Regulation by the Pitcher Plant Sarracenia purpurea of the Structure of its Inquiline Food

Web

AARON M. ELLISON[1]

Harvard University, Harvard Forest, 324 North Main Street, Petersham, Massachusetts 01366

NICHOLAS J. GOTELLI

University of Vermont, Department of Biology, Burlington, Vermont 04505

LESZEK A. BŁĘDZKI

Mount Holyoke College, Department of Biology, South Hadley, Massachusetts 01075

AND

JESSICA L. BUTLER[2]

Harvard University, Harvard Forest, 324 North Main Street, Petersham, Massachusetts 01366

[1] Corresponding author: Telephone: (978)-756-6178; E-mail: aellison@fas.harvard.edu
[2] Present Address: Barr Engineering Company, Minneapolis, Minnesota 55435

ABSTRACT.—Phytotelmata, the water-filled habitats in pitcher plants, bromeliad tanks, and tree-holes, host multitrophic food webs that are model experimental systems for studying food-web structure and dynamics. However, the plant usually is considered simply as an inert container, not as an interacting part of the food web. We used a response-surface and factorial field experiment to determine effects of nutrient enrichment (multiple levels of NH_4NO_3, PO_4, and captured prey), top predator (removed or present), and the plant itself (with or without plastic tubes inserted into the pitchers to isolate the food web from the plant) on the macrobial food web within the modified leaves ("pitchers") of the carnivorous pitcher plant *Sarracenia purpurea*. Connection to the plant, addition of NH_4NO_3 and removal of the top predator significantly increased the food web's saturation, defined as its trophic depth and number of interactions. No effects on food-web saturation resulted from addition of PO_4 or supplemental prey. Plants such as *S. purpurea* that create phytotelmata are more than inert containers and their inhabitants are more than commensal inquilines. Rather, both the plant and the inquilines are partners in a complex network of interactions.

Understanding controls on food web structure has been an organizing focus for

community ecology for well over a century (*e.g.,* Pimm, 1982; Polis, 1994; Carpenter *et al.,*

2001; McCann, 2012; Wokovich *et al.* 2014; Layman *et al.*, 2015; Zou *et al.*, 2016). As primary

producers, plants are the bottom-up controllers of "green" food webs, whereas as "detritus" the

decomposing leaf litter of plants is the resource base for "brown" food webs (Cebrian and

Lartigue, 2004). The various roles of plants in structuring both green and brown food webs have

been studied extensively, but an additional role for plants in structuring food webs is much less

appreciated. In tree holes and modified leaves, plants create habitats for a range of aquatic food

webs. Whereas the complex, mostly detritus-based food webs in these "phytotelmata"—water-

filled habitats in, for example, tree holes, bromeliads, and pitcher plants (Maguire, 1971)—have

been used as model experimental systems for studying the dynamics of aquatic food webs (*e.g.,*

Kitching, 2000, 2001; Kneitel and Miller, 2002; Miller *et al.,* 2002; Ellison *et al.,* 2003;

Srivastava *et al.,* 2004), the phytotelmata itself normally is regarded simply as a container for its

"inquilines": animals that live commensally "within the nest or abode of another" (OED, 2019).

It has been recognized only rarely that the plant itself can mediate trophic interactions of

inquiline food webs.

Research with carnivorous pitcher plants has provided two notable examples of the

importance of phytotelmata on inquiline food webs. Bradshaw and Creelman (1984)

experimentally showed that the carnivorous pitcher plant *Sarracenia purpurea* L.

(Sarraceniaceae) and two of its obligate inquilines, larvae of the pitcher-plant mosquito

Wyeomyia smithii (Coq.) and the pitcher-plant midge *Metriocnemus knabi* Coq., are mutualists.

These two inquilines accelerate the breakdown and decomposition of arthropod prey captured by

S. purpurea (Heard, 1994). Ammonia (NH_4) and carbon dioxide (CO_2) released as bacteria

mineralize nutrients in the prey (Butler *et al.*, 2008) and NO_3, NH_4, and PO_4 regenerated by

rotifers (Błędzki and Ellison, 1998, 2002; Błędzki *et al.*, 2018) are absorbed by the plant and

used for photosynthesis, subsequent growth, and reproduction (Bradshaw and Creelman, 1984;

Joel and Gepstein, 1985; Butler and Ellison, 2007). The plant, in turn, oxygenates the pitcher

fluid (Bradshaw and Creelman, 1984; Joel and Gepstein, 1985; Sirota *et al.*, 2013), which

maintains homeostatic conditions for the inquiline food web and prevents a switch to anaerobic

conditions within the phytotelmata, which can nonetheless occur if prey are superabundant

(Krieger and Kourtev, 2012; Sirota *et al.*, 2013; Northrop *et al.*, 2017).

Similarly, one of the Asian pitcher plants, *Nepenthes bicalcarata* Hook.f. (Nepenthaceae)

hosts the ant *Colobopsis schmitzi* (Stärcke) (Formicidae) in the plant's hollow tendrils (Clarke

and Kitching, 1995). The ants move out of the tendrils to forage within the pitchers on large prey

that the plant captures. The ants also clean the pitcher's peristome, reducing fungal colonization

and increasing prey-capture efficiency (Thornham *et al.*, 2011), which in turn leads to greater

nutrient acquisition, photosynthesis, and growth of this perennial plant (Bazile *et al.*, 2012).

The lack of attention to linkages between food-web structure or dynamics and the habitat

in which the food web itself assembles is not restricted to phytotelmata. Although spatiotemporal

dynamics of food webs have been studied, documented, and modeled (*e.g.*, Poisot *et al.*, 2015;

Pilosof *et al.*, 2017; Piovia-Scott et al. 2017; Hutchinson et al., 2019; Schiaffino *et al.*, 2019),

how complete food webs assemble in a habitat that itself is changing through time and interacts

with the food web itself remains unexplored. We are unsure whether this lack of attention results

from the rarity of such situations, our failure to perceive them, or the difficulty in studying them.

However, living phytotelmata such as pitcher plants provide opportunities to study

experimentally the dynamics of a food web that assembles in a habitat that itself is a living,

growing organism.

Here, we describe how nutrient enrichment, top predators, and the living habitat of the

plant itself affect the structure of the four trophic-level food web that inhabits the modified

pitcher-shaped leaves (Arber, 1941) of *S. purpurea*. In a set of controlled field experiments, we

first examined the impact of nutrient deposition on the food web that assembles in *S. purpurea*

pitchers. Atmospheric deposition of N and P in northeastern North America provide an

immediate, biologically reactive source of nutrients (as NO_3, NH_4, and PO_4) that can be used by

both the plant and the microbial community at the base of its aquatic food web (Ellison and

Gotelli, 2002). Increasing N, but not P, significantly increased the food web's "saturation" (a

measure d indicating how many trophic levels and interactions were present in the web and

described in detail in Materials and Methods), but saturation declined at highest levels of N

addition. Second, we severed the connection between the plant and its food web by inserting

closed plastic tubes (or appropriate controls) in between pitchers and their food webs to

determine if effects of experimental nutrient enrichment through additions of prey and inorganic

nutrients resulted from direct effects on the food web or indirect effects mediated by the plant.

97   **MATERIALS AND METHODS**

98   STUDY SPECIES

Sarracenia purpurea (the northern, or purple, pitcher plant) is a long-lived (> 50 yr),

rosette-forming, carnivorous perennial plant common to bogs throughout Canada and the eastern

United States. Pitcher plants have weakly developed root systems; nutrients are obtained

principally from insect prey attracted to the brightly colored pitchers and extrafloral nectaries

(Bennett and Ellison, 2009). Unlike the other 10 species of *Sarracenia* (Mellichamp and Case,

2009)*, S. purpurea* pitchers fill with rainwater and these semi-permanent pools (phytotelmata)

are colonized by a suite of specialized invertebrates and more generalist protists and bacteria

(Addicott, 1974; Cochran-Stafira and von Ende, 1998; Błędzki and Ellison, 2003; Ellison *et al.*,

2003).

Arthropod prey captured by the plant is the basal resource for this detritus-based food

web, which shreds the prey and mineralizes the available nutrients (Butler and Ellison, 2007;

Butler *et al*., 2008). Prey capture initially increases as pitchers open, expand, and harden; reaches

a peak when the leaves are two–three weeks old; and then declines during to near zero by the end

of the growing season (Fish and Hall, 1978; Ellison *et al.*, 2003). Drowned carcasses of captured

prey (primarily ants and flies; Ellison and Gotelli, 2009) are shredded by larvae of the pitcher-

plant midge *Metriocnemus knabi* and larvae of the sarcophagid fly *Fletcherimyia fletcheri*

(Aldrich); the shredded prey are broken down further by bacteria (Cochran-Stafira and von Ende,

1998; Heard, 1994; Baiser *et al.*, 2011). The bacteria, organic matter, protozoa, and algae in turn

are consumed by the bdelloid rotifer *Habrotrocha rosa* Donner and a variety of protozoa, which,

along with the pitcher-plant mite *Sarraceniopus gibsoni* (Nesbitt), also consume detritus directly

(Wallace *et al.,* 2006). Protozoa also consume bacteria (Cochran-Stafira and von Ende, 1998).

These primary consumers are preyed on by larvae of the pitcher-plant mosquito *W. smithii* (all

these macrobes will be referred to below using their generic epithets). Late instars of

Fletcherimyia also prey on rotifers and 1st and 2nd instars of *Wyeomyia* (Błędzki and Ellison,

1998; Butler *et al.*, 2008; Baiser *et al.*, 2013). Not only does *Fletcherimyia* complete its larval

development within pitchers, but the adult flies are pollinators of *S. purpurea* (Ne'eman *et al.*,

2006).

In all the experiments described below, *food-web saturation* was our response variable.

Each macrobial taxon (rotifer, mite, midge, mosquito, sarcophagid fly) in the food web was

assigned a binary value representing its presence (Table 1; Fig. 1); we assumed all food webs

had protozoa and bacteria. The saturation of the food web in a pitcher was defined as the sum of

the equivalent decimal values of each macrobial taxon present. There are 32 possible food webs

that can be assembled from these five taxa; the decimal value for each food web ranges from 0

(no macrobial taxon present) to 31 (all macrobial taxa present), with increasing numbers

indicating more "saturated" food webs. Food webs with higher saturation values had both more

trophic levels present and more trophic links present (Fig. 1).

136   STUDY SITE

The experiments reported here were conducted at Swift River Bog, a 1.9-ha glacial kettle

bog in Belchertown, Massachusetts, USA (42.27 °N, −72.34 °W, 124 m a.s.l.; Kearsley, 1999).

This bog has a lagg (moat) around it that never exceeds 2 m in width, but there is no open water

within the bog itself. Common *Sphagnum* species present at Swift River Bog were *Sp.*

angustifolium (C.E.O. Jensen ex Russow) C.E.O. Jensen*, Sp. magellanicum* Brid., and *Sp.*

rubellum Wilson (Ellison and Gotelli, 2016). The dominant vascular plants growing on the bog

mat were shrubs in the Ericaceae: *Chamaedaphne calyculata* (L.) Moench (leatherleaf)*, Kalmia*

polifolia Wangenheim (bog laurel), *K. angustifolia* L. (sheep laurel), *Rhododendron viscosum*

(L.) Torrey (swamp rhododendron) and *Vaccinium macrocarpon* Aiton (cranberry); scattered

Picea mariana (Miller) Britton (Pinaceae; black spruce) grow across the bog. The site has a large

population (1000s of plants) of *S. purpurea* (Ne'eman *et al.,* 2006; Ellison and Gotelli, 2016).

Total nitrogen deposition ($NO_3 + NH_4$) at the National Atmospheric Deposition Program

(NADP) Quabbin Reservoir monitoring station in the same watershed (MA08: 42.3925 °N,

−72.3444 °W) ranged from 1.25–1.60 mg/L during the years of this study, and has averaged 1.19

(range: 0.58–2.12) mg/L since measurements began in 1982 (National Atmospheric Deposition

Program, 2020). Phosphorus deposition is not monitored by NADP or other agencies in the US,

but P deposition may have significant effects in P-limited ecosystems or alter stoichiometric

ratios in otherwise N-limited ones (Mahowald *et al.*, 2008; Tipping *et al.*, 2014). Although *S.*

purpurea is stoichiometrically N- or N+P-limited (Ellison, 2006), field experiments have

suggested that *S. purpurea* preferentially takes up P from supplemental prey (Wakefield *et al.*,

2005).

159   EXPERIMENT 1 – IMPACTS OF N AND P ON FOOD-WEB SATURATION

The first experiment investigated the effects of inorganic N and P (as would be

accumulated in the phytotelmata by atmospheric deposition) on food-web saturation in situ. We

used a factorial response-surface design (Cottingham *et al.*, 2005) in a "pulse" experiment (sensu

Bender *et al.*, 1984) in which 5 levels each of NH_4NO_3 (4.687, 9.375, 18.75, 37.5, and 75.0 mM)

and PO_4 (0.625, 1.25, 2.5, 5.0, and 10.0 mM) and controls (distilled, deionized water: dH_2O)

were added biweekly to each pitcher of 130 non-flowering plants that were randomly assigned to

one of five groups of 26 plants. The levels of N used bracketed the observed deposition of N

measured by NADP in the years before the study, whereas the levels of P were chosen to span

the N:P ratios previously used by Ellison and Gotelli (2002) in other nutrient addition

experiments with *S. purpurea*.

Plants were located at least 10 m (linear distance) from the lagg surrounding the bog,

separated from one another by at least 50 cm, and atop the *Sphagnum* mat but not under shrubs

or trees. Each plant within a group was randomly assigned a nutrient treatment (one of the 25

possible N × P combinations or the dH$_2$O control). Beginning on 4 June 2001, 20 ml of nutrient

solution was added once to each pitcher 2-3 days after it had opened and fully hardened. Plants

were checked every three days for new leaf production. Every 3 wk (25 June, 16 July, 6 August,

27 August, and 17 September), a complete set of 26 plants was harvested. At each harvest,

pitcher fluid was pipetted into sterile tubes (one tube/leaf) and the presence or absence of each

member of the food web was determined.

We ran a parallel "press" experiment (sensu Bender *et al.*, 1984) in which we

haphazardly located 26 non-flowering plants and randomly assigned each of them to one of the

same 26 nutrient treatments as in the pulse experiment. 2-3 d after each pitcher had opened and

hardened, 20 ml of nutrient solution was added to it, and the 20-ml level was marked on the side

of the pitcher with indelible ink. Every two weeks, the food web was pipetted out of each pitcher

and into a petri dish. Presence of each macrobe was determined in the field, and all were returned

to the pitcher. Rotifers and protozoa were counted in the lab in 1-ml subsamples taken at the

same time. After field counts of invertebrates, the pitcher fluid was poured back into the pitcher

and additional nutrient solution was added to maintain the fluid level at 20 ml. This experiment

ran from 4 June–17 September 2001.

EXPERIMENT 2 – JOINT INFLUENCE OF THE PLANT, NUTRIENTS, AND PREDATION ON FOOD-WEB

STRUCTURE

The second experiment investigated how changes in food-web saturation resulted from

interactions between bottom-up effects of nutrient addition, top-down effects of predation, and

effects of the plant itself. We used a four-level factorial/response-surface experiment with 212

non-flowering plants located at least 10 m (linear distance) from the lagg surrounding the bog,

separated from one another by at least 50 cm, and atop the *Sphagnum* mat but not under shrubs

or trees. As in Experiment 1, plants were assigned randomly to unique combinations of

treatments that included physically isolating the inquiline food web within the phytotelmata.

The first treatment of the factorial assessed the influence of the plant itself on the food

web and had three levels. Pitcher interiors were: [1] fitted with intact polyethylene tubes (2.2-cm

diameter) (inquilines and their fluid isolated from effects of pitchers; n = 64 plants); [2] fitted

with perforated (ten 6.35-mm diameter drilled holes) polyethylene tubes (polyethylene tube

control; n = 64 plants); or [3] left without polyethylene tubes (unmanipulated control; n = 64

plants).

The second treatment was a continuous (ordinal) effector that examined the effects of N

deposition. One of eight different concentrations of inorganic nitrogen (NH_4NO_3: 0, 1.56, 3.1,

6.25, 12.5, 25, 50, or 100 mg N/L) was added to a randomly selected plant; there were n = 8

replicates of each N concentration in each set of 64 tube/tube-control/no tube treatments.

Because there was a constant uptake of NH_4NO_3 in control and tube-control pitchers, we

supplied an equivalent NH_4NO_3-N load each wk into each pitcher. Plant liquid volume was

maintained at two-thirds full by adding nutrient solutions or deionized water as necessary. Plants

with intact tubes were not able to assimilate NH_4NO_3 supplied to the tubes; we assumed loss

from volatilization and bacterial utilization was small compared to plant assimilation. Therefore,

we simply topped off each tube to two-thirds full every week with NH_4NO_3 solution.

The third treatment examined effects of prey availability (organic nutrients).

Supplemental prey (10 lab-raised fruit flies/wk) were added to one-half of each inorganic N ×

tube combination.

Finally, because of the importance of *Fletcherimyia* at multiple points in this food web

(Błędzki and Ellison, 1998; Butler *et al.*, 2008; Baiser *et al.*, 2013), the fourth treatment assessed

the influence of this top predator on the development of the pitcher-plant food web in the context

of bottom-up effects of prey and inorganic N addition and habitat effects of the pitcher itself.

Second instar *Fletcherimyia* larvae were either removed from pitchers (one-half of each N × tube

× fruit-fly treatment combination) or added to pitchers (one-half of each N × tube × fruit-fly

treatment combination).

In total, this factorial/response-surface design resulted in $N = 2$ plants in each tube × N ×

fruit-fly × *Fletcherimyia* treatment combination. We also haphazardly chose 20 additional plants

growing nearby in the bog as unmanipulated controls. One treated plant from each treatment

combination (96 pitchers) and half the unmanipulated controls were harvested after 4 wk, and the

remainder after 8 wk. Food-web saturation in this experiment was computed based on all

inquilines present including *Fletcherimyia*, except in the *Fletcherimyia* addition treatments (to

avoid simply documenting the addition in our calculation).

233   STATISTICAL ANALYSIS AND DATA AVAILABILITY

Data were analyzed using standard linear regression, analysis of variance or analysis of

covariance using functions in the base package of the R statistical software system, version 3.6.3

(R Core Team 2020).

All data from this experiment are available from the Harvard Forest Data Archive, dataset

HF193 (https://harvardforest.fas.harvard.edu/exist/apps/datasets/showData.html?id=hf193) and

the Environmental Data Initiative

(https://dx.doi.org/doi:10.6073/pasta/3bd2c920cf431442d34227f3f5fc7c2a).

RESULTS

EXPERIMENT 1

In the pulse experiment, which established the baseline response of the inquiline food web to bottom-up control by inorganic nutrient additions, food-web saturation peaked at intermediate levels of N added ($P = 0.001$), was significantly higher in leaves produced earlier in the season ($P = 0.025$), and in leaves harvested in mid-summer ($P = 0.033$) (Fig. 2). Although the amount of P added did not alter food web saturation ($P = 0.42$), there was a significant N × P interaction ($P = 0.036$).

Food-web saturation in the coincident press experiment also peaked at intermediate levels of N added ($P = 0.05$) and was highest in mid-summer ($P < 0.001$) in those leaves that were produced earliest in the season ($P = 0.04$) (Fig. 2). As in the pulse experiment, the amount of P added did not alter food web saturation ($P = 0.54$), but in contrast to the results of the pulse experiment, there was no significant N × P interaction in the press experiment ($P = 0.58$).

EXPERIMENT 2

Inquilines colonized all the plants, but *Wyeomyia* was observed less frequently in pitchers with intact tubes than in pitchers with perforated tubes or no tubes at all (Table 2). Similarly, *Fletcherimyia* colonized only two pitchers with intact tubes and the added *Fletcherimyia* survived in only one pitcher with an intact tube (Table 2). Occurrences of the other macrobial inquilines were unrelated to tube treatment or *Fletcherimyia* addition (Table 2).

Connections with the plant increased food-web saturation: food webs in pitchers without tubes or with perforated tubes were significantly more saturated than food webs in pitchers with

tubes (Fig. 3, right panel; P = 0.0003). Food webs in tubes rarely had predators (*Wyeomyia* or

Fletcherimyia), as these Diptera would not oviposit in intact tubes. Removal of *Fletcherimyia*

from plants led to an increase in food web saturation (Fig. 3, left panel; P = 0.0003), as this

predator consumes both rotifers and *Wyeomyia* larvae. Neither additions of prey nor inorganic

nitrogen (NH_4NO_3-N) alone altered food-web saturation (P = 0.37; P = 0.36, respectively).

However, there was a nutrient × *Fletcherimyia* interaction (P = 0.03): in the presence of

Fletcherimyia, increasing NH_4NO_3 led to decreased saturation of the food web. In contrast, in the

absence of *Fletcherimyia*, increasing NH_4NO_3 led to increased saturation of the food web. None

of the other two-way, three-way, or four-way interaction terms was significant (P > 0.18, all

cases).

275   DISCUSSION

Plant productivity, expressed as new biomass eaten by herbivores or shed litter

decomposed by detritivores, exerts bottom-up control of most food webs (McCann 2012;

Wokovich *et al.* 2014; Zou *et al.*, 2016). As phytotelmata (Maguire, 1971), plants produce

habitat for aquatic food webs (Kitching, 2000, 2001; Ellison *et al.,* 2003; Srivastava *et al.,* 2004),

but other effects of phytotelmata on food webs have been studied only rarely (Bradshaw and

Creelman, 1984; Joel and Gepstein, 1985; Clarke and Kitching, 1995; Thornham *et al.*, 2011;

Bazile *et al.*, 2012). The experiments described here illustrate that the phytotelmata of northern

pitcher plants are not simply inert containers in which complex aquatic food webs assemble and

thrive. Rather, the plant itself interacts with, and determines the strength of, the bottom-up and

top-down controls of the food web.

Unlike pitchers in other species of *Sarracenia* that are covered by a reflexed 'hood',

pitchers of *S. purpurea* are open to the sky and accumulate water from precipitation. This has

two consequences. First, as pitchers accumulate water, they create the habitat for the inquiline

food web. Second, any nutrients that accompany precipitation can enter the food web through

bacterial mineralization and immobilization.

Independent of addition of inorganic nutrients, supplemental prey, or presence of top

predators, the plant itself had a net positive effect on inquiline food-web saturation. Food webs in

containers isolated from the plant were 10–20% less saturated than food webs in perforated tubes

or unmanipulated controls of comparable size. This result agrees with previous observations of,

and experiments with, *S. purpurea* and its inquilines, in which the plant oxygenated the pitcher

fluid, maintaining homeostatic conditions for the inquiline food web and preventing a switch to

anaerobic conditions (Bradshaw and Creelman, 1984; Joel and Gepstein, 1985; Sirota *et al.*,

2013).

Further addition of inorganic N, as is occurring through atmospheric deposition,

increased food-web saturation at intermediate levels, but saturation declined as N addition was

increased further (Fig. 2). We hypothesize that this result may reflect bottom-up effects of N on

microbial productivity (Moorhead and Sinsabaugh, 2006), shifts in how *S. purpurea* uses

different forms of N (Karagatzides *et al.*, 2009), or interactions between the plant and the

bacteria that mineralize nitrogen for it (Butler *et al.*, 2008; Mouquet *et al.*, 2008). Furthermore, N

addition alters pitcher morphology, reducing pitcher size (volume) while increasing the area of

its photosynthetic keel (Ellison and Gotelli, 2002; Bott *et al.*, 2008). Although smaller pitchers

have less complex food webs (Gotelli and Ellison, 2006), this effect was not observed or

measured here because we worked only with pitchers that were fully opened, not those that were

produced after nutrients were added to early-season pitchers and that would have expressed

morphological differences following nutrient translocation and use in new pitcher production

(Butler and Ellison, 2007).

Top-down effects of predation in the inquiline food web of *S. purpurea* have been

described repeatedly (*e.g.,* Addicott, 1974; Kneitel and Miller, 2002; Hoekman, 2007, 2011;

Peterson *et al.*, 2008), but in our experiments, these effects were manifest only when the food

web was isolated from the plant. Food webs without the predator *Fletcherimyia* were more

saturated (up to the level below *Fletcherimyia*) than food webs with this predator. The

interactions that we observed between top-down effects of *Fletcherimyia* and bottom-up effects

of inorganic N additions are mirrored by modelled trade-offs in overall system nitrogen

mineralization modulated by interactions between plants and mineralizing bacteria and between

plants and higher trophic levels (Mouquet *et al.*, 2008).

Overall, the results presented here together with those from other studies on pitcher-plant

food webs illustrate that top-down and bottom-up effects on food-web structure are dependent on

the habitat context in which they occur. Anthropogenic processes such as fossil-fuel combustion

leading to atmospheric deposition of inorganic nutrients can alter habitat structure and shift

control of food web structure from strictly bottom-up control to a mixture of top-down and

bottom-up controls. These consequences are especially evident in habitats that themselves are

living structures that respond to the same forcing processes at similar time scales. Plants such as

S. purpurea that create phytotelmata are more than inert containers and their inhabitants are more

than commensal inquilines. Rather, both the plant and the 'inquilines' are partners in complex

networks of interactions. Our work and that of others illustrate clearly that the microecosystem

created by *S. purpurea* can provide a model experimental system for exploring such interactions.

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LITERATURE CITED

ADDICOTT, J.F. 1974. Predation and prey community structure: an experimental study of the effect of mosquito larvae on the protozoan communities of pitcher plants. *Ecology*, **55**:475–492.

ARBER, A. 1941. On the morphology of the pitcher-leaves in *Heliamphora, Sarracenia, Darlingtonia, Cephalotus*, and *Nepenthes*. *Ann. Bot.,* **5**:563–578.

BAISER, B., R. ARDESHIRI, AND A.M. ELLISON. 2011. Species richness and trophic diversity increase decomposition in a co–evolved food web. *PLoS ONE,* **6**:e20672.

———, H.L. BUCKLEY, N.J. GOTELLI, AND A.M. ELLISON. 2013. Predicting food-web structure with metacommunity models. *Oikos,* **122**:492–506.

BAZILE, V., J.A. MORAN, G. LE MOGUEDEC, D.J. MARSHALL, AND L. GAUME. 2012. A carnivorous plant fed by its ant symbiont: a unique multi–faceted nutritional mutualism. *PLoS ONE,* **7**:e36179.

BENDER, E.A., T.J. CASE, AND M.E. GILPIN. 1984. Perturbation experiments in community ecology: theory and practice. *Ecology,* **65**:1–13.

BŁĘDZKI, L.A. AND A. M. ELLISON. 1998. Population growth and production of *Habrotrocha rosa* Donner (Rotifera: Bdelloidea) and its contribution to the nutrient supply of its host, the northern pitcher plant, *Sarracenia purpurea* L. (Sarraceniaceae). *Hydrobiologia,* **385**:193–200.

——— AND ———. 2002. Nutrient regeneration by rotifers in new England (USA) bogs. *Verh. Int. Ver. Limnol.,* **28**:1328–1331.

——— AND ———. 2003. Diversity of rotifers from Northeastern USA bogs with new species records for North America and New England. *Hydrobiologia*, **497**:53–62.

———, J.L. BUBIER, A.M. ELLISON, AND T.R. MOORE. 2018. Ecology of rotifers and their

unappreciated source of nitrogen and phosphorus in temperate northeastern American bogs.

Fundam. Appl. Limnol., **191**:277–287.

BOTT, T., G.A. MEYER, AND E.B. YOUNG. 2008. Nutrient limitation and morphological plasticity

of the carnivorous pitcher plant *Sarracenia purpurea* in contrasting wetland environments.

New Phytol., **180**:631–641.

BRADSHAW, W.E. AND R. A. CREELMAN. 1984. Mutualism between the carnivorous purple

pitcher plant *Sarracenia purpurea* and its inhabitants. *Am. Midl. Nat.,* **112**:294–304.

BUTLER, J.L. AND A.M. ELLISON. 2007. Nitrogen cycling dynamics in the carnivorous northern

pitcher plant, *Sarracenia purpurea. Func. Ecol.,* **21**:835–843.

———, N. J. GOTELLI, AND A. M. ELLISON. 2008. Linking the brown and green: nutrient

transformation and fate in the *Sarracenia* microecosystem. *Ecology,* **89**:898–904.

CARPENTER, S.R., J.J. COLE, J.R. HODGSON, J.F. KITCHELL, M.L. PACE, D. BADE, K.L.

COTTINGHAM, T.E. ESSINGTON, J.N. HOUSER, AND D.D. SCHINDLER. 2001. Trophic cascades,

nutrients, and lake productivity: whole-lake experiments. *Ecol. Mono.,* **71**:163–186.

CEBRIAN, J. AND J. LARTIGUE. 2004. Patterns of herbivory and decomposition in aquatic and

terrestrial ecosystems. *Ecol. Mono.,* **74**:237–259.

CLARKE, C.M. AND R.L. KITCHING. 1995. Swimming ants and pitcher plants: a unique ant-plant

interaction from Borneo. *J. Trop. Ecol.,* **11**:589–602.

COCHRAN-STAFIRA, D.L. AND C.N. VON ENDE. 1998. Integrating bacteria into food webs: studies

with *Sarrcenia purpurea* inquilines. *Ecology,* **79**:880–898.

COTTINGHAM, K.L., J.T. LENNON, AND B.L. BROWN. 2005. Knowing when to draw the line:

designing more informative ecological experiments. *Front. Ecol. Env.,* **3**:145–152.

ELLISON, A.M. 2006. Nutrient limitation and stoichiometry of carnivorous plants. *Plant Biol.,*

8:740–747.

——— AND N.J. GOTELLI. 2002. Nitrogen availability alters the expression of carnivory in the

northern pitcher plant *Sarracenia purpurea. Proc. Nat. Acad. Sci.,* **99**:4409–4412.

——— AND ———. 2009. Energetics and the evolution of carnivorous plants – Darwin's "most

wonderful plants in the world." *J. Exp. Bot.,* **60**:19–42.

——— AND ———. 2016. Moths, ants and pitcher plants in bogs at Harvard Forest and

Belchertown MA 2007–2013. Harvard Forest Data Archive: HF159.

dx.doi.org/10.6073/pasta/37ee70bbf38b3f95bc955dc555c63188 [date of access: 12 January

2020].

———, ———, J.S. BREWER, L. COCHRAN-STAFIRA, J. KNEITEL, T.E. MILLER, A.S. WORLEY,

AND R. ZAMORA. 2003. The evolutionary ecology of carnivorous plants. *Adv. Ecol. Res.,*

33:1–74.

FISH D. AND D.W. HALL. 1978. Succession and stratification of aquatic insects inhabiting the

leaves of the insectivorous pitcher plant *Sarracenia purpurea. Am. Midl. Nat.,* **99**:172–183.

GOTELLI, N.J. AND A.M. ELLISON. 2006. Food-web models predict species abundance in

response to habitat change. *PLoS Bio.,* **44**:e324.

HEARD, S.B. 1994. Pitcher-plant midges and mosquitoes: a processing chain commensalism.

Ecology, **75**:1647–1660.

HOEKMAN, D. 2007. Top-down and bottom-up regulation in a detritus-based aquatic food web: a

repeated field experiment using the pitcher plant (*Sarracenia purpurea*) inquiline

community. *Am. Midl. Nat.,* **157**:52–62.

———. 2011. Relative importance of top-down and bottom-up forces in food webs of

Sarracenia pitcher communities at a northern and a southern site. *Oecologia* (*Berl.*),

165:1073–1082.

HUTCHINSON, M.C., B.B. MORA, S. PILOSOF, A.K. BARNER, S. KÉFI, E. THÉBAULT, P. JORDANO,

AND D.B. STOUFFER. 2019. Seeing the forest for the trees: putting multilayer networks to

work for community ecology. *Func. Ecol.,* **33**:206–217.

JOEL, D.M AND S. GEPSTEIN. 1985. Chloroplasts in the epidermis of *Sarracenia* (the American

pitcher plant) and their possible role in carnivory – an immunocytochemical approach.

Physiol. Plant., **63**:71–75.

KARAGATZIDES, J.D., J.L. BUTLER, AND A.M. ELLISON. 2009. The pitcher plant *Sarracenia*

*purpure*a can directly acquire organic nitrogen and short-circuit the inorganic nitrogen

cycle. *PLoS ONE,* **4**:e6164.

KEARSLEY, J. 1999. *Non-forested acidic peatlands of Massachusetts: a statewide inventory and*

vegetation classification. Westborough, MA, USA: Massachusetts Natural Heritage and

Endangered Species Program.

KITCHING, R.L. 2000. Food webs and container habitats: the natural history and ecology of

phytotelmata. Cambridge University Press. 429 p.

———. 2001. Food webs in phytotelmata: "bottom-up" and "top-down" explanations for

community structure. *Ann. Rev. Entomol.,* **46**:729–760.

KNEITEL, J.M. AND T.E. MILLER. 2002. Resource and top-predator regulation in the pitcher plant

(*Sarracenia purpurea*) inquiline community. *Ecology,* **83**:680–688.

KRIEGER, J.R. AND P.S. KOURTEV. 2012. Detection of methanogenic archaea in the pitchers of

the northern pitcher plant (*Sarracenia purpurea*). *Can. J. Microbiol.,* **58**:189–194.

LAYMAN, C.A., S.T. GIERY, S. BUHLER, R. ROSSI, T. PENLAND, M.N. HENSON, A.K. BOGDANOFF,

M.V. COVE, A.D. IRIZARRY, C.M. SCHALK, AND S.K. ARCHER. 2015. A primer on the history

of food web ecology: fundamental contributions of fourteen researchers. *Food Webs,* **4**:14–

24.

MAGUIRE, B.J. 1971. Phytotelmata: biota and community structure in plant held waters. *Ann.*

Rev. Ecol. Syst., **2**:439–464.

MAHOWALD, N., T.D. JICKELLS, A.R. BAKER, P. ARTAXO, C.R. BENITEZ-NELSON, G.

BERGAMETTI, T.C. BOND, Y. CHEN, D.D. COHEN, B. HERUT, N. KUBILAY, R. LOSNO, C. LUO,

W. MAENHAUT, K.A. MCGEE, G.S. OKIN, R.L. SIEFERT, AND S. TSUKUDA. 2008. Global

distribution of atmospheric phosphorus sources, concentrations and deposition rates, and

anthropogenic impacts. *Glob. Biogeochem. Cycles,* **22**: GB4026.

MCCANN, K.S. 2012. Food webs. Princeton University Press. 256 p.

MELLICHAMP, T.L. AND F.W. CASE. 2009. *Sarracenia*. p. 350–363. *In*: Flora of North America

Editorial Committee (ed.). Flora of North America north of Mexico volume 8. Oxford

University Press.

MILLER, T.E., L. HORTH, AND R.H. REEVES. 2002. Trophic interactions in the phytotelmata

communities of the pitcher plant, *Sarracenia purpurea. Comm. Ecol.,* **3**:109–116.

MOORHEAD, D.L. AND R.L. SINSABAUGH. 2006. A theoretical model of litter decay and microbial

interaction. *Ecol. Mono.,* **76**:151–174.

MOUQUET, N., T. DAUFRESNE, S.M. GRAY, AND T.E. MILLER. 2008. Modelling the relationship

between a pitcher plant (*Sarracenia purpurea*) and its phytotelma community: mutualism or

parasitism? *Func. Ecol.,* **22**:728–737.

NATIONAL ATMOSPHERIC DEPOSITION PROGRAM (NRSP–3). 2020. NADP Program Office,

Wisconsin State Laboratory of Hygiene, Madison, Wisconsin. nadp.slh.wisc.edu/NADP/

[date of access: 11 January 2020].

NE'EMAN, G., R. NE'EMAN, AND A.M. ELLISON. 2006. Limits to reproductive success of

Sarracenia purpurea (Sarraceniaceae). *Am. J. Bot.* **93**:1660–1666.

NORTHROP, A.C., R.K. BROOKS, A.M. ELLISON, N.J. GOTELLI, AND B.A. BALLIF. 2017.

Environmental proteomics reveals taxonomic and functional changes in an enriched aquatic

ecosystem. *Ecosphere* **8**:e01954.

OED ONLINE. 2019. "inquiline, n. (and adj.)". Oxford, UK: Oxford University Press.

www.oed.com/view/Entry/96507 [date of access: 14 January 2020].

PETERSON, C.N., S. DAY, B.E. WOLFE, A.M. ELLISON, R. KOLTER, AND A. PRINGLE. 2008. A

keystone predator controls bacterial diversity in the pitcher plant (*Sarracenia purpurea*)

microecosystem. *Env. Microbiol.,* **10**:2257–2266.

PILOSOF, S., M.A. PORTER, M. PASCUAL, AND S. KÉFI. 2017. The multilayer nature of ecological

networks. *Nature Ecol. Evol.* **1**:0101.

PIMM, S. 1982. Food webs. Chapman & Hall. 258 p.

PIOVIA–SCOTT, J., L.H. YANG, AND A.N. WRIGHT. 2017. Temporal variation in trophic cascades.

Ann. Rev. Ecol., Evol., Syst., **48**:281–300.

POISOT, T., E. CANARD, D. MOUILLOT, N. MOUQUET, AND D. GRAVEL. 2012. The dissimilarity of

species interaction networks. *Ecol. Lett.,* **15**:1353–1361.

POLIS, G.A. 1994. Food webs, trophic cascades and community structure. *Aust. J. Ecol.* **19**:121–

136.

R CORE TEAM. 2020. R: A Language and Environment for Statistical Computing. R Foundation

for Statistical Computing, Vienna. https://www.R-project.org.

SCHIAFFINO, M.R., N. DIOVISALVI, D. MARFETÁN MOLINA, P. FERMANI, C. LI PUMA, L.

LAGOMARSINO, M.V.QUIROGA, AND G.L. PÉREZ. 2019. Microbial food-web components in

two hypertrophic human-impacted Pampean shallow lakes: interactive effects of

environmental, hydrological, and temporal drivers. *Hydrobiologia* **830**:255–276.

SIROTA, J., B. BAISER, N.J. GOTELLI, AND A.M. ELLISON. 2013. Organic-matter loading

determines regime shifts and alternative states in an aquatic ecosystem. *Proc. Nat. Acad. Sci.,*

110:7742–7747.

SRIVASTAVA, D.S., J. KOLASA, J. BENGTSSON, A. GONZALEZ, S.P. LAWLER, T.E. MILLER, P.

MUNGUIA, T. ROMANUK, D.C. SCHNEIDER, AND M.K. TRZCINSKI. 2004. Are natural

microcosms useful model systems for ecology? *Trends Ecol. Evol.,* **19**:379–384.

THORNHAM, D.G., J.M. SMITH, T.U. GRAFE, AND W. FEDERLE. 2012. Setting the trap: cleaning

behaviour of *Camponotus schmitzi* ants increases long-term capture efficiency of their

pitcher plant host, *Nepenthes bicalcarata*. *Func. Ecol.,* **26**:11–19.

TIPPING, E., S. BENHAM, J.F. BOYLE, P. CROW, J. DAVIES, U. FISCHER, H. GUYATT, R.

HELLIWELL, L. JACKSON-BLAKE, A.J. LAWLOR, D.T. MONTEITH, E.C. ROWE, AND H.

TOBERMAN. 2014. Atmospheric deposition of phosphorus to land and freshwater. *Environ*

Sci.: Process. Impacts, **16**:1608–1617.

WAKEFIELD, A.E., N.J. GOTELLI, S.E. WITTMAN, AND A.M. ELLISON. 2005. The effect of prey

addition on nutrient stoichiometry, nutrient limitation, and morphology of the carnivorous

plant *Sarracenia purpurea* (Sarraceniaceae). *Ecology,* **86**:1737–1743.

WALLACE, R.L., T.W. SNELL, C. RICCI, AND T. NOGRADY. 2006. Rotifera. Volume 1: Biology,

ecology and systematics, 2nd edition. Backhuys Publishers, 299 p.

WOLKOVICH, E.M., S. ALLESINA, K.L. COTTINGHAM, J.C. MOORE, S.A. SANDIN, AND C. DE

MAZANCOURT. 2014. Linking the green and brown worlds:sthe prevalence and effect of

multichannel feeding in food webs. *Ecol. Lett.* **95**:3376–3386.

ZOU, K., E. THÉBAULT, G. LACROIX, AND S. BAROT. 2016. Interactions between the green and

brown food web determine ecosystem functioning. *Func. Ecol.* **30**:1454–1465.

Table 1 Values assigned to each inquiline taxon for calculation of a food-web saturation index.

Occurrence of taxa in a given food web is given by the sum of the binary values of all taxa

present. The saturation index is the sum of the decimal values of all taxa present. See Fig. 1 for

an illustrative example of these calculations

Taxon	Binary value	Decimal value
Metriocnemus	1	1
Habrotrocha	10	2
Sarraceniopus	100	4
Wyeomyia	1000	8
Fletcherimyia	10000	16

Table 2 Inquilines collected from pitchers in the four-level full factorial experiment in which

food webs were isolated from the plant with polyethylene tubes, inorganic and organic nutrients

were added to assess bottom-up effects, and the top predator *Fletcherimyia fletcheri* was added

to assess top-down effects. This table illustrates the effects on inquiline occurrence of the two

factors—the polyethylene tubes and the addition of the top predator—that had significant effects

on food-web saturation (Fig. 3). Values in each cell are the number of pitchers in which each

inquiline was collected, pooled over inorganic and organic nutrient addition treatments and both

harvests. The maximum possible number of occurrences for any given inquiline in a Tube ×

Fletcherimyia treatment combination is 32. Because cell entries within each column are

occurrences summed over each Tube × *Fletcherimyia* treatment combination, all entries within a

single row do not imply co-occurrences. Occurrences of the other macrobial inquilines within a

tube treatment were unrelated to the addition of *Fletcherimyia*

Tube treatment	*Fletcherimyia* treatment	Number of pitchers in which inquiline occurred				
		Metriocnemus	*Habrotrocha*	*Sarraceniopus*	*Wyeomyia*	*Fletcherimyia*
Intact	Added	27	6	15	13	1
	Not added	26	8	11	20	2
Perforated	Added	31	7	8	28	13
	Not added	29	6	12	28	10
None	Added	24	2	11	28	14
	Not added	28	1	6	24	6

a

b

Fig. 1 Complete, fully saturated inquiline food web of *Sarracenia purpurea* (**a**; modified from

Baiser *et al.*, 2013) and an example of incomplete, unsaturated one (**b**). Values adjacent to each

of the macrobial taxa are those used to compute food-web saturation (see Table 1). Prey,

bacteria, protozoa, and particulate organic matter were assumed to occur in all sampled food

webs, and each of the possible combinations of macrobes gives a unique saturation value. In

these two examples, the saturation value of the complete web (a) = 31, whereas that of the food

web lacking *Fletcherimyia* and *Sarraceniopus* (b) = 11

Fig. 2 Response surface of food-web saturation (defined in Table 1) in pitchers fed one of 25

combinations of nitrogen (N) and phosphorus (P) or distilled water once, three days after the

pitcher opened (the "pulse" experiment; left panel); or three days after the pitcher opened and

every two weeks thereafter (the "press" experiment; right panel). This figure illustrates data from

the first leaf of the season (fed on or beginning the week of 4 June 2001) at the mid-summer

540   harvest (6 August 2001 for the pulse experiment, 23 July 2001 for the press experiment), when

541   saturation was highest

Fig. 3 Saturation in the *Sarracenia purpurea* macrobial food webs in the four-level tube ×

inorganic nitrogen × fruit-fly × *Fletcherimyia* manipulation. The right panel shows the effects of

the plant itself on the food web, whereas the left panel shows the effects of the top predator,

Fletcherimyia, on the food web. Bars are means ± one standard deviation of the mean of data

from all leaves and both harvests pooled over all other treatments.