

DDRP: real-time phenology and climatic suitability modeling of invasive insects

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Short title: DDRP: a new modeling tool for invasive insects

Abstract

1 Rapidly detecting and responding to new invasive species and the spread of those that are
2 already established is essential for reducing their potential threat to food production, the
3 economy, and the environment. We describe a new multi-species spatial modeling platform that
4 integrates mapping of phenology and climatic suitability in real-time to provide timely and
5 comprehensive guidance for stakeholders needing to know both where and when invasive insect
6 species could potentially invade the conterminous United States. The Degree-Days, Risk, and
7 Phenological event mapping (DDRP) platform serves as an open-source and relatively easy-to-
8 parameterize decision support tool to help detect new invasive threats, schedule monitoring and
9 management actions, optimize biological control, and predict potential impacts on agricultural
10 production. DDRP uses a process-based modeling approach in which degree-days and
11 temperature stress are calculated daily and accumulate over time to model phenology and
12 climatic suitability, respectively. Products include predictions of the number of completed
13 generations, life stages present, dates of phenological events, and climatically suitable areas
14 based on two levels of climate stress. Species parameters can be derived from laboratory and
15 field studies, and from published and newly fitted CLIMEX models. DDRP is written entirely in
16 R, making it flexible and extensible, and capitalizes on multiple R packages to generate gridded
17 and graphical outputs. We illustrate the DDRP modeling platform and the process of model
18 parameterization using two invasive insect species as example threats to United States
19 agriculture: the light brown apple moth, *Epiphyas postvittana*, and the small tomato borer,
20 *Neoleucinodes elegantalis*. We then discuss example applications of DDRP as a decision support
21 tool, review its potential limitations and sources of model error, and outline some ideas for future
22 improvements to the platform.

23 **Introduction**

24 Invasive insects in the United States are a significant threat to the economy, environment, food
25 security, and human health [1–3]. They cause billions of dollars in damage to forests each year
26 [1,2], and their potential cost to food crop production is among the highest of any country [3].
27 Insect invasions in the United States also reduce the abundance and diversity of native species,
28 which negatively impacts ecosystem functions and services such as soil health, nutrient cycling,
29 and wildlife habitat [1,2]. Rapidly detecting and responding to new invasive insects and the
30 spread of those that are already established before they can cause significant economic and
31 environmental damage has therefore become a major priority [2,4].

32 Modeling climatic suitability (risk of establishment) and the timing of seasonal activities
33 (phenology) of invasive insect species can help stakeholders including farmers, natural resource
34 managers, and surveillance teams detect and prevent their establishment, slow their spread, and
35 manage existing populations more sustainably and economically [5]. Estimates of climatic
36 suitability identify areas to concentrate surveillance or management resources and efforts [6–8],
37 whereas real-time (i.e. current) or forecasted predictions of phenology can improve the timing of
38 surveillance and integrated pest management (IPM) efforts such as monitoring device
39 installation, pesticide applications, and biological control release [9–11]. Additionally, estimates
40 of climatic suitability, phenology, and voltinism (number of generations per year) can help
41 growers predict the impact of pests and diseases on agricultural production and associated
42 economic losses [12].

43 Degree-day models that predict insect phenology are an established tool for decision support
44 systems that assist stakeholders with scheduling surveillance, monitoring or IPM operations for
45 numerous pest species over the growing season [13–16]. Most degree-day models predict

46 phenology by measuring linear relationships between temperature and development rate, and
47 they employ daily time steps to estimate degree-days using daily minimum and maximum
48 temperature (T_{min} and T_{max} , respectively) data. In the daily time step, degree-days accumulate if
49 heat exceeds the lower developmental temperature threshold of a species (and below its upper
50 threshold for some calculation methods) during a 24-hour period [13,14,16]. Several web-based
51 platforms host degree-day models for insect pest species in the United States, offering users
52 opportunities to model phenology of multiple species at single locations (site-based model) or
53 across a certain area (spatialized model). These platforms include but are not limited to Michigan
54 State University's Enviroweather (<https://www.enviroweather.msu.edu>), Oregon State
55 University's USPEST.ORG (<https://uspest.org/wea/>), and the USA National Phenology Network
56 (<https://www.usanpn.org>) [17,18].

57 Despite their widespread use, currently available degree-day modelling platforms are in need
58 of improvements. None of them integrate predictions of phenology and climatic suitability,
59 which would provide guidance on the question of both where and when—e.g. is an area at high
60 risk of establishment, and if so, then when will the species emerge or begin a specific activity?
61 For most species, addressing this two-part question would require finding, potentially
62 purchasing, and learning how to use two separate platforms. Additionally, many phenology
63 modeling platforms use oversimplified models that make broad assumptions about insect
64 biology, such as assuming a single lower developmental temperature threshold for multiple
65 species, or assuming that an entire population emerges from overwintering at a single time.
66 However, developmental temperature thresholds may vary widely across insect species, and
67 development rates often vary within populations [19–21]. A biologically unrealistic model may
68 produce inaccurate predictions of phenological events (e.g. spring emergence, first adult flight,

69 egg-hatching) or voltinism. Moreover, most platforms are capable of forecasting phenology only
70 a week or two into the future in specific states or regions. However, stakeholders may need to
71 plan operations several weeks in advance, potentially in areas that are outside the geographic
72 bounds of predictive models.

73 In this study, we introduce a new spatial modeling platform, DDRP (short for Degree-Days,
74 Establishment Risk, and Phenological event maps) that generates real-time and forecast
75 predictions of phenology and climatic suitability (risk of establishment) of invasive insect
76 species in the conterminous United States (CONUS). The objective of DDRP is to improve the
77 efficiency and effectiveness of programs that aim to detect new or spreading invasive insect
78 species in the United States, or to monitor and manage species that are already well-established.
79 The platform is written entirely in the R statistical programming language (R Development Core
80 Team 2019), making it flexible and extensible, and has a simple command-line interface that can
81 be readily implemented for online use. Gridded temperature data for DDRP may include the
82 entire CONUS or a specific region or state, and may be at any spatial resolution that can be
83 handled by the user's computing system. DDRP will generally use observed and future (forecast
84 or recent average) temperature data because it was designed to be run as a within-season decision
85 support tool that can provide guidance on where and when to expect the pest to appear each year,
86 but it will accept temperature data for any time period. Model products include gridded (raster)
87 and graphical (map) outputs of life stages present, number of generations, phenological events,
88 and climatic suitability.

89 First, we describe the modeling process and workflow of DDRP, summarize types of model
90 products (Fig 1), and review its system and software requirements. Next, we demonstrate its
91 capabilities and the process of model parameterization using two invasive insect species which

92 threaten agricultural biosecurity in the United States: the small tomato borer, *Neoleucinodes*
93 *elegantalis* [Guenée (Lepidoptera: Crambidae)], and the light brown apple moth, *Epiphyas*
94 *postvittana* [Walker, 1863 (Lepidoptera: Tortricidae)]. These species were chosen because they
95 have been well-studied in terms of their developmental requirements, and previous climatic
96 suitability studies provide a basis for parameterizing the climatic suitability model in DDRP.
97 Additionally, models for these species are intended to aid surveillance teams at the Cooperative
98 Agricultural Pest Survey (CAPS) pest detection program, which supports the USDA Animal and
99 Plant Health Inspection Service (APHIS) as it works to safeguard agricultural and environmental
100 resources in the United States. The DDRP platform will be a useful decision support tool for
101 preventing, monitoring, and managing new and emerging invasive pests in the United States.

102

103 **Methods**

104 **1) Model inputs**

105 *Temperature data*

106 DDRP requires daily T_{min} and T_{max} data in a gridded format for an area of interest in CONUS.
107 For real-time modeling, we have been using daily T_{min} and T_{max} data at a 4 km spatial resolution
108 from the PRISM (Parameter-elevation Relationships on Independent Slopes Model) database
109 (available at <http://www.prism.oregonstate.edu>) [22]. Daily PRISM data become available *ca.* 1
110 day after weather station observations are reported, and are typically updated and improve in
111 quality as more observations are added (see PRISM website for details). The phenology mapping
112 system of the USA National Phenology Network [18] uses Real-Time Mesoscale Analysis
113 (RTMA) weather data at a 2.5 km resolution, which are available within hours after data are
114 observed. The daily T_{min} and T_{max} RTMA data set could potentially be used in DDRP; however,

115 the RTMA methodology lacks PRISM's update and quality control regimes [22]. Another
116 alternative is Daymet v3, which offers daily climate data for North America, Hawaii, and Puerto
117 Rico at a very high spatial resolution (1 km) (<https://daymet.ornl.gov>) [23]. However, Daymet
118 data are released months after the end of each year, so they would be less useful for within-
119 season modeling and decision support.

120 For forecast modeling, DDRP is currently configured to use either monthly-updated, daily-
121 downscaled NMME (North American Multi-Model Ensemble) 7-month forecasts at a 4 km
122 resolution [24], or recent 10-year average PRISM daily data that are calculated on a bi-monthly
123 basis. We consider 10-year average data to be an improvement over 30-year climate normals for
124 producing forecasts because temperatures in CONUS have significantly increased over the past
125 30 years [25,26]. The match of mean forecasts of the NMME model's ensemble to the observed
126 value (i.e. skill) varies both spatially and temporally due to topography, season, and the presence
127 of an El Niño-Southern Oscillation (ENSO) signal [27,28]. It may therefore be more
128 conservative, and provide more consistent predictions, to use 10-year averages instead of NMME
129 data to avoid potential issues with skill. However, we caution that the 10-year average data do
130 not have simulated variation in daily T_{min} and T_{max} , which may result in the under-prediction of
131 degree-day accumulation in the spring or fall as daily T_{max} only slightly exceeds the lower
132 developmental threshold of a species, or for cooler sites that have temperatures that are often
133 near the threshold. We have also prepared and plan to use the National Weather Service gridded
134 National Digital Forecast Database (NDFD) 7-day forecasts
135 (https://www.weather.gov/mdl/ndfd_info) [29] for use in DDRP.

136

137 ***Phenology modeling: species data and parameters***

138 The life history and behavior of a target species must be considered for appropriateness to model
139 in DDRP. In its current form, the platform can model four separate life stages (the egg, the larva
140 or nymph, the pupa or pre-oviposition, and the adult) plus a separately parameterized
141 overwintering stage. As movement and migration are not handled by DDRP, it is currently
142 limited in its ability to model migratory species, such as those that may establish in southern
143 areas of their potential range and migrate yearly to more northern areas. Species that lack an
144 overwintering stage, which are common in tropical and subtropical areas, may be difficult to
145 model because the timing of first spring activities and stages present cannot be accurately
146 estimated. Currently DDRP is entirely temperature-driven, so species whose growth and
147 reproduction are strongly influenced by additional environmental factors such as day length or
148 moisture may not be accurately modeled.

149 DDRP requires data on the developmental temperature thresholds (in either degrees Celsius
150 or Fahrenheit) and durations for each life stage of an insect species in degree-days (Fig 1 and
151 Table 1). These data are typically collected in the laboratory by measuring how temperature
152 influences the rate of development, although data derived from season-long monitoring studies
153 are also used [15,30]. A different developmental threshold may be assigned to each stage,
154 although we typically solve for a common threshold if differences across the stages are minimal.
155 Presently, the model depends upon a fixed starting date such as January 1, specified by the user
156 for the entire region of interest. The duration of the overwintering stage represents the number of
157 degree-days that must accumulate from the start of the year for the stage to complete, and
158 indicates the start time for the model since other stages would then commence develop.

159 Users must specify the number of degree-days that are required for the overwintering stage to
160 complete development and emerge for the growing season. These data are typically gathered

161 using field monitoring studies, whereby the temporal distribution of emergence times and
162 number of individuals that emerge on a given date is documented [e.g. 32]. Assigning a single
163 value to the overwintering stage duration parameter would assume that an entire population
164 develops simultaneously, which may not be biologically realistic because several intrinsic (with
165 a genetic basis) and extrinsic (e.g. microclimate, nutrition) factors can produce variation in
166 development rates within a species [21,32]. Indeed, phenology models that incorporate
167 developmental variability in a population may have increased predictive power [19,21,33].
168 DDRP therefore allows the duration of the overwintering stage to vary across a user-defined
169 number of cohorts (groups of individuals in a population). Much of the intrinsic variability in
170 insect development during a generation often occurs in the first stadium after overwintering [34],
171 although developmental variation may occur in any life stage [19,35,36]. DDRP uses five
172 parameters to generate a frequency distribution of emergence times: the mean, variance, low
173 bound, and high bound of emergence times, and the shape of the distribution (Gaussian or
174 lognormal; Table 1). The platform uses these data to estimate the relative size of the population
175 represented by each cohort, which initializes the population distribution that is maintained during
176 subsequent stages and generations. Individuals within each cohort develop in synchrony.

177 Users may specify the timing (in degree-days) of phenological events that are important to
178 their target system to generate phenological event maps in DDRP, which depict the estimated
179 calendar dates of the event over a time frame of interest. We typically generate phenological
180 event maps based on temperature data for an entire year so that events for multiple generations of
181 each of the five life stages are modeled. For example, phenological event maps that depict when
182 the overwintering stage would emerge may be useful for identifying start dates for surveillance
183 operations for a species, whereas maps for subsequent generations could help with planning

184 operations later in the year. The timing of phenological events may be based on life stage
185 durations (e.g. the end of the egg stage signifies egg-hatching), or on occurrences within a stage
186 such as midpoint (to approximate peak) of oviposition or midpoint of adult flight. Currently, one
187 user-defined phenological event for each life stage for up to four generations may be modeled,
188 although the platform could be modified to predict multiple events for each stage (e.g. first,
189 midpoint, and end of the stage) for any numbers of generations.

190

191 *Climate suitability modeling: species data and parameters*

192 Climatic suitability modeling in DDRP is based on cold and heat stress accumulation and
193 requires data on temperature stress threshold and limits of a species (Fig 1 and Table 1). While
194 estimates of these parameters may be estimated from laboratory or field experiments, such data
195 are lacking for most species. Additionally, extrapolating laboratory data to the field to project
196 accumulation of stress is difficult due to oversimplification of the number of variables and the
197 temporal and spatial variation in natural environments [37]. We have been using the CLIMEX
198 software [38] (Hearne Scientific Software, Melbourne, Australia) to assist with climatic
199 suitability model parameterization in DDRP, which is one of the most widely used species
200 distribution modeling tools for agricultural, livestock and forestry pests and non-pests [6,7].
201 Laboratory collected data may help with parameterizing a CLIMEX model; however, model
202 parameters are fine-tuned and the model is fitted using observations from the species' known
203 geographical distribution [38,39]. CLIMEX uses a similar process-based modeling approach as
204 DDRP, wherein climatic suitability of an area to a species is influenced by climate stress
205 accumulation. Model products generated by the two programs, including maps of temperature
206 stress accumulation, are therefore directly comparable.

207 DDRP was designed to be complementary to CLIMEX in several ways to facilitate climatic
208 suitability model parameterization, but the two programs also differ in several respects (Table 2).
209 Importantly, DDRP was designed to model climatic suitability based on daily current or forecast
210 temperature data at fine spatial scales (e.g. a single state or region), which would give users
211 insight into the potential risk of establishment or spread during a particular season or year. In
212 contrast, CLIMEX is normally used to estimate a species' potential distribution using coarse-
213 scale 10' and 30' resolution global gridded 30-year monthly climate normals centered on 1975
214 (1961–1990) or future projections from selected global circulation models (GCMs) [40]. In
215 theory CLIMEX could be used for real-time climatic suitability, but it has no native ability to
216 import and process common gridded formats and is incapable of using daily resolution climate
217 data. Thus, DDRP's climate suitability models are intended to improve the efficiency of
218 surveillance and trap deployment at a relatively small focal area for a current or near-future time
219 period, whereas CLIMEX models provide a more general and coarse-scale assessment of
220 suitability based on averaged climate data.

221 Relying on real-time climatic suitability models for decision support on where to employ pest
222 management and eradication operations for a given year or season is preferable to using models
223 based on 30-year climate normals. A model that uses current climate data is more biologically
224 relevant because the risk of establishment in an area would be affected by the conditions that a
225 species physically experiences, not by averages of historical climate. Additionally, climate in
226 CONUS is changing rapidly, so models based on climate normals may produce unrealistic
227 predictions of present-day climatic suitability. Over the past *ca.* 30 years, the average annual
228 temperature in CONUS has increased by 1.2°F (0.7°C), the number of freezing days has
229 declined, and extreme temperature events have increased in frequency and intensity [25,26].

230 Nonetheless, DDRP is not currently capable of including moisture factors in the modeling
231 process like CLIMEX, so model predictions for moisture-sensitive species in very arid or wet
232 areas should be interpreted with caution. We present a more detailed comparison of the features
233 of each platform in Table 2, and discuss the potential implications of generating a climate
234 suitability model based solely on temperature in the ‘Discussion.’

235 We compare CLIMEX’s predictions of temperature stress accumulation and overall climate
236 suitability to similar products in DDRP to help parameterize a DDRP climate suitability model.
237 Temperature stress thresholds may be calibrated so that predictions of cold and heat stress
238 accumulation at the end of the year are spatially concordant with CLIMEX’s predictions.
239 Climatic suitability in CLIMEX is estimated with the Ecoclimatic Index (EI), which is scaled
240 from 0 to 100, and integrates the annual growth index and climate stress indices to give an
241 overall measure of favorableness of a location or year for long-term occupation by the target
242 species [38,39]. Typically an EI approaching 0 indicates an unfavorable climate for long-term
243 survival, while an EI exceeding 20 or 30 (depending on the species) indicates a highly suitable
244 climate [38,39]. As discussed in more detail in ‘Case Studies’, temperature stress limits in DDRP
245 can be adjusted so that areas predicted to have highly suitability according to CLIMEX are also
246 included in DDRP’s prediction of the potential distribution.

247 Comparing DDRP climatic suitability model products to those of CLIMEX for model fitting
248 purposes is naturally more appropriate when temperature data are derived from the same time
249 period. We have therefore been using a PRISM T_{min} and T_{max} 30-year average dataset centered on
250 1975 (1961–1990) to match the time-schedule of the CliMond CM10 (also 1961–1990) world
251 climate dataset currently supplied with CLIMEX [40]. We temporally downscaled monthly
252 PRISM estimates for 1961–1990 because DDRP requires daily data and PRISM daily

253 temperature data for years prior to 1980 are not available. For each month of a given year, a
254 bilinear interpolation method was used to assign each day an average temperature value that was
255 iteratively smoothed and then adjusted so that the monthly averages were correct.

256

257 **2) Daily time step**

258 DDRP models insect phenology and climatic suitability by stepping through each day of a
259 specified time period and calculating degree-day and temperature stress accumulation at each
260 grid cell of a focal area (Fig 1). The time period may span the entire year of interest, or include
261 only a subset of days such as those during the growing season. Users may sample and save daily
262 modeling results every 30 days, 14 days, 10 days, seven days, two days, or one day. Results are
263 saved in multi-layer rasters that are processed and analyzed after the daily time step to produce
264 final model products. We describe the phenology and climatic suitability modeling process and
265 products in more detail in the following sections.

266

267 *Phenology model*

268 DDRP calculates daily degree-days over the specified time period using developmental
269 temperature threshold information and gridded temperature data that have been cropped to the
270 extent of the focal area (Fig 1). Currently DDRP has three methods to calculate degree-days: a
271 simple average, the simple average using an upper threshold with a horizontal cutoff, and the
272 single triangle method with upper threshold [41–43]. All three methods calculate degree-days
273 from the daily T_{min} and T_{max} data and a specified base (lower developmental threshold)
274 temperature, but the latter two methods also include the upper developmental threshold in

275 calculations. The single triangle method is also used as a close approximation to the more
276 complex sine-curve calculation method [43].

277 With the exception of phenological event maps, which are computed only for the last day of
278 the daily time step, DDRP saves the following phenology model results for each sampled day:

- 279 1. *Accumulated degree-days*. While daily degree days are calculated for each life stage, the
280 cumulative degree-days are summed only for the first cohort of the larval stage, as these
281 degree-day maps are representative for all cohorts and life stages. Accumulated degree-days
282 calculated for larva will be the same for other life stages if common developmental
283 thresholds are used.
- 284 2. *Life stages*. The life stage present (overwintering stage, eggs, larvae, pupae, and adults) for
285 each cohort.
- 286 3. *Number of generations*. The current generation count for each cohort. If the model is run for
287 an entire year, then the output for the last day of the year would represent the potential
288 voltinism of the species. The generation count increases when adults progress to the egg
289 stage (i.e. oviposition occurs).
- 290 4. *Phenological event maps (optional)*. The timing of phenological events is estimated by
291 computing daily degree-day totals from the gridded temperature data, and storing the day of
292 year when an event threshold is reached. Event results are generated only on the last day of
293 the daily time step (typically, the last day of the year) because the entire time period must be
294 analyzed for all potential event days to be considered.

295

296 *Climatic suitability model*

297 Independently from the phenology model simulations, DDRP steps through each day and
298 calculates cold and heat stress accumulation to predict the potential distribution of the species
299 (Fig 1). To simplify model parameterization and increase processing efficiency, it assumes that
300 all life stages have the same climatic tolerances. Cold stress units are calculated as the difference
301 between daily T_{min} and the cold stress temperature threshold, whereas heat stress units are
302 calculated as the difference between daily T_{max} and the heat stress temperature threshold. Stress
303 units accumulate across the entire time period of interest (i.e. across all life stages and
304 generations) and are presumed to kill individuals either directly or indirectly when they exceed
305 the moderate or severe stress unit limits. Stress could indirectly kill individuals by restricting
306 their activity, or directly cause mortality through extreme cold or heat events such as a hard
307 freeze. For CONUS, the northern range limit is typically delineated by cold stress and the
308 southern range limit, if any, is delineated by heat stress.

309 We opted to use moderate and severe stress limits to reflect two distinct themes. First, they
310 may provide a way to depict the potential for short term vs. longer term establishment. For most
311 species, the potential distribution could be represented by areas where cold and heat stress have
312 not exceeded the severe or moderate stress limits, as these should allow for long-term population
313 persistence. DDRP depicts these areas with maps of cold stress exclusion, heat stress exclusion,
314 and all stress exclusion (cold plus heat stress exclusions; Fig 1). Areas under moderate stress
315 exclusion may represent temporary zones of establishment in which a species establishes only
316 during a favorable season, such as after an annual migration event. Conversely, areas under
317 severe stress exclusion do not allow for even short-term establishment. Typically we visualize
318 exclusion maps calculated for the last day of the year (day 365) under investigation to provide
319 insight into the potential distribution for an entire growing season.

320 Second, using two levels of stress may provide a way to represent uncertainty for estimating
321 the potential distribution. As discussed in more detail in the ‘Discussion,’ several sources of
322 uncertainty and error in the modeling process may bias model predictions, such as applying
323 inappropriate parameter values, using climate data with low skill or poor spatial resolution,
324 ignoring biotic factors such as species interactions, or ignoring non-temperature abiotic factors
325 such as microclimate effects, moisture, and photoperiod [32,44]. Defining the potential
326 distribution as areas under severe stress only would typically provide a broader estimate than a
327 definition based on both stress levels. While this approach may over-predict the risk of
328 establishment, conducting surveys over too broad an area is probably better than surveying too
329 small of an area, which may allow a new invasive species to establish and spread.

330

331 **3. Post time step processing**

332 After the daily time step has completed, DDRP combines and analyzes results across cohorts and
333 generates final multi-layer rasters and summary maps (“.png” image files) for each sampled day.
334 If multiple cohorts were modeled, then DDRP uses estimates of the relative size of the
335 population represented by each cohort to calculate the relative size of the population (totaling
336 100%) in any given life stage and generation. For phenological event maps, the earliest and
337 average day of year that an event occurs across cohorts is calculated.

338 DDRP integrates mapping of phenology and climate suitability so that users can use a single
339 model product to obtain guidance on their “where” and “when” questions (Fig 1). For example, a
340 user involved with planning surveys may want to know where a target species may establish, and
341 within those areas, when populations may emerge from over-wintering. Each output of the
342 phenology model with the exception of accumulated degree-days will be associated with two

343 additional products for each sampled day (or the last day for a phenological event map): 1) one
344 that includes severe stress exclusion only, and 2) one that includes both severe and moderate
345 stress exclusions. For example, a phenological event map with severe and moderate stress
346 exclusions for 2018 (all 365 days) would present predicted dates of the selected event only in
347 areas where long-term establishment is predicted (Fig 2).

348

349 *System and software requirements*

350 DDRP requires the R statistical software platform and can be run from the command line or
351 within RStudio [45]. It takes advantage of functions from several R packages for data
352 manipulation, analysis, and post-model processing. The ‘raster’ package [46] is used to crop
353 daily temperature rasters to the focal area, store and manipulate daily loop raster results, and
354 process and further analyze results for each cohort. Many non-spatial data manipulations are
355 conducted with functions in the ‘dplyr,’ ‘tidyr,’ and ‘stringr’ packages [47–49]. The ‘ggplot2’
356 package [50] is used to generate and save summary maps of raster outputs, and options from the
357 command line argument are parsed using the ‘optparse’ package [51].

358 DDRP capitalizes on the multi-processing capabilities of modern servers to run multiple
359 operations in parallel, which is made possible with the ‘parallel’ and ‘doParallel’ packages in R
360 [52]. This significantly reduces computation times, particularly in cases where modeling is
361 conducted with multiple cohorts and across large areas. For example, parallel processing is used
362 to crop rasters for multiple dates, run multiple cohorts in the daily time step, and to analyze time
363 step outputs for multiple days or files simultaneously. For very large areas (currently defined as
364 the Eastern United States and CONUS), temperature rasters are split into four tiles and both the
365 tiles and cohorts are run in parallel in the daily time step.

366 We recommend running DDRP on a Linux OS because its multicore functionality supports
367 the fork system calls of the ‘parallel’ package in R. The script can easily be modified to be run
368 on a Windows OS; however, certain processes are very memory intensive and may execute
369 slowly or stall without parallel processing. Additionally, DDRP was designed to run on a Linux
370 server, and has not been sufficiently tested on Windows servers to know how well it would
371 perform.

372

373 **Case studies**

374 ***Climatic suitability, voltinism, and phenological events in *Epiphyas postvittana****

375 The light brown apple moth (LBAM), *E. postvittana* (Walker 1863) (Lepidoptera: Tortricidae),
376 is a leafroller pest native to southeastern Australia, including Tasmania [53]. The species invaded
377 Western Australia, New Zealand, New Caledonia, England, and Hawaii more than 100 years ago
378 [54–56], and has been established in coastal California since 2006 [57,58]. It poses a significant
379 threat to agricultural production in the United States because it feeds on more than 360 host
380 plants, including economically important fruits such as apple, pear, citrus and grapes [53,59,60].
381 For example, an economic risk analysis of *E. postvittana* to four major fruit crops (apple, grape,
382 orange, and pear) in CONUS estimated an annual mean cost of US\$105 million associated with
383 damage to crops and control, quarantine, and research [59]. The CAPS program at APHIS
384 conducts annual surveys for *E. postvittana* at various counties across CONUS.

385 A summary of phenology and climatic suitability model parameters used for *E. postvittana* in
386 DDRP is reported in Table 2. We assigned all life stages a lower developmental threshold of
387 7.1°C (45°F) and an upper developmental threshold of 31.1°C (88°F). Although DDRP allows
388 for different temperature thresholds for each stage, the site-based phenology modeling tool that

389 we use at USPEST.ORG requires common thresholds, and these are presented as whole numbers
390 in Fahrenheit scale for easy interpretation by end-users. Building the model for both platforms
391 keeps models simpler and facilitates cross-comparison. Additionally, laboratory studies revealed
392 small differences in the lower developmental threshold ($< 1^{\circ}\text{C}$) across different life stages
393 [53,60]. The upper developmental threshold value is based on studies showing that all life stages
394 cease development between $31\text{--}32^{\circ}\text{C}$ [53,60,61]. We derived life stage durations (in degree-days
395 $^{\circ}\text{C}$; hereafter DDC) for *E. postvittana* based on our analysis of published research [53,60,62],
396 which resulted in 127, 408, 128, and 71 DDC for eggs, larvae (females on young apple foliage),
397 pupae, and adults to 50% egg-laying, respectively. This analysis is presented in S1 Appendix.

398 We set the overwintering stage to larva because the predominant overwintering stage of *E.*
399 *postvittana* in the United States are the late larval instars [31,63]. We applied seven cohorts to
400 approximate a normal distribution of emergence times that spanned 100 to 320 DDC (average =
401 210 DDC) based on a report that overwintering larvae at four sites in California required
402 between 102 and 318 degree-days to finish development [31]. This would correspond to the time
403 required for mid-stage (3rd-5th instars, average 4th instar) female larval feeding on old foliage
404 ($0.45 \times 494 \text{ DDC} = 210 \text{ DDC}$), after a January 1 start date.

405 We generated a CLIMEX model for *E. postvittana* using CLIMEX version 4.0 [38] to help
406 parameterize the climatic suitability model in DDRP. The model applied a combination of
407 parameter values (Table 3) derived from two previous CLIMEX studies of this species [64,65].
408 However, we used a cold stress threshold (TTCS) of 3°C , which is lower than He *et al.*'s (2012)
409 value (5°C)[64], and higher than Lozier and Mill's (2011) value (1.5°C) [65]. We applied a top-
410 up irrigation (additional simulated rainfall) rate of 2.5 mm day^{-1} for the winter and summer
411 season because irrigation mitigates the hot-dry climate that limits distribution of *E. postvittana*

412 within CLIMEX. We assessed the fit of the CLIMEX model to observation-based data from 530
413 locality records for the species from Australia (N = 317), New Zealand (N = 76), and California
414 (N = 137), which were obtained from GBIF.org (18th July 2019; GBIF Occurrence Download
415 <https://doi.org/10.15468/dl.a4ucei>) and Nick Mills (pers. comm.). The model correctly predicted
416 suitable conditions ($EI > 20$) at all 530 localities where the species is known to occur (S1 Fig).

417 In DDRP, we generated a climatic suitability model for *E. postvittana* using the daily
418 downscaled PRISM T_{max} and T_{min} estimates for 1961–1990 and calibrated model parameters in
419 accordance with the CLIMEX model (Fig 3). We compared maps of temperature stress
420 accumulation, and adjusted temperature stress limits so that most areas predicted to be under
421 moderate and severe climate stress by DDRP had low ($20 > EI > 0$) or zero ($EI = 0$) suitability
422 according to CLIMEX, respectively. Finally, we modeled phenology and climatic suitability for
423 *E. postvittana* in 2018 to provide insight into its potential voltinism, seasonal activities, and risk
424 of invasion in particularly warm temperatures. The summer of 2018 in the United States was the
425 warmest since 2012 and tied for the fourth-warmest on record (NOAA website
426 <https://www.noaa.gov/news/summer-2018-ranked-4th-hottest-on-record-for-us> last accessed
427 11/21/19). We generated a phenological event map that depicted the date of first egg laying by
428 first generation females, because this activity is relevant to monitoring both eggs and the
429 emergence of adults, which typically occurs two to three days prior to egg laying.

430

431 ***Climatic suitability, voltinism, and phenological events in Neoleucinodes elegantalis***

432 The small tomato borer (STB), *N. elegantalis* (Guenée) (Lepidoptera: Crambidae), is native to
433 South America and is distributed throughout the Neotropics including in Mexico, Central
434 America, and the Caribbean [66,67]. A major insect pest of tomato (*Solanum lycopersicum*), it

435 also attacks fruits of other plants belonging to the family Solanaceae including eggplant, paprika,
436 naranjilla, and green and red pepper [67]. There are at least 1175 recorded interceptions of the
437 species from the United States, where it is considered a serious threat to agricultural biosecurity
438 because it lowers tomato production in South America [68]. The CAPS program has conducted
439 surveillance for *N. elegantalis* since at least 2011.

440 A summary of phenology and climatic suitability model parameters used for *N. elegantalis* in
441 DDRP is reported in Table 2. We re-analyzed data from a laboratory study on the development
442 of *N. elegantalis* on hybrid tomato (Paronset) at five temperatures [69] to estimate a common
443 lower temperature threshold for all life stages, which involved adding a point to force the x-
444 intercept to an integer value in degrees Fahrenheit. We weighted the analysis to select a common
445 lower threshold for immature stages, which are the longest in duration, because this should
446 produce the lowest error for the overall life cycle. The lower threshold values for immature
447 stages were very similar to the overall egg-to-adult value of 8.89°C (48°F), so we chose 8.89°C
448 as the common threshold instead of a higher one solved for the adult pre-oviposition stage
449 (11.5°C). We estimated the duration for eggs, larvae, pupae, and adults to peak oviposition as 86,
450 283, 203, and 96 DDC, respectively. This analysis is presented in S2 Appendix.

451 We compared the results of our method to estimate a common threshold to the alternative,
452 whereby separate thresholds for each life stage are derived and used in the DDRP model (S2
453 Appendix). The error resulting from a slight forcing of the linear regression model was nominal.
454 For example, the values for the x-intercept and 1/slope for the egg-to-adult interval of *N.*
455 *elegantalis* without forcing the regression were 8.64°C (47.6°F) and 1048.3, respectively. After
456 constructing the model with forcing, these values were 8.89°C (48.0°F) and 1030.8, respectively.
457 Further, models that used egg-to-adult development times calculated by the forced versus

458 unforced method were run with various start dates including January 1 and April 1, and
459 predictions differed by only 1 day.

460 *Neoleucinodes elegantalis* has no apparent photoperiodic response, diapause, or specific
461 overwintering stage. In subtropical climates in Brazil, the insect remains active throughout the
462 year if host plants are available [69]. We used January 1 as the model start date for CONUS
463 because few host plants would be available for immature stages at this time, leaving adults as the
464 overwintering stage. We assumed that adult feeding and host search activities could begin
465 immediately if temperatures are suitable, and that first egg-laying would subsequently occur after
466 the estimated pre-oviposition period of *ca.* 55 DDC. The durations of later events (1st to peak
467 oviposition, immature development, etc.) were estimated from previously published data [69,70].
468 We applied seven cohorts to approximate a normal distribution of emergence times that spanned
469 0 to 111 DDC (average = 50 DDC) because over-wintered adults begin finding hosts over this
470 time frame.

471 We parameterized the DDRP climatic suitability model by working with previously
472 published CLIMEX models for *N. elegantalis*, which were constructed using data on its
473 development at different temperatures, its known distribution in the Neotropics, and a sensitivity
474 analysis of CLIMEX parameters [71–73]. We produced a CLIMEX model for *N. elegantalis* in
475 CONUS that applied the “best-fit” parameter values proposed by da Silva et al. 2018 [71](Table
476 3) and a top-up irrigation rate of 2.5 mm day⁻¹ for the winter and summer season. However, we
477 applied a cold stress threshold of 2°C because the species may experience this temperature at
478 high-elevation areas where it occurs in Colombia (Bogota, elevation = *ca.* 2600 m) [74] and in
479 Ecuador (El Chaco, elevation = 1600 m), as documented in records of the lowest monthly

480 temperatures for these areas (available for 2017 to the present-day at <https://rp5.ru/> and
481 <https://www.worldweatheronline.com>, respectively) [75].

482 We calibrated climatic suitability parameters for *N. elegantalis* in DDRP in accordance with
483 CLIMEX results (Fig 4) using the same approach taken for *E. postvittana*. Next, we modeled
484 phenology and climatic suitability in DDRP using temperature data from 2018. We generated a
485 phenological event map for the average date of the beginning of egg hatch of the overwintered
486 generation. Predictions of egg hatch could enhance population control of *N. elegantalis* because
487 this species is most vulnerable to pesticides before larvae enter the fruit of host plants [76].

488

489 **Results**

490 Cold stress was the major determinant of the potential distribution of *E. postvittana* and *N.*
491 *elegantalis* according to the DDRP analyses based on 30-year climate normals (1961–1990).
492 Both species were excluded from the northern half of CONUS by cold stress, with the exception
493 of (mostly) western parts of Oregon and Washington (Figs 3D and 4D). Heat stress excluded *E.*
494 *postvittana* from most of the Southwest and from southern parts of Gulf Coast states (Fig 3E),
495 whereas *N. elegantalis* was excluded only from the hottest parts of the Southwest in southern
496 California, Arizona, and Texas (Fig 4E). When considering both cold and heat stress exclusions,
497 the potential distribution of both species included western parts of the Pacific states (California,
498 Oregon, and Washington), most of the Southeast, and southern parts of the Northeast (in
499 Delaware, Maryland, New Jersey, and Rhode Island) and the Midwest (in Kansas, Missouri,
500 Illinois, and Indiana). The potential distribution for *N. elegantalis* included a greater area of
501 Oklahoma and Texas than the distribution for *E. postvittana*.

502 DDRP may be over-predicting the potential distribution of *E. postvittana* and *N. elegantalis*
503 in particularly arid regions of the Southwest. According to CLIMEX, high levels of dry stress
504 (S2 Fig) were associated with low climatic suitability ($EI < 10$) throughout most of western
505 Texas, southern New Mexico, and Arizona (Figs 3C and 4C). Conversely, DDRP did not exclude
506 the species from some of these same areas (Figs 3F and 4F), which is most likely due to the
507 platform's current inability to consider moisture factors in the modeling process.

508 DDRP predicted a smaller potential distribution for *E. postvittana* and *N. elegantalis* in 2018
509 compared to 1961–1990 (Figs 2 and 5). High levels of heat stress in 2018 (S3 and S4 Figs)
510 excluded both species from warm areas of CONUS that were included in the potential
511 distribution under historical conditions, including parts of Arizona, New Mexico, Texas, and the
512 Central Valley of California. Additionally, *E. postvittana* was excluded from several areas in the
513 Southeast that were historically suitable (e.g. in South Carolina, Louisiana, Alabama, and
514 Arkansas). In the East, moderate stress exclusion resulting from cold stress (S3 and S4 Figs)
515 pushed the northern range limit for each species slightly farther south, although this shift was
516 more pronounced for *N. elegantalis*.

517 Predictions of potential dates for phenological events and voltinism for *E. postvittana* and *N.*
518 *elegantalis* in 2018 varied substantially by latitude (Figs 2 and 5). The earliest date of egg-laying
519 for the first generation of *E. postvittana* was predicted to be as early as February in Florida to as
520 late as December in the Pacific Northwest (Fig 2A). The average date of first generation
521 beginning of egg hatch for *N. elegantalis* was predicted to begin in the first week of January in
522 Florida but not until late June in the Pacific Northwest (Fig 2B). Thus, the timing of monitoring
523 trap installation to detect ovipositing adults and eggs of *E. postvittana*, or larvae of *N.*
524 *elegantalis*, could vary widely across CONUS. For both species, DDRP predicted as many as

525 seven to nine generations in coastal areas of the Southeast, compared to only one or two
526 generations in parts of the Pacific Northwest (Fig 5). Three to six generations were predicted for
527 most other regions of CONUS. These findings may indicate that the Southeast would experience
528 the longest duration of pest pressure.

529

530 **Discussion**

531 **DDRP as a decision support tool**

532 DDRP is a new spatial modeling platform that generates real-time and forecast predictions of
533 phenology and climatic suitability (risk of establishment) of invasive insect pests in CONUS.
534 These predictions may help with detecting the presence of invasive species in the shortest time
535 possible after they arrive and reproduce, which increases the chance of eradication success and
536 makes other rapid response measures (e.g. deployment of biological control) possible and less
537 costly [4]. In particular, DDRP's unique ability to produce integrated climatic suitability and
538 phenology model products can provide biosurveillance personnel with insights into both where
539 and when to focus detection efforts for a given year or growing season. For example,
540 phenological event maps for *E. postvittana* and *N. elegantalis* (Fig 2) identify high-risk areas
541 where surveillance activities could be concentrated, in addition to providing estimated dates of
542 activities that can ensure timely trap placement. Thus, users can use a single program to address
543 decision support needs for early pest detection.

544 DDRP was designed to be a multi-species platform, which makes it broadly applicable to
545 monitoring and managing populations of IPM pests and classical biological control agents. For
546 example, growers have used predictions of the timing of first egg hatch for codling moth [*Cydia*
547 *pomonella* (Linnaeus)], first emergence of western cherry fruit fly [*Rhagoletis indifferens*

548 (Curran)], and first spring oviposition of spotted wing drosophila [*Drosophila suzukii*
549 (Matsumura)] to help monitor and reduce populations of these major crop pests [77–80].
550 Phenology models for biological control insects can help managers schedule sampling trips to
551 coincide with insect presence on the target organism, and to estimate the synchrony of insect and
552 host phenology at a given location [81,82]. DDRP’s estimates of voltinism may provide insights
553 into expected pressure on target organisms, as higher voltinism should translate to greater agent
554 population growth and biocontrol success.

555 Our models for *E. postvittana* and *N. elegantalis* indicated that heat stress excluded
556 populations from a greater area of the Southwest compared to 1961–1990, which is consistent
557 with studies showing that global warming may reduce species’ distributions in warmer parts of
558 their range [83]. However, determining whether these putative range shifts are persistent or
559 temporary would require combining model runs for multiple recent years or seasons. For
560 example, trends in the geographic distribution of stress exclusions over several years or seasons
561 could be visualized with a probability surface map. Estimating the direction of range shifts may
562 also provide insights into the response of the species to future climate change. Nonetheless, the
563 differences that we documented in predictions of climatic suitability based on climate data for
564 1961–1990 compared to 2018 suggests that an area’s contemporary risk of establishment is
565 different than it was *ca.* 30 years ago. DDRP’s ability to produce climatic suitability models in
566 real-time may provide more meaningful insights into areas that are presently suitable for an
567 invasive species, and therefore allow for more effective placement of surveillance operations.

568 DDRP considers several aspects of insect development in the modeling process, which
569 should increase the realism of model predictions compared to simpler platforms that are based
570 solely on generation time degree-days. For example, DDRP’s estimates of voltinism and spring

571 activity will likely be more accurate if knowledge of the biology of a given species is sufficient,
572 because different temperature thresholds for multiple life stages may be used, and the
573 overwintering stage is parameterized separately from the post-winter stage (e.g. overwintering
574 adult vs. adult). Additionally, it accounts for developmental variation within populations by
575 generating and combining results across multiple cohorts that complete the overwintering stage
576 at different times, which is more realistic than assuming a single population cohort and single
577 model start time.

578 From a historical perspective, DDRP could be considered a partial descendant and spatialized
579 version of the PETE (Predictive Extension Timing Estimator) phenological modeling platform
580 that was established as a standard in the mid-1970s [84]. Features in common include a cohort
581 approach to population phenological modeling using daily degree-days as the main input,
582 provision for major insect life stages and a separately parameterized overwintering distribution,
583 an open-source non-proprietary standard for sharing, and a focus on agricultural extension
584 (applied decision support). Unlike PETE, DDRP is spatialized and therefore able to produce a
585 variety of mapping products including phenological event maps, and it also includes options to
586 use separate thresholds for different life stages and to generate climatic suitability models. DDRP
587 could be improved by adding certain features of the PETE platform including the use of a
588 diapause trigger, and a distributed delay function that would allow the generation time to
589 lengthen with each subsequent generation.

590 DDRP is one of the first open-source platforms that can model both phenology and climatic
591 suitability of insect pests. The Insect Life Cycle Modelling (ILCYM) software [85,86] is another
592 open-source R program that can model temperature-based phenology and establishment risk for
593 insects in a spatial context. However, ILCYM is a full population dynamics modeling platform

594 that requires life table data at constant and variable temperatures, which are seldom available for
595 anticipated but not yet present invasive insect species. ILCYM has primarily been used to
596 simulate models with climate normals or GCMs at global or regional scales [85,87], whereas
597 DDRP was designed to use real-time and forecast climate data for within-season decision
598 support. PhenModel is another R program that can model insect phenology, but it is unclear how
599 adaptable the program would be to other insect species besides leaf-feeding beetles, and it is not
600 spatialized or capable of modeling climatic suitability [88].

601 There are numerous opportunities for improving and extending the applications of DDRP,
602 especially given that it is written in R, an open source programming language. For example, the
603 program could be modified to use climate data for a region outside of CONUS, or to model other
604 types of organisms such as non-insect invertebrates, plants, plant-pathogenic bacteria and fungi,
605 and insect plant and animal virus vectors. The platform has been tested for and could be used
606 through an on-line web interface, although there is the potential for overload issues on a server
607 host. This issue, as yet untested, could be addressed by using a cloud computing platform. We
608 describe some additional features that could be added to potentially improve model accuracy and
609 expand the list of products in more detail below. The most recent code for DDRP is available at
610 GitHub (https://github.com/bbarker505/ddrp_v2.git), where we invite scientists and practitioners
611 to jointly develop the platform and database of species models.

612

613 **Uncertainties, limitations, and other considerations**

614 *Linear (degree-day) modeling*

615 DDRP uses a relatively simple degree-day modeling approach, whereas some platforms
616 including ILCYM, phenModel, and devRate [89] offer complex functions to model nonlinear

617 responses of insects to temperature. Degree-day models are ideal for multi-species platforms like
618 DDRP because there are sufficient data to parameterize a degree-day model for most insect pest
619 species of economic importance in the United States [20,90]. Linear degree-day models are also
620 readily calibrated and sometimes constructed entirely using field data, making them more
621 practical for extension and decision support use [15]. Additionally, degree-day models require
622 only daily T_{min} and T_{max} data (as opposed to hourly data for most nonlinear models), which are
623 available at a high spatial resolution for CONUS from multiple sources including PRISM and
624 RTMA. Nonetheless, it is important for users to recognize potential sources of error and lack of
625 precision in degree-day models, such as their limited ability to accurately model development at
626 supra-optimal temperatures [15,91].

627

628 *Environmental inputs*

629 DDRP is intentionally parameterized in a simple, conservative manner, which will hopefully
630 achieve the goal of a parsimonious balance of both model simplicity and accuracy [15,92].
631 Nonetheless, DDRP is driven entirely by temperature, and therefore ignores other factors that
632 may affect the development and distribution of insects such as photoperiod, moisture, dispersal,
633 resources, disturbance, and biotic interactions [7,93]. The potential consequences of this
634 limitation will depend on the biology of the organism under study. For example, dry stress is the
635 major factor restricting the current distribution of *N. elegantalis* in its native range [72,74,94],
636 and it limits the distribution of *E. postvittana* both in its native range [53,95] and in Southern
637 California and Arizona [62]. The absence of moisture factors in DDRP most likely explains why
638 it predicted higher climatic suitability for these species in arid regions of CONUS compared to
639 CLIMEX, which considers wet and dry stress in the modeling process. However, this

640 conservative-leaning error may in fact better reflect human manipulation of the landscape (e.g.
641 greenhouse and irrigation usage) that may allow the species to exist in such regions. Future
642 versions of DDRP that can process gridded moisture data and incorporate moisture stress factors
643 into climatic suitability models may help overcome our current limitations in matching CLIMEX
644 models, and they may improve predictions for moisture-sensitive species such as *E. postvittana*
645 and *N. elegantalis*. Additionally, we are developing a version of DDRP that incorporates
646 photoperiodically induced life history events such as winter diapause and summer aestivation,
647 which builds on earlier phenology modeling work that estimated voltinism of photoperiod-
648 sensitive insects [96].

649

650 ***Presumptive models***

651 Uncertainties regarding the accuracy of temporal or spatial predictions of invasive species that
652 are not yet established is inevitable, in part because no validation data are yet available, and
653 species interceptions do not imply establishment [7,97]. DDRP models for species for which
654 only presumptive models exist should therefore be used conservatively. For example,
655 surveillance or management actions could be implemented in advance of predicted phenological
656 events as a precautionary measure (e.g. installing traps even earlier than estimates for the earliest
657 date of overwintering adult emergence). To potentially avoid under-predicting the risk of
658 establishment, the potential distribution could be defined as areas not under severe climate stress
659 as opposed to defining it using both stress levels. Additionally, climatic suitability models
660 generated by DDRP could be combined with those produced using different modeling methods
661 (e.g. correlative, semi-mechanistic, mechanistic) to create a “hybrid” model, which may increase
662 the reliability of predictions [7,93].

663 Web platforms that support sharing of pest observations and related expert commentary will
664 be valuable resources for validating and increasing the predictive performance of DDRP models.
665 For example, the iPIPE and its sister platforms (<http://www.ipipe.org>, <https://ipmpipe.org>) have
666 created a national information technology infrastructure for sharing pest observations in near
667 real-time and contributing them to a national depository [98]. Similarly, the USA National
668 Phenology Network provides a depository of plant and insect phenology observations
669 contributed by citizen scientists [9]. The National Agricultural Pest Information System (NAPIS;
670 <https://napis.ceris.purdue.edu/home>) currently has over 5.17 million records from pest detection
671 surveys, and is another potential source of validation data. Unfortunately, there were insufficient
672 data from these sources to validate our DDRP model for *E. postvittana*, which we hope will
673 change in the near future.

674

675 ***Geographic variation***

676 A major challenge for insect phenology modeling in general is how to account for geographic
677 variation in insect responses to temperature [32,44]. Populations of an invading species may
678 diverge in thermal physiology traits when genetically divergent individuals are introduced to
679 different areas, or when evolutionary changes including rapid adaptation to new environments or
680 hybridization occurs [99,100]. If sufficient data on the amount and geographic distribution of
681 variation in relevant traits exists, then model accuracy may be improved by building separate
682 models for each genotype. For example, an egg hatch phenology model for a subspecies of the
683 Asian gypsy moth, *Lymantria dispar asiatica* (Vnukovskij), had reduced error compared to a
684 similar model constructed for the European subspecies that has invaded North America,
685 *Lymantria dispar dispar* (Linnaeus), which has a markedly different predominant phenotype

686 [101]. An alternative approach may be to run several models, each with a different value for the
687 parameter of interest, and present a range of model predictions. Conversely, DDRP could be
688 modified to accept a grid of parameter values so that geographic variation would be accounted
689 for in a single model run.

690 A lack of knowledge on how early-season environmental conditions or events that initiate the
691 first spring activity of a species (biofix) vary geographically may be a source of error because the
692 model start date affects all downstream predictions. For example, how does first spring activity
693 vary across the wide range of warming conditions possibly encountered for a large region such
694 as CONUS? As a case in point, our phenology model for *N. elegantalis* assumes that moths have
695 only 55 DDC before egg-laying behaviors may occur. This assumption may not be valid for sub-
696 tropical zones of the United States, where flight and reproduction could occur even earlier.

697 Conversely, a much longer spring warm-up may be needed in temperate zones because
698 commercial tomatoes are transplanted much later in the year. Studying how first spring activity
699 (adult flight) in *N. elegantalis* potentially varies geographically in Central or South America
700 would help to refine a range of model start times. The phenology model for DDRP could then be
701 parameterized using a necessarily conservative selection of start dates or by inputting a grid of
702 start dates. Using a broad distribution of emergence times to initiate the cohorts could be another
703 approach to accommodate uncertainty in first spring activity.

704

705 ***Distributed delay***

706 There is currently no distributed delay function in DDRP, meaning that the overlap in
707 generations and life stages of cohorts does not increase over multiple generations. Modeling
708 distributed delay can be important because the timing of surveillance activities or IPM events

709 like spraying or trapping need to be adjusted for later generations [35,36,84]. Not accounting for
710 distributed delay may be a greater issue for species that continue to develop throughout winter
711 months or lack a temperature or photoperiodic event that synchronize populations, such as *E.*
712 *postvittana* [31,56,60]. For many multivoltine species, DDRP may accurately predict peak events
713 in each generation, but inaccurately predict the first appearance of one or more life stages after
714 the first or second generations because of increasing overlap in generation cohorts. Thus,
715 phenological event maps produced for species that have significant overlap in generations should
716 be most reliable for the first few generations. This will be among the high priority issues in
717 development of future versions of the platform.

718

719 **Conclusion**

720 DDRP is a new modeling tool that can integrate mapping of phenology and climatic suitability in
721 real-time to provide timely and comprehensive guidance for stakeholders needing to know
722 where and when invasive species may establish. When used for surveillance, the platform will
723 hopefully increase chances for early detection of new or spreading invasive threats in the United
724 States, and therefore help pest management programs mitigate their potential damage to
725 agricultural and environmental resources. Additionally, DDRP may help plan monitoring and
726 management efforts for IPM pests and biological control insects, and to predict pest pressure on
727 host plants.

728 The case studies we presented provided examples of how models for DDRP may be
729 parameterized and then run to produce various products including gridded and graphical
730 predictions of the number of generations, life stages present, dates of phenological events, and
731 areas of climatic suitability based on two levels of climate stress. We encourage users of DDRP

732 to consider the limitations of the platform, to report the conditions that their model was designed
733 to work under (e.g. a particular region, life stage event, or generation), and to document and any
734 known sources of model error that could not be accounted for when providing validation and
735 other feedback reports. The flexible and open-source nature of DDRP will facilitate making
736 modifications and improvements, such as adding new environmental factors, using it for other
737 regions besides CONUS, modeling non-insect organisms, expanding the types of model
738 products, and adding features to improve model accuracy.

739

740 **Supporting information**

741 **S1 Appendix Estimating phenology model parameters for *Epiphyas postvittana*.**

742 (DOC)

743

744 **S2 Appendix Estimating a common lower temperature threshold and other phenology
745 model parameters for *Neoleucinodes elegantalis*.**

746 (DOC)

747

748 **S1 Fig Predictions of climatic suitability for *Epiphyas postvittana* in Australia, New**

749 **Zealand, and California according to CLIMEX (a, b) and DDRP (c) models based on**

750 **1961–1990 climate normals.** Climatic suitability of an area in CLIMEX is represented by the

751 Ecoclimatic Index (EI), where EI = 0 indicates unsuitable conditions. In DDRP, long-term

752 establishment is indicated by areas not under moderate or severe climate stress. Blue circles

753 depict known localities for the species (Australia $N = 317$; New Zealand $N = 76$; California $N =$

754 137).

755 (DOC)

756

757 **S2 Fig CLIMEX predictions of dry stress in CONUS for *Epiphyas postvittana* and**

758 ***Neoleucinodes elegantalis* based on 1961–1990 climate normals.**

759 (DOC)

760

761 **S3 Fig DDRP predictions of cold and heat stress for *Epiphyas postvittana* for 2018.**

762 (DOC)

763

764 **S4 Fig DDRP predictions of cold and heat stress for *Neoleucinodes elegantalis* for 2018.**

765 (DOC)

766

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777

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793

794 **Data availability**

795 The most current R code for DDRP is available at github.com/bbarker505/ddrp_v2. The version

796 of DDRP used for this study has been archived at Zenodo

797 (<https://doi.org/10.5281/zenodo.3832731>). Perl and Octave scripts used for temporal

798 downscaling of monthly climate averages to daily averages are at GitHub

799 (github.com/bbarker505/dailynorms) and Zenodo (<https://doi.org/10.5281/zenodo.3601671>).

800 Output data produced by this analysis are at GitHub (github.com/bbarker505/1990_daily_30yr)
801 and Zenodo (<https://doi.org/10.5281/zenodo.3833053>).

802

803 **References**

- 804 1. Pimentel D, Zuniga R, Morrison D. Update on the environmental and economic costs
805 associated with alien-invasive species in the United States. *Ecol Econ.* 2005; 52: 273–288.
806 <https://doi.org/10.1016/j.ecolecon.2004.10.002>
- 807 2. Lovett GM, Weiss M, Liebhold AM, Holmes TP, Leung B, Lambert KF, et al. Nonnative
808 forest insects and pathogens in the United States: Impacts and policy options. *Ecol Appl.*
809 2016; 26: 1437–1455. <https://doi.org/10.1890/15-1176>
- 810 3. Paini DR, Sheppard AW, Cook DC, Barro PJ De, Worner SP, Thomas MB. Global threat
811 to agriculture from invasive species. *Proc Natl Acad Sci.* 2016; 113: 7575–7579.
812 <https://doi.org/10.1073/pnas.1602205113>
- 813 4. United States Department of the Interior. Safeguarding America’s lands and waters from
814 invasive species: a national framework for early detection and rapid response. Washington
815 D.C., USA; 2016. p. 55.
- 816 5. Tonnang HEZ, Hervé BDB, Biber-freudenberger L, Salifu D, Subramanian S, Ngowi VB,
817 et al. Advances in crop insect modelling methods — Towards a whole system approach.
818 *Ecol Modell.* 2017; 354: 88–103. <https://doi.org/10.1016/j.ecolmodel.2017.03.015>
- 819 6. Venette RC. Climate analyses to assess risks from invasive forest insects: simple matching
820 to advanced models. *Curr For Reports.* 2017; 3: 255–268. [https://doi.org/10.1007/s40725-](https://doi.org/10.1007/s40725-017-0061-4)
821 [017-0061-4](https://doi.org/10.1007/s40725-017-0061-4)

- 822 7. Lantschner MV, de la Vega G, Corley JC. Predicting the distribution of harmful species
823 and their natural enemies in agricultural, livestock and forestry systems: an overview. *Int J*
824 *Pest Manag.* 2019; 65: 190–206. <https://doi.org/10.1080/09670874.2018.1533664>
- 825 8. Kriticos DJ, Venette RC, Koch F, Rafoss T, Van der Werf W, Worner S. Invasive alien
826 species in the food chain: advancing risk assessment models to address climate change,
827 economics and uncertainty. *NeoBiota.* 2013; 18: 1–7.
828 <https://doi.org/10.3897/neobiota.18.6108>
- 829 9. Crimmins TM, Crimmins MA, Gerst KL, Rosemartin AH, Weltzin JF. USA National
830 Phenology Network’s volunteer contributed observations yield predictive models of
831 phenological transitions. *PLoS One.* 2017; 12: e0182919.
832 <https://doi.org/10.1371/journal.pone.0182919>
- 833 10. Prasad YG, Prabhakar M. Pest monitoring and forecasting. In: Abrol D, Shankar U,
834 editors. *Integrated Pest Management: Principles and Practice.* Wallingford, United
835 Kingdom: CABI; 2012. pp. 41–57. <https://doi.org/10.1079/9781845938086.0041>
- 836 11. Orlandini S, Magarey RD, Park EW, Kroschel J. Methods of agroclimatology: modeling
837 approaches for pests and diseases. In: Hatfield J, Sivakuma M, Prueger J, editors.
838 *Agroclimatology: Linking Agriculture to Climate.* American Society of Agronomy, Crop
839 Science Society of America, and Soil Science Society of America, Inc.; 2017. pp. 1–36.
840 <https://doi.org/10.2134/agronmonogr60.2016.0027>
- 841 12. Donatelli M, Magarey RD, Bregaglio S, Willocquet L, Whish JPM, Savary S. Modelling
842 the impacts of pests and diseases on agricultural systems. *Agric Syst.* 2017; 155: 213–224.
843 <https://doi.org/10.1016/j.agsy.2017.01.019>

- 844 13. Pruess KP. Degree-day methods for pest management. *Environ Entomol.* 1983; 12: 613–
845 619. <https://doi.org/10.1093/ee/12.3.613>
- 846 14. Higley LG, Pedigo LP, Ostlie R. DEGDAY: a program for calculating degree-days, and
847 assumptions behind the degree-day approach. *Environ Entomol.* 1986; 15: 999–1016.
848 <https://doi.org/10.1093/ee/15.5.999>
- 849 15. Coop LB, Barker BS. Advances in understanding species ecology: phenological and life
850 cycle modeling of insect pests. In: Kogan M, Heinrichs E, editors. *Integrated Management*
851 *of Insect Pests: Current and Future Developments*. Sawston, England; 2020. pp. 43–96.
852 <https://doi.org/10.19103/AS.2019.0047.02>
- 853 16. Herms DA. Using degree-days and plant phenology to predict pest activity. In: Krischik
854 V, Davidson J, editors. *IPM (Integrated Pest Management) of Midwest Landscapes*
855 *Minnesota Agricultural Experiment Station Publication SB-07645*. St. Paul, MN; 2004.
856 pp. 49–59.
- 857 17. Crimmins TM, Marsh RL, Switzer JR, Crimmins MA, Gerst KL, Rosemartin AH, et al.
858 USA National Phenology Network gridded products documentation. *US Geol Surv Open-*
859 *File Rep 2011-1263*. 2017. <https://doi.org/10.3133/ofr20171003>
- 860 18. Crimmins TM, Gerst KL, Huerta DG, Marsh RL, Posthumus EE, Rosemartin AH, et al.
861 Short-term forecasts of insect phenology inform pest management. *Ann Entomol Soc Am.*
862 2020; 113: 139–148. <https://doi.org/10.1093/aesa/saz026>
- 863 19. Struelens Q, Rebaudo F, Quispe R, Dangles O. Thermal pace-of-life strategies improve
864 phenological predictions in ectotherms. *Sci Rep.* 2018; 8: 1–9.

- 865 <https://doi.org/10.1038/s41598-018-34274-1>
- 866 20. Jarošík V, Honěk A, Magarey RD, Skuhrovec J. Developmental database for phenology
867 models: related insect and mite species have similar thermal requirements. *J Econ*
868 *Entomol.* 2011; 104: 1870–1876. <https://doi.org/10.1603/ec11247>
- 869 21. Régnière J, Powell JA. Animal life cycle models (Poikilotherms). In: Schwartz MD,
870 editor. *Phenology: An Integrative Environmental Science*. Dordrecht, The Netherlands:
871 Springer Netherlands; 2013. pp. 295–316. https://doi.org/10.1007/978-94-007-6925-0_16
- 872 22. Daly C, Halbleib H, Smith J, Gibson W, Doggett M, Taylor G, et al. Physiographically
873 sensitive mapping of climatological temperature and precipitation across the conterminous
874 United States. *Int J Climatol.* 2008; 28: 2031–2064. <https://doi.org/10.1002/joc.1688>
- 875 23. Thornton PE, Thornton M., Mayer BW, Wei Y, Devarakonda R, Vose RS, et al. Daymet:
876 Daily Surface Weather Data on a 1-km Grid for North America, Version 3. Oak Ridge,
877 Tennessee, USA: ORNL DAAC; 2018. <https://doi.org/10.3334/ORNLDAAC/1328>
- 878 24. Kirtman BP, Min D, Infanti JM, Kinter JL, Paolino DA, Zhang Q, et al. The North
879 American multimodel ensemble: Phase-1 seasonal-to-interannual prediction; phase-2
880 toward developing intraseasonal prediction. *Bull Am Meteorol Soc.* 2014; 95: 585–601.
881 <https://doi.org/10.1175/BAMS-D-12-00050.1>
- 882 25. Vose RS, Easterling ER, Kunkel KE, LeGrande EN, Wehner MF. Temperature changes in
883 the United States. In: Wuebbles DJ, Fahey DW, Hibbard KA, Dokken DJ, Stewart BC,
884 Maycock TK, editors. *Climate Science Special Report: Fourth National Climate*
885 *Assessment, Volume I*. Washington D.C., USA; 2017. pp. 185–206.

- 886 <https://doi.org/10.7930/J0N29V45>
- 887 26. Wuebbles D, Meehl G, Hayhoe K, Karl TR, Kunkel K, Santer B, et al. CMIP5 climate
888 model analyses: Climate extremes in the United States. *Bull Am Meteorol Soc.* 2014; 95:
889 571–583. <https://doi.org/10.1175/BAMS-D-12-00172.1>
- 890 27. Barbero R, Abatzoglou JT, Hegewisch KC. Evaluation of statistical downscaling of North
891 American Multimodel Ensemble Forecasts over the western United States. *Weather*
892 *Forecast.* 2017; 32: 327–341. <https://doi.org/10.1175/waf-d-16-0117.1>
- 893 28. Roundy JK, Yuan X, Schaake J, Wood EF. A framework for diagnosing seasonal
894 prediction through canonical event analysis. *Mon Weather Rev.* 2015; 143: 2404–2418.
895 <https://doi.org/10.1175/MWR-D-14-00190.1>
- 896 29. Glahn HR, Ruth DP. The new digital forecast database of the national weather service.
897 *Bull Am Meteorol Soc.* 2003; 84: 195–201. <https://doi.org/10.1175/BAMS-84-2-195>
- 898 30. Régnière J, Powell J, Bentz B, Nealis V. Effects of temperature on development, survival
899 and reproduction of insects: experimental design, data analysis and modeling. *J Insect*
900 *Physiol.* 2012; 58: 634–647. <https://doi.org/10.1016/j.jinsphys.2012.01.010>
- 901 31. Bürgi LP, Roltsch WJ, Mills NJ. Abundance, age structure, and voltinism of light brown
902 apple moth populations in California. *Environ Entomol.* 2011; 40: 1370–1377.
903 <https://doi.org/10.1603/en11165>
- 904 32. Rebaudo F, Rabhi VB. Modeling temperature-dependent development rate and phenology
905 in insects: review of major developments, challenges, and future directions. *Entomol Exp*
906 *Appl.* 2018; 166: 607–617. <https://doi.org/10.1111/eea.12693>

- 907 33. Yonow T, Zalucki MP, Sutherst RW, Dominiak BC, Maywald GF, Maelzer DA, et al.
908 Modelling the population dynamics of the Queensland fruit fly, *Bactrocera (Dacus)*
909 *tryoni*: a cohort-based approach incorporating the effects of weather. *Ecol Modell.* 2004;
910 173: 9–30. [https://doi.org/10.1016/S0304-3800\(03\)00306-5](https://doi.org/10.1016/S0304-3800(03)00306-5)
- 911 34. Régnière J. A method of describing and using variability in development rates for the
912 simulation of insect phenology. *Can Entomol.* 1984; 116: 1367–1376.
913 <https://doi.org/10.4039/Ent1161367-10>
- 914 35. Severini M, Baumgärtner J, Limonta L. Parameter estimation for distributed delay based
915 population models from laboratory data: egg hatching of *Oulema duftschmidi*
916 Redthenbacher (Coleoptera, Chrysomelidae) as an example. *Ecol Modell.* 2003; 167: 233–
917 246. [https://doi.org/10.1016/S0304-3800\(03\)00188-1](https://doi.org/10.1016/S0304-3800(03)00188-1)
- 918 36. Wagner TL, Wu H-I, Feldman RM, Sharpe PJH, Coulson RN. Multiple-cohort approach
919 for simulating development of insect populations under variable temperatures. *Ann*
920 *Entomol Soc Am.* 1985; 78: 691–704. <https://doi.org/10.1093/aesa/78.6.691>
- 921 37. Sutherst RW. Pest species distribution modelling: origins and lessons from history. *Biol*
922 *Invasions.* 2014; 16: 239–256. <https://doi.org/10.1007/s10530-013-0523-y>
- 923 38. Kriticos DJ, Maywald GF, Yonow T, Zurcher EJ, Herrmann N, Sutherst RW. CLIMEX
924 Version 4: Exploring the effects of climate on plants, animals and diseases. CSIRO,
925 Canberra, Australia; 2016. p. 156. <https://publications.csiro.au/pu>.
- 926 39. Sutherst RW, Maywald GF. A computerised system for matching climates in ecology.
927 *Agric Ecosyst Environ.* 1985; 13: 281–299. [https://doi.org/10.1016/0167-8809\(85\)90016-](https://doi.org/10.1016/0167-8809(85)90016-)

- 928 7
- 929 40. Kriticos DJ, Webber BL, Leriche A, Ota N, Macadam I, Bathols J, et al. CliMond: global
930 high-resolution historical and future scenario climate surfaces for bioclimatic modelling.
931 *Methods Ecol Evol.* 2012; 3: 53–64. <https://doi.org/10.1111/j.2041-210X.2011.00134.x>
- 932 41. Lindsey A, Newman J. Use of official weather data in spring time: temperature analysis of
933 an Indiana phenological record. *Ecology.* 1956; 4: 812–823.
934 <https://doi.org/10.2307/1933072>
- 935 42. Sevacherian V, Stern VM, Mueller AJ. Heat accumulation for timing *Lygus* control
936 measures in a safflower-cotton complex. *J Econ Entomol.* 1977; 70: 399–402.
937 <https://doi.org/10.1093/jee/70.4.399>
- 938 43. Roltsch W, Zalom A, Strand JF, Pitcairn MJ. Evaluation of several degree-day estimation
939 methods in California climates. *Int J Biometeorol.* 1999; 42: 169–176.
940 <https://doi.org/10.1007/s004840050101>
- 941 44. Chuine I, Régnière J. Process-based models of phenology for plants and animals. *Annu*
942 *Rev Ecol Evol Syst.* 2017; 48: 159–182. [https://doi.org/10.1146/annurev-ecolsys-110316-](https://doi.org/10.1146/annurev-ecolsys-110316-022706)
943 [022706](https://doi.org/10.1146/annurev-ecolsys-110316-022706)
- 944 45. RStudio Team. RStudio: Integrated Development for R. Boston, MA, USA; 2018.
- 945 46. Hijmans RJ. raster: Geographic Data Analysis and Modeling. R package version 2.8-4.
946 <https://CRAN.R-project.org/package=raster>. 2018.
- 947 47. Wickham H, Henry L. tidyr: easily tidy data with “spread()” and “gather()” functions. R
948 package version 0.8.1. <https://CRAN.R-project.org/package=tidyr>. 2018.

- 949 48. Wickham H. stringr: simple, consistent wrappers for common string operations. R
950 package version 1.3.1. <https://CRAN.R-project.org/package=stringr>. 2018.
- 951 49. Wickham H, François R, Henry L, Müller K. dplyr: a grammar of data manipulation. R
952 package version 0.7.8. <https://CRAN.R-project.org/package=dplyr>. 2018.
- 953 50. Wickham H. ggplot2: elegant graphics for data analysis. New York.
954 <https://ggplot2.tidyverse.org>: Springer-Verlag; 2016.
- 955 51. Davis TL. optparse: Command Line Option Parser. R package version 1.6.1.
956 <https://CRAN.R-project.org/package=optparse>. 2019.
- 957 52. Weston S, Calaway R. Getting started with doParallel and foreach. Vignette, CRAN, URL
958 <http://cran.r-project.org/web/packages/doParallel/index.html>. 2018.
- 959 53. Geier PW, Briese DT. The light-brown apple moth, *Epiphyas postvittana* (Walker): a
960 native leafroller fostered by European settlement. In: Kitching PL, Jones RE, editors. The
961 Ecology of Pests: Some Australian Case Histories. Canberra, Australia: CSIRO; 1981. pp.
962 130–155.
- 963 54. Geier PW, Springett BP. Population characteristics of Australian leafrollers (*Epiphyas*
964 *spp.*, Lepidoptera) infesting orchards. *Aust J Ecol.* 1976; 1: 127–144.
965 <https://doi.org/10.1111/j.1442-9993.1976.tb01102.x>
- 966 55. Suckling DM, Brockerhoff EG. Invasion biology, ecology, and management of the light
967 brown apple moth (Tortricidae). *Annu Rev Entomol.* 2010; 55: 285–306.
968 <https://doi.org/10.1146/annurev-ento-112408-085311>
- 969 56. Dumbleton L. The apple leaf-roller (*Tortrix postvittana* Walker). *New Zeal J Sci Technol.*

- 970 1932; 14: 83–92.
- 971 57. Varela LG, Johnson MW, Strand L, Wilen CA, Pickel C. Light brown apple moth’s
972 arrival in California worries commodity groups. *Calif Agric.* 2008; 62: 57–61.
973 <https://doi.org/10.3733/ca.v062n02p57>
- 974 58. Suckling DM, Stringer LD, Baird DB, Butler RC, Sullivan TES, Lance DR, et al. Light
975 brown apple moth (*Epiphyas postvittana*) (Lepidoptera: Tortricidae) colonization of
976 California. *Biol Invasions.* 2014; 16: 1851–1863. [https://doi.org/10.1007/s10530-013-](https://doi.org/10.1007/s10530-013-0631-8)
977 0631-8
- 978 59. Fowler G, Garrett L, Neeley A, Borchert D, Spears B. Economic analysis: risk to U.S.
979 apple, grape, orange and pear production from the light brown apple moth, *Epiphyas*
980 *postvittana* (Walker). USDA-APHIS-PPQ-CPHST-PERAL. Raleigh, NC; 2007.
- 981 60. Danthanarayana W. The bionomics, distribution and host range of the light brown apple
982 moth, *Epiphyas postvittana* (Walk.) (Tortricidae). *Aust J Zool.* 1975; 23: 419–437.
983 <https://doi.org/10.1071/ZO9750419>
- 984 61. Danthanarayana W, Gu H, Ashley S. Population growth of *Epiphyas postvittana*, the
985 lightbrown apple moth (Lepidoptera: Tortricidae) in relation to diet, temperature and
986 climate. *Aust J Zool.* 1995; 43: 381–394. <https://doi.org/10.1071/ZO9950381>
- 987 62. Gutierrez AP, Mills NJ, Ponti L. Limits to the potential distribution of light brown apple
988 moth in Arizona-California based on climate suitability and host plant availability. *Biol*
989 *Invasions.* 2010; 12: 3319–3331. <https://doi.org/10.1007/s10530-010-9725-8>
- 990 63. Bürgi LP, Mills NJ. Cold tolerance of the overwintering larval instars of light brown apple

- 991 moth *Epiphyas postvittana*. J Insect Physiol. 2010; 56: 1645–1650.
- 992 <https://doi.org/10.1016/j.jinsphys.2010.06.009>
- 993 64. He S, Worner SP, Ikeda T. Modeling the potential global distribution of light brown apple
994 moth *Epiphyas postvittana* (Lepidoptera: Tortricidae) using CLIMEX. J Asia Pac
995 Entomol. 2012; 15: 479–485. <https://doi.org/10.1016/j.aspen.2012.01.004>
- 996 65. Lozier JD, Mills NJ. Predicting the potential invasive range of light brown apple moth
997 (*Epiphyas postvittana*) using biologically informed and correlative species distribution
998 models. Biol Invasions. 2011; 13: 2409–2421. <https://doi.org/10.1007/s10530-011-0052-5>
- 999 66. Díaz-Montilla A, Suárez-Baron H, Gallego-Sánchez G, Saldamando-Benjumea C, Tohme
1000 J. Geographic differentiation of Colombian *Neoleucinodes elegantalis* (Lepidoptera:
1001 Crambidae) haplotypes: evidence for Solanaceae host plant association and Holdridge life
1002 zones for genetic differentiation. Ann Entomol Soc Am. 2013; 106: 586–597.
1003 <https://doi.org/10.1603/AN12111>
- 1004 67. Bulletin OEPP/EPPO. *Neoleucinodes elegantalis*. 2015; 45: 9–13.
1005 <https://doi.org/10.1111/epp.12189>
- 1006 68. Díaz-Montilla A, Solis MA, Kondo T. The tomato fruit borer, *Neoleucinodes elegantalis*
1007 (Guenée) (Lepidoptera: Crambidae), an insect pest of neotropical solanaceous fruits. In:
1008 Peña J, editor. Potential Invasive Pests of Agricultural Crops. Boston, MA: CABI; 2013.
1009 pp. 137–159.
- 1010 69. Moraes CP, Foerster LA. Thermal requirements, fertility, and number of generations of
1011 *Neoleucinodes elegantalis* (Guenée) (Lepidoptera: Crambidae). Neotrop Entomol. 2015;

- 1012 44: 338–344. <https://doi.org/10.1007/s13744-015-0293-6>
- 1013 70. Moraes CP, Foerster LA. Development and reproduction of *Neoleucinodes elegantalis*
1014 (Lepidoptera: Crambidae) on tomato (*Solanum lycopersicum*) cultivars. Rev Colomb
1015 Entomol. 2014; 40: 40–43.
- 1016 71. da Silva RS, Kumar L, Shabani F, Picanço MC. An analysis of sensitivity of CLIMEX
1017 parameters in mapping species potential distribution and the broad-scale changes observed
1018 with minor variations in parameter values: an investigation using open-field *Solanum*
1019 *lycopersicum* and *Neoleucinodes elegantalis*. Theor Appl Climatol. 2018; 132: 135–144.
1020 <https://doi.org/10.1007/s00704-017-2072-2>
- 1021 72. da Silva RS, Kumar L, Shabani F, Vieira A. Dry stress decreases areas suitable for
1022 *Neoleucinodes elegantalis* (Lepidoptera: Crambidae) and affects its survival under climate
1023 predictions in South America. Ecol Inform. 2018; 46: 103–113.
1024 <https://doi.org/10.1016/j.ecoinf.2018.06.003>
- 1025 73. da Silva RS, Kumar L, Shabani F, da Silva EM, da Silva Galdino TV, Picanço MC.
1026 Spatio-temporal dynamic climate model for *Neoleucinodes elegantalis* using CLIMEX.
1027 Int J Biometeorol. 2017; 61: 785–795. <https://doi.org/10.1007/s00484-016-1256-2>
- 1028 74. Díaz AEM, Solis A, Brochero HL. Distribución geográfica de *Neoleucinodes elegantalis*
1029 (Lepidoptera: Crambidae) en Colombia. Rev Colomb Entomol. 2011; 37: 71–76.
- 1030 75. Noboa M, Viera W, Díaz A, Vásquez W, Ron L. Genitalic differentiations in
1031 *Neoleucinodes elegantalis* (Gueneé) (Lepidoptera: Crambidae) associated with Solanaceae
1032 crops in Ecuador. Insects. 2017; 8: 1–11. <https://doi.org/10.3390/insects8030091>

- 1033 76. Eiras AE, Blackmer JL. Eclosion time and larval behavior of the tomato fruit borer,
1034 *Neoleucinodes elegantalis* (Guenée) (Lepidoptera: Crambidae). *Sci Agric.* 2003; 60: 195–
1035 197. <https://doi.org/10.1590/S0103-90162003000100030>
- 1036 77. van Kirk JR, AliNiazee MT. Determining low-temperature threshold for pupal
1037 development of the western cherry fruit fly for use in phenology models. *Environ*
1038 *Entomol.* 1981; 10: 968–971. <https://doi.org/10.1093/ee/10.6.968>
- 1039 78. Brunner J, Hoyt S. Codling moth control - a new tool for timing sprays. *Washingt State*
1040 *Univ Coop Ext Bull.* 1987; 1072.
- 1041 79. Jones VP, Alston DG, Brunner JF, Davis DW, Shelton MD. Phenology of the western
1042 cherry fruit fly (Diptera: Tephritidae) in Utah and Washington. *Ann Entomol Soc Am.*
1043 1991; 84: 488–492. <https://doi.org/10.1093/aesa/84.5.488>
- 1044 80. Zerulla FN, Augel C, Zebitz CPW. Oviposition activity of *Drosophila suzukii* as mediated
1045 by ambient and fruit temperature. *PLoS One.* 2017; 12: e0187682.
1046 <https://doi.org/10.1371/journal.pone.0187682>
- 1047 81. Gramig GG, Burns EE, Prischmann-Voldseth DA. Predicting developmental timing for
1048 immature Canada thistle stem-mining weevils, *Hadroplontus litura* (Coleoptera:
1049 Curculionidae). *Environ Entomol.* 2015; 44: 1085–1094.
1050 <https://doi.org/10.1093/ee/nvv089>
- 1051 82. Herrera AM, Dahlsten DD, Tomic-Carruthers N, Carruthers RI. Estimating temperature-
1052 dependent developmental rates of *Diorhabda elongata* (Coleoptera: Chrysomelidae), a
1053 biological control agent of saltcedar (*Tamarix* spp.). *Environ Entomol.* 2005; 34: 775–784.

- 1054 <https://doi.org/10.1603/0046-225x-34.4.775>
- 1055 83. Menéndez R. How are insects responding to global warming? *Tijdschr voor Entomol.*
1056 2007; 150: 355–365.
- 1057 84. Welch SM, Croft B, Brunner J, Michels M. PETE: an extension phenology modeling
1058 system for management of multi-species pest complex. *Environ Entomol.* 1978; 7: 482–
1059 494. <https://doi.org/10.1093/ee/7.4.487>
- 1060 85. Sporleder M, Juarez H, Simon R. ILCYM-Insect life cycle modeling: software for
1061 developing temperature-based insect phenology models with applications for regional and
1062 global pest risk assessments and mapping. *Proceedings of the 15th Triennial ISTRC*
1063 *Symposium of the International Society for Tropical Root Crops (ISTRC)*. Lima, Peru;
1064 2009. pp. 216–223.
- 1065 86. Sporleder M, Tonnang HEZ, Carhuapoma P, Gonzalez JC, Juarex J, Kroschel J. Insect life
1066 cycle modeling (ILCYM) software - a new tool for regional and global insect pest risk
1067 assessments under current and future climate. In: Peña JE, editor. *Potential Invasive Pests*
1068 *of Agricultural Crops*. CAB eBooks; 2013. p. 464.
1069 <https://doi.org/10.1079/9781845938291.0412>
- 1070 87. Fand BB, Tonnang HEZ, Kumar M, Bal SK, Singh NP, Rao DVKN, et al. Predicting the
1071 impact of climate change on regional and seasonal abundance of the mealybug
1072 *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae) using temperature-driven
1073 phenology model linked to GIS. *Ecol Modell.* 2014; 288: 62–78.
1074 <https://doi.org/10.1016/j.ecolmodel.2014.05.018>

- 1075 88. Pollard CP, Griffin CT, Andrade Moral R de, Duffy C, Chuche J, Gaffney MT, et al.
1076 phenModel: A temperature-dependent phenology/voltinism model for a herbivorous insect
1077 incorporating facultative diapause and budburst. *Ecol Modell.* 2020; 416.
1078 <https://doi.org/10.1016/j.ecolmodel.2019.108910>
- 1079 89. Rebaudo F, Struelens Q, Dangles O. Modelling temperature-dependent development rate
1080 and phenology in arthropods: The devRate package for R. *Methods Ecol Evol.* 2018; 9:
1081 1144–1150.
- 1082 90. Nietschke BS, Magarey RD, Borchert DM, Calvin DD, Jones E. A developmental
1083 database to support insect phenology models. *Crop Prot.* 2007; 26: 1444–1448.
1084 <https://doi.org/10.1016/j.cropro.2006.12.006>
- 1085 91. Moore JL, Remais J V. Developmental models for estimating ecological responses to
1086 environmental variability: structural, parametric, and experimental issues. *Acta Biotheor.*
1087 2014; 62: 69–90. PMID: 24443079
- 1088 92. Logan JA. In defense of big ugly models. *Am Entomol.* 1994; 40: 202–207.
1089 <https://doi.org/10.1093/ae/40.4.202>
- 1090 93. Srivastava V, Lafond V, Griess VC. Species distribution models (SDM): Applications,
1091 benefits and challenges in invasive species management. *CAB Rev Perspect Agric Vet Sci*
1092 *Nutr Nat Resour.* 2019; 14. <https://doi.org/10.1079/PAVSNNR201914020>
- 1093 94. da Silva RS, Kumar L, Shabani F, Picanço MC. Potential risk levels of invasive
1094 *Neoleucinodes elegantalis* (small tomato borer) in areas optimal for open-field *Solanum*
1095 *lycopersicum* (tomato) cultivation in the present and under predicted climate change. *Pest*

- 1096 Manag Sci. 2017; 73: 616–627. <https://doi.org/10.1002/ps.4344>
- 1097 95. Danthanarayana W. Population ecology of the light brown apple moth, *Epiphyas*
1098 *postvittana* (Lepidoptera: Tortricidae). J Anim Ecol. 1983; 52: 1–33.
1099 <https://doi.org/10.2307/4585>
- 1100 96. Grevstad FS, Coop LB. The consequences of photoperiodism for organisms in new
1101 climates. Ecol Appl. 2015; 25: 1506–1517.
- 1102 97. Barbet-Massin M, Rome Q, Villemant C, Courchamp F. Can species distribution models
1103 really predict the expansion of invasive species? PLoS One. 2018; 13: 1–14.
1104 <https://doi.org/10.1371/journal.pone.0193085>
- 1105 98. Isard SA, Russo JM, Magarey RD, Golod J, Vankirk JR. Integrated Pest Information
1106 Platform for Extension and Education (iPiPE): progress through sharing. J Integr Pest
1107 Manag. 2015; 6: 1–8. <https://doi.org/10.1093/jipm/pmv013>
- 1108 99. Moran E V., Alexander JM. Evolutionary responses to global change: Lessons from
1109 invasive species. Ecol Lett. 2014; 17: 637–649. <https://doi.org/10.1111/ele.12262>
- 1110 100. Sinclair BJ, Williams CM, Terblanche JS. Variation in thermal performance among insect
1111 populations. Physiol Biochem Zool. 2012; 85: 594–606. <https://doi.org/10.1086/665388>
- 1112 101. Gray DR, Keena MA. A phenology model for Asian gypsy moth egg hatch. Environ
1113 Entomol. 2019; 48: 903–910. <https://doi.org/10.1093/ee/nvz051>
- 1114

1115

Tables and Figures

1116

1117 **Table 1. Species-specific parameters used in DDRP with corresponding values for *Epiphyas postvittana* (light brown apple**
 1118 **moth) and *Neoleucinodes elegantalis* (small tomato borer).** For both species, the phenological events for egg, larvae, and adults are
 1119 beginning of egg hatch, mid-larval development, and first egg-laying, respectively. The phenological event for pupae is first adult
 1120 emergence for *E. postvittana* and mid-pupal development for *N. elegantalis*. The duration and timing of the phenological event for the
 1121 overwintering stage will vary according to the number of cohorts applied in model runs (see text for details).

1122

Parameter	Code	<i>E. postvittana</i>	<i>N. elegantalis</i>
Lower developmental thresholds (°C)			
Egg	eggLDT	7.2	8.89
Larvae	larvaeLDT	7.2	8.89
Pupae	pupaeLDT	7.2	8.89
Adult	adultLDT	7.2	8.89
Upper developmental thresholds (°C)			
Egg	eggUDT	31.1	30

Table 1 cont.

Larvae	larvaeUDT	31.1	30
Pupae	pupaeUDT	31.1	30
Adult	adultUDT	31.1	30
Stage durations (°C degree-days)			
Egg	eggDD	127	86
Larvae	larvaeDD	408	283
Pupae	pupDD	128	203
Adult	adultDD	71	96
Overwintering larvae	OWlarvaeDD	varies	–
Overwintering adult	OWadultDD	–	varies
Phenological events (°C degree-days)			
Overwintering stage event	OWEventDD	varies	varies
Egg event	eggEventDD	126	80
Larvae event	larvaeEventDD	203	140
Pupae event	pupaeEventDD	128	100

Table 1 cont.

Adult event	adultEventDD	22	55
Cold stress			
Cold stress temperature threshold (°C)	coldstress_threshold	3	-1
Cold degree-day (°C) limit when most individuals die	coldstress_units_max1	875	300
Cold degree-day (°C) limit when all individuals die	coldstress_units_max2	1125	700
Heat stress			
Heat stress temperature threshold (°C)	heatstress_threshold	31	32
Heat stress degree-day (°C) limit when most individuals die	heatstress_units_max1	160	180
Heat stress degree-day (°C) limit when all individuals die	heatstress_units_max2	220	340
Cohorts			
Degree-days (°C) to emergence (average)	distro_mean	210	50
Degree-days (°C) to emergence (variation)	distro_var	2500	1500
Minimum degree-days (°C) to emergence	xdist1	100	0
Maximum degree-days (°C) to emergence	xdist2	320	111
Shape of the distribution	distro_shape	normal	normal

1123 **Table 2. Comparison of the characteristics, parameters, and products of climatic suitability models in CLIMEX and DDRP.**

1124 For simplicity, we do not show CLIMEX parameters related to interaction stress indices (hot-wet stress, hot-dry stress, cold-wet stress,
 1125 and cold-dry stress) or to radiation, substrate, light and diapause indices.

1126

Attributes	DDRP	CLIMEX
Temporal range (time frame)	Any – historical, real-time, near forecast, and climate change forecasts	Historical (1961–1990), and future climate change forecasts for 2030, 2050, 2070, 2080, 2090, and 2100
Temporal scale (time step)	1-day (daily) for PRISM data – others potentially accommodated	Typically weekly values interpolated from monthly data
Spatial scale	Any – default is 2' (~4 km) for PRISM data	CliMond data at a 30' (~55 km at equator) or 10' (~20 km) resolution; others potentially accommodated
Factors influencing climatic suitability	Cold and heat stress only	Cold, heat, dry, and wet stress plus population growth

Table 2 cont.

Modeling process overview	Estimates daily cold and heat stress accumulation and determines whether total accumulations exceed the moderate (max1) or severe (max2) cold and heat stress limits	Estimates weekly population growth and the accumulation of stress (cold, heat, dry, and wet); population growth is reduced when daily accumulations are too low or too high to maintain metabolism
Climate stress parameters		
Temperature thresholds	Upper and lower cold and heat stress temperature thresholds, beyond which stress accumulates as cold and heat stress units	Upper and lower cold and heat stress thresholds in Celsius or degree-day units
Temperature stress rates	Cold and heat stress accumulation limits (max1 and max2); stress units accumulate linearly over time (consecutive days not weighted higher than non-consecutive weeks)	Type 1 – weekly cold and heat stress accumulation rate (similar to DDRP) Type 2 – stress accumulates exponentially in cases of consecutive weeks of stress

Table 2 cont.

Moisture stress thresholds	None	Upper and lower dry and wet stress thresholds
Moisture stress rates	None	Weekly dry and wet stress accumulation rate for species
Total no. of parameters possible	6	38
Total no. of parameters typically used	6	21
Depiction of climatic suitability	Areas not under moderate or severe cold and heat stress exclusions	Typically areas with a high ecoclimatic index (> 20 or 30), which is calculated using annual growth and stress indices
Products	Gridded and summary map outputs of 1) cold and heat stress unit accumulation, and 2) cold, heat, and all (cold plus heat) stress exclusions	Tabular and summary map outputs of 1) cold, heat, dry, and wet stress unit accumulation, and 2) the temperature, moisture, growth, and ecoclimatic index

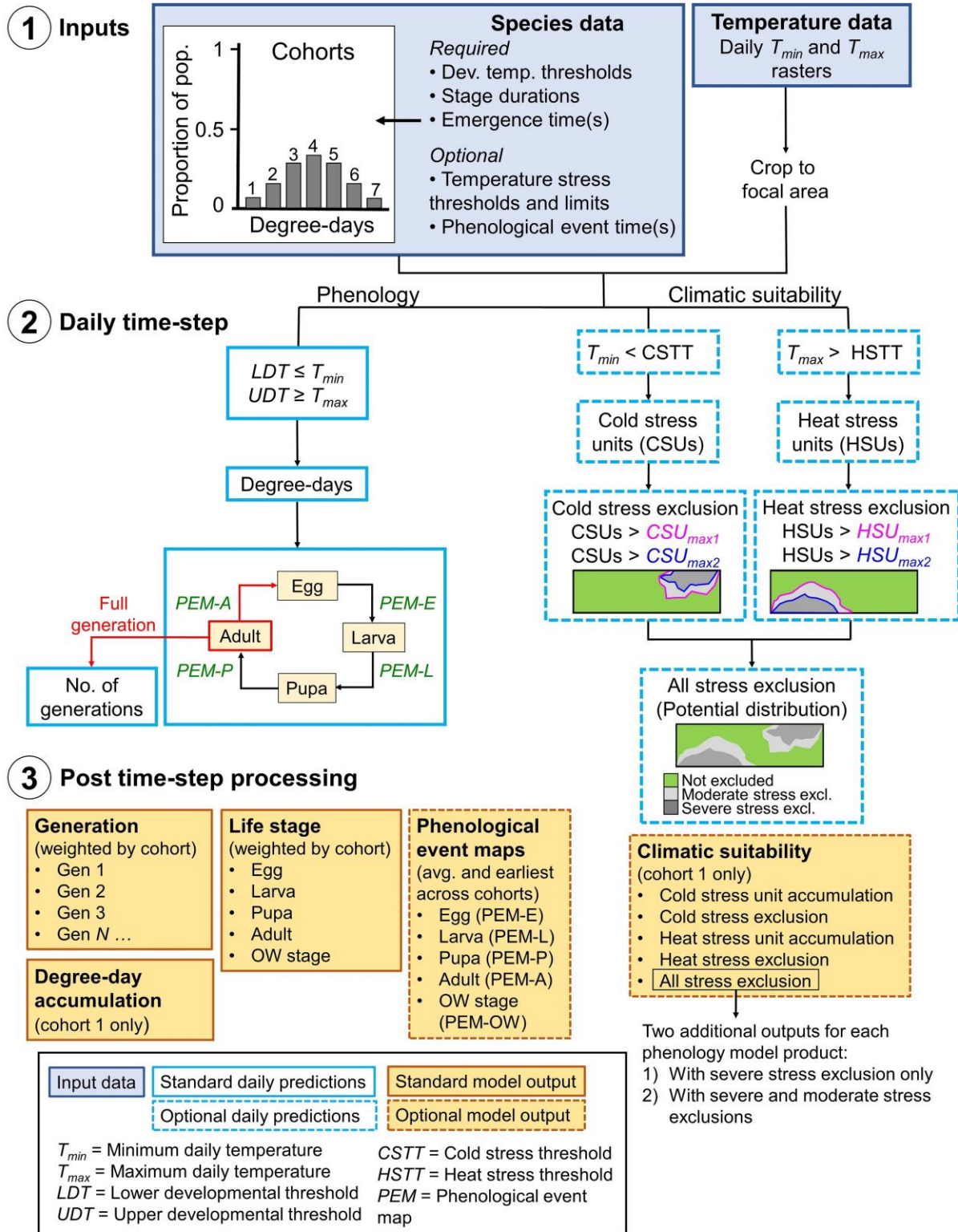
1127 **Table 3. Parameter values used to produce a CLIMEX model for *Epiphyas postvittana* (light brown apple moth) and**
 1128 ***Neoleucinodes elegantalis* (small tomato borer).**
 1129

CLIMEX parameter	Code	<i>E. postvittana</i>			<i>N. elegantalis</i>	
		Lozier & Mills (2011)	He et al. (2012)	This study	da Silva et al. (2018)	This study
Temperature						
Lower temperature threshold (°C)	DV0	7.5	7	7	8.8	8.8
Lower optimal temperature (°C)	DV1	15	13	13	15	15
Upper optimal temperature (°C)	DV2	25	23	23	27	27
Upper temperature threshold (°C)	DV3	31	30	31	30	30
Degree-days per generation (°C days)	PDD	673.6	673.6	673.6	588.2	588.2
Moisture						
Lower soil moisture threshold	SM0	0.15	0.25	0.15	0.35	0.35
Lower optimal soil moisture	SM1	0.5	0.8	0.5	0.7	0.7
Upper optimal soil moisture	SM2	0.8	1.5	0.8	1.5	1.5

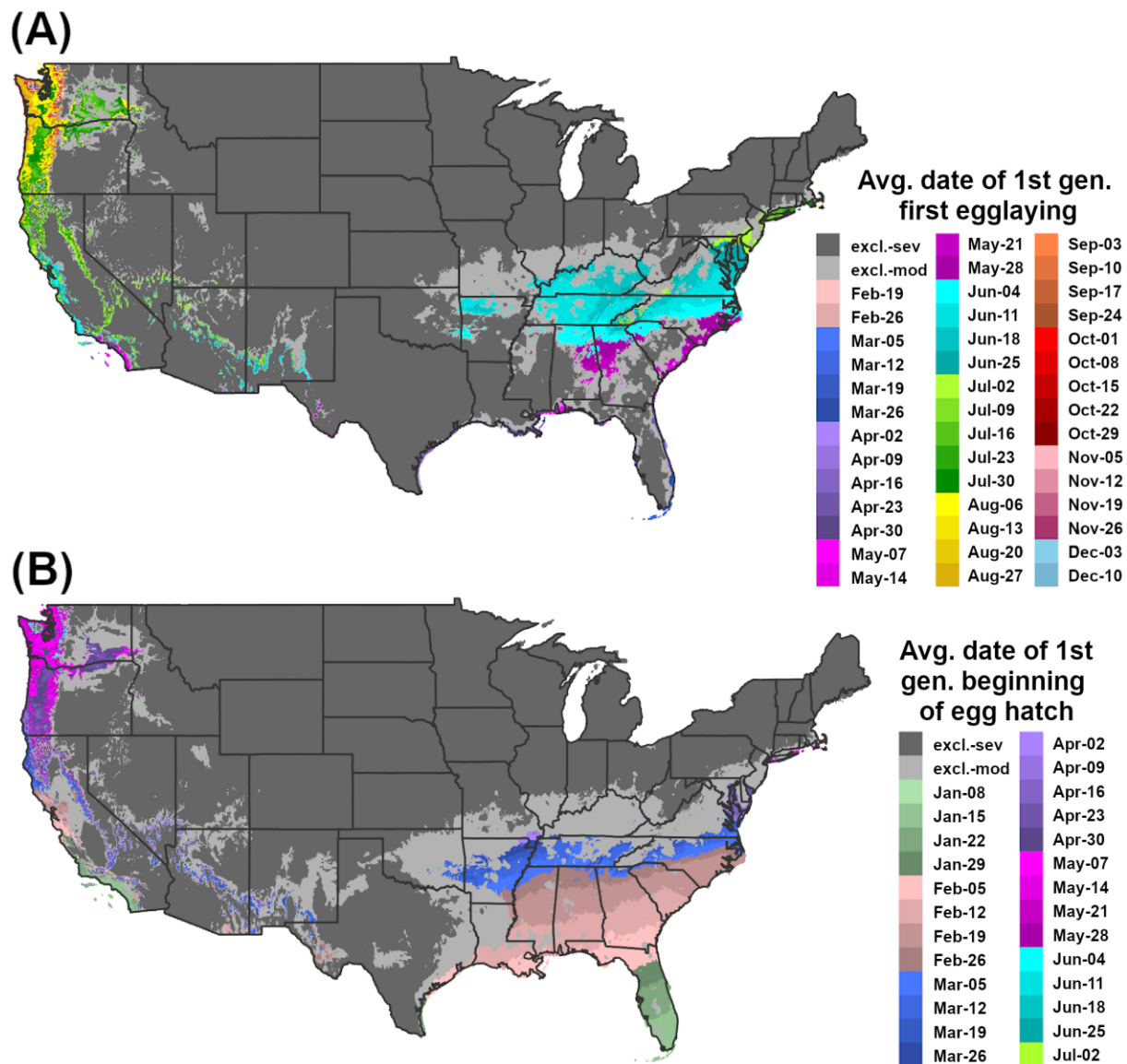
Table 3 cont.

Upper soil moisture threshold	SM3	1.4	2.5	1.4	2.5	2.5
Cold stress						
Cold stress temperature threshold (°C)	TTCS	1.5	5	3	–	2
Cold stress temperature rate (week ⁻¹)	THCS	0.005	0.0005	0.0005	–	0.001
Cold stress degree-day threshold (°C days)	DTCS	–	–	–	15	–
Cold stress degree-day rate (week ⁻¹)	DHCS	–	–	–	0.001	–
Heat stress						
Heat stress temperature threshold (°C)	TTHS	31	31	31	30	30
Heat stress temperature rate (week ⁻¹)	THHS	0.0045	0.01	0.0045	0.0007	0.0007
Dry stress						
Dry stress threshold	SMDS	0.15	0.2	0.15	0.35	0.35
Dry stress rate (week ⁻¹)	HDS	0.005	0.01	0.005	0.001	0.001
Wet stress						
Wet stress threshold	SMWS	1.4	2.5	1.4	2.5	2.5
Wet stress rate (week ⁻¹)	HWS	0.001	0.002	0.001	0.002	0.002

1130 **Fig 1. Schematic of the DDRP model framework.** 1) Input datasets (blue shaded boxes)
1131 include a) data on the developmental requirements, climatic tolerances (optional), and emergence
1132 time(s) of population cohort(s) of a species (Table 1), and b) daily minimum and maximum
1133 temperature data (T_{min} and T_{max} , respectively). 2) Hollow blue boxes indicate calculations
1134 conducted on each daily time step, where a dashed outline represents calculations for climatic
1135 suitability. Phenological event map (PEM) calculations for each life stage (E = egg, L = larva, P
1136 = pupa, A = adult) are shown in green font. A full generation is counted when adults lay eggs (in
1137 red), and the number of generations subsequently increases. 3) After the daily time step
1138 completes, DDRP combines the results across all cohorts and exports the model outputs as multi-
1139 layer raster and summary map (“.png”) files (orange shaded boxes). Orange shaded boxes with a
1140 dashed line represent model outputs for PEMs and climatic suitability.

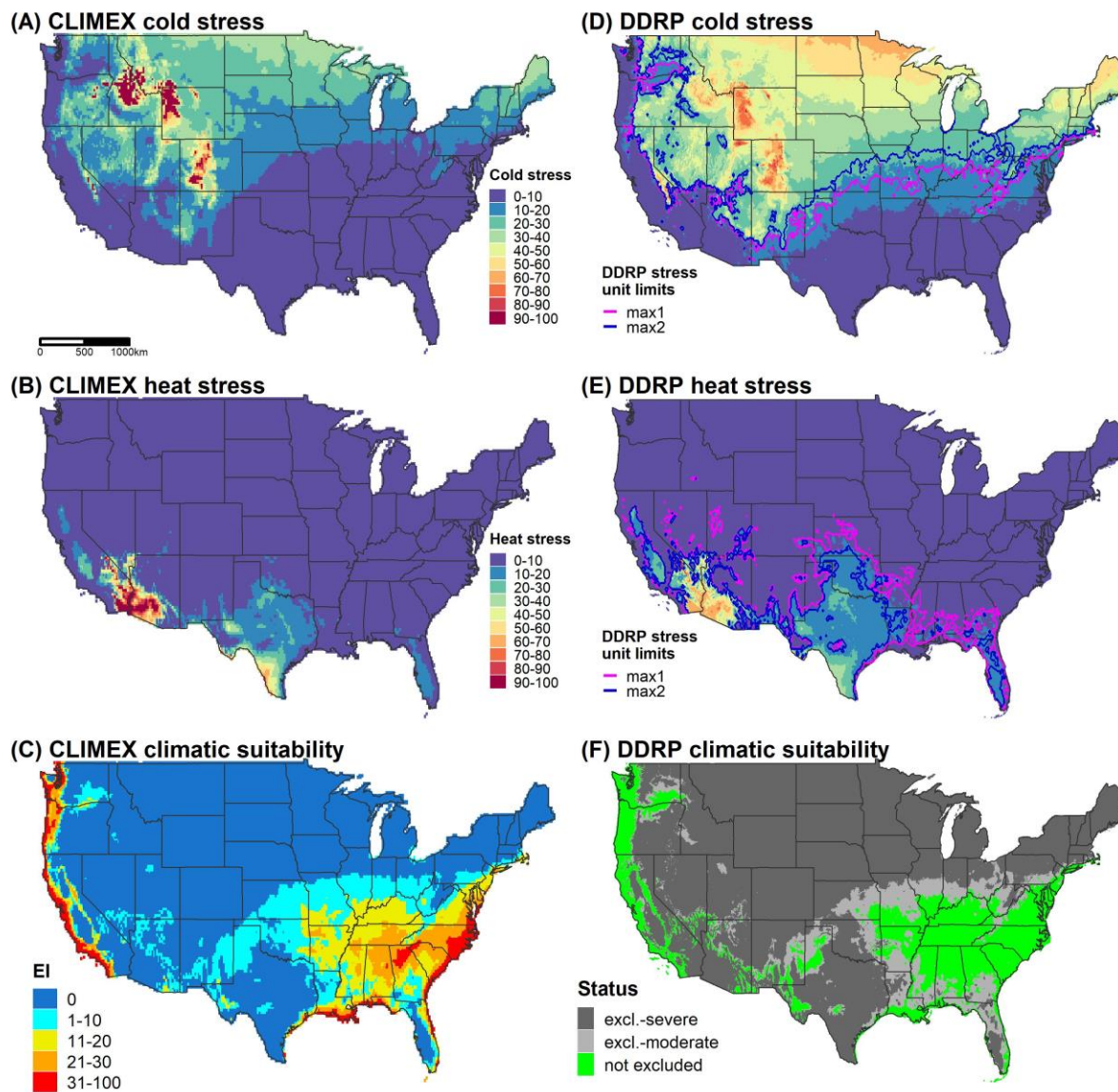


1142 **Fig 2. Phenological event maps generated by DDRP for (A) *Epiphyas postvittana* (light**
1143 **brown apple moth) and (B) *Neoleucinodes elegantalis* (small tomato borer) in CONUS in**
1144 **2018.** The map for *E. postvittana* shows the average date of egg-laying by first generation
1145 females, whereas the map for *N. elegantalis* shows the average date of first generation beginning
1146 of egg hatch. Both maps include estimates of climatic suitability, where long-term establishment
1147 is indicated by areas not under moderate (excl.-moderate) or severe (excl.-severe) climate stress
1148 exclusion.



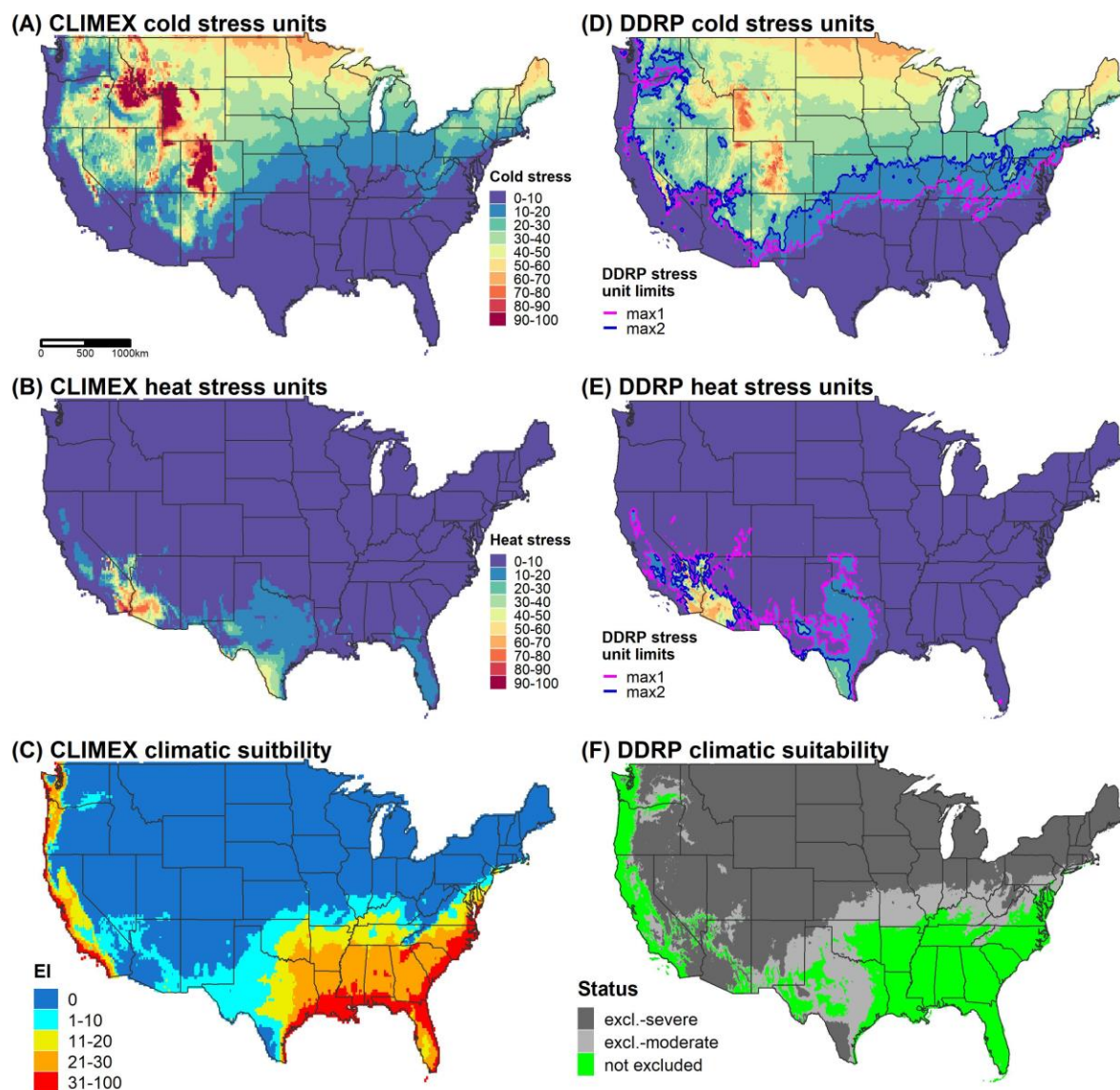
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1150 **Fig 3. Predictions of cold stress, heat stress, and climatic suitability for *Epiphyas postvittana***
1151 **(light brown apple moth) in CONUS produced by CLIMEX (A–C) and DDRP (D–F) based**
1152 **on 1961–1990 climate normals. Climatic suitability is estimated by the Ecoclimatic Index (EI)**
1153 **in CLIMEX, and by combining cold and heat stress exclusions in DDRP. In DDRP, long-term**
1154 **establishment is indicated by areas not under moderate (excl.-moderate) or severe (excl.-severe)**
1155 **climate stress exclusion. Cold and heat stress units in DDRP were scaled from 0 to 100 to match**
1156 **the scale in CLIMEX.**



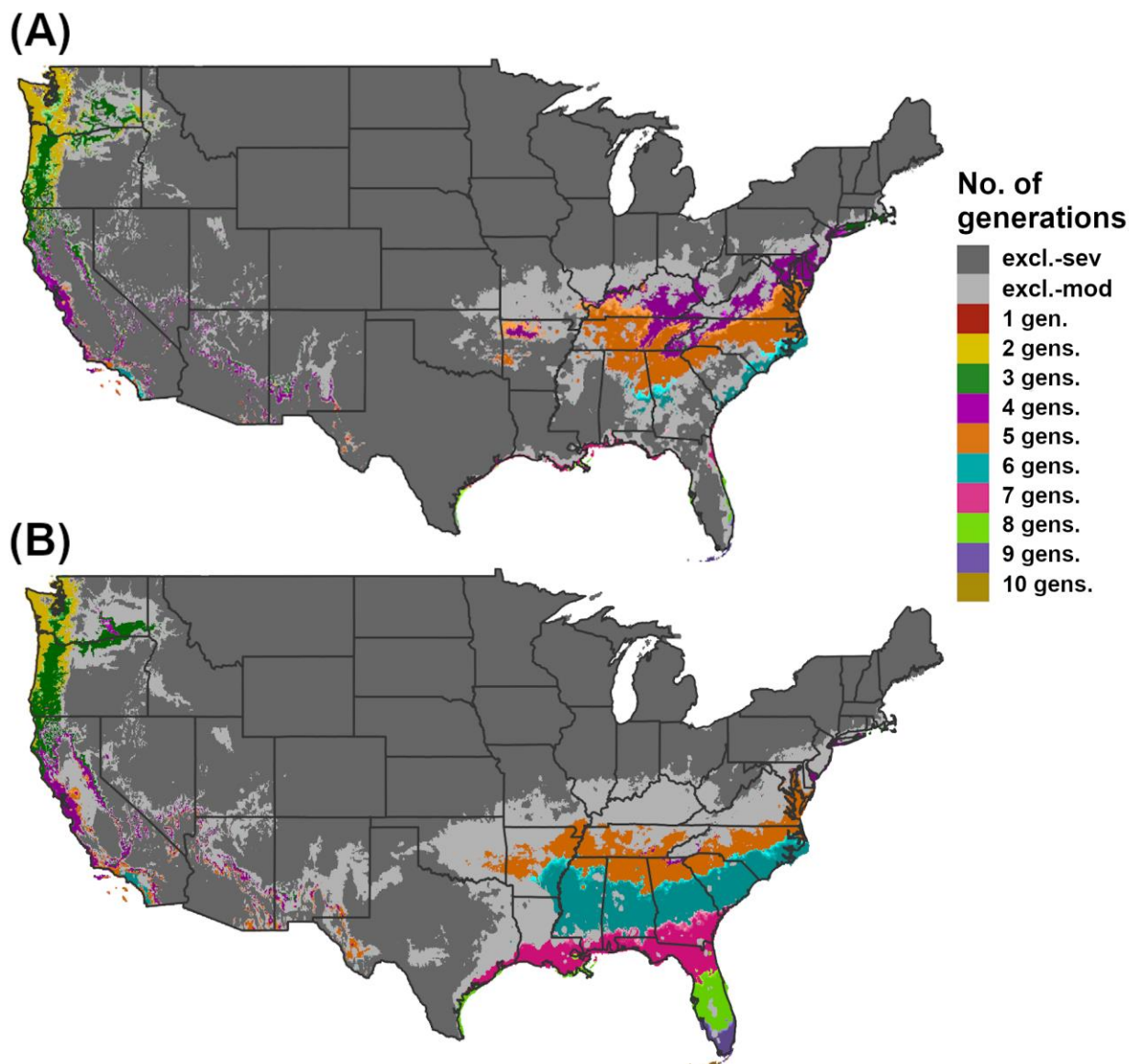
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1158 **Fig 4. Predictions of cold stress, heat stress, and climatic suitability for *Neoleucinodes***
1159 ***elegantalis* (small tomato borer) in CONUS produced by CLIMEX (A–C) and DDRP (D–F)**
1160 **based on 1961–1990 climate normals.** Climatic suitability is estimated by the Ecoclimatic
1161 Index (EI) in CLIMEX, and by combining cold and heat stress exclusions in DDRP. In DDRP,
1162 long-term establishment is indicated by areas not under moderate (excl.-moderate) or severe
1163 (excl.-severe) climate stress exclusion. Cold and heat stress units in DDRP were scaled from 0 to
1164 100 to match the scale in CLIMEX.



1165

1166 **Fig 5. DDRP model predictions of voltinism (number of generations per year) in (A)**
1167 ***Epiphyas postvittana* (light brown apple moth) and (B) *Neoleucinodes elegantalis* (small**
1168 **tomato borer) in CONUS for 2018. Maps include estimates of climatic suitability, where long-**
1169 **term establishment is indicated by areas not under moderate (excl.-moderate) or severe (excl.-**
1170 **severe) climate stress exclusion.**



1171