

1 **Multidimensional trophic niche revealed by complementary approaches: gut content,**  
2 **digestive enzymes, fatty acids and stable isotopes in soil fauna**

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## 17 **Abstract**

- 18 1. The trophic niche of an organism is tightly related to its role in the ecosystem and to  
19 interactions with other species. Thousands of species of soil animals feed on detritus and  
20 co-exist with apparently low specialisation in food resource use. Trophic niche  
21 differentiation may explain species coexistence in such a cryptic environment. However,  
22 most of the existing studies provide only few and isolated evidence on food resources,  
23 thus simplifying the multidimensional nature of the trophic niches available in soil.
- 24 2. Focusing on one of the most diverse soil taxa – springtails (Collembola) – we aimed to  
25 reveal the additional value of information provided by four complementary methods:  
26 visual gut content-, digestive enzyme-, fatty acid- and stable isotope analyses, and to  
27 demonstrate the multidimensional nature of trophic niches.
- 28 3. From 40 studies, we compiled fifteen key trophic niche parameters for 125 species, each  
29 analysed with at least one method. Focusing on interspecific variability, we explored  
30 correlations of trophic niche parameters and described variation of these parameters in  
31 different Collembola species, taxonomic groups and life forms.
- 32 4. Correlation between trophic niche parameters of different methods was weak in 45 out of  
33 64 pairwise comparisons, reflecting the complementarity of the multidimensional trophic  
34 niche approach. Gut content and fatty acids provided comparable information on  
35 fungivory and plant feeding in Collembola. Information provided by digestive enzymes  
36 differed from that gained by the other methods, suggesting its high additional value.  
37 Stable isotopes were mainly related to plant versus microbial feeding. Many parameters  
38 were affected by taxonomic affiliation but not life form. Furthermore, we showed

39 evidence of bacterial feeding, which may be more common in Collembola than usually  
40 assumed.

41 5. Different methods reveal different feeding dimensions, together drawing a  
42 comprehensive picture of the trophic niche in taxa with diverse feeding habits. Food web  
43 studies will benefit from simultaneously applying several joint approaches, allowing to  
44 trace trophic complexity. Future studies on the multidimensional trophic niche may  
45 improve understanding of food-web functioning and help to explain species coexistence  
46 in cryptic environments such as soil.

47

48 **Keywords**

49 diet tracing, food webs, biomarkers, trophic interactions, springtails, method comparison,  
50 Collembola

## 51 **Introduction**

52           Trophic interactions between organisms influence structure and stability of ecological  
53 communities, channeling of energy through food webs, and functioning of ecosystems (Barnes et  
54 al., 2018; Rooney & McCann, 2012). The trophic interactions of an organism with other  
55 coexisting species can be perceived as its ‘trophic niche’, stemming from the Hutchinsonian  
56 ‘ecological niche’ concept (Holt, 2009). Trophic niche can be defined in multidimensional space,  
57 where each axis represents an aspect, or dimension, of trophic interactions (Hutchinson, 1978;  
58 Machovsky-Capuska, Senior, Simpson, & Raubenheimer, 2016; Newsome, Martinez del Rio,  
59 Bearhop, & Phillips, 2007). Such dimensions could represent direct evidence of food objects,  
60 describe feeding behavior of individuals, and imply basal resources and trophic level of species  
61 in food webs with potential interactions with other species.

62           Soil food webs rely on dead plant and animal material, soil organic matter, phototrophic  
63 microorganisms, roots and root exudates, inseparably mixed with bacteria and fungi, and  
64 altogether generalized as ‘detritus’ (Moore et al., 2004). Thousands of soil animal species,  
65 known as ‘detritivores’ and ‘microbivores’, feed on this mixture and locally co-exist with  
66 apparently low specialisation, which inspired J.M. Anderson to formulize the ‘enigma of soil  
67 animal diversity’ (Anderson, 1975). Trophic niche differentiation is one mechanism explaining  
68 species coexistence in soil. Understanding of trophic niche differentiation in soil invertebrates,  
69 however, was for a long time constrained by the small size and cryptic lifestyle of these animals,  
70 and only few trophic niche dimensions are known. In natural soil habitats, four common methods  
71 are used, each with its advantages and drawbacks:

72           (1) Visual gut content analysis provides reliable data on ingested food materials by  
73 microscopic observations of gut content and counting different types of particles (Anderson &

74 Healey, 1972; Hagvar & Kjondal, 1981; Ponge, 2000). Fungal spores and hyphae in the gut  
75 indicate fungivory of soil animals, while coarse plant detritus, roots and shoots suggest herbivory  
76 and litter grazing, and amorphous material such as fine detritus may imply feeding on soil  
77 organic matter and faecal pellets. However, visual gut content analysis only presents a snapshot  
78 of the ingested materials, overestimates poorly digestible particles and provides limited  
79 information in case of feeding on fluids.

80 (2) Digestive enzymes, such as cellulase, chitinase and trehalase, represent the ability of  
81 an animal to decompose corresponding types of organic compounds, and provide a way to assess  
82 which ingested materials may be digested (C. O. Nielsen, 1962; Parimuchová et al., 2018; Siepel  
83 & de Ruiter-Dijkman, 1993). Cellulose is a major component of cell walls of green plants and  
84 algae; cellulase activity suggests herbivory, algivory or litter grazing of soil animals. Chitin is a  
85 major component of fungal cell walls and chitinase activity suggests fungivory. Trehalose, by  
86 contrast, is a storage component of fungal, lichen and plant cells; trehalase activity can be used  
87 as a proxy for fungivory and herbivory. Furthermore, foraging strategies of soil animals can be  
88 inferred by a combination of the three digestive enzyme analyses : ‘grazers’, which can digest  
89 both cell-walls and cell-contents, have a higher activity of cellulase and chitinase to degrade  
90 structural polysaccharides, while ‘browsers’, which can only digest cell-contents, have a higher  
91 activity of trehalase to degrade storage polysaccharides (Siepel & de Ruiter-Dijkman, 1993).  
92 However, digestive enzymes provide information on potential, rather than real assimilation of  
93 food compounds.

94 (3) Neutral lipid fatty acid (FA) analysis, by contrast, detects assimilated compounds that  
95 are retained in the fat body of consumers, a phenomenon called ‘dietary routing’ ( Chamberlain,  
96 Bull, Black, Ineson, & Evershed, 2005a; Ruess & Chamberlain, 2010). Plants, fungi and

97 different groups of prokaryotes synthesise specific membrane lipids and these compounds can be  
98 tracked in animal consumers over a period of time (usually few weeks for mesofauna; Haubert,  
99 Pollierer, & Scheu, 2011) and across trophic levels (Pollierer, Scheu, & Haubert, 2010). An  
100 extensive review of the fatty acid method for soil food web analysis can be found in Ruess &  
101 Chamberlain (2010). Despite being informative, FA analysis does not provide estimation of a  
102 species trophic level in the soil food web. Quantitative comparisons among contributions of  
103 different food origins are also limited (Kühn, Schweitzer, & Ruess, 2019).

104 (4) Similar to FA analysis, stable isotope analysis provides information on assimilated  
105 food resources of soil animals integrated over time, but the method is quantitative and allows for  
106 trophic level estimation (Tiunov, 2007). Low  $^{13}\text{C}$  concentration in animal body tissue indicates  
107 utilisation of freshly fixed plant carbon (e.g. herbivory), while high  $^{13}\text{C}$  concentration suggests  
108 consumption of microbially processed organic matter (e.g. soil feeding, bacterivory or fungivory)  
109 (A. M. Potapov, Tiunov, & Scheu, 2019). The  $^{15}\text{N}$  concentration, by contrast, infers trophic  
110 levels of animals in the food web, being low in primary consumers but high in predators and  
111 mycorrhizal fungi feeders (A. M. Potapov, Tiunov, & Scheu, 2019). However, bulk natural  
112 stable isotopes provide only rough information on the trophic position and rarely allow to  
113 reconstruct exact feeding interactions in soil.

114 Different dietary methods provide information on different trophic niche dimensions of  
115 consumers over different time scales. A recent review revealed that only few studies have  
116 conducted quantitative comparisons among different methods, and none of them simultaneously  
117 applied multiple methods (J. M. Nielsen, Clare, Hayden, Brett, & Kratina, 2018). This motivated  
118 us to compile a trophic trait dataset across the four abovementioned methods from field studies  
119 and to analyse trophic niche differentiation among soil animals. We chose springtails

120 (Collembola) as an example, since they are one of the most abundant and diverse soil  
121 invertebrates and traditionally considered as generalistic fungivores (Hopkin, 1997). However,  
122 detritivorous Collembola may assimilate only a small percentage of the ingested food (Jochum et  
123 al., 2017). Although most of Collembola are ‘herbo-fungivorous grazers’, having cellulase,  
124 chitinase and trehalase activity in digestive system (Berg, Stoffer, & van den Heuvel, 2004), they  
125 in fact occupy different trophic positions in the soil food web, spanning from algivores to high-  
126 level consumers, as indicated by the stable isotope  $^{15}\text{N}$  values (Chahartaghi, Langel, Scheu, &  
127 Ruess, 2005; Rusek, 1998). Different species of Collembola also differ in FA compositions,  
128 suggesting that they rely on food resources of different origins (Ferlian, Klarner, Langeneckert,  
129 & Scheu, 2015; T.-W. Chen, Sandmann, Schaefer, & Scheu, 2017). In particular, trophic niches  
130 of Collembola species likely correlate with taxonomic position and life form. The former  
131 correlation may suggest phylogenetic constraints, while the latter implies microhabitat  
132 specialisation in Collembola trophic niches (A. M. Potapov, Semenina, Korotkevich,  
133 Kuznetsova, & Tiunov, 2016). In this study we aimed to (1) quantitatively assess the  
134 complementarity provided by different dietary methods; (2) describe multidimensional trophic  
135 niches among different species, taxonomic groups and life forms of a model soil animal group  
136 (Collembola).

137

## 138 **Materials and methods**

139 We compiled trophic data on Collembola from field studies that used visual gut content,  
140 digestive enzyme, FA and stable isotope analyses. Data were collected from the personal  
141 libraries of the authors and complemented with searching for published literature in the Web of  
142 Science. A complete list of studies can be found in Supplementary Materials. Most of the

143 published studies applied only one method and only two used a combination of two methods  
144 (Haubert et al. 2009; Ferlian et al. 2015). For each study we averaged individual measurements  
145 by species and ecosystem for fifteen trophic niche parameters derived from the four methods  
146 (**Table 1**).

147

148 **Table 1.** List of the trophic niche parameters used in this study. Units are given in square  
149 brackets.

<b>Parameter</b>	<b>Provided information</b>
<b>1. Visual gut content analysis</b> [proportion of total particles found]	
Proportion of fungal particles	A proxy for fungivory
Proportion of plant particles	A proxy for herbivory and litter grazing
Proportion of amorphous material	A proxy for soil organic matter and faecal pellets feeding
<b>2. Digestive enzyme analysis</b> [reaction product $\mu\text{g mg}^{-1}$ body weight $\text{hour}^{-1}$ ]	
Cellulase activity	A proxy for herbivory, algivory and litter grazing
Trehalase activity	A proxy for fungivore (and herbivore) browsing strategy
Chitinase activity	A proxy for fungivory
<b>3. Fatty acid (FA) analysis</b> [proportion of total FA]	
Sum of gram-positive bacteria biomarkers	A proxy for bacterial feeding
Sum of gram-negative bacteria biomarkers	A proxy for bacterial feeding
Sum of non-specific bacterial biomarkers	A proxy for bacterial feeding
Relative fungal biomarker	A proxy for fungivory



Relative and non-specific plant biomarkers	A proxy for herbivory
Non-specific biomarker for arbuscular-mycorrhizal fungi	A (potential) proxy for mycorrhiza feeding
Animal-synthesized FAs	A (potential) proxy for predation (e.g. on nematodes)

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#### 4. Bulk stable isotope analysis [‰]

$\Delta^{13}\text{C}$	A proxy for herbivory (low values) versus microbial feeding (high values)
$\Delta^{15}\text{N}$	A proxy for algivory and litter grazing (low values) versus microbial and animal feeding (high values)

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150

#### 151 *Visual gut content*

152 Primary screening of literature yielded 31 studies that reported data on the gut content of  
153 Collembola. We selected those that provided quantitative estimates from natural environments,  
154 mostly temperate forests and grasslands. We took the most commonly reported categories and  
155 defined gut content parameters as (1) particles of fungal origin, including hyphae and spores, (2)  
156 particles of plant origin, mostly coarse plant detritus (excluding pollen and algae), and (3)  
157 amorphous material of unknown nature (i.e. fine detritus such as soil organic matter). The final  
158 dataset for the three gut content parameters included 77 records on 56 species from 15 studies  
159 (**Table S1**). Raw data were expressed as proportion of certain type of particles among the total  
160 particles ingested.

161

#### 162 *Digestive enzymes*

163 To our knowledge, cellulase, trehalase and chitinase activities in Collembola were reported only  
164 in three studies (Berg et al., 2004; Parimuchová et al., 2018; Urbášek & Rusek, 1994). Despite  
165 using the same conceptual method, these studies used different chemical protocols and ways of  
166 glucose detection, which resulted in evident differences in absolute mean values of substrate  
167 production per unit of animal body mass. Thus, we excluded the study of (Urbášek & Rusek,  
168 1994). The final dataset included 45 records on 27 species (**Table S2**). Raw data of digestive  
169 enzyme activity were expressed as mg of reaction products per g of animal mass per hour.

170

### 171 *Fatty acids*

172 Screening of literature yielded 10 studies that reported neutral lipid FA compositions of  
173 Collembola. The dataset was complemented with unpublished data collected by Melanie M.  
174 Pollierer. Studies varied in completeness of FA profiles, but most of them reported data on  
175 16:1 $\omega$ 7 and 18:1 $\omega$ 7 as general bacteria biomarkers, 18:2 $\omega$ 6,9 as relative fungal biomarker,  
176 18:1 $\omega$ 9 (in addition 21:0, 22:0, 23:0, 24:0) as relative plant biomarkers and several gram-positive  
177 bacterial biomarkers (including i15:0, a15:0, i16:0, i17:0, a17:0) and gram-negative bacterial  
178 biomarkers (including 2-OH 10:0, 2-OH 12:0, 3-OH 12:0, 2-OH 14:0, 3-OH 14:0, 2-OH 16:0,  
179 cy17:0, cy19:0). If reported, we also used 16:1 $\omega$ 5 as the biomarker related to feeding on  
180 arbuscular-mycorrhizal fungi (Ngosong, Gabriel, & Ruess, 2012) and sum of 20:1 $\omega$ 9, 22:6 $\omega$ 3,  
181 22:2 $\omega$ 6, 22:1 $\omega$ 9 and 24:1 as the FAs in metazoan animals (e.g. nematodes) ( Chamberlain &  
182 Black, 2005; Chamberlain, Bull, Black, Ineson, & Evershed, 2005b; J. Chen, Ferris, Scow, &  
183 Graham, 2001; Tanaka et al., 1996). Raw data was compiled as proportions of the total neutral  
184 lipid FAs of the organism. Individual biomarker FAs were summed up to generate seven  
185 conventional FA parameters (**Table 1**). The final dataset included 130 records on 47 species

186 from 13 studies (**Table S3**).

187

188 *Stable isotopes*

189 The dataset was based on the previous compilation of Potapov et al. (2016) and complemented  
190 with data on grassland and forest communities, including recent publications. For each study,  
191 isotopic baseline (i.e. plant litter) was used to calculate litter-calibrated  $\Delta^{13}\text{C}$  and  $\Delta^{15}\text{N}$  values of  
192 Collembola (A. M. Potapov, Tiunov, & Scheu, 2019). The final dataset for the two stable isotope  
193 parameters included 378 records on 96 species from 10 studies (**Table S4**).

194

195 *Data analysis*

196 For the fifteen trophic niche parameters derived from the four methods (**Table 1**) we in total  
197 included 125 species, each analysed with at least one method (**Table S1-4**; Dryad Digital  
198 Repository, doi:10.5061/dryad.xxxxx). All calculations were based on these datasets and  
199 conducted in R 3.5.3 (R Core Team, 2019).

200 Data on digestive enzyme activity were added with  $0.02 \mu\text{g product mg}^{-1} \text{h}^{-1}$  (the minimum  
201 positive value observed) for all values and then  $\log_{10}$ -transformed. We normalized data of  
202 chitinase activity separately for Berg et al. (2004) and Parimuchová et al. (2018) by subtracting  
203 mean and dividing by standard deviation to account for the differences in chemical protocols.  
204 Proportional data (gut content and FA) were logit-transformed, with 0 and 1 proportions adjusted  
205 to 0.025 and 0.975, respectively, using the *logit* function in the *car* package.

206 Focusing on interspecific variability across trophic parameters, we conducted species-based  
207 analyses using the species names following GBIF (<https://www.gbif.org/tools/species-lookup>).

208 For each trophic parameter and each species, we averaged data across ecosystems and studies.  
209 To have the same data representation across all fifteen parameters, each parameter was scaled  
210 between 0 (lowest observed value of the parameter) and 1 (highest observed value of the  
211 parameter). All the following analyses and results were based on the scaled data.

212 We performed three analyses: First, we tested pairwise correlations among all trophic niche  
213 parameters. Spearman rank correlation was applied using the *cor* function. The number of points  
214 for each correlation varied among parameters according to the number of shared species of the  
215 paired parameters (**Table S6**). Correlation tests were conducted for those with a minimum of six  
216 data points.

217 Second, we explored the association of species with their trophic parameters and visualised  
218 interspecific differentiation in multidimensional trophic niches using principal component  
219 analysis (PCA) with the *prcomp* function. We selected species according to their common  
220 trophic parameters available. Only two species had data across all fifteen parameters. Thus, we  
221 excluded parameters with fewer species, and finally chose six species that had data across seven  
222 parameters, with a focus on gut content and fatty acid parameters.

223 Third, we tested the effect of taxonomic affiliation (as the proxy of phylogenetic group) and life  
224 form (as the proxy of microhabitat preference) on each of the fifteen trophic niche parameters  
225 using linear models (the *lm* function). Groups with fewer than three species were excluded from  
226 the analysis. Species of Dicyrtomidae, Katiannidae, Sminthuridae, Sminthurididae,  
227 Bourletiellidae and Arrhopalitidae and species of Onychiuridae and Tullbergiidae were pooled  
228 at a higher taxonomic rank, Symphyleona and Onychiuroidea, respectively, due to low number  
229 of species in each family and trophic similarity among families (A. M. Potapov, Semenina, et al.,

230 2016). Definition of life form followed Gisin (1943) as interpreted by A. M. Potapov, Semenina,  
231 et al., (2016) and species were categorised into atmobiotic (aboveground and surface dwellers),  
232 epedaphic (surface and upper litter dwellers), hemiedaphic (litter dwellers) and euedaphic (lower  
233 litter and soil dwellers). We further divided the best replicated family Isotomidae in epedaphic  
234 versus hemiedaphic and euedaphic species to assess the effect of life form on trophic niche  
235 parameters within this family. We reported  $R^2$  and p values from the model output to compare  
236 predictability of different trophic niche parameters among taxonomic groups and among life  
237 forms. For each parameter we reported median values for taxonomic groups and life forms. We  
238 then used PCA to visualise multi-dimensional trophic niches among taxonomic groups.

239

## 240 **Results**

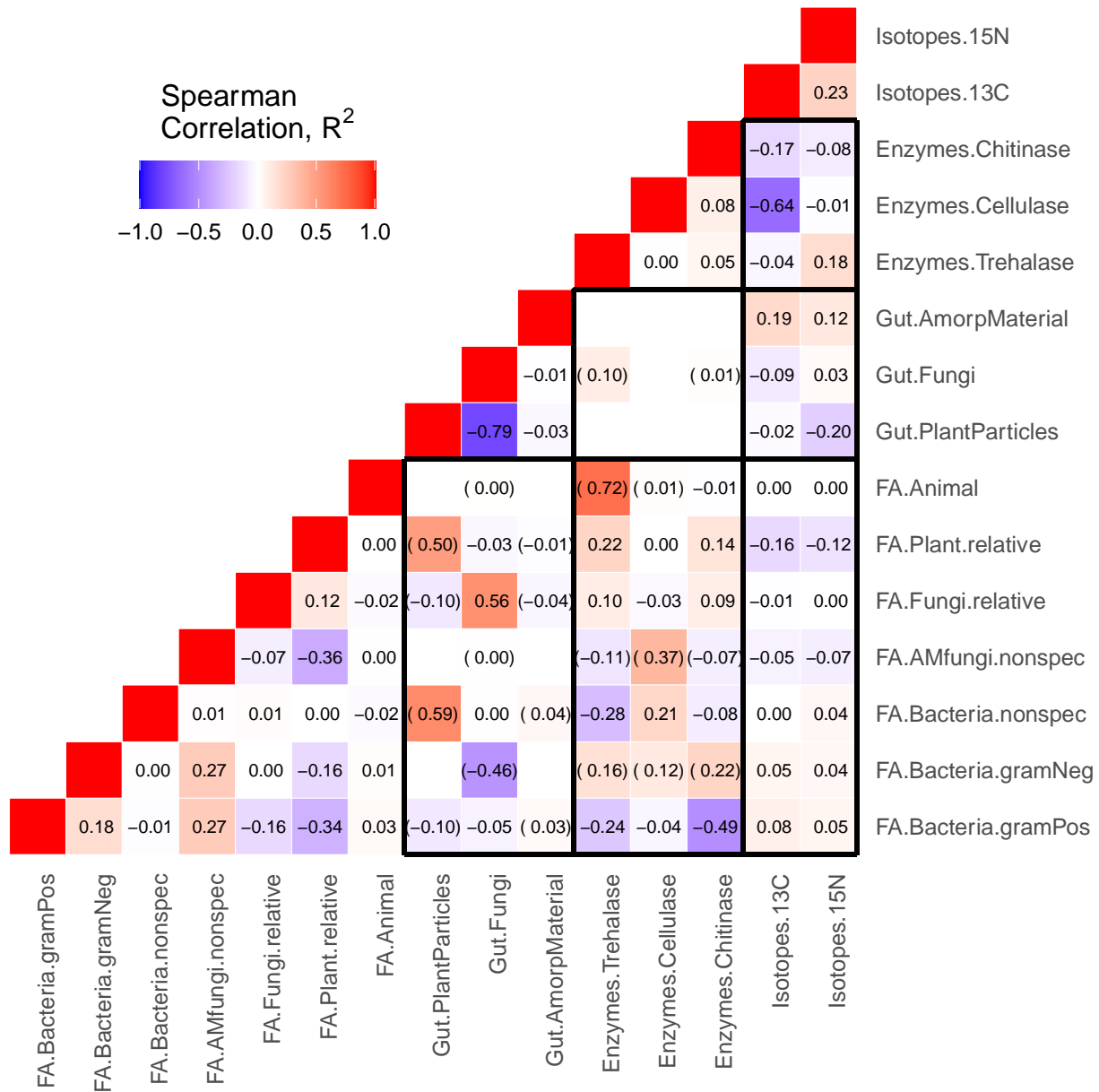
### 241 *Correlation between trophic niche parameters*

242 In 45 out of total 64 performed pairwise tests (excluding within-method tests), correlation  
243 between trophic niche parameters was weak or absent ( $R^2 < 0.15$ ; **Fig. 1**; for more details see the  
244 Supplementary **Fig. S1**). Among strong positive correlations, species with high proportion of  
245 fungal and plant particles in their guts also had high proportions of fungal-synthesised ( $R^2 =$   
246 0.56) and plant-synthesised FAs ( $R^2 = 0.50$ ), respectively. A high proportion of fungal particles  
247 in the gut, however, negatively correlated with retained FA synthesised by gram-negative  
248 bacteria ( $R^2 = 0.46$ ).

249 Trehalase activity in guts positively correlated with the proportions of retained FAs synthesized  
250 by fungi ( $R^2 = 0.10$ ), gram-negative bacteria ( $R^2 = 0.16$ ), plants ( $R^2 = 0.22$ ) and animals ( $R^2 =$   
251 0.72) in the bodies, but negatively with the proportions of FAs synthesised by gram-positive

252 bacteria ( $R^2 = 0.24$ ) and non-specific bacteria biomarkers ( $R^2 = 0.28$ ). Cellulase activity  
253 positively correlated with the proportion of arbuscular-mycorrhizal fungi ( $R^2 = 0.37$ ) and non-  
254 specific bacteria FAs ( $R^2 = 0.21$ ). Similar to trehalase activity, chitinase activity negatively  
255 correlated with the proportion of FAs synthesised by gram-positive bacteria ( $R^2 = 0.49$ ) and  
256 positively with the proportion of FAs synthesised by gram-negative bacteria ( $R^2 = 0.22$ ).

257 The  $\Delta^{13}\text{C}$  values negatively correlated with cellulase activity ( $R^2 = 0.64$ ), chitinase activity ( $R^2 =$   
258  $0.17$ ) and proportion of plant-synthesised FAs ( $R^2 = 0.16$ ). The  $\Delta^{15}\text{N}$  values negatively correlated  
259 with proportion of plant particles in gut ( $R^2 = 0.20$ ) and plant-synthesised FAs ( $R^2 = 0.12$ ), but  
260 positively with trehalase activity ( $R^2 = 0.18$ ).



261

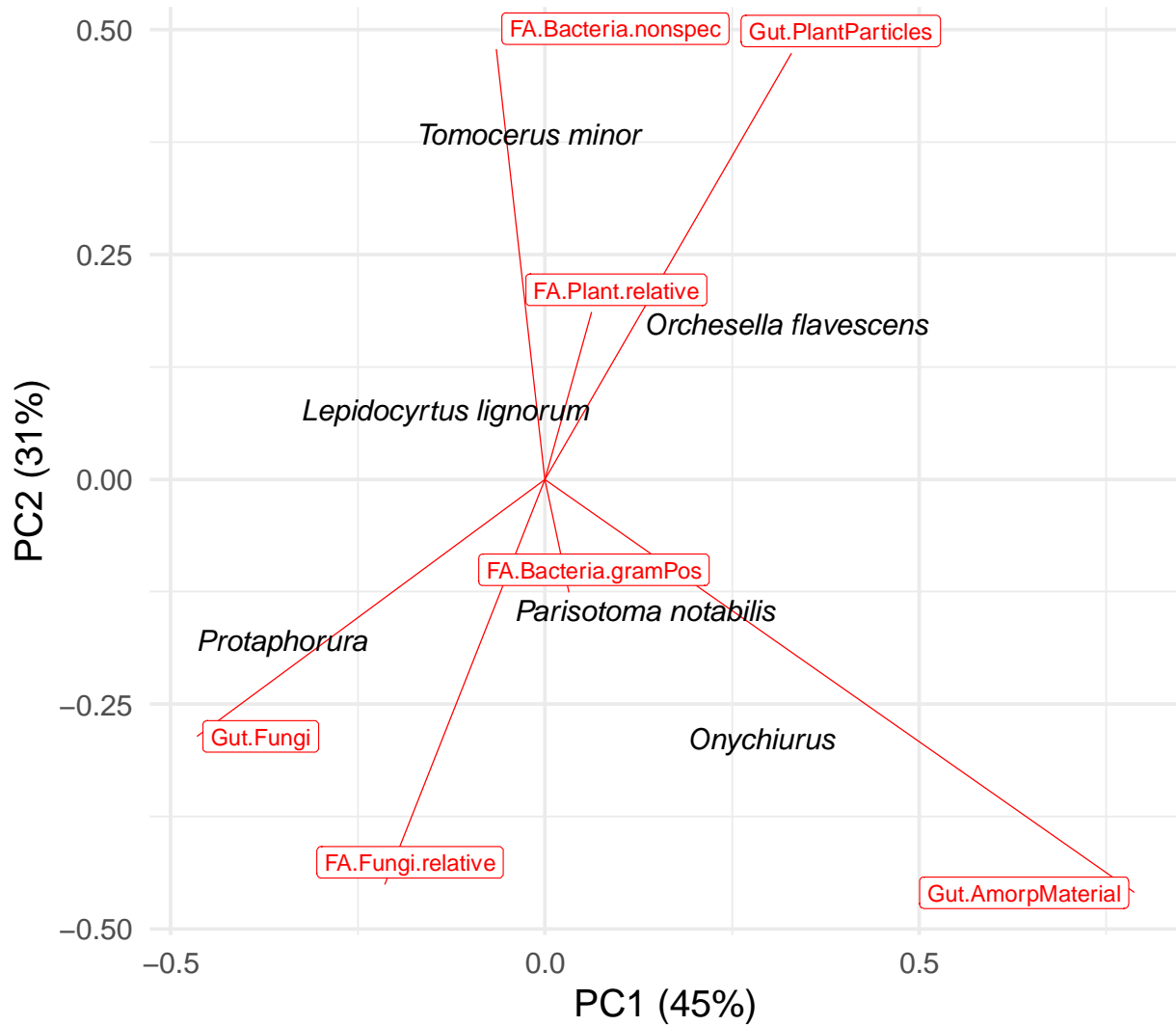
262 **Figure 1.** Correlation among fifteen trophic niche parameters in Collembola. Red colour  
 263 represents positive correlations, blue colour represents negative correlations; to display negative  
 264 correlations  $R^2$  was multiplied by -1. Correlations based on < 6 species were excluded,  
 265 correlations based on 6–9 species are given in brackets; in other cases,  $n = 10–88$  (**Table S5**).  
 266 Correlations among parameters derived from different methods are framed within black  
 267 rectangles.

268

269 *Multidimensional trophic niches of Collembola species*

270 The strongest distinction in multidimensional trophic niche space was observed between  
271 *Orchesella flavescens* (surface-dwelling species of Entomobryidae) and *Protaphorura* (soil-  
272 dwelling genus of Onychiuridae; **Fig. 2**). The former species was associated with high  
273 proportions of plant-synthesised FAs and plant particles in the gut, and the latter with high  
274 proportions of fungi-synthesised FAs and fungi in the gut. *Tomocerus minor* (Tomoceridae) was  
275 associated with plant and non-specific bacteria parameters. *Lepidocyrtus lignorum* (litter-  
276 dwelling species of Entomobryidae) was related to fungi in the gut. *Parisotoma notabilis* (litter-  
277 dwelling species of Isotomidae) and *Onychiurus* (soil-dwelling genus of Onychiuridae) were  
278 associated with gram-positive bacteria FAs and amorphous material in the gut.





279

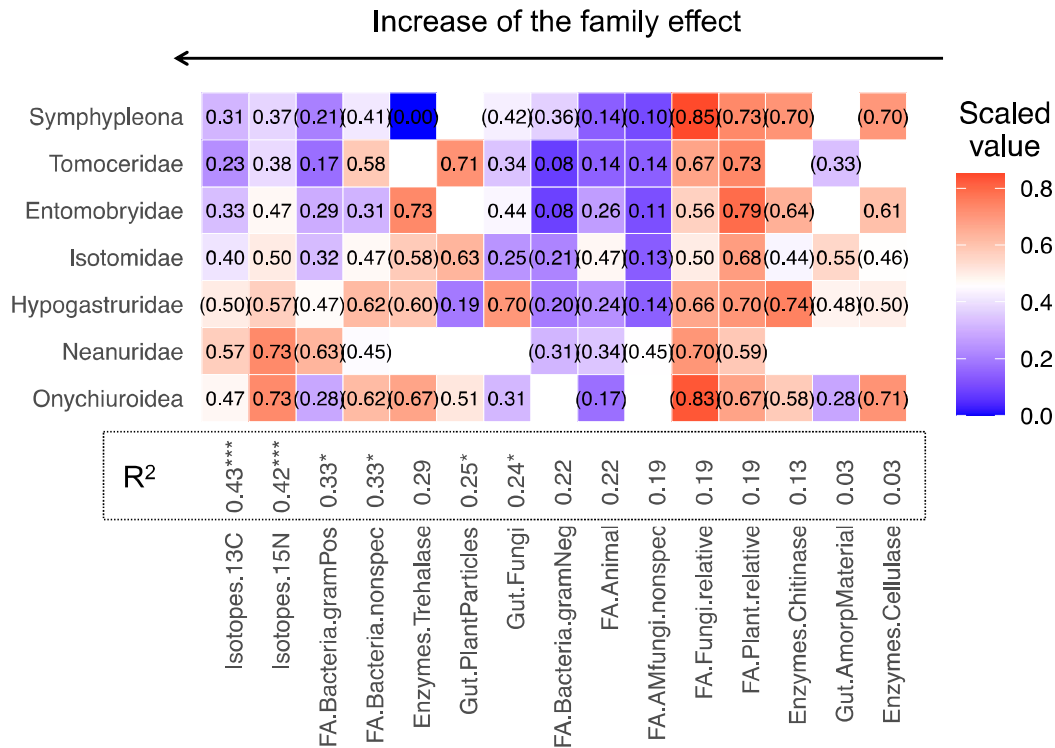
280 **Figure 2.** Principal component analysis based on seven selected trophic niche parameters of six  
281 common Collembola species. Species are shown with grey dots; trophic parameters are shown  
282 with red vectors.

283

284 *Taxonomic and life form effects on trophic niche parameters of Collembola*

285 Taxonomic affiliation best explained stable isotope  $\Delta^{13}\text{C}$  and  $\Delta^{15}\text{N}$ , and also explained part of the  
286 variation in gram-positive and non-specific bacterial FA parameters, proportion of plant and

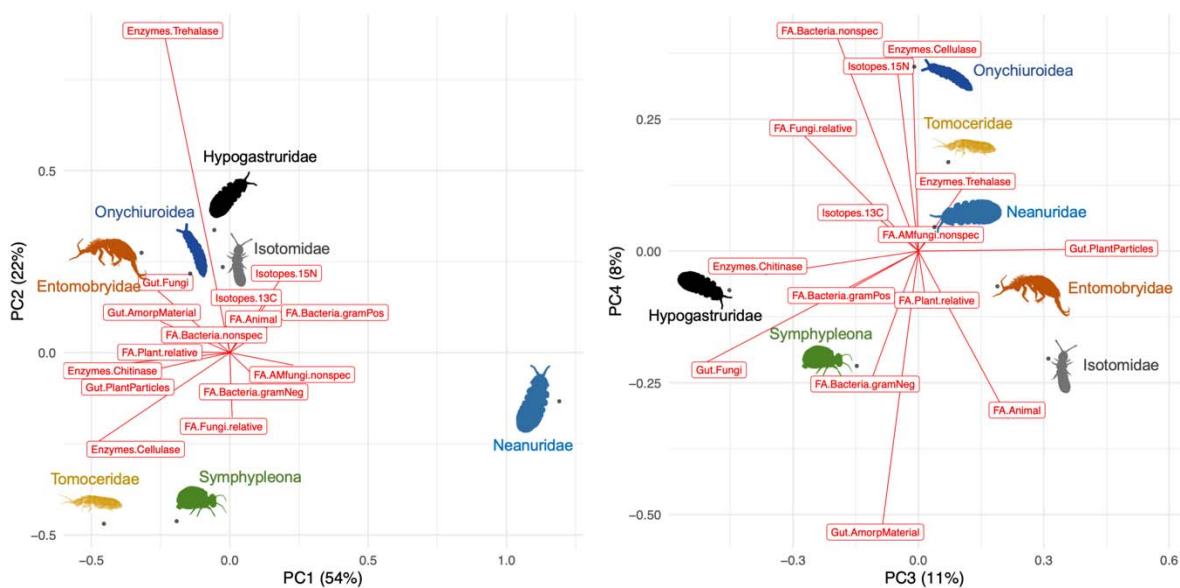
287 fungi in the gut, and trehalase activity (all  $R^2 \geq 0.24$ ,  $p < 0.05$  except for trehalase **Fig. 3**).  
 288 Cellulase activity and proportion of amorphous material in gut were poorly related to the  
 289 taxonomic affiliation ( $R^2 = 0.03$ ).



290  
 291 **Figure 3.** Median values of trophic niche parameters in Collembola taxonomic groups. All  
 292 parameters were scaled between 0 (minimum, intense blue) and 1 (maximum, intense red).  
 293 Parameter values based on < 3 species were excluded; values based on 3–5 species are given in  
 294 brackets; in other cases,  $n = 6–20$ . Numbers next to parameters indicate  $R^2$  explained by  
 295 taxonomic affiliation; \*\*\* $p < 0.001$ , \*\* $p < 0.01$ , \* $p < 0.05$ . Taxonomic groups are sorted  
 296 according to the orders; parameters are sorted according to the  $R^2$  values.

297

298 Analysed together, the seven Collembola taxonomic groups differed in their trophic niches (**Fig.**  
299 **4**). Symphypleona had high average proportion of FAs synthesised by gram-negative bacteria  
300 (PC4) and fungi (PC2), high chitinase and cellulase (PC1), but very low trehalase activity (PC2).  
301 Tomoceridae were characterised by high average proportions of plant particles in the gut (PC1)  
302 and low  $\Delta^{13}\text{C}$  and  $\Delta^{15}\text{N}$  values (PC1, PC2). Entomobryidae had the highest trehalase activity  
303 (PC2) but low values of bacteria-related parameters (PC1). By contrast, Isotomidae had lower  
304 values of fungi-related parameters and enzyme activities (PC2, PC3, PC4). A further separation  
305 of Isotomidae into epedaphic and (hemi)edaphic life forms indicated that epedaphic Isotomidae  
306 had lower  $\Delta^{13}\text{C}$  values (0.35) than edaphic ones (0.49), a lower proportion of plants (epedaphic:  
307 0.55; edaphic: 0.72) but a higher proportion of fungi in the gut (epedaphic: 0.42; edaphic: 0.20).  
308 Hypogastruridae had a remarkably high proportion of fungi in the gut and high chitinase activity,  
309 but a low proportion of plant particles in the gut (PC3). Neanuridae had the highest  $\Delta^{13}\text{C}$  and  
310  $\Delta^{15}\text{N}$  values and proportions of FAs synthesized by gram-positive bacteria and arbuscular-  
311 mycorrhizal fungi (PC1). Onychiuroidea had high  $\Delta^{15}\text{N}$  values, FAs synthesized by non-specific  
312 bacteria and fungi, and high cellulase activity (PC4).



313

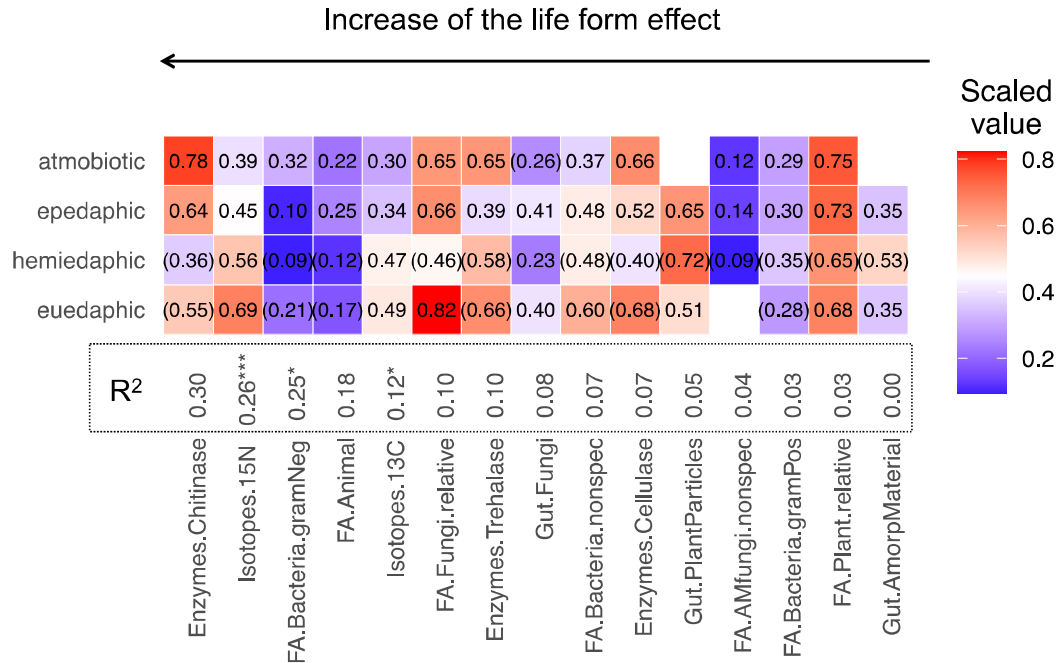
314 **Figure 4.** Principal component analysis based on median values of the fifteen trophic niche  
 315 parameters of Collembola taxonomic groups. Collembola groups are shown with dots and  
 316 silhouettes; trophic parameters are shown with red vectors. Most of the silhouettes were taken  
 317 from <http://phylopic.org>, credit goes to Birgit Lang and Kamil S. Jaron.

318

319 Life form best explained chitinase activity, but the effect was not significant due to a low number  
 320 of replicates. Life form also well explained  $\Delta^{15}\text{N}$  values and proportion of FA synthesised by  
 321 gram-negative bacteria (both  $R^2 \geq 0.25$ ; **Fig. 5**). Most of the other parameters were moderately or  
 322 not at all related to life form.

323 Atmobiotic species had high average activity of all digestive enzymes and a high proportion of  
 324 FAs synthesized by gram-negative bacteria and plants. Epedaphic species had in general  
 325 intermediate values for all parameters, but high proportions of fungi in the gut and high

326 proportions of animal-synthesized FAs. Hemiedaphic species had high  $\Delta^{13}\text{C}$  values and  
 327 proportion of FAs synthesized by gram-positive bacteria, but low activity of chitinase and  
 328 cellulase. Euedaphic species had the highest  $\Delta^{15}\text{N}$  values, high trehalase activity, high  
 329 proportions of FAs synthesised by fungi and non-specific bacteria biomarker FA.



330

331 **Figure 5.** Median values of trophic niche parameters in Collembola life forms. All parameters  
 332 were scaled between 0 (minimum) and 1 (maximum). Parameter values based on < 3 species  
 333 were removed, values based on 3–5 species are given in brackets; in other cases, n = 6–36.  
 334 Numbers next to parameters show R<sup>2</sup> explained by life form identity; \*\*\*p < 0.001, \*\*p < 0.01,  
 335 \*p < 0.05. Life forms are sorted according to the species vertical stratification along the soil  
 336 profile; parameters are sorted according to the R<sup>2</sup> values.

## 337 **Discussion**

338 Different dietary methods track different processes of animal feeding at different resolution and  
339 time scales, providing information on the multidimensional nature of trophic niches. However,  
340 studies quantitatively comparing different methods are scarce, and usually involve one or two  
341 methods only (Nielsen et al., 2018). Here, we compared fifteen trophic niche parameters derived  
342 from four methods using Collembola – a model group of the cryptic and diverse soil animals.  
343 First, we showed that trophic niche parameters were weakly correlated, reflecting  
344 complementarity, rather than redundancy, of different methods. Second, we outlined the trophic  
345 niche parameters that vary with taxonomic clusters and microenvironments of species. Finally,  
346 we presented the multidimensional trophic niche of Collembola, and provided the most detailed  
347 trophic information to date on this animal group.

348

### 349 *The additional value of method combination*

350 We found weak correlations among trophic niche parameters derived from different  
351 methods in 70% of the performed tests. As already shown, outcomes provided by different  
352 dietary methods only in part overlap (Nielsen et al., 2018). Pessimistically, this reflects biases  
353 embedded in each method – optimistically, it means that different methods inform on more  
354 trophic niche dimensions (Hambäck, Weingartner, Dalén, Wirta, & Roslin, 2016). Our study  
355 demonstrated various types of interactions among the methods and parameters, including  
356 confirmation, controversy, complementarity and clarification. For instance, proportion of plant  
357 and fungal particles in gut correlated with retention of plant and fungal FAs, respectively. Visual  
358 gut content analysis and FA analysis confirm each other in detection of herbivory and fungivory

359 in Collembola; applying one of the methods may sufficiently trace these feeding strategies.  
360 These two methods explore the trophic niches at different stages in dietary processes. Since  
361 visual gut content analysis detects ingested food particles and the FA method traces assimilated  
362 compounds, it also implies that in these two food categories, Collembola digest mostly the same  
363 material they ingest. Further,  $\Delta^{15}\text{N}$  and  $\Delta^{13}\text{C}$  values were negatively correlated with several  
364 plant-related parameters provided by the other methods, confirming (1) that high  $\Delta^{15}\text{N}$  values  
365 reflect high trophic level as evident from fewer plant particles in the gut, and (2) that high  $\Delta^{13}\text{C}$   
366 values imply feeding on microbially-decomposed organic matter but not freshly-fixed plant  
367 carbon, thus having a lower need for cellulase (A. M. Potapov, Tiunov, & Scheu, 2019; A. M.  
368 Potapov, Tiunov, Scheu, Larsen, & Pollierer, 2019).

369 Controversy was found in fungi- or plant-related parameters between the digestive  
370 enzyme method and the FA method. Although we would expect positive correlations between  
371 chitinase activity (allowing to degrade chitin in fungal cell walls) and fungal FA proportions, or  
372 between cellulase activity (allowing to degrade cellulose in plant and algae cell walls) and plant  
373 FA proportions, neither was the case. To verify correlations of the fungal or plant parameters  
374 derived from the digestive enzyme method with those from visual gut content analysis, however,  
375 more data are needed.

376 Complementarity of the methods was shown in gut fungi and plant- and fungi-  
377 synthesized FAs that were positively correlated with trehalase activity. Since trehalose is the  
378 storage component of fungal, lichen and plant cells, these correlations suggest that Collembola  
379 feeding on plant material and fungi get energy and nutrients from storage, rather than structural,  
380 polysaccharides. In relation to the foraging behaviour, Collembola are more like browsers rather  
381 than grazers (Siepel & de Ruiter-Dijkman, 1993). Another example for complementarity of the

382 methods was negative correlations of activity of trehalase and chitinase with the FAs synthesised  
383 by non-specific and gram-positive bacteria. Apparently, certain Collembola species have a  
384 specific feeding strategy where they rely more on bacterial feeding and thus invest less in the  
385 fungi-related digestive enzymes. Detritivores may largely rely on amino acids synthesised by  
386 free-living or gut symbiotic bacteria if the food quality is low (Larsen et al., 2016; A. M.  
387 Potapov, Tiunov, Scheu, et al., 2019). When repeatedly consuming decomposing litter and soil,  
388 soil animals may collaborate with microorganisms that produce digestive enzymes to release  
389 nutrients from recalcitrant organic compounds – a strategy termed “external rumen” (Swift, Heal,  
390 & Anderson, 1979). This is further supported by positive correlations of  $\Delta^{15}\text{N}$  values and FAs  
391 synthesised by non-specific and gram-positive bacteria, pointing to the utilization of bacterial  
392 symbionts when Collembola feed on soil organic matter. Higher  $\Delta^{15}\text{N}$  values in soil-dwelling  
393 Collembola species likely are due to trophic level inflation by repeated ingestion of soil organic  
394 matter (A. M. Potapov, Semenina, et al., 2016; Steffan et al., 2017). Gram-positive bacteria such  
395 as Actinobacteria and Firmicutes associate with small soil fractions and inhabit small pores  
396 (Hemkemeyer, Dohrmann, Christensen, & Tebbe, 2018; Mummey, Holben, Six, & Stahl, 2006).  
397 These bacteria are thus likely accessible for the soil feeders. Indeed, Actinobacteria dominate in  
398 the gut of *Folsomia candida*, a euedaphic Collembola species specifically adapted to the soil  
399 layer (Zhu et al., 2018).

400 We avoid to further discuss correlations since many of them were based only on few data  
401 points. In summary, our results showed that FA and visual gut content analyses both indicate  
402 herbivory and fungivory but reveal different stages of the dietary processes in Collembola.  
403 Digestive enzyme analysis has a high additional value when combined with others. It gives  
404 further insights in foraging behaviour and animal-microbial interactions: Collembola behave



405 more like browsers, rather than grazers, by feeding on storage polysaccharides from plant  
406 material and fungi. Furthermore, stable isotope analysis estimates trophic level and plant versus  
407 microbial feeding of soil animals and the complementarity of these parameters and the other  
408 trophic parameters indicates that bacterial feeding in Collembola may be more common than  
409 usually assumed.

410

#### 411 *Taxonomic and life form effects on trophic niche parameters*

412 We further explored how the trophic niche parameters vary across different Collembola  
413 taxa and life forms. Taxonomic affiliation explained variation in six out of the fifteen trophic  
414 niche parameters, suggesting that some but not all trophic niche dimensions in Collembola are  
415 phylogenetically structured (A. M. Potapov, Semenina, et al., 2016). The taxonomic groups  
416 explained most variation in stable isotope and in bacteria-related FA parameters. These  
417 parameters are related to biochemical processes in assimilation and thus may in part be  
418 constrained by physiology of phylogenetically-related species (T.-W. Chen et al., 2017). By  
419 contrast, cellulase activity was not related to taxonomic groups, suggesting that the ability of  
420 cellulose degradation might be evolutionary labile in Collembola.

421 Life form, as a proxy for the microenvironment species live in, explained variation only  
422 in three parameters with relatively low  $R^2$  values. Most trophic niche parameters were poorly  
423 related to life form, suggesting that various feeding strategies may be used by species living in  
424 the same microenvironments. However, we found higher values of stable isotopes  $\Delta^{15}\text{N}$  and  $\Delta^{13}\text{C}$   
425 in (hemi)edaphic species, suggesting that dwelling mainly in soil microhabitats results in feeding  
426 strategies that rely mainly on microbially-decomposed organic matter and less on plant materials.

427 By contrast, higher chitinase activity in atmobiotic and epedaphic Collembola suggests that they  
428 need to digest fungal cell walls. Interestingly, atmobiotic species, which mainly inhabit  
429 macrophytes (e.g. on grasses, bushes, trunks and branches of trees), had higher proportions of  
430 gram-negative bacteria FAs, as compared to epedaphic species living in the upper layer of litter.  
431 Bacterial communities in these two microhabitats clearly differ from each other. Collembola  
432 consume gram-negative bacteria such as cyanobacteria (Buse, Ruess, & Filser, 2013; Hao, Chen,  
433 Wu, Chang, & Wu, 2020). The atmobiotic Collembola might feed on corticolous cyanobacteria  
434 and lichens associated with tree bark (A. Singh, Tyagi, & Kumar, 2017). Furthermore, even  
435 within the same Isotomidae family, differences in trophic niche parameters between epedaphic  
436 and (hemi)edaphic species point to environmental determinants of trophic traits in soil animals  
437 (Ponge, 2000; A. M. Potapov, Tiunov, & Scheu, 2019).

438

#### 439 *Multidimensional trophic niches of Collembola*

440 As practical demonstration of the additional value of method combination, below we  
441 attempt to describe the multidimensional trophic niches for high-rank Collembola taxa (**Fig. 3,**  
442 **4**). The order Symphypleona (here representing a conglomerate of several surface-dwelling  
443 families) on average had high proportions of fungal FAs, high chitinase activity in fungal cell  
444 wall degradation, and a high proportion of FAs synthesised by gram-negative bacteria. Lichens  
445 are comprised of fungi and microalgae or cyanobacteria. Cyanobacteria are gram-negative  
446 bacteria able to synthesise hydroxylated FAs (Dembitsky, Shkrob, & Go, 2001; Gugger, 2002).  
447 Together with low  $\Delta^{15}\text{N}$  values typically found in algivores (A. M. Potapov, Korotkevich, &  
448 Tiunov, 2018), our results suggest that lichen grazing, or a combination of fungivory and  
449 herbivory may be widespread among Symphypleona.

450 Tomoceridae had a relatively well-defined trophic niche; the combination of parameters  
451 points to consumption of plant material with the help of bacteria. They had the highest average  
452 proportion of plant particles in the gut among all other Collembola and a relatively high  
453 proportion of plant-synthesised FAs. In addition, they had a high proportion of non-specific  
454 bacteria FAs, suggesting that they may graze on freshly fallen litter and assimilate it with the  
455 help of bacterial symbionts. Clarification of their trophic niche could be advanced with enzyme  
456 analysis, but only one record of *Tomocerus minor* was present in our database – this species had  
457 the highest cellulase activity among all records in the database, confirming plant litter grazing.

458 Entomobryidae include many species that are morphologically resembling Tomoceridae;  
459 however, these two families differed in a number of trophic niche parameters. Entomobryidae  
460 had more fungi in the gut, remarkably high trehalase activity, but lower proportion of non-  
461 specific bacteria FAs. Overall, these differences suggest that Entomobryidae are likely browsers,  
462 rather than grazers. *Lepidocyrtus lignorum* is a good illustration – this species preferred fungi  
463 (**Fig. 2**), having high trehalase, but low cellulase and limited chitinase activity (**Table S2**).

464 Isotomidae had lower average values of fungi-related parameters and lower enzymatic  
465 activity than the previous two families. Isotomidae also had high  $\Delta^{13}\text{C}$  values, pointing to  
466 consumption of organic material in advanced stages of decomposition (A. M. Potapov, Tiunov,  
467 & Scheu, 2019), potentially including invertebrate faeces. All these features were the most  
468 expressed in hemiedaphic and euedaphic species that inhabit decomposing litter and soil (e.g.  
469 *Parisotoma notabilis*; **Fig. 2**). It is likely, that edaphic Isotomidae species rely more on bacteria  
470 via using the “external rumen” feeding strategy. They are usually small and their gut often  
471 contains humus (i.e. soil and faecal material) (Ponge, 2000; Poole, 1959). Interestingly,  
472 hemiedaphic and euedaphic Isotomidae such as species of the genera *Isotomiella*, *Folsomia*,

473 *Parisotoma*, *Folsomides*, dominate in Collembola communities in many ecosystems (M.  
474 Potapov, 2001). Bacterial feeding may be more common in Collembola than it is assumed in  
475 traditional soil food web models (de Vries et al., 2013; Hunt et al., 1987).

476 Hypogastruridae had the highest proportion of fungi and the lowest proportion of plants  
477 in the gut across all other Collembola and low activity of cellulase, suggesting that they feed  
478 selectively on microorganisms and not on plant material. Species of this family were reported to  
479 live and feed on fungi (Sawahata, Soma, & Ohmasa, 2001), and to have relatively high  
480 proportions of fungi-synthesised FAs (Ferlian et al., 2015). Taken together, Hypogastruridae are  
481 fungivores, or, more broadly, microbivores.

482 Neanuridae have no molar plate and are thus unable to chew the food. This family was  
483 long recognised to have a distinct trophic niche from other families (Berg et al., 2004;  
484 Chahartaghi et al., 2005; S. B. Singh, 1969). However, their exact food objects remain enigmatic.  
485 Early studies hypothesised that Neanuridae feed by sucking up the content of fungal hyphae  
486 (Poole, 1959; S. B. Singh, 1969). Stable isotope analysis discovered that Neanuridae have an  
487 outstandingly high trophic level, suggesting that they may feed on other animals, such as  
488 nematodes (Chahartaghi et al., 2005). More recently, Neanuridae were shown to successfully  
489 breed on slime moulds (protists with mycelial stage) (Hoskins, Janion-Scheepers, Chown, &  
490 Duffy, 2015). Slime moulds are abundant in various ecosystems (Swanson, Vadell, & Cavender,  
491 1999) and tend to live in the rotten wood where Neanuridae are also found. Slime moulds,  
492 therefore, potentially serve as food for some species of this family in natural environments. In  
493 our study we showed that Neanuridae have high average values of bacteria- and fungi-related  
494 parameters, high  $\Delta^{13}\text{C}$  and  $\Delta^{15}\text{N}$  values, but very low plant supplementation, supporting the  
495 abovementioned hypotheses. Interestingly, Neanuridae also had a high proportion of FAs

496 synthesized by arbuscular mycorrhizal fungi; however, the same biomarkers can also be  
497 synthesized by bacteria. Neanuridae, as high-level consumers among Collembola, feed on  
498 various groups of microfauna and microorganisms and receive energy from both bacterial and  
499 fungal origins.

500 Onychiuroidea are soil-adapted Collembola without eyes, pigment and furca. They are  
501 associated with plant roots, presumably by feeding on root tips or mycorrhizae (Endlweber,  
502 Ruess, & Scheu, 2009; Fujii et al., 2016; A. M. Potapov, Goncharov, Tsurikov, Tully, & Tiunov,  
503 2016). They had intermediate proportions of plant and fungal particles in their gut in comparison  
504 to other Collembola. However, they had high proportions of FAs synthesised by fungi and non-  
505 specific bacteria and a high cellulase activity. Potentially, Onychiuroidea may feed on both roots  
506 and soil. However, the small-sized Tullbergiidae, which were analysed with Onychiuridae in our  
507 study, may rely less on the root-derived resources (Li et al., 2020). Food resources of this  
508 superfamily call for more studies.

509 Using comparisons across multiple methods, we provide a general overview of  
510 Collembola trophic niches. Our study is the first cross-method compilation of available data on  
511 trophic traits in Collembola that clearly shows advantages of the multidimensional trophic niche  
512 approach. When combined, different methods are not redundant but rather have a high additional  
513 value by compensating drawbacks of each other. Simultaneous application of several methods to  
514 the same population across different groups and ecosystems may improve our understanding of  
515 functioning of the food webs and help to explain species coexistence in cryptic environments,  
516 such as soil.

517

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526

## 527 **Authors' contributions**

528 AMP and TWC developed the idea and study design. SS compiled data on the gut content  
529 analysis, VS compiled data on the digestive enzyme analysis, MMP compiled data on the fatty  
530 acid analysis, AMP compiled data on the stable isotope analysis. AMP and TWC did the analysis  
531 and drafted the manuscript. All authors critically revised the analysis and text.

532

## 533 **Data accessibility**

534 Raw data supporting the results of the study are provided in the Supplementary materials (Table  
535 S1-4) and we intend to archive it to the Dryad digital repository, should the manuscript be  
536 accepted for publication.

537

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## 728 **Supplementary materials**

- 729 • Table S1 – dataset, gut content data (provided separately)
- 730 • Table S2 – dataset, enzyme data (provided separately)
- 731 • Table S3 – dataset, FA data (provided separately)
- 732 • Table S4 – dataset, stable isotope data (provided separately)
- 733 • Table S5 – overlap between different trophic-niche parameters
- 734 • Figure S1 – pairwise correlation between trophic-niche parameters, full version
- 735 • Reference list for the Supplementary materials