri *et al.* 2020; Torres *et al.* 2020). As shown in Figure 5, with a population

size decline, the SFS of derived alleles is more skewed towards rare variants when BGS is

operating, especially when B is initially small, since the effects of BGS work in opposition to the

effects of the population size reduction. This difference in the left skew of the SFS with and

without BGS is much less noticeable in the case of population expansion, since here the effects

of BGS and the expansion act in a similar direction.

As with the estimates of the apparent B values discussed above, analytical predictions of

the expected SFS after an instantaneous / exponential change in population size can be made,

using the values of B at equilibrium before the population size change using the formulae of

Polanski and Kimmel (2003) and Polanski *et al.* (2003) for the purely neutral case, as described

in the Methods section. Although the shape of the SFS is affected by both demography and BGS,

the impact of BGS is often comparatively small. Figure 5 shows that the overall shape of the SFS

is predicted reasonably well by the analytical results, although deviations are to be expected for

the rare allele classes, which are the most sensitive to demographic change and selection.

Overall, the results imply that BGS is more likely to bias demographic inference post-decline

compared with post-increase, consistent with the performance of the methods described above.

However, it should be noted that the exact patterns observed will depend on the timing of

population size changes relative to the time of sampling, as well as the value of B prior to the

size change. The patterns described here thus represent only a small subset of the possibilities. In

addition, all else being equal, in populations with large long-term population sizes (and thus low

values of B), BGS would be expected to result in even larger biases than those observed here.

A potential solution: averaging across all possible DFEs

As shown above, demographic inference can be strongly affected by uncorrected for BGS effects, as well as by direct purifying selection. A potential solution is thus to correct for these effects when performing inference of population history. A widely-used approach to estimating direct selection effects, DFE-alpha, takes a stepwise approach to inferring demography, by using a presumed neutral class (synonymous sites); conditional on that demography, it then estimates the parameters of the DFE (Keightley and Eyre-Walker 2007; Eyre-Walker and Keightley 2009; Schneider et al. 2011; Keightley and Jackson 2018). However, this approach does not include the possibility of effects of selection at linked sites, which can result an over-estimate of population growth, and while the DFE may not be mis-inferred strongly (Kim et al. 2017), there is substantial mis-inference of the DFE if synonymous sites experience direct selection (Johri et al. 2020).

Building on this idea, Johri et al. (2020) recently proposed an approach that includes both direct and background effects of purifying selection, simultaneously inferring the deleterious DFE and demography. By utilizing the decay of BGS effects around functional regions, they demonstrated high accuracy under the simple demographic models examined. Moreover, the method makes no assumptions about the neutrality of synonymous sites, and can thus be used to estimate selection acting on these sites, as well as in non-coding functional elements. However, this computationally-intensive approach is specifically concerned with jointly inferring the DFE and demographic parameters. As such, if an unbiased characterization of the population history is the sole aim, this procedure may be needlessly involved. We thus here examine the possibility of instead treating the DFE as an unknown nuisance parameter, averaging across all possible DFE shapes, in order to assess whether demographic inference may be improved simply by correcting for these selection effects without inferring their underlying parameter values. This approach utilizes functional (*i.e.*, directly selected) regions, a potential advantage in populations for which only coding data may be available (*e.g.*, exome-capture data; see Jones and Good 2016), or more generally in organisms with largely functional genomes.

In order to illustrate this approach, a functional genomic element was simulated under demographic equilibrium, 2-fold exponential population growth and 2-fold exponential population decline with four different DFE shapes (as described previously, and shown in Figure 6). A number of summary statistics were calculated (see Methods) for the entire functional

region. Inference was first performed assuming strict neutrality, and inferring a one-epoch size

change (thus estimating the ancestral (N_{anc}) and current population sizes (N_{cur})). As was found

with the other inference approaches examined, population sizes were underestimated and a false

inference of population growth was observed in almost all cases when selective effects are

ignored (Figure 6).

Next, the assumption of neutrality was relaxed, and mutations were simulated with fitness

effects characterized by a discrete DFE, with the same fitness classes given above (f_0, f_1, f_2, f_3).

Values for f_i were drawn from a uniform prior between 0 and 1, such that $\sum f_i = 1$. Note that no

assumptions were made about which sites in the genomic region were functionally important, or

regarding the presence/absence of a neutral class. These directly selected sites were then used to

infer demographic parameters. We found that, by varying the shape of the DFE, averaging across

all realizations, and estimating only parameters related to population history, highly accurate

inference of modern and ancestral population sizes is possible (Figure 6). These results

demonstrate that, even if the true DFE of a population is unknown (as will always be the case in

reality), it is possible to infer demographic history with reasonable accuracy by approximately

correcting for these selective effects.

This proposed method is most applicable to organisms in which recombination rates are

reasonably well known. If the assumed recombination rate is 2-fold lower than the true rate, the

ABC approach infers growth by over-estimating the current population size; correspondingly, if

the assumed recombination rate is higher than the true rate, the current population size is under-

estimated (Supp Figure 19). Interestingly, in both cases the ancestral population sizes are

correctly inferred, consistent with previous results (Johri *et al*. 2020).

CONCLUSION

While commonly used approaches for inferring demography assume neutrality and independence

among segregating sites, it is very difficult to verify those assumptions empirically. In addition,

there is considerable evidence for wide-spread effects of selection on linked sites in many

commonly studied organisms (Hernandez et al. 2011; Cutter and Payseur 2013; Williamson et al.

2014; Elyashiv et al. 2016; Campos et al. 2017; Booker and Keightley 2018; Pouyet et al. 2018;

Torres et al. 2018; Castellano et al. 2020). Accordingly, we have explored how violations of the

assumption of neutrality may affect demographic inference, particularly with regard to the

underlying strength of purifying selection and the genomic density of directly selected sites.

Generally speaking, the neglect of these effects (*i.e.*, background selection) results in an

inference of population growth, with the severity of the growth model roughly scaling with

selection strength and density. Thus, when the true underlying model is in fact growth,

demographic mis-inference is not particularly severe; when the true underlying model is constant

size or decline, the mis-inference can be extreme, with a many-fold underestimation of

population size.

It is important to note that BGS effects extend to genomic distances in a way that is

positively related to the strength of purifying selection. For instance, strongly and moderately

deleterious mutations affect patterns of diversity at large genomic distances, whereas mildly

deleterious mutations primarily skew allele frequencies at adjacent sites. Thus, if intergenic

regions further away from exons are used to perform demographic inference, it is predominantly

strongly deleterious mutations that are likely to bias inferences. In contrast, if synonymous sites

are used to infer demographic history, mildly deleterious mutations will be most important. Thus,

as we here focused on relatively sparsely-coding genomes (resembling human-like gene

densities) and used intergenic sites for inference, moderately and strongly deleterious mutations

resulted in more severe mis-inference. In addition, the effect of decay of BGS due to mildly

deleterious mutations depends on multiple parameters. For instance, with an exon of length 500

bp and *Drosophila*-like parameters (*e.g.,* $N_e = 10^6$; recombination rate = mutation rate = 10^{-8} /

site / generation), *B* will increase from 0.53 (at 10 bases from the end of the exon) to 0.94 at a

distance of 1000 bases. On the other hand, with human-like parameters ($N_e = 10^4$; recombination

rate = mutation rate = 10^{-8} / site / generation) the corresponding change in *B* is only 0.981 to

0.982 (Supp Table 4). Thus, mildly deleterious mutations will have drastically different effects in

this regard, depending on the underlying population parameters. Hence, a large effect of

selection is expected when such approaches are applied to synonymous sites in smaller and more

compact genomes with higher gene densities, with a larger input of deleterious mutations at

closely linked sites.

Comparing the two inference methods investigated here, it appears that *fastsimcoal2* is

less prone to inferring false fluctuations in population size. However, both methods falsely infer

growth in the presence of BGS, with increasing severity as the density of coding regions

increases. The times of population growth inferred by both methods appear to be affected in

unpredictable ways when the inferred model is incorrect. When the general model is correctly

identified, BGS leads to inference of more recent growth, and more ancient decline, than the

reality. Further, we observed little difference in performance when using all SNPs relative to

when SNPs are thinned to be separated by 5kb (Supp Figure 20), presumably because the models

investigated do not generate strong LD. Overall, the degree of mis-inference caused by a neglect

of BGS is largely similar between the two methods. Although variation in mutation and

recombination rates across the genome alone did not strongly affect demographic inference by

either method, our evaluations in the current study are restricted to a specific parameter space

resembling those of human populations. The effects of this variation on organisms with more

extreme rate fluctuations remain in need of investigation.

It is noteworthy that, even when all sites are strictly neutral or only 5% of the genome

experiences direct selection, demographic equilibrium is mis-estimated by MSMC as a series of

size changes. The pattern of these erroneous size changes lend a characteristic shape to the

MSMC curve (*i.e.*, ancient decline and recent growth) which appears to resemble the

demographic history previously inferred for the Yoruban population (Schiffels and Durbin

2014), including the time at which changes in population size occurred (Supp Figure 21).

Previous work has demonstrated that the resulting demographic model does not in fact fit the

observed SFS in the Yoruban population (Beichman et al. 2017; Lapierre et al. 2017). A similar

shape has also been inferred in the vervet subspecies (Warren *et al.* 2015; Figure 4), in passenger

pigeons (Hung *et al.* 2014; Figure 2), in elephants (Palkopoulou *et al.* 2018; Figure 4), in

Arabidopsis (Fulgione *et al.* 2018; Figure 3), and in grapevines (Zhou *et al.* 2017; Figure 2A).

Although the inferred population size fluctuations under simulated neutrality are only

~1.2-fold, in most empirical applications the fluctuations are of a somewhat larger magnitude (~

2-fold in pigeons, *Arabidopsis*, and grapevines). Nonetheless, this neutral performance of

MSMC under demographic equilibrium is concerning, and adds to the other previously published

cautions concerning the interpretation of MSMC results. For example, Mazet *et al*. (2016) and

Chikhi *et al*. (2018) demonstrated that, under constant population size with hidden structure,

MSMC may suggest false size changes (see also Orozco-terWengel 2016). In addition, MSMC

has been reported to falsely infer growth prior to instantaneous bottlenecks (Bunnefeld et al.

2015). In addition, we observed that if insufficient genomic data is utilized, or more than one

diploid genome is used to perform inference, MSMC falsely infers recent growth of varying

magnitudes (as previously observed by Beichman et al. 2017).

In sum, we find that the effects of purifying and background selection result in similar

demographic mis-inference across approaches, and that masking functional sites does not yield

accurate parameter estimates. In order to side-step many of these difficulties, our proposed

approach of inferring demography by averaging selection effects across all possible DFE shapes

within an ABC framework appears to be promising. Utilizing only functional regions, we found

a great improvement in accuracy, without making any assumptions regarding the true underlying

shape of the DFE or the neutrality of particular classes of sites. As such, this approach represents

a more computationally efficient avenue if only demographic parameters are of interest, and

ought to be particularly useful in the great majority of organisms in which unlinked neutral sites

either do not exist, or are difficult to identify and verify.

METHODS

Simulations: Simulations were performed using SLiM 3.1 (Haller and Messer 2019) with 10

replicates per evolutionary scenario. For every replicate, 22 chromosomes of 150Mb each were

simulated, totaling ~3 Gb of information per individual genome (similar to the amount of

information in a human genome). Within each chromosome, 3 different types of regions were

simulated, representing non-coding intergenic, intronic, and exonic regions. Based on the NCBI

RefSeq human genome annotation, downloaded from the UCSC genome browser for hg19

(http://genome.ucsc.edu/; Kent *et al.* 2002), mean values of exon sizes and intron numbers per

gene were calculated. To represent mean values for the human genome (Lander et al. 2001), each

gene was comprised of 8 exons and 7 introns, and exon lengths were fixed at 350 bp. By varying

the lengths of the intergenic and intronic regions, three different genomic configurations with

varying densities of functional elements were simulated and compared - with 5%, 10% and 20%

of the genome being under direct selection - hereafter referred to as genome5, genome10, and

genome20, respectively. Genome5 was comprised of introns of 3000 bp and intergenic sequence

of 31000 bp, genome10 of introns of 1500 bp and intergenic sequence of 15750 bp, while

genome20 was comprised of introns of 600 bp and intergenic sequence of 6300 bp. The total

chromosome sizes of these genomes were approximately 150 Mb (150,018,599 bp, 150,029,949

bp, 150,003,699 bp in genome5, 10, and 20, respectively) with 2737, 5164, and 11278 genes per

chromosome in genome5, 10 and 20, respectively. In order to be conservative with respect to the

performance of existing demographic estimators, intronic and intergenic regions were assumed

to be neutral.

Recombination and mutation rates were assumed to be equal at 1×10^{-8} /site / generation.

Neither crossover interference (see the discussion in Campos and Charlesworth 2019) nor gene

conversion were modeled. Exonic regions in the genomes experienced direct purifying selection

given by a discrete DFE comprised of 4 fixed classes (Johri et al. 2020), whose frequencies are

denoted by f_i: f_0, with $0 \leq 2N_e s < 1$ (*i.e.*, effectively neutral mutations), f_1, with $1 \leq 2N_e s < 10$

(*i.e.*, weakly deleterious mutations), f_2, with $10 \leq 2N_e s < 100$ (*i.e.*, moderately deleterious

mutations), and f_3, with $100 \leq 2N_e s < 2N_e$ (*i.e.*, strongly deleterious mutations), where N_e is the

effective population size and s is the reduction in fitness of the mutant homozygote relative to

wild-type. Within each bin, the distribution of s was assumed to be uniform. All mutations were

assumed to be semi-dominant. In all cases, the N_e corresponding to the DFE refers to the

ancestral effective population size. Six different types of DFE were simulated: DFE1: a DFE

skewed largely towards mildly deleterious mutations, given by f_0=0.1, f_1=0.7, f_2=0.1, f_3=0.1;

DFE2: a DFE skewed towards moderately deleterious mutations, f_0=0.1, f_1=0.1, f_2=0.7, f_3=0.1;

DFE3: a DFE skewed towards strongly deleterious mutations, f_0=0.1, f_1=0.1, f_2=0.1, f_3=0.7;

DFE4: a DFE with equal proportions of all mutations, f_0=0.25, f_1=0.25, f_2=0.25, f_3=0.25;

DFE5: a DFE with equal proportions of neutral and strongly deleterious mutations, f_0=0.5,

f_1=0.0, f_2=0.0, f_3=0.5; and DFE6: a DFE with a majority of neutral mutations and a minority of

strongly deleterious mutations, f_0=0.7, f_1=0.0, f_2=0.0, f_3=0.3.

Three different demographic models were tested for each of these DFEs (Supp Table 1):

1) demographic equilibrium, 2) recent exponential 30-fold growth, resembling that estimated for

the human CEU population (Gutenkunst et al. 2009), and 3) ~6-fold instantaneous decline,

resembling the out-of-Africa bottleneck in humans (Gutenkunst et al. 2009). For simulations of

demographic equilibrium and decline, population sizes and time of change were scaled down by

a factor of 10 (with corresponding scaling of the recombination rate, mutation rate, and selection

coefficients), while simulations of growth were not scaled.

Running MSMC: In order to quantify the effect of purifying selection on demographic

inference, we used entire chromosomes generated by SLiM to generate input files for MSMC.

For comparison, and in order to quantify the effect of BGS alone on demographic inference, we

masked the exonic regions to generate input files. For all parameters, MSMC was performed on

a single diploid genome, as the results for this case were the most accurate (Supp Figure 1). Input

files were made using the script ms2multihetsep.py provided in the msmc-tools-Repository

downloaded from https://github.com/stschiff/msmc-tools. MSMC1 and 2 were run as follows:

msmc_1.1.0_linux64bit -t 5 -r 1.0 -o output_genomeID input_chr1.tab input_chr2.tab ...

input_chr22.tab. Population sizes obtained from MSMC were plotted up to the maximum

number of generations obtained from MSMC, and the final value of the ancestral population size

was extended indefinitely as a dashed line.

Running Fastsimcoal2: In order to minimize the effects of linkage disequilibrium (LD), only

SNPs separated by 5 kb were used for inference, following Excoffier et al. (2013). Inference was

also performed by including all SNPs in order to assess the impact of violating the assumption of

no LD on inference. Site frequency spectra (SFS) were obtained for both sets of SNPs for all 10

replicates of every combination of demographic history and DFE. SNPs from all 22

chromosomes were pooled together to calculate the SFS. *Fastsimcoal2* was used to fit each SFS

to 4 distinct models: (a) equilibrium, which estimates only a single population size parameter

(N); (b) instantaneous size change (decline/growth), which fits 3 parameters - ancestral

population size (N_{anc}), current population size (N_{cur}), and time of change (T); (c) exponential size

change (decline/growth), which also estimates 3 parameters - N_{anc}, N_{cur} and T; and (d) an

instantaneous bottleneck model with 3 parameters – N_{anc}, intensity, and time of bottleneck. The

parameter search ranges for both ancestral and current population sizes in all cases were

specified to be uniformly distributed between 100-500000 individuals, while the parameter range

for time of change was specified to be uniform between 100-10000 generations in all models.

The intensity of the bottleneck, as specified by the population size during the reduction, was also

sampled uniformly from the range 100-500000 individuals. The following command line was

used to run *fastsimcoal2*:

fsc26 -t demographic_model.tpl -n 150000 -d -e demographic_model.est -M -L 50 -q,

Model selection was performed as recommended by Excoffier et al. (2013). For each

demographic model, the maximum of maximum likelihoods from all replicates was used to

calculate the Akaike Information Criterion (AIC) = 2 × number of parameters – 2 ×

ln(likelihood). The relative likelihoods (Akaike's weight of evidence) in favor of the ith model

was then calculated by:

562
$$w(i) = \frac{e^{-0.5\Delta_i}}{\sum_{j=1}^{4} e^{-0.5\Delta_j}}$$

where $\Delta_i = AIC_i - AIC_{min}$. The model with the highest relative likelihood was selected as the

best model, and the parameters estimated using that model were used to plot the final inferred

demography.

Simulations of variable recombination and mutation rates, and repeat masking: In order to

simulate variation in recombination and mutation rates, all 22 chromosomes were simulated by

mimicking chromosome 6 (~171Mb) of the human genome. Recombination rates (HapMap)

obtained from Yoruban populations (McVean et al. 2004; Myers et al. 2005) were obtained from

the UCSC genome browser, while the mutation rate map

(https://molgenis26.target.rug.nl/downloads/gonl_public/mutation_rate_map/release2/) was

assumed to correspond to estimates obtained from *de novo* mutations (Francioli et al. 2015), as in

Castellano *et al.* (2020). Absolute values of mutation rates were normalized in order to maintain

the mean mutation rate across the genome at ~ 1.0×10^{-8} per site per generation. Recombination

and mutation rate estimates were taken from positions of approximately 10 Mb to 160 Mb, with

the recombination map starting at 10010063 bp and the mutation map starting at 10010001 bp.

Regions with missing data for either of the two estimates were simulated with rates

corresponding to the previous window, except for the case of centromeres in which no

recombination was assumed. In order to understand the effect of excluding centromeric regions

in empirical studies, the 4Mb region corresponding to the centromere was masked,

corresponding to 48.5 to 52.5 Mb of the simulated 150Mb chromosomes. In order to evaluate the

effect of masking repeat regions, random segments comprising 10% of each chromosome were

masked. The lengths of these segments were drawn from the lengths of repeat regions found in

the human genome (Supp Figure 12), as obtained from the repeat regions in the *hg19* assembly

of the human genome from the UCSC genome browser.

Performing inference by approximate Bayesian computation (ABC): ABC was performed

using the R package "abc" (Csilléry et al. 2010), and non-linear regression aided by a neural net

(used with default parameters as provided by the package) was used to correct for the

relationship between parameters and statistics (Johri et al. 2020). To infer posterior estimates, a

tolerance of 0.1 was applied (*i.e.*, 10% of the total number of simulations were accepted by ABC

in order to estimate the posterior probability of each parameter). The weighted medians of the

posterior estimates for each parameter were used as point estimates. ABC inference was

performed under two conditions: (1) complete neutrality, or (2) the presence of direct purifying

selection. In both cases only 2 parameters were inferred - ancestral (N_{anc}) and current (N_{cur})

population sizes. However, in scenario 2, the shape of the DFE was also varied. Specifically, the

parameters f_0, f_1, f_2, and f_3 were treated as nuisance parameters and were sampled such that $0 \leq f_i$

≤ 1, and $\Sigma_i f_i = 1$, for $i = 0$ to 3. In addition, in order to limit the computational complexity

involved in the ABC framework, values of f_i were restricted to multiples of 0.05 (*i.e.*, $f_i \in \{0.0,$

$0.05, 0.10, \ldots, 0.95, 1.0\}$ ∀ i), which allowed us to sample 1,771 different DFE realizations.

Simulations were performed with functional genomic regions, and the demographic model was

characterized by 1-epoch changes in which the population either grows or declines exponentially

from ancestral to current size, beginning at a fixed time in the past.

For the purpose of illustration, and for a contrast with the human-like parameter set

above, parameters for ABC testing were selected to resemble those of *D. melanogaster* African

populations. Priors on ancestral and current population sizes were drawn from a uniform

distribution between 10^5-10^7 diploid individuals, while the time of change was fixed at 10^6

($\sim N_e$) generations. In order to simulate functional regions, 94 single-exon genes, as described in

Johri *et al*. (2020) and provided in

https://github.com/paruljohri/BGS_Demography_DFE/blob/master/DPGP3_data.zip, were

simulated with recombination rates specific to those exons (https://petrov.stanford.edu/cgi-

bin/recombination-rates_updateR5.pl) (Fiston-Lavier et al. 2010; Comeron et al. 2012). Mutation

rates were assumed to be fixed at 3×10^{-9} per site per generation (Keightley et al. 2009;

Keightley et al. 2014).

All parameters were scaled by the factor 320 in order to decrease computational time,

using the principle first described by Hill and Robertson (1966), and subsequently employed by

others (Comeron and Kreitman 2002; Hoggart et al. 2007; Kaiser and Charlesworth 2009; Kim

and Wiehe 2009; Uricchio and Hernandez 2014; Campos and Charlesworth 2019). The scaled

population sizes thus ranged between ~300–30000 and were reported as scaled values in the

main text. One thousand replicate simulations were performed for every parameter combination

(N_{anc}, N_{cur}, f_0, f_1, f_2, f_3); for performing ABC inference, 50 diploid genomes were randomly

sampled without replacement, and summary statistics were calculated using pylibseq 0.2.3

(Thornton 2003). The following summary statistics were calculated across the entire exonic

region for every exon: nucleotide site diversity (π), Watterson's θ, Tajima's D, Fay and Wu's H

(both absolute and normalized), number of singletons, haplotype diversity, LD-based statistics

(r^2, D, D'), and divergence (*i.e.*, number of fixed mutations per site per generation after the burn-

in period). Means and variances (between exons) of all of the above (a total of 22) were used as

final summary statistics to perform ABC. As opposed to the above examples, in this inference

scheme only exonic data (*i.e.*, directly selected sites) were utilized. Test datasets were generated

in exactly the same fashion as described above.

Analytical expectations for the relative site frequencies: To compute the expected relative

frequencies of site frequency classes, the approach of Polanski and Kimmel (2003) was

followed. They describe a method for computing the "probability that a SNP has b mutant

bases", which is equivalent to the expected site frequency spectrum (SFS) of derived variants.

This method (their equations 3-10) allows for the specification of arbitrary population size

histories and sample sizes. For reasons of computational precision, a sample size of 10 diploid

genomes was chosen. The demographic scenarios detailed in Supplementary Table 1 were

implemented as piecewise functions of the effective population size (counting haploid genomes),

and the effect of BGS was included by scaling these functions by values of B before population

size change as obtained from the forwards-in-time simulations described above. A Mathematica

notebook detailing these results, will be made available upon publication (see data availability

statement). In addition, analytical expressions can be obtained for pairwise diversity values when

there are step changes or exponential growth in population size, as described in the Appendix

and in an example program that calculates diversity values after exponential growth.

Data availability: The following data will be made publicly available upon acceptance of the

manuscript: (1) Scripts used to perform simulations; (2) Input files used to run *fastsimcoal2*; (3)

Scripts used for plotting; (4) Plotted results of MSMC and *fastsimcoal2* for all models and

scenarios tested in this work; (5) A Mathematica (version 12.1) notebook detailing calculations

of analytical expectations for the relative SFS; (6) An example program (Fortran script)

demonstrating how to obtain analytical expressions for values of B after exponential growth. All

supplemental files will be made publicly available at

https://github.com/paruljohri/demographic_inference_with_selection.

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APPENDIX

There are two simple scenarios for population size change for which simple explicit expressions for the

expected pairwise coalescent time or diversity can be obtained, without using the methodology of

Polanski and Kimmel (2003) and Polanski *et al.* (2003) – a step change in N or an exponential growth

in N. First consider the coalescent process for a step change, where the current and initial effective

population sizes are denoted by N_{e1} and N_{e0}, respectively. Let B be the background selection parameter

at the start of the process of change, corresponding to effective size N_{e0}. For convenience, time is

scaled in units of $2N_{e1}$ generations, and the time of the change in population size on this scale is

denoted by T_0, counting back from the present time, $T = 0$. T_0 is assumed to be sufficiently small that B

remains approximately constant during the period since the change in size. Denote the ratio N_{e0}/N_{e1} by

R. The derivation for the case of a step change in population size is similar to that given by Pool and

Nielsen (2009) for the purpose of comparing X chromosomes and autosomes.

Between times T and T_0, coalescence occurs at a rate B^{-1} on the chosen timescale, so that the

contribution from this period to the net coalescent time for a pair of alleles sampled at $T = 0$ is:

$$B^{-1} \int_0^{T_0} T \exp(-B^{-1}T) \, dT = B - B \exp(-B^{-1}T_0) - T_0 \exp(-B^{-1}T_0)$$

There is a probability of $\exp(-B^{-1}T_0)$ that there is no coalescence when T lies between 0 and T_0,

after which coalescence occurs at a rate $1/BR$, giving a net contribution to the coalescence time of:

$$(BR + T_0) \exp(-B^{-1}T_0)$$

The net coalescence time for the stepwise change with BGS is given by the sum of these

two expressions:

$$B[1 + (R - 1) \exp(-B^{-1}T_0)] \qquad \text{(1a)}$$

If this expression is compared to the corresponding equation with $B = 1$, the apparent value of B

at the time of sampling of the pair of alleles is given by:

$$B_S = \frac{B[1+(R-1)\exp(-B^{-1}T_0)]}{[1+(R-1)\exp(-T_0)]} \qquad \text{(1b)}$$

Next, consider a process of exponential change in population size, starting at an initial effective

size of N_{e0} at t_0 generations in the past and ending at size N_{e1}, such that the instantaneous growth rate r

per generation is r=equal to $\ln(N_{e1}/N_{e0})/t_0$. The effective population size at time t in the past is $N_e(t) =$

$N_{e1}\exp(-rt)$; with BGS, the rate of coalescence at time t is $1/BN_e(t)$. As before, the BGS parameter is

assumed to remain constant over the period of population size change. It follows that the probability of

no coalescence by generation t in the past is:

$$P_{nc}(t) = \exp\left[-\int_0^t (2BN_{e1})^{-1} \exp(rt)\,dt\right] = \exp[c^{-1}(1 - e^{rt})] \qquad (2)$$

where $c = 2BN_{e1}r$.

The pre-growth period with $t > t_0$ contributes an expected coalescent time of

$(2BN_{e0} + t_0)P_{nc}(t_0)$, on the scale of generations.

Following Slatkin and Hudson (1991), to obtain the contribution from the period with $t >$

t_0, it is convenient to measure time as $\tau = rt$. The probability of coalescence between τ and $\tau +$

$d\tau$ is then given by:

$$P_c(\tau) = c^{-1}e^\tau \exp[c^{-1}(1 - e^\tau)]\,d\tau \qquad (3)$$

716   The contribution from this period to the expected coalescent time is given by the integral

717   of $\tau\,P_c(\tau)$ between 0 and τ_0. Following Slatkin and Hudson (1991), by transforming to $u =$

718    $\exp(\tau)$, this contribution can be expressed as the following integral:

$$\bar{\tau}_1 = c^{-1}e^{c^{-1}} \int_1^{u_0} \ln(u)e^{-uc^{-1}}\,du \qquad (4)$$

722   This integral can easily be evaluated numerically. The corresponding mean coalescent

723   time on the scale of generations is obtained by division by r, and the result can be added to

724    $(2BN_{e0} + t_0)P_{nc}(t_0)$, yielding the net expected coalescent time. By dividing the resulting

725   expression by the corresponding expression with $B = 1$, the apparent BGS effect at the time of

726   sampling can be obtained, in the same way as for the step change model.

FIGURES AND TABLES

Table 1: Proportion (f_i) of mutations in each class of the discrete distribution of fitness effects (DFE) simulated in this study.

	f_0	f_1	f_2	f_3
DFE1	0.1	0.7	0.1	0.1
DFE2	0.1	0.1	0.7	0.1
DFE3	0.1	0.1	0.1	0.7
DFE4	0.25	0.25	0.25	0.25
DFE5	0.5	0.0	0.0	0.5
DFE6	0.7	0.0	0.0	0.3

Figure 1: Inference of demography by MSMC (red lines; 10 replicates) and *fastsimcoal2* (blue lines; 10 replicates) with and without BGS, under demographic equilibrium (left column), 30-fold exponential growth (middle column), and ~6-fold instantaneous decline (right column). The true demographic models are depicted as black lines, with the x-axis origin representing the present day. (a) All genomic sites are strictly neutral. (b) Exonic sites experience purifying selection specified by a DFE comprised largely of weakly deleterious mutations (DFE1: $f_0 = 0.1$, $f_1 = 0.7$, $f_2 = 0.1$, $f_3 = 0.1$). (c) Exonic sites experience purifying selection specified by a DFE comprised largely of moderately deleterious mutations (DFE2: $f_0 = 0.1$, $f_1 = 0.1$, $f_2 = 0.7$, $f_3 = 0.1$). (d) Exonic sites experience purifying selection specified by a DFE comprised largely of strongly deleterious mutations (DFE3: $f_0 = 0.1$, $f_1 = 0.1$, $f_2 = 0.1$, $f_3 = 0.7$). Exons represent 20% of the genome, and exonic sites were masked/excluded when performing demographic inference, quantifying the effects of BGS alone. The dashed lines represent indefinite extensions of the ancestral population sizes.

Figure 2: Inference of demography by MSMC (red lines; 10 replicates) and *fastsimcoal2* (blue lines; 10 replicates) in the presence of BGS generated by strongly deleterious mutations. Directly selected sites comprised 20% of the genome and were masked when performing demographic inference. (a) Exons experience purifying selection, with 30% of new mutations being strongly deleterious, and the remaining being neutral (specified by DFE6). (b) Exons experience purifying selection, with 50% of new mutations being strongly deleterious, and the remainder being neutral (specified by DFE5). The true demographic models are given as black lines, with the x-axis origin representing the present day. The dashed lines represent indefinite extensions of the ancestral population sizes.

Figure 3: Inference of demography by MSMC (red lines; 10 replicates) and *fastsimcoal2* (blue lines; 10 replicates) in the presence of BGS with varying proportions of the genome under selection, for demographic equilibrium (left column), exponential growth (middle column), and instantaneous decline (right column). Exonic sites were simulated with purifying selection with all f_i values equal to 0.25 (DFE4), and were masked when performing inference. Directly selected sites comprise (a) 20% of the simulated genome, (b) 10% of the simulated genome, and (c) 5% of the simulated genome. The true demographic models are given by the black lines, with the x-axis origin representing the present day. The dashed lines represent indefinite extensions of the ancestral population sizes.

Figure 4: B is the nucleotide site diversity (π) with BGS relative to its purely neutral expectation (π_0). The results are shown for (a) demographic equilibrium, (b) population growth, and (c) population decline. All cases refer to size changes forward in time, the ancestral B (*i.e.*, B pre-change in population size) is shown in white bars, B post-change in population size is shown in solid gray bars, and the analytical expectations for the post-size change B is shown as red bars. Various DFEs were used: **DFE1**: $f_0 = 0.1$, $f_1 = 0.7$, $f_2 = 0.1$, $f_3 = 0.1$. **DFE2**: $f_0 = 0.1$, $f_1 = 0.1$, $f_2 = 0.7$, $f_3 = 0.1$. **DFE3**: $f_0 = 0.1$, $f_1 = 0.1$, $f_2 = 0.1$, $f_3 = 0.7$. **DFE4**: $f_0 = f_1 = f_2 = f_3 = 0.25$. **DFE5:** $f_0 = 0.5$, $f_1 = 0.0$, $f_2 = 0.0$, $f_3 = 0.5$. **DFE6:** $f_0 = 0.7$, $f_1 = 0.0$, $f_2 = 0.0$, $f_3 = 0.3$. Exonic sites comprised ~10% of the genome, roughly mimicking the density of the human genome.

Figure 5: The site frequency spectrum (SFS) of derived allele frequencies at neutral sites from
10 diploid genomes under (a) demographic equilibrium, (b) population growth, and (c)
population decline, under the same DFEs as shown in Figure 4. The x-axis indicates the number
of sample alleles (out of 20) carrying the derived variant. Exonic sites comprised ~10% of the
genome, roughly mimicking the density of the human genome. The red solid circles give the
values predicted analytically with a purely neutral model, but correcting for BGS by using the *B*
values of the ancestral population (*i.e.*, pre-change in population size) obtained from simulations,
in order to quantify the effective population size.

Figure 6: Comparison of estimates of ancestral (N_{anc}) and current (N_{cur}) population sizes when assuming neutrality *vs* when varying the DFE shape as a nuisance parameter, using an ABC framework. Inference is shown for demographic equilibrium (left column), 2-fold exponential growth (middle column), and 2-fold population decline (right column), for five separate DFE shapes that define the extent of direct purifying selection acting on the genomic segment for which demographic inference is performed: (a) neutrality ($f_0 = 1$, $f_1 = 0$, $f_2 = 0$, $f_3 = 0$), (b) weak purifying selection (DFE1: $f_0 = 0.1$, $f_1 = 0.7$, $f_2 = 0.1$, $f_3 = 0.1$), (c) moderately strong purifying selection (DFE2: $f_0 = 0.1$, $f_1 = 0.1$, $f_2 = 0.7$, $f_3 = 0.1$), (d) strong purifying selection (DFE3: $f_0 = 0.1$, $f_1 = 0.1$, $f_2 = 0.1$, $f_3 = 0.7$), and (e) a DFE in which all classes of mutations are equally frequent (DFE4: $f_0 = f_1 = f_2 = f_3 = 0.25$). In each, the horizontal lines give the true values (black for N_{anc}; and gray for N_{cur}) and the box-plots give the estimated values. Black and gray boxes represent estimates when assuming neutrality, while red boxes represent estimates when the DFE is treated as a nuisance parameter.

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Supplementary Information

Supp Table 1: Parameters underlying the human-like demographic models considered.

	Demographic models	Ancestral population size	Current population size	Time of change in generations
1	Equilibrium	10,000	10,000	NA
2	Exponential growth	1000	30,000	850
3	Instantaneous decline	12,300	2,100	4,750

Supp Table 2: The site frequency spectrum (SFS) obtained from 50 diploid individuals from the simulated data (which was provided to *fastsimcoal2* for inference) and the SFS corresponding to the best model inferred by *fastsimcoal2*, in the case of 30-fold exponential growth under neutrality.

Frequency of SNP (out of 100)	*Fastsimcoal2* best model		Simulated data
	Mean	SD	Mean
1	0.529851	0.007443	0.533701
2	0.144710	0.005779	0.135345
3	0.062517	0.005029	0.059647
4	0.034133	0.003670	0.034636
5	0.021766	0.002589	0.023198
6	0.015518	0.001808	0.017067
7	0.011996	0.001255	0.013212
8	0.009808	0.000862	0.010737
9	0.008347	0.000576	0.009145
10	0.007304	0.000369	0.007903
11	0.006519	0.000214	0.006843
12	0.005909	0.000105	0.006222
13	0.005408	0.000039	0.005552
14	0.004991	0.000064	0.005050
15	0.004640	0.000109	0.004844
16	0.004329	0.000141	0.004410
17	0.004065	0.000172	0.004148
18	0.003838	0.000190	0.003860
19	0.003627	0.000209	0.003663
20	0.003441	0.000221	0.003489
21	0.003273	0.000230	0.003236
22	0.003122	0.000236	0.003155
23	0.002984	0.000240	0.003042
24	0.002854	0.000242	0.002901
25	0.002736	0.000244	0.002807
26	0.002625	0.000243	0.002701
27	0.002536	0.000245	0.002560
28	0.002437	0.000244	0.002451
29	0.002356	0.000244	0.002338
30	0.002273	0.000241	0.002308
31	0.002199	0.000239	0.002150
32	0.002127	0.000237	0.002133
33	0.002067	0.000236	0.002146
34	0.002004	0.000233	0.002032
35	0.001944	0.000230	0.001900

36	0.001891	0.000227	0.001876
37	0.001839	0.000226	0.001894
38	0.001790	0.000221	0.001761
39	0.001741	0.000219	0.001758
40	0.001698	0.000216	0.001709
41	0.001657	0.000213	0.001663
42	0.001616	0.000212	0.001645
43	0.001580	0.000210	0.001583
44	0.001543	0.000205	0.001493
45	0.001508	0.000203	0.001524
46	0.001475	0.000201	0.001444
47	0.001439	0.000197	0.001480
48	0.001414	0.000196	0.001398
49	0.001386	0.000193	0.001367
50	0.001353	0.000191	0.001323
51	0.001330	0.000189	0.001380
52	0.001302	0.000186	0.001284
53	0.001274	0.000182	0.001237
54	0.001254	0.000181	0.001213
55	0.001232	0.000178	0.001239
56	0.001210	0.000177	0.001180
57	0.001189	0.000175	0.001181
58	0.001167	0.000172	0.001176
59	0.001147	0.000170	0.001131
60	0.001127	0.000169	0.001135
61	0.001108	0.000167	0.001097
62	0.001092	0.000165	0.001115
63	0.001075	0.000163	0.001135
64	0.001057	0.000161	0.000993
65	0.001039	0.000160	0.001078
66	0.001027	0.000158	0.001054
67	0.001011	0.000156	0.001010
68	0.000991	0.000154	0.000995
69	0.000978	0.000152	0.000986
70	0.000964	0.000150	0.000946
71	0.000952	0.000150	0.000927
72	0.000937	0.000147	0.000922
73	0.000926	0.000146	0.000966
74	0.000910	0.000144	0.000898
75	0.000897	0.000143	0.000916
76	0.000888	0.000141	0.000910
77	0.000877	0.000140	0.000844
78	0.000866	0.000139	0.000866

79	0.000853	0.000137	0.000869
80	0.000845	0.000136	0.000810
81	0.000831	0.000135	0.000819
82	0.000824	0.000134	0.000792
83	0.000812	0.000132	0.000753
84	0.000801	0.000131	0.000735
85	0.000793	0.000130	0.000803
86	0.000783	0.000128	0.000790
87	0.000775	0.000128	0.000767
88	0.000768	0.000127	0.000754
89	0.000757	0.000125	0.000771
90	0.000749	0.000124	0.000776
91	0.000742	0.000124	0.000737
92	0.000734	0.000123	0.000713
93	0.000727	0.000122	0.000724
94	0.000717	0.000120	0.000724
95	0.000709	0.000119	0.000708
96	0.000702	0.000118	0.000687
97	0.000696	0.000117	0.000674
98	0.000688	0.000116	0.000653
99	0.000682	0.000116	0.000679

Supp Table 3: Inference of ancestral population size (Nanc), current population size (Ncur), and time of change, using instantaneous and exponential size change models in *fastsimcoal2*.

Fold Change	Exponential Model			Instantaneous Model			True Values		
	Ncur	Nanc	Time	Ncur	Nanc	Time	Ncur	Nanc	Time
2x	4414	2134	222	1844	964	485	2000	1000	850
5x	8057	2883	425	3990	1005	488	5000	1000	850
10x	11569	3425	530	6704	1020	522	10000	1000	850
20x	16670	4104	585	11018	1037	544	20000	1000	850
30X	21149	4624	611	14272	1033	567	30000	1000	850

Supp Table 4: Nucleotide diversity in the presence of BGS relative to that under neutrality (B), calculated for a neutral site distance y bases from the end of a gene/exon of length 500 bp. The exon experiences purifying selection with strength $2N_e s=10$, where N_e is the effective population size and s is the reduction in fitness. Shown below is $t=hs$ where h is the dominance coefficient (assumed to be 0.5 here), r is the recombination rate per site per generation, and u is the mutation rate per site per generation. B was calculated using Equation 2 of Johri *et al*. (2020).

N_e	$t (2N_e s =10)$	r	u	$B (y=1)$	$B (y=10)$	$B (y=1000)$
10^4	0.00025	1.00×10^{-8}	1.00×10^{-8}	0.9805	0.9806	0.9820
		1.00×10^{-6}	1.00×10^{-6}	0.5152	0.5312	0.9444
10^6	2.5×10^{-6}	1.00×10^{-8}	1.00×10^{-8}	0.5152	0.5312	0.9445
		1.00×10^{-6}	1.00×10^{-6}	0.4920	0.8227	0.9992

Supp Figure 1: Performance of MSMC under neutrality and demographic equilibrium when using 1, 2, and 4 diploid individuals for inference, for varying chromosome sizes: (a) 10 Mb, (b) 50 Mb, (c) 200 Mb, (d) 1 Gb. The numbers of replicates for each panel varied slightly around 100. MSMC runs with 4 diploid individuals could not be obtained due to the long computational times required. As can be seen, MSMC has a tendency to take a common shape, often falsely indicating recent population growth.

Supp Figure 2: Performance of *fastsimcoal2* under neutrality and demographic equilibrium with varying chromosome sizes: (a) 10 Mb, (b) 50 Mb, (c) 200 Mb, (d) 1 Gb, when different model choices are provided: *Left column*: the correct model of constant population size is specified; *Middle column*: model selection was performed with 3 models – equilibrium, instantaneous size change, and exponential size change; *Right column*: model selection was performed with 4 models – equilibrium, instantaneous size change, exponential size change, and instantaneous bottleneck. Inference was performed using 50 diploid individuals and the inferred population size estimates of the best model are plotted (blue lines). The numbers of replicates for each panel varied slightly around 100. The true model is shown in black.

a

Test Model / Size	Equilibrium	Instantaneous Change	Exponential Change
10 Mb	38	56	6
50 Mb	34	62	4
200 Mb	24	72	4
1 Gb	19	75	6

b

Test Model / Size	Equilibrium	Instantaneous Change	Exponential Change	Instant Bottleneck
10 Mb	33	46	5	16
50 Mb	29	48	2	21
200 Mb	19	51	2	28
1 Gb	11	47	5	37

Supp Figure 3: Model selection for 100 replicates (corresponding to Supp Figure 2) in which the true model was neutral equilibrium. (a) Model selection was performed with 3 models: equilibrium, instantaneous size change, and exponential size change; (b) Model selection was performed with 4 models: equilibrium, instantaneous size change, exponential size change, and instantaneous bottleneck.

Supp Figure 4: Inferred demography from MSMC (red lines) and *fastsimcoal2* (blue lines) in the presence of background selection, with the true DFE shown to the left of the panel, for 2-fold instantaneous decline (right column) and 2-fold exponential growth (left column). In this case, 20% of the genome experienced direct selection. The true demographic models are shown as black lines.

Supp Figure 5: Inference of demography by MSMC (red lines; 10 replicates) and *fastsimcoal2* (blue lines; 10 replicates) under demographic equilibrium (left column), 30-fold exponential growth (middle column), and ~6-fold instantaneous decline (right column) in the presence of direct purifying selection (*i.e.*, directly selected sites are not masked). The true demographic model is depicted in black lines. Exonic sites experience purifying selection specified by the following DFEs: (a) DFE1: $f_0 = 0.1$, $f_1 = 0.7$, $f_2 = 0.1$, $f_3 = 0.1$, (b) DFE2: $f_0 = 0.1$, $f_1 = 0.1$, $f_2 = 0.7$, $f_3 = 0.1$, (c) DFE3: $f_0 = 0.1$, $f_1 = 0.1$, $f_2 = 0.1$, $f_3 = 0.7$, (d) DFE4: $f_0 = 0.25$, $f_1 = 0.25$, $f_2 = 0.25$, $f_3 = 0.25$, (e) DFE5: $f_0 = 0.7$, $f_1 = 0.0$, $f_2 = 0.0$, $f_3 = 0.3$, (f) DFE6: $f_0 = 0.5$, $f_1 = 0.0$, $f_2 = 0.0$, $f_3 = 0.5$. In this case, 20% of the genome was under selection.

True Model	Test Model / DFE Shape	5%				10%				20%			
	Functional Density	Equilibrium	Instantaneous Change	Exponential Change	Instant Bottleneck	Equilibrium	Instantaneous Change	Exponential Change	Instant Bottleneck	Equilibrium	Instantaneous change	Exponential Change	Instant Bottleneck
Equilibrium	DFE0	0	10	0	0	0	10	0	0	0	10	0	0
	DFE1	0	7	3	0	0	8	2	0	0	10	0	0
	DFE2	0	9	1	0	0	10	0	0	0	10	0	0
	DFE3	0	9	1	0	0	10	0	0	0	0	10	0
	DFE4	0	10	0	0	0	9	1	0	0	10	0	0
	DFE5	0	10	0	0	0	10	0	0	0	0	10	0
	DFE6	0	7	3	0	0	10	0	0	0	9	1	0
Instantaneous Decline (6x)	DFE0	0	10	0	0	0	10	0	0	0	10	0	0
	DFE1	0	10	0	0	0	10	0	0	0	10	0	0
	DFE2	0	10	0	0	0	10	0	0	0	10	0	0
	DFE3	0	10	0	0	0	10	0	0	0	9	1	0
	DFE4	0	10	0	0	0	10	0	0	0	10	0	0
	DFE5	0	10	0	0	0	10	0	0	0	10	0	0
	DFE6	0	10	0	0	0	10	0	0	0	10	0	0
Exponential Growth (30x)	DFE0	0	0	10	0	0	0	10	0	0	0	10	0
	DFE1	0	0	10	0	0	0	10	0	0	0	10	0
	DFE2	0	0	10	0	0	0	10	0	0	0	10	0
	DFE3	0	0	10	0	0	0	10	0	0	0	10	0
	DFE4	0	0	10	0	0	0	10	0	0	0	10	0
	DFE5	0	0	10	0	0	0	10	0	0	0	10	0
	DFE6	0	0	10	0	0	0	10	0	0	0	10	0
Instantaneous Decline (2x)	DFE0	0	10	0	0	0	10	0	0	0	10	0	0
	DFE1	0	10	0	0	0	10	0	0	0	10	0	0
	DFE2	0	10	0	0	0	10	0	0	0	4	6	0
	DFE3	0	10	0	0	0	10	0	0	0	5	5	0
	DFE4	0	10	0	0	0	10	0	0	0	7	3	0
	DFE5	0	10	0	0	0	10	0	0	0	10	0	0
	DFE6	0	10	0	0	0	10	0	0	0	8	2	0
Exponential Growth (2x)	DFE0	0	10	0	0	0	10	0	0	0	10	0	0
	DFE1	0	10	0	0	0	10	0	0	0	10	0	0
	DFE2	0	10	0	0	0	10	0	0	0	10	0	0
	DFE3	0	10	0	0	0	10	0	0	0	10	0	0
	DFE4	0	10	0	0	0	10	0	0	0	10	0	0
	DFE5	0	10	0	0	0	10	0	0	0	10	0	0
	DFE6	0	10	0	0	0	10	0	0	0	10	0	0

Supp Figure 6: Model selection by *fastsimcoal2* in the presence of BGS, when chosen from four possible models: equilibrium, instantaneous size change, exponential size change, and instantaneous bottleneck. Results are shown when SNPs separated at a distance of 5 kb were used for inference, and when 5%, 10% or 20% of the genomes were exonic.

True Model	Functional Density / Behavior / DFE Shape	5%		10%		20%	
		Growth	Decline	Growth	Decline	Growth	Decline
Equilibrium	DFE0	10	0	10	0	10	0
	DFE1	10	0	10	0	10	0
	DFE2	10	0	10	0	10	0
	DFE3	10	0	10	0	10	0
	DFE4	10	0	10	0	10	0
	DFE5	10	0	10	0	10	0
	DFE6	10	0	10	0	10	0
Instantaneous Decline (6x)	DFE0	0	10	0	10	0	10
	DFE1	0	10	0	10	0	10
	DFE2	0	10	0	10	10	0
	DFE3	0	10	10	0	10	0
	DFE4	0	10	0	10	10	0
	DFE5	0	10	10	0	10	0
	DFE6	0	10	0	10	10	0
Exponential Growth (30x)	DFE0	10	0	10	0	10	0
	DFE1	10	0	10	0	10	0
	DFE2	10	0	10	0	10	0
	DFE3	10	0	10	0	10	0
	DFE4	10	0	10	0	10	0
	DFE5	10	0	10	0	10	0
	DFE6	10	0	10	0	10	0
Instantaneous Decline (2x)	DFE0	0	10	0	10	0	10
	DFE1	0	10	0	10	0	10
	DFE2	0	10	0	10	10	0
	DFE3	0	10	10	0	10	0
	DFE4	0	10	0	10	10	0
	DFE5	0	10	0	10	10	0
	DFE6	0	10	0	10	10	0
Exponential Growth (2x)	DFE0	10	0	10	0	10	0
	DFE1	10	0	10	0	10	0
	DFE2	10	0	10	0	10	0
	DFE3	10	0	10	0	10	0
	DFE4	10	0	10	0	10	0
	DFE5	10	0	10	0	10	0
	DFE6	10	0	10	0	10	0

Supp Figure 7: Effects of BGS on inference of growth or decline by *fastsimcoal2*. Results are shown when SNPs separated by a distance of 5 kb were used for inference, and when 5%, 10% and 20% of the genomes were exonic.

Supp Figure 8: Inference of the timing of 30-fold exponential growth by *fastsimcoal2* in the presence of BGS, when (a) 5%, (b) 10%, and (c) 20% of the genome is exonic (compared to neutrality (DFE0)). The purple line represents the true timing.

Supp Figure 9: Inference of the timing of 2-fold exponential growth by *fastsimcoal2* in the presence of BGS, when (a) 5%, (b) 10%, and (c) 20% of the genome is exonic (compared to neutrality (DFE0)). The purple line represents the true timing.

Supp Figure 10: Inference of the timing of 6-fold instantaneous decline by *fastsimcoal2* in the presence of BGS, when (a) 5%, (b) 10%, and (c) 20% of the genome is exonic (compared to neutrality (DFE0)). The purple line represents the true timing. Boxplots are presented in green if decline was inferred, and in yellow if growth was inferred.

Supp Figure 11: Inference of the timing of 2-fold instantaneous decline by *fastsimcoal2* in the presence of BGS, when (a) 5%, (b) 10%, and (c) 20% of the genome is exonic (compared to neutrality (DFE0)). The purple line represents the true timing. Boxplots are presented in green if decline was inferred, and in yellow if growth was inferred.

Supp Figure 12: Distribution of lengths of repeat regions in the human genomes (*hg19*). Shown above is the distribution of lengths up to 1000 bp, although lengths of repeat regions range between 6 to 160602 bp.

Supp Figure 13: Performance of demographic inference by MSMC (red lines) and *fastsimcoal2* (blue lines) under different scenarios of neutrality, when the true model is equilibrium: (a) there is variation in recombination and mutation rates, (b) there is variation in recombination and mutation rates and the centromeric region is masked, (c) there is variation in recombination and mutation rates, and short regions resembling repeats (comprising 10% of each chromosome) are randomly masked across the genome, and (d) there is variation in recombination and mutation rates, and the centromere as well as small-sized repeats are randomly masked across the genome. The maximum and minimum fold change detected in every scenario is indicated on the upper right corner.

Supp Figure 14 : Performance of demographic inference by MSMC (red lines) and *fastsimcoal2* (blue lines) under different scenarios of neutrality, when the true model is 30-fold exponential growth: (a) there is variation in recombination and mutation rates across the genome, (b) there is variation in recombination and mutation rates, and the centromeric region is masked, (c) there is variation in recombination and mutation rates, and short regions resembling repeats are randomly masked across the genome (comprising of 10% of each chromosome), and (d) there is variation in recombination and mutation rates, and the centromere as well as small-sized repeats are randomly masked across the genome.

Supp Figure 15: Performance of demographic inference by MSMC under different scenarios of neutrality, when the true model is 6-fold instantaneous decline: (a) there is variation in recombination and mutation rates across the genome, (b) there is variation in recombination and mutation rates, and the centromeric region is masked, (c) there is variation in recombination sand mutation rates, and short regions resembling repeats are randomly masked across the genome (comprising of 10% of each chromosome), and (d) there is variation in recombination and mutation rates, and the centromere as well as small-sized repeats are randomly masked across the genome.

Supp Figure 16: Performance of demographic inference by MSMC (red lines) and *fastsimcoal2* (blue lines) in the presence of background selection, under different scenarios when the true model is equilibrium: (a) there is variation in recombination and mutation rates, (b) there is variation in recombination and mutation rates and the centromeric region is masked, (c) there is variation in recombination and mutation rates, and short regions resembling repeats (comprising 10% of each chromosome) are randomly masked across the genome, and (d) there is variation in recombination and mutation rates, and the centromere as well as small-sized repeats are randomly masked across the genome. Exons comprise of 20% of the genome, experience purifying selection given by DFE4 ($f_0 = f_1 = f_2 = f_3 = 0.25$), and are masked when performing inference.

Supp Figure 17: Performance of demographic inference by MSMC (red lines) and *fastsimcoal2* (blue lines) in the presence of background selection, under different scenarios when the true model is 30-fold exponential growth: (a) there is variation in recombination and mutation rates, (b) there is variation in recombination and mutation rates and the centromeric region is masked, (c) there is variation in recombination and mutation rates, and short regions resembling repeats (comprising 10% of each chromosome) are randomly masked across the genome, and (d) there is variation in recombination and mutation rates, and the centromere as well as small-sized repeats are randomly masked across the genome. Exons comprise of 20% of the genome, experience purifying selection given by DFE4 ($f_0 = f_1 = f_2 = f_3 = 0.25$), and are masked when performing inference.

Supp Figure 18: Performance of demographic inference by MSMC (red lines) and *fastsimcoal2* (blue lines) in the presence of background selection, under different scenarios when the true model is a 6-fold instantaneous decline: (a) there is variation in recombination and mutation rates, (b) there is variation in recombination and mutation rates and the centromeric region is masked, (c) there is variation in recombination and mutation rates, and short regions resembling repeats (comprising 10% of each chromosome) are randomly masked across the genome, and (d) there is variation in recombination and mutation rates, and the centromere as well as small-sized repeats are randomly masked across the genome. Exons comprise of 20% of the genome, experience purifying selection given by DFE4 ($f_0 = f_1 = f_2 = f_3 = 0.25$), and are masked when performing inference.

Supp Figure 19: Performance of the ABC method when recombination rate is mis-specified. (a) The true recombination rate is 2-fold higher than that assumed, and (b) the true recombination rate is 2-fold lower than that assumed. Boxplots in green show the posterior estimates when the recombination rate is higher or lower than assumed. For comparison, boxplots in red show the posterior inferred when the corresponding recombination rate is correctly specified. The black line displays the true ancestral population size (N_{anc}) and the gray line represents the true current population size (N_{cur}).

Supp Figure 20: Scenarios in which demographic inference by *fastsimcoal2* resulted in different best models when performed using all SNPs (right column) and when SNPs were thinned to be separated by 5 kb (left column). The best model was defined as the model corresponding to the lowest AIC over all ten replicates. The DFEs are indicated on the left. (a) Direct purifying selection under DFE4 (exonic sites are not masked) in 5% of the genome; (b) Direct purifying selection under DFE5 (exonic sites are not masked) in 5% of the genome; (c) Direct purifying selection under DFE1 (exonic sites are not masked) in 10% of the genome; (d) Direct purifying selection under DFE6 (exonic sites are not masked) in 10% of the genome; (e) BGS under DFE5 (*i.e.*, exonic sites are masked) in which 5% of the genome is exonic; (f) BGS under DFE1 (*i.e.*, exonic sites are masked) in which 20% of the genome is exonic; (g) BGS under DFE2 (*i.e.*, exonic sites are masked) in which 20% of the genome is exonic; (h) BGS under DFE4 (*i.e.*, exonic sites are masked) in which 20% of the genome is exonic.

Supp Figure 21: Inference of demographic history by MSMC. Top panel / red line: simulations in which the true model is constant population size, and 50% of new mutations in exons are strongly deleterious with the remainder being neutral, where exons comprise 5% of the genome. Bottom panel / green line: the empirical estimate of population history of the YRI population inferred with MSMC by Schiffels and Durbin (2014). The x-axis is in years (assuming a generation time of 30 years). Note that the y-axes are on different scales, and the magnitude of change observed in the empirical data is considerably larger in the simulated data. Thus, this comparison is only meant to illustrate this common shape taken in MSMC plots (and see similar shapes in, for example, vervets (Warren *et al.* 2015; Figure 4) and passenger pigeons (Hung *et al.* 2014; Figure 2)).