Social structure defines spatial transmission of African swine fever in wild boar

Kim M. Pepin[1,*], Andrew Golnar[1], and Tomasz Podgórski[2,3]

1.   National Wildlife Research Center, USDA, APHIS, Wildlife Services, 4101 Laporte

Ave., Fort Collins, CO, 80526.

2.   Mammal Research Institute, Polish Academy of Sciences, Stoczek 1, 17-230 Białowieża,

Poland

3.   Department of Game Management and Wildlife Biology, Faculty of Forestry and Wood

Sciences, Czech University of Life Sciences, Kamýcká 129, 165 00 Praha 6, Czech Republic

*Corresponding author: Kim Pepin, National Wildlife Research Center, United States

Department of Agriculture, 4101 Laporte Ave., Fort Collins, CO, 80526;

kim.m.pepin@aphis.usda.gov

RH: Pepin et al. ˙Spatial spread of ASFv in wild boar

Abstract

1. Spatial spread of infectious disease is determined by spatial and social processes such as animal space use and family group structure. Yet, impacts of social processes on spatial spread remain poorly understood and estimates of spatial transmission kernels (STKs) often exclude social structure. Understanding impacts of social structure on STKs is important for obtaining robust inferences for policy decisions and optimizing response plans.

2. We fit spatially-explicit transmission models with different assumptions about contact structure to African swine fever virus (ASFv) surveillance data from Eastern Poland from 2014-2015 and evaluated how social structure affected inference of STKs and spatial spread.

3. The model with social structure provided better inference of spatial spread, predicted that ~80% of transmission events occurred within family groups, and that transmission was weakly female-biased (other models predicted weakly male-biased transmission). In all models, most transmission events were within 1.5 km, with some rare events at longer distances. Effective reproductive numbers were between 1.1 and 2.5 (maximum values between 4 and 8).

4. Social structure can modify spatial transmission dynamics. Accounting for this additional contact heterogeneity in spatial transmission models could provide more robust inferences of STKs for policy decisions, identify best control targets, and improve transparency in model uncertainty.

KEYWORDS: African swine fever, Effective reproduction number, Spatial transmission kernel, Surveillance, Wild boar, Social structure

INTRODUCTION

Social [1] and spatial processes [2] are key drivers of pathogen transmission, yet their

relative roles and influences on one another remain poorly understood [3, 4]. While both social

structure and animal space use shape contact heterogeneity, they can have fundamentally

different effects on contact structure and thus pathogen transmission dynamics [3-5]. For

example, social organization into family groups (one dimension of social structure) causes local

clustering of hosts that can alter pathogen transmission dynamics in a broad range of wildlife

disease systems, such as malaria in primates, fungal infection in termites, intestinal parasites in

African Artiodactylids, rabies in raccoons, bovine tuberculosis in badgers, and Chronic Wasting

Disease in deer [6-12]. In contrast, animal space use mainly acts on overall connectivity in a

population by limiting how far a disease can travel at each transmission event [13-16]. As such

these processes are commonly modeled using different techniques and thus not accounted for in

the same framework [3]. For example, social structure is often represented using network models

[3, 17], while space use is often represented through distance functions describing how

transmission probability changes with distance between hosts (spatial transmission kernels;

STKs) [18]. STKs are key parameters for planning disease control strategies because they can be

used to inform how surveillance and control strategies should be deployed spatially [2, 18-22].

However, without appropriately accounting for contact heterogeneities due to social structure,

STKs could be uncertain or biased. Thus frameworks that can account for the effects of social

structure on STKs are needed to better understand how different dimensions of contact

heterogeneity shape pathogen transmission dynamics in wildlife populations and to optimize

response plans using STKs.

Individual–based models (IBMs) are useful for incorporating social structure and space

use separately in the same framework [7, 23, 24] to understand the significance of both processes

on pathogen transmission dynamics. In general, IBMs can be especially useful for determining

how much complexity is important to capture pathogen transmission dynamics well enough to

effectively guide control policies [3, 25]. Previous work has used similar IBM structures to

represent social structure and space use concurrently in systems as different as rabies in

raccoons, African swine fever in wild boar, bovine tuberculosis in badgers, and foot-and mouth

disease in feral swine to inform disease management strategies [7, 24, 26, 27] , demonstrating

how IBMs that are designed to account for individual-level variation in social and spatial

parameters are widely applicable while allowing for an understanding of how individual-level

nuances affect pathogen transmission dynamics. This flexibility is particularly well-suited for

understanding how social structure modifies STKs. Secondly, social structure is usually

temporally dynamic, which has been frequently neglected in data-driven social-network models

due to the challenges with estimating changes in network structure over time. An IBM approach

allows for natural incorporation of temporal changes in social network structure due to

demographic changes.

To address gaps with understanding the impact of social structure on the spatial spread of

disease, we developed an IBM fit to weekly surveillance data from wild boar in Poland from

2014-2015. In Eastern Europe, wild boar demonstrate limited spatial movement and cluster into

family groups suggesting that both social structuring and space use are important for inferring

the dynamics of African swine fever virus (ASFv) transmission [26, 28, 29]. Thus, the ASFv

system provides an excellent opportunity to quantify effects of social structure on STKs and the

magnitude to which model uncertainty could affect policy decisions using STKs. ASFv has been

extremely difficult to control in wild boar partly due to a lack of robust predictions of spatial

spread for risk-based mitigation [30]. In countries where ASFv is so widespread in wild boar

such as Poland, robust estimates of STKs are crucial for improving response plans and risk

assessment for domestic pig producers by enabling prediction of how fast and where ASFv will

spread, and thus optimization of resource allocation across the landscape to surveillance and

control. Also, understanding how social structure defines STKs can help to identify control

targets that would minimize disease spread (e.g., the importance of controlling family groups

versus adult males).

Without detailed genetic or contact tracing data to reconstruct transmission history, STKs

are predominantly estimated indirectly by fitting pathogen transmission models to available case

data [2, 18, 31]. Common assumptions of STK estimation methods include a single introduction

event and perfectly observed data [32]. However, STK estimates could be biased if re-

introduction events from outside sources spark epizootic foci in spatially distinct areas from the

initial outbreak (i.e., being mistaken for transmission between clusters and causing artificially fat

tails in the STK). Likewise, STKs could be biased if the surveillance system is biased towards

detection of particular transmission events. Thus, methods that account for re-introduction and

surveillance design are important in cases where 'single introduction' and 'perfect detection'

assumptions are violated. Similar to many wildlife host-pathogen systems [31, 33-35], ASFv

surveillance in wild boar is mostly passive [36] with only a small proportion of cases likely being

detected (e.g., for ASFv in Poland: $\leq 30\%$ of all ASFv carcasses detected by carcass

surveillance, $\leq 1.7\%$ of all active ASFv infections detected by hunter harvest, [26]), and genetic

evidence suggests that at least two re-introductions from neighboring regions occurred during the

time frame of our study [37].

In previous work we fitted models that accounted for the realities of multiple introduction

events and partially observed data to ASFv surveillance data from wild boar in Poland to

estimate the frequency of carcass-based transmission and re-introduction in outbreak dynamics

[26]. This previous work predicted greater than 10 international introductions per year were

necessary to sustain viral persistence at low host densities [26]. Here, we extended this modeling

approach to understand how social structuring and space use assumptions affect estimates of

STKs and spatial pathogen dynamics. We tested three different assumptions about space use and

social structure: 1) neighborhood (local transmission only), 2) exponential decay (distance

distribution that includes long-distance processes), and 3) distance distribution with social

structure (social and spatial processes) (Fig. 1). We fit the different models to ASFv surveillance

data using approximate Bayesian computation (ABC) and compared the model fits using

distance metrics and R^2. We then predicted the STKs under each set of assumptions using the

fitted models and evaluated the effects of social and spatial processes on STKs and a key

epidemiological parameter – the effective reproduction number. Our framework provides a

general approach for understanding the relative role of social and spatial processes in pathogen

transmission dynamics and quantifying STKs in the presence of realistic complexities (i.e., social

structure, multiple re-introductions, partially observed case data).

METHODS

Study system and data

African swine fever virus (ASFv), a virulent virus of swine, emerged in domestic pigs in

Georgia in 2007 following introduction from Africa [38]. After its initial emergence, the virus

spread quickly to Eastern Europe becoming endemic in wild boar, which has challenged

elimination. With no effective vaccine or treatment options, control strategies are focused on

reducing swine movement, decontamination, and culling [39]. In wild boar, transmission occurs

primarily through direct contact or contact with contaminated carcasses, consumption of

contaminated food resources, and other forms of environmental or mechanical transmission are

suspected [40]. Effectiveness of these strategies depends on being able to rapidly find new cases

and target high-risk areas, thus models that can predict spatial spread are crucial tools. The index

case of ASFv in wild boar was detected in February 2014 in north-eastern Poland (53°19'33"N,

23°45'31"E), less than 1 km from the border with Belarus. Subsequent cases occurred close to

the Belarusian border [41, 42]. By the end of 2015, 139 wild boar tested positive for ASFv in the

area, with maximum distance of 27.4km west of the border and a 100km range along the border.

The affected area is dominated by a mosaic of woodlands and agricultural land (crop fields,

pastures, meadows) with several large (several hundred square kilometers), continuous forests.

On average, forest covers 53% of the area and wild boar densities range from 1.5 – 2.5 wild boar

/ km^2 across all of Poland [43]. A total of 2470 samples from hunters and 205 samples from

carcasses were collected from 8 administrative districts where ASFv occurred from Feb. 2014

through Dec. 2015. These samples were submitted to the National Reference Laboratory for

ASFv at the National Veterinary Research Institute in Puławy, Poland for viral diagnostics.

Surveillance data were used to fit the model and define spatio-temporal intensity of sampling for

this analysis. We also used similar data from Jan. 2016 through Jul. 2016 to evaluate out-of-

sample prediction. A detailed description of laboratory procedures and tests can be found in

Woźniakowski et al. (2015) and Śmietanka et al. (2016).

Process model

Overview. Previously we developed a spatially-explicit, individual-based model of ASFv

transmission dynamics in wild boar that we fit to surveillance data using Approximate Bayesian

Computation (ABC). This model accounted for: 1) social structure (independent adult males and

matrilineal family groups that varied seasonally in size and composition due to birth rates, natal

dispersal, and hunter harvest trends), and 2) spatial processes (daily space use and dispersal to

new home range centroids), and was used to infer the role of carcass-based transmission in

disease persistence. However, we did not investigate how social structure determines spatial

transmission nor whether this level of complexity was necessary for capturing spatial

transmission dynamics. Thus, here we used the same framework to evaluate three models that

differed by social and spatial transmission process assumptions (described below) (Fig. 1). We

estimated transmission parameters and some other epidemiological and demographic parameters

as described below. With parameters from the fitted models we then predicted cases over time,

spatial spread over time, STKs, effective reproductive numbers over time, and age- and sex-

structure of infected individuals. All analyses were implemented in Matlab (Version R2016b,

The MathWorks, Inc., Natick, Massachusetts, United States). A full description of the individual-

based model and code for running it is given in Pepin et al. (2020). Below is an overview of the

approach with emphasis on differences from our previous work.

Landscape. We used a gridded landscape to allow heterogeneity in population density

across the landscape through density-dependent reproduction. We also allowed grid cell densities

to affect dispersal (and thus potentially spatial spread of ASFv) by preventing dispersal to grid

cells that were at carrying capacity. We chose a grid cell resolution of 5 x 5 km (25 km^2) because

it allowed fine-scaled heterogeneity in host density (close to home range size) while maintaining

reasonable computation time. The total landscape size was 120 x 50 km (6000 km^2), similar to

the 'infected' zone in eastern Poland. Grid cells each had a carrying capacity of 0.5 or 2

boars/km^2 (average density of 1.5 wild boar / km^2), which is similar to previous estimates of

local densities that were estimated to range from very low (<0.5 boar/km^2) in poor habitats to

high (>1.5 boar/km^2) in high quality habitats [43]. This level of heterogeneous boar density fit

the surveillance data better than homogenous densities of 1, 2 or 4 wild boar / km^2 [26].

Attributes and demographic processes. Individual-boar attributes were monitored and

updated at a daily time step. These included age, unique group identification, X and Y

coordinates of the home range centroid, grid-cell ID; and status of life, reproduction, and

infection. Thus, the distribution of wild boar locations was continuous but density was controlled

at the grid-cell level. The variable attributes changed based on time, age, group size, grid-cell

density, natal dispersal timing, and the pathogen transmission process. Fixed individual-level

attributes included sex, dispersal distance, dispersal age, and age at natural death, which were all

chosen from probability distributions [26].

Individual-boar status was updated by the following order of processes: daily movement

(defined by the contact processes described below) and pathogen transmission, natural mortality

(occurring according to the pre-set age), natal dispersal (occurring according to the pre-set age),

dispersal due to other factors (i.e., family groups becoming too large, single females searching

for groups; occurring based on current family group size), surveillance sampling (permanent

removal of hunter-harvested individuals and carcasses), conception (rates dependent on current

grid cell density), and new births (occurring with gestating females reach the end of their

gestation period). Fixed parameters included longevity (a data-based distribution), litter size (6),

age at reproductive maturity (180 days), minimum time between conception and farrowing (90

days), gestation time (115 days), age of natal dispersal (~Poisson(13 months) truncated between

10-24 months), dispersal distance (~Weibull(2.5,0.5)), maximum size of family groups (10),

incubation period for ASFv (~Poisson(4 days) truncated at 1), infectious period for ASFv

(~Poisson(5 days) truncated at 1), and disease-induced mortality (assumed to be fixed at 100%

lethality for infectious individuals). There were also fixed seasonal trends that varied monthly for

conception probability and carcass persistence that were based on data [44-47]. For example,

seasonal birth pulses influenced group size and dispersal events over time (because dispersal was

age-dependent), but we assumed that average daily contact distances were constant throughout

the year. Rationale and sources for the processes and parameters were derived from ecological

studies of wild boar and are described in Pepin et al. (2020).

Epidemiological states and processes. Epidemiological states for individual boar

included: susceptible, exposed, infectious, infectious carcass, non-infectious carcass, and

removed from the landscape. Mortality only occurred from the disease (leading to an infectious

carcass) or reaching the age of longevity (leading to an non-infectious carcass). We assumed the

disease was lethal in 100% of infectious individuals. The hunting process of alive individuals

caused direct removal from the landscape (no carcass). Our model also included multiple spatio-

temporal scales of spatial processes because the dispersal process (~Weibull(2.5,0.5) allowed for

longer-distance movements and occurred less frequently relative to the contact process that

occurred daily and mostly at shorter distances.

We compared three different forms of contact structure: 1) neighborhood (local

transmission only; local-only model), 2) distance distribution (exponential decay that includes

long-distance processes; distance-only model), and 3) distance distribution with social structure

(same as 2) but also allows for different transmission probabilities for within versus between

family groups; distance-social model) (Fig. 1). We tested these three models because they

represent increasing levels of ecological complexity in constraining spatial transmission. We

viewed the local-only model as the coarsest representation of constraining transmission both

spatially and within family groups. The local-only and distance-only models are common ways

of considering contact in space at population-level scales [48], whereas the distance-social model

incorporates heterogeneity due to social groups. For the local-only model, infectious individuals

could transmit to all susceptible individuals within a fixed radius with equal probability. The

radius of the local neighborhood was constant across individuals and time – thus similar to a

queen's neighbor effect (Eq. 1). For the distance-only model, infectious individuals could

transmit to all susceptible individuals on the landscape, but the probability of transmission

decayed with distance (Eq. 2). The distance-social model was the same as the distance-only

model, except that transmission rates varied due to both group membership and space -

individuals in the same family group had higher transmission rates with each other relative to

those among family groups (Eq. 3). In general, the daily force of infection (λ) for each contact

structure was defined as follows:

$$\lambda = \sum_{k=1}^{K}\sum_{j=1}^{J} I_{k,d} \begin{cases} S_j\beta_d, & x_{k,j} < \zeta \\ 0, & otherwise \end{cases} + \sum_{k=1}^{K}\sum_{j=1}^{J} I_{k,c} \begin{cases} S_j\beta_c, & x_{k,j} < \zeta \\ 0, & otherwise \end{cases} + \beta_{0,\{j\}} \qquad (1)$$

$$\lambda = \sum_{k=1}^{K}\sum_{j=1}^{J} I_{k,d} S_j\beta_d e^{-\alpha x_{k,j}} + \sum_{k=1}^{K}\sum_{j=1}^{J} I_{k,c} S_j\beta_c e^{-\alpha x_{k,j}} + \beta_{0,\{j\}} \qquad (2)$$

239
       $$\lambda = \sum_{k=1}^{K}\sum_{j=1}^{J} I_{k,d}(S_j\beta_d\, e^{-\alpha x_{k,j}} + S_{j,w}\beta_{w,d}) + \sum_{k=1}^{K}\sum_{j=1}^{J} I_{k,c}(S_j\beta_c e^{-\alpha x_{k,j}} + S_{j,w}\beta_{w,c}) + \beta_{0,\{Sj\}} \qquad (3)$$
241

Where ζ is a fixed local neighborhood (Eq. 1) delimiting the contact radius, $x_{i,j}$ is the distance

between infectious individual k (I_k) and susceptible individual j (S_j), α is the rate at which

transmission decays with distance (Eq. 2 and 3), *d* denotes alive individuals or direct

transmission, *c* denotes infectious carcasses or carcass-based transmission, β is the transmission

rate that is specific to the transmission mechanism (*d* or *c*), $\beta_{0,\{j\}}$ is the baseline rate at which re-

introduction occurs to susceptible individuals near the Eastern border ([49]), and *w* denotes

contact within the same family group whereas absence of *w* denotes contact among family

groups (Eq. 3).

Observation model

Because surveillance sampling only tests a small proportion of the total population of

wild boar in the region (< 2% monthly) it was important to calibrate the process model with an

observation model. Thus, we sampled the true pathogen transmission dynamics according to the

surveillance process that was used in Poland, i.e., alive individuals were available to be harvested

by hunters, and carcasses were available to be found for carcass sampling; then both types of

samples were evaluated to identify whether individuals/carcasses were infectious. As observed

negative samples could not be georeferenced to the grid cell level (they were only available at

the district level), we were not able to account for the spatial distribution of sampling accurately.

However, we used the temporal trends in the surveillance data to determine the number of

samples to collect per day by sampling the landscape at random but excluding wild boar < 6

months of age in hunter harvest (because they are typically not targeted by hunters; [50]) and

wild boar < 3 months of age in carcass sampling (because they are unlikely to be found due to

their small size and more rapid decay rates [51]). To determine the temporal sampling trends, we

calculated the relative number of boar sampled by hunters and carcass-sampling from the data

(number sampled on day *t*/maximum ever sampled separately for each method) to produce daily

trends in the proportion of the population sampled. Then we multiplied the trend data for each

method by the scaling factors (ρ_h and ρ_c) to determine the daily proportion of boar that would be

sampled (detection probability) by hunter harvesting or dead carcasses across the landscape at

random.

Model fitting and evaluation

Unknown parameters were estimated based on approximate Bayesian computation

(ABC) with rejection sampling as described in Pepin et al. (2020). For all models, estimated

parameters included: frequency of introduction at the eastern border ($\beta_{0,\{j\}}$), β_d, β_c, scaling

parameters on seasonal trends of hunted hosts (ρ_h) and carcass sampling (ρ_c), a scaling parameter

on seasonal trends in the length of carcass persistence on the landscape (π), and a scaling

parameter on seasonal patterns of host birth probabilities (θ). In addition, we estimated spatial

parameters that describe three different contact structures: 1) ξ (nearest-neighbor), 2) α (the

decay of contact probability with distance, and 3) $\beta_{w,d}$ and $\beta_{w,c}$ (direct and carcass-based

transmission rates for within-group contacts). Prior distributions are listed in Table S1 (with

restrictions: $\beta_d > \beta_c$, $\beta_{w,d} > \beta_d$, $\beta_{w,c} > \beta_c$) and were informed by movement and contact data [28, 29,

52, 53].

To sample across parameter space efficiently we used a Latin hypercube algorithm to

generate 979,592 parameter sets and then ran the model twice on each parameter set (for a total

of 1,959,184 iterations; or 2 chains of 979,592). β_d, β_c, and ρ_c were sampled on a log scale. To

improve computational efficiency, a two-tiered approach was used to estimate posterior

distributions of parameters. Simulations were terminated early if they were highly unrealistic

compared to observed data, specific criteria and rationale were: 1) landscape-wide host density <

20% of the initial density because observed changes in wild boar density were only minor in the

study during 2014-2016, 2) > 150 new cases per day because the maximum number observed per

month was < 20; 3) no new cases sampled for 6 months because there was only 1 month with no

cases detected after the first detection was made, or 4) > 300 total cases because that is more than

double the actual number of observed cases. We then only considered parameter sets for which

the simulation reached the end of the two-year time frame as candidate values for the posterior

distributions. The posterior distributions consisted of all unique parameter sets (considering both

chains) that were within the absolute distance of three metrics: the sum of absolute differences

between observed and simulated surveillance data for monthly cases from live and dead animals

(considered separately), and the maximum monthly Euclidian distance of cases from the eastern

border. Distance metric tolerance values were 48 for monthly cases from carcasses, 24 for

monthly cases from hunter-harvest samples, and 120 for maximum distance from the border.

This allowed average error rates of 2 (carcass) and 1 (hunter harvest) cases, and 5 km from the

border per month on average. These error rates represent levels of uncertainty that we expected

from the data sources in our system, sensitivity analyses revealed that less stringent error rates

would affect the posterior distribution estimates (data not shown), and more stringent error rates

would require restrictively large computational resources unless prior distributions are more

informed.

Average distance metrics for parameter sets from the posterior distribution were used to

evaluate goodness of fit along with R^2 values (squared correlation of observed and predicted case

and spatial distance trajectories, Table S1) and mean absolute error (MAE, Fig. 2). The

combination of these metrics were used for model selection and performance relative to one

another, although we were also interested in comparing how the different model structures

impacted parameter inference thus we examined output from all three models. For each fitted

model we predicted outbreak dynamics using 100 random samples from the posterior

distribution. The average of the 100 predictions was used to calculate R^2 and MAE. We also

tested the ability of our models to forecast ASFv dynamics by using the parameters estimated

from fits to the 2014-2015 data to predict the first 7 months of 2016 (Jan.-Jul.). We predicted

underlying STKs, effective reproductive number over time (R_e), and age-sex structure of cases

by simulating from the fitted models. Thus, STKs and R_e were model outputs. We calculated

STKs through tracking the distance between each transmission event and summarizing the

resulting distributions descriptively. Similarly, we calculated R_e by tracking the number of

transmissions to new susceptible hosts that each live infectious individual and infectious carcass

transmitted throughout their infectious periods and summarized daily R_e as means for individuals

at the start of their infectious period.

RESULTS

Parameter inference and model fit

The model with both social and spatial processes (distance-social model; Eq. 3)

qualitatively captured spatial spread better than the local-only (Eq. 1) and distance-only (Eq. 2)

models of spatial transmission, and the local-only model largely overestimated cases during the

largest peak (Fig. 2). Also, the posterior distribution of transmission probabilities were much

lower and more realistic for both distance models relative to the local-only model (see β_d and β_c

in Table S1). The inferred STK for each model revealed two distinct peaks symbolic of within-

and between-group transmission (Fig. 3) but predicted different amounts of within-group

transmission when within- and between-group transmission probabilities were allowed to vary.

The distance-social model predicted the highest amount of within group transmission (80%),

followed by the distance-only (60%) and local-only (30%) models (Fig. 3). For both distance

models, between-group transmission peaked between 0.5 km and 1 km, with a peak amount of

transmission events reaching 5% and 2% for distance-only and distance-social models,

respectively (Fig. 3). For the local-only model, between-group transmission events plateaued

between 1-1.5 km at a frequency of 0.2 before dropping rapidly to 0 around 1.5km (Fig. 3). The

inferred STKs for both distance models had long tails that indicated a low frequency of long-

distance pathogen dispersal (Fig. 3).

Impacts of model structure on epidemiological processes

The specification of spatial and social transmission processes in the model structure

resulted in different inferences of R_e. The local-only model predicted higher average R_e over

time (mean: 2.5 with 95% confidence interval: [1.8, 3.2]), followed by the distance-social (1.5

[1.1-2.0]), and then the distance-only (1.1 [1.0-1.3]) models, including both direct and carcass-

based transmission (Fig. 4). However, predictions from the distance-only model suggested R_e is

relatively homogenous over time, while the local-only and distance-social models predicted

much more variability, with R_e values reaching above a value of 4 on multiple occasions, and

above a value of 8 at least once (Fig. 4). The distance-social model predicted higher R_e during

annual birth pulses (Fig. 4). The local-only and distance-only models predicted lower

contributions of carcass-based transmission in overall R_e whereas the distance-social model

predicted more similar levels of each transmission mechanism (with carcass-based transmission

being slightly lower on average). All models predicted that the infected class is predominantly

composed of juveniles (<6 months of age; Fig. S1-S3), reflecting the age-structure in the

population. However, the local-only and distance-only models predicted a slight male-bias in

infected individuals while the distance-social model predicted a slight female bias (Fig. S1-S3).

DISCUSSION

Prediction of spatial pathogen dynamics is often challenged by partially observed data, multiple

pathogen introductions, and a limited understanding of host contact processes. Our approach

accounted for partially observed surveillance data and re-introductions while examining how

different dimensions of host contact heterogeneity (social and spatial processes) affect inference

of spatial transmission. Not accounting for social structure in spatial transmission led to very

different inferences of STKs and R_e, as well as the role of sex, birth pulses, and transmission

mechanisms in driving spatial spread. As STKs are used to plan resource allocation to

surveillance and control, reducing uncertainty in STK estimates is crucial for maximizing

efficiency and effectiveness of response plans [54]. Our results suggest that robust estimates of

STKs (and other key epidemiological quantities such as R_e) may require consideration of both

social and spatial processes for accurate prediction of spatial spread. Secondly, the striking

differences among model predictions in the role of carcass-based transmission and magnitude

and variation in R_e suggests that models that exclude social structure may provide different

ecological insights about which factors are most important in transmission and persistence of

ASFv (e.g., carcass versus direct transmission processes, males versus females, or family groups

versus independent males).

Spatially-explicit disease models often predict that local variations in group size can be

extremely important in driving pathogen transmission due to threshold effects [55], but that the

magnitude of these effects is context-specific. For example, an analysis that evaluated the impact

of social-network organization on disease spread in 47 species suggests that outbreaks of highly

contagious pathogens may be larger and persist longer in highly fragmented, gregarious species

when compared to species with hierarchical social systems that are more socially connected,

which in contrast, are more prone to epidemic outbreaks of low to moderately transmissible

pathogens due to high connectivity [1]. This suggests that highly fragmented social organization

might slow the rate of spatial disease spread. Our work supports this idea because the distance-

only model predicted more rapid spatial spread relative to the distance-social model, despite

similar natal dispersal distances. Also, in order for the distance-only model to fit the case data as

well as possible, a larger median decay rate parameter on the distance function was estimated

relative to the distance-social model, explaining the faster rate of spatial spread and less locally

dense STK relative to the distance-social model. However, similar types of social behavior can

impact disease spread differently [56]. For example, territorial behaviors have been shown to

reduce the spread of bovine tuberculosis in European badgers to adjacent social groups [57, 58],

increase the risk of macroparasite infection in African antelope [11], and play only a minor role

in the spread of pathogens like feline calcivirus or canine distemper virus in Serengeti lions [56].

This suggests that social structure may not always be important for accurately capturing spatial

spread dynamics, and that it is important to identify the most important dimensions (if any) of

social behavior that might modify spatial spread.

Evaluating the impact of social structure on disease spread is facilitated when social

interactions among hosts are known [24, 59], but in many cases social structure must be captured

through representing processes such as connectivity structure or group size because individual-

level interactions are unknown [60]. Previous work on raccoon rabies demonstrates the value of

integrating social structure and space use (natal dispersal and home range size) into predictions

of spatial disease spread and evaluating the effectiveness of control [7, 23]. Similarly, a

simulation study of pathogens that are epidemiologically similar to foot-and-mouth disease and

classical swine fever showed how realistic social structure can slow the spatial spread of disease

and lower persistence, and that this effect is disproportionately strong for acute infections [27].

Spatial disease spread can be represented using a number of modeling frameworks (spatially-

explicit deterministic models, correlation models, individual-based lattice models, and reaction-

diffusion models, etc.), but methods to account for social structure and space use currently in

estimates of STKs (as demonstrated here) remain limited, especially in wildlife disease systems

[3, 55, 61].

One way to account for social structure and space use concurrently is to use spatial network

models [17], but this approach makes it difficult to disentangle the relative effects of social

versus spatial processes on pathogen transmission, which is important for optimizing control

strategies (but see [60] for a novel approach that addresses this issue). In contrast, the field of

movement ecology has developed new strategies to account for heterogeneities due to animal

movement [3, 4] and social behavior [62, 63], but these methods remain underdeveloped for

application in disease ecology [3], and present an opportunity for future work. Development of

conventional methods that allow separate inference and understanding of the role of spatial and

social drivers of pathogen transmission dynamics is important not only for an improved

mechanistic understanding of spatial pathogen dynamics but also for improved risk assessment

and optimal control strategies by highlighting which processes (e.g., host clustering, movement

patterns) at what scale should be targeted for control [61].

Although IBMs can be computationally intensive, they can capture the heterogeneous traits

that make each population, disease, and sampling design unique [25]. As such, they are

advantageous for informing policy in many fields of epidemiology and public health that are

influenced by individuals, their behaviors, and the landscape [64]. The IBM framework

presented here can be used to improve predictions of spatial pathogen dynamics and risk

assessment through incorporating important real-world complexities such as social clustering,

sampling design, seasonal demographics, and pathogen re-introductions (see Figure 1). Though

we focus on the ASFv system, this framework can be readily applied to predict spatial pathogen

dynamics and understand the importance of social structure in other host-pathogen systems by

using system-specific data streams. In addition to computational time, a second challenge with

fitting IBMs to data can be their level of complexity and amount of stochastic variation. This can

lead to wide posterior distributions on parameters, without an extremely high number of

iterations (and in some cases even with a high number of iterations). As with any complex

Bayesian model, this risk can be reduced through using a combination of informative prior

distributions and screening results that are biologically unrealistic. During preliminary analyses

we allowed more vague prior distributions on the distance decay parameter (λ) and found that

the model could also fit the data well with a large value of λ but that these λ values were

unrealistically high for wild boar movement and also led to unrealistic values for transmission

probability and detection probabilities (based on expert opinion) thus we excluded these ranges

in the prior distributions of our final model fitting.

The distance-social model performed best at capturing both the case and spatial spread

dynamics while the distance-only model performed the worst because most spatial spread was

extremely local and the distance-only model was constrained to a monotonic distribution of

transmission distances. Thus, not accounting for social structure (i.e., allowing for more

transmission within than between groups) while allowing for some amount of long-distance

transmission (distance-only model) does a poorer job at inferring the STK because there was not

enough flexibility to capture the bi- (or multi-) modal STK that best describes the spatial

pathogen dynamics. Interestingly, the distance-social model performed the best and predicted the

highest proportion of within-group transmission events (~80%) while the local-only model

performed the next best but predicted the lowest proportion of within-group transmission events

(~30%). This suggests that local-only model was better than the distance-only model because it

restricted long-distance transmission. STKs from all models had two peaks symbolic of within-

group and between-group transmission even though the local-only and distance-only models did

not allow for different contact probabilities for within- versus among-group transmission because

all models included spatial clustering due to family groups. Although the local-only model

underestimated within-group transmission, and allowed much more between-group transmission,

all of the between-group transmission was very local which allowed the model predictions to be

closer to the distance-social model relative to the distance-only model. Together our results

suggest that long-distance transmission is important for capturing spatial spread dynamics but

that it is very rare.

Almost all transmission events for all models were within 1.5 km, with some rare events at

longer distances. These STK estimates can be used to establish control and surveillance zones for

wild boar in Poland. For example, carcass removal could be intensified within 1.5 km of a case

detection where most transmission is occurring, with depopulation and surveillance intensified

out to further distances (i.e., the tail of the STK) to contain further spread. A useful approach

could be to employ an adaptive radius that focuses intervention efforts within 99% (or more –

this should be validated with modeling) of the STK, but adapts surveillance based on real-time

surveillance. However, the precise recommendations will depend on how soon a detection is

made relative to where the infection front is currently. With ASFv travelling at 1-2 km per month

[29], the radii for high-intensity culling and surveillance would need to be increased by 1-2 km

for each month that detection has lagged behind the infection front, highlighting the importance

of accurate predictions of spatial spread. A longer lag time for detection will also amplify

challenges that arise from long-distance jumps highlighting that this process is especially

important to understand. One approach to account for detection lags and anticipated spread

would be to pre-determine multiple fixed radii from the STKs to delineate surveillance and

control zones. This would allow for more rapid re-distribution of control resources and better

targeting of surveillance resources for optimizing adaptive change based on current surveillance

data.

Hunting, culling, and other anthropogenic factors could influence wild boar movement and

disrupt social structure [65], thus altering STKs. While our model assumed that daily movement

dynamics (contact) remained constant throughout the study period, it did account for social

disruption. In cases where hunting left family group members alone, these individuals dispersed

to join the nearest group (i.e., implicitly accounting for the effects of social disruption on disease

spread). In terms of the potential effects of hunting or culling on daily movement, the behavioral

response of wild boar to hunting activities in Europe remains nuanced, with space-use changes

varying by ecological and hunting context [65]. However, feral swine culling practices in the US

have shown short-term increases in movement and home range shifts, which could transiently

increase ASFv transmission [66, 67], and suggests that their effects could merit further

exploration.

Long-distance jumps (> 100 km from the epizootic region) were observed on multiple

occasions in Poland after 2015 [37] and are thought to be due to human-mediated activities as

these distances are well beyond those travelled by wild boar naturally [68, 69]. However, our

model only included processes describing natural movements of wild boar (which explained

most of the transmission events during our study period) because we did not have data to

describe sources of human-induced long-distance transmission. Thus, developing estimates of

STKs that account for mechanisms or risk factors of long-distance dispersal remains an

important objective that can help target disease control efforts. Our approach allowed for some

longer-distance events (on the scale of natural wild boar movements) but we assumed a

monotonic functional form for contact distances, such that we did not infer the effects of spatial

contact processes occurring on multiple spatial scales (beyond within- versus between-group

transmission differences in the distance-social model). In order to infer spatial spread with later

surveillance data (i.e., 2016-present when longer-distance events occurred multiple times), it will

be important to incorporate other spatial mechanisms in the inference of the STKs, perhaps using

covariate data that can inform these long-distance processes. Such an approach would provide

refined recommendations for surveillance and control targets at longer distances.

We assumed that ASFv is lethal in 100% of cases, but recent evidence indicates that wild

boar may survive infection [36, 81, 82]. If some hosts survive ASFv they could facilitate ASFv

persistence by producing new susceptible hosts. However, the fraction of survivors is extremely

small for virulent strains of ASFv (as in our study area) thus it is unclear if these individuals

could realistically affect persistence. Modeling research to understand the potential role of

survivors in the enzootic dynamics of ASFv could be useful for identifying conditions where

survivors may alter the enzootic dynamics. We also assumed that all infections were acute (with

an average infectious period of 5 days). If some individuals were to become persistently infected,

those individuals would have increased opportunity to transmit the virus within and among social

groups and could presumably increase the average distance of pathogen transmission because

there would be more opportunity for contact among wild boar at further distances [83, 84].

However, for the virulent ASFv strains circulating in Poland, there is no evidence of persistently

infected wild boar that can shed ASFv in high enough concentrations to infect other wild boar

[83]. Nonetheless, surveillance for changes in virulence are important because they could change

STKs over time.

Our models inferred substantial variation in R_e over time due to seasonally varying carcass

persistence, with the distance-social model predicting the largest peaks in R_e due to increases in

carcass and ASFv persistence in cold weather [45, 74, 75]. Prior research the Baltic States and

Poland also indicated that the seasonality of ASFv cases in wild boar peaked in winter and

summer [76]. Although this trend was observed across the entire European Union, these studies

could not prove causality [77]. Results of our analysis suggest that carcass surveillance and

removal may be especially important during the colder months due to the potentially increased

role of carcass-based transmission.

Our estimates of R_e (ranging from 1.1-2.5 on average across models) due to different STK

assumptions were similar to an estimate of R_0 of ASFv in wild boar (1.13-3.77) in Russia [70].

However, differences between our models in estimates of R_e over time suggest carcass-based

transmission has different roles in driving ASFv transmission, which could influence identifying

the optimal control strategy considering that carcass search and removal is intensive [71]. The

local-only and distance-only models suggest that carcass-based transmission is lower than direct

transmission while the distance-social model suggests that the two types of transmission

occurred at similar frequencies. Although carcasses have longer infectious periods than the alive

phase during the cooler months, transmission probability given contact is lower and contact rates

are lower relative to live individuals because carcasses do not move [45, 72, 73]. These

limitations balance the advantage of potentially longer infectious periods. The distance-social

models lead to higher rates of carcass-based transmission because they predict that most

transmission events are extremely local. The fact that models with commonly used spatial

contact structures produce different inferences of the importance of different transmission

mechanisms when social structure is added emphasizes: 1) the importance of further developing

inference methods that account for social structure in spatial spreading, and 2) the need for

approaches that aim to reduce uncertainty in estimates of STKs, e.g., [9].

Interestingly, our models also predicted differences in the role of sex in transmission over

time. The distance-social model predicted a slight female bias because this model predicted that

most transmission events occurred within family groups, which are female-biased. In contrast,

the local-only and distance-only models predicted a slight male bias because they predicted more

between-group transmission and with a 50:50 sex ratio there are more independent males relative

to family groups. Although it is known that males will travel longer distances than females [78],

especially during mating season to seek out females, we did not account for this temporal

heterogeneity in dispersal. Considering these types of movement heterogeneities in future work

could be important for improving our understanding of which sex might present a higher risk of

ASFv transmission and persistence.

Although our STK estimates are based on wild boar ecology in eastern Poland they can

provide baseline guidance for planning control in other settings. Response plans in areas that are

ASFv-free often rely only on local host movement ecology because there are no local data on

how ASFv might transmit. Our estimates of STKs suggest that transmission of ASFv in wild

boar (or pigs) in other ecological settings might mainly occur over the closer daily movement

estimates within home ranges due to the social processes in this host species. To further refine

STK estimates for specific ecological contexts, parameters for the local demographic (e.g. litter

size, age at reproductive maturity, gestation time, longevity, seasonal birth patterns, landscape

carrying capacity), movement (e.g. natal dispersal distance and timing, daily home range

exploration), and social processes (maximum group size) can be used as inputs in our simulation

model with the Poland-derived estimates for transmission rates to explore how local ecological

factors might affect STKs or their uncertainty.

Considered separately, spatial and social processes can have similar impacts on pathogen

transmission dynamics. For example, social aggregation (e.g., family groups, herd living) and

spatial structuring can both restrict the spread of pathogens [1, 79]. However, our results

highlight that spatial and social processes can also have quite different impacts on

epidemiological quantities, especially estimates of STKs, R_e, the frequency of different

transmission mechanisms, and potential risk factors such as sex. Being able to appropriately infer

the role of these quantities is crucial for optimizing disease control strategies. When these

contact heterogeneities are inappropriately accounted for it can bias inference (e.g., [80]) and

potentially misguide policy decisions [54]. Moving forward, the field of disease ecology should

emphasize the development and use of methods that account for spatial and social contact

processes separately so that their relative roles in driving pathogen transmission dynamics can be

inferred and understood. This will allow uncertainties in contact processes to be appropriately

evaluated and incorporated into predictions of spatial spread [54], for monitoring designs to be

optimized, and for risk factors to be identified more accurately so that controls can be targeted to

the most important risk factors.

Acknowledgements

We would like to thank 5 anonymous reviewers that provided helpful comments that improved

clarity of the manuscript. KMP was supported by the United States Department of Agriculture,

Animal and Plant Health Inspection Service's National Feral Swine Damage Management

Program. AG was supported by the United States Department of Agriculture, Animal and Plant

Health Inspection Service's APHIS Science Fellowship. TP was supported by the National

Science Centre, Poland (grant number 2014/15/B/NZ9/01933) and Ministry of Agriculture,

Czech Republic (grant number QK1910462). We thank M. Łyjak, A. Kowalczyk, K. Śmietanka,

and G. Woźniakowski from Department of Swine Diseases, National Veterinary Research

Institute in Pulawy, Poland, for surveillance data. N. Selva provided valuable information on

carcass persistence time.

Figures

Fig 1. Schematic of methods and contact structures. Three different forms of contact structure

were fitted to ASFv surveillance data (1. and 2.): A) neighborhood (local transmission only;

local-only model), B) distance distribution (exponential decay that includes long-distance

processes; distance-only model), and C) distance distribution with social structure (same as B.

but also allows for different transmission probabilities for within versus between family groups;

distance-social model). Goodness-of-fit was determined for each of the three fitted models (3.).

Then the fitted models were used to simulate pathogen transmission dynamics (4.) and

epidemiological metrics were derived from the simulated output (5.). ζ is a fixed local

neighborhood (Eq. A) delimiting the contact radius, $x_{i,j}$ is the distance between infectious

individual *k* (*I$_k$*) and susceptible individual *j (S$_j$)*, α is the rate at which transmission decays with

distance (Eq. B and C), *d* denotes alive individuals or direct transmission, *c* denotes infectious

carcasses or carcass-based transmission, β is the transmission rate that is specific to the

transmission mechanism (*d* or *c*), and *w* denotes contact within the same family group whereas

absence of *w* denotes contact among family groups (Eq. C).

Fig. 2. Model fits to the observed surveillance data (black). Lines are the mean predictions from

100 simulations with each fitted model (see legend in B for color code). Shading around the lines

areas are 95% prediction intervals of the means. Shading on the right side of plots indicates the

time frame for out-of-sample predictions. MAE is mean absolute error between the trajectories of

626 observed and predicted monthly cases. Plots between the top and bottom panels show trends as a

627 proportion of maximum values for total samples collected (purple), births (light blue), and

628 carcass persistence time (green).

Fig. 3. Inferred spatial transmission kernels (STKs) for each model (see legend). The X-axis is

632 the distance between home range centroids of infectious and susceptible individuals for which

633 transmission occurred. Y-axis is the proportion of all transmission events. Lines are the means of

634 100 simulations using random samples from the posterior distributions of the fitted models.

Fig. 4. Effective reproduction number (R_e) over time for each model. Effective reproduction

number at a given time point was calculated as the average number of transmissions made

throughout the infectious period for individuals that initially became infectious on day *t* (where *t*

is a day on the X-axis). Dark lines are the means of 100 simulations using random samples from

the posterior distributions of the fitted models; shading indicates 95% prediction intervals of the

means. Overall means with 95% prediction intervals for each model and each transmission

642   mechanism (black: direct, red: carcass-based) were: A) 1.9 [1.4, 2.3], 0.6 [0.4, 0.9], B) 0.9 [0.8

643   1.0], 0.2 [0.2, 0.3], C) 0.9 [0.7, 1.1], 0.6 [0.4, 0.9]. Month 1 on the x-axis is January, 2014.

References

1.   Sah, P., J. Mann, and S. Bansal, *Disease implications of animal social network structure:*

a synthesis across social systems. Journal of Animal Ecology, 2018. **87**(3): p. 546-558.

2.   Riley, S., *Large-scale spatial-transmission models of infectious disease.* Science, 2007.

316(5829): p. 1298-1301.

3.   Dougherty, E.R., et al., *Going through the motions: incorporating movement analyses*

into disease research. Ecology letters, 2018. **21**(4): p. 588-604.

4.   Emch, M., et al., *Integration of spatial and social network analysis in disease*

transmission studies. Annals of the Association of American Geographers, 2012. **102**(5):

p. 1004-1015.

5.   Arthur, R.F., et al., *Contact structure, mobility, environmental impact and behaviour: the*

importance of social forces to infectious disease dynamics and disease ecology.

Philosophical Transactions of the Royal Society B: Biological Sciences, 2017.

372(1719): p. 20160454.

6.   Woodroffe, R., et al., *Social group size affects Mycobacterium bovis infection in*

European badgers (Meles meles). Journal of Animal Ecology, 2009: p. 818-827.

7.   McClure, K.M., et al., *Variation in host home range size decreases rabies vaccination*

effectiveness by increasing the spatial spread of rabies virus. Journal of Animal Ecology,

2020: p. 1375-1386.

8.   Habib, T.J., et al., *Modelling landscape effects on density–contact rate relationships of*

deer in eastern Alberta: implications for chronic wasting disease. Ecological Modelling,

2011. **222**(15): p. 2722-2732.

9.   Nunn, C.L., et al., *Infectious disease and group size: more than just a numbers game.*

Philosophical Transactions of the Royal Society B: Biological Sciences, 2015.

370(1669): p. 20140111.

10.   Daviews, C., et al., *Malaria infection rate of Amazonian primates increases with body*

weight and group size. Functional Ecology, 1991: p. 655-662.

11.   Ezenwa, V.O., *Host social behavior and parasitic infection: a multifactorial approach.*

Behavioral Ecology, 2004. **15**(3): p. 446-454.

12.   Rosengaus, R.B., et al., *Disease resistance: a benefit of sociality in the dampwood*

termite Zootermopsis angusticollis (Isoptera: Termopsidae). Behavioral Ecology and

Sociobiology, 1998. **44**(2): p. 125-134.

13.   Benincà, E., et al., *Trade-off between local transmission and long-range dispersal drives*

infectious disease outbreak size in spatially structured populations. PLOS Computational

Biology, 2020. **16**(7): p. e1008009.

14.   Boender, G.J., et al., *Risk maps for the spread of highly pathogenic avian influenza in*

poultry. PLoS Comput Biol, 2007. **3**(4): p. e71.

15.   Boender, G.J., et al., *Transmission risks and control of foot-and-mouth disease in The*

Netherlands: spatial patterns. Epidemics, 2010. **2**(1): p. 36-47.

16.   Gubbins, S., et al., *Inferences about the transmission of lumpy skin disease virus between*

herds from outbreaks in Albania in 2016. Preventive veterinary medicine, 2018: p. 1-6.

17.   Craft, M.E., *Infectious disease transmission and contact networks in wildlife and*

livestock. Philosophical Transactions of the Royal Society B: Biological Sciences, 2015.

370(1669): p. 20140107.

18.   Salje, H., D.A. Cummings, and J. Lessler, *Estimating infectious disease transmission distances using the overall distribution of cases.* Epidemics, 2016. **17**: p. 10-18.

19.   Ferguson, N.M., C.A. Donnelly, and R.M. Anderson, *The foot-and-mouth epidemic in Great Britain: pattern of spread and impact of interventions.* Science, 2001. **292**(5519): p. 1155-1160.

20.   Ferguson, N.M., C.A. Donnelly, and R.M. Anderson, *Transmission intensity and impact of control policies on the foot and mouth epidemic in Great Britain.* Nature, 2001. **413**(6855): p. 542-548.

21.   Keeling, M.J., et al., *Dynamics of the 2001 UK foot and mouth epidemic: stochastic dispersal in a heterogeneous landscape.* Science, 2001. **294**(5543): p. 813-817.

22.   Woolhouse, M., *How to make predictions about future infectious disease risks.* Philosophical Transactions of the Royal Society B: Biological Sciences, 2011. **366**(1573): p. 2045-2054.

23.   Rees, E.E., et al., *Modelling the effect of landscape heterogeneity on the efficacy of vaccination for wildlife infectious disease control.* Journal of Applied Ecology, 2013. **50**(4): p. 881-891.

24.   Shirley, M.D., et al., *Investigating the spatial dynamics of bovine tuberculosis in badger populations: evaluating an individual-based simulation model.* Ecological Modelling, 2003. **167**(1-2): p. 139-157.

25.   Willem, L., et al., *Lessons from a decade of individual-based models for infectious disease transmission: a systematic review (2006-2015).* BMC infectious diseases, 2017. **17**(1): p. 612.

26. Pepin, K.M., et al., *Ecological drivers of African swine fever virus persistence in wild*
boar populations: insight for control. Ecol Evolv., 2020. **10**: p. 2846–2859.

27. Pepin, K.M. and K.C. VerCauteren, *Disease-emergence dynamics and control in a*
socially-structured wildlife species. Scientific reports, 2016. **6**(1): p. 1-13.

28. Pepin, K.M., et al., *Contact heterogeneities in feral swine: implications for disease*
management and future research. Ecosphere, 2016. **7**(3): p. e01230.

29. Podgórski, T., M. Apollonio, and O. Keuling, *Contact rates in wild boar populations:*
Implications for disease transmission. The Journal of Wildlife Management, 2018. **82**(6):
p. 1210-1218.

30. Arias, M., et al., *Gaps in African swine fever: Analysis and priorities.* Transboundary and
emerging diseases, 2018. **65**: p. 235-247.

31. Pepin, K.M., et al., *Predicting spatial spread of rabies in skunk populations using*
surveillance data reported by the public. PLoS neglected tropical diseases, 2017. **11**(7):
p. e0005822.

32. Chipperfield, J.D., et al., *On the approximation of continuous dispersal kernels in*
discrete-space models. Methods in Ecology and Evolution, 2011. **2**(6): p. 668-681.

33. Ypma, R.J., et al., *Unravelling transmission trees of infectious diseases by combining*
genetic and epidemiological data. Proceedings of the Royal Society B: Biological
Sciences, 2012. **279**(1728): p. 444-450.

34. Kamath, P.L., et al., *Genomics reveals historic and contemporary transmission dynamics*
of a bacterial disease among wildlife and livestock. Nature Communications, 2016. **7**(1):
p. 1-10.

35.   Price, S.J., et al., *Reconstructing the emergence of a lethal infectious disease of wildlife*

supports a key role for spread through translocations by humans. Proceedings of the

Royal Society B: Biological Sciences, 2016. **283**(1839): p. 20160952.

36.   Frant, M., et al., *African swine fever virus (ASFV) in Poland: Prevalence in a wild boar*

population (2017–2018). Veterinární medicína, 2020. **65**(4): p. 143-158.

37.   Mazur-Panasiuk, N. and G. Woźniakowski, *The unique genetic variation within the*

O174L gene of Polish strains of African swine fever virus facilitates tracking virus origin.

Archives of virology, 2019. **164**(6): p. 1667-1672.

38.   Chapman, D.A., et al., *Genomic analysis of highly virulent Georgia 2007/1 isolate of*

African swine fever virus. Emerging infectious diseases, 2011. **17**(4): p. 599.

39.   Authority, E.F.S., et al., *Epidemiological analyses of African swine fever in the European*

Union (November 2018 to October 2019). EFSA Journal, 2020. **18**(1): p. e05996.

40.   Guinat, C., et al., *Transmission routes of African swine fever virus to domestic pigs:*

current knowledge and future research directions. The Veterinary Record, 2016.

178(11): p. 262.

41.   Śmietanka, K., et al., *African swine fever epidemic, Poland, 2014–2015.* Emerging

infectious diseases, 2016. **22**(7): p. 1201.

42.   Woźniakowski, G., et al., *Current status of African swine fever virus in a population of*

wild boar in eastern Poland (2014-2015). Archives of virology, 2016. **161**(1): p. 189-

195.

43.   Borowik, T., T. Cornulier, and B. Jędrzejewska, *Environmental factors shaping ungulate*

abundances in Poland. Acta Theriologica, 2013. **58**(4): p. 403-413.

44. Rosell, C., F. Navas, and S. Romero, *Reproduction of wild boar in a cropland and coastal wetland area: implications for management.* Animal Biodiversity and Conservation, 2012. **35**(2): p. 209-217.

45. Selva, N., et al., *Factors affecting carcass use by a guild of scavengers in European temperate woodland.* Canadian Journal of Zoology, 2005. **83**(12): p. 1590-1601.

46. Ježek, M., et al., *Reproductive and morphometric characteristics of wild boar (Sus scrofa) in the Czech Republic.* Journal of forest science, 2011. **57**(7): p. 285-292.

47. Bieber, C. and T. Ruf, *Population dynamics in wild boar Sus scrofa: ecology, elasticity of growth rate and implications for the management of pulsed resource consumers.* Journal of Applied Ecology, 2005. **42**(6): p. 1203-1213.

48. Keeling, M.J. and P. Rohani, *Modeling infectious diseases in humans and animals*. 2008, NJ, USA: Princeton University Press.

49. Sjöberg, M., B. Albrectsen, and J. Hjältén, *Truncated power laws: a tool for understanding aggregation patterns in animals?* Ecology letters, 2000. **3**(2): p. 90-94.

50. Massei, G., et al., *Wild boar populations up, numbers of hunters down? A review of trends and implications for Europe.* Pest management science, 2015. **71**(4): p. 492-500.

51. Matuszewski, S., et al., *Effect of body mass and clothing on decomposition of pig carcasses.* International journal of legal medicine, 2014. **128**(6): p. 1039-1048.

52. Podgórski, T., et al., *Spatiotemporal behavioral plasticity of wild boar (Sus scrofa) under contrasting conditions of human pressure: primeval forest and metropolitan area.* Journal of Mammalogy, 2013. **94**(1): p. 109-119.

53. Kay, S.L., et al., *Quantifying drivers of wild pig movement across multiple spatial and temporal scales.* Movement ecology, 2017. **5**(1): p. 14.

54. Shea, K., et al., *Adaptive management and the value of information: learning via*

intervention in epidemiology. PLoS biology, 2014. **12**(10): p. e1001970.

55. Gudelj, I. and K. White, *Spatial heterogeneity, social structure and disease dynamics of*

animal populations. Theoretical population biology, 2004. **66**(2): p. 139-149.

56. Craft, M.E., et al., *Disease transmission in territorial populations: the small-world*

network of Serengeti lions. Journal of the Royal Society Interface, 2011. **8**(59): p. 776-

786.

57. Donnelly, C.A., et al., *Positive and negative effects of widespread badger culling on*

tuberculosis in cattle. Nature, 2006. **439**(7078): p. 843-846.

58. McDonald, J.L., A. Robertson, and M.J. Silk, *Wildlife disease ecology from the*

individual to the population: Insights from a long-term study of a naturally infected

European badger population. Journal of Animal Ecology, 2018. **87**(1): p. 101-112.

59. Firestone, S.M., et al., *Adding the spatial dimension to the social network analysis of an*

epidemic: Investigation of the 2007 outbreak of equine influenza in Australia. Preventive

veterinary medicine, 2012. **106**(2): p. 123-135.

60. Manlove, K., et al., *The ecology of movement and behaviour: a saturated tripartite*

network for describing animal contacts. Proceedings of the Royal Society B: Biological

Sciences, 2018. **285**(1887): p. 20180670.

61. Riley, S., et al., *Five challenges for spatial epidemic models.* Epidemics, 2015. **10**: p. 68-

71.

62. Langrock, R., et al., *Modelling group dynamic animal movement.* Methods in Ecology

and Evolution, 2014. **5**(2): p. 190-199.

63.   Spiegel, O., et al., *Socially interacting or indifferent neighbours? Randomization of movement paths to tease apart social preference and spatial constraints.* Methods in Ecology and Evolution, 2016. **7**(8): p. 971-979.

64.   Belsare, A.V., et al., *An agent-based framework for improving wildlife disease surveillance: A case study of chronic wasting disease in Missouri white-tailed deer.* Ecological Modelling, 2020. **417**: p. 108919.

65.   Merli, E., et al., *Wild boar under fire: the effect of spatial behaviour, habitat use and social class on hunting mortality.* Journal of Zoology, 2017. **303**(2): p. 155-164.

66.   Campbell, T.A., D.B. Long, and B.R. Leland, *Feral swine behavior relative to aerial gunning in southern Texas.* The Journal of Wildlife Management, 2010. **74**(2): p. 337-341.

67.   Sodeikat, G. and K. Pohlmeyer, *Temporary home range modifications of wild boar family groups (Sus scrofa L.) caused by drive hunts in Lower Saxony (Germany).* Zeitschrift für Jagdwissenschaft, 2002. **48**(1): p. 161-166.

68.   Chenais, E., et al., *Epidemiological considerations on African swine fever in Europe 2014–2018.* Porcine health management, 2019. **5**(1): p. 1-6.

69.   Authority, E.F.S., et al., *Epidemiological analyses of African swine fever in the Baltic States and Poland: (Update September 2016–September 2017).* EFSA Journal, 2017. **15**(11): p. e05068.

70.   Iglesias, I., et al., *Reproductive ratio for the local spread of African swine fever in wild boars in the Russian Federation.* Transboundary and emerging diseases, 2016. **63**(6): p. e237-e245.

71.   Lange, M., V. Guberti, and H.H. Thulke, *Understanding ASF spread and emergency*

825   *control concepts in wild boar populations using individual-based modelling and spatio-*

826   *temporal surveillance data.* EFSA Supporting Publications, 2018. **15**(11): p. 1521E.

827   72.   Probst, C., et al., *Behaviour of free ranging wild boar towards their dead fellows:*

828   *potential implications for the transmission of African swine fever.* Royal Society open

829   science, 2017. **4**(5): p. 170054.

830   73.   Eble, P., et al., *Transmission of African Swine Fever Virus via carrier (survivor) pigs*

831   *does occur.* Veterinary microbiology, 2019. **237**: p. 108345.

832   74.   Probst, C., et al., *The potential role of scavengers in spreading African swine fever*

833   *among wild boar.* Scientific reports, 2019. **9**(1): p. 1-13.

834   75.   Davies, K., et al., *Survival of African swine fever virus in excretions from pigs*

835   *experimentally infected with the Georgia 2007/1 isolate.* Transboundary and emerging

836   diseases, 2017. **64**(2): p. 425-431.

837   76.   Authority, E.F.S., et al., *Epidemiological analyses of African swine fever in the Baltic*

838   *States and Poland: (Update September 2016–September 2017).* EFSA Journal, 2017.

839   **15**(11): p. e05068.

840   77.   Authority, E.F.S., et al., *Epidemiological analyses of African swine fever in the European*

841   *Union (November 2017 until November 2018).* EFSA Journal, 2018. **16**(11): p. e05494.

842   78.   Podgórski, T., M. Scandura, and B. Jędrzejewska, *Next of kin next door–philopatry and*

843   *socio-genetic population structure in wild boar.* Journal of Zoology, 2014. **294**(3): p.

844   190-197.

845   79.   Keeling, M.J., *The effects of local spatial structure on epidemiological invasions.*

846   Proceedings of the Royal Society of London. Series B: Biological Sciences, 1999.

847   **266**(1421): p. 859-867.

848   80.   Ames, G.M., et al., *Using network properties to predict disease dynamics on human*

849   *contact networks.* Proceedings of the Royal Society B-Biological Sciences, 2011.

850   **278**(1724): p. 3544-3550.

851   81.   Pautienius, A., et al., *Prevalence and spatiotemporal distribution of African swine fever*

852   *in Lithuania, 2014–2017.* Virology journal, 2018. **15**(1): p. 177.

853   82.   Pejsak, Z., et al., *Four years of African swine fever in Poland. New insights into*

854   *epidemiology and prognosis of future disease spread.* Polish Journal of Veterinary

855   Sciences, 2018: p. 835–841-835–841.

856   83.   Ståhl, K., et al., *Lack of evidence for long term carriers of African swine fever virus-a*

857   *systematic review.* Virus research, 2019. **272**: p. 197725.

858   84.   O'Neill, X., et al., *Modelling the transmission and persistence of African swine fever in*

859   *wild boar in contrasting European scenarios.* Scientific reports, 2020. **10**(1): p. 1-10.

860   85.   Wilkinson, D., et al., *Cost-benefit analysis model of badger (Meles meles) culling to*

861   *reduce cattle herd tuberculosis breakdowns in Britain, with particular reference to*

862   *badger perturbation.* Journal of Wildlife Diseases, 2009. **45**(4): p. 1062-1088.

863   86.   Becker, D.J. and R.J. Hall, *Heterogeneity in patch quality buffers metapopulations from*

864   *pathogen impacts.* Theoretical ecology, 2016. **9**(2): p. 197-205.

865   87.   White, L.A., J.D. Forester, and M.E. Craft, *Dynamic, spatial models of parasite*

866   *transmission in wildlife: Their structure, applications and remaining challenges.* Journal

867   of Animal Ecology, 2018. **87**(3): p. 559-580.

Supplemental Material - **Social structure defines spatial transmission of African swine fever in wild boar**

Kim M. Pepin[1,*], Andrew Golnar[1], and Tomasz Podgórski[2,3]

4.   National Wildlife Research Center, USDA, APHIS, Wildlife Services, 4101 Laporte Ave., Fort Collins, CO, 80526.
5.   Mammal Research Institute, Polish Academy of Sciences, Stoczek 1, 17-230 Białowieża, Poland
6.   Department of Game Management and Wildlife Biology, Faculty of Forestry and Wood Sciences, Czech University of Life Sciences, Kamýcká 129, 165 00 Praha 6, Czech Republic

*Corresponding author: Kim Pepin, National Wildlife Research Center, United States Department of Agriculture, 4101 Laporte Ave., Fort Collins, CO, 80526; kim.m.pepin@aphis.usda.gov

Contents

Table S1.	Contact structure:	Local-only	Distance-only	Distance-social
	R^2			
Monthly cases	in sample	0.57 ± 0.0059	0.51 ± 0.0054	0.55 ± 0.0060
(Median of the R^2's for 1000 individual time series $\pm$ 95% confidence interval)	all	0.48 ± 0.0054	0.43 ± 0.0046	0.47 ± 0.0049
Monthly distance from border	in sample	0.28 ± 0.018	0.29 ± 0.017	0.31 ± 0.018
(Median of the R^2's for 1000 individual time series $\pm$ 95% confidence interval)	all	0.19 ± 0.016	0.14 ± 0.011	0.28 ± 0.011
Monthly cases	in sample	0.65	0.57	0.63
(R^2 of median values from 1000 points at each month)	all	0.55	0.48	0.53
Monthly distance from border	in sample	0.53	0.49	0.53
(R^2 of median values from 1000 points at each month)	all	0.49	0.38	0.55
	Distance metrics[a]			
Median absolute error in monthly live cases $\pm$ 95% confidence interval	in sample	20 ± 0.8	20 ± 0.9	20 ± 0.5
Median absolute error in monthly carcass cases $\pm$ 95% confidence interval	in sample	44 ± 1.4	43 ± 2.8	44 ± 0.8
Median absolute error in monthly distance from border $\pm$ 95% confidence interval	in sample	109.5 ± 5.0	111.7 ± 5.7	106.6 ± 3.0
Number of values in posterior distribution		16 / 1,959,184 = 0.00082%	7 / 1,959,184 = 0.00036%	53 / 1,959,184 = 0.0027%
	Uniform Priors	95% credible intervals of posterior distributions		
Probability of direct transmission given proximity (β^d)	$[0.0001,1]$[b]	$[0.0046, 1.0]$	$[0.039, 0.15]$	$[0.0003, 0.18]$
Probability of carcass-based transmission given proximity (β^c)	$[0.0001,1]$[b]	$[0.0002, 0.53]$	$[0.0038, 0.029]$	$[0.0001, 0.029]$
Annual frequency of spillover from neighboring country (ϕ)	$[0,60]$[c]	$[13, 59]$	$[9, 60]$	$[6, 59]$
Detection probability in hunted boar samples (ρ^h)	$[0.0005,0.1]$[b]	$[0.0006, 0.012]$	$[0.0007, 0.021]$	$[0.0007, 0.017]$
Detection probability in carcass samples (ρ^c)	$[0.0005,0.8]$[b]	$[0.012, 0.080]$	$[0.017, 0.045]$	$[0.014, 0.30]$
Scaling parameter on seasonal trends in carcass persistence (π)	$[0.1,1.5]$	$[0.58, 1.47]$	$[0.77, 1.41]$	$[0.17, 1.47]$
Scaling parameter on seasonal trends in conception probability (θ)	$[0.5,6]$	$[0.93, 5.73]$	$[1.31, 5.87]$	$[0.67, 5.88]$
Constant contact radius (ζ)	$[0.5,5]$	$[1.09, 3.35]$	NA	NA
Rate parameter for decay of contact probability with distance (λ)	$[0.1,2.5]$	NA	$[1.62, 2.49]$	$[0.05, 2.49]$
Probability of direct transmission given contact is in the same group ($\beta^{w,d}$)	$[0.01,1]$	NA	NA	$[0.077, 0.98]$
Probability of carcass-based transmission given contact is in the same group ($\beta^{w,c}$)	$[0.001,1]$	NA	NA	$[0.14, 0.97]$

[a]Median distance metrics $\pm$ 95% confidence intervals for 1000 simulations from the posterior distribution.

[b]These prior distributions were sampled on a natural log scale.

[c]0 indicates one introduction ever whereas values > 0 indicate the number of introductions / year.

Figure S1. The demographic dynamics of African swine fever virus based on the local-only model. Top plots show the frequency of infection in wild boar of different age classes (left) and sexes (right). Each line is a prediction from a separate sample of the posterior distribution of the fitted exponential decay & social structure model (100 trajectories in total). Bottom plots show the corresponding age class and sex distributions of infection over all time.

Figure S2. The demographic dynamics of African swine fever virus based on the distance-only model. Top plots show the frequency of infection in wild boar of different age classes (left) and sexes (right). Each line is a prediction from a separate sample of the posterior distribution of the fitted exponential decay & social structure model (100 trajectories in total). Bottom plots show the corresponding age class and sex distributions of infection over all time.

Figure S3. The demographic dynamics of African swine fever virus based on the distance-social model. Top plots show the frequency of infection in wild boar of different age classes (left) and sexes (right). Each line is a prediction from a separate sample of the posterior distribution of the fitted exponential decay & social structure model (100 trajectories in total). Bottom plots show the corresponding age class and sex distributions of infection over all time.