Regulation by the Pitcher Plant Sarracenia purpurea of the Structure of its Inquiline Food

Web

AARON M. ELLISON[1]

Harvard University, Harvard Forest, 324 North Main Street, Petersham, Massachusetts 01366

NICHOLAS J. GOTELLI

University of Vermont, Department of Biology, Burlington, Vermont 04505

LESZEK A. BŁĘDZKI

Mount Holyoke College, Department of Biology, South Hadley, Massachusetts 01075

AND

JESSICA L. BUTLER[2]

Harvard University, Harvard Forest, 324 North Main Street, Petersham, Massachusetts 01366

[1] Corresponding author: Telephone: (978)-756-6178; E-mail: aellison@fas.harvard.edu
[2] Present Address: Barr Engineering Company, Minneapolis, Minnesota 55435

ABSTRACT.—Phytotelmata, the water-filled habitats in pitcher plants, bromeliad tanks,

and tree-holes, host multitrophic food webs that are model experimental systems for studying

food-web structure and dynamics. However, the plant usually is considered simply as an inert

container, not as an interacting part of the food web. We used a manipulative field experiment

with a response-surface design to determine effects of nutrient enrichment (multiple levels of

NH_4NO_3, PO_4, and captured prey), top predator (removed or present), and the plant itself (with

or without plastic tubes inserted into the pitchers to isolate the food web from the plant) on the

macrobial food web within the modified leaves ("pitchers") of the carnivorous pitcher plant

Sarracenia purpurea. Connection to the plant, addition of NH_4NO_3 and removal of the top

predator significantly increased the food web's saturation, defined as its trophic depth and

number of interactions. No effects on food-web saturation resulted from addition of PO_4 or

supplemental prey. Plants such as *S. purpurea* that create phytotelmata are more than inert

containers and their inhabitants are more than commensal inquilines. Rather, both the plant and

the inquilines are partners in a complex network of interactions.

INTRODUCTION

Understanding controls on food web structure has been an organizing focus for

community ecology for well over a century (*e.g.,* Pimm, 1982; Polis, 1994; Carpenter *et al.,*

2001; McCann, 2012; Wokovich *et al.* 2014; Layman *et al.*, 2015; Zou *et al.*, 2016). As primary

producers, plants are the bottom-up controllers of "green" food webs, whereas as "detritus" the

decomposing leaf litter of plants is the resource base for "brown" food webs (Cebrian and

Lartigue, 2004). The bottom-up roles of plants as producers of fresh biomass and necromass in

structuring, respectively green and brown food webs, have been studied extensively, but other

roles for plants in structuring food webs through habitat creation and indirect interactions is

much less appreciated. For example, in tree holes and modified leaves, plants create habitats for

a range of aquatic food webs. The complex, mostly detritus-based food webs in these

"phytotelmata"—water-filled habitats in, for example, tree holes, bromeliads, and pitcher plants

(Maguire, 1971)—have been used as model experimental systems for studying the dynamics of

aquatic food webs (*e.g.,* Kitching, 2000, 2001; Kneitel and Miller, 2002; Miller *et al.,* 2002;

Ellison *et al.,* 2003; Srivastava *et al.,* 2004). However, the phytotelmata itself normally is

regarded simply as a container for its "inquilines": animals that live commensally "within the

nest or abode of another" (OED, 2019). It has been recognized only rarely that the plant itself

can mediate trophic interactions of inquiline food webs.

Research with carnivorous pitcher plants has provided two notable examples of the

importance of phytotelmata on inquiline food webs. Bradshaw and Creelman (1984)

experimentally showed that the carnivorous pitcher plant *Sarracenia purpurea* L.

(Sarraceniaceae) and two of its obligate inquilines, larvae of the pitcher-plant mosquito

Wyeomyia smithii (Coq.) and the pitcher-plant midge *Metriocnemus knabi* Coq., are mutualists.

These two inquilines accelerate the breakdown and decomposition of arthropod prey captured by

S. purpurea (Heard, 1994). Ammonium (NH_4) and carbon dioxide (CO_2) released as bacteria

mineralize nutrients in the prey (Butler *et al.*, 2008) and NO_3, NH_4, and PO_4 regenerated by

rotifers (Błędzki and Ellison, 1998, 2002; Błędzki *et al.*, 2018) are absorbed by the plant and

used for photosynthesis, subsequent growth, and reproduction (Bradshaw and Creelman, 1984;

Joel and Gepstein, 1985; Butler and Ellison, 2007). The plant, in turn, oxygenates the pitcher

fluid (Bradshaw and Creelman, 1984; Joel and Gepstein, 1985; Sirota *et al.*, 2013), which

maintains homeostatic conditions for the inquiline food web and prevents a switch to anaerobic

conditions within the phytotelmata, which can nonetheless occur if prey are superabundant

(Krieger and Kourtev, 2012; Sirota *et al.*, 2013; Northrop *et al.*, 2017).

Similarly, one of the Asian pitcher plants, *Nepenthes bicalcarata* Hook.f. (Nepenthaceae)

hosts the ant *Colobopsis schmitzi* (Stärcke) (Formicidae) in the plant's hollow tendrils (Clarke

and Kitching, 1995). The ants move out of the tendrils to forage within the pitchers on large prey

that the plant captures. The ants also clean the pitcher's peristome, reducing fungal colonization

and increasing prey-capture efficiency (Thornham *et al.*, 2011), which in turn leads to greater

nutrient acquisition, photosynthesis, and growth of this perennial plant (Bazile *et al.*, 2012).

The lack of attention to linkages between food-web structure or dynamics and the habitat

in which the food web itself assembles is not restricted to phytotelmata. Although spatiotemporal

dynamics of food webs have been studied, documented, and modeled (*e.g.*, Poisot *et al.*, 2015;

Pilosof *et al.*, 2017; Piovia-Scott et al. 2017; Hutchinson et al., 2019; Schiaffino *et al.*, 2019),

how complete food webs assemble in a habitat that itself is changing through time and interacts

with the food web itself remains unexplored. We are unsure whether this lack of attention results

from the rarity of such situations, our failure to perceive them, or the difficulty in studying them.

However, living phytotelmata such as pitcher plants provide opportunities to study

experimentally the dynamics of a food web that assembles in a habitat that itself is a living,

growing organism. The experiments we describe here were designed to test whether pitcher

plants are more than simply "containers" for "inquilines" and how the interactions between

plants, the habitat it creates, and the food web that assembles within it change through time.

Here, we describe how nutrient enrichment, top predators, and the living habitat of the

plant itself affect the structure of the four trophic-level macrobial food web that inhabits the

modified pitcher-shaped leaves (Arber, 1941) of *S. purpurea*. In a set of controlled field

experiments, we first examined the impact of nutrient deposition on the food web that assembles

in *S. purpurea* pitchers. Second, we severed the connection between the plant and its food web

by inserting closed plastic tubes (or appropriate controls) in between pitchers and their food webs

to determine if effects of experimental nutrient enrichment through additions of prey and

inorganic nutrients resulted from direct effects on the food web or indirect effects mediated by

the plant. We recognize that a focus on the macrobes will overlook many important processes

and interactions of the microbes present in *S. purpurea* food webs. This is a general issue with

studies of food webs (e.g., Pringle and Hutchinson, 2020) that will be able to be addressed as the

microbial assemblages of *S. purpurea* continue to be better characterized (e.g., Peterson *et al.*

2008, Miller and terHorst 2012, Paise *et al.* 2014, Canter *et al.* 2018, Boynton *et al.* 2019, Zhang

et al. 2020a, 2020b).

95   **MATERIALS AND METHODS**

96   STUDY SPECIES

Sarracenia purpurea (the northern, or purple, pitcher plant) is a long-lived (> 50 yr),
rosette-forming, carnivorous perennial plant common to bogs throughout Canada and the eastern
United States. Pitcher plants flower early in the growing season (May in Massachusetts) before
any new leaves are produced; flower buds for the following season are set in August –
September (Shreve 1906). Pitcher plants have weakly developed root systems; nutrients are
obtained principally from insect prey attracted to the brightly colored pitchers and extrafloral
nectaries (Bennett and Ellison, 2009). Unlike the other 10 species of Sarracenia (Mellichamp
and Case, 2009), S. purpurea pitchers fill with rainwater and these semi-permanent pools
(phytotelmata) are colonized by a suite of specialized invertebrates and more generalist protists
and bacteria (Addicott, 1974; Cochran-Stafira and von Ende, 1998; Błędzki and Ellison, 2003;
Ellison et al., 2003, Peterson et al. 2008, Miller and terHorst 2012, Paise et al. 2014, Canter et
al. 2018, Boynton et al. 2019, Zhang et al. 2020a, 2020b).

Arthropod prey captured by the plant is the basal resource for this detritus-based food
web, which shreds the prey and mineralizes the available nutrients (Butler and Ellison, 2007;
Butler et al., 2008). Prey capture initially increases as pitchers open, expand, and harden; reaches
a peak when the leaves are two–three weeks old; and then declines during to near zero by the end
of the growing season (Fish and Hall, 1978; Ellison et al., 2003). Drowned carcasses of captured
prey (primarily ants and flies; Ellison and Gotelli, 2009) are shredded by larvae of the pitcher-
plant midge Metriocnemus knabi and larvae of the sarcophagid fly Fletcherimyia fletcheri
(Aldrich); the shredded prey are broken down further by bacteria (Cochran-Stafira and von Ende,
1998; Heard, 1994; Baiser et al., 2011). The bacteria, organic matter, protozoa, and algae in turn
are consumed by the bdelloid rotifer Habrotrocha rosa Donner and a variety of protozoa, which,
along with the pitcher-plant mite Sarraceniopus gibsoni (Nesbitt), also consume detritus directly

(Wallace *et al.,* 2006). Protozoa also consume bacteria (Cochran-Stafira and von Ende, 1998).

These primary consumers are preyed on by larvae of the pitcher-plant mosquito *W. smithii* (all

these macrobes will be referred to below using their generic epithets). Late instars of

Fletcherimyia also prey on rotifers and 1st and 2nd instars of *Wyeomyia* (Błędzki and Ellison,

1998; Butler *et al.*, 2008; Baiser *et al.*, 2013). Not only does *Fletcherimyia* complete its larval

development within pitchers, but the adult flies are pollinators of *S. purpurea* (Ne'eman *et al.*,

2006).

STUDY SITE

The experiments reported here were conducted at Swift River Bog, a 1.9-ha glacial kettle

bog in Belchertown, Massachusetts, USA (42.27 °N, −72.34 °W, 124 m a.s.l.; Kearsley, 1999).

This bog has a lagg (moat) around it that never exceeds 2 m in width, but there is no open water

within the bog itself. Common *Sphagnum* species present at Swift River Bog were *Sp.*

angustifolium (C.E.O. Jensen ex Russow) C.E.O. Jensen*, Sp. magellanicum* Brid., and *Sp.*

rubellum Wilson (Ellison and Gotelli, 2016). The dominant vascular plants growing on the bog

mat were shrubs in the Ericaceae: *Chamaedaphne calyculata* (L.) Moench (leatherleaf)*, Kalmia*

polifolia Wangenheim (bog laurel)*, K. angustifolia* L. (sheep laurel)*, Rhododendron viscosum*

(L.) Torrey (swamp rhododendron) and *Vaccinium macrocarpon* Aiton (cranberry); scattered

Picea mariana (Miller) Britton (Pinaceae; black spruce) grow across the bog. The site has a large

population (1000s of plants) of *S. purpurea* (Ne'eman *et al.,* 2006; Ellison and Gotelli, 2016).

Total nitrogen deposition ($NO_3 + NH_4$) at the National Atmospheric Deposition Program

(NADP) Quabbin Reservoir monitoring station in the same watershed (MA08: 42.3925 °N,

−72.3444 °W) ranged from 1.25–1.60 mg/L during the years of this study (2001 – 2004), and has

averaged 1.19 (range: 0.58–2.12) mg/L since measurements began in 1982 (National

Atmospheric Deposition Program, 2020). Phosphorus deposition is not monitored by NADP or

other agencies in the US, but P deposition may have significant effects in P-limited ecosystems

or alter stoichiometric ratios in otherwise N-limited ones (Mahowald *et al.*, 2008; Tipping *et al.*,

2014). Although *S. purpurea* is stoichiometrically N- or N+P-limited (Ellison, 2006), field

experiments have suggested that *S. purpurea* preferentially takes up P from supplemental prey

(Wakefield *et al.*, 2005).

151   FOOD-WEB SATURATION: DEFINITION OF THE RESPONSE VARIABLE

In all the experiments described below, *food-web saturation* was our response variable.

We defined food-web saturation as follows: Each macrobial taxon (rotifer, mite, midge,

mosquito, sarcophagid fly) in the food web was assigned a binary value representing its presence

(Table 1; Fig. 1); we assumed all food webs had protozoa and bacteria. The saturation of the

food web in a pitcher was defined as the sum of the equivalent decimal values of each macrobial

taxon present. There are 32 possible food webs that can be assembled from these five taxa; the

decimal value for each food web ranges from 0 (no macrobial taxon present) to 31 (all macrobial

taxa present), with increasing numbers indicating more "saturated" food webs. Food webs with

higher saturation values had both more trophic levels present and more trophic links present (Fig.

1), but the food-web saturation index does not reflect any particular trophic relationship. Further,

the index can be easily extended if any other taxa (e.g., microbes) were to be included.

164   EXPERIMENT 1 – IMPACTS OF N AND P ON FOOD-WEB SATURATION

The first experiment investigated the effects of inorganic N and P (as would be

accumulated in the phytotelmata by atmospheric deposition) on food-web saturation in situ. We

used a factorial response-surface design (Cottingham *et al.*, 2005) in a "pulse" experiment (sensu

Bender *et al.*, 1984) in which 5 levels each of NH_4NO_3 (4.687, 9.375, 18.75, 37.5, and 75.0 mM)

and PO_4 (0.625, 1.25, 2.5, 5.0, and 10.0 mM) and controls (distilled, deionized water: dH_2O)

were added biweekly to each pitcher of 130 non-flowering plants that were randomly assigned to

one of five groups of 26 plants. The levels of N used bracketed the observed deposition of N

measured by NADP in the years before the study, whereas the levels of P were chosen to span

the N:P ratios previously used by Ellison and Gotelli (2002) in other nutrient addition

experiments with *S. purpurea* without any evidence of nutrient toxicity (*sensu* Camargo *et al.*

2005).

Plants were located at least 10 m (linear distance) from the lagg surrounding the bog,

separated from one another by at least 50 cm, and atop the *Sphagnum* mat but not under shrubs

or trees. Each plant within a group was randomly assigned a nutrient treatment (one of the 25

possible N × P combinations or the dH_2O control). On 4 June 2001, 20 ml of nutrient solution

was added once to each open pitcher on each plant. Plants were monitored every three d for new

pitcher production, and nutrient solutions were added (again, only once) to each new pitcher 2-3

days after it had opened and fully hardened. Every 3 wk (25 June, 16 July, 6 August, 27 August,

and 17 September), a complete set of 26 plants was harvested. At each harvest, pitcher fluid was

pipetted into sterile tubes (one tube/leaf), the leaves further rinsed to ensure that all rotifers and

mites were collected, and the presence or absence of each member of the food web was

determined.

We ran a parallel "press" experiment (sensu Bender *et al.*, 1984) in which we

haphazardly located 26 non-flowering plants and randomly assigned each of them to one of the

same 26 nutrient treatments as in the pulse experiment. 2-3 d after each pitcher had opened and

hardened, 20 ml of nutrient solution was added to it, and the 20-ml level was marked on the side

of the pitcher with indelible ink. Every two weeks, the food web was pipetted out of each pitcher

and into a petri dish; the volume of the flued was recorded. Presence of each macrobe was

determined in the field, and all were returned to the pitcher. Rotifers and protozoa were counted

in the lab in 1-ml subsamples taken at the same time; total counts of rotifers and protozoa were

extrapolated to the full volume by simple multiplication. After field counts of invertebrates, the

pitcher fluid was poured back into the pitcher and additional nutrient solution was added to

maintain the fluid level at 20 ml. This experiment ran from 4 June–17 September 2001.

EXPERIMENT 2 – INTERACTIONS BETWEEN THE PLANT, NUTRIENTS, AND PREDATION ON FOOD-WEB

200   STRUCTURE

The second experiment investigated how changes in food-web saturation resulted from

interactions between bottom-up effects of nutrient addition, top-down effects of predation, and

effects of the plant itself. We used a four-level factorial/response-surface experiment with 212

non-flowering plants located at least 10 m (linear distance) from the lagg surrounding the bog,

separated from one another by at least 50 cm, and atop the *Sphagnum* mat but not under shrubs

or trees. As in Experiment 1, plants were assigned randomly to unique combinations of

treatments that included physically isolating the inquiline food web within the phytotelmata.

The first treatment (factor) assessed the influence of the plant itself on the food web and

had three levels. Pitcher interiors were: [1] fitted with intact polyethylene tubes (2.2-cm

diameter) (inquilines and their fluid isolated from effects of pitchers; n = 64 plants, 1

pitcher/plant); [2] fitted with perforated (ten 6.35-mm diameter drilled holes) polyethylene tubes

(polyethylene tube control; n = 64 plants); or [3] left without polyethylene tubes (unmanipulated

control; n = 64 plants).

The second treatment (ordinal factor used in the response surface) examined the effects

of N deposition. One of eight different concentrations of inorganic nitrogen (NH_4NO_3: 0, 1.56,

3.1, 6.25, 12.5, 25, 50, or 100 mg N/L) was added to a randomly selected plant; there were n = 8

replicates of each N concentration in each set of 64 tube/tube-control/no tube treatments.

Because we assumed a constant uptake of NH_4NO_3 by pitcher tissue in control and tube-control

pitchers, and volatilization and uptake by bacteria in all pitchers, we replaced fluid and nutrients

with the appropriate NH_4NO_3 solution every two weeks.

The third treatment (factor) examined effects of prey availability (organic nutrients).

Supplemental prey (10 lab-raised fruit flies/wk) were added to one-half of each inorganic N ×

tube combination.

Finally, because of the importance of *Fletcherimyia* at multiple points in this food web

(Błędzki and Ellison, 1998; Butler *et al.*, 2008; Baiser *et al.*, 2013), the fourth treatment (factor)

assessed the influence of this top predator on the development of the pitcher-plant food web in

the context of bottom-up effects of prey and inorganic N addition and habitat effects of the

pitcher itself. Second instar *Fletcherimyia* larvae were either removed from pitchers (one-half of

each N × tube × fruit-fly treatment combination) or added to pitchers (one-half of each N × tube

× fruit-fly treatment combination).

In total, this factorial/response-surface design resulted in $N = 2$ plants in each tube × N ×

fruit-fly × *Fletcherimyia* treatment combination. We also haphazardly chose 20 additional plants

growing nearby in the bog as unmanipulated controls. One treated plant from each treatment

combination (96 pitchers) and half the unmanipulated controls were harvested after 4 wk, and the

remainder after 8 wk. Food-web saturation in this experiment was computed based on all

inquilines present including *Fletcherimyia*, except in the *Fletcherimyia* addition treatments (to

avoid simply documenting the addition in our calculation).

239   STATISTICAL ANALYSIS AND DATA AVAILABILITY

Data were analyzed using standard linear regression, analysis of variance, or analysis of

covariance using functions for generalized linear models (glm) in the base and stats packages of

the R statistical software system, version 3.6.3 (R Core Team 2020).

Experiment 1 (pulse and press experiments) were analyzed using a linear regression

model (i.e., linear in the regression parameters) with which we fit a polynomial surface of food-

web saturation as function of N and P concentrations and their interaction. Experiment 2 (four

treatment factorial/response-surface design) was analyzed using a glm with main effects of the

tubes (3 levels), supplemental prey (2 levels), *Fletcherimyia* addition (2 levels), inorganic N

addition (8 ordered levels treated as a continuous variable in the model), and all of their two- or

three-way interactions. Initial analyses revealed no significant interaction effects among the three

discrete factors ($P > 0.20$ all cases), so we report only the results of the main effects and the

interactions with the inorganic N additions. Throughout, F statistics and associated P values are

based on Type III sums of squares.

All data from this experiment are available from the Harvard Forest Data Archive, dataset

HF193 (https://harvardforest.fas.harvard.edu/exist/apps/datasets/showData.html?id=hf193) and

the Environmental Data Initiative

(https://dx.doi.org/doi:10.6073/pasta/3bd2c920cf431442d34227f3f5fc7c2a).

258   RESULTS

259   EXPERIMENT 1

In the pulse experiment, which established the baseline response of the inquiline food

web to bottom-up control by inorganic nutrient additions, food-web saturation peaked at

intermediate levels of N added ($F_{1,227} = 10.7$. $P < 0.01$), was significantly higher in pitchers

produced earlier in the season ($F_{1,227} = 5.0$, $P = 0.03$), and in leaves harvested in mid-summer

($F_{1,227} = 4.4$, $P = 0.04$) (Fig. 2). Although the amount of P added did not alter food web saturation

($F_{1,227} = 0.7$, $P = 0.42$), there was a significant N × P interaction ($F_{1,227} = 4.4$, $P = 0.04$).

Food-web saturation in the coincident press experiment also peaked at intermediate levels

of N added ($F_{1,292} = 3.7$, $P = 0.05$) and was highest in mid-summer ($F_{1,292} = 26.9$, $P < 0.01$) in

those pitchers that were produced earliest in the season ($F_{1,203} = 4.1$, $P = 0.04$) (Fig. 2). As in the

pulse experiment, the amount of P added did not alter food web saturation ($F_{1,292} = 0.4$, $P =$

0.54), but in contrast to the results of the pulse experiment, there was no significant N × P

interaction in the press experiment ($F_{1,292} = 0.3$, $P = 0.58$). We did not observe any pitcher

mortality from the higher levels of N additions.

274   EXPERIMENT 2

Inquilines colonized all the plants, but *Wyeomyia* was observed somewhat less frequently

in pitchers with intact tubes than in pitchers with perforated tubes or no tubes at all ($\chi^2_{[2]} = 28.5$,

$P < 0.01$; Table 2). Similarly, *Fletcherimyia* colonized only two pitchers with intact tubes and the

added *Fletcherimyia* survived in only one pitcher with an intact tube ($\chi^2_{[1]} = 20.6$, P < 0.01;

Table 2). Occurrences of the other macrobial inquilines were unrelated to tube treatment or

Fletcherimyia addition (respectively $\chi^2_{[2]} = 3.4$, P = 0.18 and $\chi^2_{[1]} = 0.03$, P = 0.87; Table 2).

Connections with the plant increased food-web saturation: food webs in pitchers without

tubes or with perforated tubes were significantly more saturated than food webs in pitchers with

intact tubes (Fig. 3, right panel; $F_{2,173} = 8.4$, P < 0.01). Removal of *Fletcherimyia* from plants led

to an increase in food web saturation (Fig. 3, left panel; $F_{1,173} = 13.2$, P < 0.01), as this predator

consumes both rotifers and *Wyeomyia* larvae. Neither additions of prey nor inorganic nitrogen

(NH_4NO_3-N) alone altered food-web saturation ($F_{1,173} = 0.8$, P = 0.37; $F_{1,173} = 0.0$, P = 0.88,

respectively). However, there was a nutrient × *Fletcherimyia* interaction ($F_{1,182} = 4.9$, P = 0.03):

in the presence of *Fletcherimyia*, increasing NH_4NO_3 led to decreased saturation of the food

web. In contrast, in the absence of *Fletcherimyia*, increasing NH_4NO_3 led to increased saturation

of the food web. None of the other two-way, three-way, or four-way interaction terms was

significant (P > 0.18, all cases).

293   DISCUSSION

Plant productivity, expressed as new biomass eaten by herbivores or shed litter

decomposed by detritivores, exerts bottom-up control of most food webs (McCann 2012;

Wokovich *et al.* 2014; Zou *et al.*, 2016). As phytotelmata (Maguire, 1971), plants produce

habitat for aquatic food webs (Kitching, 2000, 2001; Ellison *et al.,* 2003; Srivastava *et al.,* 2004),

but other effects of phytotelmata on food webs have been studied only rarely (Bradshaw and

Creelman, 1984; Joel and Gepstein, 1985; Clarke and Kitching, 1995; Thornham *et al.*, 2011;

Bazile *et al.*, 2012). The experiments described here illustrate that the phytotelmata of northern

pitcher plants are not simply inert containers in which complex aquatic food webs assemble and

thrive. Rather, the plant itself interacts with, and determines the strength of, the bottom-up and

top-down controls of the food web.

Unlike pitchers in other species of *Sarracenia* that are covered by a reflexed 'hood',

pitchers of *S. purpurea* are open to the sky and accumulate water from precipitation. This has

two consequences. First, as pitchers accumulate water, they create the habitat for the inquiline

food web. Second, any nutrients that accompany precipitation can enter the food web through

bacterial mineralization and immobilization.

Independent of addition of inorganic nutrients, supplemental prey, or presence of top

predators, the plant itself had a net positive effect on inquiline food-web saturation. Food webs in

containers isolated from the plant were 10–20% less saturated than food webs in perforated tubes

or unmanipulated controls of comparable size. This result agrees with previous observations of,

and experiments with, *S. purpurea* and its inquilines, in which the plant oxygenated the pitcher

fluid, maintaining homeostatic conditions for the inquiline food web and preventing a switch to

anaerobic conditions (Bradshaw and Creelman, 1984; Joel and Gepstein, 1985; Sirota *et al.*,

2013).

Further addition of inorganic N, as is occurring through atmospheric deposition,

increased food-web saturation at intermediate levels, but saturation declined as N addition was

increased further (Fig. 2). We hypothesize that this result may reflect bottom-up effects of N on

microbial productivity (Moorhead and Sinsabaugh, 2006), shifts in how *S. purpurea* uses

different forms of N (Karagatzides *et al.*, 2009), or interactions between the plant and the

bacteria that mineralize nitrogen for it (Butler *et al.*, 2008; Mouquet *et al.*, 2008). These studies,

and others with excess prey addition show shifts in the macrobial food web or bacterial

functional groups (see Northrop *et al.* 2021 for the latter) but no evidence of nutrient toxicity

(*sensu* Camargo *et al.* 2005). Furthermore, N addition alters pitcher morphology, reducing

pitcher size (volume) while increasing the area of its photosynthetic keel (Ellison and Gotelli,

2002; Bott *et al.*, 2008). Although smaller pitchers have less complex food webs (Gotelli and

Ellison, 2006), this effect was not observed or measured here because we worked only with

pitchers that were fully opened, not those that were produced after nutrients were added to early-

season pitchers and that would have expressed morphological differences following nutrient

translocation and use in new pitcher production (Butler and Ellison, 2007).

Top-down effects of predation in the inquiline food web of *S. purpurea* have been

described repeatedly (*e.g.,* Addicott, 1974; Kneitel and Miller, 2002; Hoekman, 2007, 2011;

Peterson *et al.*, 2008), but in our experiments, these effects were manifest only when the food

web was isolated from the plant. Food webs in tubes rarely had predators (*Wyeomyia* or

Fletcherimyia), as these Diptera would not oviposit in intact tubes and food webs without the

predator *Fletcherimyia* were more saturated (up to the level below *Fletcherimyia*) than food

webs with this predator. The interactions that we observed between top-down effects of

Fletcherimyia and bottom-up effects of inorganic N additions are mirrored by modelled trade-

offs in overall system nitrogen mineralization modulated by interactions between plants and

mineralizing bacteria and between plants and higher trophic levels (Mouquet *et al.*, 2008; Ellison

and Gotelli, 2021). Finally, the potential importance of digestive enzymes in mediating any of

these produced by the plant remains unknown. When digestive enzymes are produced during

pitcher development and at what concentrations remain open questions (*e.g.,* Gallie and Chang,

1977; Luciano and Newell, 2017). However, our results do illustrate that the pitcher itself does

more than simply maintain a habitat for symbionts (*cf.* Luciano and Newell, 2017).

Overall, the results presented here together with those from other studies on pitcher-plant food webs illustrate that top-down and bottom-up effects on food-web structure are dependent on the habitat context in which they occur. Anthropogenic processes such as fossil-fuel combustion leading to atmospheric deposition of inorganic nutrients can alter habitat structure and shift control of food web structure from strictly bottom-up control to a mixture of top-down and bottom-up controls. These consequences are especially evident in habitats that themselves are living structures that respond to the same forcing processes at similar time scales. Plants such as *S. purpurea* that create phytotelmata are more than inert containers and their inhabitants are more than commensal inquilines. Rather, both the plant and the 'inquilines' are partners in complex networks of interactions. Our work and that of others illustrate clearly that the microecosystem created by *S. purpurea* can provide a model experimental system for exploring such interactions (Ellison and Gotelli, 2021).

Acknowledgement.—We thank Clarisse Hart, Callan Ordoyne, and Ali Rosenberg for significant assistance with the fieldwork, and two reviewers for excellent comments on an early version of the ms. This research was supported by NSF grants DEB 98-05722, 98-08504, 02-35128, and 02-34710 to AME, and permitted by the Massachusetts Department of Hatcheries.

LITERATURE CITED

ADDICOTT, J.F. 1974. Predation and prey community structure: an experimental study of the effect of mosquito larvae on the protozoan communities of pitcher plants. *Ecology*, **55**:475–492.

ARBER, A. 1941. On the morphology of the pitcher-leaves in *Heliamphora, Sarracenia, Darlingtonia, Cephalotus*, and *Nepenthes*. *Ann. Bot.,* **5**:563–578.

BAISER, B., R. ARDESHIRI, AND A.M. ELLISON. 2011. Species richness and trophic diversity increase decomposition in a co–evolved food web. *PLoS ONE,* **6**:e20672.

———, H.L. BUCKLEY, N.J. GOTELLI, AND A.M. ELLISON. 2013. Predicting food-web structure with metacommunity models. *Oikos,* **122**:492–506.

BAZILE, V., J.A. MORAN, G. LE MOGUEDEC, D.J. MARSHALL, AND L. GAUME. 2012. A carnivorous plant fed by its ant symbiont: a unique multi–faceted nutritional mutualism. *PLoS ONE,* **7**:e36179.

BENDER, E.A., T.J. CASE, AND M.E. GILPIN. 1984. Perturbation experiments in community ecology: theory and practice. *Ecology,* **65**:1–13.

BŁĘDZKI, L.A. AND A. M. ELLISON. 1998. Population growth and production of *Habrotrocha rosa* Donner (Rotifera: Bdelloidea) and its contribution to the nutrient supply of its host, the northern pitcher plant, *Sarracenia purpurea* L. (Sarraceniaceae). *Hydrobiologia,* **385**:193–200.

——— AND ———. 2002. Nutrient regeneration by rotifers in new England (USA) bogs. *Verh. Int. Ver. Limnol.,* **28**:1328–1331.

——— AND ———. 2003. Diversity of rotifers from Northeastern USA bogs with new species records for North America and New England. *Hydrobiologia*, **497**:53–62.

———, J.L. BUBIER, A.M. ELLISON, AND T.R. MOORE. 2018. Ecology of rotifers and their unappreciated source of nitrogen and phosphorus in temperate northeastern American bogs. *Fundam. Appl. Limnol.,* **191**:277–287.

BOTT, T., G.A. MEYER, AND E.B. YOUNG. 2008. Nutrient limitation and morphological plasticity of the carnivorous pitcher plant *Sarracenia purpurea* in contrasting wetland environments. *New Phytol.,* **180**:631–641.

BOYNTON, P., C.N. PETERSON, AND A. PRINGLE. 2019. Superior dispersal ability can lead to persistent ecological dominance throughout succession. *Appl. Env. Microbiol.*, **85**:e02421–18.

BRADSHAW, W.E. AND R. A. CREELMAN. 1984. Mutualism between the carnivorous purple pitcher plant *Sarracenia purpurea* and its inhabitants. *Am. Midl. Nat.,* **112**:294–304.

BUTLER, J.L. AND A.M. ELLISON. 2007. Nitrogen cycling dynamics in the carnivorous northern pitcher plant, *Sarracenia purpurea. Func. Ecol.,* **21**:835–843.

———, N. J. GOTELLI, AND A. M. ELLISON. 2008. Linking the brown and green: nutrient transformation and fate in the *Sarracenia* microecosystem. *Ecology,* **89**:898–904.

CANTER, E.J., C. CUELLAR-GEMPELER, A.I. PASTORE, T.E. MILLER, AND O.U. MASON. 2018. Predator identity more than predator richness structures aquatic microbial assemblages in *Sarracenia purpurea* leaves. *Ecology,* **99**:652–660.

CAMARGO, J.A., A. ALONSO, AND A. SALAMANCA. 2005. Nitrate toxicity to aquatic animals: a review with new data for freshwater invertebrates. *Chemosphere*, **58**:1255–1267.

CARPENTER, S.R., J.J. COLE, J.R. HODGSON, J.F. KITCHELL, M.L. PACE, D. BADE, K.L. COTTINGHAM, T.E. ESSINGTON, J.N. HOUSER, AND D.D. SCHINDLER. 2001. Trophic cascades, nutrients, and lake productivity: whole-lake experiments. *Ecol. Mono.,* **71**:163–186.

CEBRIAN, J. AND J. LARTIGUE. 2004. Patterns of herbivory and decomposition in aquatic and

terrestrial ecosystems. *Ecol. Mono.,* **74**:237–259.

CLARKE, C.M. AND R.L. KITCHING. 1995. Swimming ants and pitcher plants: a unique ant-plant

interaction from Borneo. *J. Trop. Ecol.,* **11**:589–602.

COCHRAN-STAFIRA, D.L. AND C.N. VON ENDE. 1998. Integrating bacteria into food webs: studies

with *Sarrcenia purpurea* inquilines. *Ecology,* **79**:880–898.

COTTINGHAM, K.L., J.T. LENNON, AND B.L. BROWN. 2005. Knowing when to draw the line:

designing more informative ecological experiments. *Front. Ecol. Env.,* **3**:145–152.

ELLISON, A.M. 2006. Nutrient limitation and stoichiometry of carnivorous plants. *Plant Biol.,*

8:740–747.

——— AND N.J. GOTELLI. 2002. Nitrogen availability alters the expression of carnivory in the

northern pitcher plant *Sarracenia purpurea*. *Proc. Nat. Acad. Sci.,* **99**:4409–4412.

——— AND ———. 2009. Energetics and the evolution of carnivorous plants – Darwin's "most

wonderful plants in the world." *J. Exp. Bot.,* **60**:19–42.

——— AND ———. 2016. Moths, ants and pitcher plants in bogs at Harvard Forest and

Belchertown MA 2007–2013. Harvard Forest Data Archive: HF159.

dx.doi.org/10.6073/pasta/37ee70bbf38b3f95bc955dc555c63188 [date of access: 12 January

2020].

——— AND ———. 2021. *Scaling in Ecology with a Model System*. Princeton, NJ, USA:

Princeton University Press,

———, ———, J.S. BREWER, L. COCHRAN-STAFIRA, J. KNEITEL, T.E. MILLER, A.S. WORLEY,

AND R. ZAMORA. 2003. The evolutionary ecology of carnivorous plants. *Adv. Ecol. Res.,*

33:1–74.

FISH D. AND D.W. HALL. 1978. Succession and stratification of aquatic insects inhabiting the

leaves of the insectivorous pitcher plant *Sarracenia purpurea*. *Am. Midl. Nat.,* **99**:172–183.

GALLIE, D.R. AND S.-C. CHANG. 1997. Signal transduction in the carnivorous plant *Sarracenia*

purpurea. *Plant Physiol.*, **115**:1461–1471.

GOTELLI, N.J. AND A.M. ELLISON. 2006. Food-web models predict species abundance in

response to habitat change. *PLoS Bio.,* **44**:e324.

HEARD, S.B. 1994. Pitcher-plant midges and mosquitoes: a processing chain commensalism.

Ecology, **75**:1647–1660.

HOEKMAN, D. 2007. Top-down and bottom-up regulation in a detritus-based aquatic food web: a

repeated field experiment using the pitcher plant (*Sarracenia purpurea*) inquiline

community. *Am. Midl. Nat.,* **157**:52–62.

———. 2011. Relative importance of top-down and bottom-up forces in food webs of

Sarracenia pitcher communities at a northern and a southern site. *Oecologia* (*Berl.*)*,*

165:1073–1082.

HUTCHINSON, M.C., B.B. MORA, S. PILOSOF, A.K. BARNER, S. KÉFI, E. THÉBAULT, P. JORDANO,

AND D.B. STOUFFER. 2019. Seeing the forest for the trees: putting multilayer networks to

work for community ecology. *Func. Ecol.,* **33**:206–217.

JOEL, D.M AND S. GEPSTEIN. 1985. Chloroplasts in the epidermis of *Sarracenia* (the American

pitcher plant) and their possible role in carnivory – an immunocytochemical approach.

Physiol. Plant., **63**:71–75.

KARAGATZIDES, J.D., J.L. BUTLER, AND A.M. ELLISON. 2009. The pitcher plant *Sarracenia*

*purpure*a can directly acquire organic nitrogen and short-circuit the inorganic nitrogen

cycle. *PLoS ONE,* **4**:e6164.

KEARSLEY, J. 1999. *Non-forested acidic peatlands of Massachusetts: a statewide inventory and*

vegetation classification. Westborough, MA, USA: Massachusetts Natural Heritage and

Endangered Species Program.

KITCHING, R.L. 2000. Food webs and container habitats: the natural history and ecology of

phytotelmata. Cambridge University Press. 429 p.

———. 2001. Food webs in phytotelmata: "bottom-up" and "top-down" explanations for

community structure. *Ann. Rev. Entomol.,* **46**:729–760.

KNEITEL, J.M. AND T.E. MILLER. 2002. Resource and top-predator regulation in the pitcher plant

(*Sarracenia purpurea*) inquiline community. *Ecology,* **83**:680–688.

KRIEGER, J.R. AND P.S. KOURTEV. 2012. Detection of methanogenic archaea in the pitchers of

the northern pitcher plant (*Sarracenia purpurea*). *Can. J. Microbiol.,* **58**:189–194.

LAYMAN, C.A., S.T. GIERY, S. BUHLER, R. ROSSI, T. PENLAND, M.N. HENSON, A.K. BOGDANOFF,

M.V. COVE, A.D. IRIZARRY, C.M. SCHALK, AND S.K. ARCHER. 2015. A primer on the history

of food web ecology: fundamental contributions of fourteen researchers. *Food Webs,* **4**:14–

24.

LUCIANO, C.S. AND S.J. NEWELL. 2017. Effects of prey, pitcher age, and microbes on acid

phosphatase activity in fluid from pitchers of *Sarracenia purpurea* (Sarraceniaceae). *PLoS*

ONE, **12**:e0181252.

MAGUIRE, B.J. 1971. Phytotelmata: biota and community structure in plant held waters. *Ann.*

Rev. Ecol. Syst., **2**:439–464.

MAHOWALD, N., T.D. JICKELLS, A.R. BAKER, P. ARTAXO, C.R. BENITEZ-NELSON, G.

BERGAMETTI, T.C. BOND, Y. CHEN, D.D. COHEN, B. HERUT, N. KUBILAY, R. LOSNO, C. LUO,

W. MAENHAUT, K.A. MCGEE, G.S. OKIN, R.L. SIEFERT, AND S. TSUKUDA. 2008. Global

distribution of atmospheric phosphorus sources, concentrations and deposition rates, and

anthropogenic impacts. *Glob. Biogeochem. Cycles,* **22**: GB4026.

MCCANN, K.S. 2012. Food webs. Princeton University Press. 256 p.

MELLICHAMP, T.L. AND F.W. CASE. 2009. *Sarracenia*. p. 350–363. *In*: Flora of North America

Editorial Committee (ed.). Flora of North America north of Mexico volume 8. Oxford

University Press.

MILLER, T.E. AND C.P. TERHORST. 2012. Testing successional hypotheses of stability,

heterogeneity, and diversity in pitcher-plant inquiline communities. *Oecologia*, **170**:243–251.

———, L. HORTH, AND R.H. REEVES. 2002. Trophic interactions in the phytotelmata

communities of the pitcher plant, *Sarracenia purpurea. Comm. Ecol.,* **3**:109–116.

MOORHEAD, D.L. AND R.L. SINSABAUGH. 2006. A theoretical model of litter decay and microbial

interaction. *Ecol. Mono.,* **76**:151–174.

MOUQUET, N., T. DAUFRESNE, S.M. GRAY, AND T.E. MILLER. 2008. Modelling the relationship

between a pitcher plant (*Sarracenia purpurea*) and its phytotelma community: mutualism or

parasitism? *Func. Ecol.,* **22**:728–737.

NATIONAL ATMOSPHERIC DEPOSITION PROGRAM (NRSP–3). 2020. NADP Program Office,

Wisconsin State Laboratory of Hygiene, Madison, Wisconsin. nadp.slh.wisc.edu/NADP/

[date of access: 11 January 2020].

NE'EMAN, G., R. NE'EMAN, AND A.M. ELLISON. 2006. Limits to reproductive success of

Sarracenia purpurea (Sarraceniaceae). *Am. J. Bot.* **93**:1660–1666.

NORTHROP, A.C., R.K. BROOKS, A.M. ELLISON, N.J. GOTELLI, AND B.A. BALLIF. 2017.

Environmental proteomics reveals taxonomic and functional changes in an enriched aquatic

ecosystem. *Ecosphere* **8**:e01954.

———, V. AVALONE, A.M. ELLISON, B.A. BALLIF, AND N.J. GOTELLI. 2021. Clockwise and counterclockwise hysteresis characterize changes in the same aquatic ecosystem. *Ecol. Lett.*, **24**:94-101.

OED ONLINE. 2019. "inquiline, n. (and adj.)". Oxford, UK: Oxford University Press. www.oed.com/view/Entry/96507 [date of access: 14 January 2020].

PAISE, T.K., T.E. MILLER, AND O.U. MASON. 2014. Effects of a ciliate protozoa predator on microbial communities in pitcher plant Sarracenia purpurea leaves. *PLoS ONE,* **9**:e113384.

PETERSON, C.N., S. DAY, B.E. WOLFE, A.M. ELLISON, R. KOLTER, AND A. PRINGLE. 2008. A keystone predator controls bacterial diversity in the pitcher plant (*Sarracenia purpurea*) microecosystem. *Env. Microbiol.,* **10**:2257–2266.

PILOSOF, S., M.A. PORTER, M. PASCUAL, AND S. KÉFI. 2017. The multilayer nature of ecological networks. *Nature Ecol. Evol.* **1**:0101.

PIMM, S. 1982. Food webs. Chapman & Hall. 258 p.

PIOVIA–SCOTT, J., L.H. YANG, AND A.N. WRIGHT. 2017. Temporal variation in trophic cascades. *Ann. Rev. Ecol., Evol., Syst.,* **48**:281–300.

POISOT, T., E. CANARD, D. MOUILLOT, N. MOUQUET, AND D. GRAVEL. 2012. The dissimilarity of species interaction networks. *Ecol. Lett.,* **15**:1353–1361.

POLIS, G.A. 1994. Food webs, trophic cascades and community structure. *Aust. J. Ecol.* **19**:121–136.

PRINGLE, R.M. AND M.C. HUTCHINSON. 2020. Resolving food-web structure. *Ann. Rev. Ecol., Evol., Syst.,* **51**:55–80.

R CORE TEAM. 2020. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna. https://www.R-project.org.

SCHIAFFINO, M.R., N. DIOVISALVI, D. MARFETÁN MOLINA, P. FERMANI, C. LI PUMA, L.

LAGOMARSINO, M.V.QUIROGA, AND G.L. PÉREZ. 2019. Microbial food-web components in

two hypertrophic human-impacted Pampean shallow lakes: interactive effects of

environmental, hydrological, and temporal drivers. *Hydrobiologia* **830**:255–276.

SHREVE, F. 1906. The development and anatomy of *Sarracenia purpurea*. *The Botanical Gazette*

(Old Series) **42**:107–126.

SIROTA, J., B. BAISER, N.J. GOTELLI, AND A.M. ELLISON. 2013. Organic-matter loading

determines regime shifts and alternative states in an aquatic ecosystem. *Proc. Nat. Acad. Sci.,*

110:7742–7747.

SRIVASTAVA, D.S., J. KOLASA, J. BENGTSSON, A. GONZALEZ, S.P. LAWLER, T.E. MILLER, P.

MUNGUIA, T. ROMANUK, D.C. SCHNEIDER, AND M.K. TRZCINSKI. 2004. Are natural

microcosms useful model systems for ecology? *Trends Ecol. Evol.,* **19**:379–384.

THORNHAM, D.G., J.M. SMITH, T.U. GRAFE, AND W. FEDERLE. 2012. Setting the trap: cleaning

behaviour of *Camponotus schmitzi* ants increases long-term capture efficiency of their

pitcher plant host, *Nepenthes bicalcarata*. *Func. Ecol.,* **26**:11–19.

TIPPING, E., S. BENHAM, J.F. BOYLE, P. CROW, J. DAVIES, U. FISCHER, H. GUYATT, R.

HELLIWELL, L. JACKSON-BLAKE, A.J. LAWLOR, D.T. MONTEITH, E.C. ROWE, AND H.

TOBERMAN. 2014. Atmospheric deposition of phosphorus to land and freshwater. *Environ*

Sci.: Process. Impacts, **16**:1608–1617.

WAKEFIELD, A.E., N.J. GOTELLI, S.E. WITTMAN, AND A.M. ELLISON. 2005. The effect of prey

addition on nutrient stoichiometry, nutrient limitation, and morphology of the carnivorous

plant *Sarracenia purpurea* (Sarraceniaceae). *Ecology,* **86**:1737–1743.

WALLACE, R.L., T.W. SNELL, C. RICCI, AND T. NOGRADY. 2006. Rotifera. Volume 1: Biology,

ecology and systematics, 2nd edition. Backhuys Publishers, 299 p.

WOLKOVICH, E.M., S. ALLESINA, K.L. COTTINGHAM, J.C. MOORE, S.A. SANDIN, AND C. DE

MAZANCOURT. 2014. Linking the green and brown worlds:sthe prevalence and effect of

multichannel feeding in food webs. *Ecol. Lett.* **95**:3376–3386.

ZHANG, S., S.R. CHALUVADI, AND J.L. BENNETZEN. 2020a. Draft genome sequence of a

Enterobacter sp. C6, found in the pitcher fluids of *Sarracenia rosea*. *Microbiol. Res.*

Announce., **9**:e01214–19.

———, ———, AND ———. 2020b. Draft genome sequence of a *Serratia marcescens* strain

isolated from the pitcher fluids of a *Sarracenia* pitcher plant. *Microbiol. Res. Announce.*,

9:e01216–19.

ZOU, K., E. THÉBAULT, G. LACROIX, AND S. BAROT. 2016. Interactions between the green and

brown food web determine ecosystem functioning. *Func. Ecol.* **30**:1454–1465.

Table 1 Values assigned to each inquiline taxon for calculation of a food-web saturation index.

Occurrence of taxa in a given food web is given by the sum of the binary values of all taxa

present. The saturation index is the sum of the decimal values of all taxa present. See Fig. 1 for

an illustrative example of these calculations

Taxon	Binary value	Decimal value
Metriocnemus	1	1
Habrotrocha	10	2
Sarraceniopus	100	4
Wyeomyia	1000	8
Fletcherimyia	10000	16

Table 2 Inquilines collected from 64 pitchers in the four-level full factorial experiment in which

food webs were isolated from the plant with polyethylene tubes, inorganic and organic nutrients

were added to assess bottom-up effects, and the top predator *Fletcherimyia fletcheri* was added

to assess top-down effects. This table illustrates the effects on inquiline occurrence of the two

factors—the polyethylene tubes and the addition of the top predator—that had significant effects

on food-web saturation (Fig. 3). Values in each cell are the number of pitchers in which each

inquiline was collected, pooled over inorganic and organic nutrient addition treatments and both

harvests. The maximum possible number of occurrences for any given inquiline in a Tube ×

Fletcherimyia treatment combination is 32. Because cell entries within each column are

occurrences summed over each Tube × *Fletcherimyia* treatment combination, all entries within a

single row do not imply co-occurrences. Occurrences of the other macrobial inquilines within a

tube treatment were unrelated to the addition of *Fletcherimyia*

Tube treatment	*Fletcherimyia* treatment	Number of pitchers in which inquiline occurred				
		Metriocnemus	*Habrotrocha*	*Sarraceniopus*	*Wyeomyia*	*Fletcherimyia*
Intact	Added	27	6	15	13	1
	Not added	26	8	11	20	2
Perforated	Added	31	7	8	28	13
	Not added	29	6	12	28	10
None	Added	24	2	11	28	14
	Not added	28	1	6	24	6

Fig. 1 Complete, fully saturated inquiline food web of *Sarracenia purpurea* (**a**; modified from

Baiser *et al.*, 2013) and an example of incomplete, unsaturated one (**b**). Black type indicates the

"macrobes" we studied here; grey type the microbes and detritus that we did not quantify. Values

adjacent to each of the macrobial taxa are those used to compute food-web saturation (see Table

1). Prey, bacteria, protozoa, and particulate organic matter were assumed to occur in all sampled

food webs, and each of the possible combinations of macrobes gives a unique saturation value.

In these two examples, the saturation value of the complete web (a) = 31, whereas that of the

food web lacking *Fletcherimyia* and *Sarraceniopus* (b) = 11

Fig. 2 Response surface of food-web saturation (defined in Table 1) in pitchers fed one of 25

combinations of nitrogen (N) and phosphorus (P) or distilled water once, three days after the

pitcher opened (the "pulse" experiment; left panel); or three days after the pitcher opened and

every two weeks thereafter (the "press" experiment; right panel). This figure illustrates data from

the first leaf of the season (fed on or beginning the week of 4 June 2001) at the mid-summer

harvest (6 August 2001 for the pulse experiment, 23 July 2001 for the press experiment), when

saturation was highest

Fig. 3 Effects of tube treatment manipulation of the top predator on food-web saturation in the

Sarracenia purpurea macrobial food webs in the four-level tube × inorganic nitrogen × fruit-fly

× *Fletcherimyia* manipulation. Data are pooled among nutrient and addition treatments, for

which the main effects were not statistically significant (see main text for ANCOVA results). In

each panel, the effects of the three levels of the inserted tubes (intact, perforated, absent) are

ordered along the *x*-axis; the two different panels illustrate effects of the tube treatment without

(left) or with (right) *Fletcherimyia* present in the pitcher. Box-and-whisker plots illustrate

median (dark horizontal line), upper and lower quartiles (top and bottom of the box), and upper

609 and lower hinges (length of vertical "whiskers" = 1.5×interquartile range). Individual points

610 represent data from each pitcher harvested in mid-summer (grey points) or at the end of the

611 growing season (black points).