CHARACTERIZATION OF ANCESTRAL ORIGIN OF CYSTIC FIBROSIS OF PATIENTS WITH NEW REPORTED MUTATIONS IN CFTR

César Paz-y-Miño^{1*}, Ana Karina Zambrano^{1*}, Juan Carlos Ruiz-Cabezas^{234,} Isaac Armendáriz-Castillo¹, Jennyfer M. García-Cárdenas¹, Santiago Guerrero¹, Andrés López-Cortés¹, Andy Pérez-Villa¹, Patricia Guevara-Ramírez¹, Verónica Yumiceba¹, Paola E. Leone¹

¹ Centro de Investigación Genética y Genómica, Facultad de Ciencias de la Salud Eugenio Espejo, Universidad UTE, Av. Mariscal Sucre and Mariana de Jesús, Block I, Quito 170129, Ecuador.

² Universidad de Especialidades Espíritu Santo (UEES). Guayaquil-Ecuador.
 ³ Instituto de Biomedicina, Universidad Católica de Santiago de Guayaquil.
 Guayaquil - Ecuador.

⁴ Instituto Oncológico Nacional de la Sociedad de Lucha Contra el Cáncer (ION-SOLCA). Guayaquil- Ecuador.

* César Paz-y-Miño y Ana Karina Zambrano contributed equally in the paper.

César Paz-y-Miño: cesar.pazymino@ute.edu.ec Ana Karina Zambrano: anazambrano17@hotmail.com Juan Carlos Ruiz-Cabezas: jcruizc@hotmail.com Isaac Armendáriz-Castillo: fabian.armendariz@ute.edu.ec Jennyfer García-Cárdenas: jennyfer.garcia@ute.edu.ec Santiago Guerrero: santiago.guerrero@ute.edu.ec Andrés López-Cortés: andresa.lopez@ute.edu.ec Andy Pérez-Villa: andy.perez@ute.edu.ec Patricia Guevara-Ramírez: alexandra.guevara@ute.edu.ec Verónica Yumiceba: veronica.yumiceba@ute.edu.ec Paola E. Leone: paola.leone@ute.edu.ec

Corresponding Author:

César-Paz-y-Miño Address: Centro de Investigación Genética y Genómica, Facultad de Ciencias de la Salud Eugenio Espejo, Universidad UTE, Av. Mariscal Sucre and Mariana de Jesús, Block I, Quito 170129, Ecuador. Telephone: +593 99 502 6485 E-mail address: <u>cesar.pazymino@ute.edu.ec</u>

Abstract

The incidence of Cystic fibrosis (CF) and the frequency of the variants for CFTR depend on the population; furthermore, CF symptomatology is characterized by obstructive lung disease, pancreatic insufficiency among others, reliant on the individual genotype. Ecuadorian population is a mixture of Native Americans, Europeans, and Africans. That population admixture could be the reason for the new mutations reported in a previous study by Ruiz et al. (2019). A panel of 46 Ancestry Informative Markers was used to estimate the ancestral proportions of each available sample (12 samples in total). As a result, the Native American ancestry proportion was the most prevalent in almost all individuals, except for three patients from Guayaquil with the mutation [c.757G>A:p.Gly253Arg; c.1352G>T:p.Gly451Val] who had the highest European composition.

Keywords: Cystic Fibrosis, Native American, Ecuadorian, CFTR.

Introduction

Cystic fibrosis (CF) is an autosomal recessive disorder that has been extensively studied among populations (1). It is characterized by obstructive lung disease, pancreatic insufficiency, diabetes, liver disease, among others (2). The most frequent worldwide mutation in Cystic Fibrosis Transmembrane Conductance Regulator (CFTR) protein gene is c.1521_1523delCTT (p.Phe508del) (3) which originated between 11 000 and 34 000 years ago in Europeans, then it spread across all Europe (4). CF occurs in 1 out of 2 500 live births with high prevalence in the European ancestry, and the frequency of the heterozygotes has been reported as 1 in 25 in Europeans (5) (6). There are plenty of studies in CF, yet the majority in Europeans, underrepresenting the Latin Americans (4) (6) (7) (8). In the United States, a study reported the CF incidence to be 1 in 9 200 Hispanics and 1 in 10 900 Native Americans, yet the USA has a different population structure to South America (1) (6) (9). In general, in Latin America the incidence is 1 per 6 000 newborn alive; specifically, Ecuador exhibits an incidence of 1 in 11 252 newborns (10) (11) (12).

Ecuadorian population, located in the Northwest of South America is a mixed population conformed by Native Americans, Europeans who arrived in the 16th century during the conquest, and Africans who came with them as slaves. According to the last census, the population projection for 2020 was estimated as 17 510 643 Ecuadorians. Moreover, Ecuadorian self–identified as: "mestizos" 71.9%, "montubios" 7.4%, Afro-Ecuadorian 7.2%, "indígenas" 7%, "blancos" 6.1% and others (0.4%) (13). There are also reports of the Ecuadorian ancestry using AIMs in mestizo population where Native American was the most prevalent ancestry (59.6%), followed by European (28.8%) and lastly African (11.6%) (14) (15).

Like other South American studies, Ecuador is underrepresented in cystic fibrosis research, and none of them involve the comparison of the mutations with the population's origin. Paz-y-Miño et.al (1999) reported 10 cases of Ecuadorian CF patients, at least 60% of the mutations differ from c.1521_1523delCTT (p.Phe508del) (16). Valle et al. (2007), analyzed 62 Ecuadorian CF patients, the most prevalent mutation was F508del (37.1%) (12). The last report by Ortiz et al. (2017), included 48 Ecuadorian individuals with CF, reported F508del with the highest frequency (20.27%)(17). These studies, however, are mainly focused on the particular F508 mutation, revealing that the percentage is not relatively high as in Europeans. The incidence and the frequency of the CF mutation depend on the population under study, Ecuadorians are a mestizo population, and the population's composition is not clear yet.

Here we provide the ancestry origin data of 46 Ancestry Informative Markers of the individuals with the new mutations reported in a previous study of CF patients from Ecuador (18). We aimed to elucidate if the mutations reported are mainly from European ancestry, due to the previous data of the main incidence.

Main Text

Methods

Samples and DNA extraction

Twelve CF patients from Guayaquil (coast) and Cuenca (highland) who were available and had new CFTR disease-causing variants reported in a previous study were selected: one patient from Guayaquil with *c.1473T>A:p.Cys491**; one patient from Guayaquil and two from Cuenca with *c.2672del:p.Asp891Alafs*15*; one patient from Cuenca with *c.1486T>C:p.Trp496Arg*; six patients from Guayaquil and one from Cuenca with [*c.757G>A:p.Gly253Arg; c.1352G>T:p.Gly451Val*], were selected (18). DNA was extracted using Chelex 100 (Bio-Rad) (10%) from peripheral blood samples collected on FTA cards (GE Healthcare Life Sciences) and quantified using NanoDrop (Thermo Scientific). To protect the identity of the individuals, the samples were anonymized.

DNA amplification

PCR amplification of the twelve CF samples and controls (positive: 2800 and negative) was performed using 46 AIMs-INDELs: MID-1470, MID-777, MID-196, MID-881, MID-3122, MID-548, MID-659, MID-2011, MID-2929, MID-593, MID-798, MID-1193, MID-1871, MID-17, MID-2538, MID-1644, MID-3854, MID-2275, MID-94, MID-3072, MID-772, MID-2313, MID-397, MID-1636, MID-51, MID-2431, MID-2264, MID-2256, MID-128, MID-15, MID-2241, MID-419, MID-943, MID-159, MID-2005, MID-250, MID-1802, MID-1607, MID-1734, MID-406, MID-1386, MID-1726, MID-3626, MID-360, MID-1603, MID-2719 (19), in one multiplex reaction and following the standardized protocol of the laboratory. The fragment separation was carried out in 3500 Genetic Analyzers (Applied Biosystems). Data were collected with Data collection v3 and visualized with Gene Mapper v5.

Statistical Analyses

Data were analyzed with Structure v2.3.4 in order to estimate the ancestral proportions in the population, the runs consisted of a burn-in length of 10 000 followed by 10 000 Markov Chain Monte Carlo (MCMC) interactions. The option used was the admixture model ("Use population information to test for migrants"). The cluster considered for the analysis was one to three (k=1, k=2, and k=3) due to the historical background of the Ecuadorian population and according to the cluster identification by Evanno *et al* (20) (14).

Principal component analysis (PCA) was built with RStudio v1.1.453 to visualize the CF individuals' structure: the correlation between individuals under analysis and the reference population from HGDP-CEPH (Native Americans, Europeans, and Africans) subset H952 (19).

Results

The DNA quantification was optimal to perform the PCR (5-20ng/ul). After the amplification, complete profiles were obtained. A total of 339 individuals (reference population and samples), were analyzed, assuming a clustering of three using the information to test for migrants, permuting 10 000 burn-in periods and 10 000 interactions and a bar plot was obtained showing the main ancestral population analyzed (Fig 1).

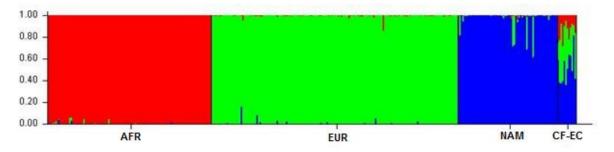
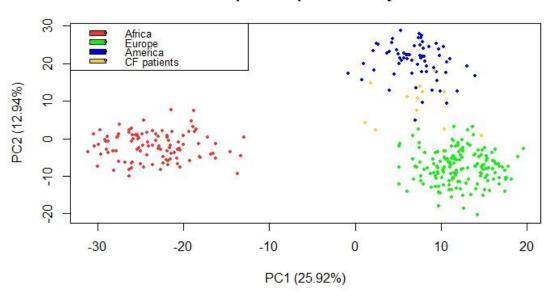


Fig 1.- Bar plot grouped by population identification (AFR: African ancestry, EUR: European ancestry, NAM: Native American ancestry, CF-EC: Ecuadorian Cystic Fibrosis patients). Three inferred clusters (K=3).

Principal component analysis (PCA) results showed the three reference populations clearly differentiate between them. The CF Ecuadorian population is in the middle of them but mainly between European and Native American reference population. The two main principal components represented 38.86% of the total (Fig2).



Principal Component Analysis

Fig 2.- Principal component analysis of cystic fibrosis Ecuadorian patients.

A percentage of the ancestral composition of each individual was obtained, as a result, a heterogeneous percentage was found depending on the individual and the region under study. Hence, clearly showing the admixture of the Ecuadorian population according to history (Table 1). The global ancestry composition of CF patients was: The Native American 50% (standard deviation of 14.03), the European 35% (standard deviation of 15.5) and the African 11.5% (standard deviation of 7.82). The Native American ancestry was the first origin of almost all individuals, except for three patients from Guayaquil with the mutation [c.757G>A:p.Gly253Arg; c.1352G>T:p.Gly451Val] with the highest European composition.

PATIENT	AFR	EUR	NAM	MUTATION	MUTATIONS' REFERENCE
01	24.6	16.7	58.7	c.1473T>A:p.Cys491*	(18)
02	7.6	43.5	48.9	c.2672del:p.Asp891Alafs*15	(18)
03	21.6	14.8	63.6	c.2672del:p.Asp891Alafs*15	(18)
04	16	42.2	41.8	c.2672del:p.Asp891Alafs*15	(18)
05	9.1	8.9	82	c.1486T>C:p.Trp496Arg	(18)
06	22.5	39.6	37.9	[c.757G>A:p.Gly253Arg; c.1352G>T:p.Gly451Val]	(18)

Table 1.- Percentage of the ancestral composition of each individual under study.

07	7.2	55.3	37.5	[c.757G>A:p.Gly253Arg; c.1352G>T:p.Gly451Val]	(18)
08	28.2	31.9	39.9	[c.757G>A:p.Gly253Arg; c.1352G>T:p.Gly451Val]	(18)
09	5	58.7	36.3	[c.757G>A:p.Gly253Arg; c.1352G>T:p.Gly451Val]	(18)
10	11.9	37.2	50.9	[c.757G>A:p.Gly253Arg; c.1352G>T:p.Gly451Val]	(18)
11	9.3	32.1	58.6	[c.757G>A:p.Gly253Arg; c.1352G>T:p.Gly451Val]	(18)
12	11	25.7	63.3	[c.757G>A:p.Gly253Arg; c.1352G>T:p.Gly451Val]	(18)

* AFR: African ancestry, EUR: European Ancestry and NAM: Native American Ancestry.

Discussion

The present study is the first report of the ancestral composition of CF Ecuadorian patients with new CFTR mutations. There are plenty of CF studies among different populations that revealed the differences between gender, age, and symptoms in CF patients. Some studies compared CF patients of different ages and gender describing that the incidence of CF in Europe is higher in children than in adults, approximately 4 CF children per 3 adults and around 1.1 males per each female (21) (22).

There are reports that evidenced CF prevalence, differ depending on ethnicity. For instance, the incidence in Native Americans, whites and black individuals is 37.2, 38.8 and 17.1 per 100 000, respectively (23). Moreover, other studies revealed the incidence among diverse ethnicities: as an example, the prevalence of CF reported by Rohlfs et al. (2011) is 1 in 242 in Asian, 1 in 28 Caucasian, 1 in 59 Hispanic and 1 in 70 Native American (2). That study clearly revealed the ethnic differences in the incidence and the distribution of CF worldwide.

In Ecuador, there are studies about the ancestral origin, for instance, the Ecuadorian was reported to be composed of 59.6% of Native American, 28.8% of European and lastly 11.6% of African origin (14) (15).

In addition to the variable predisposition of CF among populations, reports exhibit a total of 2 063 mutations listed on the CFTR mutation database (24); while in the CFTR2 database, the most recent file updated on 8 December 2017, shows a total of 374 variants (25). Those variants were identified in different populations in diverse frequencies. For instance, the frequencies of the most common variant c.1521_1523delCTT (p.Phe508del) depend on ethnicity, it was reported as 72% in US Caucasians, ~41% in African Americans and 18% in Iranians, yet it also differs among Caucasians (26) (2) (27) (1) (28).

There are some mutations that have been commonly reported in ethnic groups, as examples, c.1624G>T (p.Gly542X) was reported in 43% of Turkish origin (29), while in a study in Peruvian patients the frequency was 6.9% (30); c.3846G>A (p.Trp1282Ter) was reported in 43% of Ashkenazi patients (31); c.2988+1G>A (3120+1G>A) reported in 12.3% of native African CF patients (32) (33); c.3909C>G (p.Asn1303Lys) described in 1.7% of the total number of CF analyzed from Europeans and the United States population (34), while in Algerian population the frequency was 20% (35); c.1652G>A (p.Gly551Asp) presented a frequency of 3% in North Brazilian population (36). Furthermore, some mutations have been found in a specific ethnic group, like, c.16C>G (p.Leu6Val) was found in one Argentinian, c.3294G>C (p.Trp1098Cys) was found in one Mexican, among other variations described (4); c.3276C>G (p.Tyr1092Ter) was found in Jews from Iraq (31) (2).

In conclusion, the identification of ethnicity-dependent mutations would be an important aspect of CF testing in Ecuador. The present study exhibited a greater ancestral composition of Native American, followed by European and lastly African, the mixed population origin could possibly explain the new CF mutations reported.

Limitations

Although we have found the ancestral proportions of the majority CF patient with new mutations previously reported, we could not access all the samples due to the available conditions of the patients. Moreover, a larger CF patients study with the commonly reported mutation should be conducted to better approximation to the ancestral proportions of the patients.

List of Abbreviations

CF: cystic fibrosis

CFTR: Cystic Fibrosis Transmembrane Conductance Regulator

AIMs: Ancestry Informative Markers

INDELs: Insertions Deletions

PCA: Principal Component Analysis

AFR: African ancestry

- EUR: European ancestry
- NAM: Native American ancestry

Declarations

Ethics approval and consent to participate

The patients were registered with the Ecuadorian Cystic Fibrosis Foundation at Guayaquil and Cuenca. Patients' informed consent, including the paternal signed

authorization to participate in the study. The investigation was approved with the number 2018-127E by "Comité de Ética de Investigación en Seres Humanos Universidad San Francisco de Quito".

Consent for publication

Not applicable.

Availability of data

The data is presented in the manuscript.

Competing interests

The authors declare that they have no competing interests.

Funding

The article was not funded.

Authors' contributions

César Paz-y-Miño: coordination and followed up with the development of the article. cesar.pazymino@ute.edu.ec

Ana Karina Zambrano: design, experimental procedure, data analysis and writing. anazambrano17@hotmail.com

Juan Carlos Ruiz-Cabezas: written edition. jcruizc@hotmail.com

Isaac Armendáriz-Castillo: written edition. fabian.armendariz@ute.edu.ec

Jennyfer García-Cárdenas: written edition. jennyfer.garcia@ute.edu.ec

Santiago Guerrero: written edition. santiago.guerrero@ute.edu.ec

Andrés López-Cortés: written edition. andresa.lopez@ute.edu.ec

Andy Pérez-Villa: writing and formatting andy.perez@ute.edu.ec

Patricia Guevara-Ramírez: written edition. alexandra.guevara@ute.edu.ec

Verónica Yumiceba: written edition. veronica.yumiceba@ute.edu.ec

Paola E. Leone: written edition. paola.leone@ute.edu.ec

Acknowledgments

The authors are grateful to the patients for their selfless participation in the present study. To all the people and institutions involved in the development of the present study.

Bibliography

1. Heim RA, Sugarman EA, Allitto BA. Improved detection of cystic fibrosis

mutations in the heterogeneous U.S. population using an expanded, panethnic mutation panel. Genet Med [Internet]. 2001 Jun 1 [cited 2019 Feb 25];3(3):168–76. Available from: http://www.nature.com/doifinder/10.1097/00125817-200105000-00004

- Rohlfs EM, Zhou Z, Heim RA, Nagan N, Rosenblum LS, Flynn K, et al. Cystic fibrosis carrier testing in an ethnically diverse US population. Clin Chem [Internet]. 2011 [cited 2018 Jun 19];57(6):841–8. Available from: http://clinchem.aaccjnls.org/content/clinchem/57/6/841.full.pdf
- Raskin S, Pereira-Ferrari L, Caldeira Reis F, Abreu F, Marostica P, Rozov T, et al. Incidence of cystic fibrosis in five different states of Brazil as determined by screening of p.F508del, mutation at the CFTR gene in newborns and patients. J Cyst Fibros [Internet]. 2008 [cited 2018 May 28];7(1):15–22. Available from: https://www.cysticfibrosisjournal.com/article/S1569-1993(07)00036-7/pdf
- 4. Pérez MM, Luna MC, Pivetta OH, Keyeux G. CFTR gene analysis in Latin American CF patients: Heterogeneous origin and distribution of mutations across the continent. J Cyst Fibros. 2007;6(3):194–208.
- 5. Pereira L, Raskin S, Freund A a., Ribas PD, Castro RMV, Pignatti PF, et al. Cystic fibrosis mutations R1162X and 2183AA®G in two southern Brasilian states. Genet Mol Biol [Internet]. 1999 [cited 2018 May 28];22(3):291–4. Available from: http://www.scielo.br/pdf/%0D/gmb/v22n3/22n3a02.pdf
- 6. World Health Organization. The molecular genetic epidemiology of cystic fibrosis [Internet]. 1983. p. 1–24. Available from: https://www.who.int/genomics/publications/en/HGN_WB_04.02_report.pdf
- 7. Ioannou L, Mcclaren BJ, Massie J, Lewis S, Metcalfe SA, Forrest L, et al. Population-based carrier screening for cystic fibrosis: A systematic review of 23 years of research. Genet Med. 2014;16(3):207–16.
- 8. Ogino S, Flodman P, Wilson RB, Gold B, Grody WW. Risk calculations for cystic fibrosis in neonatal screening by immunoreactive trypsinogen and CFTR mutation tests. Genet Med. 2005;7(5):317–27.
- 9. Hamosh A, FitzSimmons SC, Macek M. J, Knowles MR, Rosenstein BJ, Cutting GR. Comparison of the clinical manifestations of cystic fibrosis in black and white patients. J Pediatr. 1998;132(2):255–9.
- 10. Hernández P, González-Andrade F. Guía de Práctica Clínica (GPC) y Manual de Procedimientos Fibrosis quística [Internet]. 2013 [cited 2018 Jun 12]. 94 p. Available from: http://instituciones.msp.gob.ec/documentos/Guias/fibrosis.pdf
- 11. Silva Filho LVRF, Castaños C, Ruíz HH. Cystic fibrosis in Latin America— Improving the awareness. J Cyst Fibros [Internet]. 2016;15(6):791–3. Available from: http://dx.doi.org/10.1016/j.jcf.2016.05.007
- 12. Valle E, Burgos R, Valle J, Egas D, Ruiz-Cabezas J. Analysis of CFTR gene

mutations and Cystic Fibrosis incidence in the Ecuatorian population . Invest Clin. 2007;48(1):91–8.

- INEC. Poster INEC [Internet]. Censo de Población y Vivienda. 2010. p. 2010. Available from: http://www.ecuadorencifras.gob.ec/censo-de-poblacion-yvivienda
- Zambrano AK, Gaviria A, Cobos-Navarrete S, Gruezo C, Rodríguez-Pollit C, Armendáriz-Castillo I, et al. The three-hybrid genetic composition of an Ecuadorian population using AIMs-InDels compared with autosomes, mitochondrial DNA and Y chromosome data. Sci Rep [Internet]. 2019 [cited 2019 Jul 11];9(1):9247. Available from: www.nature.com/scientificreports
- Zambrano AK, Gaviria A, Vela M, Cobos S, Leone PE, Gruezo C, et al. Ancestry characterization of Ecuador's Highland mestizo population using autosomal AIM-INDELs. Forensic Sci Int Genet Suppl Ser [Internet]. 2017;6(August):e477–8. Available from: https://doi.org/10.1016/j.fsigss.2017.09.191
- Paz-y-Mino C, Perez JC, Burgos R, Davalos M V., Leone PE. The ΔF508 mutation in Ecuador, South America. Hum Mutat [Internet]. 1999 Oct [cited 2018 Jun 14];14(4):348–50. Available from: http://doi.wiley.com/10.1002/%28SICI%291098-1004%28199910%2914%3A4%3C348%3A%3AAID-HUMU11%3E3.0.CO%3B2-8
- Ortiz SC, Aguirre SJ, Flores S, Maldonado C, Mejía J, Salinas L. Spectrum of CFTR gene mutations in Ecuadorian cystic fibrosis patients: the second report of the p.H609R mutation. Mol Genet Genomic Med [Internet]. 2017 [cited 2018 Jun 14];5(6):751–7. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5702561/pdf/MGG3-5-751.pdf
- Ruiz-Cabezas JC, Barros F, Sobrino B, García G, Burgos R, Farhat C, et al. Mutational analysis of CFTR in the Ecuadorian population using nextgeneration sequencing. Gene [Internet]. 2019 May [cited 2019 Feb 26];696:28–32. Available from: https://linkinghub.elsevier.com/retrieve/pii/S0378111919301246
- 19. Pereira R, Phillips C, Pinto N, Santos C, dos Santos SEB, Amorim A, et al. Straightforward inference of ancestry and admixture proportions through ancestry-informative insertion deletion multiplexing. PLoS One. 2012;7(1).
- 20. Evanno G, Regnaut S, Goudet J. Detecting the number of clusters of individuals using the software STRUCTURE: A simulation study. Mol Ecol. 2005;14(8):2611–20.
- 21. di Sant'Agnese PA, Davis PB. Cystic fibrosis in adults: 75 cases and a review of 232 cases in the literature. Am J Med [Internet]. 1979 Jan 1 [cited 2019 Feb 7];66(1):121–32. Available from: https://www.sciencedirect.com/science/article/pii/0002934379904911?via%3

Dihub

- 22. McCormick J, Green MW, Mehta G, Culross F, Mehta A. Demographics of the UK cystic fibrosis population: Implications for neonatal screening. Eur J Hum Genet [Internet]. 2002 [cited 2018 May 28];10(10):583–90. Available from: https://www.nature.com/articles/5200850.pdf
- Spoonhower KA, Davis PB. Epidemiology of Cystic Fibrosis [Internet]. Vol. 37, Clinics in Chest Medicine. Elsevier; 2016 [cited 2019 Feb 12]. p. 1–8. Available from: https://www.sciencedirect.com/science/article/pii/S0272523115001367?via% 3Dihub
- 24. Dorfman R. Cystic Fibrosis Mutation Database: Statistics. Cyst Fibros Mutat Database [Internet]. 2015 [cited 2019 Apr 8]; Available from: http://www.genet.sickkids.on.ca/StatisticsPage.html
- 25. The Hospital for Sick Children, Johns Hopkins University, The Hospital for Sick Children. CFTR2 Clinical and Functional Translation if CFTR [Internet]. [cited 2019 Apr 8]. Available from: https://www.cftr2.org/
- 26. Palomaki GE, FitzSimmons SC, Haddow JE. Clinical sensitivity of prenatal screening for cystic fibrosis via CFTR carrier testing in a United States panethnic population. Genet Med [Internet]. 2004 [cited 2019 Feb 11];6(5):405–14. Available from: http://www.
- Sugarman EA, Rohlfs EM, Silverman LM, Allitto BA. CFTR mutation distribution among U.S. Hispanic and African American individuals: Evaluation in cystic fibrosis patient and carrier screening populations [Internet]. Vol. 6, Genetics in Medicine. Nature Publishing Group; 2004 [cited 2019 Feb 25]. p. 392–9. Available from: http://www.nature.com/doifinder/10.1097/01.GIM.0000139503.22088.66
- Alibakhshi R, Kianishirazi R, Cassiman J-J, Zamani M, Cuppens H. Analysis of the CFTR gene in Iranian cystic fibrosis patients: Identification of eight novel mutations. J Cyst Fibros [Internet]. 2008 Mar 1 [cited 2019 Feb 25];7(2):102–9. Available from: https://www.sciencedirect.com/science/article/pii/S1569199307000896?via% 3Dihub
- 29. Lerer I, Sagi M, Cutting GR, Abeliovich D. Cystic fibrosis mutations Δ F508 and G542X in Jewish patients. J Med Genet. 1992;29(2):131–3.
- 30. Aquino R, Protzel A, Rivera J, Abarca H, Dueñas M, Nestarez C, et al. Frequency of the most common mutations of the CFTR gene in Peruvian patients with cystic fibrosis using the ARMS-PCR technique. Rev Peru Med Exp Salud Publica. 2017;34(1):62–9.
- 31. Quint A, Lerer I, Sagi M, Abeliovich D. Mutation spectrum in jewish cystic fibrosis patients in israel: Implication to carrier screening. Am J Med Genet. 2005;136 A(3):246–8.

- 32. Padoa C, Goldman A, Jenkins T, Ramsay M. Cystic fibrosis carrier frequencies in populations of African origin. J Med Genet [Internet]. 1999 Jan 1 [cited 2018 May 28];36(1):41–4. Available from: http://www.ncbi.nlm.nih.gov/pubmed/9950364
- 33. Macek M, Mackova A, Hamosh A, Hilman BC, Selden RF, Lucotte G, et al. Identification of common cystic fibrosis mutations in African-Americans with cystic fibrosis increases the detection rate to 75%. Am J Hum Genet. 1997;60(5):1122–7.
- 34. Osborne L, Santis G, Schwarz M, Klinger K, Dörk T, McIntosh I, et al. Incidence and expression of the N1303K mutation of the cystic fibrosis (CFTR) gene. Hum Genet. 2004;89(6):653–8.
- 35. Loumi O, Baghriche M, Delpech M, Kaplan JC, Bienvenu T. Analysis of the complete coding region of the CFTR gene in ten Algerian cystic fibrosis families. Hum Hered. 1999;49(2):81–4.
- 36. de Araújo FG, Novaes FC, dos Santos NPC, Martins VC, de Souza SM, dos Santos SEB, et al. Prevalence of ΔF508, G551D, G542X, and R553X mutations among cystic fibrosis patients in the North of Brazil. Brazilian J Med Biol Res. 2005;38(1):11–5.