1 Using singleton densities to detect recent

2 selection in *Bos taurus*

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19 Abstract: Many quantitative traits are subject to polygenic selection, where 20 several genomic regions undergo small, simultaneous changes in allele 21 frequency that collectively alter a phenotype. The widespread availability of 22 genome data, along with novel statistical techniques, has made it easier to detect these changes. We apply one such method, the 'Singleton Density Score', 23 to the Holstein breed of *Bos taurus* to detect recent selection (arising up to 24 25 around 740 years ago). We identify several genes as candidates for targets of 26 recent selection, including some relating to cell regulation, catabolic processes, 27 neural-cell adhesion and immunity. We do not find strong evidence that three traits that are important to humans - milk protein content, milk fat content, and 28 29 stature – have been subject to directional selection. Simulations demonstrate 30 that since *B. taurus* recently experienced a population bottleneck, singletons are 31 depleted so the power of SDS methods are reduced. These results inform on 32 which genes underlie recent genetic change in *B. taurus*, while providing 33 information on how polygenic selection can be best investigated in future 34 studies.

36 **Impact statement:** Many traits of ecological or economic importance (including height, 37 disease propensity, climatic adaptation) are 'polygenic'. That is, they are affected by a 38 large number of genetic variants, with each one only making a small contribution to a 39 trait, but collectively influence variation. As selection acts on all of these variants 40 simultaneously, it only changes the frequency of each one by a small amount, making it hard to detect such selection from genome data. This situation has changed in recent 41 42 years, with the proliferation of whole-genome data from many individuals, along with 43 the development of methods to detect the subtle effects of polygenic selection. Here, 44 we use data from 102 genomes from domesticated cattle (Bos taurus) that has 45 experienced intense artificial selection since domestication, and test whether we can 46 detect signatures of recent selection (arising up to 740 years ago). Domesticated 47 species are appealing for this kind of study, as they are subject to extensive genome 48 sequencing studies, and genetic variants can be related to traits under selection. We 49 carried out our analysis in two parts. We first performed a genome-wide scan to find 50 individual genetic regions that show signatures of recent selection. We identify some 51 relating to cell regulation, catabolic processes, neural-cell adhesion and immunity. In 52 the second part, we then analysed genetic regions associated with three key traits: 53 milk protein content, milk fat content, and stature. We tested whether these regions 54 collectively showed a signature of selection, but did not find a significant result in either 55 case. Simulations suggest that the domestication history of cattle affected the power 56 of these methods. We end with a discussion on how to best detect polygenic selection 57 in future studies.

58 Introduction

59 Determining which genomic regions have been subject to selection is a major 60 research goal in evolutionary genetics. Traditional methods have focused on 61 detecting strong selection affecting individual genes (Nielsen, 2005; Vitti et al., 2013; Stephan, 2019). An alternative process is 'polygenic selection', where many loci 62 63 contribute to genetic variation in a trait, so selection acting on it is expected to 64 generate small and simultaneous allele frequency changes at multiple loci (Pritchard & Di Rienzo, 2010; Pritchard et al., 2010). Many polygenic models have been 65 66 formulated to account for both the response to phenotypic selection, and the 67 maintenance of genetic variance in guantitative traits [reviewed by Sella & Barton 68 (2019); Barghi et al. (2020)]. Among them is Fisher's infinitesimal model, which is 69 important for its historical role in uniting population and quantitative genetics, and its 70 recent renaissance in the context of genome-wide association studies (Fisher, 1918; 71 Barton & Keightley, 2002; Barton et al., 2017; Charlesworth & Edwards, 2018; 72 Visscher & Goddard, 2019). However, whereas it has been possible to identify which 73 genetic regions contribute to trait variation, it has historically been hard to infer which 74 alleles have been involved in the polygenic selection response. Extensive theoretical 75 studies of how alleles at multiple loci act when a population adapts to a new optimum 76 generally find that 'large-effect' alleles, which strongly affect a trait, are the first to spread and fix while 'small-effect' alleles take much longer to reach high frequencies 77 78 (de Vladar & Barton, 2014; Wollstein & Stephan, 2014; Jain & Stephan, 2015, 2017a, 79 2017b; Stetter et al., 2018; Thornton, 2019; Hayward & Sella, 2019). Furthermore, if epistasis exists between variants, many selected alleles do not reach fixation as they 80 81 eventually become deleterious (de Vladar & Barton, 2014; Jain & Stephan, 2017b).

82 The spread of large-effect alleles may also be impeded if a faster adaptive response 83 can be otherwise realised through changes at many small-effect alleles (Lande, 84 1983; Chevin & Hospital, 2008; Pavlidis et al., 2012; Chevin, 2019). Alternatively, if 85 the optimum shift is sufficiently big, then large-effect mutations that first go to fixation 86 can subsequently be replaced by small-effect variants over longer timescales (on the 87 order of the population size; Hayward and Sella (2019)). Overall, only a small 88 proportion of loci affected by polygenic selection are expected to fix sufficiently 89 quickly to leave selection signatures in genomic data (Pavlidis et al., 2012; Thornton, 90 2019).

91 Due to this difficulty, earlier methods for detecting polygenic selection focused 92 on cases where selection favours distinct phenotypes in different populations, so trait 93 differentiation amongst populations will be greater than expected under neutral drift. 94 Tests for this form of selection relied on comparing Q_{st} and F_{st} statistics, which 95 respectively measured mean genetic differentiation at the trait itself and a set of 96 neutral loci (Whitlock, 2008; Le Corre & Kremer, 2012; Savolainen et al., 2013). Yet 97 these methods do not determine which genomic regions are subject to selection. This 98 situation has now changed with the increased number of genome-wide association 99 study (GWAS) data that link genotypes and phenotypes, as exemplified by the 100 development of large cohort studies [e.g., the UK Biobank; Bycroft et al. (2018)]. The 101 release of these data spurred a series of studies and new methods designed 102 specifically to detect polygenic selection. These methods usually involve determining, 103 which SNPs affecting a phenotype show correlated changes in frequency (Berg & 104 Coop, 2014; Racimo et al., 2018; Sanjak et al., 2018; Josephs et al., 2019; Berg et 105 al., 2019a, 2019b; Uricchio et al., 2019; Edge & Coop, 2019; Kreiner et al., 2020;

106 Wieters et al., 2021; Gramlich et al., 2021); which sets of alleles are associated with 107 certain environmental or climatic variations (Coop et al., 2010; Turchin et al., 2012; 108 Robinson et al., 2015; Yeaman et al., 2016; Exposito-Alonso et al., 2018; Zan & 109 Carlborg, 2018; Exposito-Alonso et al., 2019; MacLachlan et al., 2021; Ehrlich et al., 110 2021; Fuhrmann et al., 2021; Rowan et al., 2021); or determining which SNPs or 111 genetic regions explain a large fraction of phenotypic variance and trait heritability 112 (Zhou et al., 2013; Yang et al., 2015; Gazal et al., 2017; Zeng et al., 2018; Schoech 113 et al., 2019; Exposito-Alonso et al., 2020; Duntsch et al., 2020; Zeng et al., 2021). 114 Some of these approaches use overlapping methods. 115 Detecting recent polygenic selection is much harder, as long periods of time 116 (number of generations on the order of the population size: Hayward and Sella, 2019: 117 Thornton, 2019) may be needed to cause detectable frequency changes in alleles 118 with small effect sizes. Over shorter timescales, these frequency changes are 119 expected to be more modest and harder to detect (Stephan, 2016; Jain & Stephan, 120 2017a). A recent breakthrough in detecting these subtle changes was the 121 development of the 'Singleton Density Score' (SDS), a statistic tailored to detect 122 recent and coordinated allele frequency changes over many SNPs (Field et al., 123 2016). Recent selection at a locus favouring one variant will lead to a reduction in the 124 number of singletons (i.e., variants that are only observed once) around it. The SDS 125 detects regions that exhibit a reduction in the density of singletons, to determine 126 candidate regions that have been subject to recent selection. Using this approach, 127 Field et al. (2016) found correlations between SDS scores at SNPs and their 128 associated GWAS effect sizes for several polygenic traits in the modern UK human 129 population, including increased height, infant head circumference and fasting insulin.

Their findings suggested that these traits have been subject to recent selection
during the most recent 75 or so generations (about 2,000 years). However, these
(and other) results that detect selection for increased height may instead reflect
previously unaccounted–for population structure (Novembre and Barton, 2018;
Barton et al., 2019; Sohail et al., 2019; Berg et al., 2019; Uricchio et al., 2019; Edge
and Coop, 2019).

136 The SDS method is ideally suited to organisms where large amount of whole-137 genome data are available, along with QTL or GWAS information that link genotypes 138 to phenotypes, Domesticated species are attractive systems for studying recent 139 selection, as selected phenotypes are often already known and these species are 140 subject to large-scale sequencing studies. Investigating the genetic architecture 141 underlying rapid selection in these species is also important to determine how they 142 respond to agricultural practices, and uncover selection targets that can be used to 143 improve breeding programs (Georges et al., 2018). Domestic cattle Bos taurus has 144 been subject to intensive genomics analyses to improve artificial selection for traits 145 that are important for human use, including milk protein yield, milk fat content, and 146 stature (Hayes et al., 2009; Meuwissen et al., 2013; Wray et al., 2019). These traits 147 are influenced in part by an individual's genome, with significant heritability estimates 148 being recorded, some as high as 80% (Soyeurt et al., 2007; Haile-Mariam et al., 149 2013; Buitenhuis et al., 2016). Previous selection scans on B. taurus reported 150 individual regions that were likely to be subject to recent selection, some of which were close to genetic regions for stature and milk protein content (Lemay et al., 2009; 151 152 MacEachern et al., 2009; Qanbari et al., 2010; Boitard & Rocha, 2013; Qanbari et al., 153 2014; Zhao et al., 2015; Boitard et al., 2016a; Bouwman et al., 2018). However,

154	stature and milk protein content are polygenic traits, with several genetic regions and
155	QTLs associated with each (Lemay et al., 2009; Boitard et al., 2016a; Bouwman et
156	al., 2018; van den Berg et al., 2020). While recent methods have been developed to
157	detect polygenic environmental adaptation (Rowan et al., 2021), there has yet to be a
158	formal test of whether these intrinsic traits show evidence of polygenic selection.
159	Here, we applied the SDS method to whole-autosome sequencing data from
160	102 B. taurus Holstein individuals. We first determined genetic regions that have
161	been subject to recent directional selection, and subsequently tested whether
162	evidence exists for recent selection acting on a set of QTLs underlying either milk
163	protein content, milk fat content, or stature in this breed.
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164

165 *Results*

166 Methods outline

167 We filtered the data to retain only bi-allelic SNPs that had a sensible level of 168 coverage and did not lie in putatively over-assembled regions (i.e., duplicated 169 sections that caused many reads to assemble at a specific genetic location). Over-170 assembled regions appear as highly heterozygous with elevated coverage, and can 171 exhibit false signatures of recent selection. We also obtained a set of singletons and 172 filtered them to retain high-quality variants where both alleles were equally well 173 covered to remove potentially erroneous calls. We polarised test SNPs using 174 outgroup sequences and applied the SDS test of Field et al. (2016) to detect recent 175 selection, with increased SDS values reflecting selection favouring derived SNPs 176 over ancestral variants. We standardised SDS scores with those of a similar 177 frequency, so they are normally distributed [similar normalisation was also carried out

by Field et al. (2016)]. These values are denoted sSDS for 'standardised SDS'.

179 Further details are available in the *Methods* in the Supplementary Text.

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181 Estimating timescale of selection

182 We first determined the timescale over which we expect to detect selection in 183 *B. taurus* using the SDS method. SDS measures the changes in singleton numbers 184 around putatively selected SNPs, relative to background numbers in the absence of 185 selection. As singletons arise on the tips of the underlying gene trees, the average tip 186 length in the genealogy of sequenced samples determines the timescale over which 187 the SDS detects a signal (Field et al., 2016). As more haploid genomes are included 188 in the study, the time to first coalescence between two samples decreases, reducing 189 the tip lengths and therefore shortening the timescale over which SDS detects 190 selection (Field *et al.*, 2016). We hence simulate tip-ages over a range of sample 191 sizes to investigate how this timescale changes accordingly.

192 To calculate the mean tip age, we simulated gene genealogies under two 193 scenarios. We first simulated the Holstein population demography inferred by Boitard 194 et al. (2016b), which suggested that this population experienced a sudden decline in 195 effective population size (N_e) since domestication, but with a present-day N_e (~793) 196 that is much larger than that inferred from pedigree data [~49; Sørensen et al. 197 (2005)] or from temporal variation in SNP frequencies (~48; Jiménez–Mena et al. 198 2016). Hence, we also simulated genealogies under a second model that used the 199 Boitard et al. (2016b) demographic model, but with the present-day N_e set to 49.

200 These scenarios will be referred to as the 'High N_0 ' and 'Low N_0 ' models,

201 respectively.

202 Figure 1 shows simulation results. Depending on the assumed present-day 203 N_e , the tip length in our sample of 204 alleles (i.e., assuming two per diploid 204 individual) goes back either 65 or 148 generations. Assuming 5 years per generation 205 (Boitard *et al.*, 2016b), this timescale corresponds to between 325 and 740 years 206 ago. Since *B. taurus* domestication started around 10,000 years ago (Zeder, 2008) 207 the sample size used in this study will only capture selection acting in the very recent 208 past that is more relevant for breed formation, rather than selection during *B. taurus* 209 domestication. Sample sizes and tip-ages are linearly related on a log-log scale, 210 meaning that an increase in sample size will greatly decrease the timescale over 211 which SDS detects selection. For example, with 500 haplotypes then SDS will detect 212 selection acting no more than 50 generations ago, depending on the underlying 213 demographic model.

We will focus on detecting selection signatures assuming the high N_0 model. Results using the low N_0 model to calibrate scores were broadly similar. They are outlined in the Supplementary Text; we will highlight when differences arise.

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218 *Genome–wide sSDS*

Figure 2 plots sSDS values (at SNPs with minor allele frequency greater than 5%) across all autosomes, excluding chromosome 25 (due to an insufficient number of singletons needed to obtain SDS scores after filtering). Many SNPs have elevated sSDS scores (158 SNPs at *FDR* < 0.05; 306 for the low *N*₀ model). Several regions contain SNPs with significantly high sSDS values (Bonferroni–corrected nominal *P* < 0.05; actual *P* < ~2.7 x 10⁻⁸). To further investigate potential selection targets, we looked for genes that either overlapped significant SNPs or lay 10kb up– or

226 downstream of them. Linkage disequilibrium (LD), as measured by r^2 , decays to 227 around 0.2 over 50kb in Danish Holstein breeds (Buitenhuis et al., 2016), so genes 228 within 10kb should be in LD with regions harbouring high sSDS scores. Table 1 lists 229 these genes, with more targets present under the low N_0 model. Most of these genes 230 are of unknown function (as listed on UniProt); the list also includes an snRNA. 231 PPM1L is involved with cellular regulation and the activation of stress-activated 232 protein kinases. TDO2 is involved in tryptophan-related catabolic processes, while 233 *NTM* is implicated in neural cell adhesion. SNPs with significantly elevated scores 234 are also found on chromosome 23 near the MHC region, which may reflect over-235 dominant selection. All Bonferroni-significant SNPs were removed from subsequent 236 tests of recent polygenic selection to prevent directional selection from skewing the 237 underlying sSDS distributions. Figure S1 shows results for the low No model.

238

239 Testing for polygenic selection acting on milk protein and stature

240 If polygenic selection were acting on specific traits, we expect a positive 241 correlation between the effect size of variant underpinning it, and selection acting on 242 it as measured by sSDS. We collated sSDS scores of SNPs that lie close to QTLs 243 reported for either milk fat percentage, milk protein percentage (van den Berg et al., 244 2020), or those that lie close to stature QTLs (Bouwman et al., 2018). The latter were 245 inferred from a meta-analysis of GWAS studies conducted in seven Holstein 246 populations, but not every QTL had an effect size reported in each population. We 247 hence investigated two overlapping consensus QTL sets, where an effect size was either reported in at least 6 of 7 populations (yielding 42 QTLs with sSDS scores 248 249 associated with them), or where effect sizes were reported in at least 5 of 7

populations (58 QTLs had sSDS scores). We re-polarised SDS scores so that a
positive score reflected a trait-increasing effect; we denote these values 'tSDS'
following Field *et al.* (2016). We then determine if there was a positive correlation
between the absolute log₁₀-value of the QTL *P*-value (a proxy for the effect size) and
tSDS.

255 Figure 3 shows the relationship between QTL *P*-values and tSDS for SNPs 256 that lie close to QTLs. Although positive trends are observed, they all exhibit non-257 significant correlations (milk fat percentage Spearman $\rho = 0.0990$, P = 0.603; milk 258 protein percentage Spearman $\rho = 0.0354$, P = 0.758; stature from 6 breeds 259 Spearman ρ = -0.0739, P = 0.642; stature from 5 breeds Spearman ρ = -0.00966, P 260 = 0.943). Relationships remain non-significant after removing an outlier point for the 261 milk traits whose QTL has an extremely low *P*-value (Figure S2), and also under the 262 low N_0 model (Figure S3; see figure legends for regression *P*-values).

263 sSDS (and tSDS) can become correlated along the genome if focal SNPs are 264 in LD with one another, which was not accounted for in the preceding analyses. To 265 determine whether LD could have affected these correlations, we randomly 266 subsampled sSDS scores from SNPs that shared the same chromosome and bin of 267 derived-allele frequency as the SNPs used in the above analyses, and re-polarised 268 them to transform them into tSDS values. We then determined the Spearman's ρ 269 associated with these permuted values to determine whether that for the true data 270 was significantly elevated (see Methods for details). In all cases, the observed value 271 was not significantly higher than for permuted values (see Figure S4 for histograms 272 and exact *P*-values, which all exceed 0.05). We therefore conclude that these QTL 273 datasets do not harbour SNPs with significantly different tSDS scores compared to

the rest of the genome.

275

276 *Discussion*

277 Summary of results

278 We analysed an extensive *B. taurus* genomic dataset to identify signatures of 279 recent selection in the Holstein breed, and to determine whether the data contained a 280 signal of polygenic selection acting on milk proteins and QTLs underlying phenotypic 281 variation in stature. Given the sample size and the demographic history of Holsteins, 282 the SDS method can detect very recent selection events arising no more than 283 approximately 740 years ago (Figure 1). A whole-genome scan for sSDS scores 284 identified several targets of recent directional selection that overlap or lie close to 285 protein-coding genes (Figure 2; Table 1). The genes whose functions are known are 286 involved in protein regulation, catabolic processes, and neural-cell adhesion. 287 Significant values were also observed near the MHC region. We subsequently 288 investigated whether either milk protein genes or SNPs near stature QTLs 289 collectively showed evidence of polygenic selection. We did so by testing whether 290 there is a relationship between the QTL effect size, as measured by its *P*-value, and 291 tSDS values to SNPs near them. However, no relationship was observed, even after 292 performing a permutation test (Figures 3, S2-S4). Hence, while sSDS could reveal 293 specific instances of recent selection, tests based on collective scores of variants 294 associated with known selected traits yielded no signal of polygenic selection.

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296 Potential reasons for a lack of polygenic selection signal

297 Impact of Holstein demographic history

298 While the SDS method detected individual candidate genes for very recent 299 selection, we were unable to find strong evidence for polygenic selection acting on 300 three traits that were subject to artificial selection since domestication. This result is a 301 priori surprising, given that these traits have been subject to recent intense artificial 302 selection. Recent studies generally find non-zero heritability estimates for them, 303 indicating that there should be the potential for genetic variants underpinning them to 304 change in response to artificial selection (Soveurt *et al.*, 2007; Haile-Mariam *et al.*, 305 2013; Buitenhuis et al., 2016). In addition, the ratio of the mutation and recombination 306 rates in cattle is just over three (Boitard et al., 2016b; Harland et al., 2018), indicating 307 that several informative SNPs exist per haplotypes that should improve the power of 308 the SDS method [in contrast, this ratio is approximately equal to one in humans 309 (Field et al., 2016)].

310 One potential reason for this lack of signal is due to the population history of 311 Bos taurus. The effective population size of many *B. taurus* breeds appears to have 312 undergone a decline since domestication (Sørensen et al., 2005; Boitard et al., 313 2016b), which likely reflects successive bottlenecks due to domestication, breed 314 formation and intense recent selection. Population size reductions are known to 315 reduce the number of low-frequency variants and increases the prevalence of 316 intermediate-frequency variants (Harpending *et al.*, 1998), which can affect the power 317 of the SDS method. To understand if the history of *B. taurus* affects the detection of 318 recent selection in Holstein cattle using SDS, we ran coalescent simulations to 319 determine its ability to detect ongoing selection, given realistic Holstein population 320 history and genetic parameters (see Methods for details). We simulated a partial

321 sweep occurring in the middle of a 10Mb region, either assuming a mutation rate in 322 line with what has been inferred for Holstein, or one 10-fold higher to replicate 323 diversity expected in a genetic region with an elevated mutation rate. 324 For the standard mutation rate, no SDS scores were produced for any 325 simulations. After inspecting the simulation results, we see that there is a large skew 326 in the distribution of singleton numbers per individual with a large number of 327 individuals (over 20 on average) that do not carry singletons at the end of 328 simulations, preventing the calculation of a local SDS score (Figure 4). This fraction 329 remained the same irrespective of whether the simulated region was neutral or 330 subject to selection; the main effect of a sweep was to reduce the mean number of 331 singletons per individual, which is the signal measured by SDS (Field et al., 2016). 332 This reduction in overall singleton numbers is consistent with the known effects of 333 population size contraction on reducing tip lengths (Harpending et al., 1998). 334 With a 10-fold higher mutation rate, there were fewer cases where no 335 individual harboured singletons (Figure 4). Accordingly, SDS scores could be 336 calculated for 65 and 66 out of 100 simulations for the neutral and selective cases 337 respectively. In these cases, sSDS values were significantly higher in the selected 338 case than for the neutral case (Figure 5; two-sided Wilcox Test $P = 1.1 \times 10^{-5}$). 339 However, note that sSDS values is less than one for the selected case, which does 340 not exceed the FDR threshold in our study (for the high N_0 case, the smallest sSDS 341 value with FDR < 0.05 is 4.46).

Although singleton numbers differ between the two cases, a reduction in power could also be caused by a more general reduction in diversity due to the small recent effective population sizes of cattle. To investigate this effect, we estimated the

345 fixed Ne that would yield the same number of segregating sites in simulations using 346 the standard mutation rate, based on Watterson's estimator (Watterson, 1975; 347 Hudson, 1990; see Methods for details). In both cases where selection is present or 348 absent, Ne estimates lie at around 25,000, which is that inferred at approximately 349 halfway between the onset of domestication and the present day. (Boitard et al., 350 2016b; Figure S5). Given that estimates are similar irrespective of whether a sweep 351 was present or not, the reduced population size caused by domestication could have 352 also affected power due to limiting genetic variation and thus the potential to detect 353 subtle sweep signatures associated with polygenic selection. 354 Overall, these simulations are consistent with population size reductions in B. 355 taurus both reducing the overall genetic diversity and the number of singletons, which 356 limits its ability to detect partial sweeps. SDS is more likely to detect signals in 357 regions of elevated mutation rate, suggesting there will likely be an ascertainment 358 bias in where signals are detected in the genome. The reduction in singletons also 359 reduces the power to investigate SDS values in telomeric regions. SDS values are 360 calculated using the distance up- and downstream from a SNP to the nearest 361 singleton, and are undefined if a certain number of samples do not harbour 362 singletons in either direction (Field et al., 2016). SDS values are hence less likely to 363 be defined in telomeric regions, as it is generally less feasible to observe singletons 364 up until the end of the chromosome. This problem is exacerbated if there are few

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367 Other potential reasons for a lack of signal

singletons overall.

368 Another potential reason for a lack of signal is that the selection response on

369 these traits may have been driven by large-effect variants that have already fixed in 370 the population, with a smaller contribution from small-effect mutations. Theoretical 371 models have shown that more major-effect QTLs are likely to fix if the population lies 372 further from a fitness optimum (Lande, 1983; Jain & Stephan, 2017b; Thornton, 373 2019). Domesticated species, which experience strong and sustained directional 374 artificial selection, especially in recent generations, could thereby fix more adaptive 375 mutation via sweep-like processes compared to populations evolving in more stable 376 environments (Lande, 1983; Jain & Stephan, 2017a). Furthermore, once a population 377 has adapted to a new environment (the domestication phenotype in this case), then 378 any remaining major-effect mutations are likely to be superseded by variants with 379 weaker effects, which are harder to detect (Hayward & Sella, 2019). The response to 380 polygenic selection will be further weakened in smaller populations (John & Stephan, 381 2020), which could be a factor given the reduced effective population sizes of B. 382 taurus (Sørensen et al., 2005; Boitard et al., 2016b). There is some evidence of this 383 explanation; selective sweeps signatures are associated with stature QTLs 384 (Bouwman et al., 2018), and the study of van den Berg et al. (2020) was more likely 385 to identify milk QTLs that had a moderate to high minor allele frequency, suggesting 386 reduced power to detect low-frequency variants that are potential contributors to 387 polygenic selection. Conversely, the stature meta-analysis by Bouwman et al. (2018) 388 found significant SNPs that explained up to 13.8% of the variance in stature, which is 389 similar to that explained by significant SNPs for human height (16%), which is a 390 classic trait for polygenic selection studies. Hence, there may be sufficient polygenic 391 SNPs present to test for polygenic selection, but the power will still be reduced due to 392 the demographic history of Holstein cattle.

393 Potential solutions to increase power include increasing sample sizes; using 394 alternative methods; or analysing different kinds of genome data to detect polygenic 395 selection. Applying SDS to a larger sample size would increase the power to detect 396 selection acting in the recent past [Figure 1; see also Field et al. (2016)], but overall 397 power will still be limited by the tip-length of neutral genealogies. Recent 398 developments in methodology involve directly inferring trees from genome data, and 399 using these to identify subtle sweep signatures associated with trait variants (Edge & 400 Coop, 2019; Speidel et al., 2019; Stern et al., 2021). These methods have greater 401 power to detect weakly-selected mutations that may be segregating for longer than 402 the tip-length of the population.

403 Another approach would be to look beyond sequence data and focus on gene 404 networks [reviewed by Fagny & Austerlitz (2021)]. The recently-proposed 'omnigenic' 405 model (Boyle et al., 2017; Liu et al., 2019) posits that variation in guantitative traits is 406 principally affected by a plethora of 'peripheral' genes that indirectly affect them, 407 rather than a limited set of 'core' genes that directly modify a trait. These numerous 408 peripheral genes may exert their influence via regulatory effects (e.g., gene 409 expression changes), but are also expected to be highly pleiotropic. Fully testing the 410 omnigenic model will require larger datasets and novel experimental designs (Wray 411 et al., 2018). A recent example is from an experiment with Drosophila melanogaster. 412 where gene knockouts that do not pass a GWAS significance threshold for pupal 413 length still significantly affect it (Zhang et al., 2021). There is also nascent evidence 414 that gene regulation may underlie directional polygenic selection. Boitard et al. 415 (2016a) found that some adaptive signatures of *B. taurus* are located in intergenic 416 regions; regulatory changes were also proposed to guide polygenic selection in

425	Materials and Methods
424	
423	impact of polygenic selection.
422	and a broader range of gene-sets could be a promising approach to determine the
421	selection (Castellano et al., 2019). Further investigations using regulatory information
420	be exceptional cases, as they are more likely to contain genes subject to very strong
419	primates (Daub et al., 2013, 2017; Svardal et al., 2017). Immunity gene-sets might
418	responses or immunity also found evidence for polygenic selection in humans and
417	Arabidopsis (He et al., 2016). Analyses of gene-sets associated with infection

- 426 Full methods are available in the Supplementary Text.
- 427

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437

Author contributions. All authors contributed to the study design. NAP and
BG provided data. MH performed the analyses and wrote the manuscript, with
feedback from NAP, BG and TB.

- 442 *Data archiving.* Raw SDS scores and polarisation information has been
- 443 deposited on Dryad (https://doi.org/10.5061/dryad.547d7wm8q). Data analysis and
- 444 simulation scripts are available on GitHub
- 445 (https://github.com/MattHartfield/CattleSDS).

446 *References*

- 447 Barghi, N., Hermisson, J. & Schlötterer, C. (2020) Polygenic adaptation: a unifying
- 448 framework to understand positive selection. *Nat. Rev. Genet.*, **21**, 769–781.
- 449 Barton, N.H., Etheridge, A.M. & Véber, A. (2017) The infinitesimal model: Definition,
- 450 derivation, and implications. *Theor. Popul. Biol.*, **118**, 50–73.
- Barton, N.H., Hermisson, J. & Nordborg, M. (2019) Why structure matters. *eLife*, 8,
 e45380.
- 453 Barton, N.H. & Keightley, P.D. (2002) Understanding quantitative genetic variation.
- 454 Nat. Rev. Genet., **3**, 11–21.
- 455 Berg, I. van den, Xiang, R., Jenko, J., Pausch, H., Boussaha, M., Schrooten, C., et
- 456 *al.* (2020) Meta-analysis for milk fat and protein percentage using imputed sequence
- 457 variant genotypes in 94,321 cattle from eight cattle breeds. *Genet. Sel. Evol.*, **52**, 37.
- 458 Berg, J.J. & Coop, G. (2014) A Population Genetic Signal of Polygenic Adaptation.
- 459 *PLoS Genet.*, **10**, e1004412.
- 460 Berg, J.J., Harpak, A., Sinnott-Armstrong, N., Joergensen, A.M., Mostafavi, H., Field,
- 461 Y., *et al.* (2019a) Reduced signal for polygenic adaptation of height in UK Biobank.
- 462 *eLife*, **8**, e39725.
- Berg, J.J., Zhang, X. & Coop, G. (2019b) Polygenic Adaptation has Impacted Multiple
 Anthropometric Traits. *bioRxiv*, 167551.
- 465 Boitard, S., Boussaha, M., Capitan, A., Rocha, D. & Servin, B. (2016a) Uncovering
- 466 Adaptation from Sequence Data: Lessons from Genome Resequencing of Four
- 467 Cattle Breeds. *Genetics*, **203**, 433–450.
- 468 Boitard, S. & Rocha, D. (2013) Detection of signatures of selective sweeps in the
- Blonde d'Aquitaine cattle breed. *Anim. Genet.*, **44**, 579–583.

- 470 Boitard, S., Rodríguez, W., Jay, F., Mona, S. & Austerlitz, F. (2016b) Inferring
- 471 Population Size History from Large Samples of Genome-Wide Molecular Data An
- 472 Approximate Bayesian Computation Approach. *PLoS Genet.*, **12**, e1005877.
- 473 Bouwman, A.C., Daetwyler, H.D., Chamberlain, A.J., Ponce, C.H., Sargolzaei, M.,
- 474 Schenkel, F.S., et al. (2018) Meta-analysis of genome-wide association studies for
- 475 cattle stature identifies common genes that regulate body size in mammals. *Nat.*
- 476 *Genet.*, **50**, 362–367.
- 477 Boyle, E.A., Li, Y.I. & Pritchard, J.K. (2017) An Expanded View of Complex Traits:
- 478 From Polygenic to Omnigenic. *Cell*, **169**, 1177–1186.
- 479 Buitenhuis, B., Poulsen, N.A., Gebreyesus, G. & Larsen, L.B. (2016) Estimation of
- 480 genetic parameters and detection of chromosomal regions affecting the major milk
- 481 proteins and their post translational modifications in Danish Holstein and Danish
- 482 Jersey cattle. *BMC Genet.*, **17**, 114.
- 483 Bycroft, C., Freeman, C., Petkova, D., Band, G., Elliott, L.T., Sharp, K., et al. (2018)
- 484 The UK Biobank resource with deep phenotyping and genomic data. *Nature*, **562**,
- 485 203–209.
- 486 Castellano, D., Uricchio, L.H., Munch, K. & Enard, D. (2019) Viruses rule over
- 487 adaptation in conserved human proteins. *bioRxiv*, 555060.
- 488 Charlesworth, B. & Edwards, A.W.F. (2018) A century of variance. *Significance*, **15**,
 489 20–25.
- 490 Chevin, L.-M. (2019) Selective Sweep at a QTL in a Randomly Fluctuating
- 491 Environment. *Genetics*, **213**, 987–1005.
- 492 Chevin, L.-M. & Hospital, F. (2008) Selective Sweep at a Quantitative Trait Locus in
- the Presence of Background Genetic Variation. *Genetics*, **180**, 1645–1660.

- 494 Coop, G., Witonsky, D., Di Rienzo, A. & Pritchard, J.K. (2010) Using Environmental
- 495 Correlations to Identify Loci Underlying Local Adaptation. *Genetics*, **185**, 1411–1423.
- 496 Daub, J.T., Hofer, T., Cutivet, E., Dupanloup, I., Quintana-Murci, L., Robinson-
- 497 Rechavi, M., et al. (2013) Evidence for Polygenic Adaptation to Pathogens in the
- 498 Human Genome. *Mol. Biol. Evol.*, **30**, 1544–1558.
- 499 Daub, J.T., Moretti, S., Davydov, I.I., Excoffier, L. & Robinson-Rechavi, M. (2017)
- 500 Detection of Pathways Affected by Positive Selection in Primate Lineages Ancestral
- 501 to Humans. *Mol. Biol. Evol.*, **34**, 1391–1402.
- 502 Duntsch, L., Tomotani, B.M., Villemereuil, P. de, Brekke, P., Lee, K.D., Ewen, J.G., et
- 503 al. (2020) Polygenic basis for adaptive morphological variation in a threatened
- Aotearoa I New Zealand bird, the hihi (*Notiomystis cincta*). *Proc. R. Soc. B.*, 287,
 20200948.
- 506 Edge, M.D. & Coop, G. (2019) Reconstructing the History of Polygenic Scores Using
- 507 Coalescent Trees. *Genetics*, **211**, 235–262.
- 508 Ehrlich, M.A., Wagner, D.N., Oleksiak, M.F. & Crawford, D.L. (2021) Polygenic
- 509 Selection within a Single Generation Leads to Subtle Divergence among Ecological
- 510 Niches. *Genome Biol. Evol.*, **13**, evaa257.
- 511 Exposito-Alonso, M., 500 Genomes Field Experiment Team, Burbano, H.A.,
- 512 Bossdorf, O., Nielsen, R. & Weigel, D. (2019) Natural selection on the Arabidopsis
- 513 *thaliana* genome in present and future climates. *Nature*, **573**, 126–129.
- 514 Exposito-Alonso, M., Vasseur, F., Ding, W., Wang, G., Burbano, H.A. & Weigel, D.
- 515 (2018) Genomic basis and evolutionary potential for extreme drought adaptation in
- 516 Arabidopsis thaliana. Nat. Ecol. Evol., **2**, 352–358.
- 517 Exposito-Alonso, M., Wilton, P. & Nielsen, R. (2020) Non-additive polygenic models

- 518 improve predictions of fitness traits in three eukaryote model species. *bioRxiv*,
- 519 2020.07.14.194407.
- 520 Fagny, M. & Austerlitz, F. (2021) Polygenic Adaptation: Integrating Population
- 521 Genetics and Gene Regulatory Networks. *Trends Genet.*, **37**, 631–638.
- 522 Field, Y., Boyle, E.A., Telis, N., Gao, Z., Gaulton, K.J., Golan, D., et al. (2016)
- 523 Detection of human adaptation during the past 2000 years. *Science*, **354**, 760–764.
- 524 Fisher, R.A. (1918) The correlation between relatives on the supposition of
- 525 Mendelian inheritance. *Trans. R. Soc. Edinb.*, **52**, 399–433.
- 526 Fuhrmann, N., Prakash, C. & Kaiser, T.S. (2021) Polygenic adaptation from standing
- 527 genetic variation allows rapid ecotype formation. *bioRxiv*, 2021.04.16.440113.
- 528 Gazal, S., Finucane, H.K., Furlotte, N.A., Loh, P.-R., Palamara, P.F., Liu, X., et al.
- 529 (2017) Linkage disequilibrium–dependent architecture of human complex traits
- shows action of negative selection. *Nat. Genet.*, **49**, 1421–1427.
- 531 Georges, M., Charlier, C. & Hayes, B. (2018) Harnessing genomic information for
- 532 livestock improvement. Nat. Rev. Genet.
- 533 Gramlich, S., Liu, X., Favre, A., Alex Buerkle, C. & Karrenberg, S. (2021) A polygenic
- architecture with conditionally neutral effects underlies ecological differentiation in
- 535 *Silene. bioRxiv*, 2021.07.06.451304.
- 536 Haile-Mariam, M., Nieuwhof, G.J., Beard, K.T., Konstatinov, K.V. & Hayes, B.J.
- 537 (2013) Comparison of heritabilities of dairy traits in Australian Holstein-Friesian cattle
- 538 from genomic and pedigree data and implications for genomic evaluations:
- 539 Implication of genomic heritability for genomic evaluation. J. Anim. Breed. Genet.,
- 540 **130**, 20–31.
- 541 Harland, C., Coppieters, W., Mullaart, E., Charlier, C. & Georges, M. (2018) Rate of

- 542 *de novo mutation in dairy cattle and potential impact of reproductive technologies.*
- 543 Proc. World Congr. Genet. Appl. to Livest. Prod.
- 544 Harpending, H.C., Batzer, M.A., Gurven, M., Jorde, L.B., Rogers, A.R. & Sherry, S.T.
- 545 (1998) Genetic traces of ancient demography. Proc. Natl. Acad. Sci. USA, 95, 1961–
- 546 1967.
- 547 Hayes, B.J., Bowman, P.J., Chamberlain, A.J. & Goddard, M.E. (2009) Invited review:
- 548 Genomic selection in dairy cattle: Progress and challenges. J. Dairy Sci., 92, 433–
- 549 443.
- 550 Hayward, L.K. & Sella, G. (2019) Polygenic adaptation after a sudden change in
- 551 environment. *bioRxiv*, 792952.
- He, F., Arce, A.L., Schmitz, G., Koornneef, M., Novikova, P., Beyer, A., et al. (2016)
- 553 The Footprint of Polygenic Adaptation on Stress-Responsive Cis-Regulatory
- 554 Divergence in the *Arabidopsis* Genus. *Mol. Biol. Evol.*, **33**, 2088–2101.
- 555 Hudson, R.R. (1990) Gene Genealogies and the Coalescent Process. In Oxford
- 556 *Surveys in Evolutionary Biology* (ed. by Futuyma, D.J. & Antonovics, J.). Oxford Univ.
- 557 Press, Oxford, pp. 1–42.
- Jain, K. & Stephan, W. (2015) Response of Polygenic Traits Under Stabilizing
- 559 Selection and Mutation When Loci Have Unequal Effects. *G3*, **5**, 1065–1074.
- Jain, K. & Stephan, W. (2017a) Modes of Rapid Polygenic Adaptation. *Mol. Biol.*
- 561 *Evol.*, **34**, 3169–3175.
- Jain, K. & Stephan, W. (2017b) Rapid Adaptation of a Polygenic Trait After a Sudden
- 563 Environmental Shift. *Genetics*, **206**, 389–406.
- Jiménez-Mena, B., Tataru, P., Brøndum, R.F., Sahana, G., Guldbrandtsen, B. &
- 565 Bataillon, T. (2016) One size fits all? Direct evidence for the heterogeneity of genetic

- 566 drift throughout the genome. *Biol. Lett.*, **12**.
- 567 John, S. & Stephan, W. (2020) Important role of genetic drift in rapid polygenic
- 568 adaptation. *Ecol. Evol.*, **10**, 1278–1287.
- Josephs, E.B., Berg, J.J., Ross-Ibarra, J. & Coop, G. (2019) Detecting Adaptive
- 570 Differentiation in Structured Populations with Genomic Data and Common Gardens.
- 571 *Genetics*, **211**, 989–1004.
- 572 Kreiner, J.M., Tranel, P.J., Weigel, D., Stinchcombe, J.R. & Wright, S.I. (2020) The
- 573 genetic architecture and genomic context of glyphosate resistance in *Amaranthus*
- 574 *tuberculatus. bioRxiv*, 2020.08.19.257972.
- 575 Lande, R. (1983) The response to selection on major and minor mutations affecting a
- 576 metrical trait. *Heredity*, **50**, 47–65.
- 577 Le Corre, V. & Kremer, A. (2012) The genetic differentiation at quantitative trait loci 578 under local adaptation. *Mol. Ecol.*, **21**, 1548–1566.
- 579 Lemay, D.G., Lynn, D.J., Martin, W.F., Neville, M.C., Casey, T.M., Rincon, G., et al.
- 580 (2009) The bovine lactation genome: insights into the evolution of mammalian milk.
- 581 *Genome Biol.*, **10**, R43.
- 582 Liu, X., Li, Y.I. & Pritchard, J.K. (2019) Trans Effects on Gene Expression Can Drive
- 583 Omnigenic Inheritance. *Cell*, **177**, 1022-1034.e6.
- 584 MacEachern, S., Hayes, B., McEwan, J. & Goddard, M. (2009) An examination of
- 585 positive selection and changing effective population size in Angus and Holstein cattle
- 586 populations (*Bos taurus*) using a high density SNP genotyping platform and the
- 587 contribution of ancient polymorphism to genomic diversity in Domestic cattle. BMC
- 588 *Genom.*, **10**, 181.
- 589 MacLachlan, I.R., McDonald, T.K., Lind, B.M., Rieseberg, L.H., Yeaman, S. & Aitken,

- 590 S.N. (2021) Genome-wide shifts in climate-related variation underpin responses to
- selective breeding in a widespread conifer. *Proc. Natl. Acad. Sci. USA*, **118**,
- 592 e2016900118.
- 593 Meuwissen, T., Hayes, B. & Goddard, M. (2013) Accelerating Improvement of
- 594 Livestock with Genomic Selection. *Annu. Rev. Anim. Biosci.*, **1**, 221–237.
- 595 Nielsen, R. (2005) Molecular Signals of Natural Selection. *Annu. Rev. Genet.*, **39**,
 596 197–218.
- 597 Novembre, J. & Barton, N.H. (2018) Tread Lightly Interpreting Polygenic Tests of
- 598 Selection. *Genetics*, **208**, 1351–1355.
- 599 Pavlidis, P., Metzler, D. & Stephan, W. (2012) Selective Sweeps in Multilocus Models
- 600 of Quantitative Traits. *Genetics*, **192**, 225–239.
- 601 Pritchard, J.K. & Di Rienzo, A. (2010) Adaptation not by sweeps alone. *Nat. Rev.*
- 602 *Genet.*, **11**, 665–667.
- 603 Pritchard, J.K., Pickrell, J.K. & Coop, G. (2010) The Genetics of Human Adaptation:
- Hard Sweeps, Soft Sweeps, and Polygenic Adaptation. *Curr. Biol.*, **20**, R208–R215.
- 605 Qanbari, S., Pausch, H., Jansen, S., Somel, M., Strom, T.M., Fries, R., et al. (2014)
- 606 Classic Selective Sweeps Revealed by Massive Sequencing in Cattle. PLoS Genet.,
- 607 **10**, e1004148.
- 608 Qanbari, S., Pimentel, E.C.G., Tetens, J., Thaller, G., Lichtner, P., Sharifi, A.R., et al.
- 609 (2010) A genome-wide scan for signatures of recent selection in Holstein cattle.
- 610 Anim. Genet.
- 611 Racimo, F., Berg, J.J. & Pickrell, J.K. (2018) Detecting Polygenic Adaptation in
- 612 Admixture Graphs. *Genetics*, **208**, 1565–1584.
- 613 Robinson, M.R., Hemani, G., Medina-Gomez, C., Mezzavilla, M., Esko, T.,

- 614 Shakhbazov, K., et al. (2015) Population genetic differentiation of height and body
- 615 mass index across Europe. *Nat. Genet.*, **47**, 1357.
- 616 Rowan, T.N., Durbin, H.J., Seabury, C.M., Schnabel, R.D. & Decker, J.E. (2021)
- 617 Powerful detection of polygenic selection and evidence of environmental adaptation
- 618 in US beef cattle. *PLoS Genet.*, **17**, e1009652.
- 619 Sanjak, J.S., Sidorenko, J., Robinson, M.R., Thornton, K.R. & Visscher, P.M. (2018)
- 620 Evidence of directional and stabilizing selection in contemporary humans. *Proc. Natl.*
- 621 Acad. Sci. USA, 115, 151–156.
- 622 Savolainen, O., Lascoux, M. & Merila, J. (2013) Ecological genomics of local
- 623 adaptation. *Nat. Rev. Genet.*, **14**, 807–820.
- 624 Schoech, A.P., Jordan, D.M., Loh, P.-R., Gazal, S., O'Connor, L.J., Balick, D.J., et al.
- 625 (2019) Quantification of frequency-dependent genetic architectures in 25 UK Biobank
- traits reveals action of negative selection. *Nat. Commun.*, **10**, 790.
- 627 Sella, G. & Barton, N.H. (2019) Thinking About the Evolution of Complex Traits in the
- 628 Era of Genome-Wide Association Studies. Annu. Rev. Genom. Hum. Genet., 20,
- 629 461–493.
- 630 Sohail, M., Maier, R.M., Ganna, A., Bloemendal, A., Martin, A.R., Turchin, M.C., et al.
- 631 (2019) Polygenic adaptation on height is overestimated due to uncorrected
- 632 stratification in genome-wide association studies. *eLife*, **8**, e39702.
- 633 Sørensen, A.C., Sørensen, M.K. & Berg, P. (2005) Inbreeding in Danish Dairy Cattle
- 634 Breeds. J. Dairy Sci., 88, 1865–1872.
- 635 Soyeurt, H., Gillon, A., Vanderick, S., Mayeres, P., Bertozzi, C. & Gengler, N. (2007)
- 636 Estimation of Heritability and Genetic Correlations for the Major Fatty Acids in Bovine
- 637 Milk. J. Dairy Sci., **90**, 4435–4442.

- 638 Speidel, L., Forest, M., Shi, S. & Myers, S.R. (2019) A method for genome-wide
- 639 genealogy estimation for thousands of samples. *Nat Genet.*, **51**, 1321–1329.
- 640 Stephan, W. (2016) Signatures of positive selection: from selective sweeps at
- 641 individual loci to subtle allele frequency changes in polygenic adaptation. *Mol. Ecol.*,
- 642 **25**, 79–88.
- 643 Stephan, W. (2019) Selective Sweeps. *Genetics*, **211**, 5–13.
- 644 Stern, A.J., Speidel, L., Zaitlen, N.A. & Nielsen, R. (2021) Disentangling selection on
- 645 genetically correlated polygenic traits via whole-genome genealogies. *Am. J. Hum.*
- 646 *Genet.*, **108**, 219–239.
- 647 Stetter, M.G., Thornton, K. & Ross-Ibarra, J. (2018) Genetic architecture and
- selective sweeps after polygenic adaptation to distant trait optima. *PLoS Genet.*, **14**,
 e1007794.
- 650 Svardal, H., Jasinska, A.J., Apetrei, C., Coppola, G., Huang, Y., Schmitt, C.A., et al.
- 651 (2017) Ancient hybridization and strong adaptation to viruses across African vervet
- 652 monkey populations. *Nat. Genet.*, **49**, 1705–1713.
- 653 Thornton, K.R. (2019) Polygenic Adaptation to an Environmental Shift: Temporal
- 654 Dynamics of Variation Under Gaussian Stabilizing Selection and Additive Effects on a
- 655 Single Trait. *Genetics*, **213**, 1513–1530.
- 656 Turchin, M.C., Chiang, C.W., Palmer, C.D., Sankararaman, S., Reich, D., Genetic
- 657 Investigation of ANthropometric Traits (GIANT) Consortium, *et al.* (2012) Evidence of
- 658 widespread selection on standing variation in Europe at height-associated SNPs. *Nat*
- 659 *Genet.*, **44**, 1015–1019.
- 660 Uricchio, L.H., Kitano, H.C., Gusev, A. & Zaitlen, N.A. (2019) An evolutionary
- 661 compass for detecting signals of polygenic selection and mutational bias. *Evol. Lett.*,

- 662 **3**, 69–79.
- 663 Visscher, P.M. & Goddard, M.E. (2019) From R.A. Fisher's 1918 Paper to GWAS a
- 664 Century Later. *Genetics*, **211**, 1125–1130.
- 665 Vitti, J.J., Grossman, S.R. & Sabeti, P.C. (2013) Detecting Natural Selection in
- 666 Genomic Data. Annu. Rev. Genet., **47**, 97–120.
- 667 Vladar, H.P. de & Barton, N. (2014) Stability and Response of Polygenic Traits to
- 668 Stabilizing Selection and Mutation. *Genetics*, **197**, 749–767.
- 669 Watterson, G.A. (1975) On the number of segregating sites in genetical models
- 670 without recombination. *Theor. Popul. Biol.*, **7**, 256–276.
- 671 Whitlock, M.C. (2008) Evolutionary inference from Fst. *Mol. Ecol.*, **17**, 1885–1896.
- Wieters, B., Steige, K.A., He, F., Koch, E.M., Ramos-Onsins, S.E., Gu, H., et al.
- 673 (2021) Polygenic adaptation of rosette growth in Arabidopsis thaliana. *PLoS Genet.*,
- 674 **17**, e1008748.
- 675 Wollstein, A. & Stephan, W. (2014) Adaptive Fixation in Two-Locus Models of
- 676 Stabilizing Selection and Genetic Drift. *Genetics*, **198**, 685–697.
- 677 Wray, N.R., Kemper, K.E., Hayes, B.J., Goddard, M.E. & Visscher, P.M. (2019)
- 678 Complex Trait Prediction from Genome Data: Contrasting EBV in Livestock to PRS in
- 679 Humans. *Genetics*, **211**, 1131–1141.
- 680 Wray, N.R., Wijmenga, C., Sullivan, P.F., Yang, J. & Visscher, P.M. (2018) Common
- 681 Disease Is More Complex Than Implied by the Core Gene Omnigenic Model. *Cell*,
- 682 **173**, 1573–1580.
- Yang, J., Bakshi, A., Zhu, Z., Hemani, G., Vinkhuyzen, A.A.E., Lee, S.H., et al. (2015)
- 684 Genetic variance estimation with imputed variants finds negligible missing heritability
- for human height and body mass index. *Nat. Genet.*, **47**, 1114–1120.

- 686 Yeaman, S., Hodgins, K.A., Lotterhos, K.E., Suren, H., Nadeau, S., Degner, J.C., et
- *al.* (2016) Convergent local adaptation to climate in distantly related conifers.
- 688 *Science*, **353**, 1431.
- Zan, Y. & Carlborg, Ö. (2018) A Polygenic Genetic Architecture of Flowering Time in
- the Worldwide Arabidopsis thaliana Population. *Mol. Biol. Evol.*, **36**, 141–154.
- End Seder, M.A. (2008) Domestication and early agriculture in the Mediterranean Basin:
- 692 Origins, diffusion, and impact. Proc. Natl. Acad. Sci. USA, 105, 11597–11604.
- End Seng, J., Vlaming, R. de, Wu, Y., Robinson, M.R., Lloyd-Jones, L.R., Yengo, L., et al.
- 694 (2018) Signatures of negative selection in the genetic architecture of human complex
- 695 traits. *Nat. Genet.*, **50**, 746–753.
- 696 Zeng, J., Xue, A., Jiang, L., Lloyd-Jones, L.R., Wu, Y., Wang, H., et al. (2021)
- 697 Widespread signatures of natural selection across human complex traits and
- functional genomic categories. *Nat. Commun.*, **12**, 1164.
- 2699 Zhang, W., Reeves, G.R. & Tautz, D. (2021) Testing Implications of the Omnigenic
- 700 Model for the Genetic Analysis of Loci Identified through Genome-wide Association.
- 701 *Curr. Biol.*, **31**, 1092-1098.e6.
- 702 Zhao, F., McParland, S., Kearney, F., Du, L. & Berry, D.P. (2015) Detection of
- selection signatures in dairy and beef cattle using high-density genomic information.
- 704 Genet. Sel. Evol., **47**, 49.
- Zhou, X., Carbonetto, P. & Stephens, M. (2013) Polygenic Modeling with Bayesian
- 706 Sparse Linear Mixed Models. *PLoS Genet.*, **9**, e1003264.
- 707
- 708

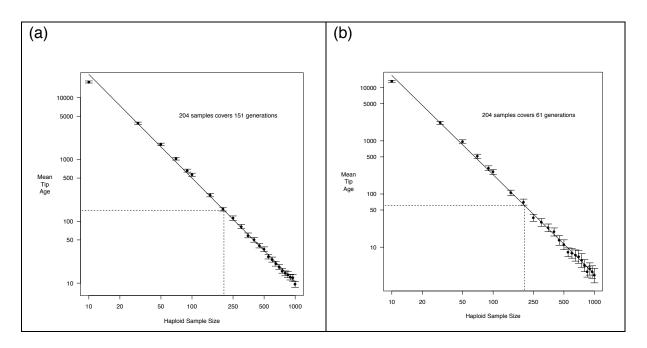
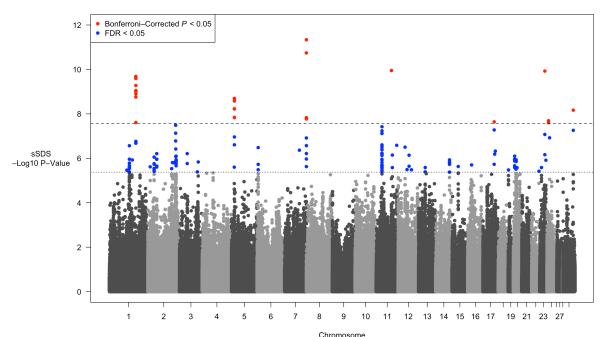
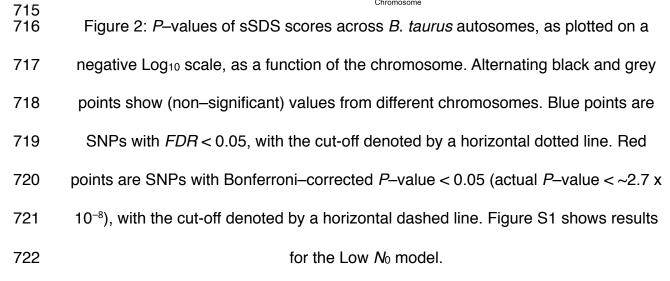


Figure 1: Simulated mean tip age for *B. taurus*, as a function of the number of haploid samples. Simulations assumed either (a) demography as inferred by Boitard et al. (2016b) (the 'High N_0 ' model), or (b) the same but with a smaller present–day N_e of 49 (the 'Low N_0 ' model). Points are the mean values; bars show 95% confidence intervals. The solid line is the best linear fit to the log of both values; dotted lines show the predicted tip age for 204 alleles.



sSDS Results for Bos taurus Autosomes



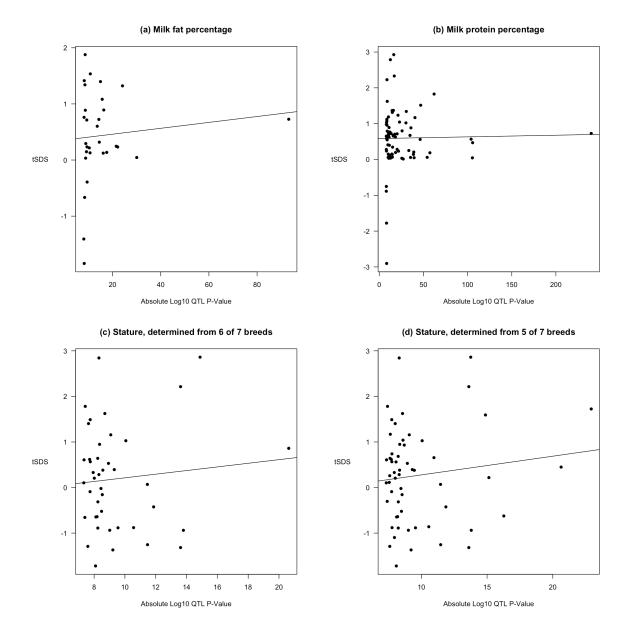


Figure 3: Relationship between tSDS scores near milk or stature QTLs, as noted in
the subheadings, and the absolute log *P*-value of QTLs. Lines show a linear model
regression fit. Figure S3 shows results assuming a low *N*₀ model.

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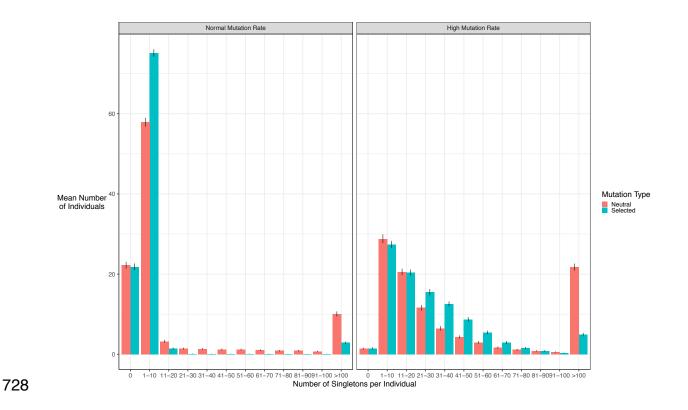
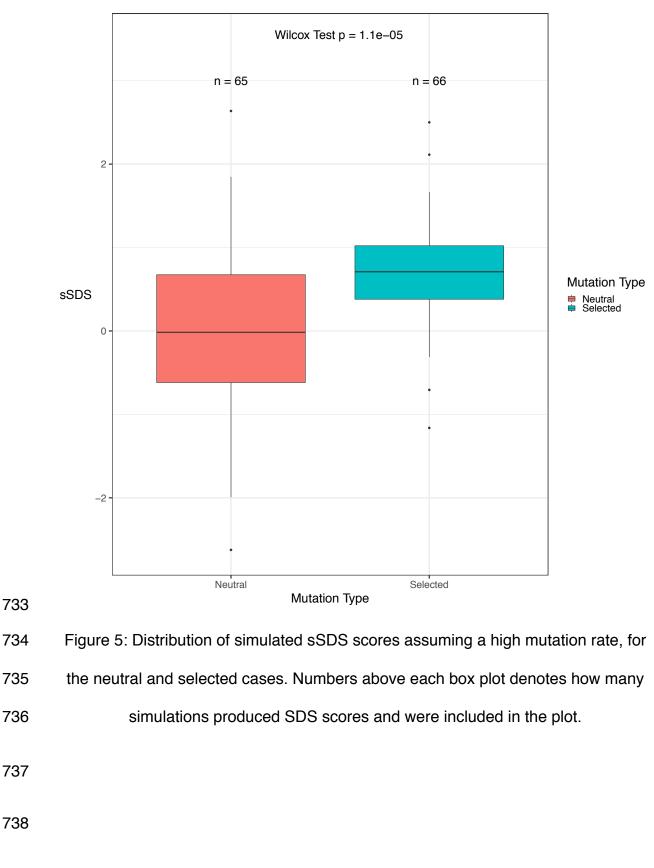


Figure 4: Mean distribution of singleton numbers per individual for each simulation,

730 wither assuming a standard mutation rate (left) or a 10-fold higher mutation rate

731 (right). Bars represent 95% confidence intervals.



Chromosome	Gene Name	Start Position	End Position	Gene Biotype	High, Low N_0
1	PPM1L	106405113	106727070	Protein Coding	High, Low
5	TMCC3	24306913	24595494	Protein Coding	High, Low
5	CEP83	24070404	24345243	Protein Coding	High, Low
17	U6	43381106	43381209	snRNA	Low
17	СТЅО	43364999	43381605	Protein Coding	Low
17	TDO2	43386894	43403747	Protein Coding	High, Low
23	OR12D2H	29291787	29292713	Protein Coding	High, Low
23	OR12D2E	29305933	29309785	Protein Coding	High, Low
24	GAREM1	24694637	24927333	Protein Coding	High, Low
29	NTM	34576918	34994005	Protein Coding	High, Low

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741 Table 1: Genes that overlap or lie close to Bonferroni–significant sSDS regions. The

'High, Low N_0 ' column specifies which genes are close to significant SNPs for each

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*N*₀ model.