Plant Co-expression Annotation Resource: a webserver for identifying targets for genetically modified crop breeding pipelines

Marcos José Andrade Viana[1,3], Adhemar Zerlotini[2], Mauricio de Alvarenga Mudadu[2,*]

[1] Postgraduate Program in Bioinformatics, ICB, UFMG, Belo Horizonte, Minas Gerais, Postcode 486, Brazil

[2] Development and Research Department, Embrapa Agricultural Informatics , Campinas, São Paulo, Postcode 6041, Brazil

[3] Information Technology Department, Embrapa Maize and Sorghum, Sete Lagoas, Minas Gerais, Postcode 285, Brazil

* To whom correspondence should be addressed. Tel: +55 19 32115807; Fax: +55 19 3211-5754; Email:mauricio.mudadu@embrapa.br

Keywords: proteins of unknown function, annotation, abiotic stress, database

ABSTRACT

Background

The development of genetically modified crops (GM) includes the discovery of candidate genes through bioinformatics analysis using genomics data, gene expression, and others. Proteins of unknown function (PUFs) are interesting targets for GM crops breeding pipelines for the novelty associated to such targets and also to avoid copyright protections. One method of inferring the putative function of PUFs is by relating them to factors of interest such as abiotic stresses using orthology and co-expression networks, in a guilt-by-association manner.

Results

In this regard, we have downloaded, analyzed, and processed genomics data of 53 angiosperms, totaling 1,862,010 genes and 2,332,974 RNA. Diamond and InterproScan were used to discover 72,266 PUFs for all organisms. RNA-seq datasets related to abiotic stresses were downloaded from

NCBI/GEO. The RNA-seq data was used as input to the LSTrAP software to construct co-expression networks. LSTrAP also created clusters of transcripts with correlated expression, whose members are more probably related to the molecular mechanisms associated to abiotic stresses in the plants. Orthologous groups were created (OrhtoMCL) using all 2,332,974 proteins in order to associate PUFs to abiotic stress related clusters of co-expression and therefore infer their function in a guilt-by-association manner.

Conclusion

A freely available web resource named "Plant Co-expression Annotation Resource" (https://www.machado.cnptia.embrapa.br/plantannot), *Plantannot*, was created to provide indexed queries to search for PUF putatively associated to abiotic stresses. The web interface also allows browsing, querying and retrieving of public genomics data from 53 plants. We hope *Plantannot* to be useful for researchers trying to obtain novel GM crops resistant to climate change hazards.

BACKGROUND

In the last decades, the ability to genetically engineer plants with success showed the potential to create genetically modified (GM) crops with favourable economic outcomes [1]. As well, in the last decades, the main achievements in this area were genetically improved plants tolerant to herbicide and resistant to insects. Others, like nutritional composition improvements are ongoing [2]. Furthermore, new mechanisms for genome editing are improving the accuracy and speed of genome modifications in plants, such as the CRISPR/CAS system [3,4].

Regarding climate change and environmental factors, plants are being genetically modified to become resilient to abiotic stresses, such as drought, high temperature, rising atmospheric CO_2, to potentially overcome the yield losses due to these factors [5,6].

Intellectual property rights (IPR) are vastly used by biotechnology enterprises for their GM plants, to allow exclusive rights and provide better returns for the high investments in research and development [7]. In this way, over the last years many patents applications for genetically improved crops regarding stress tolerance were filled [8].

The first phase for creating GM crops is the candidate gene discovery, which relies on bioinformatics analyses that uses huge volumes of genomics data available on public resources [9,10]. To avoid intellectual property rights over already patented genes, its molecular mechanisms and products, it might be desirable to start researching genes and proteins with no function yet described. These proteins of unknown function (PUF) are very prevalent in eukaryotic genomes and may play roles in determining differences between species [11] and also may be related to resistance to abiotic stresses [12].

Resistance to abiotic stresses is a complex and multigenic trait. Tools and analyses such as QTL, GWAS, gene expression and regulatory networks can be used to find the genes and molecular mechanisms that may play a role in these conditions [13–15] with some results already available [6,16,17].

It is known that differences in the pattern of gene expression, allied to environmental influences, lead to differences in the morphology and phenotype of animals and plants [18]. It is also well established that organs and tissues with the same evolutionary origin have correlated gene expression patterns [19]. To perform molecular comparisons between different species, the focus are genes with the same evolutionary origin, and therefore, with homolog functions, i.e. orthologs [20]. One approach for studying the regulatory functions of a network of genes over different species is to align the co-expression networks using using ortholog genes [21].

In this work we present a web resource named “Plant co-expression annotation resource” (https://www.machado.cnptia.embrapa.br/plantannot) which uses plant genomics and RNA sequencing data, orthology and co-expression networks that allows the selection of PUFs as abiotic stress related candidates to enter GM crop breeding pipelines.

METHODS

Raw Data

Genome data (sequence assembly in FASTA formatted files and annotationin GFF files) for 53 angiosperms (Table 1), including *Glycine max* (Gma), *Zea mays* (Zma), *Arabidopsis thaliana* (Ath) and

Oryza sativa (Osa), were obtained from Phytozome v12 [22] and one from NCBI (*Boea hygrometrica*). The total number of genes and mRNA stored was 1,862,010 and 2,332,974, respectively, together with their translated proteins.

Table 1: Organisms, genome versions and PUF Quantification.

Organism	Genome version	PUF Quantification					
		Protocol A	Protocol B	Protocol C	Protocol D	Protocol E	Protocol F
Amaranthus hypochondriacus	v1.0	873	3	3	4	0	2
Amborella trichopoda	v1.0	52	0	4	0	0	3
Ananas comosus	v3	1790	0	7	4	0	3
Aquilegia coerulea	v3.1	2214	10	38	2	0	25
Arabidopsis halleri	v1.1	362	0	13	7	0	8
Arabidopsis lyrata	v2.1	609	0	4	4	0	3
Arabidopsis thaliana	TAIR10	322	0	150	17	0	128
Boea hygrometrica	GCA_001598015.1	37	0	2	0	0	0
Boechera stricta	v1.2	557	4	14	18	0	10
Brachypodium distachyon	v3.1	2018	2	73	6	0	49
Brachypodium stacei	v1.1	1060	1	41	2	1	33
Brassica oleracea capitata	V1.0	390	0	11	2	0	0
Brassica rapa	FPsc	565	1	21	7	0	13
Capsella grandiflora	v1.1	202	0	14	9	0	9
Capsella rubella	v1.0	2	0	10	0	0	10
Carica papaya	ASGPBv0.4	3333	0	0	5	0	0
Citrus clementenina	v1.0	7	0	24	0	0	20
Citrus sinensis	v1.1	5	0	27	1	0	23
Cucumis sativus	v1.0	995	0	20	5	0	18
Daucus carota	v2.0	8	0	0	0	0	0
Eucalyptus grandis	v2.0	56	0	23	0	0	21
Eutrema salsugineum	v1.0	3	0	8	0	0	8
Fragaria vesca	v1.1	3142	20	1	2	0	0
Glycine max	Wm82.a2.v1	20	0	103	5	0	98
Gossypium raimondii	v2.1	18	0	62	0	0	46
Kalanchoe fedtschenkoi	v1.1	1933	14	53	5	1	40
Kalanchoe laxiflora	v1.1	1576	9	99	7	1	71
Linum usitatissimum	v1.0	1542	27	8	7	1	3
Malus domestica	v1.0	5025	5	48	7	0	27
Manihot esculenta	v6.1	20	0	40	0	0	35
Medicago truncatula	Mt4.0v1	229	0	50	0	0	37
Mimulus guttatus	v2.0	715	2	36	9	0	27
Musa acuminata	v1	3759	2	2	11	0	0
Oropetium thomaeum	v1.0	2551	8	7	10	1	4
Oryza sativa	v7_JGI	709	0	17	82	0	17
Panicum hallii	v2.0	22	0	63	2	0	45
Panicum virgatum	v1.1	10211	6	117	31	1	59
Phaseolus vulgaris	v2.1	123	0	36	5	0	35
Populus trichocarpa	v3.0	1466	0	124	8	0	94

Prunus persica	v2.1	16	0	42	2	0	34
Ricinus communis	v0.1	18	0	0	1	0	0
Salix purpurea	v1.0	1539	0	0	10	0	0
Setaria italica	v2.2	1492	1	59	0	1	38
Setaria viridis	v1.1	1896	1	64	1	1	40
Solanum lycopersicum	iTAG2.4	2694	0	1	1	0	0
Solanum tuberosum	v4.03	3353	2265	3303	4	4	887
Sorghum bicolor	v3.1.1	14	0	18	0	0	11
Spirodela polyrhiza	v2	1104	13	17	11	0	8
Theobroma cacao	v1.1	151	4	1448	0	0	25
Trifolium pratense	v2	1630	6	12	8	0	10
Vitis vinifera	Genoscope.12X	123	1	1	0	0	0
Zea mays	284_AGPv3	9674	3	67	1042	1	60
Zostera marina	v2.2	41	1	164	0	0	143
TOTAL	**53**	**72266**	**2409**	**6569**	**1364**	**13**	**2280**

RNA-seq data related to abiotic stresses (heat, drought, dehydration and osmotic stress) were downloaded from NCBI/GEO in a total of 17 different GEO Series, 53 GEO Samples and 60 SRA short read files only for Gma, Zma, Gma and Ath (Table 2). The data was obtained by searching GEO datasets for the given organisms using the keywords “stress” and filtering the study type by "Expression profiling by high throughput sequencing". The raw reads, correspoding to the GEO Samples, were obtained from NCBI/SRA automatically using the sratoolkit v2.9.2 [23].

Table 2. GEO experiments, GEO samples and SRA identifiers used to obtain RNA-seq data.

Organism*	GEO series	GEO samples	SRA	Condition	Tissue	Date
Arabidopsis thaliana	GSE85653	GSM2280286	SRR4033018	Heat stress rep1	leaves	May-30-2018
Arabidopsis thaliana	GSE85653	GSM2280287	SRR4033019	Heat stress rep2	leaves	May-30-2018
Arabidopsis thaliana	GSE85653	GSM2280288	SRR4033020	Heat stress rep3	leaves	May-30-2018
Arabidopsis thaliana	GSE93979	GSM2466002	SRR5196729	WT drought rep1	leaf	Jun-13-2017
Arabidopsis thaliana	GSE93979	GSM2466003	SRR5196730	WT drought rep1	leaf	Jun-13-2017
Arabidopsis thaliana	GSE93420	GSM2453038	SRR5167847	WT_dehydration1	leaf	Apr-11-2017
Arabidopsis thaliana	GSE93420	GSM2453039	SRR5167848	WT_dehydration2	leaf	Apr-11-2017
Arabidopsis thaliana	GSE93420	GSM2453040	SRR5167849	WT_dehydration3	leaf	Apr-11-2017
Arabidopsis thaliana	GSE94015	GSM2467113	SRR5197907	WT RL3h rep1 heat stress (treated at 37C for 3h)	rosette leaves at flower stages 1-9	Mar-15-2017
Arabidopsis thaliana	GSE94015	GSM2467114	SRR5197908	WT RL3h rep2 heat stress (treated at 37C for 3h)	rosette leaves at flower stages 1-9	Mar-15-2017
Arabidopsis thaliana	GSE94015	GSM2467115	SRR5197909	WT RL3h rep3 heat stress (treated at 37C for 3h)	rosette leaves at flower stages 1-9	Mar-15-2017
Arabidopsis thaliana	GSE72806	GSM1872392	SRR2302914	Col h-1R heat stress (44oC for 1h)	leaves	Oct-24-2016
Arabidopsis thaliana	GSE72806	GSM1872393	SRR2302915	Col h-2R heat stress (44oC for 1h)	leaves	Oct-24-2016
Arabidopsis thaliana	GSE72806	GSM1872394	SRR2302916	Col h-3R heat stress (44oC for 1h)	leaves	Oct-24-2016
Arabidopsis thaliana	GSE72806	GSM1872389	SRR2302911	Col s-1R salinity stress	leaves	Oct-24-2016
Arabidopsis thaliana	GSE72806	GSM1872390	SRR2302912	Col s-2R salinity stress	leaves	Oct-24-2016
Arabidopsis thaliana	GSE72806	GSM1872391	SRR2302913	Col s-3R salinity stress	leaves	Oct-24-2016

Oryza sativa	GSE101734	GSM2714235	SRR5856930	Salt	Seedling leaf	Jul-22-2017
Oryza sativa	GSE101734	GSM2714236	SRR5856931	Salt	Seedling leaf	Jul-22-2017
Oryza sativa	GSE101734	GSM2714237	SRR5856932	Salt	Seedling leaf	Jul-22-2017
Oryza sativa	GSE77510	GSM2053502	SRR3140959	Heat stress (45oC) - 12h	leaf	Dec-21-2017
Oryza sativa	GSE78972	GSM2082859	SRR3209771	Long Day Drought_S3	leaf	Mar-01-2017
Oryza sativa	GSE78972	GSM2082860	SRR3209772	Long Day Drought_S4	leaf	Mar-01-2017
Oryza sativa	GSE78972	GSM2082863	SRR3209775	Short Day Drought_S7	leaf	Mar-01-2017
Oryza sativa	GSE78972	GSM2082864	SRR3209776	Short Day Drought_S8	leaf	Mar-01-2017
Oryza sativa	GSE78972	GSM2082866	SRR3209778	Long Day Drought_S10	leaf	Mar-01-2017
Oryza sativa	GSE78972	GSM2082868	SRR3209780	Short Day Drought_S12	leaf	Mar-01-2017
Oryza sativa	GSE80811	GSM2137964	SRR3466960	drought - 1 d	leaves	Feb-14-2017
Oryza sativa	GSE80811	GSM2137964	SRR3466961	drought - 1 d	leaves	Feb-14-2017
Oryza sativa	GSE80811	GSM2137965	SRR3466962	drought - 2 d	leaves	Feb-14-2017
Oryza sativa	GSE80811	GSM2137965	SRR3466963	drought - 2 d	leaves	Feb-14-2017
Oryza sativa	GSE80811	GSM2137966	SRR3466964	drought - 3 d	leaves	Feb-14-2017
Oryza sativa	GSE80811	GSM2137966	SRR3466965	drought - 3 d	leaves	Feb-14-2017
Oryza sativa	GSE95668	GSM2520922	SRR5311340	heat - 35oC - 6h	leaf	Nov-07-2017
Oryza sativa	GSE95668	GSM2520923	SRR5311341	heat - 35oC - 6h	leaf	Nov-07-2017
Zea mays	GSE71723	GSM1843772	SRR2144414	drought	leaf V12	Feb-04-2016
Zea mays	GSE71723	GSM1843780	SRR2144422	drought	leaf V14	Feb-04-2016
Zea mays	GSE71723	GSM1843788	SRR2144430	drought	leaf V16	Feb-04-2016
Zea mays	GSE71723	GSM1843796	SRR2144438	drought	leaf R1	Feb-04-2016
Zea mays	GSE71377	GSM1833214	SRR2129983	drought	leaf	Jan-22-2016
Zea mays	GSE71046	GSM1826061	SRR2106186	wt Salt T7 Rep1	youngest wrapped leaf	Jan-14-2016
Zea mays	GSE71046	GSM1826073	SRR2106198	wt Salt T0 Rep2+Rep3	youngest wrapped leaf	Jan-14-2016
Zea mays	GSE71046	GSM1826077	SRR2106202	wt Salt T7 Rep2+Rep3	youngest wrapped leaf	Jan-14-2016
Glycine max	GSE98958	GSM2628302	SRR5569810	dehydrated	leaf	May-31-2018
Glycine max	GSE98958	GSM2628302	SRR5569811	dehydrated	leaf	May-31-2018
Glycine max	GSE98958	GSM2628303	SRR5569812	dehydrated	leaf	May-31-2018
Glycine max	GSE98958	GSM2628303	SRR5569813	dehydrated	leaf	May-31-2018
Glycine max	GSE69571	GSM1704043	SRR2051086	salt stress	leaves	Jul-11-2017
Glycine max	GSE69571	GSM1704044	SRR2051087	salt stress	leaves	Jul-11-2017
Glycine max	GSE69571	GSM1704045	SRR2051088	salt stress	leaves	Jul-11-2017
Glycine max	GSE69571	GSM1704046	SRR2051089	salt stress	leaves	Jul-11-2017
Glycine max	GSE70310	GSM1723542	SRR2079645	drought (15 days)	leaf r2 stage	Aug-31-2015
Glycine max	GSE70310	GSM1723542	SRR2079646	drought (15 days)	leaf r2 stage	Aug-31-2015
Glycine max	GSE70310	GSM1723542	SRR2079647	drought (15 days)	leaf r2 stage	Aug-31-2015
Glycine max	GSE69469	GSM1701586	SRR2048167	drought (3 days ZT0-8h R1)	leaves v1 stage	Jul-07-2015
Glycine max	GSE69469	GSM1701592	SRR2048173	drought (3 days ZT4-12h R1)	leaves v1 stage	Jul-07-2015
Glycine max	GSE69469	GSM1701598	SRR2048179	drought (3 days ZT8-16h R1)	leaves v1 stage	Jul-07-2015
Glycine max	GSE69469	GSM1701604	SRR2048185	drought (3 days ZT12-20h R1)	leaves v1 stage	Jul-07-2015
Glycine max	GSE69469	GSM1701610	SRR2048191	drought (3 days ZT16-24h R1)	leaves v1 stage	Jul-07-2015
Glycine max	GSE69469	GSM1701616	SRR2048197	drought (3 days ZT20-4h R1)	leaves v1 stage	Jul-07-2015

** Gma (Glycine max), Zma (Zea mays), (Ath) Arabidopsis thaliana and Osa (Oryza sativa)*

Analyses

The RNA-seq data was used as input to the *LSTrAP* v1.3 software [15] to construct co-expression networks. Only leaf tissue expression data was used to obtain the networks, to avoid adding noise to

the data. LSTrAP was also used to create groups of co-expression, that are clusters of transcripts with correlated expression by using the software MCL version 14-137.

To characterize PUFs, *Diamond v0.9.24 [24] was used* to align all proteins against the NCBI's *nr* database (downloaded in January 2018). Diamond BLAST was run with the flag --max-target-seqs 5 and the best hit was selected. *InterproScan* v5.26-65.0 [25] was used to annotate the proteins from the 53 genomes with "Panther" analyses disabled. All other softwares were run using default parameters. Homolog groups were created using *OrhtoMCL* v2.0.9 [26] and the 53 genome's proteins as input, with default options.

Framework interface

The *Machado* software [27] was used to store all data and results, and also provide a web server as interface for fast data browsing.

Filter Protocols

The *Plantannot* software provides several filters and a text search box that allows searching for molecules by its desired annotation features. These filters are needed to obtain PUFs and to try to relate them to abiotic stresses using RNA-seq expression data and co-expression networks. The Filters menu is separated in 8 fields, of those we are going to use only five: "Organism", "Feature type", "Orthology", "Orthologs_coexpression" and "Analyses". The "Feature Type" filter has three molecule types, from those the polypeptide box is the only that is going to be always checked and the others blank. By using the other 4 remaining filters, 6 protocols were created as examples of different ways to selecting PUFs. Protocol A: using lack of both homology and protein domain signatures. Protocol B: using lack of homology, presence of domain signatures - trying to select Domains of Unknown Function (DUF) from PFAM, and the text search "Unknown function". Protocol C: using homology, lack of protein domain signatures and the text search "Unknown function". Protocol D-F: same protocols of A-C but using ortholog groups to find homolog proteins with co-expression data related to abiotic stress. The protocols are explained in Table 3.

Table 3: Protocols used to characterize PUFs.

Name	Objective	Filters (checked boxes only)*
Protocol A [28]	Find PUFs from organisms whose proteins are not yet in the NCBI's "nr" database and have no protein domain signatures found by InterproScan.	• Analyses: no diamond matches. • Analyses: no interproscan matches.
Protocol B [29]	The same as A but trying to select proteins with the DUF domains from PFAM.	• Analyses: no diamond matches. • Analyses: interproscan matches. • Text search: "Unknown function".
Protocol C [30]	Find PUFs from organisms whose proteins are already public in the "nr" database.	• Analyses: diamond matches. • Analyses: no interproscan matches. • Text search: "Unknown function".
Protocol D [31]	Same as A but using ortholog groups and co-expression networks to relate proteins to abiotic stress	• Analyses: no diamond matches. • Analyses: no interproscan matches. • Orthology: orthology. • Orthologs_coexpression: co-expression.
Protocol E [32]	Same as B but using ortholog groups and co-expression networks to relate proteins to abiotic stress	• Analyses: no diamond matches. • Analyses: interproscan matches. • Text search: "Unknown function". • Orthology: orthology. • ORetryrtholog s_coexpression: co-expression.
Protocol F [33]	Same as C but using ortholog groups and co-expression networks to relate proteins to abiotic stress	• Analyses: diamond matches. • Analyses: no interproscan matches. • Text search: "Unknown function". • Orthology: orthology. • Orthologs_coexpression: co-expression.

** For all protocols "Feature type: polypeptide" is always checked.*

Availability of Supporting Source Code and Requirements

Project name: Plantannot

Project home page: https://www.machado.cnptia.embrapa.br/plantannot

Operating system(s): Platform independent

Programming language: Python 3; Machado (https://github.com/lmb-embrapa/machado)

Other requirements: None.

License: Free to use

bio.tools id: plantannot

RRID: SCR_018429

OVERVIEW

An overview of the component processes of the system covering all data and analysis results used as input to the *Machado* framework, can be seen in Figure 1A and are described below in details.

Homolog groups

The 2,332,974 proteins were used as input to the *OrhtoMCL* software to produce 164,267 clusters, or groups of homolog proteins (putative orthologs). All groups comprise 1,900,313 proteins, and the mean cluster size was 11.57 protein members, ranging from 1 to 4,587 members. It is worth mentioning that 8,535 clusters (5,19%) were left with only 1 protein and 75% of all clusters are composed of up to 6 proteins. The ortholog groups are automatically shown in the “Results” frame of the software.

Co-expression networks

To construct co-expression networks, the 53 GEO Samples (Table 2) were filtered to get expression data only from “leaf” tissue (17, 8, 13 and 15 for Ath, Zma, Gma and Osa respectively). Four co-expression networks were constructed for each of the four organisms (Ath, Zma, Gma and Osa), using the default filters and options of LSTrAP. Groups of co-expression were created using the MCL software following the default instructions in LSTrAP. The MCL software clusters the transcripts with more correlated expression. In this way, the groups of co-expression are supposedly correlated to the molecular mechanisms regarding abiotic stress. 524 groups were obtained (169, 36, 177 and 142 for Ath, Zma, Gma and Osa respectively), with mean size of 140, 113, 282 and 225 for Ath, Zma, Gma e Osa transcript members each, ranging from 1 to 7097 members for Ath, 1 to 4786 for Zma, 1 to 6927 for Gma and 1 to 6636 for Osa.

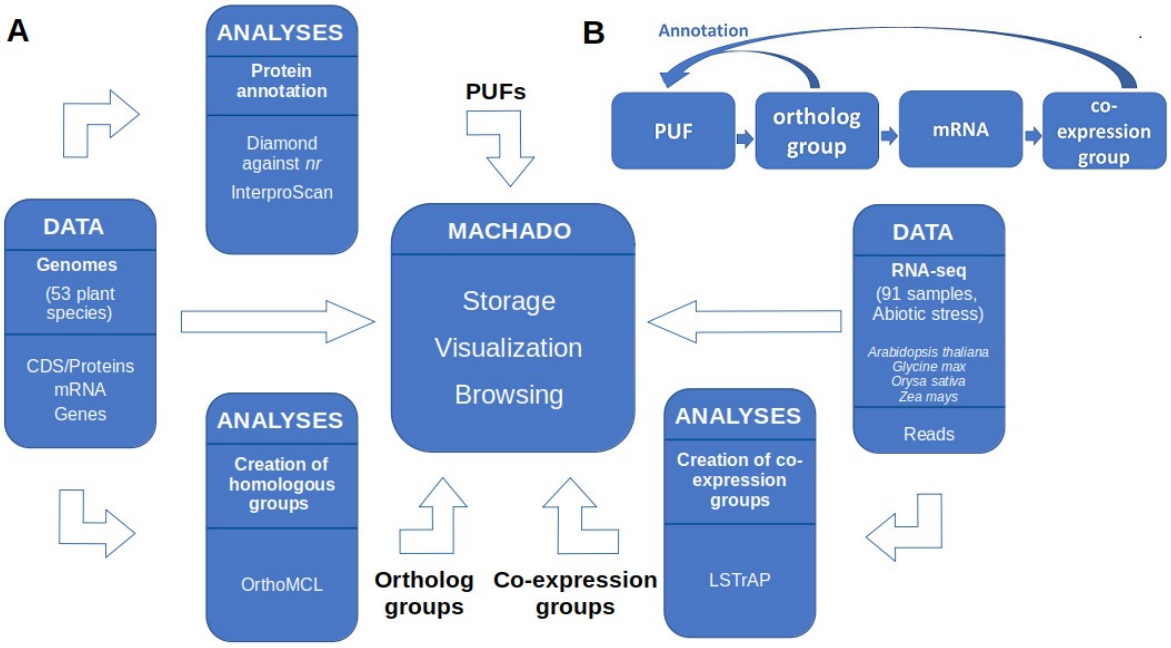

Figure 1. Overview of the Plant Co-expression Annotation Resource processes. B. Guilt-by-association algorithm used to transfer function annotation to PUFs.

PUF characterization

After analyzing all 2,332,974 proteins with *Diamond* and *InterproScan, 72,266* PUFs were characterized (Table 1 – Protocol A) as sequences with no annotation using either *Diamond* or *InterproScan*. Another less sensitive way to find PUFs is to text search for "Unknown proteins" and filter for *InterproScan* matches (e.g.: trying to select PFAM's DUF domains) only or *Diamond* matches only (e.g.: trying to find proteins with uninformative function annotations), which leads to 2,409 and 6,569 PUFs respectively (Table 1 – Protocols B and C respectively).

PUF annotation

As there are no information regarding the function of PUFs, one way to infer function is to link PUFs to other molecules by using orthology groups using a guilt-by-association algorithm (Figure 1B). In this way, members from a given ortholog group which already have annotation and/or have protein domains characterized, can be used as a proxy to infer function for the PUF proteins by association. There are 21,895 PUFs as members of ortholog groups which could be a source of functional

information and annotation (Protocol A, plus adding the filter “Orthology: orthologs”). Furthermore, whenever a given PUF is part of an ortholog group in which some member, necessarily one of Ath, Gma, Osa or Zma, have its mRNA composing a co-expression group, then by association, the initial PUF is supposedly also related to response to abiotic stresses in plants by inference (see Figure 2). 1364 PUFs were related to co-expression groups using filters that were created to automate this selection (Table 3, Protocol D). This method of searching for PUFs was found to be very strict, since it only retrieves proteins that have no annotations whatsoever. However there are many cases in which PUFs have uninformative annotations, such as: “protein with unknown function”, “putative” or “hypothetical” for example. By modifying Protocol D and text searching for “Unknown function” plus filtering for InterproScan matches only or Diamond matches only, we could annotate 13 and 2,280 PUFs respectively (Table 3, Protocols E and F respectively).

Case Study: PUF annotations of desiccation tolerant species

We used two species known to be tolerant to desiccation as a pilot study for *Plantannot* as we believe there can be interesting target PUFs related to abiotic stresses to be encountered in these organisms.

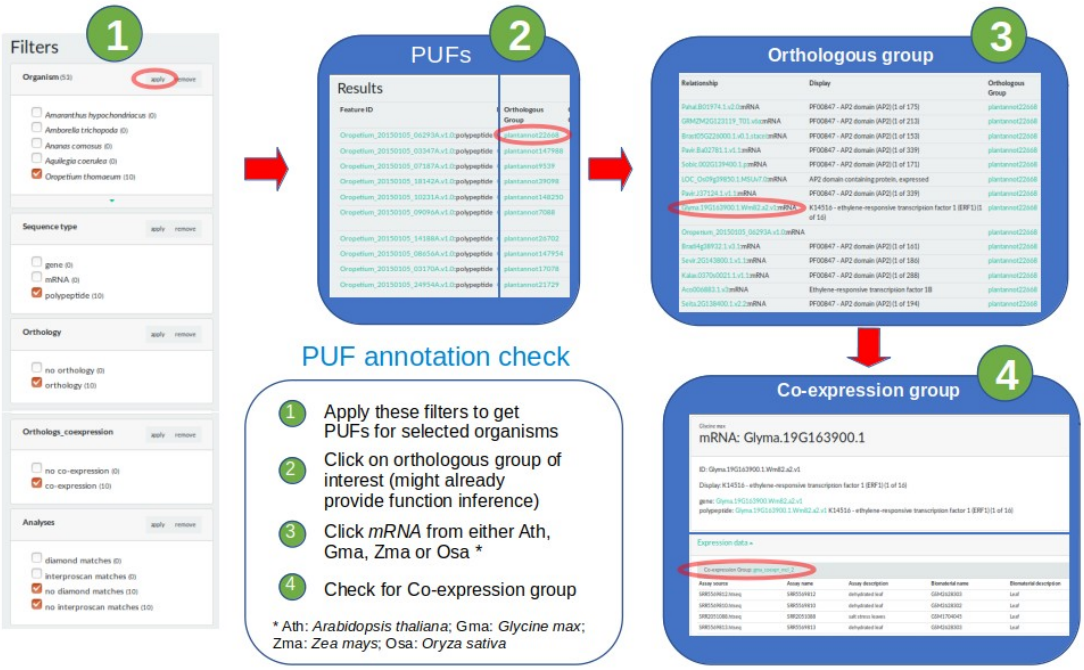

Figure 2: Procotol to check PUF annotation using orthology and co-expression data.

Oropetium thomaeum

Recently added to the Phytozome database, *Oropetium thomaeum [34]* is a good candidate to discover genes related to abiotic stress. This grass is resilient to extreme and prolonged drying and must have genes involved in the molecular mechanisms related to the control of this phenotype. To find PUFs for *Oropetium thomaeum* one could use Protocol D as described in Table 1. By doing this one will see 10 PUFs in the "Results" page. As there is no annotation for these proteins (although there is one protein that was already annotated as "PTHR13020:SF36 – EXPRESSED PROTEIN (1 of 1" that is not much informative of a function), one can survey the homologous sequences present in the orthologous groups to check for other annotations. In this regard, one can click, for example, on the first member of the "Plantannot22668" group ID, in the "Orthologous Group" column of which the PUF "Oropetium_20150105_06293A.v1.0" is a member. By doing this a new "Results" page will show all members of the "Plantannot22668" group (https://www.machado.cnptia.embrapa.br/plantannot/find/?selected_facets=orthologous_group:plantannot22668). Interestingly the majority of the members are annotated as having an "AP2 domain (PFAM – PF00847)". By investigating the function of this PFAM domain PF00847 (https://pfam.xfam.org/family/AP2), one can discover that AP2 is a transcription factor that have a major role in hormone regulation [35] and one study shows that there is a binding factor DBF1 that binds AP2 and is related to osmotic stress tolerance and abiotic stress responses in *Arabidopsis thaliana [36]*. By association, it is possible to infer that the PUF "Oropetium_20150105_06293A.v1.0" have a function possibly related to "AP2", and that orthology could be useful to give novel information for the PUFs. Going further, the "Orthologs_coexpression" box checked before, filtered for orthologous groups of which at least one member participates in a co-expression group. In this way, and also by associative inference, this adds up more evidence that the PUF "Oropetium_20150105_06293A.v1.0" is a good candidate to be related to abiotic stresses and should be further investigated. To check for the co-expression group related to to this PUF, one can follow the procedure in Figure 2 showing that one member of the ortholog group "Plantannot22668" is a protein from Ath, Osa, Zma or Gma, and whose respective mRNA participate in a co-expression group (in this case, the protein from Gma and its mRNA with the same ID: Glyma.19G163900.1.Wm82.a2.v1). This case study can be performed by checking the tutorial session in *Plantannot*'s initial page.

Boea hygrometrica (Dorcoceras hygrometricum)

“Drying without dying” is an essential feature in the evolution of earthly plants and *Boea hygrometrica* is an important model of resurrection plant that survives the drying of its leaves and roots without dying [37] . By using a modified version of Protocol F from Table 3 in which we used the text search word "hypothetical", we recovered 414 PUFs. From these we obtained possible annotations for 199 PUFs (48% of the total) by surveying the orthologous group members as described above. By manually inspecting all 193 annotations we found that 153 (36.95% of the total) had references to abiotic stresses. From these we chose 3 interesting PUFs to describe the possible efficiency of our protocol. The first is the protein KZV45975.1, member of the ortholog group “plantannot11681”, which had members related to “E3 ubiquitin ligase family of proteins”. This family of proteins seems to enhance drought tolerance in *Arabidopsis thaliana* [38]. Another interesting example is the KZV43328.1 protein, member of “plantannot19415” ortholog group, which have 5 members with the PFAM domain “PF00642 - Zinc finger C-x8-C-x5-C-x3-H type (and similar) (zf-CCCH)”. This domain apparently plays roles in abiotic stress response in maize [39]. The final example is the KZV34923.1 protein, who is member of the “plantannot11601” ortholog group which have 17 members that have the PFAM domain “PF05349 - GATA-type transcription activator, N-terminal (GATA-N) (1 of 1)”. It is has been shown that GATA like transcription factors are related to abiotic stress responses in rice [40]. It is worth mentioning that some annotations found refer to abiotic stress that were not part of our RNA-seq data set experimental conditions, like resistance to Aluminum and Cadmium. This could be due to the fact that drought and desiccation tolerance involves a complex process to avoid oxidative damage [41] and we speculate if it may share molecular mechanisms with other kinds of abiotic stresses. The full Boea’s PUF survey can be retrieved from the Supplemental Sheet 1.

DISCUSSION

Many web servers and online tools available allow navigation and comparative search of expression and co-expression data in plants. Some tools only work online and are not open source like PLAZA 3.0 [42], others are more generic and seek any type of annotation such as CoNeKT [43] and many use also microarray data like the Genevestigator [44]. *Plantannot* has a very specific role of surveying for proteins with unknown function possibly related to abiotic stresses in plants and one of its great differentials is the large number of organisms involved (53 angiosperm species). In addition, the algo-

rithm used to search for PUF annotation includes meta analyses and data relations that involve searches for similarities of sequences, orthology and networks of gene co-expression that are specific and unique.

To demonstrate the potentials of *Plantannot* we devised 6 protocols for filtering sequences of interest. From all the 6 protocols, Protocol A was the most permissive, as it seems that most of the organisms have many proteins that do not return as Diamond best hits against the “nr” database. These sequences were selected by the “no diamond matches” filter and could be retrieved (see table 1). By modifying protocol A and inserting the textual search filter “Unknown function”, led to Protocols B and C.

It is important to mention that genome projects end up having proteins of unknown function annotated in several different ways, by using terms like “hypothetical”, “putative”, “unknown protein”, etc. Therefore, there should be specific text searches for each organism to obtain the best results for selecting PUFs. For example, we needed to adapt the filtering protocols for *Boea hygrometrica,* whose PUFs were best retrieved using the text search “hypothetical”. Other examples can be cited, such as the text search "putative protein" used more efficiently to select PUFs from the organism *Ricinus communis.*

Protocol B uses InterproScan results to search for “Domains of Unknown Function”, or DUFs, from PFAM, that are annotations that could result in more PUFs selected. Protocol C uses the text search to filter Diamond hits and also the original sequence annotations to filter out more PUFs.

The Protocols D-F are more complex protocols that refer to modifications of the Protocols A-C, respectively. They were created by adding filters that could retrieve PUFs that were in the same group of homologous proteins, whose mRNA participate in co-expression network clusters, related do abiotic stresses. This guilt-by-association algorithm explained in Figure 2 led to filtering of many interesting PUFs that would not be highlighted using protocols A-C, such as those described in the study case section.

Protocol D is quite stringent and after applying it, 15 organisms out of 53 involved did not show any results. The reason for this result is that many organisms already have their proteins deposited in the “nr” database and the Diamond best hits would retrieve their own sequence leading them to be filtered out. This occurred with *Boea hygrometrica* but did not occur with *Oropetium tomaeum,* both described in our case studies above.

Many other protocols can still be created, for example, modifying Protocols D-F filtering only by groups of orthologs (filter “Orthology: orthology”) and not by co-expression. This filter selected 21,895 PUFs that belonged to any group of orthologs. This simpler filter could allow one to infer possible functions to these PUFs by just relating them to the annotations found in the members of their common groups of orthologs. Similarly, after applying Protocol D for all organisms, we could manually curate the 1364 PUFs selected, supposedly related to abiotic stress. By conducting a manual search in the groups of orthologs that these PUFs belong, we were able to confirm 159 PUFs with functions possibly related to abiotic stress, found in annotations of ortholog co-members of these PUFs. This result equals 11.6% of the initial PUFs (check the Supplemental Sheet 2 for a complete list of PUFs and annotations for all organisms using this methodology).

CONCLUSION

We believe that the Plant Co-expression Annotation Resource can be a valuable bioinformatics tool to be used for the search of proof of concept targets to enter pipelines for the creation of genetic modified crops resistant to abiotic stresses and adapted to climate change.

AVAILABILITY

The Plant Co-expression Annotation Resource is freely available at https://www.machado.cnptia.embrapa.br/plantannot

ACKNOWLEDGMENT

Many thanks for Embrapa’s Multiuser Bioinformatics Laboratory (LMB - Laboratório Multiusuário de Bioinformática da Embrapa), UMiP GenClima and Embrapa Agricultural Informatics (Embrapa Informática Agropecuária) for all the support.

COMPETING INTERESTS

The authors declare that they have no competing interests.

FUNDING

Embrapa 13.16.04.010.00.00 - *Plantannot* - Implementation of a bioinformatics pipeline for gene discovery related to abiotic stresses in plants.

REFERENCES

1. Vincelli P, Jackson-Smith D, Holsapple M, Grusak MA, Harsh M, Klein T, et al. National Academies report has broad support. Nat Biotechnol [Internet]. 2017;35:304–6. Available from: http://www.nature.com/articles/nbt.3842

2. Napier JA, Haslam RP, Tsalavouta M, Sayanova O. The challenges of delivering genetically modified crops with nutritional enhancement traits. Nat Plants [Internet]. 2019;5:563–7. Available from: http://www.nature.com/articles/s41477-019-0430-z

3. Hilscher J, Bürstmayr H, Stoger E. Targeted modification of plant genomes for precision crop breeding. Biotechnol J [Internet]. 2017;12:1600173. Available from: http://doi.wiley.com/10.1002/biot.201600173

4. Zafar SA, Zaidi SS-A, Gaba Y, Singla-Pareek SL, Dhankher OP, Li X, et al. Engineering abiotic stress tolerance via CRISPR/ Cas-mediated genome editing. Foyer C, editor. J Exp Bot [Internet]. 2020;71:470–9. Available from: https://academic.oup.com/jxb/article/71/2/470/5603722

5. Bailey-Serres J, Parker JE, Ainsworth EA, Oldroyd GED, Schroeder JI. Genetic strategies for improving crop yields. Nature [Internet]. 2019;575:109–18. Available from: http://www.nature.com/articles/s41586-019-1679-0

6. Nutan KK, Rathore RS, Tripathi AK, Mishra M, Pareek A, Singla-Pareek SL. Integrating the dynamics of yield traits in rice in response to environmental changes. Om D, editor. J Exp Bot [Internet]. 2020;71:490–506. Available from: https://academic.oup.com/jxb/article/71/2/490/5549718

7. Woźniak E, Waszkowska E, Zimny T, Sowa S, Twardowski T. The Rapeseed Potential in Poland and Germany in the Context of Production, Legislation, and Intellectual Property Rights. Front Plant Sci [Internet]. 2019;10. Available from: https://www.frontiersin.org/article/10.3389/fpls.2019.01423/full

8. Kaur R, Kumar Bhunia R, Kumar Ghosh A. Molecular Genetic Approaches for Environmental Stress Tolerant Crop Plants: Progress and Prospects. Recent Pat Biotechnol [Internet]. 2016;10:12–29. Available from: http://www.eurekaselect.com/openurl/content.php?genre=article&issn=1872-2083&volume=10&issue=1&spage=12

9. Prado JR, Segers G, Voelker T, Carson D, Dobert R, Phillips J, et al. Genetically Engineered Crops: From Idea to Product. Annu Rev Plant Biol [Internet]. 2014;65:769–90. Available from: http://www.annualreviews.org/doi/10.1146/annurev-arplant-050213-040039

10. Scheben A, Edwards D. Bottlenecks for genome-edited crops on the road from lab to farm. Genome Biol [Internet]. 2018;19:178. Available from: https://genomebiology.biomedcentral.com/articles/10.1186/s13059-018-1555-5

11. Gollery M, Harper J, Cushman J, Mittler T, Girke T, Zhu J-K, et al. What makes species unique? The contribution of proteins with obscure features. Genome Biol [Internet]. 2006;7:R57. Available from: http://www.ncbi.nlm.nih.gov/pubmed/16859532

12. Luhua S, Hegie A, Suzuki N, Shulaev E, Luo X, Cenariu D, et al. Linking genes of unknown function with abiotic stress responses by high-throughput phenotype screening. Physiol Plant [Internet]. 2013;148:322–33. Available from: http://doi.wiley.com/10.1111/ppl.12013

13. Nogué F, Mara K, Collonnier C, Casacuberta JM. Genome engineering and plant breeding: impact on trait discovery and development. Plant Cell Rep [Internet]. 2016;35:1475–86. Available from: http://link.springer.com/10.1007/s00299-016-1993-z

14. Nuccio ML, Paul M, Bate NJ, Cohn J, Cutler SR. Where are the drought tolerant crops? An assessment of more than two decades of plant biotechnology effort in crop improvement. Plant Sci [Internet]. 2018;273:110–9. Available from: https://linkinghub.elsevier.com/retrieve/pii/S016894521731213X

15. Proost S, Krawczyk A, Mutwil M. LSTrAP: efficiently combining RNA sequencing data into co-expression networks. BMC Bioinformatics [Internet]. 2017;18:444. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29017446

16. Dahal K, Li X-Q, Tai H, Creelman A, Bizimungu B. Improving Potato Stress Tolerance and Tuber Yield Under a Climate Change Scenario – A Current Overview. Front Plant Sci [Internet]. 2019;10. Available from: https://www.frontiersin.org/article/10.3389/fpls.2019.00563/full

17. Stanford BCM, Rogers SM. R(NA)-tistic expression: The art of matching unknown mRNA and proteins to environmental response in ecological genomics. Mol Ecol [Internet]. 2018;27:827–30. Available from: http://doi.wiley.com/10.1111/mec.14419

18. Roux J, Rosikiewicz M, Robinson-Rechavi M. What to compare and how: Comparative transcriptomics for Evo-Devo. J Exp Zool Part B Mol Dev Evol [Internet]. 2015;324:372–82. Available from: http://doi.wiley.com/10.1002/jez.b.22618

19. Sudmant PH, Alexis MS, Burge CB. Meta-analysis of RNA-seq expression data across species, tissues and studies. Genome Biol [Internet]. 2015;16:287. Available from: https://genomebiology.biomedcentral.com/articles/10.1186/s13059-015-0853-4

20. Sonnhammer ELL, Gabaldon T, Sousa da Silva AW, Martin M, Robinson-Rechavi M, Boeckmann B, et al. Big data and other challenges in the quest for orthologs. Bioinformatics [Internet]. 2014;30:2993–8. Available from: https://academic.oup.com/bioinformatics/article-lookup/doi/10.1093/bioinformatics/btu492

21. Serin EAR, Nijveen H, Hilhorst HWM, Ligterink W. Learning from Co-expression Networks: Possibilities and Challenges. Front Plant Sci [Internet]. 2016;7. Available from: http://journal.frontiersin.org/Article/10.3389/fpls.2016.00444/abstract

22. Goodstein DM, Shu S, Howson R, Neupane R, Hayes RD, Fazo J, et al. Phytozome: a comparative platform for green plant genomics. Nucleic Acids Res [Internet]. 2012;40:D1178–86. Available from: https://academic.oup.com/nar/article-lookup/doi/10.1093/nar/gkr944

23. NCBI. The SRA Toolkit [Internet]. Available from: https://github.com/ncbi/sra-tools

24. Buchfink B, Xie C, Huson DH. Fast and sensitive protein alignment using DIAMOND. Nat Methods [Internet]. 2015;12:59–60. Available from: http://www.nature.com/articles/nmeth.3176

25. Quevillon E, Silventoinen V, Pillai S, Harte N, Mulder N, Apweiler R, et al. InterProScan: protein domains identifier. Nucleic Acids Res [Internet]. 2005;33:W116–20. Available from: https://academic.oup.com/nar/article-lookup/doi/10.1093/nar/gki442

26. Fischer S, Brunk BP, Chen F, Gao X, Harb OS, Iodice JB, et al. Using OrthoMCL to Assign Proteins to OrthoMCL-DB Groups or to Cluster Proteomes Into New Ortholog Groups. Curr Protoc Bioinforma [Internet]. Hoboken, NJ, USA: John Wiley & Sons, Inc.; 2011. Available from: http://doi.wiley.com/10.1002/0471250953.bi0612s35

27. Mudadu M de A, Zerlotini A. Machado: open source genomics data integration framework. bioRxiv [Internet]. 2020;2020.05.08.084731. Available from: http://biorxiv.org/content/early/2020/05/10/2020.05.08.084731.abstract

28. Viana M, Zerlotini A, Mudadu M. Protocol A - Plantannot [Internet]. Available from: https://dx.doi.org/10.17504/protocols.io.bgcvjsw6

29. Viana M, Zerlotini A, Mudadu M. Protocol B - Plantannot [Internet]. Available from: https://dx.doi.org/10.17504/protocols.io.bgdgjs3w

30. Viana M, Zerlotini A, Mudadu M. Protocol C - Plantannot [Internet]. Available from: https://dx.doi.org/10.17504/protocols.io.bgdijs4e

31. Viana M, Zerlotini A, Mudadu M. Protocol D - Plantannot. Available from: https://dx.doi.org/10.17504/protocols.io.bgd6js9e

32. Viana M, Zerlotini A, Mudadu M. Protocol E - Plantannot. Available from: https://dx.doi.org/10.17504/protocols.io.bgdjjs4n

33. Viana M, Zerlotini A, Mudadu M. Protocol F - Plantannot. Available from: https://dx.doi.org/10.17504/protocols.io.bgdkjs4w

34. VanBuren R, Wai CM, Keilwagen J, Pardo J. A chromosome-scale assembly of the model desiccation tolerant grass Oropetium thomaeum. Plant Direct [Internet]. 2018;2:e00096. Available from: http://doi.wiley.com/10.1002/pld3.96

35. Ogawa T, Uchimiya H, Kawai-Yamada M. Mutual Regulation of Arabidopsis thaliana Ethylene-responsive Element Binding Protein and a Plant Floral Homeotic Gene, APETALA2. Ann Bot [Internet]. 2007;99:239–44. Available from: http://academic.oup.com/aob/article/99/2/239/2390906/Mutual-Regulation-of-Arabidopsis-thaliana

36. Saleh A, Lumbreras V, Lopez C, Kizis, Eva Dominguez-Puigjaner D, Pagès M. Maize DBF1-interactor protein 1 containing an R3H domain is a potential regulator of DBF1 activity in stress responses. Plant J [Internet]. 2006;46:747–57. Available from: http://doi.wiley.com/10.1111/j.1365-313X.2006.02742.x

37. Xiao L, Yang G, Zhang L, Yang X, Zhao S, Ji Z, et al. The resurrection genome of Boea hygrometrica : A blueprint for survival of dehydration. Proc Natl Acad Sci [Internet]. 2015;112:5833–7. Available from: http://www.pnas.org/lookup/doi/10.1073/pnas.1505811112

38. Yang L, Wu L, Chang W, Li Z, Miao M, Li Y, et al. Overexpression of the maize E3 ubiquitin ligase gene ZmAIRP4 enhances drought stress tolerance in Arabidopsis. Plant Physiol Biochem [Internet]. 2018;123:34–42. Available from: https://linkinghub.elsevier.com/retrieve/pii/S0981942817303868

39. Peng X, Zhao Y, Cao J, Zhang W, Jiang H, Li X, et al. CCCH-Type Zinc Finger Family in Maize: Genome-Wide Identification, Classification and Expression Profiling under Abscisic Acid and Drought Treatments. Defossez P-A, editor. PLoS One [Internet]. 2012;7:e40120. Available from: https://dx.plos.org/10.1371/journal.pone.0040120

40. Gupta P, Nutan KK, Singla-Pareek SL, Pareek A. Abiotic Stresses Cause Differential Regulation of Alternative Splice Forms of GATA Transcription Factor in Rice. Front Plant Sci [Internet]. 2017;8. Available from: http://journal.frontiersin.org/article/10.3389/fpls.2017.01944/full

41. Pardo J, Man Wai C, Chay H, Madden CF, Hilhorst HWM, Farrant JM, et al. Intertwined signatures of desiccation and drought tolerance in grasses. Proc Natl Acad Sci [Internet]. 2020;117:10079–88. Available from: http://www.pnas.org/lookup/doi/10.1073/pnas.2001928117

42. Vandepoele K. A Guide to the PLAZA 3.0 Plant Comparative Genomic Database. 2017. p. 183–200. Available from: http://link.springer.com/10.1007/978-1-4939-6658-5_10

43. Proost S, Mutwil M. CoNekT: an open-source framework for comparative genomic and transcriptomic network analyses. Nucleic Acids Res [Internet]. 2018;46:W133–40. Available from: https://academic.oup.com/nar/article/46/W1/W133/4990637

44. Hruz T, Laule O, Szabo G, Wessendorp F, Bleuler S, Oertle L, et al. Genevestigator V3: A Reference Expression Database for the Meta-Analysis of Transcriptomes. Adv Bioinformatics [Internet]. 2008;2008:1–5. Available from: http://www.hindawi.com/journals/abi/2008/420747/