DDRP: real-time phenology and climatic suitability modeling of invasive insects

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Short title: DDRP: a new modeling tool for invasive insects

Abstract

Rapidly detecting and responding to new invasive species and the spread of those that are

already established is essential for reducing their potential threat to food production, the

economy, and the environment. We describe a new multi-species spatial modeling platform that

integrates mapping of phenology and climatic suitability in real-time to provide timely and

comprehensive guidance for stakeholders needing to know both where and when invasive insect

species could potentially invade the conterminous United States. The Degree-Days, Risk, and

Phenological event mapping (DDRP) platform serves as an open-source and relatively easy-to-

parameterize decision support tool to help detect new invasive threats, schedule monitoring and

management actions, optimize biological control, and predict potential impacts on agricultural

production. DDRP uses a process-based modeling approach in which degree-days and

temperature stress are calculated daily and accumulate over time to model phenology and

climatic suitability, respectively. Products include predictions of the number of completed

generations, life stages present, dates of phenological events, and climatically suitable areas

based on two levels of climate stress. Species parameter values can be derived from laboratory

and field studies, and from published and newly fitted CLIMEX models. DDRP is written

entirely in R, making it flexible and extensible, and capitalizes on multiple R packages to

generate gridded and graphical outputs. We illustrate the DDRP modeling platform and the

process of model parameterization using two invasive insect species as example threats to United

States agriculture: the light brown apple moth, *Epiphyas postvittana*, and the small tomato borer,

Neoleucinodes elegantalis. We then discuss example applications of DDRP as a decision support

tool, review its potential limitations and sources of model error, and outline some ideas for future

improvements to the platform.

Introduction

Invasive insects in the United States are a significant threat to the economy, environment, food security, and human health [1–3]. They cause billions of dollars in damage to forests each year [1,2], and their potential cost to food crop production is among the highest of any country [3]. Insect invasions in the United States also reduce the abundance and diversity of native species, which negatively impacts ecosystem functions and services such as soil health, nutrient cycling, and wildlife habitat [1,2]. Rapidly detecting and responding to new invasive insects and the spread of those that are already established before they can cause significant economic and environmental damage has therefore become a major priority [2,4].

Modeling climatic suitability (risk of establishment) and the timing of seasonal activities (phenology) of invasive insect species can help stakeholders including farmers, natural resource managers, and surveillance teams detect and prevent their establishment, slow their spread, and manage existing populations more sustainably and economically [5]. Estimates of climatic suitability identify areas to concentrate surveillance or management resources and efforts [6,7], whereas real-time (i.e. current) or forecasted predictions of phenology can improve the timing of surveillance and integrated pest management (IPM) efforts such as monitoring device installation, pesticide applications, and biological control release [8–10]. Additionally, estimates of climatic suitability, phenology, and voltinism (number of generations per year) can help growers predict the impact of pests and diseases on agricultural production and associated economic losses [11].

Degree-day models that predict insect phenology are an established tool for decision support systems that assist stakeholders with scheduling surveillance, monitoring or IPM operations for numerous pest species over the growing season [12–15]. Most degree-day models predict

phenology by measuring linear relationships between temperature and development rate, and

they employ daily time steps to estimate degree-days using daily minimum and maximum

temperature (T_{min} and T_{max}, respectively) data. In the daily time step, degree-days accumulate if

heat exceeds the lower developmental temperature threshold of a species (and below its upper

threshold for some calculation methods) during a 24-hour period [12,13,15]. Several web-based

platforms host degree-day models for insect pest species in the United States, offering users

opportunities to model phenology of multiple species at single locations (site-based model) or

across a certain area (spatialized model). These platforms include but are not limited to Michigan

State University's Enviroweather (https://www.enviroweather.msu.edu), Oregon State

University's USPEST.ORG (https://uspest.org/wea/), the Spatial Analytic Framework for

Advance Risk Information System (SAFARIS; https://safaris.cipm.info/), and the USA National

Phenology Network (https://www.usanpn.org) [8,16].

Despite their widespread use, currently available degree-day modeling platforms are in need

of improvements. None of them integrate predictions of phenology and climatic suitability,

which would provide guidance on the question of both where *and* when—e.g. is an area at high

risk of establishment, and if so, then when will the species emerge or begin a specific activity?

For most species, addressing this two-part question would require finding, potentially

purchasing, and learning how to use two separate platforms. Additionally, many phenology

modeling platforms use oversimplified models that make broad assumptions about insect

biology, such as assuming a single lower developmental temperature threshold for multiple

species, or assuming that an entire population emerges from overwintering at a single time.

However, developmental temperature thresholds may vary widely across insect species, and

development rates often vary within populations [17–19]. A biologically unrealistic model may

produce inaccurate predictions of phenological events (e.g. spring emergence, first adult flight,

egg-hatching) or voltinism. Moreover, most platforms are capable of forecasting phenology only

a week or two into the future in specific states or regions. However, stakeholders may need to

plan operations several weeks in advance, potentially in areas that are outside the geographic

bounds of predictive models.

In this study, we introduce a new spatial modeling platform, DDRP (short for Degree-Days,

establishment Risk, and Phenological event maps) that generates real-time and forecast

predictions of phenology and climatic suitability (risk of establishment) of invasive insect

species in the conterminous United States (CONUS). The objective of DDRP is to improve the

efficiency and effectiveness of programs that aim to detect new or spreading invasive insect

species in the United States, or to monitor and manage species that are already well-established.

The platform is written entirely in the R statistical programming language [20], making it

flexible and extensible, and has a simple command-line interface that can be readily

implemented for online use. Gridded temperature data for DDRP may include the entire CONUS

or a specific region or state, and may be at any spatial resolution that can be handled by the

user's computing system. DDRP will generally use observed and future (forecast or recent

average) temperature data because it was designed to be run as a within-season decision support

tool that can provide guidance on where and when to expect the pest to appear each year, but it

will accept temperature data for any time period. Model products include gridded (raster) and

graphical (map) outputs of life stages present, number of generations, phenological events, and

climatic suitability.

First, we describe the modeling process and workflow of DDRP, summarize types of model

products, and review its system and software requirements. Next, we demonstrate its capabilities

and the process of model parameterization using two invasive insect species which threaten

agricultural biosecurity in the United States: the small tomato borer, *Neoleucinodes elegantalis*

[Guenée (Lepidoptera: Crambidae)], and the light brown apple moth, *Epiphyas postvittana*

[Walker, 1863 (Lepidoptera: Tortricidae)]. These species were chosen because they have been

well-studied in terms of their developmental requirements, and previous climatic suitability

studies provide a basis for parameterizing the climatic suitability model in DDRP. Additionally,

models for these species are intended to aid surveillance teams at the Cooperative Agricultural

Pest Survey (CAPS) pest detection program, which supports the USDA Animal and Plant Health

Inspection Service (APHIS) as it works to safeguard agricultural and environmental resources in

the United States. We used population monitoring data for *E. postvittana* in California to test the

hypothesis that DDRP can correctly predict the timing of first spring egg laying and the

generation length of the species. Additionally, we used a validation data set consisting of

presence localities from California (*E. postvittana*) and Brazil (*N. elegantalis*) to test the

hypothesis that DDRP can correctly predict each species' known distribution in these areas. The

DDRP platform will be a useful decision support tool for preventing, monitoring, and managing

new and existing invasive pests of agriculture and natural resources in the United States.

Methods

1) Model inputs

Temperature data

DDRP requires daily T_{min} and T_{max} data in a gridded format for an area of interest in CONUS

(Fig 1). For real-time modeling, we have been using daily T_{min} and T_{max} data at a 4 km spatial

resolution from the PRISM (Parameter-elevation Relationships on Independent Slopes Model)

database (available at https://prism.oregonstate.edu) [21]. Daily PRISM data become available

ca. 1 day after weather station observations are reported, and are typically updated and improve

in quality as more observations are added (see PRISM website for details). The phenology

mapping system of the USA National Phenology Network [8] uses Real-Time Mesoscale

Analysis (RTMA) weather data at a 2.5 km resolution, which are available within hours after

data are observed. The daily T_{min} and T_{max} RTMA data set could potentially be used in DDRP;

however, the RTMA methodology lacks PRISM's update and quality control regimes [21].

Another alternative is Daymet v3, which offers daily climate data for North America, Hawaii,

and Puerto Rico at a very high spatial resolution (1 km) (https://daymet.ornl.gov) [22]. However,

Daymet data are released months after the end of each year, so they would be less useful for

within-season modeling and decision support.

Fig 1. Schematic of the DDRP model framework. 1) Input data sets (blue shaded boxes)

include a) data on the developmental requirements, climatic tolerances (optional), and emergence

time(s) of population cohort(s) of a species (Table 1), and b) daily minimum and maximum

temperature data (T_{min} and T_{max}, respectively). 2) Hollow blue boxes indicate calculations

conducted on each daily time step, where a dashed outline represents calculations for climatic

suitability. Phenological event map (PEM) calculations for each life stage (E = egg, L = larva, P

= pupa, A = adult) are shown in green font. A full generation is counted when adults lay eggs (in

red), and the number of generations subsequently increases. 3) After the daily time step

completes, DDRP combines the results across all cohorts and exports the model outputs as multi-

layer raster and summary map (".png") files (orange shaded boxes). Orange shaded boxes with a

dashed line represent model outputs for PEMs and climatic suitability.

For forecast modeling, DDRP is currently configured to use either monthly-updated, daily-

downscaled NMME (North American Multi-Model Ensemble) 7-month forecasts at a 4 km

resolution [23], or recent 10-year average PRISM daily data that are calculated on a bi-monthly

basis. We consider 10-year average data to be an improvement over 30-year climate normals for

producing forecasts because temperatures in CONUS have significantly increased over the past

30 years [24,25]. The match of mean forecasts of the NMME model's ensemble to the observed

value (i.e. skill) varies both spatially and temporally due to topography, season, and the presence

of an El Niño-Southern Oscillation (ENSO) signal [26,27]. It may therefore be more

conservative, and provide more consistent predictions, to use 10-year averages instead of NMME

data to avoid potential issues with skill. However, we caution that the 10-year average data do

not simulate variation in daily T_{min} and T_{max}, which may result in the under-prediction of degree-

day accumulation in the spring or fall as daily T_{max} only slightly exceeds the lower

developmental threshold of a species, or for cooler sites that have temperatures that are often

near the threshold. We have also prepared and plan to use the National Weather Service gridded

National Digital Forecast Database (NDFD) 7-day forecasts

(https://www.weather.gov/mdl/ndfd_info) [28] for use in DDRP.

Phenology modeling: species data and parameters

The life history and behavior of a target species must be considered for appropriateness to model

in DDRP. In its current form, the platform can model four separate life stages (the egg, the larva

or nymph, the pupa or pre-oviposition, and the adult) plus a separately parameterized

overwintering stage. As movement and migration are not handled by DDRP, it is currently

limited in its ability to model migratory species, such as those that may establish in southern

areas of their potential range and migrate yearly to more northern areas. Species that lack an

overwintering stage, which are common in tropical and subtropical areas, may be difficult to

model because the timing of first spring activities and stages present cannot be accurately

estimated. Currently DDRP is entirely temperature-driven, so species whose growth and

reproduction are strongly influenced by additional environmental factors such as day length or

moisture may not be accurately modeled.

DDRP requires data on the developmental temperature thresholds (in either degrees Celsius

or Fahrenheit) and durations for each life stage of an insect species in degree-days (Fig 1 and

Table 1). These data are typically collected in the laboratory by measuring how temperature

influences the rate of development, although data derived from season-long monitoring studies

are also used [14,29]. A different developmental threshold may be assigned to each stage,

although we typically solve for a common threshold if differences across the stages are minimal.

Presently, the model depends upon a fixed starting date such as January 1, specified by the user

for the entire region of interest. The duration of the overwintering stage represents the number of

degree-days that must accumulate from the start of the year for the stage to complete.

Table 1. **Species-specific parameters used in DDRP with corresponding values for *Epiphyas postvittana* (light brown apple**

moth) and *Neoleucinodes elegantalis* (small tomato borer).

Parameter	Code	*E. postvittana*	*N. elegantalis*
Lower developmental thresholds (°C)			
Egg	eggLDT	7.2	8.89
Larvae	larvaeLDT	7.2	8.89
Pupae	pupaeLDT	7.2	8.89
Adult	adultLDT	7.2	8.89
Upper developmental thresholds (°C)			
Egg	eggUDT	31.1	30
Larvae	larvaeUDT	31.1	30
Pupae	pupaeUDT	31.1	30
Adult	adultUDT	31.1	30
Stage durations (°C degree-days)			
Egg	eggDD	127	86

Table 1 cont.

Larvae	larvaeDD	408	283
Pupae	pupDD	128	203
Adult	adultDD	71	96
Overwintering larvae	OWlarvaeDD	varies	–
Overwintering adult	OWadultDD	–	varies
Phenological events (°C degree-days)			
Overwintering stage event	OWEventDD	varies	varies
Egg event	eggEventDD	126	80
Larvae event	larvaeEventDD	203	140
Pupae event	pupaeEventDD	128	100
Adult event	adultEventDD	22	55
Cold stress			
Cold stress temperature threshold (°C)	coldstress_threshold	3	6
Cold degree-day (°C) limit when most individuals die	coldstress_units_max1	875	1150
Cold degree-day (°C) limit when all individuals die	coldstress_units_max2	1125	1600

Table 1 cont.

Heat stress			
Heat stress temperature threshold (°C)	heatstress_threshold	31	32
Heat stress degree-day (°C) limit when most individuals die	heatstress_units_max1	375	750
Heat stress degree-day (°C) limit when all individuals die	heatstress_units_max2	550	1000
Cohorts			
Degree-days (°C) to emergence (average)	distro_mean	210	50
Degree-days (°C) to emergence (variance)	distro_var	2500	1500
Minimum degree-days (°C) to emergence	xdist1	100	0
Maximum degree-days (°C) to emergence	xdist2	320	111
Shape of the distribution	distro_shape	normal	normal
Degree-day calculation method	calctype	triangle	triangle

Phenology model parameter values were derived from previous studies and an analysis of published data for *E. postvittana* [30–32],

and from an analysis of published data for *N. elegantalis* [33]. Climatic suitability model parameter values were estimated by

calibrating models in accordance with outputs of a CLIMEX model for each species. For both species, the phenological events for

egg, larvae, and adults are beginning of egg hatch, mid-larval development, and first egg-laying, respectively. The phenological event

for pupae is first adult emergence for *E. postvittana* and mid-pupal development for *N. elegantalis.* The duration and timing of the

phenological event for the overwintering stage will vary according to the number of cohorts applied in model runs (see text for

details). Both models applied the single triangle method ("triangle") with upper threshold to calculate degree-days.

Users must specify the number of degree-days that are required for the overwintering stage to

complete development and emerge for the growing season. These data are typically gathered

using field monitoring studies, whereby the temporal distribution of emergence times and

number of individuals that emerge on a given date is documented [e.g. 32]. Assigning a single

value to the overwintering stage duration parameter would assume that an entire population

develops simultaneously, which may not be biologically realistic because several intrinsic (with

a genetic basis) and extrinsic (e.g. microclimate, nutrition) factors can produce variation in

development rates within a species [19,35]. Indeed, phenology models that incorporate

developmental variability in a population may have increased predictive power [17,19,36].

DDRP therefore allows the duration of the overwintering stage to vary across a user-defined

number of cohorts (groups of individuals in a population). Much of the intrinsic variability in

insect development during a generation often occurs in the first stadium after overwintering [37],

although developmental variation may occur in any life stage [17,38,39]. DDRP uses five

parameters to generate a frequency distribution of emergence times: the mean, variance, low

bound, and high bound of emergence times, and the shape of the distribution (Gaussian or

lognormal; Table 1). The platform uses these data to estimate the relative size of the population

represented by each cohort, which initializes the population distribution that is maintained during

subsequent stages and generations. Individuals within each cohort develop in synchrony.

Users may specify the timing (in degree-days) of phenological events that are important to

their target system to generate phenological event maps in DDRP, which depict the estimated

calendar dates of the event over a time frame of interest. We typically generate phenological

event maps based on temperature data for an entire year so that events for multiple generations of

each of the five life stages are modeled. For example, phenological event maps that depict when

the overwintering stage would emerge may be useful for identifying start dates for surveillance

operations for a species, whereas maps for subsequent generations could help with planning

operations later in the year. The timing of phenological events may be based on life stage

durations (e.g. the end of the egg stage signifies egg-hatching), or on occurrences within a stage

such as the midpoint or peak of oviposition or adult flight. Currently, one user-defined

phenological event for each life stage for up to four generations may be modeled, although the

platform could be modified to predict multiple events for each stage (e.g. first, midpoint, and end

of the stage) for any numbers of generations.

Climatic suitability modeling: species data and parameters

Climatic suitability modeling in DDRP is based on cold and heat stress accumulation and

requires data on temperature stress threshold and limits of a species (Fig 1 and Table 1). While

parameter values may be estimated from laboratory or field experiments, such data are lacking

for most species. Additionally, extrapolating laboratory data to the field to project accumulation

of stress is difficult due to oversimplification of the number of variables and the temporal and

spatial variation in natural environments [40]. We have been using the CLIMEX software [41]

(Hearne Scientific Software, Melbourne, Australia), which is one of the most widely used

species distribution modeling tools for agricultural, livestock and forestry pests and non-pests

[6,7], to assist with climatic suitability model parameterization in DDRP. Laboratory collected

data may help with parameterizing a CLIMEX model; however, model parameters are fine-tuned

and the model is fitted using observations from the species' known geographical distribution

[41,42].

DDRP was designed to be complementary to CLIMEX in several ways to facilitate climatic

suitability model parameterization, but the two programs also differ in several respects (Table 2).

Both platforms use a process-based modeling approach in which parameters that describe the

response of a species to temperature stress are included in calculations of climatic suitability.

Certain model products, particularly maps of temperature stress accumulation, are therefore

directly comparable. DDRP uses the stress accumulation method of CLIMEX in which cold and

heat stress units begin to accumulate when temperatures exceed the cold and heat stress

temperature thresholds, respectively. In both platforms, cold stress units are calculated as the

difference between T_{min} and the cold stress temperature threshold, and heat stress units are

calculated as the difference between T_{max} and the heat stress temperature threshold. However,

DDRP uses daily temperature data and stress accumulates linearly over time, whereas CLIMEX

uses average weekly temperature data and stress accumulates at a weekly rate that becomes

exponential over time (each week's stress in multiplied by the number of weeks since the stress

first exceeded zero) [41]. These differences appear to be minor in using CLIMEX outputs and

parameters to support calibration of DDRP parameters, as demonstrated in our case studies (see

'Case Studies'). Similar to CLIMEX, DDRP uses a single cold and heat stress temperature

threshold for all life stages, and stress units accumulate across the entire time period of interest

(i.e. across all life stages and generations). We apply the cold and heat stress threshold of the life

stage that would be most likely to experience the coldest or hottest temperatures of the year,

respectively. DDRP assumes that stress could indirectly kill individuals by restricting their

activity, or directly cause mortality through extreme cold or heat events such as a hard freeze.

Table 2. Comparison of the characteristics, parameters, and products of climatic suitability models in CLIMEX and DDRP.

Attributes	DDRP	CLIMEX
Temporal range (time frame)	Any – historical, real-time, near forecast, and climate change forecasts	Historical (1961–1990), and future climate change forecasts for 2030, 2050, 2070, 2080, 2090, and 2100
Temporal scale (time step)	1-day (daily) for PRISM data – others potentially accommodated	Typically weekly values interpolated from monthly data
Spatial scale	Any – default is a 2′ resolution (~4 km) for PRISM data	CliMond data at a 30′ (~55 km at equator) or 10′ (~20 km) resolution; others potentially accommodated
Factors influencing climatic suitability	Cold and heat stress	Cold, heat, dry, and wet stress plus population growth
Modeling process overview	Estimates daily cold and heat stress accumulation and determines whether total accumulations exceed the moderate (max1) or severe (max2) cold and heat stress limits	Estimates weekly population growth and the accumulation of stress (cold, heat, dry, and wet); population growth is reduced when daily accumulations are too low or too high to maintain metabolism

Table 2 cont.

Climate stress parameters		
Temperature thresholds	Upper and lower cold and heat stress temperature thresholds in Celsius units	Upper and lower cold and heat stress thresholds in Celsius or degree-day units
Temperature stress rates	Cold and heat stress accumulation limits (max1 and max2); stress units accumulate linearly over time (consecutive days not weighted higher than non-consecutive days)	Weekly cold and heat stress accumulation rate based on thresholds in Celsius units (similar to DDRP) or degree-day units; stress units accumulate exponentially over time (consecutive weeks are weighted higher than non-consecutive weeks).
Moisture stress thresholds	None	Upper and lower dry and wet stress thresholds
Moisture stress rates	None	Weekly dry and wet stress accumulation rate for species
Total no. of parameters possible	6	38
Total no. of parameters typically used	6	21

Table 2 cont.

Depiction of climatic suitability	Areas not under moderate or severe cold and heat stress exclusions	Typically areas with a high ecoclimatic index (> 20 or 30), which is calculated using annual growth and stress indices
Products	Gridded and summary map outputs of 1) cold and heat stress unit accumulation, and 2) cold, heat, and all (cold plus heat) stress exclusions	Tabular and summary map outputs of 1) cold, heat, dry, and wet stress unit accumulation, and 2) the temperature, moisture, growth, and ecoclimatic index

For simplicity, we do not show CLIMEX parameters related to interaction stress indices (hot-wet stress, hot-dry stress, cold-wet stress,

and cold-dry stress) or to radiation, substrate, light and diapause indices.

Importantly, DDRP was designed to model climatic suitability based on daily current or

forecast temperature data at fine spatial scales (e.g. a single state or region), which would give

users insight into the potential risk of establishment or spread during a particular season or year.

In contrast, CLIMEX is normally used to estimate a species' potential distribution using coarse-

scale 10′ and 30′ resolution global gridded 30-year monthly climate normals centered on 1975

(1961–1990) or future projections from selected global circulation models (GCMs) [43]. In

theory CLIMEX could be used for real-time climatic suitability, but it has no native ability to

import and process common gridded formats and is incapable of using daily resolution climate

data. Thus, DDRP's climate suitability models are intended to improve the efficiency of

surveillance and trap deployment at a relatively small focal area for a current or near-future time

period, whereas CLIMEX models provide a more general and coarse-scale assessment of

suitability based on averaged climate data.

Relying on real-time climatic suitability models for decision support on where to employ pest

management and eradication operations for a given year or season is preferable to using models

based on 30-year climate normals. A model that uses current climate data is more biologically

relevant because the risk of establishment in an area would be affected by the conditions that a

species physically experiences, not by averages of historical climate. Additionally, climate in

CONUS is changing rapidly, so models based on climate normals may produce unrealistic

predictions of present-day climatic suitability. Over the past *ca*. 30 years, the average annual

temperature in CONUS has increased by 1.2°F (0.7°C), the number of freezing days has

declined, and extreme temperature events have increased in frequency and intensity [24,25].

Nonetheless, DDRP is not currently capable of including moisture factors in the modeling

process like CLIMEX, so model predictions for moisture-sensitive species in very arid or wet

283 areas should be interpreted with caution. We discuss the potential implications of generating a

284 climate suitability model based solely on temperature in the 'Discussion.'

285  We compare CLIMEX's predictions of temperature stress accumulation and overall climate

286 suitability to similar products in DDRP to help parameterize a DDRP climatic suitability model.

287 Temperature stress thresholds may be calibrated so that predictions of cold and heat stress

288 accumulation at the end of the year are spatially concordant with CLIMEX's predictions.

289 Climatic suitability in CLIMEX is estimated with the Ecoclimatic Index (EI), which is scaled

290 from 0 to 100, and integrates the annual growth index and climate stress indices to give an

291 overall measure of favorableness of a location or year for long-term occupation by the target

292 species [41,42]. Typically an EI approaching 0 indicates an unfavorable climate for long-term

293 survival, while an EI exceeding 20 or 30 (depending on the species) indicates a highly suitable

294 climate [41,42]. As discussed in more detail in 'Case Studies', temperature stress limits in DDRP

295 can be adjusted so that areas predicted to be suitable by CLIMEX are also included in DDRP's

296 prediction of the potential distribution.

297  Comparing DDRP climatic suitability model products to those of CLIMEX for model fitting

298 purposes is naturally more appropriate when temperature data are derived from the same time

299 period. We have therefore been using a PRISM T_{min} and T_{max} 30-year average data set centered

300 on 1975 (1961−1990) to match the time-schedule of the CliMond CM10 (also 1961−1990) world

301 climate data set currently supplied with CLIMEX [43]. We temporally downscaled monthly

302 PRISM estimates for 1961−1990 because DDRP requires daily data and PRISM daily

303 temperature data for years prior to 1980 are not available. For each month of a given year, a

304 bilinear interpolation method was used to assign each day an average temperature value that was

305 iteratively smoothed and then adjusted so that the monthly averages were correct.

2) Daily time step

DDRP models insect phenology and climatic suitability by stepping through each day of a

specified time period and calculating degree-day and temperature stress accumulation at each

grid cell of a focal area (Fig 1). The time period may span the entire year of interest, or include

only a subset of days such as those during the growing season. Users may sample and save daily

modeling results every 30 days, 14 days, 10 days, seven days, two days, or one day. Results are

saved in multi-layer rasters that are processed and analyzed after the daily time step to produce

final model products. We describe the phenology and climatic suitability modeling process and

products in more detail in the following sections.

Phenology model

DDRP calculates daily degree-days over the specified time period using developmental

temperature threshold information and gridded temperature data that have been cropped to the

extent of the focal area (Fig 1). Currently DDRP has three methods to calculate degree-days: a

simple average, the simple average using an upper threshold with a horizontal cutoff, and the

single triangle method with upper threshold [44–46]. All three methods calculate degree-days

from the daily T_{min} and T_{max} data and a specified base (lower developmental threshold)

temperature, but the latter two methods also include the upper developmental threshold in

calculations. The single triangle method is also used as a close approximation to the more

complex sine-curve calculation method [46].

With the exception of phenological event maps, which are computed only for the last day of

the daily time step, DDRP saves the following phenology model results for each sampled day:

1. *Accumulated degree-days.* While daily degree-days are calculated for each life stage, the

cumulative degree-days are summed only for the first cohort of the larval stage, as these

degree-day maps are representative for all cohorts and life stages. Accumulated degree-days

calculated for larva will be the same for other life stages if common developmental

thresholds are used.

2. *Life stages*. The life stage present (overwintering stage, eggs, larvae, pupae, and adults) for

each cohort.

3. Number of generations. The current generation count for each cohort. If the model is run for

an entire year, then the output for the last day of the year would represent the potential

voltinism of the species. The generation count increases when adults progress to the egg

stage (i.e. oviposition occurs).

4. *Phenological event maps (optional).* The timing of phenological events is estimated by

computing daily degree-day totals from the gridded temperature data, and storing the day of

year when an event threshold is reached. Event results are generated only on the last day of

the daily time step (typically, the last day of the year) because the entire time period must be

analyzed for all potential event days to be considered.

Climatic suitability model

Independently from the phenology model simulations, DDRP calculates daily cold and heat

stress accumulations and compares these estimates to user-defined moderate and severe stress

limits to delineate the potential distribution of the species (Fig 1). We opted to use moderate and

severe stress limits to reflect two distinct themes. First, they may provide a way to depict the

potential for short term vs. longer term establishment. For most species, the potential distribution

could be represented by areas where cold and heat stress have not exceeded the severe or

moderate stress limits, as these should allow for long-term population persistence. DDRP depicts

these areas with maps of cold stress exclusion, heat stress exclusion, and all stress exclusion

(cold plus heat stress exclusions; Fig 1). Areas under moderate stress exclusion may represent

temporary zones of establishment in which a species establishes only during a favorable season,

such as after an annual migration event. Conversely, areas under severe stress exclusion do not

allow for even short-term establishment. Typically we visualize exclusion maps calculated for

the last day of the year (day 365) under investigation to provide insight into the potential

distribution for an entire growing season. For CONUS, the northern range limit is typically

delineated by cold stress and the southern range limit, if any, is delineated by heat stress.

Second, using two levels of stress may provide a way to represent uncertainty for estimating

the potential distribution. As discussed in more detail in the 'Discussion,' several sources of

uncertainty and error in the modeling process may bias model predictions, such as applying

inappropriate parameter values, using climate data with low skill or poor spatial resolution,

ignoring biotic factors such as species interactions, or ignoring non-temperature abiotic factors

such as microclimate effects, moisture, and photoperiod [35,47]. Defining the potential

distribution as areas under severe stress only would typically provide a broader estimate than a

definition based on both stress levels. While this approach may over-predict the risk of

establishment, conducting surveys over too broad an area is probably better than surveying too

small of an area, which may allow a new invasive species to establish and spread.

3. Post time step processing

After the daily time step has completed, DDRP combines and analyzes results across cohorts and

generates final multi-layer rasters and summary maps ("".png" image files) for each sampled day.

If multiple cohorts were modeled, then DDRP uses estimates of the relative size of the

population represented by each cohort to calculate the relative size of the population (totaling

100%) in any given life stage and generation. For phenological event maps, the earliest and

average day of year that an event occurs across cohorts is calculated, and these are displayed as

calendar dates (month-day) on summary map outputs (Fig. 1).

DDRP integrates mapping of phenology and climate suitability so that users can use a single

model product to obtain guidance on their "where" and "when" questions (Fig 1). For example, a

user involved with planning surveys may want to know where a target species may establish, and

within those areas, when populations may emerge from over-wintering. Each output of the

phenology model with the exception of accumulated degree-days will be associated with two

additional products for each sampled day (or the last day for a phenological event map): 1) one

that includes severe stress exclusion only, and 2) one that includes both severe and moderate

stress exclusions. For example, a phenological event map with severe and moderate stress

exclusions for 2018 (all 365 days) would present predicted dates of the selected event only in

areas where long-term establishment is predicted (Fig 2).

Fig 2. Phenological event maps generated by DDRP for (A) *Epiphyas postvittana* **(light**

brown apple moth) and (B) *Neoleucinodes elegantalis* **(small tomato borer) in CONUS in**

2018. The map for *E. postvittana* shows the average date of egg-laying by first generation

females, whereas the map for *N. elegantalis* shows the average date of first generation beginning

of egg hatch. Both maps include estimates of climatic suitability, where long-term establishment

is indicated by areas not under moderate (excl.-moderate) or severe (excl.-severe) climate stress

exclusion.

System and software requirements

DDRP requires the R statistical software platform and can be run from the command line or

within RStudio [48]. It takes advantage of functions from several R packages for data

manipulation, analysis, and post-model processing. The raster package [49] is used to crop daily

temperature rasters to the focal area, store and manipulate daily loop raster results, and process

and further analyze results for each cohort. Many non-spatial data manipulations are conducted

with functions in the dplyr, tidyr, and stringr packages [50–52]. The ggplot2 package [53] is used

to generate and save summary maps of raster outputs, and options from the command line

argument are parsed using the optparse package [54].

DDRP capitalizes on the multi-processing capabilities of modern servers and computers to

run multiple operations in parallel, which is made possible with the R packages parallel and

doParallel [55]. This significantly reduces computation times, particularly in cases where

modeling is conducted with multiple cohorts and across large areas. For example, parallel

processing is used to crop rasters for multiple dates, run multiple cohorts in the daily time step,

and to analyze time step outputs for multiple days or files simultaneously. For very large areas

(currently defined as the Eastern United States and CONUS), temperature rasters are split into

four tiles and both the tiles and cohorts are run in parallel in the daily time step.

We recommend running DDRP on a server or computer with multicore functionality because

certain processes are very memory intensive and may execute slowly or stall without parallel

processing. The platform was designed to run on a Linux server, but we have also successfully

used it on a Windows OS with eight cores. As an example, a model that was run on a Linux

server with 48 cores completed *ca.* 3.5 times faster than it did on a Windows OS with eight

cores.

Case studies

Climatic suitability, voltinism, and phenological events **in Epiphyas postvittana**

The light brown apple moth, *E. postvittana* (Walker 1863) (Lepidoptera: Tortricidae), is a

leafroller pest native to southeastern Australia, including Tasmania [30]. The species invaded

Western Australia, New Zealand, New Caledonia, England, and Hawaii more than 100 years ago

[56–58], and has been established in California since 2006 [59,60]. It poses a significant threat to

agricultural production in the United States because it feeds on more than 360 host plants,

including economically important fruits such as apple, pear, citrus and grapes [30,31,61]. For

example, an economic risk analysis of *E. postvittana* to four major fruit crops (apple, grape,

orange, and pear) in CONUS estimated an annual mean cost of US$105 million associated with

damage to crops and control, quarantine, and research [61]. The CAPS program at APHIS

conducts annual surveys for *E. postvittana* at various counties across CONUS.

A summary of phenology and climatic suitability model parameters used for *E. postvittana* in

DDRP is reported in Table 2. We assigned all life stages a lower developmental threshold of

7.1°C (45°F) [30] and an upper developmental threshold of 31.1°C (88°F). Although DDRP

allows for different temperature thresholds for each stage, the site-based phenology modeling

tool that we use at USPEST.ORG requires common thresholds, and these are presented as whole

numbers in Fahrenheit scale for easy interpretation by end-users. Building the model for both

platforms keeps models simpler and facilitates cross-comparison. Additionally, laboratory

studies revealed small differences in the lower developmental threshold ($< 1°C$) across different

life stages [30,31]. The upper developmental threshold value is based on studies showing that all

life stages cease development between $31–32°C$ [30–32]. We derived life stage durations (in

degree-days Celsius; hereafter, DDC) for *E. postvittana* based on an analysis of published data

[30], which resulted in 127, 408, 128, and 71 DDC for eggs, larvae (females on young apple

foliage), pupae, and adults to 50% egg-laying, respectively (S1 Appendix).

We set the overwintering stage to larva because the predominant overwintering stage of *E.*

postvittana in the United States are the late larval instars [34,62]. We applied seven cohorts to

approximate a normal distribution of emergence times that spanned 100 to 320 DDC (average =

210 DDC) based on a report that overwintering larvae at four sites in California required

between 102 and 318 DDC to finish development [34]. This would correspond to the time

required for mid-stage (3rd−5th instars, average 4th instar) female larval feeding on old foliage

(0.45×494 DDC = 210 DDC), after a January 1 start date. The single triangle method was used

to calculate degree-days.

We tested the hypothesis that DDRP can correctly predict the timing of first spring egg

laying and generation length for *E. postvittana* by analyzing three monitoring data sets that were

collected in California over a 12 year time frame (2008–2009, 2011–2014, and 2019–2020). For

each data set, we estimated the difference between the date of the peak in first spring flight and

DDRP predictions of the average date of first spring egg laying. This event was chosen because

peak flight would likely happen at about the same time that peak egg laying is occurring [63].

Additionally, we used DDRP to calculate the number of degree-days that accumulated between

the last peak fall flight and first peak spring flight for four winters (2011–2012, 2012–2013,

2013–2014, and 2019–2020), which should serve as a rough estimate of generation time. These

estimates were compared to the generation time that is used by the DDRP model (823 DDC for

the overwintering generation; 734 DDC are used for later generations that have young foliage to

feed upon [30]). Model runs for each year used PRISM data and applied seven cohorts and a

normal distribution of emergence times. Additional details about the data sets and methods for

this analysis are presented in S2 Appendix.

We generated a CLIMEX model for *E. postvittana* using CLIMEX version 4.0 [41] to help

parameterize the climatic suitability model in DDRP. The model applied a combination of

parameter values (Table 3) derived from two previous CLIMEX studies of this species [64,65].

However, we used a cold stress threshold (TTCS) of 3°C, which is lower than He et al.'s (2012)

value (5°C)[64], and higher than Lozier and Mill's (2011) value (1.5°C) [65]. We applied a top-

up irrigation (additional simulated rainfall) rate of 2.5 mm day^{-1} for the winter and summer

season because irrigation mitigates the hot-dry climate that limits distribution of *E. postvittana*

within CLIMEX. We fit the CLIMEX model using a data set of 393 georeferenced locality

records from Australia (N = 317) and New Zealand (N = 76), which were obtained from

GBIF.org (18th July 2019; GBIF Occurrence Download https://doi.org/10.15468/dl.a4ucei) and

Nick Mills at UC Berkeley (pers. comm.).

For model validation, we used locality records from 14,949 sites in California where the

California Department Food and Agriculture detected *E. postvittana* between 2007 and 2013, and

from an additional 47 localities in California where it was positively diagnosed between 2011

and 2017. We removed localities that occurred within the same CLIMEX grid cell, which

resulted in 140 unique localities. Of these, 95% (133/140) were in areas predicted to have high

suitability (EI > 20), 4% (6/140) were in areas predicted to have low suitability (0 < EI < 20),

and 0.007% (1/140) were in unsuitable areas (S1 Fig). These findings suggest that the CLIMEX

model accurately predicted suitable climates at the majority of localities where the species is

known to be established in CONUS, and it is therefore a useful tool for calibrating a climatic

suitability model in DDRP.

Table 3. Parameter values used to produce a CLIMEX model for *Epiphyas postvittana* (light brown apple moth) and

***Neoleucinodes elegantalis* (small tomato borer)**.

CLIMEX parameter	Code	*E. postvittana*			*N. elegantalis*	
		Lozier & Mills (2011)	He et al. (2012)	This study	da Silva et al. (2018)	This study
Temperature						
Limiting low temperature (°C)	DV0	7.5	7	7	8.8	8.8
Lower optimal temperature (°C)	DV1	15	13	13	15	15
Upper optimal temperature (°C)	DV2	25	23	23	27	27
Limiting high temperature (°C)	DV3	31	30	31	30	31
Degree-days per generation (°C days)	PDD	673.6	673.6	673.6	588.2	588.2
Moisture						
Limiting low moisture	SM0	0.15	0.25	0.15	0.35	0.35
Lower optimal moisture	SM1	0.5	0.8	0.8	0.7	0.7
Upper optimal moisture	SM2	0.8	1.5	1	1.5	1.5

Table 3 cont.

Limiting high moisture	SM3	1.4	2.5	2.5	2.5	2.5
Cold stress						
Cold stress temperature threshold (°C)	TTCS	1.5	5	3	–	6
Cold stress temperature rate (week^{-1})	THCS	−0.005	−0.0005	−0.0005	–	−0.0005
Cold stress degree-day threshold (°C days)	DTCS	–	–	–	15	–
Cold stress degree-day rate (week^{-1})	DHCS	–	–	–	0.001	–
Heat stress						
Heat stress temperature threshold (°C)	TTHS	31	31	31	30	31
Heat stress temperature rate (week^{-1})	THHS	0.0045	0.01	0.002	0.0007	0.00084
Dry stress						
Dry stress threshold	SMDS	0.15	0.2	0.15	0.35	0.35
Dry stress rate (week^{-1})	HDS	−0.005	−0.01	−0.005	−0.001	−0.001
Wet stress						
Wet stress threshold	SMWS	1.4	2.5	2.5	2.5	2.5
Wet stress rate (week^{-1})	HWS	0.001	0.002	0.001	0.002	0.002

In DDRP, we generated a climatic suitability model for *E. postvittana* using the daily

downscaled PRISM T_{max} and T_{min} estimates for 1961–1990 and calibrated model parameters in

accordance with the CLIMEX model (Fig 3). Specifically, we compared maps of temperature

stress accumulation, and adjusted temperature stress limits so that most areas predicted to be

under moderate and severe climate stress by DDRP had low $(20 > EI > 0)$ or zero $(EI = 0)$

suitability according to CLIMEX, respectively. Additionally, we modeled climatic suitability for

the species in California for each year between 2010 and 2019 and calibrated climatic stress

parameters to maximize the fit of the model to 144 georeferenced records that were collected

over the same time period $(N = 144$ from GBIF, $N = 77$ from Nick Mills). For each modeled

time period, we extracted climatic stress exclusion values for presence localities in the California

Department Food and Agriculture data set to test our hypothesis that DDRP could correctly

predict the potential distribution of the species. We removed localities that occurred within the

same 4km^2 grid cell, which resulted in 872 unique validation localities.

Finally, we modeled phenology and climatic suitability for *E. postvittana* in 2018 to provide

insight into its potential voltinism, seasonal activities, and risk of invasion in particularly warm

temperatures. The summer of 2018 in the United States was the warmest since 2012 and tied for

the fourth-warmest on record (NOAA website https://www.noaa.gov/news/summer-2018-

ranked-4th-hottest-on-record-for-us last accessed 11/21/19). We generated a phenological event

map that depicted the date of first egg laying by first generation females, because this activity is

relevant to monitoring both eggs and the emergence of adults, which typically occurs two to

three days prior to egg laying.

Fig 3. Predictions of cold stress, heat stress, and climatic suitability for *Epiphyas postvittana*

(light brown apple moth) in CONUS produced by CLIMEX (A–C) and DDRP (D–F) based

on 1961–1990 climate normals. Climatic suitability is estimated by the Ecoclimatic Index (EI)

in CLIMEX, and by combining cold and heat stress exclusions in DDRP. In DDRP, long-term

establishment is indicated by areas not under moderate (excl.-moderate) or severe (excl.-severe)

climate stress exclusion. Cold and heat stress units in DDRP were scaled from 0 to 1000 to

match the scale in CLIMEX. CLIMEX maps were generated for this study based on parameters

documented in Table 3 and have not been previously published.

Climatic suitability, voltinism, and phenological events **in Neoleucinodes elegantalis**

The small tomato borer, *N. elegantalis* (Guenée) (Lepidoptera: Crambidae), is native to South

America and is distributed throughout the Neotropics including in Mexico, Central America, and

the Caribbean [66,67]. A major insect pest of tomato (*Solanum lycopersicum*), it also attacks

fruits of other plants belonging to the family Solanaceae including eggplant, paprika, naranjilla,

and green and red pepper [67]. There are at least 1175 recorded interceptions of the species from

the United States, where it is considered a serious threat to agricultural biosecurity because it

lowers tomato production in South America [68]. The CAPS program has conducted surveillance

for *N. elegantalis* since at least 2011.

A summary of phenology and climatic suitability model parameters used for *N. elegantalis* in

DDRP is reported in Table 2. We re-analyzed data from a laboratory study on the development

of *N. elegantalis* on hybrid tomato (Paronset) at five temperatures [33] to estimate a common

lower temperature threshold for all life stages, which involved adding a point to force the x-

intercept to an integer value in degrees Fahrenheit. We weighted the analysis to select a common

lower threshold for immature stages, which are the longest in duration, because this should

produce the lowest error for the overall life cycle. The lower threshold values for immature

stages were very similar to the overall egg-to-adult value of 8.89°C (48°F), so we chose 8.89°C

as the common threshold instead of a higher one solved for the adult pre-oviposition stage

(11.5°C). We estimated the duration for eggs, larvae, pupae, and adults to peak oviposition as 86,

283, 203, and 96 DDC, respectively. This analysis is presented in S3 Appendix.

Neoleucinodes elegantalis has no apparent photoperiodic response, diapause, or specific

overwintering stage. In subtropical climates in Brazil, the insect remains active throughout the

year if host plants are available [33]. We defined adults as the overwintering stage and used

January 1 as the model start date for CONUS because few host plants would be available for

immature stages at this time. We assumed that adult feeding and host search activities could

begin immediately if temperatures are suitable, and that first egg-laying would subsequently

occur after the estimated pre-oviposition period of *ca*. 55 DDC. The durations of later events (1st

to peak oviposition, immature development, etc.) were estimated from previously published data

[33,69]. We applied seven cohorts to approximate a normal distribution of emergence times that

spanned 0 to 111 DDC (average = 50 DDC) because over-wintered adults begin finding hosts

over this time frame, and used the single triangle method to calculate degree-days.

Unfortunately, we were unable to find monitoring data that were suitable for validating the

DDRP phenology model for *N. elegantalis*. Population monitoring studies of the species have

been conducted in Brazil; however, the species at these locations does not have a discrete

overwintering stage so peaks in flight are more or less sporadic and not indicative of first spring

activity or voltinism.

We used two different approaches to parameterize a DDRP climatic suitability model for *N.*

elegantalis for CONUS. First, we used a similar approach taken for *E. postvittana* in using a

CLIMEX model to help calibrate climatic suitability parameters for DDRP. As detailed in S4

Appendix, we re-parameterized a previously published CLIMEX model for *N. elegantalis* [70]

because it appeared to underpredict suitability in warmer areas where the species is known to

occur. Briefly, this involved fitting a CLIMEX model using 228 locality records from the

Neotropics and validating it using a separate set of 54 localities (S1 Table). CLIMEX correctly

predicted suitable conditions (EI > 0) at 96% of the validation localities (51/53) that fell within a

unique grid cell (S2 Fig), which suggests that the model is robust at predicting climatic

suitability. We therefore compared DDRP maps of temperature stress accumulation to those of

CLIMEX, and adjusted temperature stress limits so that most areas predicted to be suitable by

CLIMEX were also included in the potential distribution by DDRP. We considered any area that

had an EI of greater than 0 to potentially be suitable because some localities occurred in areas

that had only marginal suitability (S2 Fig).

Fig 4. Predictions of cold stress, heat stress, and climatic suitability for *Neoleucinodes*

***elegantalis* (small tomato borer) in CONUS produced by CLIMEX (A–C) and DDRP (D–F)**

based on 1961–1990 climate normals. Climatic suitability is estimated by the Ecoclimatic

Index (EI) in CLIMEX, and by combining cold and heat stress exclusions in DDRP. In DDRP,

long-term establishment is indicated by areas not under moderate (excl.-moderate) or severe

(excl.-severe) climate stress exclusion. Cold and heat stress units in DDRP were scaled from 0 to

1000 to match the scale in CLIMEX. CLIMEX maps were generated for this study based on

parameters documented in Table 3 and have not been previously published.

We further calibrated the DDRP climatic suitability model for *N. elegantalis* by fitting a

model for Brazil using 83 locality records and daily gridded T_{min} and T_{max} data for 2005 to 2016

at a 0.25° × 0.25° (*ca.* 28 km^2) spatial resolution [71]. To our knowledge, Brazil is the only

country within the range of *N. elegantalis* for which daily gridded meteorological data are

available. We did not use temperature data for earlier years (1980 to 2004) because they had

lower accuracy [71]. A random subsample of 70% of locality records were used to fit the DDRP

model (N = 58), and the remaining 30% (N = 25) were used for model validation. We modeled

climatic suitability for each year between 2005 and 2016 and iteratively adjusted heat stress

parameters to maximize model fit. Brazil represents a relatively warm part of the distribution of

N. elegantalis, so we were unable to calibrate cold stress parameters. During this process, we

also considered how well DDRP predictions of suitability and heat stress aligned with those of

CLIMEX (Fig 5). Despite DDRP's use of climate data for different time periods than CLIMEX

for this analysis (i.e. individual recent years vs. 30-year climate normals), its predictions of heat

stress and exclusions over several years were often spatially concordant with CLIMEX's

estimates (Fig. 5). This finding provided some additional reassurance beyond the validation

analysis that DDRP could correctly model climatic suitability of the species.

Fig 5. CLIMEX (A) and DDRP (B) predictions of heat stress and climatic suitability for

***Neoleucinodes elegantalis* in Brazil for all years between 2011 and 2016.** Locality records that

were used to fit (blue triangles) and validate (black circles) the DDRP climatic suitability model

are depicted. Both models predicted suitable conditions in regions where *N. elegantalis* is

widespread, including in the states of Espírito Santo (ES), Goiás (GA), Minas Gerais (MG),

Pernambuco (PE), Rio de Janeiro (RJ), Paraná (PR), Rio Grande do Sul (RS), Santa Catarina

(SC), and São Paulo (SP). The pink and blue lines in DDRP heat stress maps depict the moderate

and severe temperature stress limits, respectively.

Finally, we modeled phenology and climatic suitability in DDRP using temperature data

from 2018. We generated a phenological event map for the average date of the beginning of egg

hatch of the overwintered generation (Fig 2B). Predictions of egg hatch could enhance

population control of *N. elegantalis* because this species is most vulnerable to pesticides before

larvae enter the fruit of host plants [72].

Results

DDRP predictions of the timing of first spring egg laying for *E. postvittana* were consistent with

estimates of first spring peak flight derived from three population monitoring data sets for most

years. Predicted dates of first spring egg laying differed by fewer than six days from the peak

date for 2013, 2014, 2019, and 2020, with some occurring later than the peak (2014, 2020) and

others occurring earlier (2013, 2019; S2 Table). Model prediction error was larger for 2012 (*ca.*

3 weeks later than peak spring flight). For 2008 and 2009, DDRP predictions for three of the four

analyzed counties were consistent with the observed month of peak spring flight. Estimates of

degree-day accumulation between the last peak fall and first spring flights across four winters

(S3 Table) were similar to or slightly lower (average = 722 DDC, range = 693–827 DDC) than

the generation time assumed by the DDRP model (822 DDC).

For all modeled years, DDRP predictions of the potential distribution of *E. postvittana* were

in agreement with the vast majority of validation localities. The model based on 30-year climate

normals predicted suitable conditions at 99.7% (871/874) of validation localities. On average,

models based on climate data for each year between 2010 and 2019 predicted suitable conditions

at 97.4% (range = 92.4−99.4%) of validation localities, whereas only 2.3% (range = 0.6−6.8%)

and 0.4% (range = 0.2−0.8%) were predicted to be excluded by moderate and severe heat stress,

respectively.

Similarly, DDRP models for *N. elegantalis* in Brazil predicted suitable conditions at most

validation locations where the species is known to occur (Fig. 5B). On average, models based on

climate data for each year between 2005 and 2016 predicted suitable conditions at 92.4% (range

= 87−100%) of validation localities. Conversely, 4.7% (range = 0−13%) and 2.9% (range =

0−8.7%) were predicted to be excluded by moderate and severe heat stress, respectively. Two

localities which were incorrectly excluded by heat stress in 13 of the 15 modeled years occurred

in the Serra de Ibiapaba, which is a high-elevation region (maximum of 850 m) in the state of

Ceará where elevations change quickly across relatively short distances (e.g. *ca.* 30 km in many

areas). The coarse spatial resolution of input climate data (*ca.* 28 km^2) relative to the scale of

these elevational changes most likely explains inaccurate predictions. For relatively warm years

since 2011, heat stress was predicted to exclude *N. elegantalis* from warmer regions of Brazil

(e.g. in the states of Maranhão, Piauí, and Tocantins; Fig 5B) where the species has not been

documented to our knowledge.

Cold stress was the major determinant of the potential distribution of *E. postvittana* and *N.*

elegantalis in CONUS according to the DDRP analyses based on 30-year climate normals

(1961−1990). Both species were excluded from the northern half of CONUS by cold stress, with

the exception of (mostly) western parts of Oregon and Washington (Figs 3D and 4D). Heat stress

excluded *E. postvittana* from most of southern Arizona and California, and in parts of Texas (Fig

3E). Conversely, *N. elegantalis* was excluded only from the hottest parts of southern Arizona and

California (Fig 4E). When considering both cold and heat stress exclusions, the potential

distribution of both species included western parts of the Pacific states (California, Oregon, and

Washington), the Southeast, and southern parts of the Northeast (in Delaware, Maryland, and

New Jersey). The predicted northern range limit of *E. postvittana* extended farther north than *N.*

elegantalis and included parts of some Midwestern states (Kansas, Missouri, Illinois, and

Indiana).

DDRP predicted a smaller potential distribution for *E. postvittana* and *N. elegantalis* in 2018

compared to 1961–1990 (Figs 2 and 6). According to model runs for 2018, *E. postvittana* was

excluded by heat stress from warm areas of CONUS that were included in its potential

distribution under historical conditions, including parts of Arizona, New Mexico, Texas, and the

Central Valley of California (S3 Fig). Conversely, the more heat tolerant *N. elegantalis*

experienced only small reductions in its potential distribution in southeastern California and

southwestern Arizona due to heat stress (S3 Fig). In the East, the northern range limit for each

species was predicted to be slightly farther south in 2018 than in 1961–1990 due to higher levels

of cold stress (S3 Fig).

Predictions of potential dates for phenological events and voltinism for *E. postvittana* and *N.*

elegantalis in 2018 varied substantially by latitude in eastern CONUS (Figs 2 and 6). The earliest

date of egglaying for the first generation of *E. postvittana* was predicted to be as early as mid-

March in Florida to as late as December in the Pacific Northwest (Fig 2A). The average date of

first generation beginning of egg hatch for *N. elegantalis* was predicted to begin in the first week

of January in Florida but not until mid-June in the Pacific Northwest (Fig 2B). Thus, the timing

of monitoring trap installation to detect ovipositing adults and eggs of *E. postvittana*, or larvae of

N. elegantalis, could vary widely across CONUS. For both species, DDRP predicted as many as

seven to nine generations in coastal areas of the Southeast, compared to only one or two

generations in colder parts of the Pacific Northwest (Fig 6). Three to six generations were

predicted for most other regions of CONUS. These findings may indicate that the Southeast

would experience the longest duration of pest pressure.

Fig 6. DDRP model predictions of voltinism (number of generations per year) in (A)

***Epiphyas postvittana* (light brown apple moth) and (B) *Neoleucinodes elegantalis* (small**

tomato borer) in CONUS for 2018. Maps include estimates of climatic suitability, where long-

term establishment is indicated by areas not under moderate (excl.-moderate) or severe (excl.-

severe) climate stress exclusion.

Discussion

DDRP as a decision support tool

An improved understanding of *where* an invasive species could potentially establish as well as

when developmental stages are expected to occur have the potential to support and dramatically

improve strategic and tactical pest management decisions [6–8]. DDRP is a new spatial modeling

platform that integrates real-time and forecast predictions of phenology and climatic suitability

(risk of establishment) of invasive insect pests in CONUS, providing insights into both where

and when to focus detection efforts for a given year or growing season. These predictions may

help with detecting the presence of invasive species in the shortest time possible after they arrive

and reproduce, which increases the chance of eradication success and makes other rapid response

measures (e.g. deployment of biological control) possible and less costly [4]. For example,

phenological event maps for *E. postvittana* and *N. elegantalis* (Fig 2) identify high-risk areas

where surveillance activities could be concentrated, in addition to providing estimated dates of

activities that can ensure timely trap placement. Thus, users can use a single program to address

decision support needs for early pest detection.

DDRP was designed to be a multi-species platform, which makes it broadly applicable to

monitoring and managing populations of IPM pests and classical biological control agents. For

example, growers have used predictions of the timing of first egg hatch for codling moth [*Cydia*

pomonella (Linnaeus)], first emergence of western cherry fruit fly [*Rhagoletis indifferens*

(Curran)], and first spring oviposition of spotted wing drosophila [*Drosophila suzukii*

(Matsumura)] to help monitor and reduce populations of these major crop pests [73–76].

Phenology models for biological control insects can help managers schedule sampling trips to

coincide with insect presence on the target organism, and to estimate the synchrony of insect and

host phenology at a given location [77,78]. DDRP's estimates of voltinism may provide insights

into expected pressure on target organisms, as higher voltinism should translate to greater agent

population growth and biocontrol success.

Our hypothesis that DDRP can correctly predict the timing of first spring egg laying and the

generation length of *E. postvittana* was generally supported, although there was variation in

prediction error across years and regions. DDRP predictions were averaged across several grid

cells for two of the three monitoring data sets because they lacked precise location information,

which would likely contribute to prediction errors. Additionally, errors in predictions of spring

egg laying may reflect either differences in the effects of evening temperatures and wind speeds

on flight activity [79], differences in diet which affects development rates [80], or a lack of

consistent correspondence between male flight and peak egglaying, albeit the two events are

often found to correlate well enough at least during the springtime to use as a biofix for models

informing treatment decisions. Most estimates of generation length that we obtained by

measuring degree-day accumulation for the overwintering generation across four years were

higher than the value used by the DDRP phenology model, which may in part be due to the

model's assumption that overwintering larvae feed on old apple leaves, which slows

development [30].

DDRP predictions of the potential distribution of *E. postvittana* and *N. elegantalis* in

simulations for multiple recent years were in agreement with the vast majority of validation

localities, a finding which supports our hypothesis that DDRP can correctly predict the known

distribution of each species. Our models for 2018 indicated that heat stress excluded populations

of both species from a greater area of the Southwest compared to 1961–1990, which is consistent

with studies showing that global warming may reduce species' distributions in warmer parts of

their range [81]. However, determining whether these putative range shifts are persistent or

temporary would require combining model runs for multiple recent years or seasons. For

example, trends in the geographic distribution of stress exclusions over several years or seasons

could be visualized with a probability surface map. Estimating the direction of range shifts may

also provide insights into the response of the species to future climate change. Nonetheless, the

differences that we documented in predictions of climatic suitability based on climate data for

1961–1990 compared to 2018 suggests that an area's contemporary risk of establishment is

different than it was *ca.* 30 years ago. DDRP's ability to produce climatic suitability models in

real-time may provide more meaningful insights into areas that are presently suitable for an

invasive species, and therefore allow for more effective placement of surveillance operations.

Additionally, the ability to capture interannual variability of pest risk in space-time may allow

decision-makers in pest management to react more adaptively to risk.

There are numerous opportunities for improving and extending the applications of DDRP.

The platform can be readily modified to use daily temperature data for a different region than

CONUS, as we demonstrated in our analysis of climatic suitability for *N. elegantalis* in Brazil.

Additionally, DDRP could be modified to model other types of temperature-dependent

organisms such as non-insect invertebrates, plants, plant-pathogenic bacteria and fungi, and

insect plant and animal virus vectors. The platform has been tested for and could be used through

an on-line web interface, although there is the potential that memory intensive processes could

overload a server host. This issue, as yet untested, could be addressed by using a cloud

computing platform. We describe some additional features that could be added to potentially

improve model accuracy and expand the list of products in more detail below. The most recent

code for DDRP is available at GitHub (https://github.com/bbarker505/ddrp_v2.git), where we

invite scientists and practitioners to jointly develop the platform and database of species models.

Comparison of DDRP to other platforms

From a historical perspective, DDRP could be considered a partial descendant and spatialized

version of the PETE (Predictive Extension Timing Estimator) phenological modeling platform

that was established as a standard in the mid-1970s [82]. Features in common include a cohort

approach to population phenological modeling using daily degree-days as the main input,

provision for major insect life stages and a separately parameterized overwintering distribution,

an open-source non-proprietary standard for sharing, and a focus on agricultural extension

(applied decision support). Unlike PETE, DDRP is spatialized and therefore able to produce a

variety of mapping products including phenological event maps, and it also includes options to

use separate thresholds for different life stages and to generate climatic suitability models. DDRP

could be improved by adding certain features of the PETE platform including the use of a

diapause trigger, and a distributed delay function that would allow population spread to increase

with each subsequent generation.

Web-hosted daily degree-day maps and degree-day lookup table maps for pest management

are available from several platforms including USPEST.ORG, SAFARIS, and Enviroweather,

but none of these are capable of predicting climatic suitability like DDRP. Additionally, degree-

day lookup tables have certain underlying assumptions which may result in an oversimplified

model that is lacking in biological realism, such as assuming that multiple species or life stages

within a species have a common lower temperature threshold, and that early season development

in a population begins at the same time (i.e. no developmental variation). In contrast, DDRP uses

species specific parameters including stage-specific lower and upper developmental temperature

thresholds, and it can account for developmental variation within populations by generating and

combining results across multiple cohorts that complete the overwintering stage at different

times. Compared to a simple model based on generation time degree-days, a well-parameterized

DDRP model would likely produce more accurate predictions of voltinism and spring activity

because different temperature thresholds for multiple life stages may be used, the overwintering

stage is parameterized separately from the post-winter stage (e.g. overwintering adult vs. adult),

and the timing of spring activity is summarized across multiple cohorts.

The Insect Life Cycle Modelling (ILCYM) [83,84] and phenModel [85] software packages

are open-source R programs that use life stage specific parameters to model temperature-based

insect phenology, and both incorporate parameters that describe developmental variation within a

population. ILCYM is similar to DDRP in its ability to predict phenology and climate based

establishment risk in a spatial context. However, ILCYM is a full population dynamics modeling

platform that requires life table data at constant and variable temperatures, which are seldom

available for anticipated but not yet present invasive insect species. Additionally, published

ILCYM risk model simulations use climate normals or GCMs at global or regional scales

[83,86], whereas DDRP was designed to use real-time and forecast climate data for within-

season decision support in CONUS. The phenModel package is not spatialized nor capable of

modeling climatic suitability, and it would need to be modified to use for an insect species other

than the blue willow beetle *Phratora vulgatissima* [85].

Uncertainties, limitations, and other considerations

Linear (degree-day) modeling

DDRP uses a relatively simple degree-day modeling approach, whereas some platforms

including ILCYM, phenModel, and devRate [87] offer complex functions to model nonlinear

responses of insects to temperature. Degree-day models are ideal for multi-species platforms like

DDRP because there are sufficient data to parameterize a degree-day model for most insect pest

species of economic importance in the United States [18,88]. Linear degree-day models are also

readily calibrated and sometimes constructed entirely using field data, making them more

practical for extension and decision support use [14]. Additionally, degree-day models require

only daily T_{min} and T_{max} data (as opposed to hourly data for most nonlinear models), which are

available at a high spatial resolution for CONUS from multiple sources including PRISM and

RTMA. Nonetheless, it is important for users to recognize potential sources of error and lack of

precision in degree-day models, such as their limited ability to accurately model development at

supra-optimal temperatures [14,89].

Environmental inputs

DDRP is intentionally parameterized in a simple, conservative manner, which will hopefully

achieve the goal of a parsimonious balance of both model simplicity and accuracy [14,90].

Nonetheless, DDRP is driven entirely by temperature, and therefore ignores other factors that

may affect the development and distribution of insects such as photoperiod, moisture, dispersal,

resources, disturbance, and biotic interactions [7,91]. The potential consequences of this

limitation will depend on the biology of the organism under study. For example, dry stress is a

major factor restricting the current distribution of *N. elegantalis* in its native range [92–94], and

it limits the distribution of *E. postvittana* both in its native range [30,95] and in Southern

California and Arizona [96]. Our CLIMEX model for *N. elegantalis* predicted higher dry stress

in arid parts of the Southwest than the rest of CONUS (S4 Fig), which suggests that an absence

of moisture factors in DDRP may result in over-predictions of climatic suitability in this region.

However, this conservative-leaning error may in fact better reflect human manipulation of the

landscape (e.g. greenhouse and irrigation usage) that may allow the species to exist in such

regions. Future versions of DDRP that can process gridded moisture data and incorporate

moisture stress factors into climatic suitability models may help overcome our current limitations

in matching CLIMEX models, and may improve predictions for moisture-sensitive species such

as *N. elegantalis.* Additionally, we are developing a version of DDRP that incorporates

photoperiodically induced life history events such as winter diapause and summer aestivation,

which builds on earlier phenology modeling work that estimated voltinism of photoperiod-

sensitive insects [97].

Presumptive models

Uncertainties regarding the accuracy of temporal or spatial predictions of invasive species that

are not yet established is inevitable, in part because no validation data are yet available, and

species interceptions do not imply establishment [7,98]. DDRP models for species for which

only presumptive models exist should therefore be used conservatively. For example,

surveillance or management actions could be implemented in advance of predicted phenological

events as a precautionary measure (e.g. installing traps even earlier than estimates for the earliest

date of overwintering adult emergence). To potentially avoid under-predicting the risk of

establishment, the potential distribution could be defined as areas not under severe climate stress

as opposed to defining it using both stress levels. Additionally, climatic suitability models

generated by DDRP could be combined with those produced using different modeling methods

(e.g. correlative, semi-mechanistic, or mechanistic) to create a "hybrid" model, which may

increase the reliability of predictions [7,91].

Web platforms that support sharing of pest observations from the United States will be

valuable resources for validating and increasing the predictive performance of DDRP models for

species that are already established in CONUS. For example, the iPiPE and its sister platforms

(http://www.ipipe.org, https://ipmpipe.org) have created a national information technology

infrastructure for sharing pest observations in near real-time and contributing them to a national

depository [99]. Similarly, the USA National Phenology Network provides a depository of plant

and insect phenology observations contributed by citizen scientists [16]. The National

Agricultural Pest Information System (NAPIS; https://napis.ceris.purdue.edu/home) currently

has over 5.17 million records from pest detection surveys, and is another potential source of

validation data.

Geographic variation

Populations of an invading species may exhibit geographic variation in temperature dependent

development if genetically divergent individuals are introduced to different areas or rapid

evolutionary changes occur in new environments [100,101]. If the geographic distribution of

variation in a relevant thermal trait is well understood, then model accuracy may be improved by

building separate models for each genotype. For example, an egg hatch phenology model for a

subspecies of the Asian gypsy moth, *Lymantria dispar asiatica* (Vnukovskij), had reduced error

compared to a similar model constructed for the European subspecies that has invaded North

America, *Lymantria dispar dispar* (Linnaeus), which has a markedly different predominant

phenotype [102]. An alternative approach may be to run several models, each with a different

value for the parameter of interest, and present a range of model predictions. Conversely, DDRP

could be modified to accept a grid of parameter values so that geographic variation would be

accounted for in a single model run.

A lack of knowledge on how early-season environmental conditions or events that initiate the

first spring activity of a species (biofix) vary geographically may be a source of error because the

model start date affects all downstream predictions. For example, how does first spring activity

vary across the wide range of warming conditions possibly encountered for a large region such

as CONUS? As a case in point, our phenology model for *N. elegantalis* assumes that moths have

only 55 DDC before egg-laying behaviors may occur. This assumption may not be valid for sub-

tropical zones of the United States, where flight and reproduction could occur even earlier.

Conversely, a much longer spring warm-up may be needed in temperate zones because

commercial tomatoes are transplanted much later in the year. Studying how first spring activity

(adult flight) in *N. elegantalis* potentially varies geographically in Central or South America

would help to refine a range of model start times. The phenology model for DDRP could then be

parameterized using a necessarily conservative selection of start dates or by inputting a grid of

start dates. Using a broad distribution of emergence times to initiate the cohorts could be another

approach to accommodate uncertainty in first spring activity.

Distributed delay

There is currently no distributed delay function in DDRP, meaning that the overlap in

generations and life stages of cohorts does not increase over multiple generations. This is

particularly an issue for species that have significant overlap in generations because they

continue to develop throughout winter months or lack a temperature or photoperiodic event that

synchronize populations, such as *E. postvittana* [31,34,58]. DDRP may accurately predict peak

events in each generation for these species, but inaccurately predict the first appearance of one or

more life stages after the first or second generations because of increasing overlap in generation

cohorts. Consequently, phenological event maps should be most reliable for the first few

generations. This will be among the high priority issues in development of future versions of the

platform.

Conclusion

DDRP is a new modeling tool that can integrate mapping of phenology and climatic suitability in

real-time to provide timely and comprehensive guidance for stakeholders needing to know both

where and when an invasive insect species could potentially invade. When used for surveillance,

the platform will hopefully increase chances for early detection of new or spreading invasive

threats in the United States, and therefore help pest management programs mitigate their

potential damage to agricultural and environmental resources. Additionally, DDRP may help

plan monitoring and management efforts for IPM pests and biological control insects, and to

predict pest pressure on host plants.

The case studies we presented provided examples of how DDRP models may be

parameterized and then run to produce various products including gridded and graphical

predictions of the number of generations, life stages present, dates of phenological events, and

areas of climatic suitability based on two levels of climate stress. We encourage users of DDRP

to consider the limitations of the platform, to report the conditions that their model was designed

to work under (e.g. a particular region, life stage event, or generation), and to document any

known sources of model error that could not be accounted for when providing validation and

other feedback reports. Additionally, models should be validated whenever possible, as we did

for the *E. postvittana* and *N. elegantalis* models presented in this study. The flexible and open-

source nature of DDRP will facilitate making modifications and improvements, such as adding

new environmental factors, using it for other regions besides CONUS, modeling non-insect

organisms, expanding the types of model products, and adding features to improve model

accuracy.

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Data availability

On GitHub, we have uploaded the most current version of DDRP along with a user guide and

species parameter files (github.com/bbarker505/ddrp_v2), Perl and Octave scripts used for

temporal downscaling of monthly climate averages to daily averages

(github.com/bbarker505/dailynorms), and temporally downscaled climate data

(github.com/bbarker505/1990_daily_30yr). On Zenodo, we have archived the version of DDRP

used for this study (https://doi.org/10.5281/zenodo.3832731), and the scripts and outputs of the

temporal downscaling analysis of climate data (https://doi.org/10.5281/zenodo.3601671 and

https://doi.org/10.5281/zenodo.387962, respectively).

References

1.   Pimentel D, Zuniga R, Morrison D. Update on the environmental and economic costs

associated with alien-invasive species in the United States. Ecol Econ. 2005;52: 273–288.

https://doi.org/10.1016/j.ecolecon.2004.10.002

2.   Lovett GM, Weiss M, Liebhold AM, Holmes TP, Leung B, Lambert KF, et al. Nonnative

forest insects and pathogens in the United States: Impacts and policy options. Ecol Appl.

2016;26: 1437–1455. https://doi.org/10.1890/15-1176

3.   Paini DR, Sheppard AW, Cook DC, Barro PJ De, Worner SP, Thomas MB. Global threat

to agriculture from invasive species. Proc Natl Acad Sci. 2016;113: 7575–7579.

https://doi.org/10.1073/pnas.1602205113

4.   United States Department of the Interior. Safeguarding America's lands and waters from

invasive species: a national framework for early detection and rapid response. Washington

D.C., USA; 2016. p. 55.

5.   Tonnang HEZ, Hervé BDB, Biber-freudenberger L, Salifu D, Subramanian S, Ngowi VB,

et al. Advances in crop insect modelling methods — Towards a whole system approach.

Ecol Modell. 2017;354: 88–103. https://doi.org/10.1016/j.ecolmodel.2017.03.015

6.   Venette RC. Climate analyses to assess risks from invasive forest insects: simple matching

to advanced models. Curr For Reports. 2017;3: 255–268. https://doi.org/10.1007/s40725-

017-0061-4

7.   Lantschner MV, de la Vega G, Corley JC. Predicting the distribution of harmful species

and their natural enemies in agricultural, livestock and forestry systems: an overview. Int J

Pest Manag. 2019;65: 190–206. https://doi.org/10.1080/09670874.2018.1533664

8.   Crimmins TM, Gerst KL, Huerta DG, Marsh RL, Posthumus EE, Rosemartin AH, et al.

Short-term forecasts of insect phenology inform pest management. Ann Entomol Soc Am.

2020;113: 139–148. https://doi.org/10.1093/aesa/saz026

9.   Prasad YG, Prabhakar M. Pest monitoring and forecasting. In: Abrol D, Shankar U,

editors. Integrated Pest Management: Principles and Practice. Wallingford, United

Kingdom: CABI; 2012. pp. 41–57. https://doi.org/10.1079/9781845938086.0041

10.   Orlandini S, Magarey RD, Park EW, Kroschel J. Methods of agroclimatology: modeling

approaches for pests and diseases. In: Hatfield JL, Sivakuma MVK, Prueger JH, editors.

Agroclimatology: Linking Agriculture to Climate, Agronomy Monograph 60. Madison,

Wisconsin: American Society of Agronomy, Crop Science Society of America, and Soil

Science Society of America, Inc.; 2017.

https://doi.org/10.2134/agronmonogr60.2016.0027

11.   Donatelli M, Magarey RD, Bregaglio S, Willocquet L, Whish JPM, Savary S. Modelling

the impacts of pests and diseases on agricultural systems. Agric Syst. 2017;155: 213–224.

https://doi.org/10.1016/j.agsy.2017.01.019

12.   Pruess KP. Degree-day methods for pest management. Environ Entomol. 1983;12: 613–

619. https://doi.org/10.1093/ee/12.3.613

13.   Higley LG, Pedigo LP, Ostlie R. DEGDAY: a program for calculating degree-days, and

assumptions behind the degree-day approach. Environ Entomol. 1986;15: 999–1016.

https://doi.org/10.1093/ee/15.5.999

14.   Coop LB, Barker BS. Advances in understanding species ecology: phenological and life

cycle modeling of insect pests. In: Kogan M, Heinrichs E, editors. Integrated Management

of Insect Pests: Current and Future Developments. Sawston, England; 2020. pp. 43–96.

https://doi.org/10.19103/AS.2019.0047.02

15.   Herms DA. Using degree-days and plant phenology to predict pest activity. In: Krischik

V, Davidson J, editors. IPM (Integrated Pest Management) of Midwest Landscapes

Minnesota Agricultural Experiment Station Publication SB-07645. St. Paul, MN; 2004.

pp. 49–59.

16.   Crimmins TM, Crimmins MA, Gerst KL, Rosemartin AH, Weltzin JF. USA National

Phenology Network's volunteer contributed observations yield predictive models of

phenological transitions. PLoS One. 2017;12: e0182919.

https://doi.org/10.1371/journal.pone.0182919

17.   Struelens Q, Rebaudo F, Quispe R, Dangles O. Thermal pace-of-life strategies improve

1014   phenological predictions in ectotherms. Sci Rep. 2018;8: 1–9.

1015   https://doi.org/10.1038/s41598-018-34274-1

1016   18.   Jarošík V, Honěk A, Magarey RD, Skuhrovec J. Developmental database for phenology

1017   models: related insect and mite species have similar thermal requirements. J Econ

1018   Entomol. 2011;104: 1870–1876. https://doi.org/10.1603/ec11247

1019   19.   Régnière J, Powell JA. Animal life cycle models (Poikilotherms). In: Schwartz MD,

1020   editor. Phenology: An Integrative Environmental Science. Dordrecht, The Netherlands:

1021   Springer Netherlands; 2013. pp. 295–316. https://doi.org/10.1007/978-94-007-6925-0_16

1022   20.   R Development Core Team. R: A Language and Environment for Statistical Computing.

1023   Vienna, Austria: R Foundation for Statistical Computing; 2020.

1024   21.   Daly C, Halbleib H, Smith J, Gibson W, Doggett M, Taylor G, et al. Physiographically

1025   sensitive mapping of climatological temperature and precipitation across the conterminous

1026   United States. Int J Climatol. 2008;28: 2031–2064. https://doi.org/10.1002/joc.1688

1027   22.   Thornton PE, Thornton M., Mayer BW, Wei Y, Devarakonda R, Vose RS, et al. Daymet:

1028   Daily Surface Weather Data on a 1-km Grid for North America, Version 3. Oak Ridge,

1029   Tennessee, USA: ORNL DAAC; 2018. https://doi.org/10.3334/ORNLDAAC/1328

1030   23.   Kirtman BP, Min D, Infanti JM, Kinter JL, Paolino DA, Zhang Q, et al. The North

1031   American multimodel ensemble: Phase-1 seasonal-to-interannual prediction; phase-2

1032   toward developing intraseasonal prediction. Bull Am Meteorol Soc. 2014;95: 585–601.

1033   https://doi.org/10.1175/BAMS-D-12-00050.1

1034   24.   Vose RS, Easterling ER, Kunkel KE, LeGrande EN, Wehner MF. Temperature changes in

1035   the United States. In: Wuebbles DJ, Fahey DW, Hibbard KA, Dokken DJ, Stewart BC,

1036   Maycock TK, editors. Climate Science Special Report: Fourth National Climate

1037   Assessment, Volume I. Washington D.C., USA; 2017. pp. 185–206.

1038   https://doi.org/10.7930/J0N29V45

1039   25.   Wuebbles D, Meehl G, Hayhoe K, Karl TR, Kunkel K, Santer B, et al. CMIP5 climate

1040   model analyses: Climate extremes in the United States. Bull Am Meteorol Soc. 2014;95:

1041   571–583. https://doi.org/10.1175/BAMS-D-12-00172.1

1042   26.   Barbero R, Abatzoglou JT, Hegewisch KC. Evaluation of statistical downscaling of North

1043   American Multimodel Ensemble Forecasts over the western United States. Weather

1044   Forecast. 2017;32: 327–341. https://doi.org/10.1175/waf-d-16-0117.1

1045   27.   Roundy JK, Yuan X, Schaake J, Wood EF. A framework for diagnosing seasonal

1046   prediction through canonical event analysis. Mon Weather Rev. 2015;143: 2404–2418.

1047   https://doi.org/10.1175/MWR-D-14-00190.1

1048   28.   Glahn HR, Ruth DP. The new digital forecast database of the national weather service.

1049   Bull Am Meteorol Soc. 2003;84: 195–201. https://doi.org/10.1175/BAMS-84-2-195

1050   29.   Régnière J, Powell J, Bentz B, Nealis V. Effects of temperature on development, survival

1051   and reproduction of insects: experimental design, data analysis and modeling. J Insect

1052   Physiol. 2012;58: 634–647. https://doi.org/10.1016/j.jinsphys.2012.01.010

1053   30.   Geier PW, Briese DT. The light-brown apple moth, *Epiphyas postvittana* (Walker): a

1054   native leafroller fostered by European settlement. In: Kitching PL, Jones RE, editors. The

Ecology of Pests: Some Australian Case Histories. Canberra, Australia: CSIRO; 1981. pp.

130–155.

31.   Danthanarayana W. The bionomics, distribution and host range of the light brown apple

moth, *Epiphyas postvittana* (Walk.) (Tortricidae). Aust J Zool. 1975;23: 419–437.

https://doi.org/10.1071/ZO9750419

32.   Danthanarayana W, Gu H, Ashley S. Population growth of *Epiphyas postvittana*, the

lightbrown apple moth (Lepidoptera: Tortricdae) in relation to diet, temperature and

climate. Aust J Zool. 1995;43: 381–394. https://doi.org/10.1071/ZO9950381

33.   Moraes CP, Foerster LA. Thermal requirements, fertility, and number of generations of

Neoleucinodes elegantalis (Guenée) (Lepidoptera: Crambidae). Neotrop Entomol.

2015;44: 338–344. https://doi.org/10.1007/s13744-015-0293-6

34.   Bürgi LP, Roltsch WJ, Mills NJ. Abundance, age structure, and voltinism of light brown

apple moth populations in California. Environ Entomol. 2011;40: 1370–1377.

https://doi.org/10.1603/en11165

35.   Rebaudo F, Rabhi VB. Modeling temperature-dependent development rate and phenology

in insects: review of major developments, challenges, and future directions. Entomol Exp

Appl. 2018;166: 607–617. https://doi.org/10.1111/eea.12693

36.   Yonow T, Zalucki MP, Sutherst RW, Dominiak BC, Maywald GF, Maelzer DA, et al.

Modelling the population dynamics of the Queensland fruit fly, *Bactrocera (Dacus)*

tryoni: a cohort-based approach incorporating the effects of weather. Ecol Modell.

2004;173: 9–30. https://doi.org/10.1016/S0304-3800(03)00306-5

37.   Régnière J. A method of describing and using variability in development rates for the

1077   simulation of insect phenology. Can Entomol. 1984;116: 1367–1376.

1078   https://doi.org/10.4039/Ent1161367-10

1079   38.   Severini M, Baumgärtner J, Limonta L. Parameter estimation for distributed delay based

1080   population models from laboratory data: egg hatching of *Oulema duftschmidi*

1081   Redthenbacher (Coleoptera, Chrysomelidae) as an example. Ecol Modell. 2003;167: 233–

1082   246. https://doi.org/10.1016/S0304-3800(03)00188-1

1083   39.   Wagner TL, Wu H-I, Feldman RM, Sharpe PJH, Coulson RN. Multiple-cohort approach

1084   for simulating development of insect populations under variable temperatures. Ann

1085   Entomol Soc Am. 1985;78: 691–704. https://doi.org/10.1093/aesa/78.6.691

1086   40.   Sutherst RW. Pest species distribution modelling: origins and lessons from history. Biol

1087   Invasions. 2014;16: 239–256. https://doi.org/10.1007/s10530-013-0523-y

1088   41.   Kriticos DJ, Maywald GF, Yonow T, Zurcher EJ, Herrmann N, Sutherst RW. CLIMEX

1089   Version 4: Exploring the effects of climate on plants, animals and diseases. CSIRO,

1090   Canberra, Australia; 2016. p. 156. https://publications.csiro.au/pu.

1091   42.   Sutherst RW, Maywald GF. A computerised system for matching climates in ecology.

1092   Agric Ecosyst Environ. 1985;13: 281–299. https://doi.org/10.1016/0167-8809(85)90016-7

1093   43.   Kriticos DJ, Webber BL, Leriche A, Ota N, Macadam I, Bathols J, et al. CliMond: global

1094   high-resolution historical and future scenario climate surfaces for bioclimatic modelling.

1095   Methods Ecol Evol. 2012;3: 53–64. https://doi.org/10.1111/j.2041-210X.2011.00134.x

44.   Lindsey A, Newman J. Use of official weather data in spring time: temperature analysis of

an Indiana phenological record. Ecology. 1956;4: 812–823.

https://doi.org/10.2307/1933072

45. Sevacherian V, Stern VM, Mueller AJ. Heat accumulation for timing *Lygus* control

measures in a safflower-cotton complex. J Econ Entomol. 1977;70: 399–402.

https://doi.org/10.1093/jee/70.4.399

46. Roltsch W, Zalom A, Strand JF, Pitcairn MJ. Evaluation of several degree-day estimation

methods in California climates. Int J Biometeorol. 1999;42: 169–176.

https://doi.org/10.1007/s004840050101

47. Chuine I, Régnière J. Process-based models of phenology for plants and animals. Annu

Rev Ecol Evol Syst. 2017;48: 159–182. https://doi.org/10.1146/annurev-ecolsys-110316-

022706

48. RStudio Team. RStudio: Integrated Development for R. Boston, MA, USA; 2020.

49. Hijmans RJ. raster: Geographic Data Analysis and Modeling. R package version 2.9-5.

https://CRAN.R-project.org/package=raster. 2019.

50. Wickham H, Henry L. tidyr: easily tidy data with "spread()" and "gather()" functions. R

package version 0.8.3. https://CRAN.R-project.org/package=tidyr. 2019.

51. Wickham H. stringr: simple, consistent wrappers for common string operations. R

package version 1.4.0. https://CRAN.R-project.org/package=stringr. 2019.

52. Wickham H, François R, Henry L, Müller K. dplyr: a grammar of data manipulation. R

package version 0.8.2. https://CRAN.R-project.org/package=dplyr. 2019.

53. Wickham H. ggplot2: elegant graphics for data analysis. New York.

https://ggplot2.tidyverse.org: Springer-Verlag; 2016.

54.   Davis TL. optparse: Command Line Option Parser. R package version 1.6.1.

https://CRAN.R-project.org/package=optparse. 2019.

55.   Weston S, Calaway R. Getting started with doParallel and foreach. Vignette, CRAN, URL

http://cran.r-project.org/web/packages/doParallel/index.html. 2018.

56.   Geier PW, Springett BP. Population characteristics of Australian leafrollers (*Epiphyas*

spp., Lepidoptera) infesting orchards. Aust J Ecol. 1976;1: 127–144.

https://doi.org/10.1111/j.1442-9993.1976.tb01102.x

57.   Suckling DM, Brockerhoff EG. Invasion biology, ecology, and management of the light

brown apple moth (Tortricidae). Annu Rev Entomol. 2010;55: 285–306.

https://doi.org/10.1146/annurev-ento-112408-085311

58.   Dumbleton L. The apple leaf-roller (*Tortrix postvittana* Walker). New Zeal J Sci Technol.

1932;14: 83–92.

59.   Varela LG, Johnson MW, Strand L, Wilen CA, Pickel C. Light brown apple moth's

arrival in California worries commodity groups. Calif Agric. 2008;62: 57–61.

https://doi.org/10.3733/ca.v062n02p57

60.   Suckling DM, Stringer LD, Baird DB, Butler RC, Sullivan TES, Lance DR, et al. Light

brown apple moth (*Epiphyas postvittana*) (Lepidoptera: Tortricidae) colonization of

California. Biol Invasions. 2014;16: 1851–1863. https://doi.org/10.1007/s10530-013-

0631-8

61.   Fowler G, Garrett L, Neeley A, Borchert D, Spears B. Economic analysis: risk to U.S.

1139   apple, grape, orange and pear production from the light brown apple moth, Epiphyas

1140   postvittana (Walker). USDA-APHIS-PPQ-CPHST-PERAL. Raleigh, NC; 2007.

1141 62. Bürgi LP, Mills NJ. Cold tolerance of the overwintering larval instars of light brown apple

1142   moth *Epiphyas postvittana*. J Insect Physiol. 2010;56: 1645–1650.

1143   https://doi.org/10.1016/j.jinsphys.2010.06.009

1144 63. Tomkins AR, Penman DR, Chapman RB, Worner SP. An evaluation of a phenological

1145   model (PETE) to assist insect pest control in apple orchards in Canterbury, New Zealand.

1146   New Zeal J Exp Agric. 1987;15: 381–388.

1147   https://doi.org/10.1080/03015521.1987.10425586

1148 64. He S, Worner SP, Ikeda T. Modeling the potential global distribution of light brown apple

1149   moth *Epiphyas postvittana* (Lepidoptera: Tortricidae) using CLIMEX. J Asia Pac

1150   Entomol. 2012;15: 479–485. https://doi.org/10.1016/j.aspen.2012.01.004

1151 65. Lozier JD, Mills NJ. Predicting the potential invasive range of light brown apple moth

1152   (*Epiphyas postvittana*) using biologically informed and correlative species distribution

1153   models. Biol Invasions. 2011;13: 2409–2421. https://doi.org/10.1007/s10530-011-0052-5

1154 66. Díaz-Montilla A, Suárez-Baron H, Gallego-Sánchez G, Saldamando-Benjumea C, Tohme

1155   J. Geographic differentiation of Colombian *Neoleucinodes elegantalis* (Lepidoptera:

1156   Crambidae) haplotypes: evidence for Solanaceae host plant association and Holdridge life

1157   zones for genetic differentiation. Ann Entomol Soc Am. 2013;106: 586–597.

1158   https://doi.org/10.1603/AN12111

1159 67. Bulletin OEPP/EPPO. *Neoleucinodes elegantalis*. 2015;45: 9–13.

https://doi.org/10.1111/epp.12189

68.   Díaz-Montilla A, Solis MA, Kondo T. The tomato fruit borer, *Neoleucinodes elegantalis*

(Guenée) (Lepidoptera: Crambidae), an insect pest of neotropical solanaceous fruits. In:

Peña J, editor. Potential Invasive Pests of Agricultural Crops. Boston, MA: CABI; 2013.

pp. 137–159.

69.   Moraes CP, Foerster LA. Development and reproduction of *Neoleucinodes elegantalis*

(Lepidoptera: Crambidae) on tomato (*Solanum licopercum*) cultivars. Rev Colomb

Entomol. 2014;40: 40–43.

70.   da Silva RS, Kumar L, Shabani F, Picanço MC. An analysis of sensitivity of CLIMEX

parameters in mapping species potential distribution and the broad-scale changes observed

with minor variations in parameter values: an investigation using open-field *Solanum*

lycopersicum and *Neoleucinodes elegantalis* as an example. Theor Appl Climatol.

2018;132: 135–144. https://doi.org/10.1007/s00704-017-2072-2

71.   Xavier AC, King CW, Scanlon BR. Daily gridded meteorological variables in Brazil

(1980–2013). Int J Climatol. 2016;36: 2644–2659. https://doi.org/10.1002/joc.4518

72.   Eiras AE, Blackmer JL. Eclosion time and larval behavior of the tomato fruit borer,

Neoleucinodes elegantalis (Guenée) (Lepidoptera: Crambidae). Sci Agric. 2003;60: 195–

197. https://doi.org/https://doi.org/10.1590/S0103-90162003000100030

73.   van Kirk JR, AliNiazee MT. Determining low-temperature threshold for pupal

development of the western cherry fruit fly for use in phenology models. Environ

Entomol. 1981;10: 968–971. https://doi.org/10.1093/ee/10.6.968

74.   Brunner J, Hoyt S. Codling moth control - a new tool for timing sprays. Washington State

1182   University Cooperative Extension Bulletin. 1987;1072.

1183   75.   Jones VP, Alston DG, Brunner JF, Davis DW, Shelton MD. Phenology of the western

1184   cherry fruit fly (Diptera: Tephritidae) in Utah and Washington. Ann Entomol Soc Am.

1185   1991;84: 488–492. https://doi.org/10.1093/aesa/84.5.488

1186   76.   Zerulla FN, Augel C, Zebitz CPW. Oviposition activity of *Drosophila suzukii* as mediated

1187   by ambient and fruit temperature. PLoS One. 2017;12: e0187682.

1188   https://doi.org/10.1371/journal.pone.0187682

1189   77.   Gramig GG, Burns EE, Prischmann-Voldseth DA. Predicting developmental timing for

1190   immature Canada thistle stem-mining weevils, *Hadroplontus litura* (Coleoptera:

1191   Curculionidae). Environ Entomol. 2015;44: 1085–1094. https://doi.org/10.1093/ee/nvv089

1192   78.   Herrera AM, Dahlsten DD, Tomic-Carruthers N, Carruthers RI. Estimating temperature-

1193   dependent developmental rates of *Diorhabda elongata* (Coleoptera: Chrysomelidae), a

1194   biological control agent of saltcedar (Tamarix spp.). Environ Entomol. 2005;34: 775–784.

1195   https://doi.org/10.1603/0046-225x-34.4.775

1196   79.   Danthanarayana W. Diel and lunar flight periodicities in the light brown apple moth,

1197   *Epiphyas postvittana* (Walker) (Tortricidae) and their possible adaptive significance. Aust

1198   J Zool. 1976;24: 65–73.

1199   80.   Tomkins AR. Tortricid moth pest management in Canterbury apple orchards. Ph.D. thesis,

1200   University of Canterbury, Canterbury, New Zealand. 1984.

81.   Menéndez R. How are insects responding to global warming? Tijdschr voor Entomol.

2007;150: 355–365.

82.   Welch SM, Croft BA, Brunner J, Michels M. PETE: an extension phenology modeling

system for management of multi-species pest complex. Environ Entomol. 1978;7: 482–

494. https://doi.org/10.1093/ee/7.4.487

83.   Sporleder M, Juarez H, Simon R, Kroschel J. ILCYM-Insect life cycle modeling: software

for developing temperature-based insect phenology models with applications for regional

and global pest risk assessments and mapping. Proceedings of the 15th Triennial ISTRC

Symposium of the International Society for Tropical Root Crops (ISTRC). Lima, Peru;

2009. pp. 216–223.

84.   Sporleder M, Tonnang HEZ, Carhuapoma P, Gonzalez JC, Juarex J, Kroschel J. Insect life

cycle modeling (ILCYM) software - a new tool for regional and global insect pest risk

assessments under current and future climate. In: Peña JE, editor. Potential Invasive Pests

of Agricultural Crops. CAB eBooks; 2013. p. 464.

https://doi.org/10.1079/9781845938291.0412

85.   Pollard CP, Griffin CT, Andrade Moral R de, Duffy C, Chuche J, Gaffney MT, et al.

phenModel: A temperature-dependent phenology/voltinism model for a herbivorous insect

incorporating facultative diapause and budburst. Ecol Modell. 2020;416.

https://doi.org/10.1016/j.ecolmodel.2019.108910

86.   Fand BB, Tonnang HEZ, Kumar M, Bal SK, Singh NP, Rao DVKN, et al. Predicting the

impact of climate change on regional and seasonal abundance of the mealybug

Phenacoccus solenopsis Tinsley (Hemiptera: Pseudococcidae) using temperature-driven

phenology model linked to GIS. Ecol Modell. 2014;288: 62–78.

https://doi.org/10.1016/j.ecolmodel.2014.05.018

87. Rebaudo F, Struelens Q, Dangles O. Modelling temperature-dependent development rate

and phenology in arthropods: The devRate package for R. Methods Ecol Evol. 2018;9:

1144–1150.

88. Nietschke BS, Magarey RD, Borchert DM, Calvin DD, Jones E. A developmental

database to support insect phenology models. Crop Prot. 2007;26: 1444–1448.

https://doi.org/10.1016/j.cropro.2006.12.006

89. Moore JL, Remais J V. Developmental models for estimating ecological responses to

environmental variability: structural, parametric, and experimental issues. Acta Biotheor.

2014;62: 69–90. PMID: 24443079

90. Logan JA. In defense of big ugly models. Am Entomol. 1994;40: 202–207.

https://doi.org/10.1093/ae/40.4.202

91. Srivastava V, Lafond V, Griess VC. Species distribution models (SDM): Applications,

benefits and challenges in invasive species management. CAB Rev Perspect Agric Vet Sci

Nutr Nat Resour. 2019;14. https://doi.org/10.1079/PAVSNNR201914020

92. Díaz AEM, Solis A, Brochero HL. Distribución geográfica de *Neoleucinodes elegantalis*

(Lepidoptera: Crambidae) en Colombia. Rev Colomb Entomol. 2011;37: 71–76.

93. da Silva RS, Kumar L, Shabani F, Vieira A. Dry stress decreases areas suitable for

Neoleucinodes elegantalis (Lepidoptera: Crambidae) and affects its survival under climate

predictions in South America. Ecol Inform. 2018;46: 103–113.

https://doi.org/10.1016/j.ecoinf.2018.06.003

94.   da Silva RS, Kumar L, Shabani F, Picanço MC. Potential risk levels of invasive

1246   *Neoleucinodes elegantalis* (small tomato borer) in areas optimal for open-field *Solanum*

1247   *lycopersicum* (tomato) cultivation in the present and under predicted climate change. Pest

1248   Manag Sci. 2017;73: 616–627. https://doi.org/10.1002/ps.4344

1249   95.   Danthanarayana W. Population ecology of the light brown apple moth, *Epiphyas*

1250   *postvittana* (Lepidoptera: Tortricidae). J Anim Ecol. 1983;52: 1–33.

1251   https://doi.org/10.2307/4585

1252   96.   Gutierrez AP, Mills NJ, Ponti L. Limits to the potential distribution of light brown apple

1253   moth in Arizona-California based on climate suitability and host plant availability. Biol

1254   Invasions. 2010;12: 3319–3331. https://doi.org/10.1007/s10530-010-9725-8

1255   97.   Grevstad FS, Coop LB. The consequences of photoperiodism for organisms in new

1256   climates. Ecol Appl. 2015;25: 1506–1517.

1257   98.   Barbet-Massin M, Rome Q, Villemant C, Courchamp F. Can species distribution models

1258   really predict the expansion of invasive species? PLoS One. 2018;13: 1–14.

1259   https://doi.org/10.1371/journal.pone.0193085

1260   99.   Isard SA, Russo JM, Magarey RD, Golod J, Vankirk JR. Integrated Pest Information

1261   Platform for Extension and Education (iPiPE): progress through sharing. J Integr Pest

1262   Manag. 2015;6: 1–8. https://doi.org/10.1093/jipm/pmv013

1263   100.   Moran E V., Alexander JM. Evolutionary responses to global change: Lessons from

1264   invasive species. Ecol Lett. 2014;17: 637–649. https://doi.org/10.1111/ele.12262

1265   101.   Sinclair BJ, Williams CM, Terblanche JS. Variation in thermal performance among insect

1266   populations. Physiol Biochem Zool. 2012;85: 594–606. https://doi.org/10.1086/665388

1267   102.   Gray DR, Keena MA. A phenology model for Asian gypsy moth egg hatch. Environ

1268   Entomol. 2019;48: 903–910. https://doi.org/10.1093/ee/nvz051

1269

1270   **Supporting information**

1271   **S1 Appendix Estimating phenology model parameters for *Epiphyas postvittana*.**

1272   (PDF)

1273

1274   **S2 Appendix Data sources for validating a DDRP phenology model for *Epiphyas***

1275   ***postvittana.***

1276   (PDF)

1277

1278   **S3 Appendix Estimating a common lower temperature threshold and other phenology**

1279   **model parameters for *Neoleucinodes elegantalis*.**

1280   (PDF)

1281

1282   **S4 Appendix Methods for fitting and validating a CLIMEX model for *Neoleucinodes***

1283   ***elegantalis*.**

1284   (PDF)

1285

1286   **S1 Table Locality records used for validating the CLIMEX and DDRP climatic suitability**

1287   **models for *Neoleucinodes elegantalis*.**

1288   (PDF)

1289

1290   **S2 Table Comparison of DDRP predictions for the dates of first spring egg laying**

1291   **("Predicted") to the month (data set 1) or dates (data sets 2 and 3) of peak spring adult**

1292   **flight for *Epiphyas postvittana* in California according to three monitoring data sets**

1293   **("Observed").**

1294   (PDF)

1295

1296   **S3 Table DDRP predictions of the number of degree-days Celsius (DDC) that accumulated**

1297   **between peaks in last fall flight and first spring flight of *Epiphyas postvittana* in California.**

1298   (PDF)

1299

1300   **S1 Fig Predictions of climatic suitability for *Epiphyas postvittana* in Australia, New**

1301   **Zealand, and California based on 1961–1990 climate normals according to (A, B) CLIMEX**

1302   **and (C) DDRP.**

1303   (PDF)

1304

1305   **S2 Fig CLIMEX model for *Neoleucinodes elegantalis* in the Neotropics.**

1306   (PDF)

1307

1308   **S3 Fig DDRP predictions of cold and heat stress for (A) *Epiphyas postvittana* and (B)**

1309   ***Neoleucinodes elegantalis* for 2018.**

1310   (PDF)

1311

1312 **S4 Fig CLIMEX predictions of dry stress for *Neoleucinodes elegantalis* in CONUS.**

1313   (PDF)

Fig. 1

Fig. 2

Fig. 3

Fig. 4

Fig. 5

Fig. 6

(A) *Epiphyas postvittana*

(B) *Neoleucinodes elegantalis*

No. of
generations

- excl.-sev
- excl.-mod
- 0 gens.
- 1 gens.
- 2 gens.
- 3 gens.
- 4 gens.
- 5 gens.
- 6 gens.
- 7 gens.
- 8 gens.
- 9 gens.
- 10 gens.