Effects of seasonality and land use on the abundance and distribution of mosquitoes on St. Kitts, West Indies

Matthew J. Valentine[8], Brenda Ciraola[8], Gregory R. Jacobs[2,5], Charlie Arnot[9], Patrick J. Kelly[10], Courtney C. Murdock[1,2,3,4,5,6,7*].

[1]Department of Infectious Diseases, College of Veterinary Medicine, University of Georgia, Athens, GA 30602, U.S.A.

[2]Odum School of Ecology, University of Georgia, Athens, GA 30602, U.S.A.

[3]Department of Entomology, College of Agriculture and Life Sciences, Cornell University, Ithaca, NY, 14853, U.S.A.

[4]Center for Tropical Emerging and Global Diseases, University of Georgia, Athens, GA 30602, U.S.A.

[5]Center for Ecology of Infectious Diseases, Odum School of Ecology, University of Georgia, Athens, GA 30602, U.S.A.

[6]Center for Vaccines and Immunology, College of Veterinary Medicine, University of Georgia, Athens, GA 30602, U.S.A.

[7]River Basin Center, Odum School of Ecology, University of Georgia, Athens, Ga 30602, U.S.A.

[8]One Health Centre for Zoonoses and Tropical Veterinary Medicine. Ross University School of Veterinary Medicine, Island Main Road, West Farm, Basseterre, Saint Kitts and Nevis.

[9]CNWA Consulting, Basseterre, Saint Kitts and Nevis.

[10]Department of Clinical Sciences. Ross University School of Veterinary Medicine, Island Main Road, West Farm, Basseterre, Saint Kitts and Nevis.

*Corresponding author: cmurdock@uga.edu, ccm256@cornell.edu

Emails:

MV: mvalentine@students.rossu.edu

BC: brc9050@gmail.com

GJ: greg.jacobs@uga.edu

CA: charlie.arnot@gmail.com

PK: pkelly@rossvet.edu.kn

CM: cmurdock@uga.edu

Abstract

Background: High quality mosquito surveys that collect fine resolution local data on mosquito species' abundances provide baseline data to help us understand potential host-pathogen-mosquito relationships, accurately predict disease transmission, and target mosquito control efforts in areas at risk of mosquito borne diseases.

Methods: As part of an investigation into arboviral sylvatic cycles on the Caribbean island of St. Kitts, we carried out an island wide mosquito survey from November 2017 to March 2019. Using Biogents Sentinel 2 and miniature CDC light traps that were set monthly and run for 48 hour intervals, we collected mosquitoes from a total of 30 sites distributed across the five common land covers on the island (agricultural, mangrove, rainforest, scrub, and urban). We developed a mixed effects negative binomial regression model to predict the effects of land cover, seasonality, and precipitation on observed counts of the most abundant mosquito species we found.

Results: We captured 10 of the 14 mosquito species reported on the island, the four most abundant being *Aedes taeniorhynchus*, *Culex quinquefasciatus*, *Aedes aegpyti*, and *Deinocerites magnus*. Sampling in the mangroves yielded the most mosquitoes, with *Ae. taeniorhynchus, Cx. quinquefasciatus,* and *De. magnus* predominating. *Aedes aegypti* was recovered primarily from urban and agricultural habitats, but also at lower frequency in other land covers. *Psorophora pygmaea* and *Toxorhynchites guadeloupensis* were only captured in scrub habitat. Capture rates in rainforests were low. Our models indicated the relative abundance of the four most common species varied seasonally and with land cover. They also suggested that the extent to which monthly average precipitation influenced counts varied according to species.

Conclusions: This study demonstrates there is high seasonality in mosquito abundances and that land cover influenced the distribution and abundance of mosquito species on St. Kitts. Further, human-adapted mosquito species (e.g. *Ae. aegypti* and *Cx. quinquefasciatus*) that are known vectors for many human relevant pathogens are the most wide-spread (across land covers) and the least responsive to seasonal variation in precipitation.

Keywords: Caribbean, Land cover, Model, Mosquito, Precipitation, Season, Surveillance

Background

Mosquitoes are responsible for considerable human and animal suffering and economic losses because of their nuisance value and the diseases of high morbidity and mortality they can transmit [1–2]. Recent mosquito-borne arboviral pandemics have been able to spread through previously unaffected regions, like the Americas [3–4], due to the wide-spread presence and abundance of human-adapted mosquito species. Furthermore, mosquito-borne pathogens can become established in new areas if there are suitable animal reservoir populations and mosquito species that can transmit the organisms between these animals (and potentially to humans), as has occurred with yellow fever virus in South America [5–6].

High quality mosquito surveys are an essential tool for predicting mosquito-borne disease transmission and for mosquito control. Surveys that collect fine resolution local data on mosquito species abundances provide fundamental baseline data on the composition of mosquito communities in a given area, the relative abundances of mosquito species within the community, and how the abundance of species and the composition of mosquito communities change across space and time. The development of population abundance models that leverage count data

generated from these surveys, in turn, can be used to predict how mosquito abundances change

seasonally and across different land covers. Information of this nature is crucial for describing

potential host-pathogen-mosquito relationships in novel transmission foci, accurately predicting

disease transmission, and for targeting and assessing the efficacy of mosquito control efforts [7–

8].

St. Kitts is a small tropical island in the Caribbean where local experience shows

mosquitoes are very common and their nuisance value high. Outbreaks of mosquito-borne

arboviral infections have occurred on the island, which also has a large population of African

green monkeys (*Chlorocebus aethiops sabaeus*) that may be involved in arbovirus sylvatic

cycles as is the case in Africa [6]. Due to the large numbers of tourists visiting the region each

year, islands in the Caribbean like St. Kitts could be a source of mosquitoes, and the pathogens

they carry, for transfer into currently naïve areas of the world like the USA. Indeed, some

workers consider islands, 'hotspots' for disease emergence [9].

Historically, there has been long standing interest in the mosquito species inhabiting the

Caribbean particularly since it was discovered that malarial parasites (*Plasmodium* spp.) and

yellow fever virus are transmitted by mosquitoes. Detailed mosquito surveys from the 1970s

included several Caribbean islands including St. Kitts and Nevis [10]. The most recent survey on

St. Kitts was conducted in 2010 [11] and although this was the most comprehensive survey

performed on the island to date, it did not provide data on the how the distribution and relative

abundances of mosquito species changes seasonally and with land cover. Mosquitoes were only

collected during a single week of the dry and wet seasons and sampling did not include all land

covers. As part of an investigation into arboviral sylvatic cycles on St. Kitts, we carried out a

comprehensive survey of the mosquito populations across the various land covers on the island

on a monthly/ bimonthly basis from September 2017 to March 2019. We related mosquito

survey data to relevant biological and environmental covariates to assess the influence of land

use and seasonal variation on spatial and temporal biodiversity of mosquitoes on St. Kitts. Below

are a description of our methods and our findings.

Methods

Study Area

St. Kitts (Fig. 1) is a 168 square km, geographically isolated, volcanic, Caribbean island located

in the Lesser Antilles (17.33° N, 62.75° W). It has a population of approximately 40,000 people

mostly inhabiting Basseterre, the capital, and a string of small village communities distributed

along the main coastal road which circles the island. The climate in St. Kitts is tropical, driven by

constant sea breezes with little seasonal temperature variation (27-30°C). The wet season runs

from May to November with risk of hurricanes from June to November. Rainforest covers the

uninhabited, steep volcanic slopes in the center of the island, surrounded by lower gentler slopes

consisting mostly of abandoned sugar cane fields or arable farmlands. The south east of the

island is primarily an arid peninsula covered mainly in scrub with beaches, mangroves, and salt-

ponds.

Mosquito Sampling

To estimate the diversity and abundance of mosquito species across different land covers and

seasons, we evaluated counts and species identity of mosquitoes captured in trap arrays set

across the island. Trapping was carried out once every one to two months from September 2017

to March 2019 in each of five representative land covers (September and October 2017 were

omitted from the analysis due to severe weather). We used a randomized stratified sampling

design to select six replicate sites at least 1 km apart, in each of five distinct land cover

categories (agricultural, mangrove, rainforest, scrub and urban) producing 30 sites in total (Fig.

1). Final site selection was dependent on accessibility and landowner consent.

To increase detection of host-seeking mosquitoes we used both CDC light traps (J.W.

Hock, USA) and Biogents Sentinel 2 traps (BGS) (Biogents AG, Germany) until November

2018, thereafter only BGS traps were used. The CDC light trap was run without the light source

to decrease potential non-mosquito bycatch, and the BG-sentinel lure (Biogents AG, Germany)

was used in the BGS trap to target anthrophilic species. Carbon dioxide was generated by mixing

35 g of dried bread making yeast (Fleischmann's Active Dry Yeast, USA), 0.7 kg of unbranded

white sugar, and approximately 2.5 L water in a centrally placed 5 L water bottle. Carbon

dioxide was delivered to each trap via a 5 m length of 5 mm (internal diameter) PVC tubing [12–

14]. At each site, traps were placed 10 m apart from one another. Traps were run each month

when possible from September 2017 to March 2019 for 48 hours, with yeast-sugar solution,

batteries, and catch bags replaced every 24 hours. Trapped mosquitoes were transported to the

research laboratory of Ross University School of Veterinary Medicine (RUSVM) and stored at -

80°C for later identification. After being rehydrated on chilled damp tissue paper, mosquitoes

were identified on a chill table using morphological keys under a stereomicroscope (Cole

Palmer, USA) at 10x-40x magnification [15–17]. Counts of each mosquito species were recorded

for each sampling date, land use, and location.

Statistical Analysis

We evaluated influences of different land covers (agricultural, mangrove, rainforest, scrub,

urban) on the relative abundance of the four most common mosquito species found in our survey.

The land covers in a 1 km^2 area (565 m radius) around each sampling site were determined from

local observation, a remote sensing vegetation classification [18], assorted vector data sets

supplied by the government of St. Kitts, and the most recent Google Imagery (2019). When

discordance in ascribing land covers was found between the different methods, the Google

images were used preferentially. The percentages of each land cover at each site (Fig. 2) were

calculated and used in establishing the models.

We assessed the effects of land cover at each trap location and monthly precipitation on

the numbers of the four most common mosquito species trapped using mixed effects generalized

linear regression models [19]. Our response variable for this analysis was the number of

mosquitoes of each species captured by BGS traps during each 48 hour trapping interval. We

omitted CDC traps in the model because they were not used on all occasions and seemed to

capture fewer mosquitoes. A list of variables and general expectations of their effects on the

counts of mosquitoes of each species captured can be found in Additional file 1: Table S1. We

included two categorical variables reflecting the high affinity of some mosquito species (e.g. *Ae.*

taeniorhynchus and *De. magnus*) for mangrove habitat and crabhole habitat located in the

vicinity of mangroves [10, 20–22]. These variables included "mangrove", which described the

land cover of sites that fell within mangrove habitats regardless of surrounding land covers, and

"m_trait", which described a mosquito species preference for mangrove habitat. Monthly

precipitation measurements were obtained at the Robert L. Bradshaw International Airport and

accessed as archived data downloaded from the Weather Underground website

(www.wunderground.com: accessed August 2019).

We fitted models using the R package glmmTMB, which allows the specification of

generalized linear mixed-effects models for a variety of error distributions, including Poisson

and negative binomial distributions [23]. Preliminary analyses revealed that a negative binomial

distribution with a quadratic variance-to-mean relationship best explained our data [24]

(Additional file 2: Table S2), and we used this error distribution for all subsequent analyses. We

assumed a linear relationship between overall mosquito counts (on the log-link scale) and

monthly average precipitation at the island scale to account for intra-annual seasonality (e.g., wet

vs. dry seasons). Species-specific random slope and intercept terms for precipitation allow its

effect to vary by species, and random site intercepts account for repeat observations at each site.

The effects of land cover were allowed to vary independently by species. We evaluated 12 model

hypotheses of species-specific variation in relative abundance with land cover (Table 1). Land

cover variables incorporated in our 12 models generally include the percentage of local land

cover. We excluded any hypotheses for which variance inflation factors were greater than five

prior to model evaluation and used AIC to select our best model from the candidate set [25]. To

assess model fit, we evaluated the distribution of re-scaled model residuals from the R package

DHARMa [26] and calculated conditional and marginal R^2 values following Nakagawa et al

[27]. We used site-level predictions from our best model to show model-estimated trends in

mosquito abundance across land use and season for the duration of our study.

Results

From November 2017 to March 2019 we captured 10 of the 14 species previously recorded on

St. Kitts [10–11, 15] (Additional file 3: Table S3, Additional file 4: Table S4). Voucher

specimens were deposited in the United States National Museum (USNM) under the following

catalog numbers USNMENT01239050-74 and USNMENT01239079. The most abundant

species of mosquito was *Aedes taeniorhynchus* (n=3861)*,* which was primarily found in

mangroves (88.4%) (Figure 1, Additional file 3: Table S3). *Culex quinquefasciatus* (n=1663)

was the second most abundant species primarily captured in urban areas (48.8%) (Figure 1,

Additional file 3: Table S3). *Aedes aegypti*, *Ae. taeniorhynchus, Cx. quinquefasciatus*, and

Deinocerites magnus were species captured in all five land covers (Figure 1, Additional file 3: Table S3). There were no clear morphological differences in the *Ae. aegypti* collected across different land covers. All other species were much less abundant. *Psorophora pygmaea* and *Toxorhynchites guadeloupensis* were only captured in scrub habitat, with the remaining species being distributed across more than one land cover (Figure 1, Additional file 3: Table S3). The highest overall number of mosquitoes captured, mostly *Ae. taeniorhynchus*, were caught in November 2018 (N=3786), and monthly mean average catches were higher in general during the wet season (N=1080) than the dry season (N=177). *Aedes aegypti* was the only species to be captured during every trapping month. Only four *Anopheles albimanus*, the main vector of malaria in the Caribbean [28], were caught across the entire survey period (Additional file 4: Table S4).

The best model of temporal variation in the relative abundance of the four most common mosquito species on the Island of St. Kitts was the model H12 from Table 1. This model included the effects of monthly precipitation, a mangrove breeding trait interaction with mangrove sites, and species-specific effects driven by agriculture, urban, and rainforest land covers (Table 2). Our best model can be expressed in pseudo-code as

$$
\begin{aligned}
y_{(i,j,s)} &\sim \text{NegBin}\big(\mu_{(i,j,s)}, \phi\big) \\
\log\big(\mu_{(i,j,s)}\big) &= \beta_0 + \alpha_{0(s)} + \big(\beta_1 + \alpha_{1(s)}\big)Precip_{(j)} + \beta_2 m_{trait_{(s)}} + \beta_3 Mangrove_{(i)} + \\
&\quad \beta_{4(s)} LocalAgriculture_{(i)} + \beta_{5(s)} LocalUrban_{(i)} + \beta_{6(s)} LocalRainforest_{(i)} + \\
&\quad \beta_7 m_{trait_{(s)}} Mangrove_{(i)} + \alpha_{2(i)}
\end{aligned}
\quad \#(\text{eq. 1})
$$

where i, j, and s denote indices for site, month, and species, respectively. NegBin reflects the negative binomial distribution, and the final model includes the following: random intercepts for each species ($\beta_0 + \alpha_{0(s)}$), a main effect of precipitation with species-specific random slopes

$((\beta_1 + \alpha_{1(s)})Precip_{(j)})$, an interaction between mangrove and the mangrove breeding trait

$(\beta_2 m\_trait_{(s)} + \beta_3 Mangrove_{(i)} + \beta_7 m\_trait_{(s)} Mangrove_{(i)})$, species-specific main effects

on proportional local land cover variables $(\beta_{4(s)} LocalAgriculture_{(i)} + \beta_{5(s)} LocalUrban_{(i)} +$

$\beta_{6(s)} LocalRainforest_{(i)})$, and a site-level random intercept term $(\alpha_{2(i)})$. The variance for our

best negative binomial model scales quadratically with the mean (μ): $Var(y_{(i,j,s)}) = \mu_{(i,j,s)}(1 +$

$\frac{\mu_{(i,j,s)}}{\phi})$ [24]. Our best model's predicted relative abundance for each mosquito species, averaged

across each site's land cover, illustrated species-specific responses to surrounding land cover and

seasonal precipitation effects across the island (Table 2, Figure 3, Additional file 5: Figure S1).

Overall, we observed a positive effect of precipitation on the relative abundance of mosquitoes

(β_1), with significant among-species variation in this relationship $(\alpha_{1(s)}$, Table 2). To further

explore the effect of precipitation, we derived the best linear unbiased predictors (BLUPs) for

each species' precipitation effect. The BLUPs suggested that the effect of precipitation was

strongest for *Ae. taeniorhynchus* and to a lesser extent *Ae. aegypti* (Figure 4), and was weakest

for *Cx. quinquefasciatus* and *De. magnus*. The model also predicted significantly negative

relationships between urban land cover and *Ae. taeniorhynchus* and *De. magnus,* but only

slightly-positive, non-significant relationships between urban land cover and the urban-

associated mosquitoes, *Cx. quinquefasciatus* and *Ae. aegypti*. Rainforest and agricultural land

cover were negatively associated with the relative abundance of all species we considered except

Ae. aegypti, which exhibited no significant covariation with either rainforest or agricultural land

cover (Table 2). As expected, the model predicts the relative abundance of mosquito species with

a mangrove breeding preference (*Ae. taeniorhynchus* and *De. magnus*) to be lower than average

except when trapping sites occur within mangrove habitat where the mangrove "trait" had a net

positive effect (i.e., $(\beta_2 + \beta_7) > 0$, Table 2). Inspection of re-scaled residuals generated by

simulation from the fitted model [26] suggested uniformity in the distribution of residuals (One-

sample Kolmogorov-Smirnov test: $D = 0.018$, $P = 0.671$), indicating good concurrence between

the data and model predictions. Marginal and conditional R^2 (0.601 and 0.696, respectively) also

indicated a well-fit model (Table 2): the marginal R^2 of 0.601 suggested that the fixed effects

portion of the model explained over 60% of the variation in counts, and the conditional R^2 of

0.695 revealed the additional variance explained by accounting for additional variation

attributable to the random effects [27].

DISCUSSION

Monthly mosquito surveillance on St. Kitts from November 2017 to March 2019 enabled us to

capture a diversity of mosquito species that varied in abundance across seasons and land cover

types. We captured 10 of the 14 species (5 genera) historically recorded on the island [10–11,

15]. While our results largely confirm those of the 2010 survey, they provide higher spatial and

temporal resolution of both the relative abundance and distribution of different mosquito species

on the island.

The four most abundant species captured were *Ae. taeniorhynchus*, *Cx. quinquefasciatus*,

Ae. aegypti, and *De. magnus*. These species were detected at least once in all land cover types

over the study period. The three most abundant species captured are competent vectors of

pathogens recorded on St. Kitts (*Cx. quinquefasciatus* - West Nile virus and *Dirofilaria immitis*;

Ae. taeniorhynchus – *Dirofilaria immitis*; and *Ae. aegypti* - dengue, chikungunya, and Zika

viruses) [29–33] and other pathogens that could potentially become established on the island in

the future due to the abundance of their vectors. For example, *Cx. quinquefasciatus*, the southern

house mosquito, is widely distributed across the sub tropics and can also transmit St. Louis

encephalitis virus, Western equine encephalitis virus, Rift Valley fever virus, *Wuchereria*

bancrofti, and avian malaria [17]. *Aedes taeniorhynchus,* the black salt marsh mosquito, is

widely distributed in all islands of the Caribbean and can transmit Venezuelan, Eastern and

Western equine encephalitis virus [11, 34–35]. The eponymous yellow fever mosquito, *Ae.*

aegypti, can also transmit yellow fever virus [17]. While *An. albimanus* is a known malaria

vector in Central America, northern South America, and the Caribbean, the overall low

abundance of this species on St. Kitts (n=4 in our study and n=2 in 2010, [11]) suggests this

species is unlikely to support malaria transmission on the island.

The presence and absence, as well as overall relative abundance, of particular mosquito

species captured across the different land covers on St. Kitts broadly align with what is known

for these species in the literature. The species count model for our four most abundant

mosquitoes predicts both *Cx. quinquefasciatus* and *Ae. aegypti* to have the highest relative

abundance in an urban habitat. Both species breed most successfully in fresh water-filled man-

made containers and are therefore found primarily around houses in urban environments.

Further, *Ae. aegypti* preferentially feeds on human hosts, particularly when indoors [17, 36] and

rests inside domestic dwellings [17]. The fact that we observed these species, albeit at lower

abundances, in other land covers is not entirely surprising. The model predicted *Ae. aegypti* to be

similarly abundant across the survey period in urban as well as agricultural habitats. From local

experience, agricultural land covers provide ample breeding habitats for container breeding

mosquito species in the form of discarded tires, styrofoam containers, plastic water bottles and

bags, and agricultural equipment in which water can collect.

Interestingly, the model also predicted moderate relative abundance across the survey

period for both *Cx. quinquefasciatus* and *Ae. aegypti* in mangrove and scrub habitats, which may

be due to the presence of artificial containers suitable for breeding or an increased tolerance to

brackish water in high marsh areas of the mangrove [21, 37]. *Culex quinquefasciatus* has been

recorded previously in brackish water on St. Kitts [10] and *Ae. aegypti* is tolerant of brackish

water in other coastal regions [38–40]. Additionally, *Cx. quinquefasciatus* is an opportunistic

forager that has the ability to fly relatively long distances [41] and so adults could be found in

areas far removed from their larval breeding sites. Finally, *Ae. aegypti* was also predicted to be

abundant, albeit at lower levels, in rainforest habitat across the trapping period. Elsewhere in the

Caribbean, *Ae. aegypti* have been found breeding in more natural habitats in addition to artificial

containers [22]. These include rock holes, calabashes, tree holes, leaf axils, bamboo joints,

papaya stumps, coconut shells, bromeliads, ground pools, coral rock holes, crab holes, and conch

shells which are all also present on St. Kitts.

Species with the highest capture rates and predicted by our model to have high relative

abundance in mangrove habitats included both *Ae. taeniorhynchus* and *De. magnus*. These two

species were predicted to have relatively low abundance in scrub surrounding mangrove sites on

the island, and were not predicted to be abundant in other land cover types. *Aedes*

taeniorhynchus, the black salt marsh mosquito, is widely distributed in all islands of the

Caribbean where it also favors low lying marsh land as is the case on St. Kitts [21, 35].

Similarly, *De. magnus,* the crabhole mosquito, is found primarily in crabholes that are abundant

in the soft sands in mangrove habitats around the Caribbean [10, 42]. We also captured *Culex*

nigripalpus, *An. albimanus*. *Ps. pygmaea,* and *Aedes tortilis* in mangroves or the surrounding

scrub land cover, likely due to their preference for breeding in temporary brackish and / or fresh

water sources [10, 15]. We did not include these species in our relative abundance model due to

low capture rates.

Generally, we had very low capture rates of mosquitoes in rainforest land cover which

was consistent with our model's predictions of a negative effect of rainforest cover on the

relative abundance of the four most abundant species we trapped. This is not necessarily

reflective of the results of surveys from other regions of the Caribbean. For example, a study in

forested areas of eastern Trinidad between July 2007 and March 2009 collected 185,397

mosquitoes across 46 species [43]. Although this study was of a similar duration, our low capture

rates in rainforest land cover might reflect 1) less breeding habitat or fewer vertebrate hosts

present in the rainforest, 2) different sampling methods across the studies (e.g. CDC light traps

deployed with CO_2 lures used vs. BGS traps baited with the human lure or CDC light traps

baited with sugar-yeast CO_2 lures [12]), and 3) frequency of trapping effort (weekly vs.

monthly). It seems unlikely that our trapping at ground level may have excluded mosquito

species that thrive in tree-top habitats because several arboviral surveillance studies in forests in

Brazil [44], New Mexico [45], and other sites across the United States [46] demonstrate that

various trapping methods (e.g. entomological nets, aspirators, and CDC light traps) set in the

canopy did not catch significantly more mosquitoes than those on the ground. We did capture

one *Aedes busckii* in the rainforest during our survey. A previous survey [10] as well as some

experience with larval surveys of tree holes in rainforest habitats on St. Kitts (data not shown)

suggest *Ae. busckii* could be a rain forest habitat specialist, but more data are needed. In general,

little is known about the ecology of this mosquito other than that it is confined to the Lesser

Antilles (Dominica, Grenada, Guadeloupe, Martinique, Montserrat, St. Kitts and Nevis and Saint

Lucia) [47]. We also captured *Cx. quinquefasciatus*, *Ae. taeniorhynchus*, and *De. magnus* at

very low rates in the rainforest (Additional file 3: Table S3). We speculate that these captures

could be the result of mosquitoes either breeding in man-made containers in neighboring

agricultural habitat (*Cx. quinquefasciatus*) or mosquitoes being blown into novel habitats during

tropical storms (*Ae. taeniorhynchus* and *De. magnus*). In the case of *De. magnus,* land crabs

(*Gecarcinus ruricola*) do inhabit the rainforest, which could provide breeding and resting sites in

their crabhole burrows for this specialist mosquito species [42].

Mosquito capture rates were also strongly determined by time of season and precipitation

throughout the survey period. In general, our species count model predicted a positive effect of

precipitation on the relative abundance of our four most common mosquito species. The effects

of precipitation we found could be due to several reasons. Although excess rain may flush larvae

from their habitats and decrease adult mosquito populations [48–49], a seasonal increase in

precipitation increases the abundance and persistence of larval habitats resulting in higher

densities and overall capture rates [48, 50–52]. Additionally, increased precipitation is associated

with increased relative humidity, which has been shown to have important positive effects on

mosquito abundance [52–53], lifespan [54], activity, and questing behavior [55–57].

Interestingly, the effect of precipitation on relative abundance was species-specific among the

four most common mosquito species we found. Our post-hoc BLUPs assessment suggests the

effect of precipitation had a strong effect on the relative abundances of *Ae. taeniorhynchus*, a

moderate effect on *Ae. aegypti*, and smaller effects on *Cx. quinquefasciatus* and *De. magnus*. The

strong effect of precipitation on *Ae. taeniorhynchus* might occur because this mosquito species

relies largely on natural habitats, which are often dependent on local rainfall. While *Ae. aegypti*

utilizes artificial, and human watered containers heavily for ovipositing, dependent on rainfall it

is also known to oviposit in natural habitats on other Caribbean islands [22]. *Culex*

quinquefasciatus might be less dependent on rainfall, as most individuals were captured in urban

habitat and were most likely emerging from persistent, human-watered, artificial habitats.

Whereas, increased rainfall above a certain threshold might expand water bodies in mangroves,

flooding crabholes, that in turn could locally reduce breeding sites for *De. magnus* [42].

While the overall capture rates of *Ae. aegypti* were significantly lower across non-urban

habitats, their presence in other land covers on St. Kitts and other Caribbean islands [10, 22]

could have several implications for our understanding of the general ecology of this species and

transmission of arboviruses in the Caribbean. Unlike the situation in Africa [58], where there are

morphological differences between urban and non-urban forms, *Ae. aegypti* collected from urban

areas of St. Kitts had the same morphology as *Ae. aegypti* collected from other land covers.

Depending on the amount of gene flow that occurs across these various populations (which is

thought to be limited [59–60]), ecological heterogeneity across the common land covers of St.

Kitts and other Caribbean islands could result in substantial genetic variation across *Ae. aegypti*

populations at fine spatial scales. Variation in microclimate [53, 61], oviposition sites [10, 22],

and species of vertebrates available for blood feeding [62] are examples of potentially relevant

sources of selection on *Ae. aegypti* populations inhabiting different land covers on St. Kitts.

Genetic variation across *Ae. aegypti* populations on the island, in turn, could translate into

substantial ecological differences among these populations with potentially significant

implications for disease transmission among sylvatic reservoir hosts (e.g. non-human primates)

and human populations on the island. Population genotyping is underway in our laboratories to

identify if significant genetic structuring exists across *Ae. aegypti* populations on St. Kitts. Also,

to more precisely determine phenotypic differences between populations, we are conducting

studies to confirm the presence of reproducing adults across each land cover and using blood

meal analysis and bait trapping to identify novel mosquito-host associations.

Our study provides a more comprehensive spatial and temporal (within-year) picture of

the distribution of mosquito species on St. Kitts relative to previous surveys [10, 11, 15].

However, it suffers from several minor limitations. Due to specimen damage during capture and

transport to the laboratory, a proportion of *Aedes* spp. (N=219) and *Culex* spp. (N=1694) were

reported only to genus level. These counts differed substantially from those identified to species

level which comprised 4334 *Aedes* spp. and 1697 *Culex* spp. in total. Based on the capture

location of these specimens, the majority of these individuals are likely *Ae. taeniorhynchus*

(mangrove) and *Cx. quinquefasciatus* (urban), respectively. That being said, capture rates for *Ae.*

tortilis and *Cx. nigripalpus* are likely underestimated due to difficulty in identifying specimens

damaged from capture and handling. Finally, the CDC traps performed unexpectedly poorly

compared with BGS traps, possibly due to the lack of light source and the use of a human lure in

the BGS traps. As trap type can select for different mosquito species [63], our count data may be

biased toward species that prefer BGS traps and human lures.

Conclusion

Consistent with findings on other islands and the literature, our island wide mosquito survey has

demonstrated that the distribution and relative abundance of mosquito species on St. Kitts are

affected by land cover. Further, we found substantial seasonality in mosquito capture rates, likely

driven by variation in precipitation. Interesting insights gained from this study include the

presence of *Ae. aegypti* in all the land covers we studied, which could have important

implications for mosquito-borne disease transmission on the island. This also somewhat counters

the current literature suggesting *Ae. aegypti* is primarily found in highly urban habitats and feeds

almost exclusively on human hosts. Although *Aedes albopictus* occurs on other Caribbean

islands [64] we did not find this species in our survey. Ongoing surveillance will be important,

however, as invasive mosquito species and changing patterns of mosquito communities caused

by changes in land use and climate may facilitate mosquito-borne disease transmission in people

and animals.

Additional files

Supplementary data submitted as separate files:

Additional file 1: Table S1. Variables and associated hypotheses evaluated in statistical models.

Additional file 2: Table S2. Statistical and model methods

Additional file 3: Table S3. Counts of mosquito species caught across the five different land

covers from November 2017 to March 2019 on St. Kitts.

Additional file 4: Table S4. Counts of mosquito species per month on St. Kitts from Nov 2017

to March 2019 with the wet season highlighted in grey (May-November).

Additional file 5: Figure S1. Time series plot for each site land use category (column) and for

each species (row) in their predicted relative abundance (conditional on random effects) from our

best model. Solid lines denote the average predicted relative abundance of mosquito species

across the 6 sites within that land cover and dotted lines denote the 95% confidence interval of

that 6-site mean. Points denote the 6-site average relative abundance from the raw data. Note that

the y-axis scale varies by species (row).

Abbreviations

BGS: Biogents Sentinel 2 Trap; CO^2: carbon dioxide; CDC: Centre for Disease Control (USA);

RUSVM: Ross University School of Veterinary Medicine; USNM: United States National

Museum

Acknowledgements

Gilbert Gordon, Ross University School of Veterinary Medicine Students, David Pecor, Erica

McAlister, Michelle Evans, Mike Newberry, Bryan Giordano, Rob and Kathleen Gilbert, Anna

Becker, Moses Humphrey, Michel Vandenplas.

Declarations

Ethics approval and consent to participate

Not applicable

Consent for publication

Not applicable

Availability of data and material

The datasets supporting the conclusions of this article are included within the article and its

additional files. Voucher specimens of some of the mosquito specimens associated with this

study were deposited in the United States National Museum (USNM) under the following

catalog numbers USNMENT01239050-74 and USNMENT01239079.

Competing interests

None

Funding

The project was funded by National Institute of Allergy and Infectious Diseases R21 grant

(1R21AI128407-01) and RUSVM.

Authors' contributions

MV, BC and CM conducted the survey. PK and CM designed the study and supervised the

survey. GJ, CA and CM produced the models. CA, MV and GJ produced tables and figures for

the manuscript text. MV, CM, GJ, PK and CA prepared the manuscript text. All authors read and

approved the final manuscript.

REFERENCES

1. WHO a. World Health Organization: Global Burden of Major Vector-Borne Diseases, as of

March 2017. 2018. http://www.who.int/vector-control/burden_vector-borne_diseases.pdf

(accessed June 13th 2019); Global vector control response 2017-2030

2. WHO b. World Health Organisation: Vector Borne Diseases Factsheet.

http://www.who.int/en/news-room/fact-sheets/detail/vector-borne-diseases. Accessed 13th June

2019.

3. WHO c. World Health Organisation: Chikungunya. http://www.who.int/news-room/fact-

sheets/detail/chikungunya. Accessed 13th June 2019.

4. WHO d. World Health Organisation: Zika. http://www.who.int/news-room/fact-

sheets/detail/zika-virus. Accessed 13th June 2019.

5. Figueiredo LTM. Human urban arboviruses can infect wild animals and jump to sylvatic

maintenance cycles in South America. Front Cell Infect Microbiol. 2019;9:259.

6. Valentine MJ, Murdock CC, Kelly PJ. Sylvatic cycles of arboviruses in non-human primates.

Parasit Vectors. 2019;12(1):463.

7. Shragai T, Tesla B, Murdock CC, Harrington LC. Zika and chikungunya: mosquito-borne

viruses in a changing world. Ann N Y Acad Sci. 2017;1399(1):61–77.

8. Cornel AJ, Lee Y, Almeida APG, Johnson T, Mouatcho J, Venter M, et al. Mosquito

community composition in South Africa and some neighboring countries. Parasit Vectors.

2018;11**:**331.

9. Mavian C, Dulcey M, Munoz O, Salemi M, Vittor AY, Capua I. Islands as Hotspots for

Emerging Mosquito-Borne Viruses: A One-Health Perspective. Viruses. 2019;11(1):11.

10. Belkin JN, Heinemann SJ. Collection records of the project of "Mosquitoes of Middle

America" 4. Leeward Islands Anguilla (ANG), Antigua (ANT), Barbuda (BAB), Montserrat

(MNT), Nevis (NVS), St Kitts (KIT). Mosquito Systematics. 1976;8(2):123–62.

11. Mohammed H, Evanson J, Revan F, Lee E, Krecek RC, Smith J. A Mosquito Survey of the

Twin-Island Caribbean Nation of Saint Kitts and Nevis, 2010. J Am Mosq Control Assoc.

2015;31(4):360–3.

12. Saitoh Y, Hattori J, Chinone S, Nihei N, Tsuda Y, Kurahashi H, et al. Yeast-generated CO^2

as a convenient source of carbon dioxide for adult mosquito sampling. J Am Mosq Control

Assoc. 2004;20:261–64.

13. Smallegange RC, Schmied WH, van Roey KJ, Verhulst NO, Spitzen J, Mukabana WR, et al.

Sugar-fermenting yeast as an organic source of carbon dioxide to attract the malaria mosquito

Anopheles gambiae. Malar J. 2010;9:292.

14. Hapairai LK, Joseph H, Sang MA, Melrose W, Ritchie SA, Burkot TR, et al. Field evaluation

of selected traps and lures for monitoring the filarial and arbovirus vector, *Aedes polynesiensis*

(Diptera: Culicidae), in French Polynesia. J Med Ent. 2013;50(4):731–39.

15. Belkin JN, Heinemann SJ, Page WA. The culicadae of Jamaica. Contrib Amer Ent Inst.

1970;6:1–458.

16. Darsie RF, Ward RA. Identification and geographical distribution of the mosquitoes of North

America, north of Mexico. Gainesville, FL: University Press of Florida; 2005.

17. Burkett-Cadena, ND. Mosquitoes of the Southeastern United States. Tuscaloosa, AL: The

University of Alabama Press; 2013.

18. Helmer EH, Kennaway TA, Pedreros DH, Clark ML, Marcano-Vega, H, Tieszen LL, et al.

Land cover and forest formation distributions for St. Kitts, Nevis, St. Eustatius, Grenada and

Barbados from decision tree classification of cloud-cleared satellite imagery. Caribbean J Sci.

2008;44(2):175–98.

19. R Core Team. R: A language and environment for statistical computing. R Foundation for

Statistical Computing, Vienna, Austria. 2019.

20. Lane J. Neotropical Culicidae. Vols. I and II. Sao Paulo, Brazil: University of Sao Paulo.

1953.

21. Ritchie SA. Mosquito Control Handbook: Salt Marshes and Mangrove Forests. University of

Florida. Publication ENY70034.1992.

22. Chadee DD, Ward RA, Novak RJ. Natural habitats of *Aedes Aegypti* in the Caribbean--a

review. J Am Mosq Control Assoc. 1998;14(1):5–11.

23. Brooks ME, Kristensen K, van Benthem KJ, Magnusson A, Berg CW, Nielsen A, et al.

glmmTMB balances speed and flexibility among packages for zero-inflated generalized linear

mixed modeling. R J. 2017;9:378–400.

24. Hardin JW, Hilbe JM. Generalized linear models and extensions. 4th ed. College station, TX:

Stata press; 2007.

25. Burnham KP, Anderson DR. Practical use of the information-theoretic approach. In: Model

selection and inference, Burnham KP, Anderson DR editors. Model Selection and interference.

New York, NY: Springer; 1998. P. 75-117.

26. Hartig F. DHARMa: Residual diagnostics for hierarchical (multi-level / mixed) regression

models. R package version 0.2.4. 2019. http://florianhartig.github.io/DHARMa/.Accessed 4th

September 2019.

27. Nakagawa S, Johnson PCD, Schielzeth H. The coefficient of determination R^2 and intra-class

correlation coefficient from generalized linear mixed-effects models revisited and expanded. J R

Soc Interface. 2017;14:134.

28. Briët OJT, Impoinvil DE, Chitnis, N, Pothin E, Lemoine JF, Frederic J, et al. Models of

effectiveness of interventions against malaria transmitted by *Anopheles albimanus*. Malar J.

2019;18(1):263.

29. Conan A, Napier P, Shell L, Knobel DL, Dundas J, Scorpio D, et al. Heterogeneous

distribution of *Dirofilaria immitis* in dogs in St. Kitts, West Indies, 2014–2015. Vet Parasitol

Reg Stud Reports. 2017;10:139–142.

30. Bolfa P, Jeon I, Loftis A, Leslie T, Marchi S, Sithole F, et al. Detection of West Nile virus

and other common equine viruses in three locations from the Leeward Islands, West Indies. Acta

Tropica. 2017;174:24–28.

31. PAHO a. Pan American Health Organisation: PLISA health information platform for the

Americas. http://www.paho.org/data/index.php/en/mnu-topics/indicadores-dengue-en/dengue-

nacional-en/252-dengue-pais-ano-en.html. Accessed 6th August 2019.

32. PAHO b. Pan American Health Organisation: PLISA health information platform for the

Americas.

https://www.paho.org/hq/index.php?option=com_topics&view=rdmore&cid=5927&item=chiku

ngunya&type=statistics&Itemid=40931&lang=en. Accessed 6th August 2019

33. PAHO c. Pan American Health Organisation: PLISA health information platform for the

Americas.

https://www.paho.org/hq/index.php?option=com_content&view=article&id=12390:zika-

cumulative-cases&Itemid=42090&lang=en. Accessed 6th August 2019

34. Barrera R, MacKay A, Amador M, Vasquez J, Smith J, Díaz A, et al. Mosquito vectors of

West Nile virus during an epizootic outbreak in Puerto Rico. J Med Entomol. 2010;47(6):1185–

95.

35. Agramonte NM, Connelly CR. Black salt marsh mosquito *Aedes*

taeniorhynchus(Wiedemann) (Insecta: Diptera: Culicidae). EDIS. EENY591. 2014.

https://edis.ifas.ufl.edu/in1035#FIGURE%205. Accessed 4th April 2020.

36. Harrington LC, Edman JD, Scott TW. Why do female *Aedes aegypti* (Diptera: Culicidae)

feed preferentially and frequently on human blood? 2001;38(3):411–22.

37. Roberts D. Mosquitoes (Diptera: Culicidae) breeding in brackish water: female ovipositional

preferences or larval survival? J Med Entomol. 1996;33(4):525–30.

38. Ramasamy R, Surendran SN, Jude PJ, Dharshini S, Vinobaba M. Larval development of

Aedes aegypti and *Aedes albopictus* in peri-urban brackish water and its implications for

transmission of arboviral diseases. PLOS Negl Trop Dis. 2011;5(11):e1369.

39. Ramasamy R, Jude PJ, Veluppillai T, Eswaramohan T, Surendran SN. Biological differences

between brackish and fresh water-derived *Aedes aegypti* from two locations in the Jaffna

peninsula of Sri Lanka and the implications for arboviral disease transmission. PLoS One.

2014;9(8):e104977.

40. de Brito Arduino M, Mucci LF, Serpa LL, Rodrigues Mde M. Effect of salinity on the

behavior of *Aedes aegypti* populations from the coast and plateau of southeastern Brazil. J

Vector Borne Dis. 2015;52(1):79–87.

41. Verdonschot PFM, Besse-Lototskaya AM. Flight distance of mosquitoes (Culicidae): a

metadata analysis to support the management of barrier zones around rewetted and newly

constructed wetlands. Limnologica. 2014;45:69–79.

42. Belkin JN, Hogue CL. A review of the crabhole mosquitoes of the genus *Deinocerites*

(Diptera, Culicidae). Univ Calif publ entomol. 1959;14(6):411–58.

43. Auguste AJ, Adams AP, Arrigo NC, Martinez R, Travassos da Rosa AP, Adesiyun AA, et al.

Isolation and characterization of sylvatic mosquito-borne viruses in Trinidad: enzootic

transmission and a new potential vector of Mucambo virus. Am J Trop Med Hyg.

2010;83(6):1262–1265.

44. Lira-Vieira AR, Gurgel-Gonçalves R, Moreira IM, Yoshizawa MA, Coutinho ML, Prado PS,

et al. Ecological aspects of mosquitoes (Diptera: Culicidae) in the gallery forest of Brasília

National Park, Brazil, with an emphasis on potential vectors of yellow fever. Rev Soc Bras Med

Trop. 2013;46(5):566–74.

45. DiMenna MA, Bueno R Jr, Parmenter RR, Norris DE, Sheyka JM, Molina JL, et al.

Comparison of mosquito trapping method efficacy for West Nile virus surveillance in New

Mexico. J Am Mosq Control Assoc. 2006;22(2):246–53.

46. Andreadis TG, Armstrong PM. A two-year evaluation of elevated canopy trapping for *Culex*

mosquitoes and West Nile virus in an operational surveillance program in the northeastern

United States. J Am Mosq Control Assoc. 2007;23:137–48.

47. WRBU Walter Reed Biosystematics Unit: Systematic catalog of Culicidae. Walter Reed

Biosystematics Unit, Smithsonian Institution, Washington D.C. 2015.

http://www.mosquitocatalog.org/taxon_descr.aspx?ID=15654 accessed 4th April 2020.

48. DeGaetano AT. Meteorological effects on adult mosquito (*Culex*) populations in

metropolitan New Jersey. Int J Biometeorol. 2005;49(5), 345–53.

49. Dieng H, Rahman GM, Abu Hassan A, Che Salmah MR, Satho T, Miake F, et al. The effects

of simulated rainfall on immature population dynamics of *Aedes albopictus* and female

oviposition. Int J Biometeorol. 2012;56(1):113–20.

50. Reisen WK, Cayan D, Tyree M, Barker CM, Eldridge B, Dettinger M. Impact of climate

variation on mosquito abundance in California. J Vector Ecol. 2008;33(1):89–98.

51. De Little SC, Bowman DM, Whelan PI, Brook BW, Bradshaw CJ. Quantifying the drivers of

larval density patterns in two tropical mosquito species to maximize control efficiency. Environ

Entomol. 2009;38(4):1013–21.

52. Asigau S, Parker PG. The influence of ecological factors on mosquito abundance and

occurrence in Galápagos. J Vector Ecol. 2018;43(1):125–37.

53. Evans MV, Hintz CW, Jones L, Shiau J, Solano N, Drake JM, et al. Microclimate and larval

habitat density predict adult *Aedes Albopictus* abundance in urban areas. Am J Trop Med Hyg.

2019;101(2):362–70.

54. Hylton AR. Studies on longevity of adult *Eretmapodites chrysogaster*, *Aedes Togoi* and

Aedes (Stegomyia) Albopictus Females (Diptera: Culicidae). J Med Ent. 1969;6(2):147–9.

55. Rowley WA, Graham CL. The effect of temperature and relative humidity on the flight

performance of female *Aedes aegypti*. J Insect Physiol. 1968;14(9):1251–7.

56. Bayoh MN. Studies on the development and survival of *Anopheles gambiae sensu stricto* at

various temperatures and relative humidities. Durham: University of Durham; 2001.

57. Okech BA, Gouagna LC, Killeen GF, Knols BG, Kabiru EW, Beier JC, et al. Influence of

indoor microclimate and diet on survival of *Anopheles gambiae* (Diptera: Culicidae) in semifield

conditions in western Kenya. J Med Entomol. 2003;40(5):657–63.

58. Powell JR, Tabachnick WJ. History of domestication and spread of *Aedes aegypti*--a review.

Mem Inst Oswaldo Cruz. 2013;108(Suppl 1):11–17.

59. Harrington LC, Scott TW, Lerdthusnee K, Coleman RC, Costero A, Clark GG, et al.

Dispersal of the dengue vector *Aedes aegypti* within and between rural communities. Am J Trop

Med Hyg. 2005;72(2):209–20.

60. Winskill P, Carvalho DO, Capurro ML, Alphey L, Donnelly CA, McKemey AR. Dispersal

of engineered male *Aedes aegypti* mosquitoes. PLoS Negl Trop Dis. 2015:9(11):e0004156.

61. Murdock CC, Evans MV, McClanahan TD, Miazgowicz KL, Tesla B. Fine-scale variation in

microclimate across an urban landscape changes the capacity of *Aedes albopictus* to vector

arbovirus. PLoS Negl Trop Dis. 2017;11(5):e0005640.

62. Burkett-Cadena, ND, McClure CJW, Estep LK, Eubanks, MD. 2013. Host or habitats: What

drives the spatial distribution of mosquitoes? Ecosphere. 2013;4(2):1–16.

63. Silver JB. Mosquito ecology: field sampling methods. 3rd ed. London: Springer; 2008.

Tabachnick WJ, Munstermann LE, Powell JR. Genetic distinctness of sympatric forms of *Aedes*

aegypti in east Africa. Evolution. 1978;33:287–95.

64. Ali I, Mundle M, Anzinger JJ, Sandiford SL. Tiger in the sun: a report of *Aedes albopictus*.

Acta Trop. 2019;199:105112.

Figure legends

Fig 1. Numbers and species of mosquitoes trapped on St. Kitts at 30 sites comprising six replicates in each of five land covers

Fig 2. The proportions of the different land covers found in a 1 km^2 area (565 m radius) around each of the trapping 30 sites used in the study

Fig 3. Time series plot for each site land cover (line color) and for the four major species we captured (panel) in their predicted relative abundance (conditional on random effects) from our best model. Solid lines denote the average predicted relative abundance of mosquito species across the 6 sites located within that land cover

Fig 4. Best linear unbiased predictors (BLUPs) for the effect of precipitation on mosquito counts

Table 1 A list of main effects of all candidate models considered in analyses of land cover effects on mosquito relative abundance. Interaction-only terms only were used between indicator variables, and evaluates the effect when both indicator variables are equal to 1 (i.e., "m_trait·Mangrove" evaluates the effect of having the mangrove breeding trait when located in a mangrove)

Name	Main Effects	ΔAIC	K
H1	Precip + LocalAgriculture + LocalUrban + LocalRainforest + LocalMangrove + spp[a]Mangrove	11.4	26
H2	Precip + LocalAnthropogenic	53.6	11
H3	Precip + LocalAgriculture + LocalUrban	31	15
H4	Precip + LocalAnthropogenic + m_trait·Mangrove	38.6	14
H5	Precip + LocalAgriculture + LocalUrban + m_trait·Mangrove	14.5	18
H6	Precip + LocalScrub+ LocalAnthropogenic + m_trait·Mangrove	37.2	18
H7	Precip + LocalAgriculture + LocalUrban + LocalScrub+ m_trait·Mangrove	15	22
H8	Precip + LocalRainforest + LocalAnthropogenic + m_trait·Mangrove	25.6	18
H9	Precip + LocalAgriculture + LocalUrban + LocalRainforest + m_trait·Mangrove	3.9	20
H10	Precip + LocalRainforest + LocalScrub+ LocalAnthropogenic + m_trait·Mangrove	26.8	22
H11	Precip + LocalAgriculture + LocalUrban + LocalRainforest + LocalScrub+ m_trait·Mangrove	3.9	26
H12	Precip + LocalAgriculture + LocalUrban + LocalRainforest + m_trait[a]Mangrove	0	22

646 [a]Two variables with full factorial interaction comprised of main effects and interaction terms
647 · cases where only the interaction term is evaluated
648

Table 2 Model parameters and diagnostics from our best model of mosquito counts. Means and confidence intervals are given on the log-scale, and symbols correspond to parameters in equation 1

Predictors	Symbol	Mean	95th % CI	p
Intercept	β_0	-0.75	-1.33 – -0.18	0.01
Precipitation	β_1	0.66	0.13 – 1.19	0.015
Mangrove Breeding Trait	β_2	-1.4	-2.12 – -0.69	<0.001
Mangrove site category	β_3	0.01	-1.29 – 1.31	0.99
Local Agriculture (Ae.ae.)	$\beta_{4(Ae.ae.)}$	0.2	-0.38 – 0.77	0.503
Local Agriculture (Ae.ta.)	$\beta_{4(Ae.ta.)}$	-1.35	-1.98 – -0.71	<0.001
Local Agriculture (Cx.qu.)	$\beta_{4(Cu.qu.)}$	-1.07	-1.65 – -0.49	<0.001
Local Agriculture (De.ma.)	$\beta_{4(De.ma.)}$	-1.08	-1.80 – -0.35	0.003
Local Urban (Ae.ae.)	$\beta_{5(Ae.ae.)}$	0.13	-0.42 – 0.68	0.644
Local Urban (Ae.ta.)	$\beta_{5(Ae.ta.)}$	-2.11	-2.91 – -1.30	<0.001
Local Urban (Cx.qu.)	$\beta_{5(Cu.qu.)}$	0.32	-0.22 – 0.85	0.248
Local Urban (De.ma.)	$\beta_{5(De.ma.)}$	-0.97	-1.67 – -0.26	0.007
Local Rainforest (Ae.ae.)	$\beta_{5(Ae.ae.)}$	-0.43	-0.99 – 0.13	0.129
Local Rainforest (Ae.ta.)	$\beta_{5(Ae.ta.)}$	-1.21	-1.94 – -0.48	0.001
Local Rainforest (Cx.qu.)	$\beta_{5(Cu.qu.)}$	-1.39	-2.03 – -0.76	<0.001
Local Rainforest (De.ma.)	$\beta_{5(De.ma)}$	-0.91	-1.64 – -0.18	0.014
Mangrove Breeding x Mangrove Site	β_7	2.32	0.83 – 3.81	0.002
Overdispersion	ϕ	0.0962		

Random Effects

Site Intercept	$\alpha_{2(i)}$	0.54		
Species Intercept	$\alpha_{0(s)}$	0.02		
Species x Precipitation Slope	$\alpha_{1(s)}$	0.24		
Correlation of Species random effects		-0.69		
Number of sites		30		
Number of species		4		
Observations		1568		

Marginal R^2 / Conditional R^2	0.601 / 0.695

649 Ae.ae., *Ae. aegypti*; Ae.ta., *Ae. taeniorhynchus*; Cx.qu., *Cx. quinquefasciatus*; De.ma., *De.*

650 *magnus*