Origin of Novel Coronavirus (COVID-19): A Computational Biology Study using Artificial Intelligence

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ABSTRACT
Origin of the COVID-19 virus has been intensely debated in the scientific community since the first infected cases were detected in December 2019. The disease has caused a global pandemic, leading to deaths of thousands of people across the world and thus finding origin of this novel coronavirus is important in responding and controlling the pandemic. Recent research results suggest that either bat or pangolin might be the original host of the virus based on comparative studies using its genomic sequences. This paper investigates the COVID-19 virus origin by using artificial intelligence (AI) and raw genomic sequences of the virus. More than 300 genome sequences of COVID-19 infected cases collected from different countries are explored and analysed using unsupervised clustering methods. The results obtained from various AI-enabled experiments using clustering algorithms demonstrate that all examined COVID-19 virus genomes belong to a cluster that also contains bat and pangolin coronavirus genomes. This provides evidences strongly supporting scientific hypotheses that bats and pangolins are probable hosts for the COVID-19 novel coronavirus. At the whole genome analysis level, our findings also indicate that bats are more likely the host for the COVID-19 virus than pangolins.

KEYWORDS
Artificial intelligence; AI; machine learning; coronavirus; COVID-19; SARS-CoV-2; origin; bat; pangolin; pandemic

INTRODUCTION

The COVID-19 pandemic has rapidly spread across many countries and disturbed lives of millions of people around the globe. The race to produce vaccines and medicines to mitigate its impacts and prevent a similar pandemic in future is still ongoing and no effective results have been reported yet. Studies on understanding the virus, which was named severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), are of utmost important in facilitating the discovery of treatment medicines and vaccines. Among those studies, finding origin of the COVID-19 virus (i.e. SARS-CoV-2) is crucial because it helps to understand the virus via its evolutionary relationships with other biological organisms and species based on physical or genetic characteristics. A study by Wu et al. [1] using a complete genome obtained from a patient who was a worker at a seafood market in Wuhan city, Hubei province, China shows that the virus is closely related to a group of SARS-like CoVs that were previously found present in bats in China. It is believed that bats are the most likely reservoir for the COVID-19 virus as it is very similar to a bat coronavirus. These results are supported by a separate study by Lu et al. [2] using genome sequences acquired from nine COVID-19 patients who were among early cases in Wuhan, China. Outcomes of a phylogenetic analysis suggest that the virus belongs to the genus *Betacoronavirus*, sub-genus *Sarbecovirus*, which includes many bat SARS-like CoVs and SARS CoVs. Likewise, Zhou et al. [3] also advocate a probable bat origin of SARS-CoV-2 by using complete genome sequences of five patients at the beginning of the outbreak in Wuhan, China. One of these sequences shows 96.2% similarity to the genome sequence of a coronavirus, which was previously obtained from a *Rhinolophus affinis* bat, found in Yunnan province of China.

In a recent study, Lam et al. [4] found two related lineages of CoVs in pangolin genome sequences sampled in Guangxi and Guangdong provinces in China, which have similar genomic organizations to SARS-CoV-2. That study suggests that pangolins could be a possible host of SARS-CoV-2 although they are solitary animals in an endangered status with relatively small population sizes. These findings are supported by Zhang et al. [5] who assembled a pangolin CoV draft

genome using a reference-guided scaffolding approach based on contigs taxonomically annotated to SARS-CoV-2, SARS-CoV, and bat SARS-like CoV. On the other hand, by analysing genomic features of SARS-CoV-2, i.e. mutations in the receptor binding domain portion of the spike protein and distinct backbone of the virus, Andersen et al. [6] determined that this novel coronavirus is originated through natural processes rather than from a laboratory manipulation.

This study presents a step further to suggesting the likely origin of the COVID-19 virus by using artificial intelligence (AI) methods to explore genome sequences obtained from more than 300 COVID-19 patients across the world. We use AI-enabled unsupervised clustering methods to demonstrate and emphasize the relationships between COVID-19 virus, bat CoVs and pangolin CoVs. Through analysing the results of clustering methods, we are able to suggest the sub-genus *Sarbecovirus* of the genus *Betacoronavirus* of SARS-CoV-2 and the more likely bat origin of the virus rather than a pangolin origin.

MATERIALS AND METHODS

We downloaded 334 complete genome sequences of SARS-CoV-2 available from the GenBank database, which is maintained by the National Center for Biotechnology Information (NCBI), in early April 2020. Among these sequences, 258 were reported from USA, 49 were from China and the rest were distributed through various countries from Asia to Europe and South America. Accession numbers and detailed distribution of these genome sequences across different countries are presented in Tables 4 and 5 in Appendix 1.

Most of reference sequences, e.g. ones within the *Alphacoronavirus* and *Betacoronavirus* genera, are also downloaded from the NCBI GenBank and Virus-Host DB (https://www.genome.jp/virushostdb/) that covers NCBI Reference Sequences (RefSeq, release 99, March 2, 2020). Genome sequences of Guangxi pangolin CoVs [4] are downloaded from the GISAID database (https://www.gisaid.org) with accession numbers EPI_ISL_410538 - EPI_ISL_410543. A Guangdong pangolin CoV genome [7] is also downloaded from GISAID with accession number EPI_ISL_410721. We employ three sets of reference sequences in this study with details presented in Tables 1-3. The selection of reference genomes at different taxonomic levels is based on a study in [8] that uses the supervised decision tree method to classify novel pathogens, which include SARS-CoV-2 sequences. We aim to traverse from high to low taxonomic levels to search for the COVID-19 virus origin through discovering its genus and sub-genus taxonomy and its closest genome sequences.

Table 1. Reference viruses from major virus classes at a high taxonomic level - Set 1

Virus (Accession Number)	Taxonomy	Virus (Accession Number)	Taxonomy
Human adenovirus D8 (AB448767)	Adenoviridae	Murine leukemia virus (AB187566)	Ortervirales
TT virus sle1957 (AM711976)	Anelloviridae	Human papillomavirus type 69 (AB027020)	Papillomaviridae
Staphylococcus phage S13' (AB626963)	Caudovirales	Adeno-associated virus - 6 (AF028704)	Parvoviridae
Chili leaf curl virus-Oman (KF229718)	Geminiviridae	Cotesia plutellae polydnavirus (AY651828)	Polydnaviridae
Meles meles fecal virus (JN704610)	Genomoviridae	Aves polyomavirus 1 (AF118150)	Polyomaviridae
Chlamydia phage 3 (AJ550635)	Microviridae	Middle East respiratory syndrome (MERS) CoV (NC_019843)	Riboviria

Unsupervised clustering methods are employed to cluster datasets comprising both query sequences (SARS-CoV-2) and reference sequences into clusters. In this paper, we propose the use of hierarchical clustering algorithm [9] and density-based spatial clustering of applications with noise (DBSCAN) method [10] for this purpose. With these two methods, we perform two steps to observe the clustering results that lead to conclusions about the taxonomy and origin of SARS-CoV-2. In the first step, we apply clustering algorithms to cluster the set of reference sequences only, and then use the same settings (i.e. values of parameters) of clustering algorithms to cluster a dataset that merges reference sequences and

Table 2. Reference viruses within the Riboviria realm - Set 2

Virus (Accession Number)	Taxonomy	Virus (Accession Number)	Taxonomy
Grapevine rupestris stem pitting-associated virus (GRSPV) 1 (AF057136)	Betaflexiviridae	Lymantria dispar cypovirus 14 (AF389452)	Reoviridae
Cucumber mosaic virus (AJ276479)	Bromoviridae	Hybrid snakehead virus (KC519324)	Rhabdoviridae
Chiba virus (AB042808)	Caliciviridae	Rice tungro spherical virus (NC_-001632)	Secoviridae
Mercadeo virus (NC_027819)	Flaviviridae	Bulbul CoV HKU11-796 (FJ376620)	Coronaviridae; DeltaCoV
Bunyamwera virus (NC_001925)	Peribunyaviridae	Avian infectious bronchitis virus (AIBV) (AY646283)	Coronaviridae; GammaCoV
Rice grassy stunt tenuivirus (NC_-002323)	Phenuiviridae	Human CoV NL63 (NC_005831)	Coronaviridae; AlphaCoV
Theiler's-like virus of rats (AB090161)	Picornaviridae	SARS CoV BJ01 (AY278488)	Coronaviridae; BetaCoV
Turnip mosaic virus (AB194796)	Potyviridae		

SARS-CoV-2 sequences. Through this step, we can find out reference sequences by which SARS-CoV-2 sequences form a group with. In the second step, we vary the settings of the clustering algorithms and observe changes in the clustering outcomes. With the second step, we are able to discover the closest reference sequences to the SARS-CoV-2 sequences and compare the similarities between genomes.

In the hierarchical clustering method, the cut-off parameter C plays as a threshold in defining clusters and thus C is allowed to change during our experiments. With regard to the DBSCAN method, the neighbourhood search radius parameter ε and the minimum number of neighbours parameter, which is required to identify a core point, are crucial in partitioning observations into clusters. In our experiments, we set the minimum number of neighbours to 3 and allow only the search radius parameter ε to vary. Outputs of the DBSCAN method may also include outliers, which are normally labelled as cluster "-1". To facilitate the execution of the clustering methods, we propose the use of pairwise distances between sequences based on the Jukes-Cantor method [11] and the maximum composite likelihood method [12]. The Jukes-Cantor method estimates evolutionary distances by the maximum likelihood approach using the number of substitutions between two sequences. With nucleotide sequences, the distance is defined as $d = -3/4 * ln(1 - p * 4/3)$ where p is the ratio between the number of positions where the substitution is to a different nucleotide and the number of positions in the sequences. On the other hand, the maximum composite likelihood method considers the sum of log-likelihoods of all pairwise distances in a distance matrix as a composite likelihood because these distances are correlated owing to their phylogenetic relationships. Tamura et al. [12] showed that estimates of pairwise distances and their related substitution parameters such as those of the Tamura-Nei model [13] can be obtained accurately and efficiently by maximizing this composite likelihood. The unweighted pair group method with arithmetic mean (UPGMA) method is applied to create hierarchical cluster trees, which are used to construct dendrogram plots for the hierarchical clustering method. The UPGMA method is also employed to generate phylogenetic trees in order to show results of the DBSCAN algorithm.

RESULTS AND DISCUSSIONS

We start the experiments to search for taxonomy and origin of SARS-CoV-2 with the first set of reference genome sequences (Set 1 in Table 1). This set consists of much more diversified viruses than the other two sets as it includes representatives from major virus classes at the highest available virus taxonomic level. With a large coverage of various types of viruses, the use of this reference set minimizes the probability of missing out any known type of reference virus. Outcomes of the hierarchical clustering and DBSCAN methods are presented in Figs. 1 and 2, respectively. In these figures, we use 16 SARS-CoV-2 sequences representing 16 countries in Table 5 for the demonstration purpose.

Table 3. Reference viruses in the genus AlphaCoV and BetaCoV - Set 3

Virus (Accession Number)	Taxonomy	Virus (Accession Number)	Taxonomy
Transmissible gastroenteritis virus (TGEV) (NC_038861)	AphaCoV	SARS CoV BJ01 (AY278488)	BetaCoV; Sarbecovirus
Mink CoV WD1127 (NC_-023760)	AlphaCoV	Bat SARS CoV RsSHC014 (KC881005)	BetaCoV; Sarbecovirus
Porcine epidemic diarrhea virus (PEDV) (NC_003436)	AlphaCoV	Bat SARS CoV WIV1 (KF367457)	BetaCoV; Sarbecovirus
Rhinolophus bat CoV HKU2 (NC_009988)	AlphaCoV	Bat SARS CoV Rp3 (DQ071615)	BetaCoV; Sarbecovirus
Human CoV 229E (NC_-002645)	AlphaCoV	Bat SARS CoV Rs672/2006 (FJ588686)	BetaCoV; Sarbecovirus
Human CoV NL63 (NC_-005831)	AlphaCoV	Bat SARS CoV Rf1 (DQ412042)	BetaCoV; Sarbecovirus
Human CoV OC43 (NC_-006213)	BetaCoV; Embecovirus	Bat SARS CoV Longquan-140 (KF294457)	BetaCoV; Sarbecovirus
Murine hepatitis virus (MHV) (AC_000192)	BetaCoV; Embecovirus	Bat SARS CoV HKU3-1	BetaCoV; Sarbecovirus
Rousettus bat CoV HKU9 (NC_009021)	BetaCoV; Nobecovirus	Bat SARS CoV ZXC21 (MG772934)	BetaCoV; Sarbecovirus
Rousettus bat CoV GCCDC1 (NC_030886)	BetaCoV; Nobecovirus	Bat SARS CoV ZC45 (MG772933)	BetaCoV; Sarbecovirus
MERS CoV (NC_019843)	BetaCoV; Merbecovirus	Bat CoV RaTG13 (MN996532)	BetaCoV; Sarbecovirus
Pipistrellus bat CoV HKU5 (NC_009020)	BetaCoV; Merbecovirus	Guangxi pangolin CoV GX/P4L (EPI_ISL_410538)	BetaCoV; Sarbecovirus
Tylonycteris bat CoV HKU4 (NC_009019)	BetaCoV; Merbecovirus	Guangxi pangolin CoV GX/P1E (EPI_ISL_410539)	BetaCoV; Sarbecovirus
Bat Hp-betaCoV/Zhejiang2013 (NC_025217)	BetaCoV; Hibecovirus	Guangxi pangolin CoV GX/P5L (EPI_ISL_410540)	BetaCoV; Sarbecovirus
SARS CoV BtKY72 (KY352407)	BetaCoV; Sarbecovirus	Guangxi pangolin CoV GX/P5E (EPI_ISL_410541)	BetaCoV; Sarbecovirus
Bat CoV BM48-31/BGR/2008 (GU190215)	BetaCoV; Sarbecovirus	Guangxi pangolin CoV GX/P2V (EPI_ISL_410542)	BetaCoV; Sarbecovirus
SARS CoV LC5 (AY395002)	BetaCoV; Sarbecovirus	Guangxi pangolin CoV GX/P3B (EPI_ISL_410543)	BetaCoV; Sarbecovirus
SARS CoV SZ3 (AY304486)	BetaCoV; Sarbecovirus	Guangdong pangolin CoV (EPI_ISL_410721)	BetaCoV; Sarbecovirus
SARS CoV Tor2 (AY274119)	BetaCoV; Sarbecovirus		

The first released SARS-CoV-2 genome of each country is selected for these experiments. Clustering outcomes on all 334 sequences are presented in Fig. 9 in Appendix 2, which shows results similar to those reported here. Both clustering methods consistently demonstrate that SARS-CoV-2 sequences belong to the Riboviria cluster among 12 major virus classes (*Adenoviridae*, *Anelloviridae*, *Caudovirales*, *Geminiviridae*, *Genomoviridae*, *Microviridae*, *Ortervirales*, *Papillomaviridae*, *Parvoviridae*, *Polydnaviridae*, *Polyomaviridae*, and *Riboviria*). The Middle East respiratory syndrome (MERS) CoV, which caused the MERS outbreak in 2012, is chosen as a representative of the Riboviria realm. In hierarchical clustering (Fig. 1), when combined with reference genomes, SARS-CoV-2 genomes do not create a new cluster on their own but form a cluster with the MERS CoV, i.e. cluster "8". With the DBSCAN method (Fig. 2), SARS-CoV-2 genomes also do not create their own cluster but form the cluster "1" with the MERS CoV. These clustering results suggest that the SARS-CoV-2 belongs to the Riboviria realm.

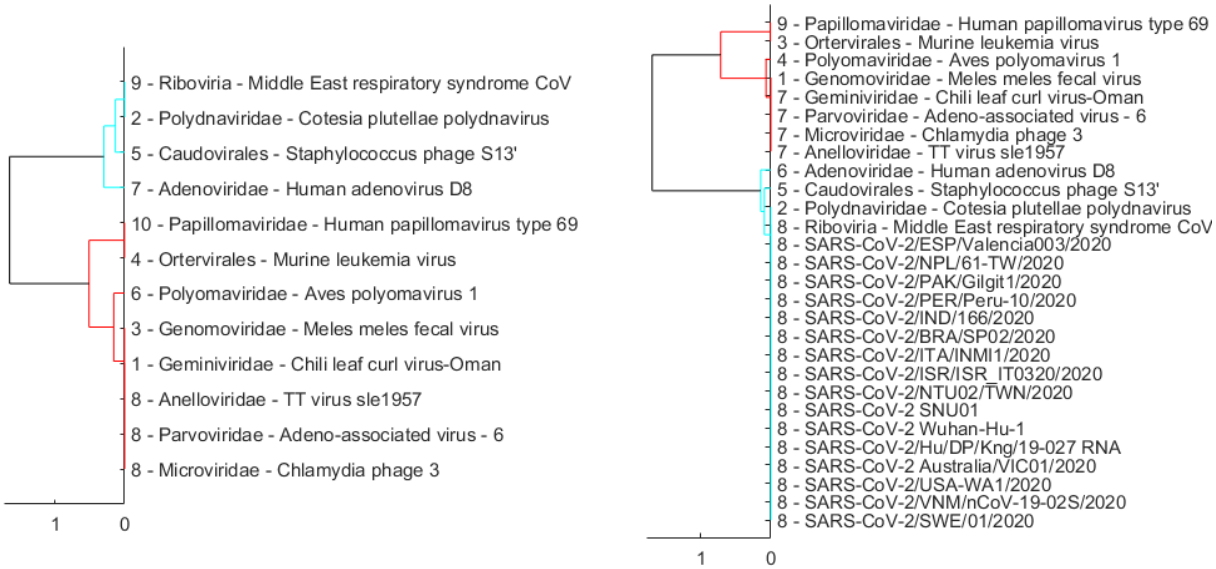

Fig. 1. Dendrogram plots showing hierarchical clustering results using only the reference sequences in Set 1 (Table 1) with the cut-off parameter C equal to $5 * 10^{-4}$ (left), and using a set that merges 16 representative SARS-CoV-2 sequences and reference sequences with C is also set to $5 * 10^{-4}$ (right). A number at the beginning of each virus name indicates the cluster that virus belongs to after clustering.

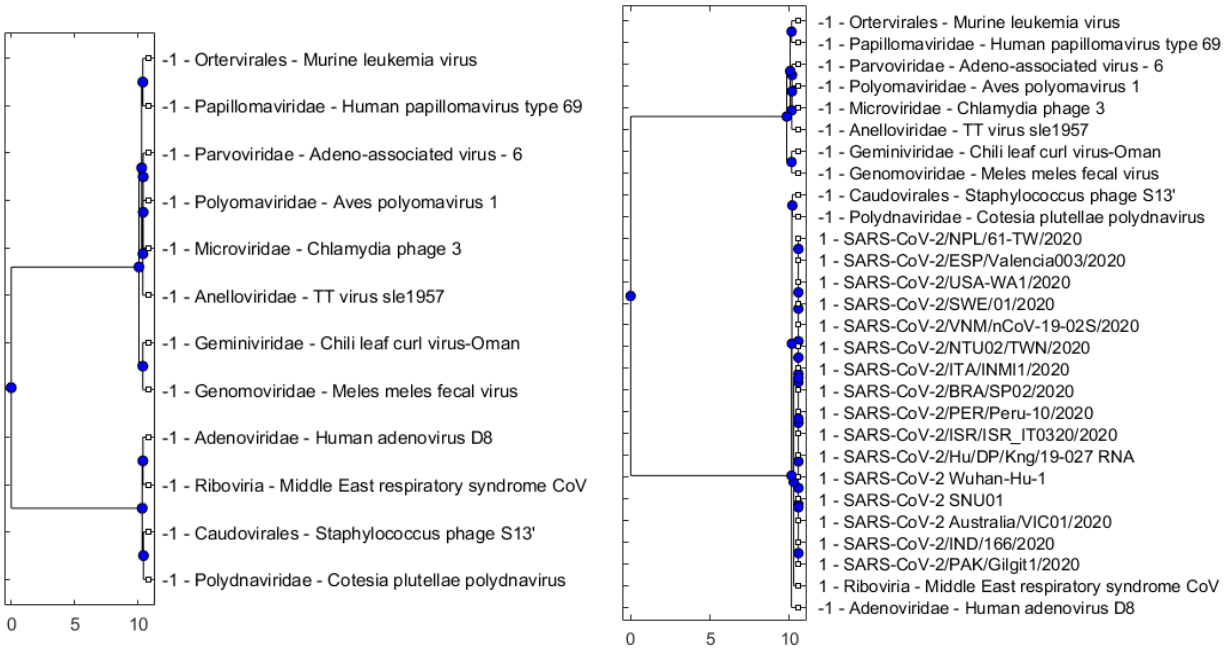

Fig. 2. Phylogenetic trees showing DBSCAN results using only the reference sequences in Set 1 (Table 1) with the search radius parameter ε equal to 0.7 (left), and using a set that merges SARS-CoV-2 sequences and reference sequences with $\varepsilon = 0.7$ (right). As Set 1 includes representatives of major virus classes and the minimum number of neighbours is set to 3 while $\varepsilon = 0.7$, DBSCAN considers individual viruses as outliers (left). When the dataset is expanded to include SARS-CoV-2 sequences, DBSCAN forms cluster "1" that includes all SARS-CoV-2 sequences and the MERS CoV, which represents the Riboviria realm (right).

Once we have been able to identify SARS-CoV-2 as belonging to the Riboviria realm, we move to the next lower taxonomic level that consists of 12 virus families within Riboviria. These families are presented in Set 2 (Table 2) that includes *Betaflexiviridae*, *Bromoviridae*, *Caliciviridae*, *Coronaviridae*, *Flaviviridae*, *Peribunyaviridae*, *Phenuiviridae*, *Picornaviridae*, *Potyviridae*, *Reoviridae*, *Rhabdoviridae*, and *Secoviridae*. Results of the two clustering methods presented in Figs. 3 and 4 show that SARS-CoV-2 sequences form a group with viruses in the *Coronaviridae* family. As we also include in Set 2 representatives of four genera in the *Coronaviridae* family (i.e. *Alphacoronavirus - AlphaCoV*, *Betacoronavirus - BetaCoV*, *Deltacoronavirus*, *Gammacoronavirus*), we are able to look further to the next taxonomic level within this experiment and observe that SARS-CoV-2 belongs to the *Betacoronavirus* genus (see Figs. 3 and 4).

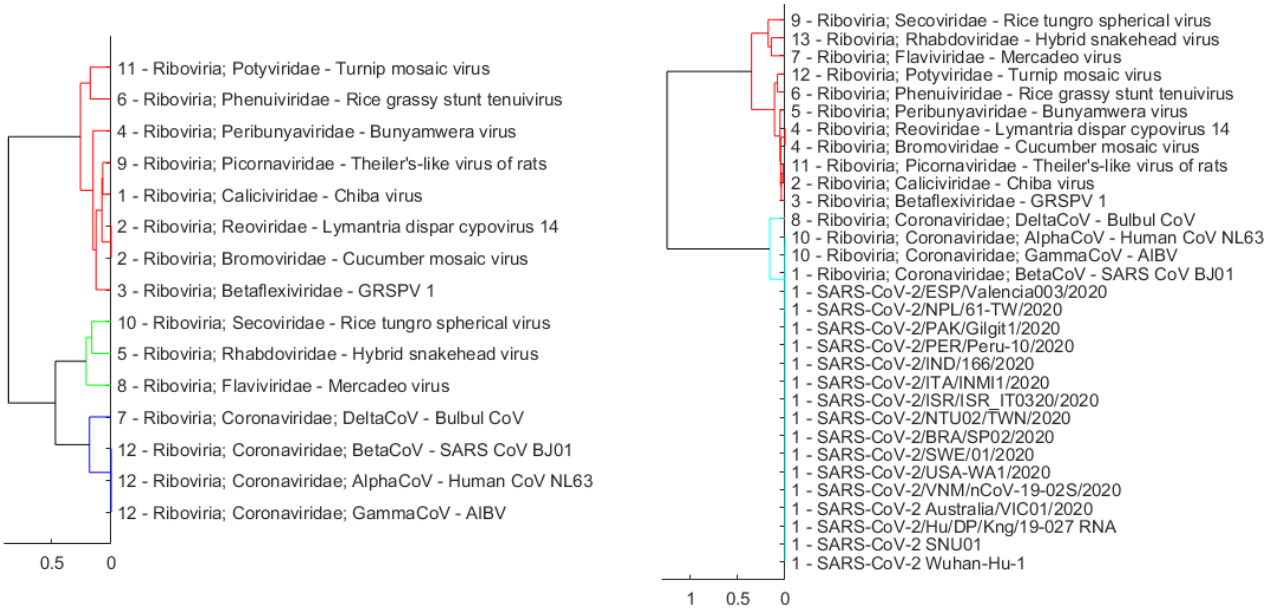

Fig. 3. Dendrogram plots showing hierarchical clustering results using only the reference sequences in Set 2 (Table 2) with the cut-off parameter C equal to 0.001 (left), and using a set that merges SARS-CoV-2 sequences and reference sequences with C also set to 0.001 (right).

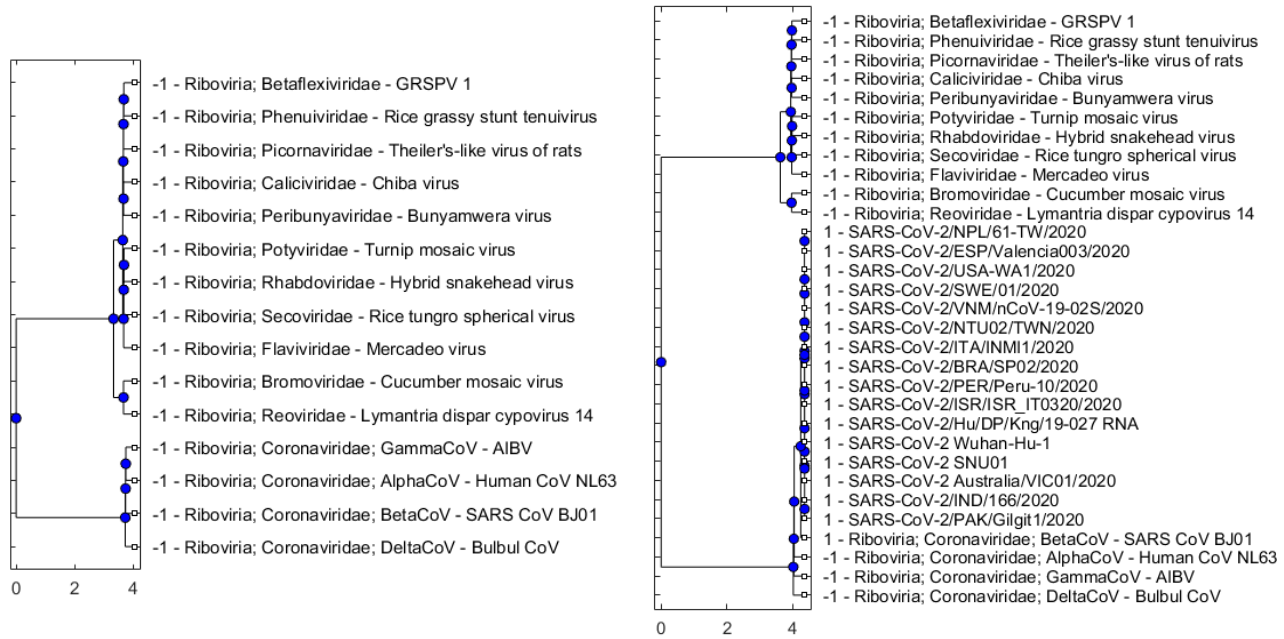

Fig. 4. Phylogenetic trees showing DBSCAN results using only the reference sequences in Set 2 (Table 2) with the search radius parameter ε equal to 0.6 (left), and using a set that merges SARS-CoV-2 sequences and reference sequences with ε also being set to 0.6 (right).

There are 5 sub-genera in the *Betacoronavirus* genus, including *Embecovirus*, *Nobecovirus*, *Merbecovirus*, *Hibecovirus*, *Sarbecovirus*. In the next lower taxonomic level, we investigate which of these sub-genera that SARS-CoV-2 belongs to or whether SARS-CoV-2 genomes create a new cluster on their own. We use Set 3 that includes 37 reference sequences (Table 3) for this investigation. Six representatives of the *Alphacoronavirus* genus, which is proximal to the *Betacoronavirus* genus, are also included in Set 3. The rest of Set 3 comprises 2 viruses of the *Embecovirus* sub-genus, 2 viruses of the *Nobecovirus* sub-genus, 3 viruses of the *Merbecovirus* sub-genus, 1 virus of the *Hibecovirus* sub-genus and 23 viruses of the *Sarbecovirus* sub-genus. Most of the representatives of the *Sarbecovirus* sub-genus are SARS CoVs and bat SARS-like CoVs. Notably, we also include in this set 5 sequences of Guangxi pangolin CoVs deposited to the GISAID database by Lam et al. [4] and a sequence of Guangdong pangolin CoV constructed in [7].

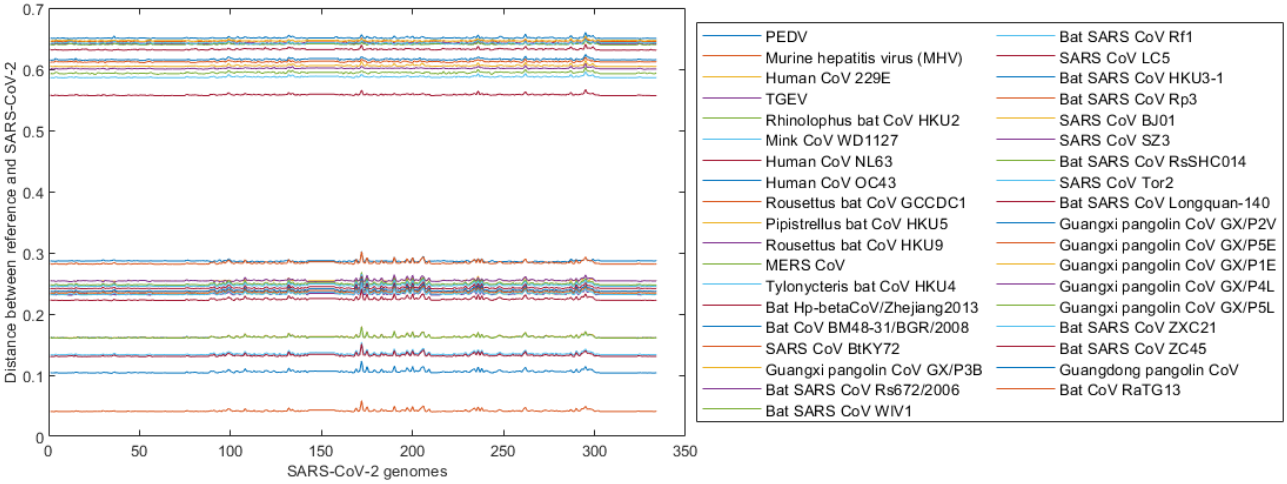

Fig. 5. Distances between each of the reference genomes in Set 3 (Table 3) with 334 SARS-CoV-2 genomes where the latter are ordered by the released date (earliest to latest) over a period of approximately 3 months, from late December 2019 to late March 2020. These pairwise distances are computed based on the Jukes-Cantor method using whole genomes. The line at the bottom, for instance, represents the distances between the bat CoV RaTG13 genome with each of the 334 SARS-CoV-2 genomes.

Evolutionary distances between each of the reference genomes in Set 3 (Table 3) to the 334 SARS-CoV-2 genomes based on the Jukes-Cantor method are presented in Fig. 5. We can observe that these distances are almost constant across 334 SARS-CoV-2 sequences, which are collected in 16 countries (Table 5) over approximately 3 months since late December 2019. This implies that not much variation of SARS-CoV-2 genomes has occurred over time and across countries. Despite this characteristic, SARS-CoV-2 is more easily spread than two other coronaviruses - SARS-CoV and MERS CoV as there have been approximately 4 million confirmed cases of COVID-19 globally, including nearly 275,000 deaths, reported to the World Health Organization in the middle of May 2020 [14]. As in Fig. 5, there are several groups of reference genomes shown via the closeness of the distance lines. For example, the top group contains genomes of *AlphaCoV* viruses (refer to the taxonomy in Table 3) that are much evolutionarily divergent from SARS-CoV-2 sequences. The middle group of lines comprises most of the *BetaCoV* viruses, especially those in the *Sarbecovirus* sub-genus. The bottom lines identify reference viruses that are closest to SARS-CoV-2, which include bat CoV RaTG13, Guangdong pangolin CoV, bat SARS CoV ZC45 and bat SARS CoV ZXC21. The bat CoV RaTG13 line at the bottom is notably distinguished from other lines while the Guangdong pangolin CoV line is the second closest to SARS-CoV-2. The similarity between bat CoV RaTG13, Guangdong pangolin CoV and Guangxi pangolin CoV GX/P4L with SARS-CoV-2/Australia/VIC01/2020, produced by the SimPlot software [15], is displayed in Fig. 6. Consistent with the results presented in Fig. 5, bat CoV RaTG13 is shown closer to SARS-CoV-2 than pangolin CoVs.

Fig. 7 shows outcomes of the hierarchical clustering method using Set 3 of reference sequences in Table 3. With the cut-off parameter C is set equal to 0.7, the hierarchical clustering algorithm separates the reference sequences into 6 clusters in which cluster "5" comprises all viruses of the *Sarbecovirus* sub-genus, including many SARS CoVs, bat SARS-like CoVs and pangolin CoVs (Fig. 7A). It is observed that the algorithm reasonably groups viruses into clusters, for example, the genus *AlphaCoV* is represented by cluster "4" while sub-genera *Embecovirus*, *Nobecovirus*, *Merbecovirus*, and *Hibecovirus* are labelled as clusters "3", "6", "2", and "1", respectively. Using the same cut-off value of 0.7, we next perform clustering on a dataset that merges reference sequences and 16 representative SARS-CoV-2 sequences (see Fig. 7B). Results on all 334 SARS-CoV-2 sequences, which are similar to those on the 16 representative sequences, are provided in Fig. 10 in Appendix 2. The outcome presented in Fig. 7B shows that the 16 representative SARS-CoV-2 sequences fall into cluster "5", which comprises the *Sarbecovirus* sub-genus. The number of clusters is still 6 and the membership structure of the clusters is the same as in the case of clustering reference sequences only (Fig. 7A), except that the *Sarbecovirus* cluster now has been expanded to also contain SARS-CoV-2 sequences. By comparing Figs. 7A and 7B, we believe that SARS-CoV-2 genomes are naturally part of the *Sarbecovirus* sub-genus. This realization is substantiated by moving to Fig. 7C that shows a clustering outcome when the cut-off C is decreased to 0.1. In Fig. 7C, while 3

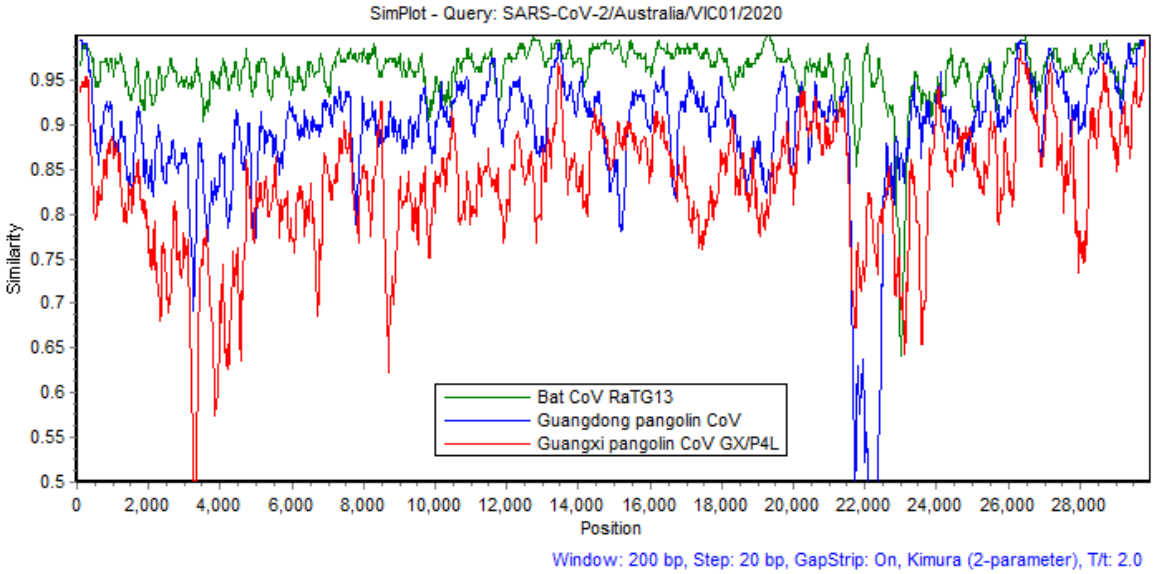

Fig. 6. Similarity between genomes of bat CoV RaTG13, Guangdong pangolin CoV and Guangxi pangolin CoV GX/P4L with the SARS-CoV-2/Australia/VIC01/2020 genome sequence.

members of the *Merbecovirus* sub-genus (i.e. *Pipistrellus* bat CoV HKU5, *Tylonycteris* bat CoV HKU4 and MERS CoV) are divided into 3 clusters ("12", "2" and "4") or members of the *Sarbecovirus* cluster are separated themselves into 2 clusters "'1" and "11", sequences of SARS-CoV-2 still join the cluster "11' with other members of *Sarbecovirus* such as 3 bat viruses (bat SARS CoV ZC45, bat SARS CoV ZXC21, bat CoV RaTG13) and 7 pangolin CoVs.

As the cut-off parameter C decreases, the number of clusters increases. This is an expected outcome because the cut-off threshold line moves closer to the leaves of the dendrogram. When the cut-off C is reduced to 0.03 (Fig. 7D), there are only 2 viruses (bat CoV RaTG13 and Guangdong pangolin CoV) that can form a cluster with SARS-CoV-2 (labelled as cluster "15"). These are 2 viruses closest to SARS-CoV-2 based on the whole genome analysis. Results in Figs. 7C and 7D therefore provide evidence that bat or pangolin could be a possible host for SARS-CoV-2. We next reduce the cut-off C to 0.01 as in Fig. 7E. At this stage, only bat CoV RaTG13 is within the same cluster with SARS-CoV-2 (cluster "17"). We thus believe that bats are the more probable host for SARS-CoV-2 than pangolins. The inference of our AI-enabled analysis is in line with a result in [16] that investigates the polyprotein 1ab of SARS-CoV-2 and suggests that this novel coronavirus has more likely been arisen from viruses infecting bats rather than pangolins.

When the cut-off C is reduced to 0.001 as in Fig. 7F, we observe that the total number of clusters now increases to 29 and more importantly, SARS-CoV-2 sequences do not combine with any other reference viruses but form its own cluster "19". Could we use this clustering result (Fig. 7F) to infer that SARS-CoV-2 may not originate from bats or pangolins? This is a debatable question because the answer depends on the level of details we use to differentiate between the species or organisms. The cut-off parameter in hierarchical clustering can be considered as the level of details. With the result obtained in Fig. 7D, we support a hypothesis that bat or pangolin is the probable origin of SARS-CoV-2. This is because we observe that the similarity between SARS-CoV-2 and bat CoV RaTG13 (or Guangdong pangolin CoV) is considerably large compared to the similarity between viruses that are originated from the same host. For example, bat SARS-like CoVs such as bat SARS CoV ZC45 and bat SARS CoV WIV1 have the same bat origin. In Fig. 7D, these viruses however are separated into 2 different clusters ("2" and "3") while all 16 SARS-CoV-2 representatives are grouped together with bat CoV RaTG13 and Guangdong pangolin CoV in cluster "15". This demonstrates that the difference between the same origin viruses (bat SARS CoV ZC45 and bat SARS CoV WIV1) is larger than the difference between SARS-CoV-2 and bat CoV RaTG13 (or Guangdong pangolin CoV). Therefore, SARS-CoV-2 is deemed very likely originated from the same host with bat CoV RaTG13 or Guangdong pangolin CoV, which is bat or pangolin, respectively.

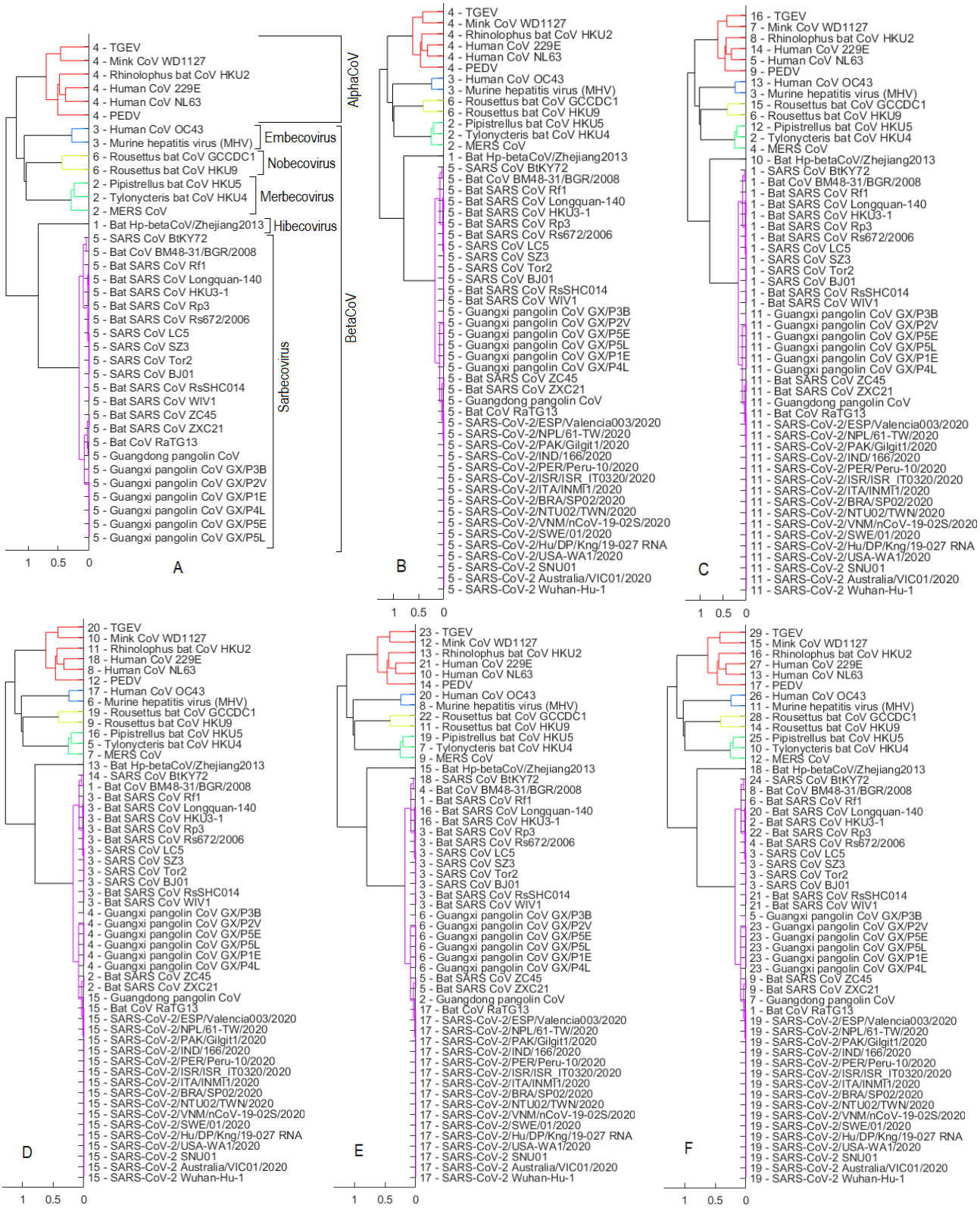

Fig. 7. Set 3 of reference sequences - results obtained by using hierarchical clustering via dendrogram plots. Numbers at the beginning of each virus label indicate the cluster that virus is a member of as a result of the clustering algorithm. (A) when the cut-off C is equal to 0.7 and using Set 3 of reference sequences only: there are 6 clusters where cluster "5" covers all examined viruses in the *Sarbecovirus* sub-genus. (B) when cut-off C is still equal to 0.7 and the dataset now merges between reference sequences and 16 representative SARS-CoV-2 sequences (merged set). (C) using the merged set with $C = 0.1$. (D) using the merged set with $C = 0.03$. (E) using the merged set with $C = 0.01$. (F) using the merged set with $C = 0.001$.

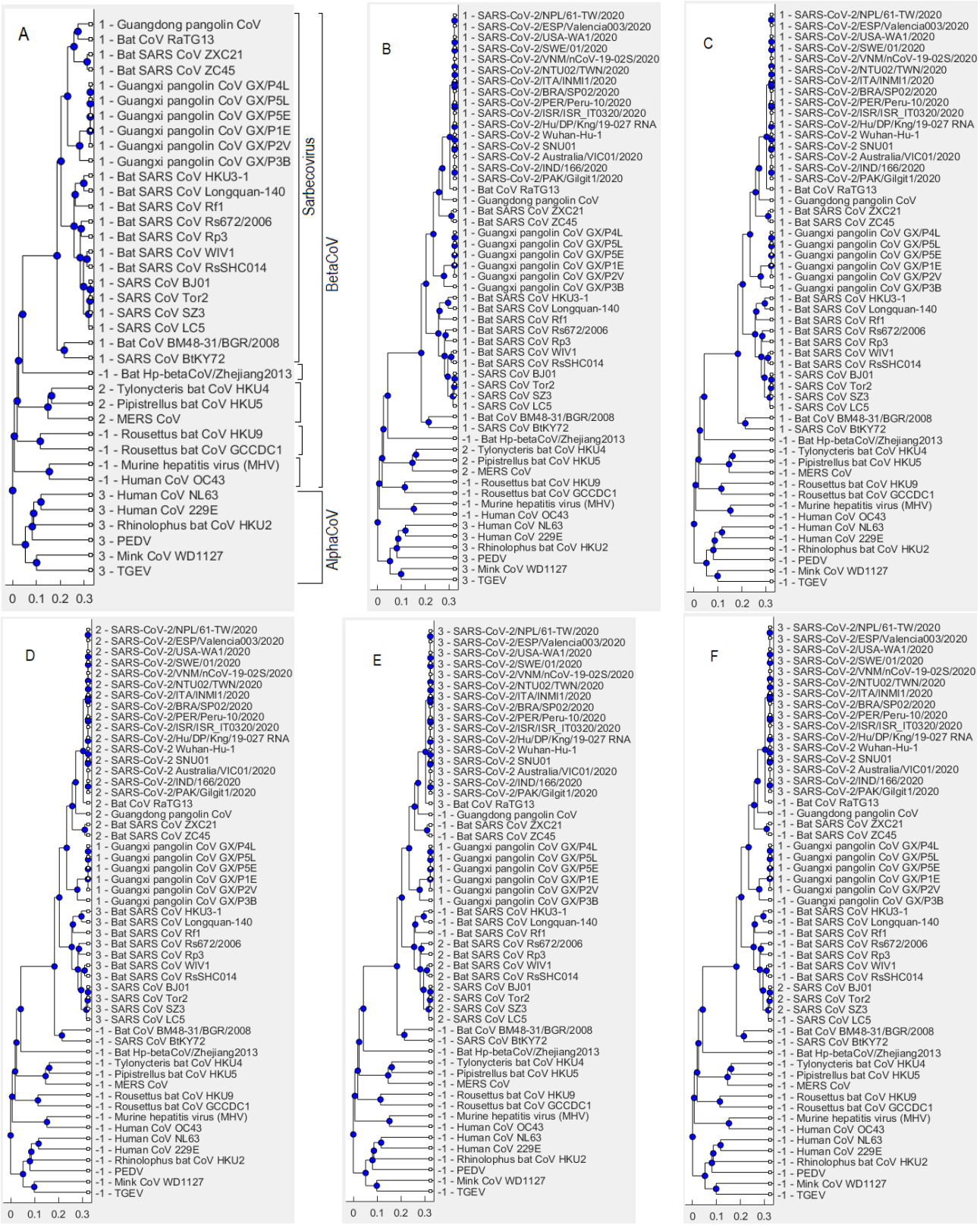

Fig. 8. Set 3 of reference sequences - results obtained by using DBSCAN via phylogenetic trees. Numbers at the beginning of each virus label indicate the cluster that virus is a member of as a result of the clustering algorithm. (A) when the search radius parameter ε is equal to 0.55 and using Set 3 of reference sequences only: there are 3 clusters where the cluster "1" covers all examined viruses in the *Sarbecovirus* sub-genus. (B) when search radius ε is still equal to 0.55 and the dataset now merges between reference sequences and 16 representative SARS-CoV-2 sequences (merged set). (C) using the merged set with $\varepsilon = 0.3$. (D) using the merged set with $\varepsilon = 0.15$. (E) using the merged set with $\varepsilon = 0.1$. (F) using the merged set with $\varepsilon = 0.01$.

Clustering outcomes of the DBSCAN method via phylogenetic trees using Set 3 of reference sequences (Table 3) are presented in Fig. 8. We first apply DBSCAN to reference sequences only, which results in 3 clusters and several outliers (Fig. 8A). The search radius parameter ε is set equal to 0.55. As we set the minimum number of neighbours parameter to 3, it is expected that viruses of the sub-genera *Embecovirus*, *Nobecovirus* and *Hibecovirus* are detected as outliers "-1" because there are only 1 or 2 viruses in these sub-genera. Three viruses of the *Merbecovirus* sub-genus (i.e. *Pipistrellus* bat CoV HKU5, *Tylonycteris* bat CoV HKU4 and MERS CoV) are grouped into the cluster "2". All examined viruses of the *Sarbecovirus* sub-genus are joined in cluster "1" while the *AlphaCoV* viruses are combined into cluster "3". Fig. 8B shows an outcome of DBSCAN with the same ε value of 0.55 and the dataset has been expanded to include 16 representative SARS-CoV-2 sequences. We observe that genomes of SARS-CoV-2 fall into the cluster "1", which includes all the examined *Sarbecovirus* viruses. When ε is decreased to 0.3 in Fig. 8C, all members of the *Merbecovirus* cluster or the *AlphaCoV* cluster become outliers while 16 SARS-CoV-2 genomes still stick with the *Sarbecovirus* cluster. In line with the results obtained by using hierarchical clustering in Fig. 7, those obtained in Fig. 8B and 8C using the DBSCAN method give us the confidence to confirm that SARS-CoV-2 is part of the *Sarbecovirus* sub-genus. Fig. 8D shows that bat CoV RaTG13 and Guangdong pangolin CoV are closest to SARS-CoV-2 as they join with 16 SARS-CoV-2 representatives in cluster "2". This again emphasizes the probable bat or pangolin origin of SARS-CoV-2. By reducing ε to 0.1 as in Fig. 8E, the Guangdong pangolin CoV becomes an outlier whilst SARS-CoV-2 sequences form a cluster ("3") with only bat CoV RaTG13. This further confirms our findings when using the hierarchical clustering in Fig. 7 that bats are more likely a reservoir host for the SARS-CoV-2 than pangolins. When ε is decreased to 0.01 as in Fig. 8F, SARS-CoV-2 genomes form its own cluster "3", which is separated with any bat or pangolin sequences. As with the result in Fig. 7F by the hierarchical clustering, this result also raises a question whether SARS-CoV-2 is really originated from bats or pangolins. In Fig. 8D, it is again observed that the similarity between SARS-CoV-2 and bat CoV RaTG13 (or Guangdong pangolin CoV) is larger than the similarity between bat SARS CoV ZC45 (cluster "2") and bat SARS CoV WIV1 (cluster "3"), which have the same bat origin. We thus suggest that SARS-CoV-2 probably has the same origin with bat CoV RaTG13 or Guangdong pangolin CoV. In other words, bat or pangolin is the probable origin of SARS-CoV-2.

All results presented above are obtained using the pairwise distances estimated by the Jukes-Cantor method. Results based on distances calculated by the maximum composite likelihood method are reported in Appendix 3, which are similar to those obtained by using the Jukes-Cantor method.

CONCLUSIONS

The severity of COVID-19 pandemic has initiated a race in finding origin of the COVID-19 virus. Studies on DNA sequences obtained from early patients in Wuhan city in China suggest the probable bat origin of the virus based on similarities between these sequences and those obtained from bat CoVs previously reported also in China. Other studies afterwards found that SARS-CoV-2 DNA sequences are also similar to pangolin CoV sequences and accordingly raised a hypothesis on the pangolin host for the COVID-19 virus. This paper has investigated origin of the COVID-19 virus using unsupervised clustering methods and more than 300 raw DNA sequences of SARS-CoV-2 collected from various countries around the world. Outcomes of these AI-enabled methods are analysed, which lead to a confirmation on the *Coronaviridae* family of the COVID-19 virus. More specifically, the SARS-CoV-2 belongs to the sub-genus *Sarbecovirus* within the genus *Betacoronavirus* that includes SARS-CoV, which caused the global SARS pandemic in 2002-2003 [17; 18]. The results of various clustering experiments show that the SARS-CoV-2 sequences are more likely to form a cluster with the bat CoV sequence RaTG13 than pangolin CoVs, which were constructed from (frozen tissue) samples collected in Guangxi and Guangdong provinces in China. This indicates that bats are more likely the reservoir for the COVID-19 virus than pangolins. This study among many AI studies in the fight against the COVID-19 pandemic [19] has shown the power and capabilities of AI in this challenging battle, especially from the computational biology and medicine perspective. The findings of this research on the large dataset of 334 SARS-CoV-2 genomic sequences provide more insights about the COVID-19 virus and thus facilitate the progress on discovering medicines and vaccines to cure and prevent this deadly virus. A further research in this direction is strongly encouraged by a recent success of AI in identifying powerful new kinds of antibiotic from a pool of more than 100 million molecules as published in [20]. As AI is capable of analysing and discovering knowledge from large datasets in an intelligent and efficient manner, finding a COVID-19 vaccine using

AI is a realistic hope [21].

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APPENDIX 1

Tables 4 and 5 in this appendix present accession numbers and detailed distribution of 334 SARS-CoV-2 complete genomes across different countries obtained from the NCBI GenBank database.

Table 4. Accession numbers of 334 SARS-CoV-2 genome sequences obtained from NCBI GenBank in early April 2020, sorted by date released

MN908947, MN985325, MN975262, MN938384, MN988713, MN997409, MN994468, MN994467, MN988669, MN988668, MN996531, MN996530, MN996529, MN996528, MN996527, MT007544, MT019533, MT019532, MT019531, MT019530, MT019529, MT020881, MT020880, MT027064, MT027063, MT027062, MT039890, MT039888, MT039887, MT039873, MT049951, MT044258, MT044257, MT066176, MT066175, MT072688, MT093631, MT093571, MT106054, MT106053, MT106052, MT118835, MT123293, MT123292, MT123291, MT123290, LC528233, LC528232, MT126808, MT135044, MT135043, MT135042, MT135041, MT152824, MT050493, MT012098, LC529905, MT159722, MT159721, MT159720, MT159719, MT159718, MT159717, MT159716, MT159715, MT159714, MT159713, MT159712, MT159711, MT159710, MT159709, MT159708, MT159707, MT159706, MT159705, MT121215, MT066156, MT163719, MT163718, MT163717, MT163716, MT184913, MT184912, MT184911, MT184910, MT184909, MT184908, MT184907, MT188341, MT188340, MT188339, MT192773, MT192772, MT192765, MT192759, MT198652, MT226610, MT233523, MT233522, MT233519, MT240479, MT246667, MT246490, MT246489, MT246488, MT246487, MT246486, MT246485, MT246484, MT246482, MT246481, MT246480, MT246479, MT246478, MT246477, MT246476, MT246475, MT246474, MT246473, MT246472, MT246471, MT246470, MT246469, MT246468, MT246467, MT246466, MT246464, MT246462, MT246461, MT246460, MT246459, MT246458, MT246457, MT246456, MT246455, MT246454, MT246453, MT246452, MT246451, MT246450, MT246449, MT233526, MT253710, MT253709, MT253708, MT253707, MT253706, MT253705, MT253704, MT253703, MT253702, MT253701, MT253700, MT253699, MT253698, MT253697, MT253696, MT251980, MT251979, MT251978, MT251977, MT251976, MT251975, MT251974, MT251973, MT251972, LC534419, LC534418, MT259287, MT259286, MT259285, MT259284, MT259282, MT259281, MT259280, MT259278, MT259277, MT259275, MT259274, MT259273, MT259271, MT259269, MT259268, MT259267, MT259266, MT259264, MT259263, MT259261, MT259260, MT259258, MT259257, MT259256, MT259254, MT259253, MT259252, MT259251, MT259250, MT259249, MT259248, MT259247, MT259246, MT259245, MT259244, MT259243, MT259241, MT259239, MT259237, MT259236, MT259235, MT259231, MT259230, MT259229, MT259228, MT259227, MT259226, MT258383, MT258382, MT258381, MT258380, MT258379, MT258378, MT258377, MT263469, MT263468, MT263467, MT263465, MT263464, MT263463, MT263462, MT263459, MT263458, MT263457, MT263456, MT263455, MT263454, MT263453, MT263452, MT263451, MT263450, MT263449, MT263448, MT263447, MT263446, MT263445, MT263444, MT263443, MT263442, MT263441, MT263440, MT263439, MT263438, MT263437, MT263436, MT263435, MT263434, MT263433, MT263432, MT263431, MT263430, MT263429, MT263428, MT263426, MT263425, MT263424, MT263423, MT263422, MT263421, MT263420, MT263419, MT263418, MT263417, MT263416, MT263415, MT263414, MT263413, MT263412, MT263411, MT263410, MT263408, MT263406, MT263405, MT263404, MT263403, MT263402, MT263400, MT263399, MT263398, MT263396, MT263395, MT263394, MT263392, MT263391, MT263390, MT263388, MT263387, MT263386, MT263384, MT263383, MT263382, MT263381, MT263074, MT262993, MT262916, MT262915, MT262914, MT262913, MT262912, MT262911, MT262910, MT262909, MT262908, MT262907, MT262906, MT262905, MT262904, MT262903, MT262902, MT262901, MT262900, MT262899, MT262898, MT262897, MT262896, MT276598, MT276597, MT276331, MT276330, MT276329, MT276328, MT276327, MT276326, MT276325, MT276324, MT276323.

Table 5. The number of COVID-19 genomic sequences collected from various countries

Countries	Number of sequences	Countries	Number of sequences
USA	258	India	2
China	49	Brazil	1
Japan	5	Italy	1
Spain	4	Peru	1
Taiwan	3	Nepal	1
Vietnam	2	South Korea	1
Israel	2	Australia	1
Pakistan	2	Sweden	1

APPENDIX 2

In this Appendix, we first present results of the hierarchical clustering method applied to the dataset that combines Set 1 of reference sequences (Table 1) with all 334 SARS-CoV-2 sequences (see Fig. 9). We then show results of the hierarchical clustering (Fig. 10) and DBSCAN (Fig. 11) on a dataset that combines all 334 SARS-CoV-2 sequences and reference sequences in Set 3 (Table 3).

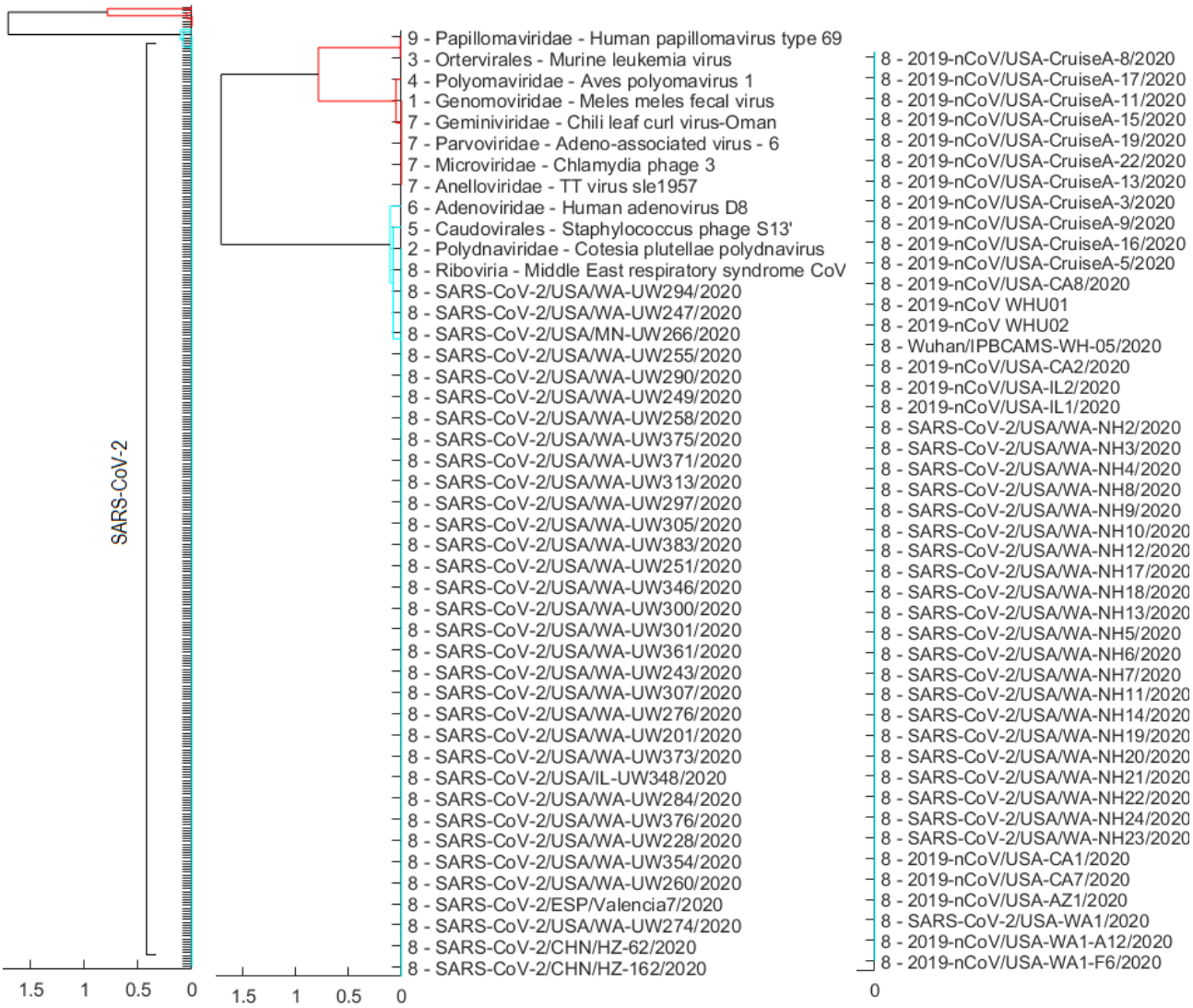

Fig. 9. Results shown via a dendrogram plot (left) of the hierarchical clustering method applied to the dataset that combines reference sequences in Set 1 (Table 1) and all 334 SARS-CoV-2 sequences. The middle figure shows in detail (zoom in) the top part of the dendrogram plot while the right figure shows the bottom part of the plot. All 334 SARS-CoV-2 sequences are grouped in cluster "8", which also includes the Middle East respiratory syndrome CoV of the Riboviria realm. This means that SARS-CoV-2 belongs to the Riboviria realm. These results are consistent with those shown in Fig. 1B that, for the demonstration purpose, employed only 16 SARS-CoV-2 genomes, which are representatives of 16 countries in Table 5.

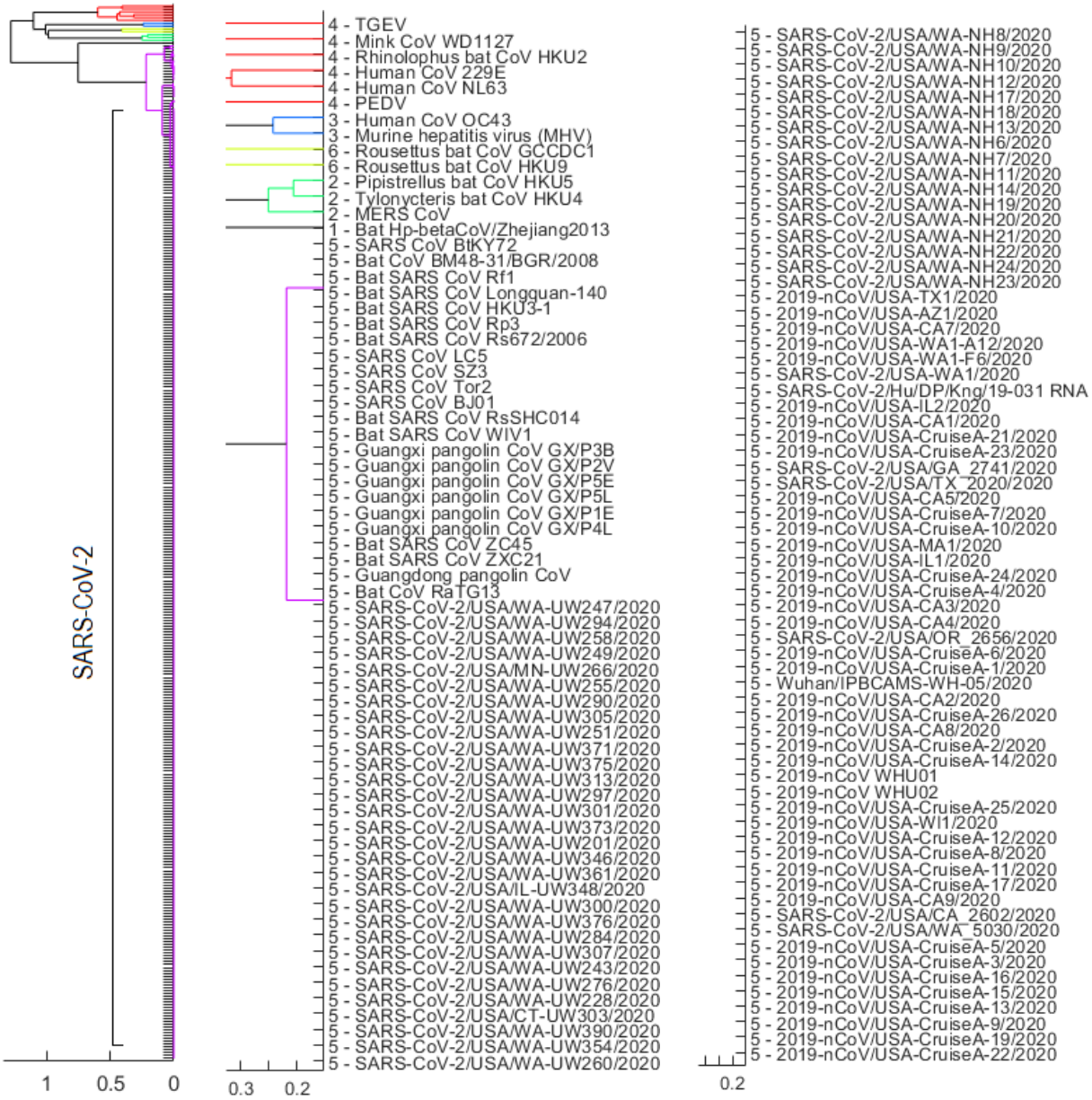

Fig. 10. Dendrogram plots using all 334 SARS-CoV-2 sequences obtained across 16 countries with the hierarchical clustering outcome when the cut-off parameter C is set equal to 0.7 (left). The middle figure shows in detail (zoom in) the top part of the dendrogram plot while the right figure shows the bottom part of the plot. All 334 SARS-CoV-2 sequences are grouped in cluster "5", which also includes SARS CoVs, bat SARS CoVs and pangolin CoVs. This cluster belongs to the *Sarbecovirus* sub-genus of the *Betacoronavirus* genus. Six viruses of the *Alphacoronavirus* genus (i.e. human CoV NL63, human CoV 229E, *Rhinolophus* bat CoV HKU2, PEDV, Mink CoV WD1127 and TGEV) are combined into cluster "4" while 3 viruses of the *Merbecovirus* sub-genus (i.e. *Pipistrellus* bat CoV HKU5, *Tylonycteris* bat CoV HKU4 and MERS CoV) are joined in cluster "2". These outcomes are in line with the results obtained in Fig. 7B, which used only 16 representative SARS-CoV-2 genomes.

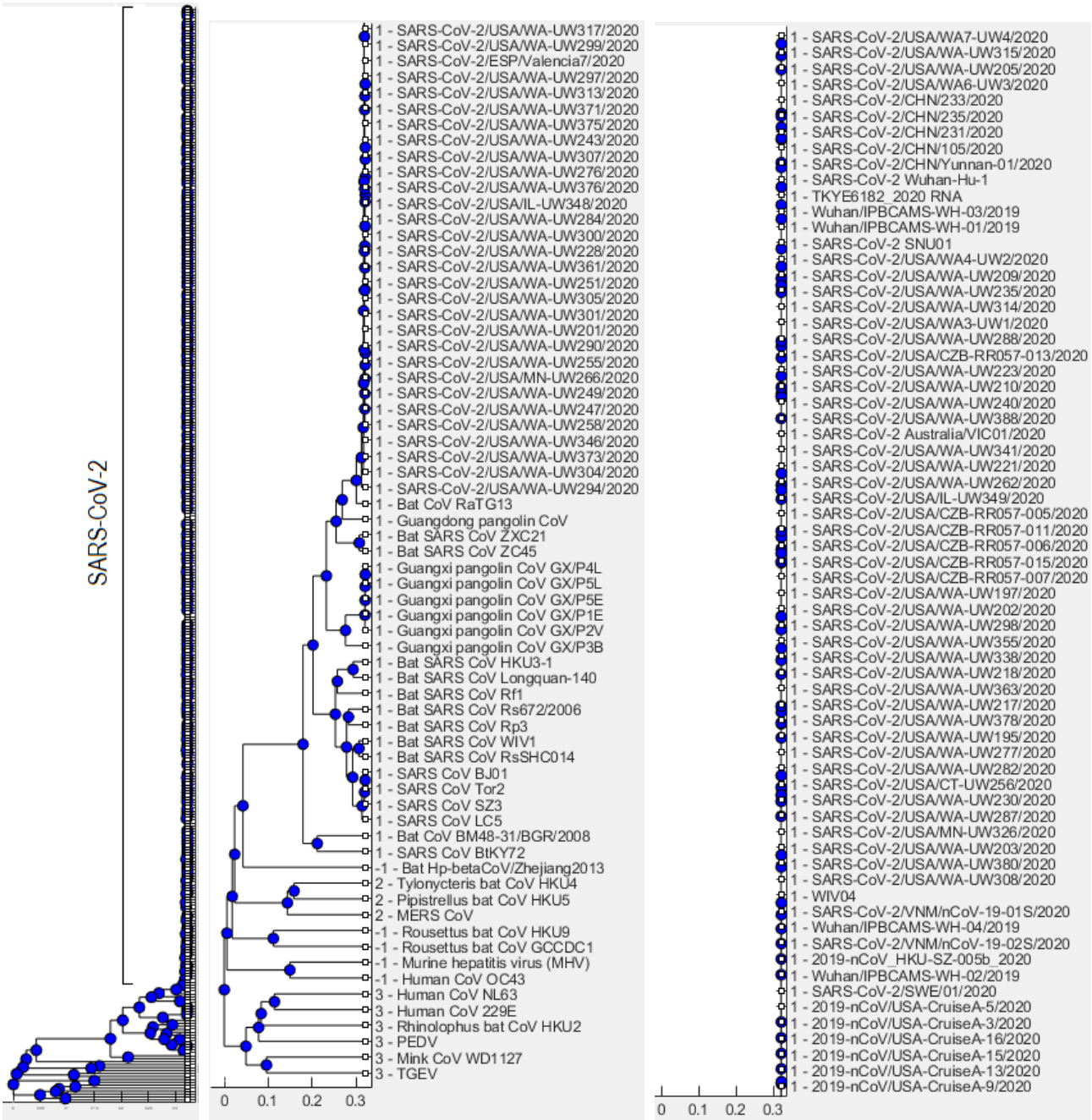

Fig. 11. A phylogenetic tree using all 334 SARS-CoV-2 sequences obtained across 16 countries with the DBSCAN clustering outcome when the search radius parameter ε is set equal to 0.55 and the minimum number of neighbours parameter equal to 3 (left). The middle figure shows in detail (zoom in) the bottom part of the tree while the right figure shows the top part of the tree. All 334 SARS-CoV-2 sequences are grouped in cluster "1", which also includes SARS CoVs, bat SARS CoVs and pangolin CoVs. This cluster belongs to the *Sarbecovirus* sub-genus of the *Betacoronavirus* genus. Six viruses of the *Alphacoronavirus* genus (i.e. human CoV NL63, human CoV 229E, *Rhinolophus* bat CoV HKU2, PEDV, Mink CoV WD1127 and TGEV) are combined into cluster "3" while 3 viruses of the *Merbecovirus* sub-genus (i.e. *Pipistrellus* bat CoV HKU5, *Tylonycteris* bat CoV HKU4 and MERS CoV) are joined in cluster "2". These results are similar to Fig. 8B, which used only 16 representative SARS-CoV-2 genomes for the demonstration purpose.

APPENDIX 3

This appendix presents results of two clustering methods, i.e. hierarchical clustering and DBSCAN, using the sequence distances computed by the maximum composite likelihood method [12], which was conducted in the MEGA X software [22]. These results are greatly similar to those obtained by using the Jukes-Cantor distance method shown throughout the paper. In these experiments, the clustering methods are applied to a dataset that combines reference sequences in Set 3 (Table 3) and 16 representative genomes of 16 countries in Table 5. When a country has more than one collected genome, the first released genome of that country is selected for this experiment. Fig. 12 demonstrates the distances estimated by the maximum composite likelihood method between each of the reference sequences and 16 representative SARS-CoV-2 genomes. The lines are almost parallel indicating that SARS-CoV-2 genome is not altered much across countries, which is in line with the results obtained using the Jukes-Cantor distance estimates in Fig. 5. The bat CoV RaTG13 is again shown much closer to SARS-CoV-2 than pangolin CoVs and other reference viruses although the distance range in Fig. 12 is larger than that in Fig. 5, i.e. [0, ∼1.6] versus [0, ∼0.7], respectively.

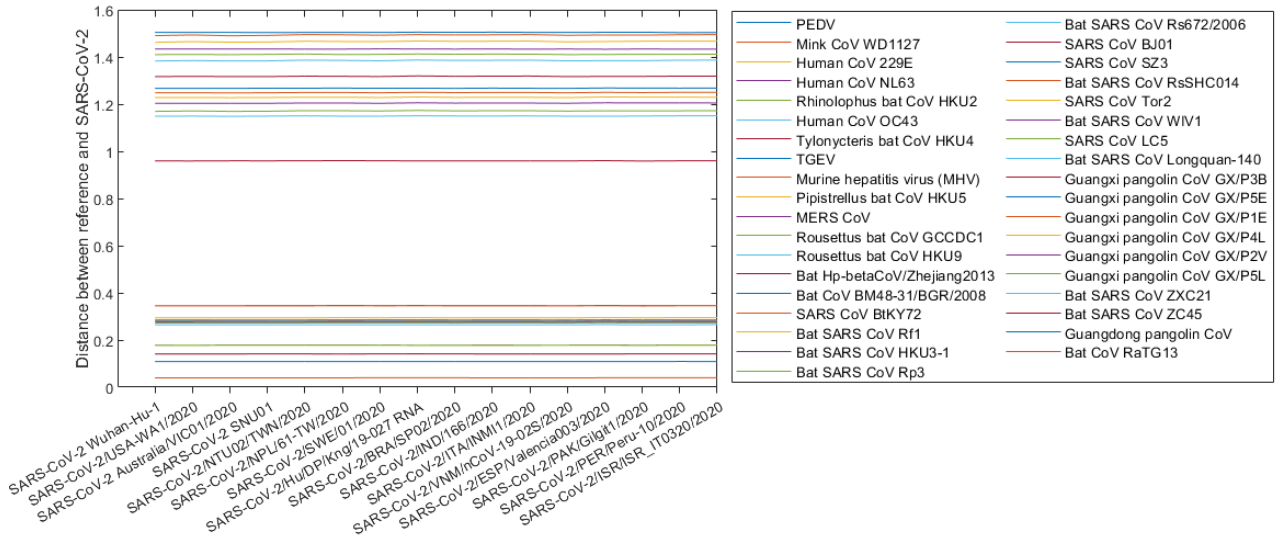

Fig. 12. Evolutionary distances between reference genomes and 16 representative SARS-CoV-2 genomes of 16 countries based on the maximum composite likelihood method. Each line is composed of 16 data points representing 16 pairwise distances. The bottom line, for instance, shows the distances between bat CoV RaTG13 and each of 16 representative genomes on the *x*-axis.

In Fig. 13A, when the hierarchical clustering cut-off parameter is set equal to 0.1, all 16 representative SARS-CoV-2 genomes are grouped into cluster "12", which also includes other viruses of the *Sarbecovirus* sub-genus of the *BetaCoV* genus. When moving from Fig. 13A to Fig. 13B, even though members of the *Sarbecovirus* cluster ("12" in Fig. 13A) are split into 2 clusters "1" and "2" in Fig. 13B, the SARS-CoV-2 sequences are still grouped into cluster "14" with other members of the *Sarbecovirus* sub-genus such as bat CoV RaTG13, Guangdong pangolin CoV, bat SARS CoV ZXC21 and bat SARS CoV ZC45. These results provide us with a confidence on confirming the *Sarbecovirus* sub-genus of the SARS-CoV-2. This is consistent with the result based on the Jukes-Cantor distances shown in Fig. 7. Fig. 13C shows that SARS-CoV-2 genomes are combined only with that of bat CoV RaTG13 when the cut-off parameter is decreased to 0.001. This again indicates that bat is the more likely origin of SARS-CoV-2 than pangolin. When we reduce the cut-off parameter to 0.0001, the SARS-CoV-2 sequences create their own cluster "22" and this questions the probable bat or pangolin origin of SARS-CoV-2. However, in Fig. 13B, we also find that the similarity between SARS-CoV-2 and bat CoV RaTG13 (or Guangdong pangolin CoV) is larger than the similarity between viruses having the same origin. For example, bat SARS CoV WIV1 and bat SARS CoV ZC45 have the same bat origin but they are divided into 2 clusters ("1" and "14") while all 16 SARS-CoV-2 representatives are grouped into a cluster ("14") with bat CoV RaTG13 and Guangdong pangolin CoV. This implies that SARS-CoV-2 may have originated from bats or pangolins.

Results of DBSCAN using distances calculated by the maximum composite likelihood method are presented in Fig. 14, which are also consistent with those obtained by the Jukes-Cantor distance method in Fig. 8.

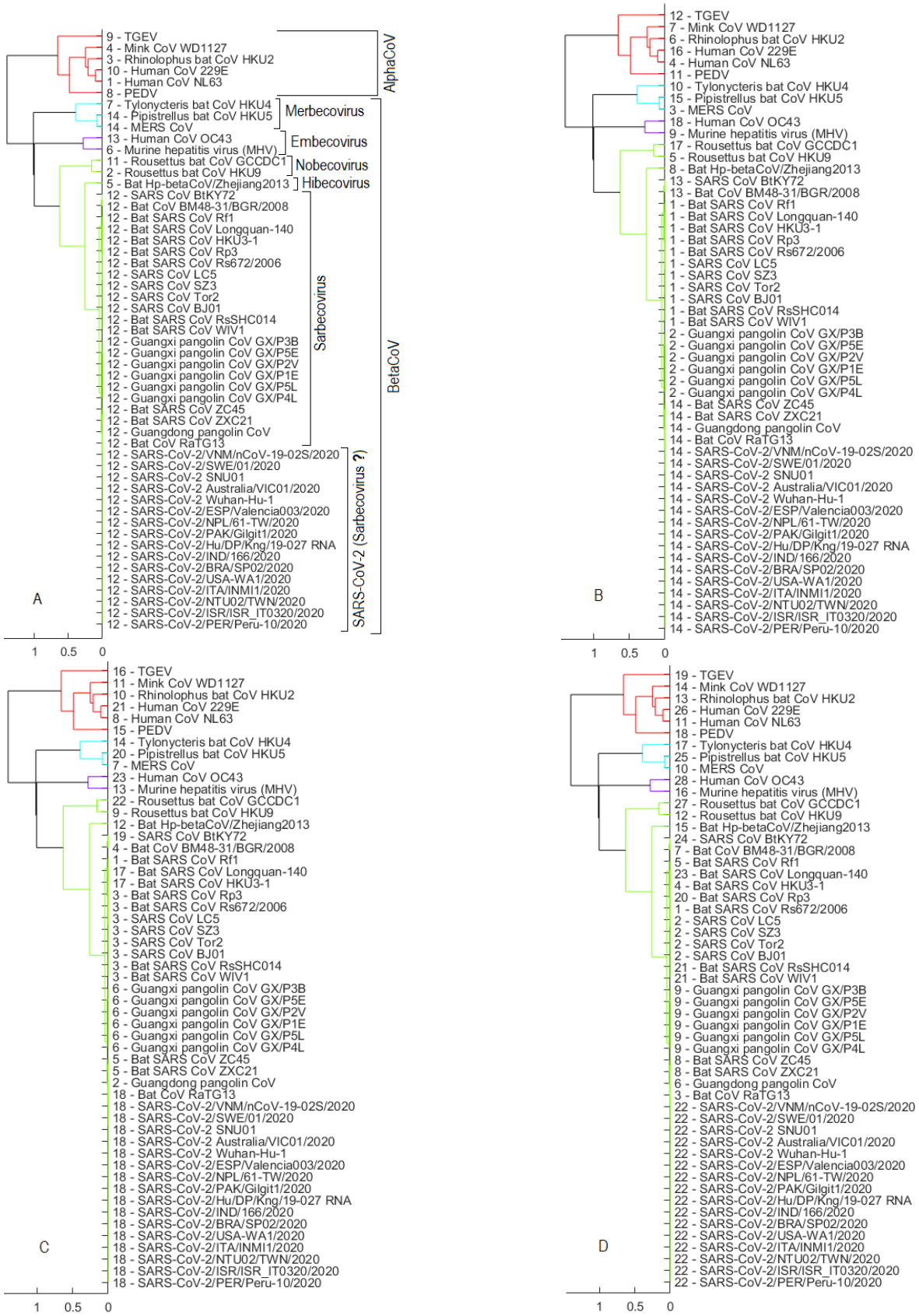

Fig. 13. Hierarchical clustering results using pairwise distances based on the maximum composite likelihood method, (A) the cut-off parameter is set to 0.1, (B) cut-off equal to 0.01, (C) cut-off is 0.001, and (D) cut-off is 0.0001.

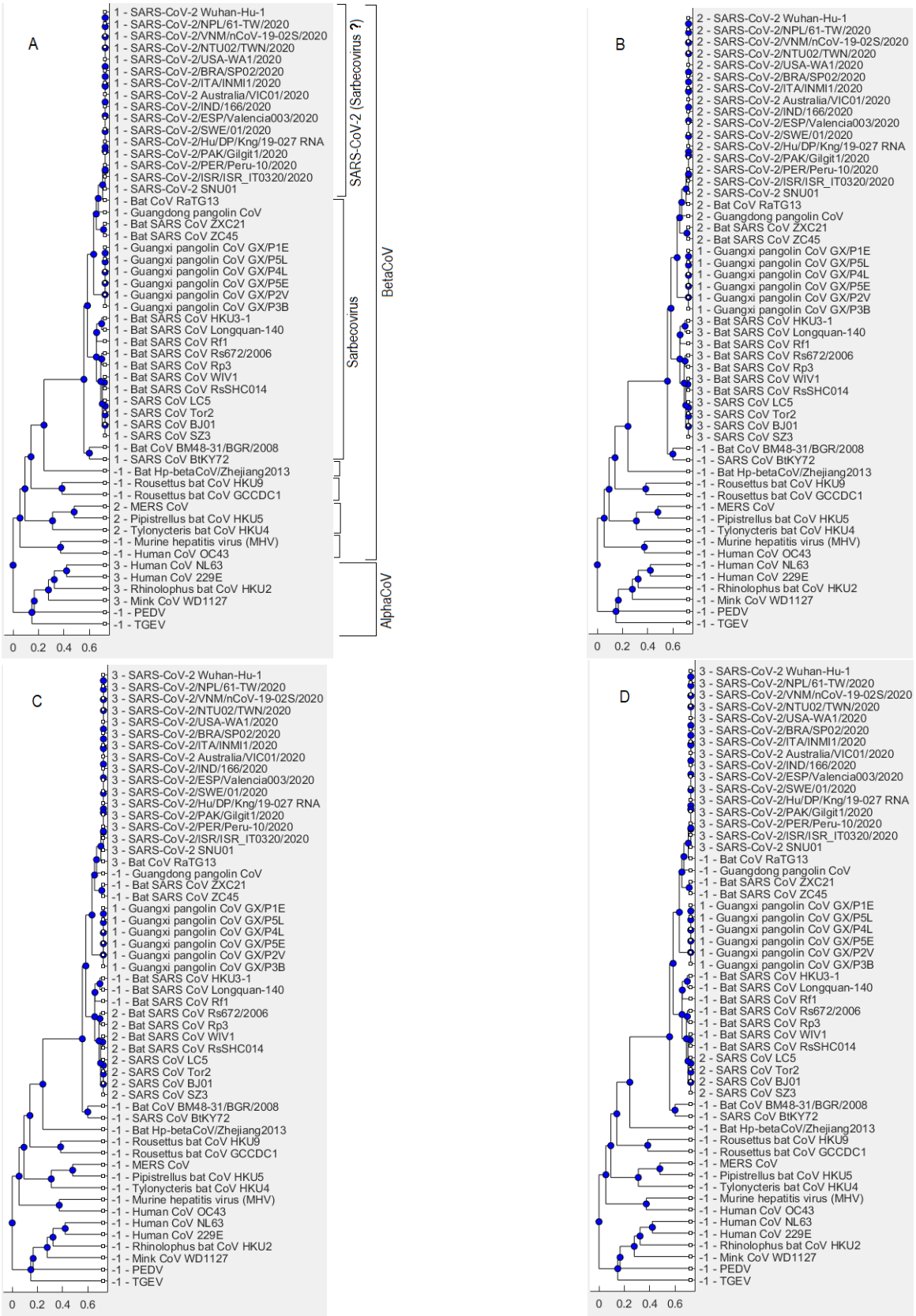

Fig. 14. Results of DBSCAN using the pairwise distances calculated by the maximum composite likelihood method, (A) the search radius parameter ε is set to 0.9, (B) ε is equal to 0.15, (C) $\varepsilon = 0.1$, and (D) $\varepsilon = 0.01$.